



HAL
open science

The status of strontium in biological apatites: an XANES/EXAFS investigation

Dominique Bazin, Arnaud Dessombz, Christelle Nguyen, Hang Korng Ea, Frédéric Lioté, John Rehr, Christine Chappard, Stephan Rouzière, Dominique Thiaudière, Solen Reguer, et al.

► To cite this version:

Dominique Bazin, Arnaud Dessombz, Christelle Nguyen, Hang Korng Ea, Frédéric Lioté, et al.. The status of strontium in biological apatites: an XANES/EXAFS investigation. *Journal of Synchrotron Radiation*, 2014, 21 (Pt 1), pp.136-42. 10.1107/S1600577513023771 . hal-00994761

HAL Id: hal-00994761

<https://hal.sorbonne-universite.fr/hal-00994761v1>

Submitted on 27 May 2014

HAL is a multi-disciplinary open access archive for the deposit and dissemination of scientific research documents, whether they are published or not. The documents may come from teaching and research institutions in France or abroad, or from public or private research centers.

L'archive ouverte pluridisciplinaire **HAL**, est destinée au dépôt et à la diffusion de documents scientifiques de niveau recherche, publiés ou non, émanant des établissements d'enseignement et de recherche français ou étrangers, des laboratoires publics ou privés.

The status of strontium in biological apatites: an XANES/EXAFS investigation

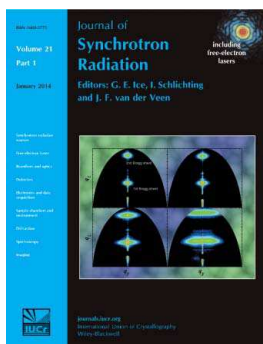
Dominique Bazin, Arnaud Dessombz, Christelle Nguyen, Hang Korng Ea,
Frédéric Lioté, John Rehr, Christine Chappard, Stephan Rouzière,
Dominique Thiaudière, Solen Reguer and Michel Daudon

J. Synchrotron Rad. (2014). **21**, 136–142

Copyright © International Union of Crystallography

Author(s) of this paper may load this reprint on their own web site or institutional repository provided that this cover page is retained. Reproduction of this article or its storage in electronic databases other than as specified above is not permitted without prior permission in writing from the IUCr.

For further information see <http://journals.iucr.org/services/authorrights.html>



Synchrotron radiation research is rapidly expanding with many new sources of radiation being created globally. Synchrotron radiation plays a leading role in pure science and in emerging technologies. The *Journal of Synchrotron Radiation* provides comprehensive coverage of the entire field of synchrotron radiation research including instrumentation, theory, computing and scientific applications in areas such as biology, nanoscience and materials science. Rapid publication ensures an up-to-date information resource for scientists and engineers in the field.

Crystallography Journals **Online** is available from journals.iucr.org

The status of strontium in biological apatites: an XANES/EXAFS investigation

Dominique Bazin,^{a,*} Arnaud Dessombz,^b Christelle Nguyen,^{c,d} Hang Korng Ea,^{c,e} Frédéric Lioté,^{c,e} John Rehr,^f Christine Chappard,^g Stephan Rouzière,^b Dominique Thiaudière,^h Solen Reguer^h and Michel Daudonⁱ

^aCNRS-LCMCP-UPMC, Collège de France, 11 place Marcelin Berthelot, 75231 Paris Cedex 05, France, ^bLaboratoire de Physique des Solides, Université Paris XI, Bâtiment 510, 91405 Orsay, France, ^cINSERM, UMR-S 606, Lariboisière Hospital, F-75010 Paris, France, ^dUniversité Paris Diderot, Sorbonne Paris Cité, F-75205 Paris, France, ^eRheumatology Department, AP-HP, Hôpital Lariboisière, Centre Viggo Petersen, Pôle Appareil Locomoteur, F-75010 Paris, France, ^fDepartment of Physics, University of Washington, Seattle, WA 98195, USA, ^gB2OA, UMR 7052 CNRS, Université Paris Diderot, 10 avenue de Verdun, F-75010 Paris, France, ^hSynchrotron SOLEIL, L'Orme des Merisiers, Saint-Aubin, BP 48, 91192 Gif-sur-Yvette, France, and ⁱService d'Explorations Fonctionnelles, AP-HP, Hôpital Tenon, 4 rue de la Chine, F-75020 Paris, France. *E-mail: dominique.bazin@u-psud.fr

Osteoporosis represents a major public health problem through its association with fragility fractures. The public health burden of osteoporotic fractures will rise in future generations, due in part to an increase in life expectancy. Strontium-based drugs have been shown to increase bone mass in postmenopausal osteoporosis patients and to reduce fracture risk but the molecular mechanisms of the action of these Sr-based drugs are not totally elucidated. The local environment of Sr²⁺ cations in biological apatites present in pathological and physiological calcifications in patients without such Sr-based drugs has been assessed. In this investigation, X-ray absorption spectra have been collected for 17 pathological and physiological calcifications. These experimental data have been combined with a set of numerical simulations using the *ab initio* FEFF9 X-ray spectroscopy program which takes into account possible distortion and Ca/Sr substitution in the environment of the Sr²⁺ cations. For selected samples, Fourier transforms of the EXAFS modulations have been performed. The complete set of experimental data collected on 17 samples indicates that there is no relationship between the nature of the calcification (physiological and pathological) and the adsorption mode of Sr²⁺ cations (simple adsorption or insertion). Such structural considerations have medical implications. Pathological and physiological calcifications correspond to two very different preparation procedures but are associated with the same localization of Sr²⁺ versus apatite crystals. Based on this study, it seems that for supplementation of Sr at low concentration, Sr²⁺ cations will be localized into the apatite network.

Keywords: Ca phosphate apatites; physiological calcifications; pathological calcifications; Fourier transform infrared spectroscopy; X-ray absorption spectroscopy; strontium environment.

© 2014 International Union of Crystallography

1. Introduction

As underlined in recent reviews and publications, osteoporosis, which mainly occurs during aging and after menopause, is characterized by an increase in bone fragility due to low bone mass and deterioration of bone quality (Meunier *et al.*, 2004; Rochefort *et al.*, 2010; Moyad, 2003). Femoral neck, humeral and vertebral fractures are the hallmark of severe

osteoporosis, although fracture risk in other bones is also increased in the presence of osteoporosis.

