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Synthesis of pyrrolidine-based analogues of 2-acetamidoglycosides as *N*-acetylglucosaminidase inhibitors

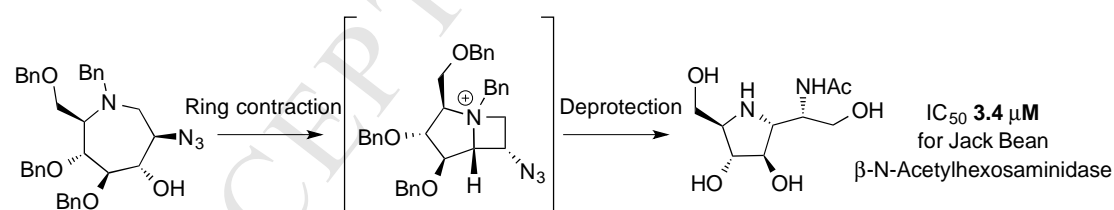
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Abstract. A ring-contraction strategy applied to β -azido, γ -hydroxyazepanes yielded after functional group manipulation new tetrahydroxylated pyrrolidines displaying an acetamido moiety, one of these iminosugars demonstrating low micromolar inhibition on *N*-acetylglucosaminidases.



Keywords. Iminosugars, Ring-contraction, Glycosidase inhibitors, Pyrrolidines

Highlights.

- A new family of amino-functionalised pyrrolidines was synthesized.
- A ring-contraction of β -azido, γ -hydroxyazepanes was used.
- A low micromolar inhibitor of β -N-acetylhexosaminidase identified.

Polyhydroxylated pyrrolidines^{1,2} are well-established powerful glycosidase inhibitors, even though their analogy with hexopyranoses, and therefore the structural basis of their inhibition, are less straightforward than for the corresponding piperidines.³ Hexosaminidases are a very important class of glycosidases that cleave the pyranosidic *N*-acetyl-D-glucosamine unit from glycoconjugates. Several pyrrolidines bearing an acetamide group have been reported as potent hexosaminidases inhibitors. Interestingly, only one naturally occurring acetamido-containing pyrrolidine was isolated so far: Pochonicine.⁴⁻⁶ The main classes of synthetic nitrogen functionalized polyhydroxylated pyrrolidines are represented in Figure 1, the most studied one being the 2,5-dideoxy-2,5-imino-hexitols **A**⁷⁻²¹ but other scaffolds such as **B**,²²⁻²⁴ **C**,²⁵⁻²⁶ **D**,²⁷ **E**²⁸ and **F**⁸ have also been prepared. It is rather striking that structure **G**, which can be seen as a combination of **A** and **E** possessing as many hydroxyl groups as the hexosaminidase substrate and product, has never been synthesized and assessed as a hexosaminidase inhibitor. The present study reports the synthesis and hexosaminidase inhibitory evaluation of molecules derived from scaffold **G**. (Figure 1)

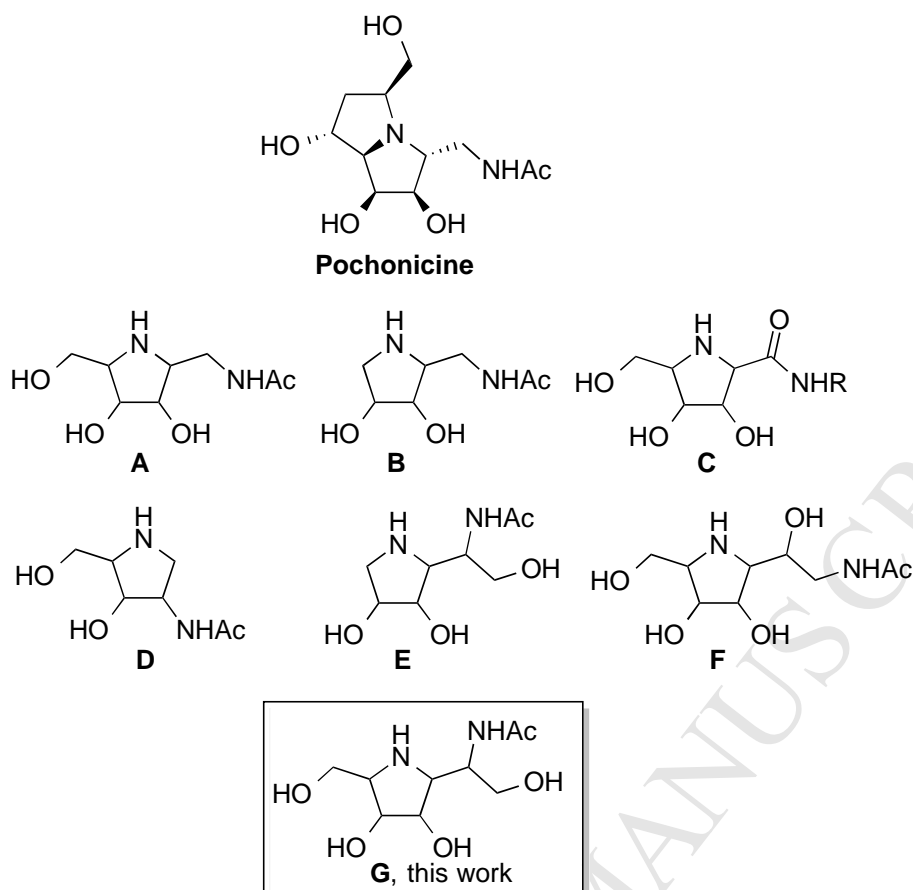
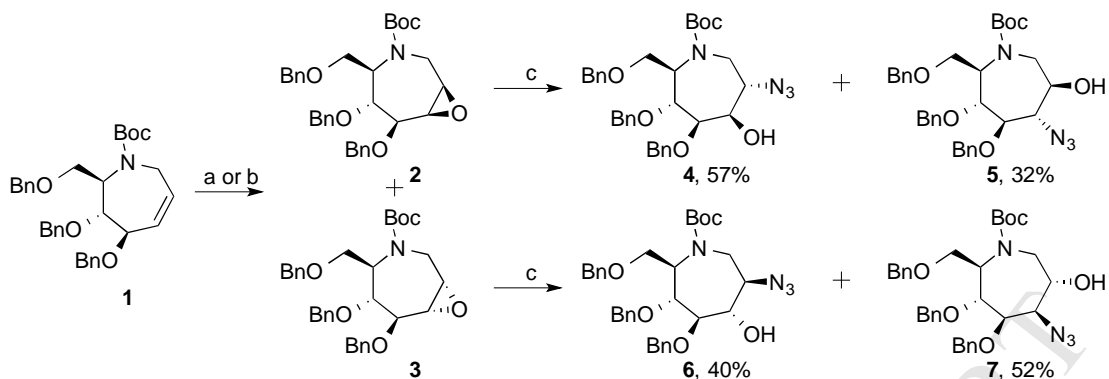


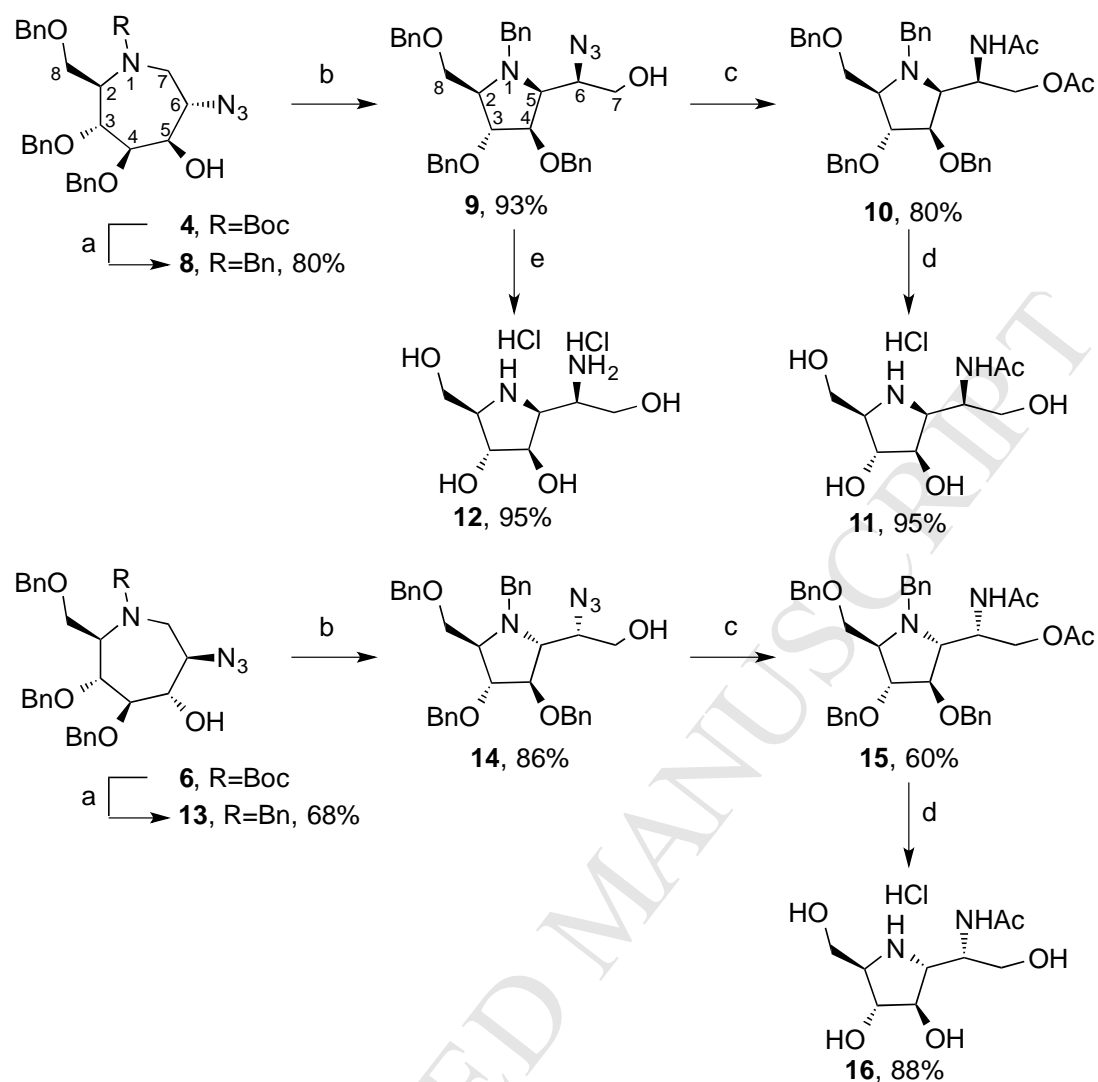
Figure 1: Structures of the existing classes of acetamido-pyrrolidines **A-F** and of the target scaffold **G**.

In the course of our studies aimed at the synthesis of GlcNAc-like piperidine homoiminosugars exploiting a ring-contraction methodology,²⁹⁻³² a 2,3-trans-2-hydroxy,3-azido-azepane was required and obtained from the unsaturated 7-membered ring **1**.³³ The obvious synthetic route transits via the formation of an epoxide, followed by its azidolysis. We observed that it was possible to operate the epoxydation with some degree of stereocontrol to afford either epoxides **2** or **3** as the main products.³³ These latter could then be opened using sodium azide to give, in both cases, a significant amount of the 2-azido derivatives **4** and **6** together with the desired 3-azido compounds **5** and **7**.³³ (Scheme 1)



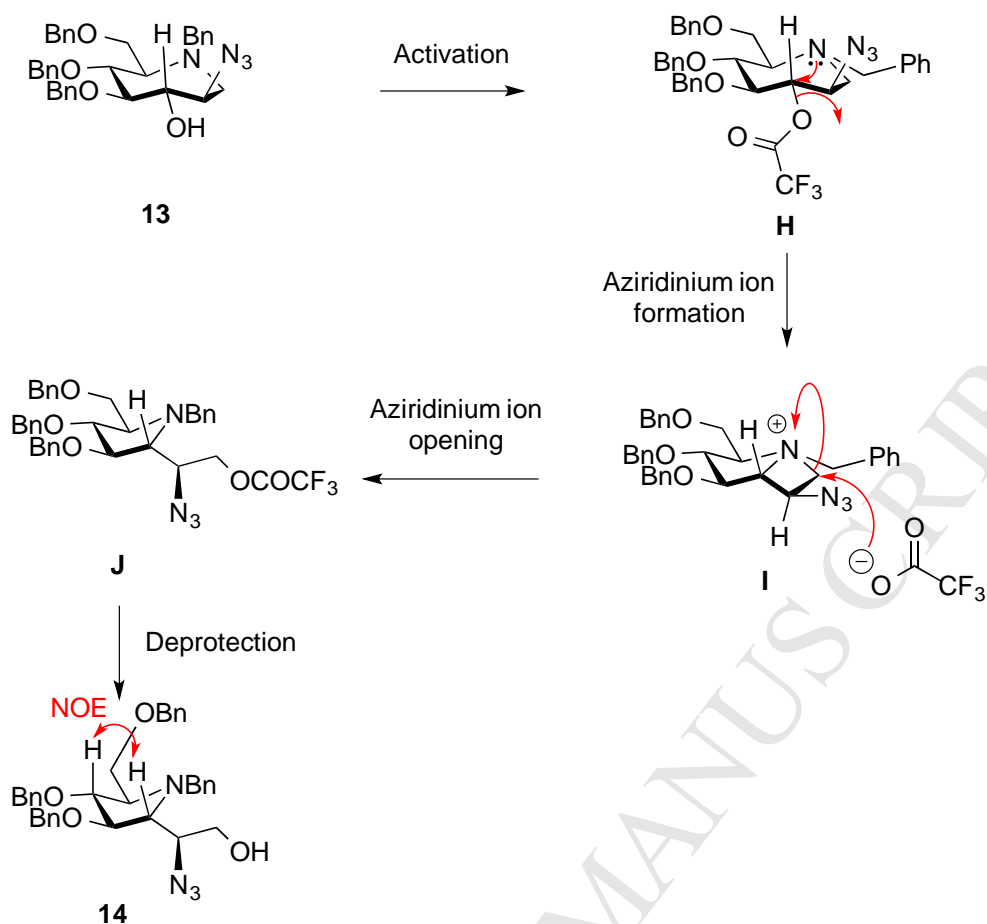
Scheme 1 : Synthesis of azidoazepanes **4-7**. Reagents and conditions: a) Oxone, D-epoxone, NaHCO₃, **2** : 54%, b) Oxone, CF₃COCH₃, NaHCO₃, **2** : 29%, **3** : 51%; c) NaN₃, NH₄Cl, DMF/H₂O, 90°C

Compounds **4** and **6**³³ are also suitable candidates for a ring contraction reaction³⁴ to give pyrrolidine derivatives³⁵ through a γ -aminoalcohol rearrangement. Hence, we decided to apply the TFAA-mediated ring contraction conditions developed by Cossy³⁶ to β -azidoazepanes **4** and **6** that were first converted into the *N*-benzyl derivatives **8** (80%) and **13** (68%) respectively, using TFA followed by *N*-benzylation (BnBr, K₂CO₃). Their ring contraction with TFAA furnished the azidopyrrolidines **9** (93%) and **14** (86%) respectively in good yield. Reduction of the azide moiety (PPh₃, THF/H₂O) followed by *N*-acetylation was achieved to provide the acetamide **10** (80%) and **15** (60%) respectively. Final *O*-deacetylation followed by hydrogenolysis yielded the target pyrrolidines **11** (95%) and **16** (88%). Compound **9** was also directly submitted to the action of hydrogen in the presence of Pd/C to give the diamine **12** in 95% yield as its hydrochloride salt (Scheme 2).



Scheme 2 : Synthesis of NHAc derived pyrrolidines **11** and **16**. Reagents and conditions: a) i) TFA, DCM; ii) BnBr, K₂CO₃, EtOAc/H₂O; b) i) Trifluoroacetic anhydride (TFAA), Et₃N, toluene, reflux, ii) 10% aq. NaOH; c) i) PPh₃, THF/H₂O, 80°C, ii) Pyridine, Ac₂O; d) i) Et₃N, MeOH, H₂O, ii) H₂, Pd/C, MeOH, HCl; e) H₂, Pd/C, MeOH, HCl

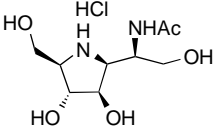
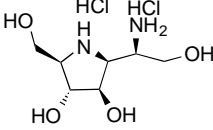
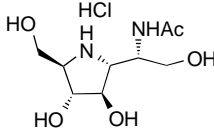
The ring contraction reaction is initiated by the esterification of the free hydroxyl group in azepane **13** to give intermediate **H**, in which the amine displaces this leaving group to produce the fused pyrrolidine-azetidinium ion **I**. Nucleophilic ring opening at the less hindered carbon affords pyrrolidine **J** which leads to the five-membered iminosugar **14** upon saponification. The stereochemistry of the ring-contracted product is the one expected by this mechanism as attested by the NOE cross-correlation between H-3 and H-5 on **14**. (Scheme 3)



Scheme 3 : Proposed mechanism for the ring contraction step.

The three pyrrolidines **11**, **12** and **16** were assayed as inhibitors of a panel of hexosaminidases and β -glucuronidases. Iminosugar **11** is a moderate inhibitor of β -*N*-acetylglucosaminidases with IC_{50} in the high micromolar range. The present work revealed that inversion of C-1 side chain in **11** to give **16** significantly enhanced its inhibition potency against these enzymes, pyrrolidine **16** demonstrating potent Jack bean β -*N*-acetylglucosaminidase inhibition, with a IC_{50} value of 3.4 μ M. In contrast, replacement of the acetamide group by an amine as in **12** is detrimental to hexosaminidase inhibition (Table 1). It is noteworthy that pyrrolidine **11** showed low micromolar inhibition against bovine liver and *E.coli* β -glucuronidase, with IC_{50} values of 26 and 15 μ M, respectively. Previous study suggested that β -glucuronidase recognized uronic acid and carboxylic acid part is required for tight binding.^{37,38} Thus, pyrrolidine **11** is an interesting case for β -glucuronidase inhibition.

Table 1 . Concentration of iminosugars giving 50 % inhibition of various glycosidases

IC ₅₀ (μM)			
			
	11	12	16
Enzyme :			
β-N-Acetylglucosaminidase			
<i>Aspergillus oryzae</i>	241	NI (0%)	323
Bovine kidney	181	NI (0%)	31
HL60	538	NI (6.6%)	18
Human placenta	597	NI (0%)	15
Jack bean	61	NI (7.3%)	3.4
α-N-Acetylgalactosaminidase			
Chicken liver	^a NI ^b (26.2%)	NI (3.3%)	NI (7.2%)
β-glucuronidase			
Bovine liver	26	NI (0%)	NI (48.5%)
<i>E.coli</i>	15	NI (17.8%)	145

^a NI : No inhibition (less than 50% inhibition at 1000 μM).

^b () : inhibition % at 1000 μM

In conclusion, a ring-contraction methodology applied to seven-membered iminosugars bearing an azido group in β position furnished a low micromolar hexosaminidase inhibitor after conversion of the azide function into an acetamide and final deprotection. This work complements previous work on the conversion of polyhydroxylated azepanes into six-membered NHAc-homoiminosugars.

1. Experimental

1.1 Material and methods

All commercial reagents were used as supplied. Solvents (DMF, THF) were distilled under anhydrous conditions. TLC plates (Macherey-Nagel, ALUGRAM[®] SIL G/UV₂₅₄, 0.2 mm silica gel 60 Å) were visualized under 254 nm UV light and/or by dipping the TLC plate into a solution of 3 g of phosphomolybdic acid in 100 mL of ethanol followed by heating with a heat gun. Flash column

chromatography was performed using Macherey-Nagel silica gel 60 (15-40 μm). NMR experiments were recorded with a Bruker AM- 400 spectrometer at 400 MHz for ^1H nuclei and at 100 MHz for ^{13}C nuclei. The chemical shifts are expressed in part per million (ppm) using residual CHCl_3 signal as internal reference ($\delta(^1\text{H}) = 7.26$ ppm and $\delta(^{13}\text{C}) = 77.16$ ppm) and the coupling constant J in hertz (Hz). NMR multiplicities are reported using the following abbreviations: b = broad, s = singlet, d = doublet, t = triplet, q = quadruplet, m = multiplet. HRMS were recorded on a Bruker microTOF spectrometer, using Tuning-Mix as reference. Optical rotations were measured on a Perkin-Elmer 341 digital polarimeter or a Jasco P-2000 polarimeter with a path length of 1 dm.

1.2 tert-butyl (2R,3R,4R,5R,6S)-6-azido-3,4-bis(benzyloxy)-2-((benzyloxy)methyl)-5-hydroxyazepane-1-carboxylate (4)

Known epoxide **2**³³ (465 mg, 0.853 mmol) was dissolved in a DMF/ H_2O mixture (9.0/1.0 mL), then NaN_3 (277 mg, 4.26 mmol) and NH_4Cl (226 mg, 4.26 mmol) were added. The resulting mixture was stirred at 90 $^\circ\text{C}$ for 3 days. After being cooled to room temperature, EtOAc and H_2O were added and the layers were separated. The aqueous layer was extracted twice with EtOAc and the combined organic layers were dried over MgSO_4 , filtered and evaporated. The residue was purified by flash chromatography (Cy/EtOAc: 9/1) to give **4** (285 mg, 57%) as colorless oil and **5**⁶ (160 mg, 32%) [α]_D +18.6 ($c = 1.0$, CHCl_3) ^1H NMR (400 MHz, CDCl_3 , 2 rotamers): 7.39-7.26 (m, 26H, H_{ar}), 7.23-7.22 (m, 4H, H_{ar}), 4.81 (d, 1H, $^2J = 11.5$ Hz, CH_2Ph), 4.75 (d, 1H, $^2J = 12.0$ Hz, CH_2Ph), 4.69-4.59 (m, 4H, CH_2Ph), 4.49-4.40 (m, 6H, CH_2Ph), 4.02-3.91 (m, 6H, $\text{H}_{8\text{a}}$, H_2 , H_3 , H_3' , H_5 , H_5), 3.87-3.53 (m, 12H, $\text{H}_{8\text{b}}$, $\text{H}_{8\text{a}'}$, $\text{H}_{8\text{b}'}$, H_2' , H_4 , H_4' , H_6 , H_6' , $\text{H}_{7\text{a}}$, $\text{H}_{7\text{b}}$, $\text{H}_{7\text{a}'}$, $\text{H}_{7\text{b}'}$), 2.57 (bs, 0.8H, OH), 2.52 (bs, 0.8H, OH'), 1.49 (s, 9H, CH_3 , *Boc*), 1.41 (s, 9H, CH_3 , *Boc*); ^{13}C NMR (100 MHz, CDCl_3 , 2 rotamers): δ 155.4, 155.1 (CO, *Boc*), 138.3, 138.1, 138.0, 138.0, 137.9, 137.8 (C_{ipso}) 128.4, 128.3, 128.2, 128.0, 127.9, 127.8, 127.7, 127.6, 127.5, 127.5 (CH_{ar}), 81.8, 81.4 (C_4 , C_4'), 80.3, 80.2 ($\text{C}(\text{CH}_3)_3$, *Boc*) 74.4, 74.3 (C_3 , C_3'), 74.0, 74.0, 73.4, 72.9 (2C), 72.8 (CH_2Ph), 72.5, 72.4 (C_5 , C_5'), 69.5, 69.0 (C_8 , C_8'), 63.1, 62.4 (C_6 , C_6'), 58.9 (2C, C_2 , C_2'), 45.1, 43.7 (C_7 , C_7'), 28.3, 28.2 (CH_3 , *Boc*); ESI-HRMS calcd. for $\text{C}_{33}\text{H}_{40}\text{N}_4\text{NaO}_6$ [$\text{M}+\text{Na}$]⁺: 611.2846, found 611.2840.