Current therapy for osteoporosis includes dietary supplementation of calcium and vitamin D, in addition to treatment with estrogens, oral or parenteral bisphosphonates, selective estrogen receptor modulators, teriparatide, strontium ranelate or, even more recently, denosumab, according to National recommendations and rules (Pinkerton & Dalkin, 2007;

Kanakamani & Tandon, 2008). The only strontium-based drug, strontium ranelate, has also shown anti-fracture efficacy in the treatment of postmenopausal osteoporosis (Marie, 2005; Cortet, 2011).

A starting point for understanding the role of strontium from a physicochemistry point of view is given by a description of the local order around Sr atoms in order to assess its localization *versus* hydroxyapatite (HAP), the mineral part of the bone (Bazin *et al.*, 2009a). The different structural hypotheses (Fig. 1) regarding the localization of Sr²⁺ cations in bone which take into account the physicochemistry of biological apatite have been presented previously (Bazin *et al.*, 2011; Elliott, 1994). These structural hypotheses take into account the different structural specificities of biological apatites (Vallet-Regi & Gonzalez-Calbet, 2002; Penel *et al.*, 1998; Rey *et al.*, 1995, 2007).

The first structural hypothesis corresponds to Sr²⁺ cations surrounded only by oxygen atoms which are adsorbed at the surface of collagen or apatite (hypothesis 1 in Fig. 1). Note that Sr²⁺ cations may have various coordination numbers with oxygen in solids and aqueous solutions (Persson *et al.*, 1995; O'Day *et al.*, 2000). Sr²⁺ cations could also be engaged in the hydrated poorly crystalline apatite region present at the surface of calcium phosphate nanocrystals (hypothesis 2 in Fig. 1). Finally, a substitution could occur between Sr²⁺ cations and Ca²⁺ cations inside Ca phosphate nanocrystals on either crystallographic site (I) or (II) (hypothesis 3 in Fig. 1).

As mentioned previously, we should recognize that, either for nanometer-scale materials or due to the low content of trace elements inside physiological or pathological apatites, diffraction techniques have to be completed by a local structural investigation through X-ray absorption spectroscopy (XAS) (Guinier, 1956; Li *et al.*, 2010; Sayers *et al.*, 1970; Bazin *et al.*, 2006, 2012; Bazin & Daudon, 2012). It should be noted that XAS is insensitive to polydispersity (Moonen *et al.*, 1995). As emphasized previously (Bazin *et al.*, 2012), XAS encompasses both X-ray absorption near-edge structure (XANES) and extended X-ray absorption fine structure (EXAFS). Several previous investigations have shown that XAS brings invaluable structural and electronic information regarding the local environment of specific elements such as Ca²⁺ (Eichert *et al.*, 2005; Sepulcre *et al.*, 2004; Carpentier *et al.*, 2010; Laur-

Table 1

Composition given by FTIR and the origin of the calcifications.

CA = carbonated calcium hydroxylapatite; MAP = magnesium ammonium phosphate (struvite); Prot = protein; C1 = whewellite; C2 = weddellite; ACCP = amorphous carbonated calcium phosphate; TGL = triglyceride; WK = whitlockite.

Sample	Origin	Composition
N15116	Kidney	87% CA, 7% Prot, 4% C2, 2% C1
N15203	Kidney	91% CA, 6% Prot, 3% C1
N13083	Kidney	90% CA, 6% Prot, 4% C1
N3289	Prostate	75% CA, 14% C2, 9% C1, 1% MAP, 1% Prot
N60592	Prostate	74% CA, 9% Prot, 7% WK, 5% ACCP, 3% C1, 2% C2
N11823	Kidney	66% CA, 30% ACCP, 4% Prot
N13066	Kidney	79% CA, 15% ACCP, 4% Prot, 2% C1
N13086	Prostate	84% CA, 12% Prot, 4% C1
N15048	Kidney	87% CA, 6% C1, 4% C2, 3% Prot
N17161	Bladder	61% CA, 30% ACCP, 8% Prot, 1% C1

encin *et al.*, 2011) or trace elements such as Zn²⁺ (Bazin *et al.*, 2009b; Yu *et al.*, 2009; Tang *et al.*, 2009; Dessombz *et al.*, 2013) or Sr²⁺ (Bazin *et al.*, 2011; Palmer *et al.*, 1996; McKeown *et al.*, 2003; Seward *et al.*, 1999; Murata *et al.*, 2012; Yachandra & Yano, 2011; Martin *et al.*, 2012) in pathological calcifications or in other materials.

In a previous study (Bazin *et al.*, 2011), the XANES part of the absorption spectra as well as numerical simulations using the *ab initio* FEFF9 X-ray spectroscopy program (Rehr *et al.*, 2009; Bazin & Rehr, 2003) have been used to assess the localization of Sr²⁺ cations in these biological apatites. This investigation seems to indicate that a simple adsorption of Sr²⁺ cations exists in the case of pathological calcifications while an insertion of Sr²⁺ is observed at least for one physiological calcification (bone 1). For the other three physiological samples, bones 2, 3 and 4, the evidence is much weaker.

In this investigation, 17 pathological and physiological calcifications have been considered. A new set of numerical simulations using the *ab initio* FEFF9 X-ray spectroscopy program have been performed in order to take into account local lattice distortion. Moreover, for selected samples, Fourier transforms of the EXAFS modulations have been performed. The ultimate aim of this study is to confirm, through X-ray absorption experiments at the Sr *K*-edge and numerical simulations, the possible relationship between the adsorption mode of Sr²⁺ cations and the nature of the calcification.