1.3 (3S,4R,5R,6R,7R)-3-azido-1-benzyl-5,6-bis(benzyloxy)-7-((benzyloxy)methyl)azepan-4-ol (8)

To a solution of **4** (46 mg, 0.078 mmol) in CH₂Cl₂ (2.0 mL) was added trifluoroacetic acid (2.0 mL) and the solution was stirred at room temperature for 1 hour. The solvents were evaporated and co-evaporated with toluene to remove completely the TFA. The obtained residue was dissolved in a mixture of EtOAc/H₂O (5.0/0.5 mL) and BnBr (13 μL, 0.101 mmol), K₂CO₃ (32 mg, 0.234 mmol) were added respectively. The mixture was refluxed for 18h. After being cooled to room temperature, H₂O and EtOAc were added and the layers were separated. The aqueous layer was extracted twice with EtOAc. Then the organic layer was dried over MgSO₄, filtered and concentrated under reduced pressure. The residue was purified by flash chromatography (Cy/EtOAc: 8.5/1.5) to give **8** as colorless oil (36 mg, 80%). [α]_D +19.0 (*c* = 1.0, CHCl₃); ¹H (400 MHz, CDCl₃): 7.37-7.19 (m, 20H, H_{ar}), 4.74 (d, 1H, ²*J* = 11.5 Hz, CH₂Ph), 4.69 (d, 1H, ²*J* = 11.5 Hz, CH₂Ph), 4.54 (d, 1H, ²*J* = 11.5 Hz, CH₂Ph), 4.41 (s, 2H, 2xCH₂Ph), 4.36 (d, 1H, ²*J* = 11.5 Hz, CH₂Ph), 4.10 (ddd, 1H, *J*_{H5-H4} = 1.5 Hz, *J*_{H5-OH} = 4.0 Hz, *J*_{H5-H6} = 6.0 Hz, H₅), 4.05 (dd, 1H, *J*_{H4-H5} = 1.5 Hz, *J*_{H4-H3} = 5.5 Hz, H₄), 4.01 (d, 1H, ²*J* = 14.5 Hz, NCH₂Ph), 3.92 (d, 1H, ²*J* = 14.5 Hz, NCH₂Ph), 3.82-3.75 (m, 2H, H₃, H₆), 3.68 (dd, 1H, *J*_{H8a-H2} = 5.0 Hz, *J*_{H8a-H8b} = 9.5 Hz, H_{8a}), 4.64 (dd, 1H, *J*_{H1b-H2} = 5.0 Hz, *J*_{H8b-H8a} = 9.5 Hz, H_{8b}), 3.33 (dd, 1H, *J*_{H7a-H6} = 4.5 Hz, *J*_{H7a-H7b} = 14.5 Hz, H_{7a}), 3.03 (dt, 1H, *J*_{H2-H1} = 5.0 Hz, *J*_{H2-H3} = 6.0 Hz, H₂), 2.69 (dd, 1H, *J*_{H7b-H6} = 6.5 Hz, *J*_{H7b-H7a} = 14.5 Hz, H_{7b}), 2.55 (d, 1H, *J*_{OH-H5} = 4.0 Hz, OH); ¹³C NMR (100 MHz, CDCl₃): δ 140.1, 138.3, 138.2, 138.1 (C_{ipso}), 128.5, 128.4, 128.4, 128.3, 128.0, 127.9, 127.7, 127.7, 127.6, 127.0 (CH_{ar}), 82.1 (C₄), 76.6 (C₃), 73.8 (CH₂Ph), 73.2 (2C, C₅, CH₂Ph), 72.7 (CH₂Ph), 68.9 (C₈), 63.9 (C₆), 63.5 (C₂), 57.2 (NCH₂Ph), 51.6 (C₇); ESI-HRMS calcd. for C₃₅H₃₉N₄O₄ [M+H]⁺: 579.2971, found 579.2975.

1.4 (R)-2-azido-2-((2S,3R,4R,5R)-1-benzyl-3,4-bis(benzyloxy)-5-((benzyloxy)methyl)pyrrolidin-2-yl)ethan-1-ol (9)

To a solution of **8** (60 mg, 0.104 mmol) in toluene (1.0 mL) were added trifluoroacetic anhydride (28 μL, 0.194 mmol) and Et₃N (26 μL, 0.194 mmol). The obtained solution was refluxed for 3h and cooled to room temperature. A solution of

NaOH (10%, 5 mL) was added and the mixture was stirred for 30 minutes. EtOAc and H₂O were added and the layers were separated. The aqueous layer was extracted twice with EtOAc and the combined organic layers were dried on Na₂SO₄, filtered and evaporated. The obtained crude was purified by flash chromatography (Cy/EtOAc: 9/1) to give compound **9** (55 mg, 92%). [α]_D +1.3 (*c* = 1.0, CHCl₃); ¹H NMR (400 MHz, CDCl₃): δ 7.38-7.27 (m, 18H, H_{ar}), 7.18-71.7 (m, 2H, H_{ar}), 4.60 (d, 1H, ²*J* = 12.0 Hz, CH₂Ph), 4.56 (d, 1H, ²*J* = 12.0 Hz, CH₂Ph), 4.53 (s, 2H, CH₂Ph), 4.27 (s, 2H, CH₂Ph), 4.15-4.09 (m, 2H, H₃, H₄), 4.07 (d, 1H, ²*J* = 14.0 Hz, NCH₂Ph), 4.97-3.81 (m, 4H, H₆, H_{7a}, H_{7b}, NCH₂Ph), 3.45 (t, 1H, *J*_{H5-H4} = *J*_{H5-H6} = 6.5 Hz, H₅), 3.36-3.31 (m, 1H, H_{8a}), 3.18-3.12 (m, 2H, H_{8b}, H₂); ¹³C NMR (100 MHz, CDCl₃): δ 138.2, 138.1, 138.1, 137.4 (C_{ipso}), 129.4, 128.4, 128.2, 127.7, 127.5, 127.5, 127.4 (CH_{ar}), 83.5 (C₄), 81.5 (C₃), 72.8, 72.4, 71.7 (CH₂Ph), 69.8 (C₈), 67.3 (C₅), 66.4 (C₂), 63.6 (C₇), 62.6 (C₆), 61.5 (NCH₂Ph); ESI-HRMS calcd. for C₃₅H₃₉N₄O₄ [M+H]⁺: 579.2971, found 579.2947.

1.5 (R)-2-acetamido-2-((2R,3R,4R,5R)-1-benzyl-3,4-bis(benzyloxy)-5-(benzyloxy)methylpyrrolidin-2-yl)ethyl acetate (**10**)

To a solution of azide **9** (28 mg, 0.044 mmol) in THF/H₂O (2.0 mL/1.0mL) was added Ph₃P (35 mg, 0.132 mmol) and the resulting solution was stirred at 65 °C for 2h. The solution was cooled to room temperature, solvents were evaporated and the crude was dried for 2h under reduced pressure. The residue was dissolved in pyridine (2.0 mL) and Ac₂O (1.0 mL) was added at 0°C. The resulting solution was then stirred for 12 h at room temperature. Pyridine and Ac₂O were removed by evaporation and co-evaporation with toluene (5 x 3 mL). The residue was purified by flash chromatography (cyclohexane/AcOEt: 6/4) to give **10** (22 mg, 76%). [α]_D -22.9 (*c* = 1.05, CHCl₃); ¹H NMR (400 MHz, CDCl₃): 7.36-7.22 (m, 16H, H_{ar}), 7.19-71.5 (m, 4H, H_{ar}), 4.73-4.67 (m, 1H, H₆), 4.62 (d, 1H, ²*J* = 12.0 Hz, CH₂Ph), 4.51 (d, 1H, ²*J* = 12.0 Hz, CH₂Ph), 4.41 (d, 1H, ²*J* = 10.5 Hz, CH₂Ph), 4.34 (d, 1H, ²*J* = 12.0 Hz, CH₂Ph), 4.31(d, 1H, ²*J* = 10.5 Hz, CH₂Ph), 4.19 (d, 1H, ²*J* = 12.0 Hz, CH₂Ph), 4.15 (dd, 1H, *J*_{H7a-H6} = 6.5 Hz, *J*_{H7a-H7b} = 10.5 Hz, H_{7a}), 4.11 (dd, 1H, *J*_{H7b-H6} = 6.5 Hz, *J*_{H7b-H7a} = 10.5 Hz, H_{7b}), 4.02-3.98 (m, 3H, H₃, H₄, NCH₂Ph), 3.55 (d, 1H, ²*J* = 13.0 Hz, NCH₂Ph), 3.27 (t, 1H, *J*_{H5-H4} = *J*_{H5-H6} = 4.5 Hz, H₅), 3.20-3.14 (m, 2H, H_{8a}, H₂), 2.91 (dd, *J*_{8b,2} = 10.0 Hz, *J*_{8b,8a} = 16.0 Hz, H_{8b}), 2.05 (s, 3H, CH₃, Ac), 1.77 (s, 3H, CH₃,

Ac); ^{13}C NMR (100 MHz, CDCl_3): δ 170.9, 170.0, (CO), 138.5, 138.5, 138.1, 137.1 (C_{ipso}), 129.5, 128.7, 128.5, 128.3, 128.3, 127.9, 127.8, 127.8, 127.5, 127.5, 127.2 (CH_{ar}), 84.4 (C_4), 80.9 (C_3), 72.9, 71.7 (CH_2Ph), 71.5 (C_8), 70.8 (CH_2Ph), 68.0 (C_2), 64.8 (C_5), 64.6 (C_7), 57.4 (NCH_2Ph), 46.7 (C_6), 23.3, 21.0 (CH_3 , Ac); ESI-HRMS calcd. for $\text{C}_{39}\text{H}_{45}\text{N}_2\text{O}_6$: $[\text{M}+\text{H}]^+$: 637.3278, found 637.3299.

1.6 N-((R)-1-((2R,3R,4R,5R)-3,4-dihydroxy-5-(hydroxymethyl)pyrrolidin-2-yl)-2-hydroxyethyl)acetamide (11)

A solution of **10** (20 mg, 0.314 mmol) in MeOH/ H_2O / Et_3N (4/0.5/0.5 mL) was stirred for 18h at room temperature. The solvents were evaporated and co-evaporated three times with toluene. The obtained residue was dissolved in MeOH (2 mL) and aqueous HCl (1M, 0.2 mL) was added under argon. After addition of Pd/C (10%, 20 mg), the argon was removed. The H_2 was introduced and the mixture was bubbled for 5 minutes. After stirring the solution for 24 under H_2 atmosphere, the mixture was filtered on micro-filter (0.3 μm). The solvent was evaporated to give compound **11** (6 mg, 82%) as a white solid. $[\alpha]_{\text{D}}^{18} +22.7$ ($c = 0.5$, MeOH); ^1H NMR (400 MHz, D_2O): 4.46 (dt, 1H, $J_{\text{H}_6-\text{H}_7\text{a}} = J_{\text{H}_6-\text{H}_7\text{b}} = 5.5$ Hz, $J_{\text{H}_6-\text{H}_5} = 10.5$ Hz, H_6), 4.23 (d, 1H, $J_{\text{H}_4-\text{H}_5} = 2.5$ Hz, H_4), 4.13 (dd, 1H, $J_{\text{H}_3-\text{H}_4} = 1.0$ Hz, $J_{\text{H}_3-\text{H}_2} = 2.0$ Hz, H_3), 4.01 (dd, 1H, $J_{\text{H}_8\text{a}-\text{H}_2} = 5.0$ Hz, $J_{\text{H}_8\text{a}-\text{H}_8\text{b}} = 12.0$ Hz, $\text{H}_{8\text{a}}$), 3.95-3.87 (m, 2H, $\text{H}_{8\text{b}}$, H_5), 3.84 (dd, 1H, $J_{\text{H}_7\text{a}-\text{H}_6} = 5.5$ Hz, $J_{\text{H}_7\text{a}-\text{H}_7\text{b}} = 12.0$ Hz, $\text{H}_{7\text{a}}$), 3.77 (dd, 1H, $J_{\text{H}_7\text{b}-\text{H}_6} = 5.5$ Hz, $J_{\text{H}_7\text{b}-\text{H}_7\text{a}} = 12.0$ Hz, $\text{H}_{7\text{b}}$), 3.71 (ddd, 1H, $J_{\text{H}_2-\text{H}_3} = 2.0$ Hz, $J_{\text{H}_2-\text{H}_8\text{a}} = 5.0$ Hz, $J_{\text{H}_2-\text{H}_8\text{b}} = 8.0$ Hz, H_2), 2.08 (s, 3H, CH_3 , Ac); ^{13}C NMR (100 MHz, D_2O): δ 174.6 (CO), 75.0 (C_4), 74.9 (C_3), 69.0 (C_2), 61.9 (C_5), 61.1 (C_7), 59.6 (C_8), 47.4 (C_6), 21.9 (CH_3 , Ac); ESI-HRMS calcd. for $\text{C}_9\text{H}_{19}\text{N}_2\text{O}_5$ $[\text{M}+\text{H}]^+$: 235.1294, found 235.1297.

1.7 N-((R)-1-((2R,3R,4R,5R)-3,4-dihydroxy-5-(hydroxymethyl)pyrrolidin-2-yl)-2-hydroxyethyl)amonium (12)

9 (10 mg, 0.017 mmol) was dissolved in MeOH (2 mL) and aqueous HCl (1M, 0.2 mL) was added under argon. After adding Pd/C (10%, 10 mg), the argon was removed. The H_2 was introduced and the mixture was bubbled for 5 minutes. After stirring the solution for 24 under H_2 atmosphere, the mixture was filtered on micro-filter (0.3 μm). The solvent was evaporated to give the desired product (4.5 mg, 95%).