2. Materials and methods

The biological samples (Table 1) analysed in the present investigation came from two different hospitals. More precisely, kidney stones and bones were obtained from Necker Hospital and Lariboisière Hospital (both Paris), respectively. Regarding an ethics statement, we followed the usual procedures (Dessombz *et al.*, 2011, 2012). All participants gave verbal consent, documented by the researchers, for use of their samples. The samples were examined without knowledge of the name of the patient or other identifying data. Ethical approval for the study was obtained from the ethics committee

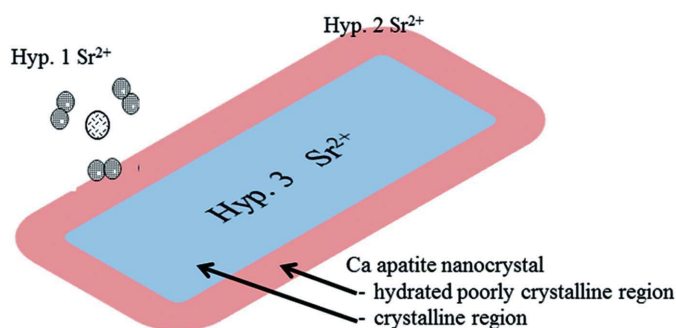


Figure 1
Schematic representation of the three structural hypotheses assessed in this study regarding the localization of Sr²⁺ cations in HAP.

of Tenon and Lariboisiere Hospital. The investigation conformed to the principles of the Declaration of Helsinki.

All of the samples have been characterized by Fourier transform infrared (FTIR) spectroscopy using the Vector 22 (Bruker Spectrospin, Wissembourg, France) FTIR spectrometer. Data were collected in the absorption mode between 4000 and 400 cm^{-1} with a resolution of 4 cm^{-1} (Estepa & Daudon, 1997). Compositions for the pathological calcifications given by FTIR spectroscopy are gathered in Table 1. For all the calcifications, apatite is the major component, and also for the bones (bone samples 1 to 7).

All of the samples were investigated on the DIFFABS beamline at synchrotron SOLEIL (France). In this experimental set-up, it is possible to combine micro-X-ray scattering, micro-X-ray absorption and micro-X-ray fluorescence spectroscopies. SOLEIL is running at 2.75 GeV with an average current of 400 mA in the new top-up mode. Details regarding the monochromator, the mirror as well as the devices used for the detection on DIFFABS, are gathered in previous studies (Bazin *et al.*, 2008; Nguyen *et al.*, 2011). In our case, the beamline was optimized in order to measure XANES spectra at the Sr *K*-edge. The energy range for the spectra acquisition was selected between 16000 and 16200 eV, with an energy step of 0.5 eV and a 3 s acquisition time. The measurements were made in the fluorescence mode, using a silicon drift detector. The size of the beam was determined by a set of slits (100–500 μm).

Regarding the analysis of the EXAFS modulations, data treatment was carried out using standard procedures (Sayers *et al.*, 1970, 1971). The background contribution, originating from the pre-edge region, was extrapolated using the Victoreen relationship and then subtracted from the experimental data. The smooth atom-like contribution $\mu_0(k)$ was obtained by cubic-spline fitting. The edge energy, E_0 , was arbitrarily chosen at the edge inflection. The EXAFS function, multiplied by k , was a Fourier transform (FT) in the photoelectron wavevector range (2.33–9.00 \AA^{-1}) after the application of a Kaiser window. Such analysis parameters have been selected in order to compare our results with a previous investigation (Terra *et al.*, 2009). In this work, all FTs are calculated and presented without phase correction.

3. Results

All the XANES spectra collected at the Sr *K*-edge on the different samples have been plotted in Fig. 2. The spectra are classified depending on the type of calcifications (physiological/pathological). For some of these experimental data, a small feature (see arrows in Fig. 2) exists (at 16135 eV) just after the white line which reflects the effective charge of Sr^{2+} ions ($4d^0$ electron configuration). We can already underline the fact that such a feature seems to be present in physiological as well as in pathological calcifications.

A set of numerical simulations have been performed regarding the local environment of $\text{Sr}(\text{I})^{2+}$. For the sake of clarity, the labels (see Fig. 3) which indicate the Sr environment are similar to the ones previously used (Bazin *et al.*,

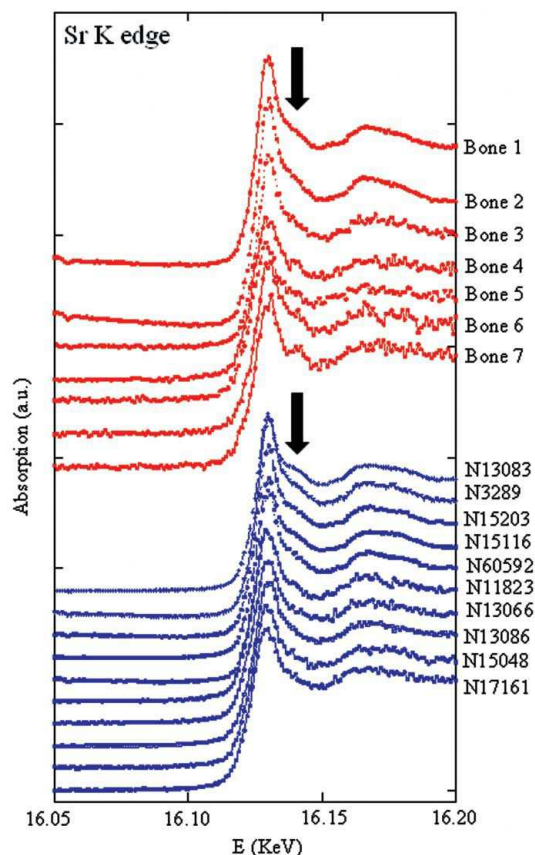


Figure 2
XANES part of X-ray absorption spectra collected at the Sr *K*-edge for different physiological (in red) and pathological (in blue) calcifications.

2011). For this new set of numerical calculations, different structural configurations have been taken into account including cations vacancies (Fig. 4), Ca/Sr substitutions (Fig. 5) and the local lattice distortion (Fig. 6).

Regarding the analysis of the EXAFS modulations beyond the Sr *K*-edge, a typical example of the X-ray absorption spectrum is shown in Fig. 7. Selected EXAFS oscillations have been plotted and have been compared with *FEFF* simulations and other experimental results in Fig. 8.