$[\alpha]_D^{18} +63.7$ ($c = 0.2$, MeOH); ^1H RMN (400 MHz, D_2O) δ 4.41 (dd, 1H, $J_{\text{H}_4\text{-H}_3} = 1.0$ Hz, $J_{\text{H}_4\text{-H}_5} = 3.0$ Hz, H_4), 4.22 (dd, 1H, $J_{\text{H}_3\text{-H}_4} = 1.0$ Hz, $J_{\text{H}_3\text{-H}_2} = 2.5$ Hz, H_3), 4.16 (dd, $J_{\text{H}_5\text{-H}_4} = 3.0$ Hz, $J_{\text{H}_5\text{-H}_6} = 9.0$ Hz, H_5), 4.07-4.00 (m, 3H, H_{8a} , H_6 , H_{7a}), 3.94-3.89 (m, 2H, H_{8b} , H_{7b}), (ddd, 1H, $J_{\text{H}_2\text{-H}_3} = 2.5$ Hz, $J_{\text{H}_2\text{-H}_{8a}} = 4.5$ Hz, $J_{\text{H}_2\text{-H}_{8b}} = 8.5$ Hz, H_2); ^{13}C RMN (100 MHz, D_2O) δ 75.6 (C_3), 74.1 (C_4), 68.8 (C_2), 59.5 (2C, C_5 , C_7), 59.0 (C_8), 48.6 (C_6), HRMS calcd for $\text{C}_7\text{H}_{17}\text{N}_2\text{O}_4$: $[\text{MH}]^+$: 193.1188 found 193.1191.

1.8 tert-butyl (2R,3R,4R,5S,6R)-6-azido-3,4-bis(benzyloxy)-2-((benzyloxy)methyl)-5-hydroxyazepane-1-carboxylate (6)

To a solution of known epoxide **3**³³ (160 mg, 0.294 mmol) in a mixture of DMF/ H_2O (2.9/0.3 mL) was added NaN_3 (88 mg, 1.358 mmol) followed by NH_4Cl (53 mg, 1.358 mmol). The mixture was stirred for 28 h at 90 °C. EtOAc (50 mL) and H_2O (50 mL) were added. The layers were separated and the aqueous layer was extracted with EtOAc (50 mL). The combined organic layers were dried on MgSO_4 , filtered and evaporated. The residue was purified by flash chromatography (Cy/ EtOAc: 96/4 - 95/5) to give compound **7**⁶ (90 mg, 52%) and **6** (70 mg, 40%) as a pale yellow oil. $[\alpha]_D -38.5$ ($c = 1.0$, CHCl_3); ^1H NMR (400 MHz, CDCl_3 , 2 rotamers): δ 7.37-7.26 (m, 26H, H_{ar}), 7.214-7.18 (m, 4H, H_{ar}), 4.78 (d, 2H, $^2J = 11.5$ Hz, CH_2Ph), 4.72, 4.71 (2s, 4H, CH_2Ph), 4.51 (d, 2H, $^2J = 11.5$ Hz, CH_2Ph), 4.47 (d, 2H, $^2J = 12.0$ Hz, CH_2Ph), 4.38 (d, 2H, $^2J = 12.0$ Hz, CH_2Ph), 4.27, 4.26 (2s, 2H, OH, OH'), 4.10-4.08 (m, 2H, H_2 , H_2'), 4.02-3.84 (m, 10H, H_3 , H_3' , H_4 , H_4' , H_5 , H_5' , H_6 , H_6' , H_{7a} , H_{7a}'), 3.67 (dd, 2H, $J_{\text{H}_{8a}\text{-H}_2} = 4.5$ Hz, $J_{\text{H}_{8a}\text{-H}_{8b}} = 9.5$ Hz, H_{8a} , H_{8a}'), 3.55 (dd, 2H, $J_{\text{H}_{8b}\text{-H}_2} = 4.5$ Hz, $J_{\text{H}_{8b}\text{-H}_{8a}} = 9.5$ Hz, H_{8b} , H_{8b}'), 3.31 (dd, $J_{\text{H}_{7b}\text{-H}_6} = 1.5$ Hz, $J_{\text{H}_{7b}\text{-H}_{7a}} = 15.5$ Hz, H_{7b} , H_{7b}'), 1.42 (s, 18H, CH_3 , *Boc*); ^{13}C NMR (100 MHz, CDCl_3 , 2 rotamers): δ 158.9 (CO, *Boc*), 138.0, 137.9, 137.7 (C_{ipso}) 128.4 128.4, 127.8, 127.7, 127.6, 127.5, 127.4 (CH_{ar}) 81.5 (2C, ($\text{C}(\text{CH}_3)_3$, *Boc*), 80.5 (2C, C_4 , C_4'), 74.4 (2C, C_3 , C_3'), 73.9, 72.9 (CH_2Ph , 72.0 (2C, C_5 , C_5') 68.9 (2C, C_8 , C_8'), 66.1 (2C, C_6 , C_6'), 57.6 (2C, C_2 , C_2'), 44.7 (2C, C_7 , C_7'), 28.2 (CH_3 , *Boc*); ESI-HRMS calcd. for $\text{C}_{33}\text{H}_{40}\text{N}_4\text{NaO}_6$ $[\text{M}+\text{Na}]^+$: 611.2846, found 611.2858.

1.9 (3R,4S,5R,6R,7R)-3-azido-1-benzyl-5,6-bis(benzyloxy)-7-((benzyloxy)methyl)azepan-4-ol (13)

To a solution of **6** (42 mg, 0.071 mmol) in CH₂Cl₂ (4 mL) was added trifluoroacetic acid (2 mL) and the obtained solution was stirred at room temperature for 1h. The solution was evaporated and co-evaporated with toluene (3x5 mL). The residue was dissolved in EtOAc/H₂O (4/0.4 mL) and K₂CO₃ (49 mg, 0.355 mmol), BnBr (13 μL, 0.107 mmol) were added respectively. The resulting mixture was stirred at 80 °C for 16 h and cooled to room temperature. EtOAc and H₂O were added and the layers were separated. The aqueous layer was extracted twice with EtOAc and the combined organic layers were dried on Na₂SO₄, filtered and evaporated. The obtained crude was purified by flash chromatography to give **13** (28 mg, 68%). [α]_D +43.7 (*c* = 1.0, CHCl₃); ¹H NMR(400 MHz, CDCl₃): 7.40-7.26 (m, 18H, H_{ar}), 7.20-7.18 (m, 2H, H_{ar}), 5.06 (d, 1H, ²*J* = 11.0 Hz, CH₂Ph), 4.93 (d, 1H, ²*J* = 11.0 Hz, CH₂Ph), 4.63 (d, 1H, ²*J* = 11.0 Hz, CH₂Ph), 4.47-4.40 (m, 3H, CH₂Ph), 3.95-3.88 (m, 2H, H₄, NCH₂Ph), 3.77 (d, 1H, ²*J* = 13.0 Hz, NCH₂Ph), 3.10-3.63 (m, 2H, H_{8a}, H₃), 3.61 (dd, 1H, *J*_{H8b-H2} = 3.5 Hz, *J*_{H8b-H8a} = 10.0 Hz, H_{8b}), 3.53 (t, 1H, *J*_{H5-H4} = *J*_{H5-H6} = 8.0 Hz, H₅), 3.31 (s, 1H, OH), 3.21 (ddd, 1H, *J*_{H6-H7b} = 4.0 Hz, *J*_{H6-H5} = 8.0 Hz, *J*_{H6-7a} = 11.5 Hz, H₆), 3.12 (dd, 1H, *J*_{H7a-H6} = 11.5 Hz, *J*_{H7a-7b} = 14.0 Hz, H_{7a}), 2.90 (dt 1H, *J*_{H2-H1a} = *J*_{H2-H1b} = 3.5 Hz, *J*_{H2-H3} = 9.0 Hz, H₂), 2.63 (dd, 1H, *J*_{H7b-H6} = 4.0, *J*_{H7b-H7a} = 14.0 Hz, H_{7b}); ¹³C NMR (100 MHz, CDCl₃): δ 139.1, 138.3, 138.1, 138.0 (C_{ipso}), 128.8, 128.6, 128.4, 128.4, 128.4, 128.1, 128.0, 127.7, 127.7, 127.4 (CH_{ar}), 83.2 (C₄), 79.2 (C₃), 78.1 (C₂), 76.1, 75.3, 73.2 (CH₂Ph), 67.8 (C₈), 64.4 (C₆), 63.7 (C₂), 59.6 (NCH₂Ph), 48.9 (C₇); ESI-HRMS calcd. for C₃₅H₃₉N₄O₄ [M+H]⁺: 579.2971, found 579.2952.

1.10 (S)-2-azido-2-((2R,3R,4R,5R)-1-benzyl-3,4-bis(benzyloxy)-5-(benzyloxy)methylpyrrolidin-2-yl)ethan-1-ol (**14**)

To a solution of **13** (28 mg, 0.048 mmol) in toluene (0.5 mL) were added trifluoroacetic anhydride (14 μL, 0.1 mmol) and Et₃N (13 μL, 0.097 mmol). The obtained solution was refluxed for 3 h and cooled to room temperature. A solution of NaOH (10%, 2 mL) was added and the mixture was stirred for 30 minutes. AcOEt and H₂O were added and the layers were separated. The aqueous layer was extracted twice with AcOEt and the combined organic layers were dried on Na₂SO₄, filtered and evaporated. The obtained crude was purified by flash chromatography (Cyclohexane/AcOEt: 95/5) to give compound **14** (24 mg, 86%). [α]¹⁹_D +16.8 (*c* = 0.5, CHCl₃); ¹H NMR (400 MHz, CDCl₃): δ 7.38-7.22 (m, 20H, H_{ar}), 4.56 (d, 1H, ²*J*

= 12.0 Hz, CH₂Ph), 4.50-4.43 (m, 5H, CH₂Ph), 4.16 (dd, 1H, $J_{\text{H4-H3}} = 2.0$ Hz, $J_{\text{H4-H5}} = 4.0$ Hz, H₄), 4.10 (t, 1H, $J_{\text{H3-H2}} = J_{\text{H3-H4}} = 2.0$ Hz, H₃), 4.05 (d, 1H, $^2J = 14.0$ Hz, NCH₂Ph), 3.86 (td, 1H, $J_{\text{H6-H5}} = 4.0$ Hz, $J_{\text{H6-H7a}} = J_{\text{H6-H7b}} = 6.5$ Hz, H₆), 3.76 (d, 1H, $^2J = 14.0$ Hz, NCH₂Ph), 3.68-3.56 (m, 4H, H_{8a}, H_{8b}, H_{7a}, H_{7b}), 3.46 (td, 1H, $J_{\text{H2-H3}} = 2.0$ Hz, $J_{\text{H2-H8a}} = J_{\text{H2-H8b}} = 6.0$ Hz, H₂), 3.27 (t, $J_{\text{H5-H4}} = J_{\text{H5-H6}} = 4.0$ Hz, H₅), 3.07 (s, 0.9 H, OH); ¹³C NMR (100 MHz, CDCl₃): δ 138.2, 138.0, 137.9, 137.6 (C_{ipso}), 128.5, 128.4, 128.3, 128.3, 127.9, 127.9, 127.7, 127.6, 127.6, 127.1 (CH_{ar}), 85.7 (C₄), 83.6 (C₃), 73.2, 71.7, 71.4 (CH₂Ph), 68.9 (C₅), 66.6 (C₈), 63.2 (C₂), 62.9 (C₇), 62.0 (C₆), 51.9 (NCH₂Ph); ESI-HRMS calcd. for C₃₅H₃₉N₄O₄ [M+H]⁺: 579.2971, found 579.2977.

1.11 (S)-2-acetamido-2-((2S,3R,4R,5R)-1-benzyl-3,4-bis(benzyloxy)-5-((benzyloxy)methyl)pyrrolidin-2-yl)ethyl acetate (15)

To a solution of azide **14** (24 mg, 0.042 mmol) in THF/H₂O (2.0 mL/1.0mL) was added Ph₃P (32 mg, 0.125 mmol) and the resulting solution was stirred at 65 °C for 2 h. The solution was cooled to room temperature, solvents were evaporated and the reaction crude was dried for 2 h under reduced pressure. The residue was dissolved in pyridine (2.0 mL) and Ac₂O (1.0 mL) was added at 0°C. The resulting solution was then stirred for 12 h at room temperature. Pyridine and Ac₂O were removed by evaporation and co-evaporation with toluene (5 x 3 mL). The residue was purified by flash chromatography (Cy/EtOAc: 6.5/3.5) to give **15** as a white solid (16 mg, 60%). $[\alpha]_{\text{D}} -12.3$ ($c = 0.5$, CHCl₃); ¹H NMR (400 MHz, CDCl₃): 7.36-7.26 (m, 20H, H_{ar}), 6.67 (d, 1H, $J_{\text{NH-H6}} = 6.5$ Hz, NHAc), 4.56 (d, 1H, $^2J = 11.0$ Hz, CH₂Ph), 4.51-4.44 (m, 5H, CH₂Ph), 4.32-4.27 (m, 1H, H₆), 4.14 (dd, 1H, $J_{\text{H7a-H6}} = 5.0$ Hz, $J_{\text{H7a-H7b}} = 11.0$ Hz, H_{7a}), 4.08 (bs, 1H, H₃), 3.99 (dd, 1H, $J_{\text{H7b-H6}} = 7.5$ Hz, $J_{\text{H7b-H7a}} = 11.0$ Hz, H_{7b}), 3.94 (bs, 1H, H₄), 3.90 (d, 1H, $^2J = 14.5$ Hz, NCH₂Ph), 3.81 (d, 1H, $^2J = 14.5$ Hz, NCH₂Ph), 3.71 (dd, 1H, $J_{\text{H8a-H2}} = 4.5$ Hz, $J_{\text{H8a-H8b}} = 9.0$ Hz, H_{8a}), 3.54 (t, 1H, $J_{\text{H8b-H2}} = J_{\text{H8b-H8a}} = 9.0$ Hz, H_{8b}), 3.50 (dd, 1H, $J_{\text{H2-H8a}} = 4.5$ Hz, $J_{\text{H2-H8b}} = 9.0$ Hz, H₂), 3.40-3.39 (m, 1H, H₅), 1.97 (s, 3H, CH₃, Ac), 1.56 (s, 3H, CH₃, Ac); ¹³C NMR (100 MHz, CDCl₃): δ 170.7, 170.5 (CO), 139.0, 138.2, 137.6, 137.1 (C_{ipso}), 128.7, 128.6, 128.5, 128.4, 128.4, 128.4, 128.3, 128.2, 128.1, 127.9, 127.7, 127.6, 127.6, 127.0 (CH_{ar}), 83.9 (C₄), 82.6 (C₃), 73.3, 71.6, 71.4 (CH₂Ph), 68.0 (C₈) 66.7 (C₅), 64.2 (C₂), 63.7

(C₇), 50.7 ((NCH₂Ph), 47.0 (C₆), 22.8, 20.7 (CH₃, Ac); ESI-HRMS calcd. for C₃₉H₄₅N₂O₆ [M+H]⁺: 637.3278 found 637.3281.

1.12 N-((S)-1-((2S,3R,4R,5R)-3,4-dihydroxy-5-(hydroxymethyl)pyrrolidin-2-yl)-2-hydroxyethyl)acetamide (**16**)

A solution of **15** (8 mg, 0.013 mmol) in MeOH/H₂O/Et₃N (2/0.25/0.25 mL) was stirred for 18 h at room temperature. The solvents were evaporated and co-evaporated three times with toluene. The obtained residue was dissolved in MeOH (1 mL) and aqueous HCl (1M, 0.1 mL) was added under argon. After adding Pd/C (10%, 10 mg), the argon was removed. The H₂ was introduced and the mixture was bubbled for 5 minutes. After stirring the solution for 24 under H₂ atmosphere, the mixture was filtered on micro-filter (0.3 μm). The solvent was evaporated to give compound **16** (3 mg, 88%). $[\alpha]_{\text{D}}^{22} = +18.3$ ($c = 0.16$, MeOH); ¹H NMR (400 MHz, D₂O): 4.44 (q, 1H, $J_{\text{H6-H7a}} = J_{\text{H6-H7b}} = J_{\text{H6-H5}} = 5.5$ Hz, H₆), 4.20 (dd, 1H, $J_{\text{H4-H3}} = 6.5$ Hz, $J_{\text{H4-H5}} = 8.0$ Hz, H₄), 4.14 (dd, 1H, $J_{\text{H3-H4}} = 6.5$ Hz, $J_{\text{H3-H2}} = 8.0$ Hz, H₃), 3.98 (dd, 1H, $J_{\text{H8a-H2}} = 3.5$ Hz, $J_{\text{H1a-H1b}} = 12.5$ Hz, H_{8a}), 3.91 (dd, 1H, $J_{\text{H8b-H2}} = 5.5$ Hz, $J_{\text{H8b-H8a}} = 12.5$ Hz, H_{8b}), 3.84 (2d, 2H, $J = 5.5$ Hz, H_{7a}, H_{7b}), 3.78 (dd, 1H, $J_{\text{H5-H6}} = 5.5$ Hz, $J_{\text{H5-H4}} = 8.0$ Hz, H₅), 3.63 (ddd, 1H, $J_{\text{H2-H8a}} = 3.5$ Hz, $J_{\text{H2-H8b}} = 5.5$ Hz, $J_{\text{H2-H3}} = 8.0$ Hz, H₂), 2.14 (s, 3H, CH₃, Ac); ¹³C NMR (100 MHz, D₂O): δ 175.9 (CO), 74.8 (C₄), 74.3 (C₃), 62.7 (C₂), 61.7 (C₅), 60.5 (C₇), 57.5 (C₈), 51.0 (C₆), 21.8 (CH₃, Ac); ESI-HRMS calcd. for C₉H₁₈N₂NaO₄ [M+Na]⁺: 257.1113, found 257.1108.

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Highlights.

- A new family of amino-functionalised pyrrolidines was synthesized.
- A ring-contraction of β -azido, γ -hydroxyazepanes was used.
- A low micromolar inhibitor of β -N-acetylhexosaminidase identified.

ACCEPTED MANUSCRIPT

SUPPLEMENTARY INFORMATION

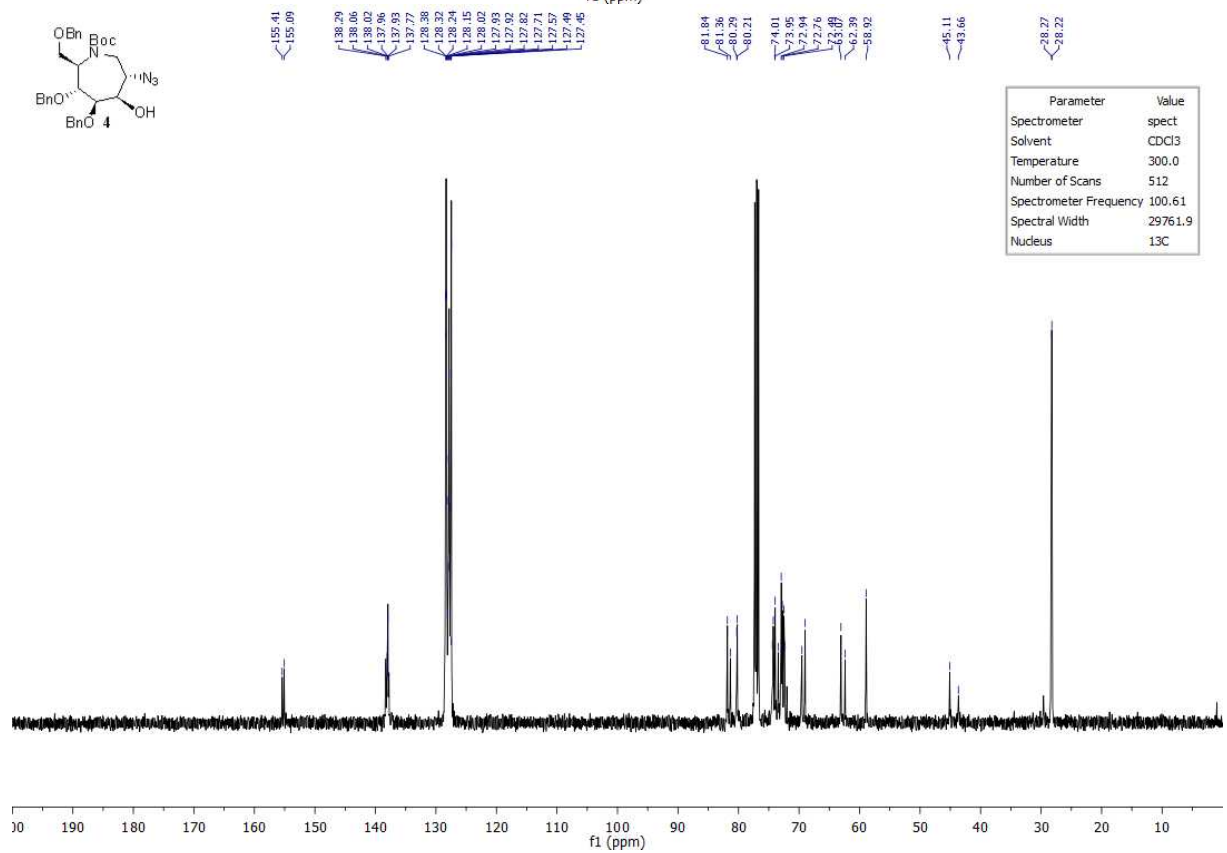
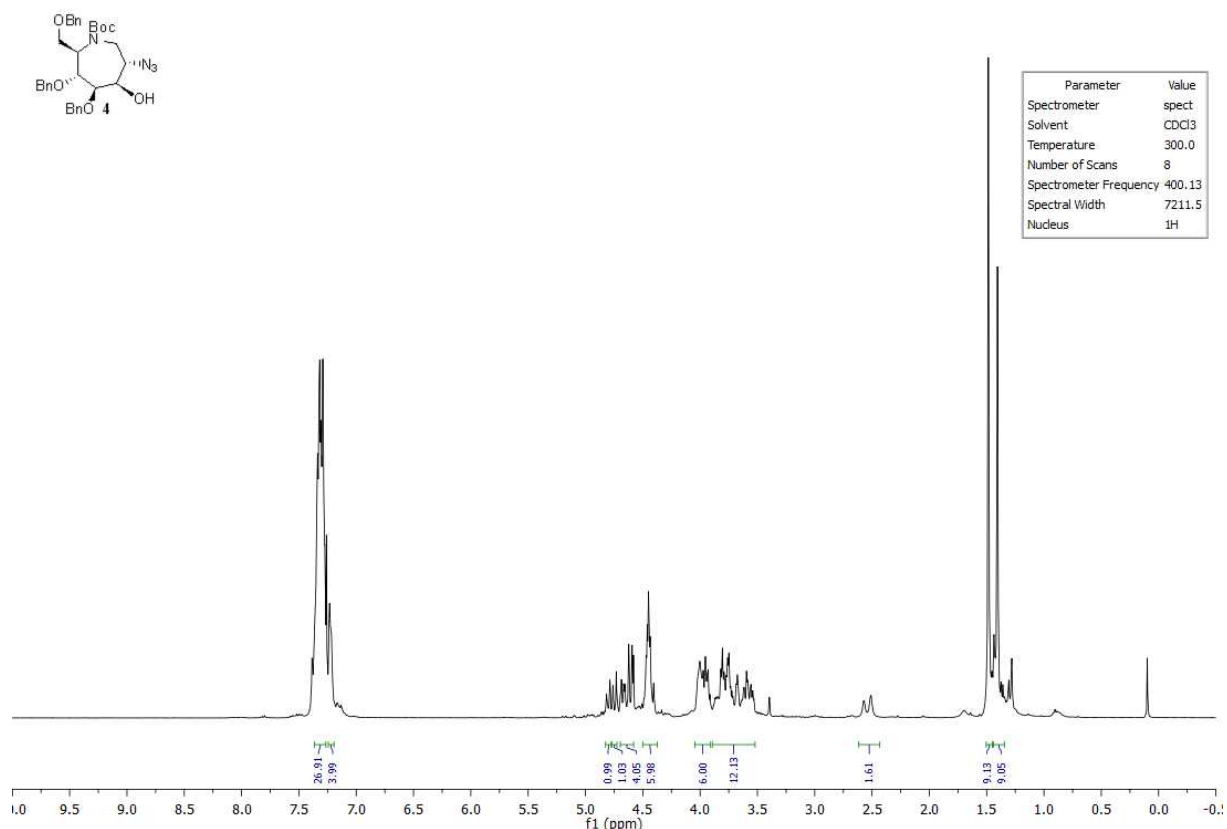
Synthesis of pyrrolidine-based analogues of 2-acetamidoglucosaminidase inhibitors