4. Discussion

Strontium-substituted apatites are used in the treatment of postmenopausal osteoporosis and have different applications owing to their antimicrobial activity (Ravi *et al.*, 2012), blue-luminescent composition (Zhang *et al.*, 2009) and radiopacity (Romieu *et al.*, 2006). This set of potential industrial applications has motivated different investigations regarding the interaction of Sr^{2+} cations and different compounds such as calcium hydrogenophosphate dehydrate, $\text{Na}_2\text{O}-\text{CaO}-\text{P}_2\text{O}_5-\text{SiO}_2$ glasses (O'Donnel & Hill, 2010) or Ca phosphate (Korbas *et al.*, 2004).

Different characterization techniques exist to assess the localization of bivalent cations in the inorganic matrix. Among them are NMR (Bonhomme *et al.*, 2012; Pizzala *et al.*, 2009), FTIR spectroscopy (Bigi *et al.*, 2007), microRaman (Gomes *et*

x	y	z	Type	Distance
0.00000	0.00000	0.00000	Sr ²⁺ (I)	0.00000
-1.44114	0.88703	-1.73195	O1	2.42144
1.48885	0.80450	-1.73195	O2	2.42148
-0.04762	-1.69168	-1.73195	O3	2.42151
-1.73357	0.24000	1.70855	O4	2.44581
1.07471	1.38127	1.70855	O5	2.44582
0.65894	-1.62141	1.70855	O6	2.44588
-0.61777	2.70175	0.45965	O7	2.80934
-2.03090	-1.88598	0.45965	O8	2.80940
2.64875	-0.81592	0.45965	O9	2.80943
0.28156	2.69750	1.70855	P1	3.20545
-2.47689	-1.10501	1.70855	P2	3.20549
2.19541	-1.59264	1.70855	P3	3.20554
0.00000	0.00000	3.41710	Ca1	3.41710
0.00000	0.00000	-3.46390	Ca2	3.46390
-2.96862	1.10491	-1.73195	P4	3.61015
2.44128	2.01840	-1.73195	P5	3.61019
0.52743	-3.12345	-1.73195	P6	3.61023
-3.41462	1.88588	-0.48305	O10	3.93059
3.34061	2.01416	-0.48305	O11	3.93063
0.07409	-3.90018	-0.48305	O12	3.93068
-0.70762	3.48932	-1.73195	Ca3	3.95926
-2.66803	-2.35758	-1.73195	Ca4	3.95932
3.37573	-1.13189	-1.73195	Ca5	3.95934
-2.77748	2.35748	1.70855	Ca6	4.02384
3.43046	1.22658	1.70855	Ca7	4.02389
-0.65290	-3.58421	1.70855	Ca8	4.02393

Figure 3
Details regarding the spatial repartition of atoms around Sr²⁺ cations located in site (I).

obtained (Mimura *et al.*, 2012). At this point we have to underline that more classical chemical techniques such as solid titration also give quite interesting results. For example, the increase in solubility with strontium content has been interpreted as a destabilization of the crystal structure (Pan *et al.*, 2009a).

Several publications show that the incorporation of foreign ions into the HAP structure can affect its crystallinity, morphology, lattice parameters and stability (Ibrahim *et al.*, 2011). All these structural parameters may have significant medical implications. For example, we have already discussed the modulations of crystal morphology with the nature of the disease (Daudon *et al.*, 2008, 2009). Among the bivalent cations, Sr²⁺, Cd²⁺ and Pb²⁺ can replace Ca²⁺ over the whole range of compositions (Bigi *et al.*, 1995). Several other ions, usually with an ionic radius smaller than that of Ca²⁺, such as Mg²⁺, Mn²⁺, Fe²⁺, Co²⁺ and Ni²⁺, are known to inhibit the synthesis of HAP and promote the formation of β -tricalcium phosphate. Regarding the interaction of

Sr with apatite, no significant incorporation of strontium in the resulting apatite was detected for [Sr] < 0.1 mM, but crystal morphology changed dramatically and crystallinity increased for [Sr] \geq 0.3 mM due to the formation of strontium-substituted apatite, with substitution increasing with an increase of solution (Pan *et al.*, 2009b). Also, from the kinetics point of view, it has been reported that the presence of strontium retarded the crystal growth of HAP on seed crystals (Rokidi &

al., 2012) small-angle X-ray scattering, X-ray diffraction (Li *et al.*, 2010), neutron powder diffraction (Gomes *et al.*, 2011) as well as techniques specific to synchrotron radiation such as anomalous wide-angle X-ray scattering (Revel *et al.*, 2000) and X-ray absorption spectroscopy. Recently, an overview of the most important microanalytical techniques currently available for the *in situ* detection of trace metals in cells and tissues has been published (McRae *et al.*, 2009). An X-ray beam focused in one direction to a width of 7 nm at 20 keV has been

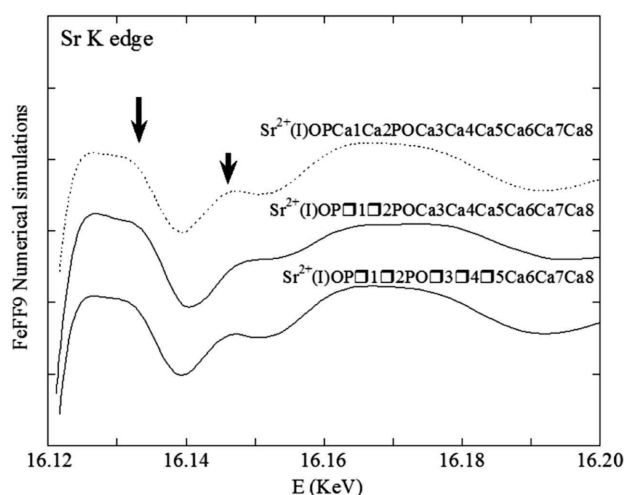


Figure 4
FEFF simulations taking into account the presence of cationic vacancies (squares) for the environment of Sr²⁺(I). The positions of vacancies in the different simulations have been given.