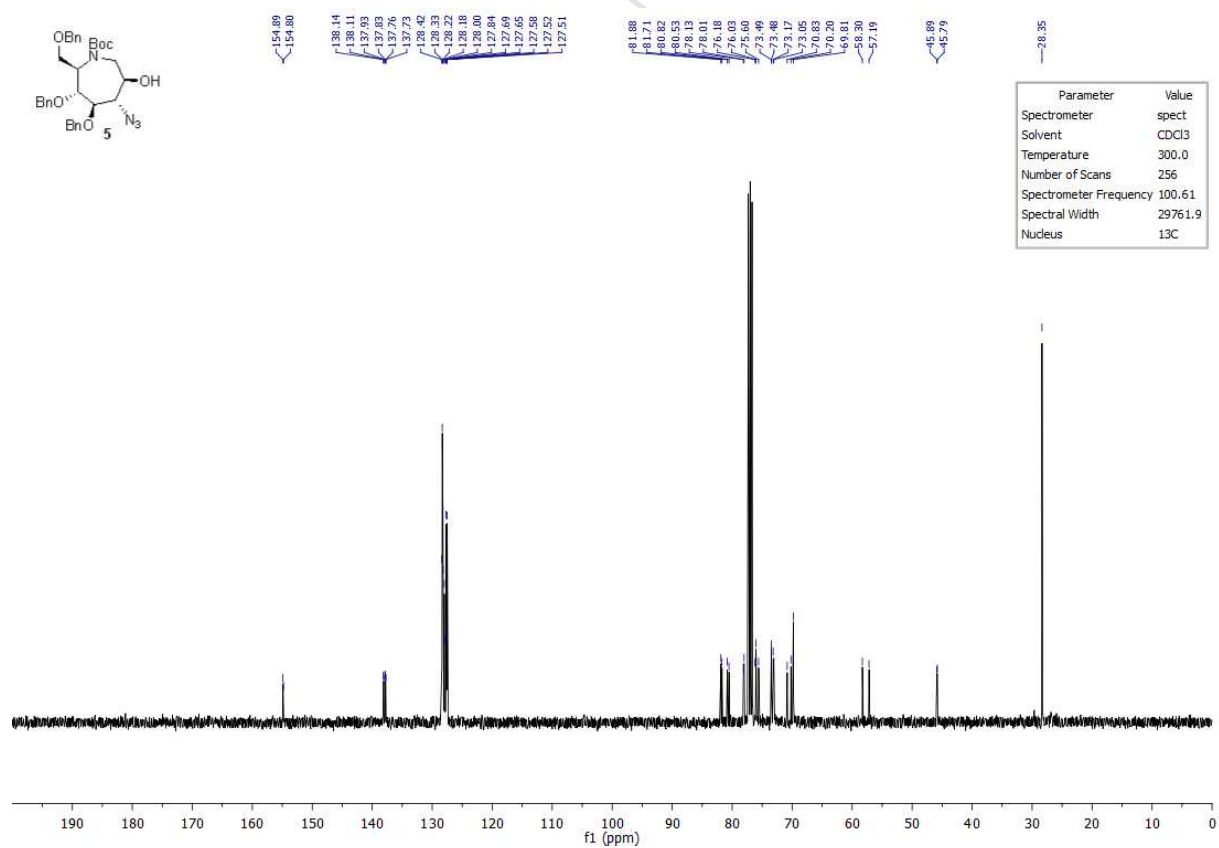
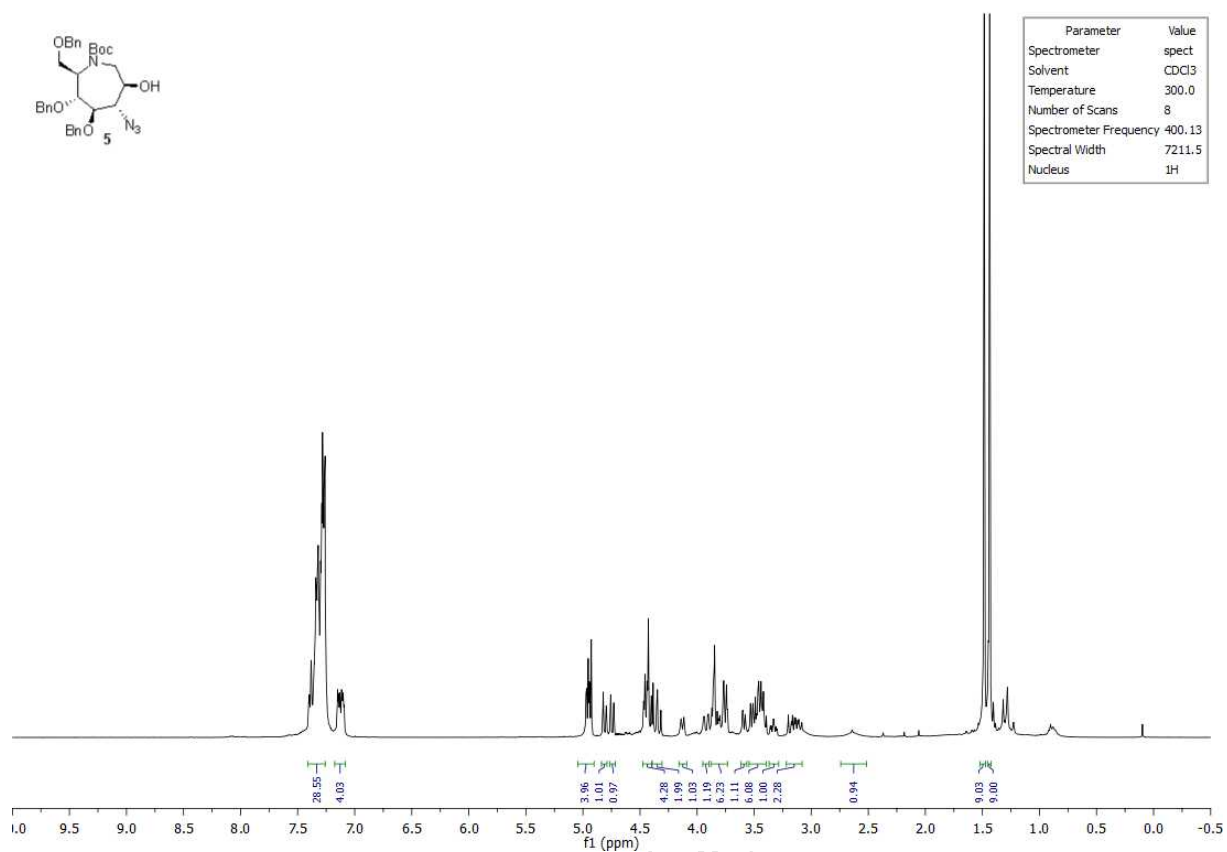
Anh Tuan Tran,^a Bo Luo,^a Jagadeesh Yerri,^b Nicolas Auberger,^b Jérôme Désiré,^b Shinpei Nakagawa,^c Atsushi Kato,^c Yongmin Zhang,^a Yves Blériot,^b * Matthieu Sollogoub^a *

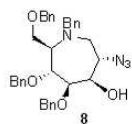
^a Sorbonne Universités, UPMC Univ Paris 06, Institut Universitaire de France, UMR-CNRS 8232, IPCM, F-75005 Paris, France. E-mail : matthieu.sollogoub@upmc.fr

^b Glycochemistry Group of “Organic Synthesis” Team, Université de Poitiers, UMR-CNRS 7285 IC2MP, 4 rue Michel Brunet, 86073 Poitiers Cedex 9, France. E-mail: yves.bleriot@univ-poitiers.fr

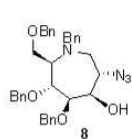
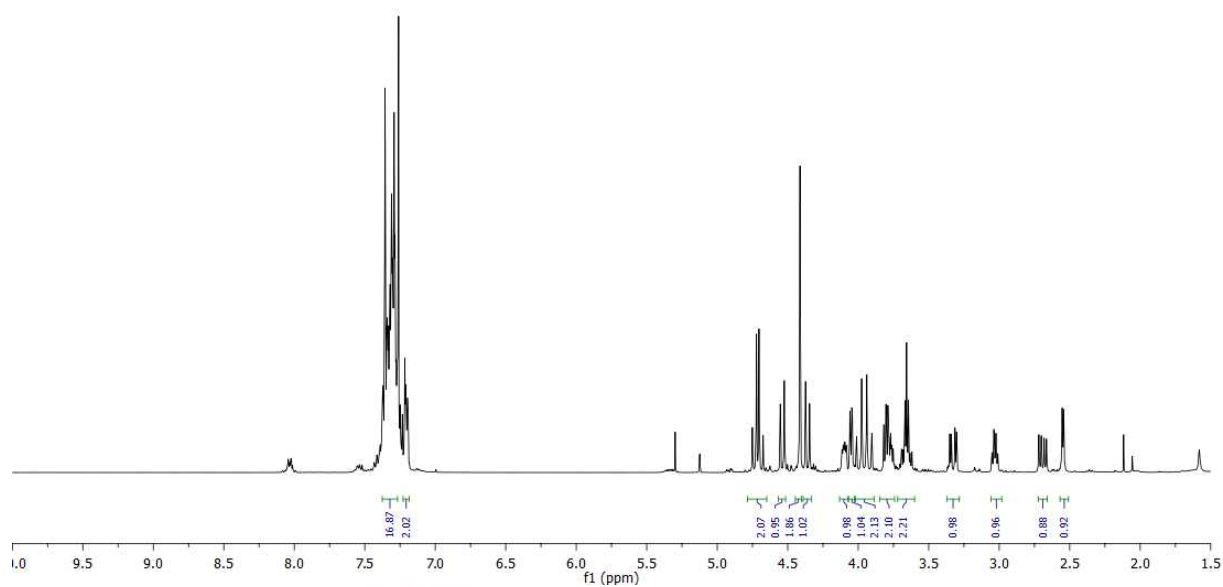
^c Department of Hospital Pharmacy, University of Toyama, 2630 Sugitani, Toyama 930-0194, Japan







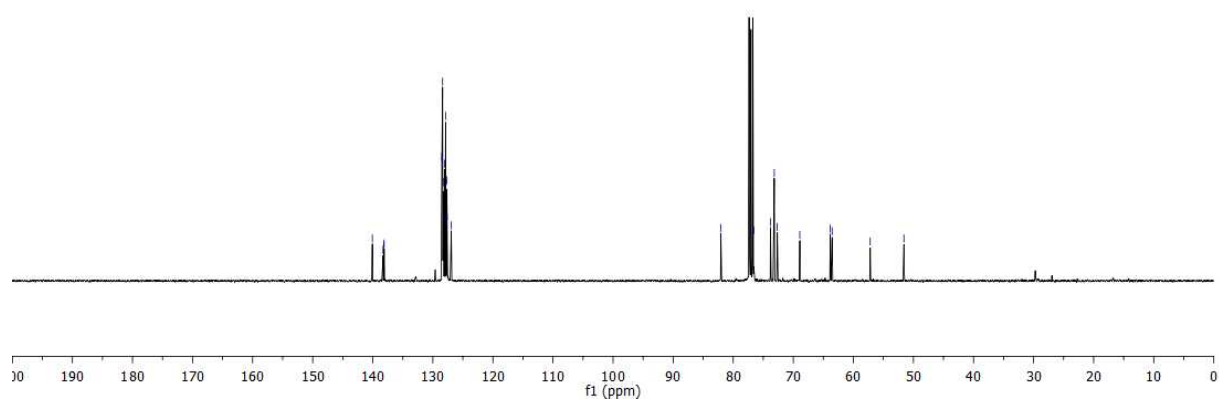
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Spectral Width	7211.5
Nucleus	¹ H