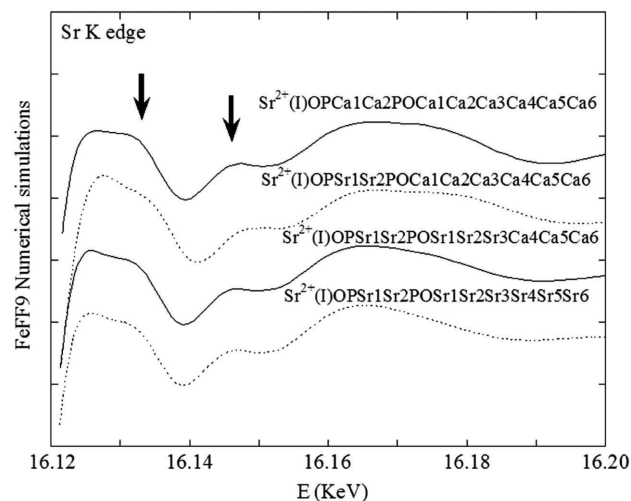


Figure 5
FEFF simulations, taking into account Ca/Sr substitution in the environment of Sr²⁺(I). The different substitutions taken into account in the simulations have been given.

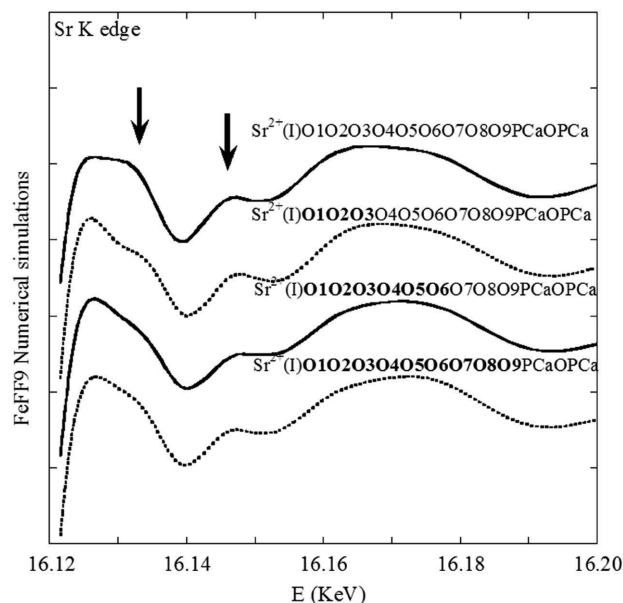


Figure 6
FEFF simulations taking into account possible distortion in the environment of $\text{Sr}^{2+}(\text{I})$. More precisely, cartesian coordinates of oxygen atoms in bold have been changed.

Koutsoukos, 2012). Previous studies clearly indicate that the insertion of cations in apatite is a complex process which is sensitive to preparation methods (Bazin *et al.*, 2011).

In this investigation, we focus on the localization of Sr^{2+} cations in Ca apatite. It should be noted that such measurements can be performed directly on the sample with minimal preparation. In order to confirm the possible relationship between the adsorption mode of Sr^{2+} cations (adsorbed at the surface or inserted inside the Ca apatite crystal) and the nature of the calcification (physiological or pathological calcifications), X-ray absorption experiments at the Sr *K*-edge as well as *ab initio* numerical simulations have been performed (Bazin *et al.*, 2011). At first, the new set of XANES data collected for 17 samples (Fig. 2) seems to indicate that such a relationship is not obvious. The XANES data corresponding to pathological and physiological calcifications are quite similar. A small feature related to the adsorption mode of Sr^{2+} cations exists (at 16135 eV) just after the white line and before the first oscillation is observed for one sample (bone 1) while for three physiological samples (bones 2–4) the evidence is much weaker (Fig. 2).

Regarding the numerical simulations of the XANES part of the absorption spectra, in this new set of *ab initio* calculations several structural configurations have been taken into account, including a possible local lattice distur-

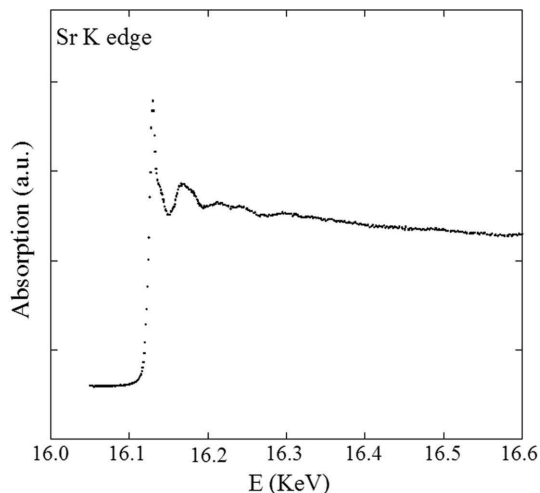


Figure 7
 X-ray absorption spectra collected beyond the Sr *K*-edge.

tion around the Sr^{2+} cations (Fig. 6), the existence of Ca/Sr substitutions (Fig. 5) or cations vacancies (Fig. 4). As observed, the existence of the small feature present just after the white line seems to be not significantly affected by such structural modifications. We can underline the fact that the local environment of $\text{Sr}(\text{I})^{2+}$ is already quite distorted and thus structural modifications such as modification of the Cartesian coordinates of oxygen atoms around Sr^{2+} cations, Ca/Sr substitution or cation vacancies seem to have a weak influence on the XANES part of the absorption spectra. Now we have to gather this new set of simulations with the previous one. The feature apparent after the white line is visible only for Sr^{2+} cations present inside the apatite network and disappears for

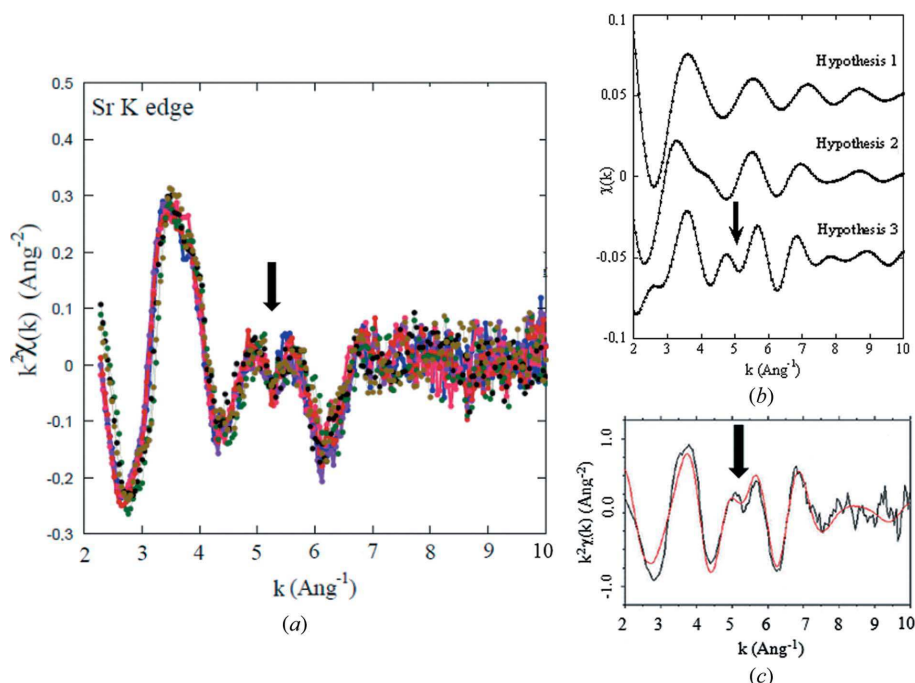


Figure 8
 (a) EXAFS modulations after the Sr *K*-edge for different samples. (b) EXAFS modulations as calculated for the three different hypotheses. (c) EXAFS modulations as measured in the case of hypothesis 3 (Terra *et al.*, 2009).

Sr^{2+} cations surrounded by oxygen and phosphorus atoms. As underlined previously (Bazin *et al.*, 2011), such a feature is not always observed for Sr^{2+} cations present inside the apatite network.

Regarding the EXAFS data measured after the Sr *K*-edge, the modulations are very similar to ones previously measured (Terra *et al.*, 2009; Korbas *et al.*, 2004) in which Sr^{2+} cations are located inside the apatite crystal (Fig. 1). In order to show what differences in Sr signal would be expected for Sr in different local environments, we have calculated the EXAFS modulations for the different structural possibilities (Fig. 8*b*). It is quite clear that our experimental measurements correspond to a substitution between Sr^{2+} cations and Ca^{2+} cations inside Ca phosphate nanocrystals (see arrow in Figs. 8*a*, 8*b* and 8*c*). Such data, in line with previous experiments (Fig. 8*c*), indicate clearly the existence of Sr–Ca or Sr–Sr interatomic distances after the contribution coming from oxygen and phosphorus atoms for the different samples selected in this investigation. There is no significant structural difference between physiological and pathological calcifications regarding the localization of Sr^{2+} cations.

Such structural considerations may have medical implications. In osteoporosis, clinical studies indicate that strontium ranelate increases bone formation and tends to decrease bone resorption in treated osteoporotic patients (Marie, 2007). Thus, different investigations have been performed to assess the effect of strontium incorporation into hydroxyapatite nanocrystals on bone cell response. For example, different experiments (Capuccini *et al.*, 2009) indicate that strontium stimulates osteoblast activity and exerts its inhibitory effect on osteoclast proliferation even when incorporated into hydroxyapatite. Similar results have been obtained (Doublier *et al.*, 2011) which showed that the quality of bone mineralization (density and heterogeneity at tissue level) was preserved after a long-term treatment with strontium ranelate. By contrast, a similar *in vitro* study underlines a potential physicochemical interference by Sr with HAP formation and crystal properties (Verberckmoes *et al.*, 2004).

In our investigation, we consider at least two very different preparation procedures related to physiological and pathological calcifications. In physiological calcifications, cells are at the core of their formation while in pathological calcifications, such as kidney stones, a simple supersaturation process explains their pathogenesis. For physiological calcifications, their formations are made in a confined space while the building process of concretions occurred under a fluid circulation. Finally, in both cases, the content of Sr is quite low. For these two kinds of calcifications, the contribution of Sr atoms in X-ray fluorescence spectra are quite small (around 500 p.p.m.). Taking into account these similarities and differences, it is quite interesting to stress that the localization of Sr^{2+} cations in biological apatites is basically the same. By contrast, the literature seems to show that for synthetic apatites, the localization of Sr^{2+} cations depends of the preparation procedure. Based on this study, we may suppose that, for supplementation of Sr at low concentration, Sr^{2+} cations are localized into the apatite network.

5. Conclusion

Even if strontium ranelate has also shown anti-fracture efficacy in the treatment of a major health public problem, namely osteoporosis, little is known regarding the environment of Sr in biological apatites. Moreover, it is quite difficult to obtain such information based on the literature regarding the relationship between Sr^{2+} cations and synthetic apatites, such a relationship being strongly dependent on the preparation procedure.

In this work, biological apatites have been investigated through XANES–EXAFS spectroscopy and a set of numerical simulations using the *FEFF9* program, taking into account local lattice distortion, $\text{Ca}^{2+}/\text{Sr}^{2+}$ substitution or the presence of cationic vacancies, have been performed. The complete set of experimental data collected on 17 samples seems to indicate that there is no relationship between the nature of the calcification (physiological and pathological) and the adsorption mode of Sr^{2+} cations (simple adsorption or insertion). Also, the existence of Sr–Ca or Sr–Sr interatomic distances seems to show that Sr^{2+} cations are inserted into the apatite network, a result in line with previous investigations performed on synthetic and biological apatites (Seward *et al.*, 1999; Martin *et al.*, 2012; Rey *et al.*, 2007; Bazin *et al.*, 2012).

This work was supported by the Physics and Chemistry Institutes of the CNRS, the Fondation pour la Recherche Médicale (grant DV020081013483 ‘Vieillesse ostéoarticulaire’ 2008–2011, Pr Lioté), the Association Rhumatisme et Travail (ART), the Association ‘Cristaux et Cartilage’, PTD, the Association pour la Recherche en Pathologie Synoviale (ARPS) and by the Centre National de la Recherche Scientifique (CalArthros project ‘Longévité et Vieillesse 2010’ CNRS interdisciplinary program) and the Agence Nationale de la Recherche (France) (contract ANR-09-BLAN-0120-02). The authors declare that the study sponsors have no role in the study design, collection, analysis and interpretation of data, in the writing of the manuscript and in the decision to submit the manuscript for publication. The authors wish to thank the other members of the DIFFABS beamline team, Filipe Alves and Cristian Mocuta, for their help during synchrotron experiments.