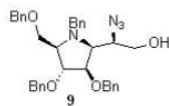


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127.62
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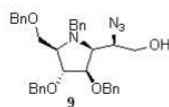
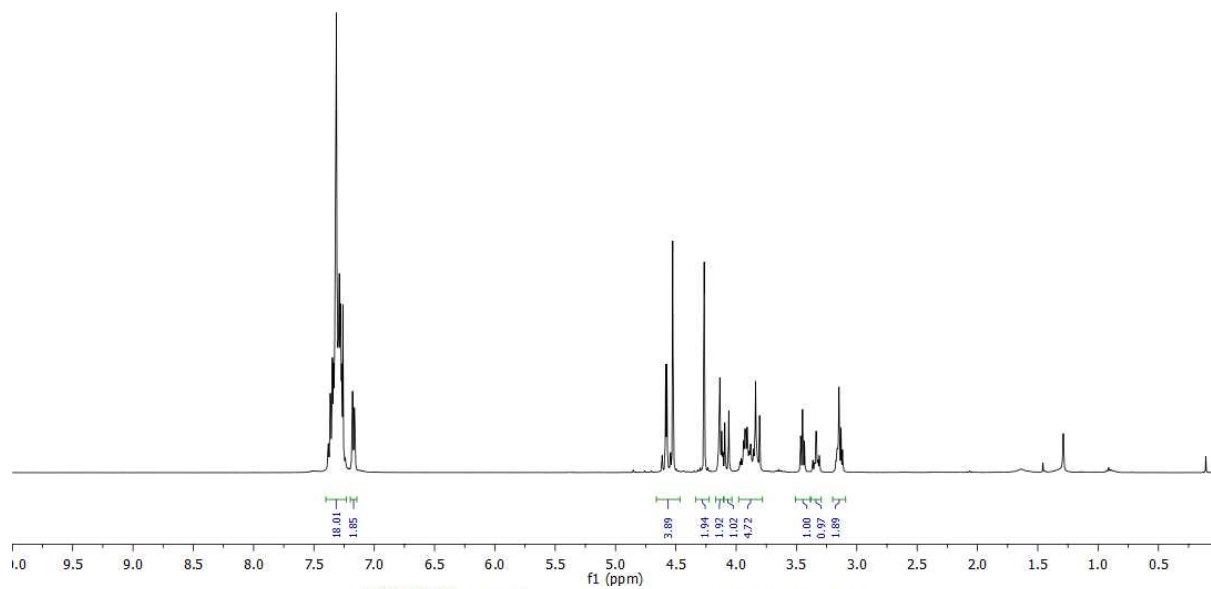
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63.95
63.53
57.20
51.58

Parameter	Value
Spectrometer	spect
Solvent	CDCl ₃
Temperature	300.0
Number of Scans	2048
Spectrometer Frequency	100.61
Spectral Width	29761.9
Nucleus	¹³ C



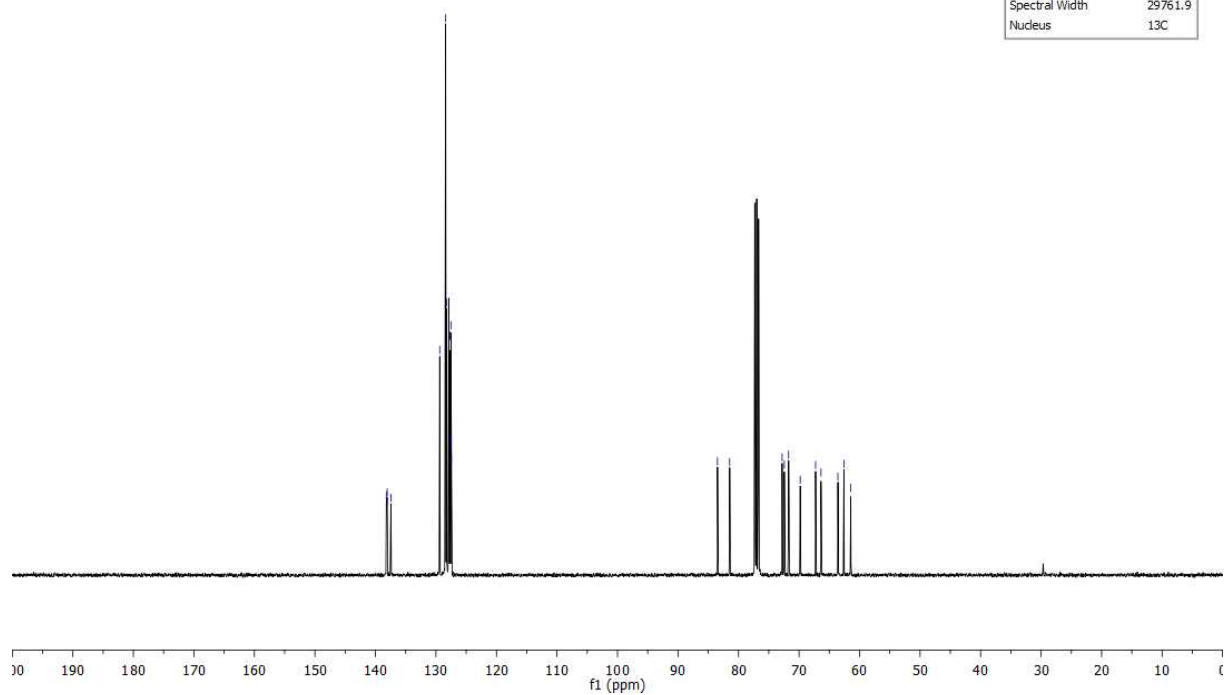


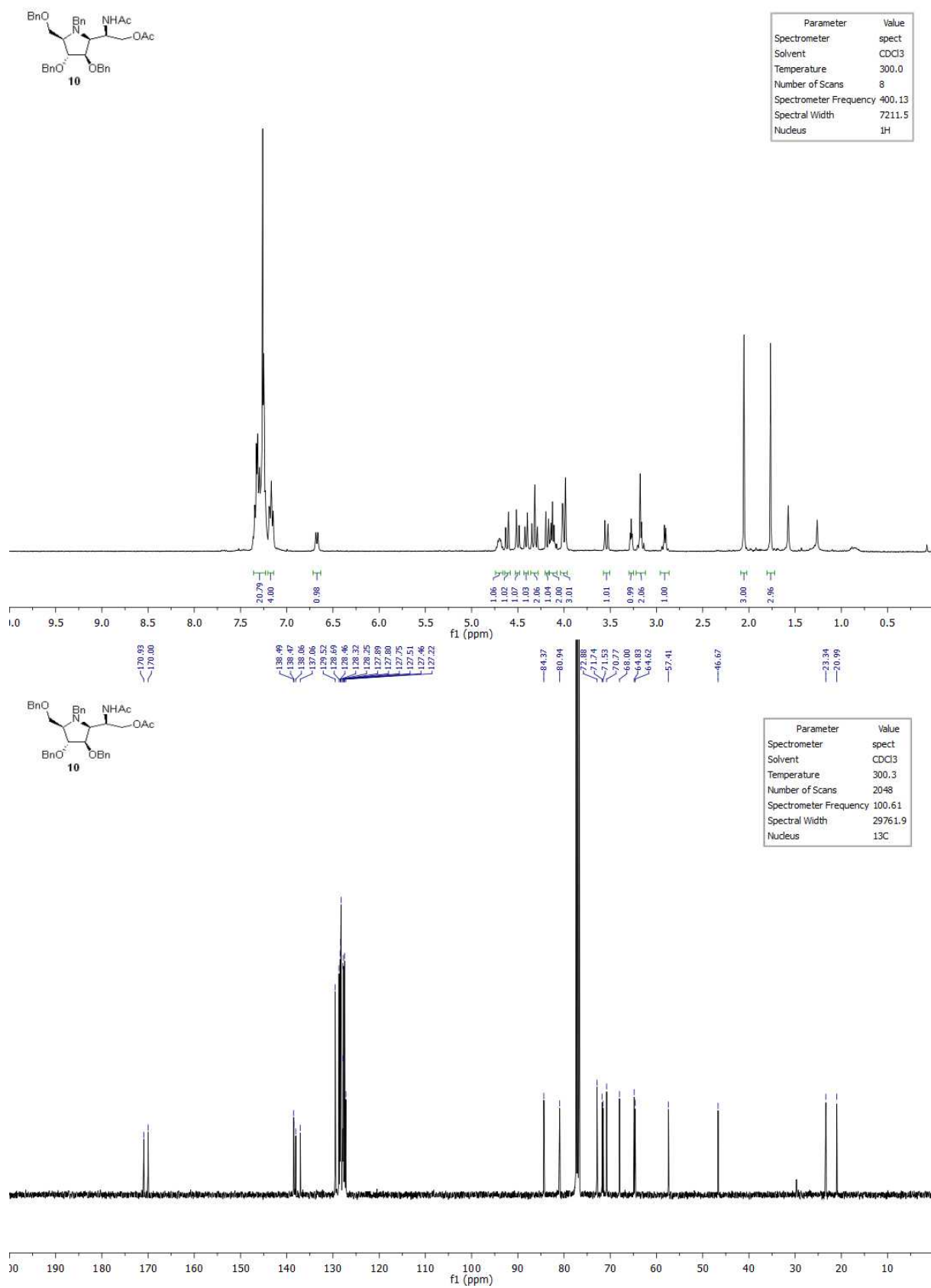
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Spectrometer	spect
Solvent	CDCl3
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Number of Scans	8
Spectrometer Frequency	400.13
Spectral Width	7211.5
Nucleus	¹ H

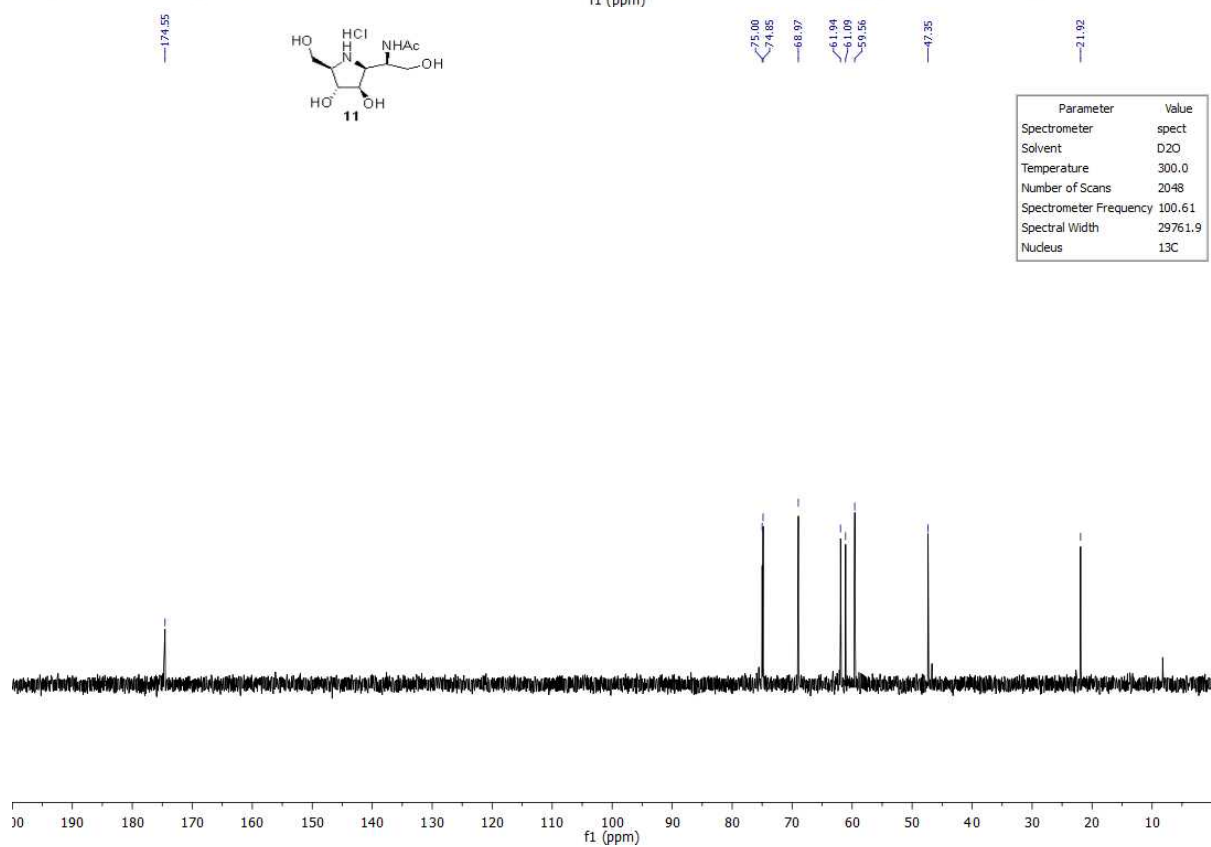
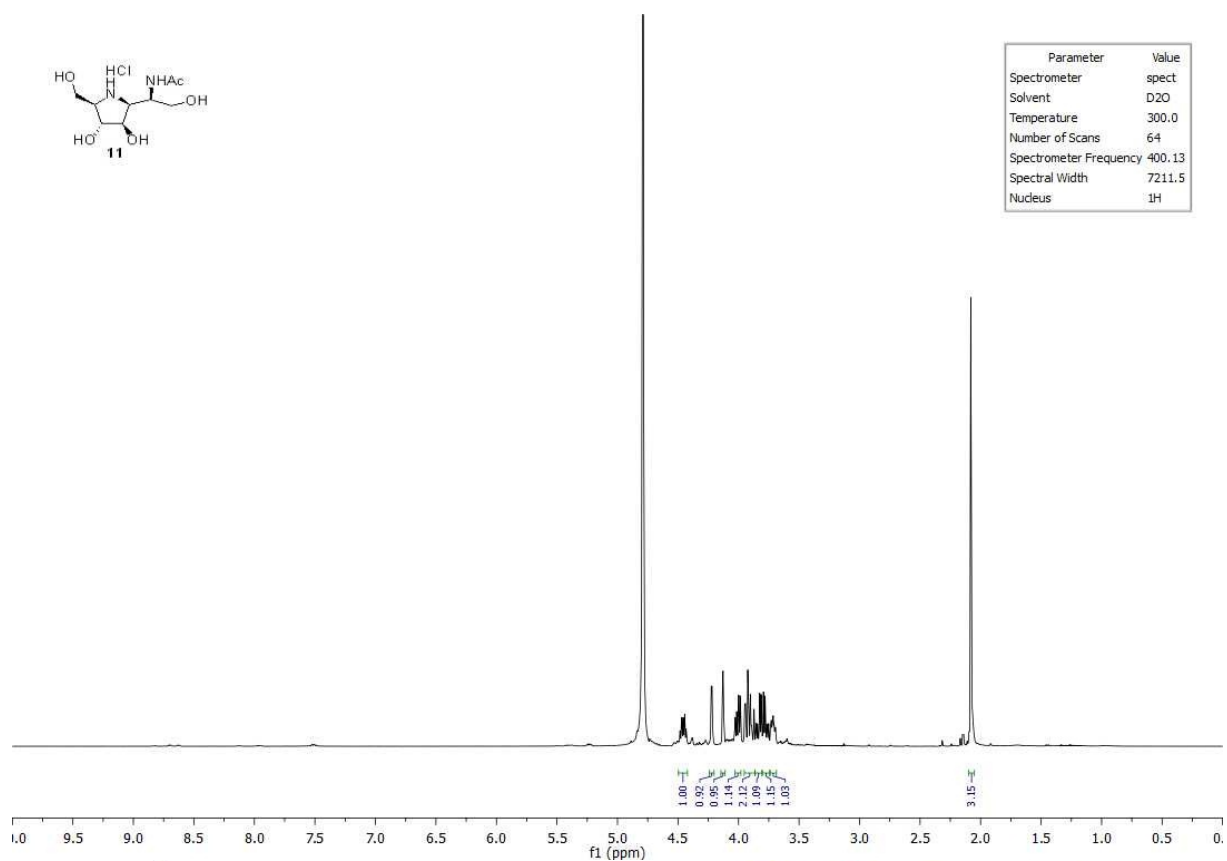