References

- Bazin, D., Carpentier, X., Brocheriou, I., Dorfmueller, P., Aubert, S., Chappard, C., Thiaudière, D., Reguer, S., Waychunas, G., Jungers, P. & Daudon, M. (2009*b*). *Biochimie*, **91**, 1294–1300.
- Bazin, D., Carpentier, X., Traxer, O., Thiaudière, D., Somogyi, A., Reguer, S., Waychunas, G., Jungers, P. & Daudon, M. (2008). *J. Synchrotron Rad.* **15**, 506–509.
- Bazin, D., Chappard, Ch., Combes, Ch., Carpentier, X., Rouzière, S., André, G., Matzen, G., Allix, M., Thiaudière, D., Reguer, S., Jungers, P. & Daudon, M. (2009*a*). *Osteoporos. Int.* **20**, 1065–1075.
- Bazin, D. & Daudon, M. (2012). *J. Phys. D.* **45**, 383001.
- Bazin, D., Daudon, M., Chappard, C., Rehr, J. J., Thiaudière, D. & Reguer, S. (2011). *J. Synchrotron Rad.* **18**, 912–918.
- Bazin, D., Daudon, M., Chevallier, P., Rouzière, S., Elkaim, E., Thiaudière, D., Fayard, B., Foy, E., Albouy, P. A., André, G., Matzen, G. & Véron, E. (2006). *Ann. Biol. Clin.* **64**, 125–139.

- Bazin, D., Daudon, M., Combes, Ch. & Rey, Ch. (2012). *Chem. Rev.* **112**, 5092–5120.
- Bazin, D. & Rehr, J. (2003). *Catal. Lett.* **87**, 85–90.
- Bigi, A., Boanini, E., Capuccini, C. & Gazzano, M. (2007). *Inorg. Chim. Acta*, **360**, 1009–1016.
- Bigi, A., Foresti, E., Gandolfi, M., Gazzano, M. & Roveri, N. (1995). *J. Inorg. Biochem.* **58**, 49–58.
- Bonhomme, C., Gervais, C., Babonneau, F., Coelho, C., Pourpoint, F., Azaïs, T., Ashbrook, S. E., Griffin, J. M., Yates, J. R., Mauri, F. & Pickard, C. J. (2012). *Chem. Rev.* **112**, 5733–5779.
- Capuccini, C., Torricelli, P., Boanini, E., Gazzano, M., Giardino, R. & Bigi, A. (2009). *J. Biomed. Mater. Res. A*, **89**, 594–600.
- Carpentier, X., Bazin, D., Jungers, P., Reguer, S., Thiaudière, D. & Daudon, M. (2010). *J. Synchrotron Rad.* **17**, 374–379.
- Cortet, B. (2011). *Curr. Osteoporos. Rep.* **9**, 25–30.
- Daudon, M., Bazin, D., André, G., Jungers, P., Cousson, A., Chevallier, P., Véron, E. & Matzen, G. (2009). *J. Appl. Cryst.* **42**, 109–115.
- Daudon, M., Jungers, P. & Bazin, D. (2008). *N. Engl. J. Med.* **359**, 100–102.
- Dessombz, A., Bazin, D., Dumas, P., Sandt, C., Sule-Suso, J. & Daudon, M. (2011). *PLoS ONE*, **6**, e28007.
- Dessombz, A., Méria, P., Bazin, D. & Daudon, M. (2012). *PLoS ONE*, **7**, e51691.
- Dessombz, A., Nguyen, Ch., Ea, H. K., Rouzière, S., Foy, E., Hannouche, D., Réguer, S. Picca, F. E., Thiaudière, D., Lioté, F., Daudon, M. & Bazin, D. (2013). *J. Trace Elem. Med. Biol.* In the press.
- Doublier, A., Farlay, D., Khebbab, M. T., Jaurand, X., Meunier, P. J. & Boivin, G. (2011). *Eur. J. Endocrinol.* **165**, 469–476.
- Eichert, D., Salomé, M., Banu, M., Susini, J. & Rey, C. (2005). *Spectrochim. Acta B*, **60**, 850–858.
- Elliott, J. C. (1994). *Structure and Chemistry of the Apatites and Other Calcium Orthophosphates*. Amsterdam: Elsevier.
- Estepa, L. & Daudon, M. (1997). *Biospectroscopy*, **3**, 347–369.
- Gomes, S., Nedelec, J. M., Jallot, E., Sheptyakov, D. & Renaudin, G. (2011). *Chem. Mater.* **23**, 3072–3085.
- Gomes, S., Nedelec, J. & Renaudin, G. (2012). *Acta Biomater.* **8**, 1180–1189.
- Guinier, A. (1956). *X-ray Diffraction in Crystals Imperfect Crystals and Amorphous Bodies*. Paris: Dunod.
- Ibrahim, D. M., Mostafa, A. A. & Korowash, S. I. (2011). *Chem. Cent. J.* **5**, 74.
- Kanakamani, J. & Tandon, N. (2008). *Ind. J. Rheumatol.* **3**, 148–157.
- Korbas, M., Rokita, E., Meyer-Klaucke, W. & Ryzek, J. (2004). *J. Biol. Inorg. Chem.* **9**, 67–76.
- Laurencin, D., Almora-Barrios, N., de Leeuw, N. H. & Gervais, Ch. (2011). *Biomaterials*, **3**, 1826–1837.
- Li, C., Paris, O., Siegel, S., Roschger, P., Paschalis, E. P., Klaushofer, K. & Fratzl, P. (2010). *J. Bone Miner. Res.* **25**, 968–975.
- McKeown, D. A., Kot, W. K. & Pegg, I. L. (2003). *J. Non-Cryst. Solids*, **317**, 290–300.
- McRae, R., Bagchi, P., Sumalekshmy, S. & Fahrni, C. J. (2009). *Chem. Rev.* **109**, 4780–4827.
- Marie, P. J. (2005). *Curr. Opin. Pharmacol.* **5**, 633–636.
- Marie, P. J. (2007). *Bone*, **40**, S5–S8.
- Martin, R. A., Twyman, H. L., Rees, G. J., Barney, E. R., Moss, R. M., Smith, J. M., Hill, R. G., Cibin, G., Charpentier, T., Smith, M. E., Hanna, J. V. & Newport, R. J. (2012). *J. Mater. Chem.* **22**, 22212.
- Meunier, P. J., Roux, C., Seeman, E., Ortolani, S., Badurski, J. E., Spector, T. D., Cannata, J., Balogh, A., Lemmel, E., Pors-Nielsen, S., Rizzoli, R., Genant, H. K. & Reginster, J. (2004). *N. Engl. J. Med.* **350**, 459–468.
- Mimura, H., Handa, S., Kimura, T., Yumoto, H., Yamakawa, D. & Yokoyama, H. (2012). *Nat. Phys.* **6**, 122–125.
- Moonen, J., Slot, J., Lefferts, L., Bazin, D. & Dexpert, H. (1995). *Physica B*, **208–209**, 689–690.
- Moyad, M. A. (2003). *Urol. Oncol. Semin. Orig. Investig.* **21**, 375–379.
- Murata, H., Shitara, K., Tanaka, I., Nakahira, A., Mizoguchi, T. & Matsunaga, K. (2012). *J. Phys. Condens. Matter*, **22**, 384213.
- Nguyen, C., Ea, H. K., Thiaudiere, D., Reguer, S., Hannouche, D., Daudon, M., Lioté, F. & Bazin, D. (2011). *J. Synchrotron Rad.* **18**, 475–480.
- O'Day, P. A., Newville, M., Neuhoff, P. S., Sahai, N. & Carroll, S. A. (2000). *J. Colloid Interface Sci.* **222**, 184–197.
- O'Donnell, M. D. & Hill, R. G. (2010). *Acta Biomater.* **6**, 2382–2385.
- Palmer, B. J., Pfund, D. M. & Fulton, J. L. (1996). *J. Phys. Chem.* **100**, 13393–13398.
- Pan, H. B., Li, Z. Y., Lam, W. M., Wong, J. C., Darvell, B. W., Luk, K. D. & Lu, W. W. (2009a). *Acta Biomater.* **5**, 1678–1685.
- Pan, H., Li, Z., Wang, T., Lam, W. M., Wong, C. T., Darvell, B. W., Luk, K. D. K., Hu, Y. & Lu, W. W. (2009b). *Cryst. Growth Des.* **9**, 3342–3345.
- Penel, G., Leroy, G., Rey, C. & Bres, E. (1998). *Calcif. Tissue Int.* **63**, 475–481.
- Persson, I., Sandström, M., Yokoyama, H. & Chaudhry, M. (1995). *Z. Naturforsch. A*, **50**, 21–30.
- Pinkerton, J. V. & Dalkin, A. C. (2007). *Am. J. Obstet. Gynecol.* **197**, 559–565.
- Pizzala, H., Caldarelli, S., Eon, J. G., Rossi, A. M., San Gil, R. A., Laurencin, D. & Smith, M. E. (2009). *J. Am. Chem. Soc.* **131**, 5145–5152.
- Ravi, N. D., Balu, R. & Sampath Kumar, T. S. (2012). *J. Am. Ceram. Soc.* **95**, 2700–2708.
- Rehr, J. J., Kas, J. J., Prange, M. P., Sorini, A. P., Takimoto, Y. & Vila, F. (2009). *C. R. Phys.* **10**, 548–559.
- Revel, R., Bazin, D., Elkaim, E., Kihn, Y. & Dexpert, H. (2000). *J. Phys. Chem. B*, **104**, 9828–9835.
- Rey, C., Combes, C., Drouet, C., Sfihi, H. & Barroug, A. (2007). *Mater. Sci. Eng.* **27**, 198–205.
- Rey, C., Miquel, J. L., Facchini, L., Legrand, A. P. & Glimcher, M. J. (1995). *Bone*, **16**, 583–586.
- Rocheffort, G. Y., Pallu, S. & Benhamou, C. L. (2010). *Osteoporos. Int.* **21**, 1457–1469.
- Rokidi, S. & Koutsoukos, P. G. (2012). *Chem. Eng. Sci.* **77**, 157–164.
- Romieu, G., Garric, X., Munier, S., Vert, M. & Boudeville, Ph. (2006). *Acta Biomater.* **6**, 3208–3215.
- Sayers, D. A., Lytle, F. W. & Stern, E. A. (1970). *Advances in X-ray Analysis*. New York: Plenum.
- Sayers, D. A., Stern, E. A. & Lytle, F. W. (1971). *Phys. Rev. Lett.* **27**, 1204–1207.
- Sepulcre, F., Proietti, M. G., Benfatto, M., Della Longa, S., García, J. & Padrós, E. (2004). *Biophys. J.* **87**, 513–520.
- Seward, T., Henderson, C., Charnock, J. & Driesner, T. (1999). *Geochim. Cosmochim. Acta*, **63**, 2409–2418.
- Tang, Y., Chappell, H. F., Dove, M. T., Reeder, R. J. & Lee, Y. J. (2009). *Biomaterials*, **30**, 2864–2872.
- Terra, J., Dourado, E. R., Eon, J. G., Ellis, D. E., Gonzalez, G. & Rossi, A. M. (2009). *Phys. Chem. Chem. Phys.* **11**, 568–577.
- Vallet-Regi, M. & Gonzalez-Calbet, J. M. (2002). *Prog. Solid State Chem.* **32**, 1–31.
- Verberckmoes, S. C., Behets, G. J., Oste, L., Bervoets, A. R., Lamberts, L. V., Drakopoulos, M., Somogyi, A., Cool, P., Dorriné, W., De Broe, M. E. & D'Haese, P. C. (2004). *Calcif. Tissue Int.* **75**, 405–415.
- Yachandra, V. K. & Yano, J. (2011). *J. Photochem. Photobiol. B*, **104**, 51–59.
- Yu, M., Tian, S., Chu, W., Chen, D., Wang, Q. & Wu, Z. (2009). *J. Synchrotron Rad.* **16**, 528–532.
- Zhang, C., Cheng, Z., Yang, P., Xu, Z., Peng, C., Li, G. & Lin, J. (2009). *Langmuir*, **25**, 13591–13598.