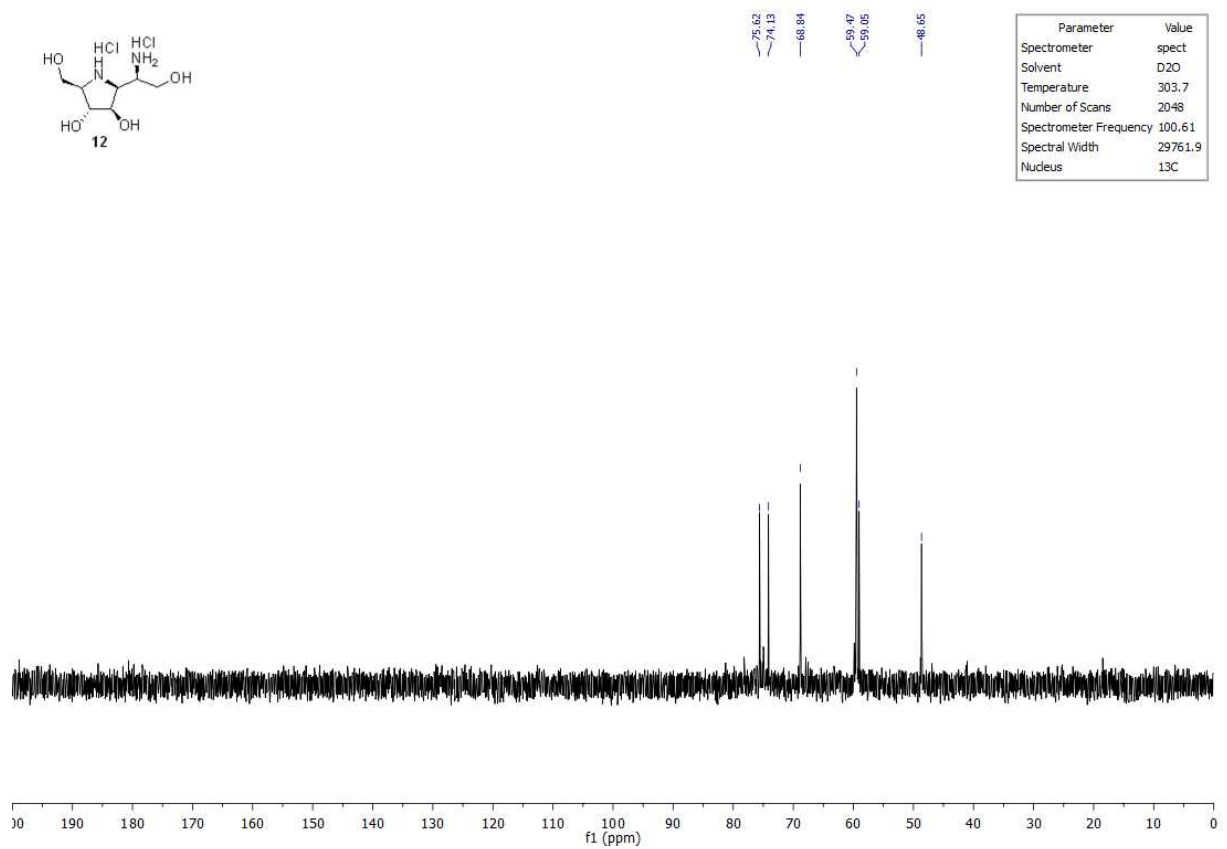
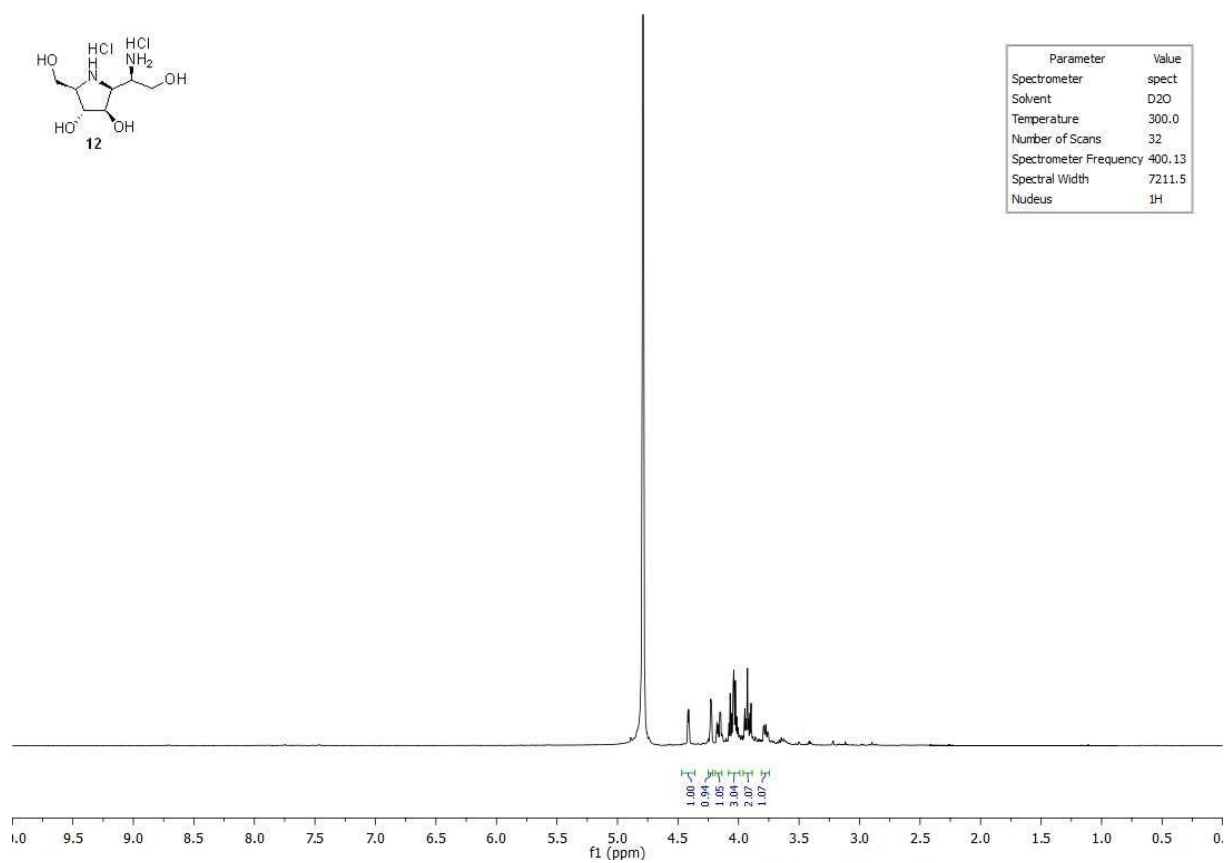
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72.43
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61.46

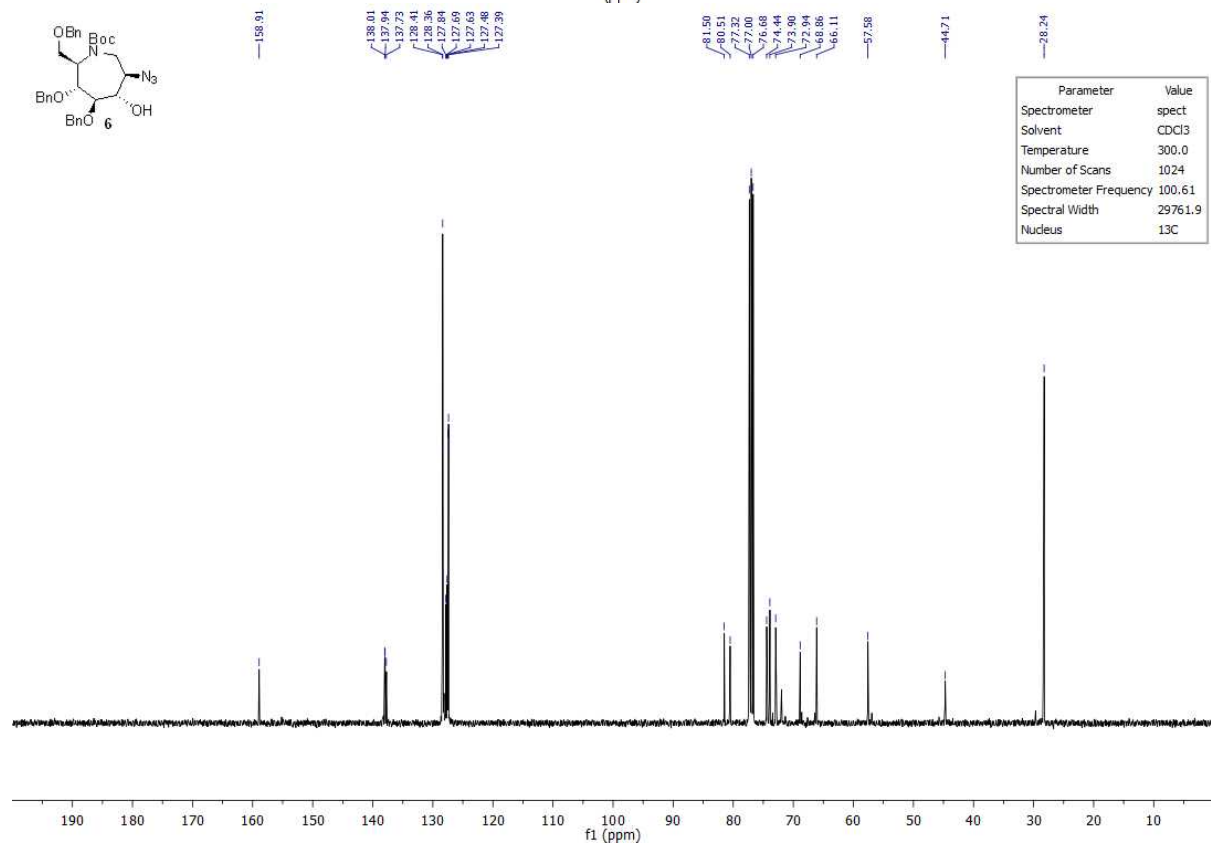
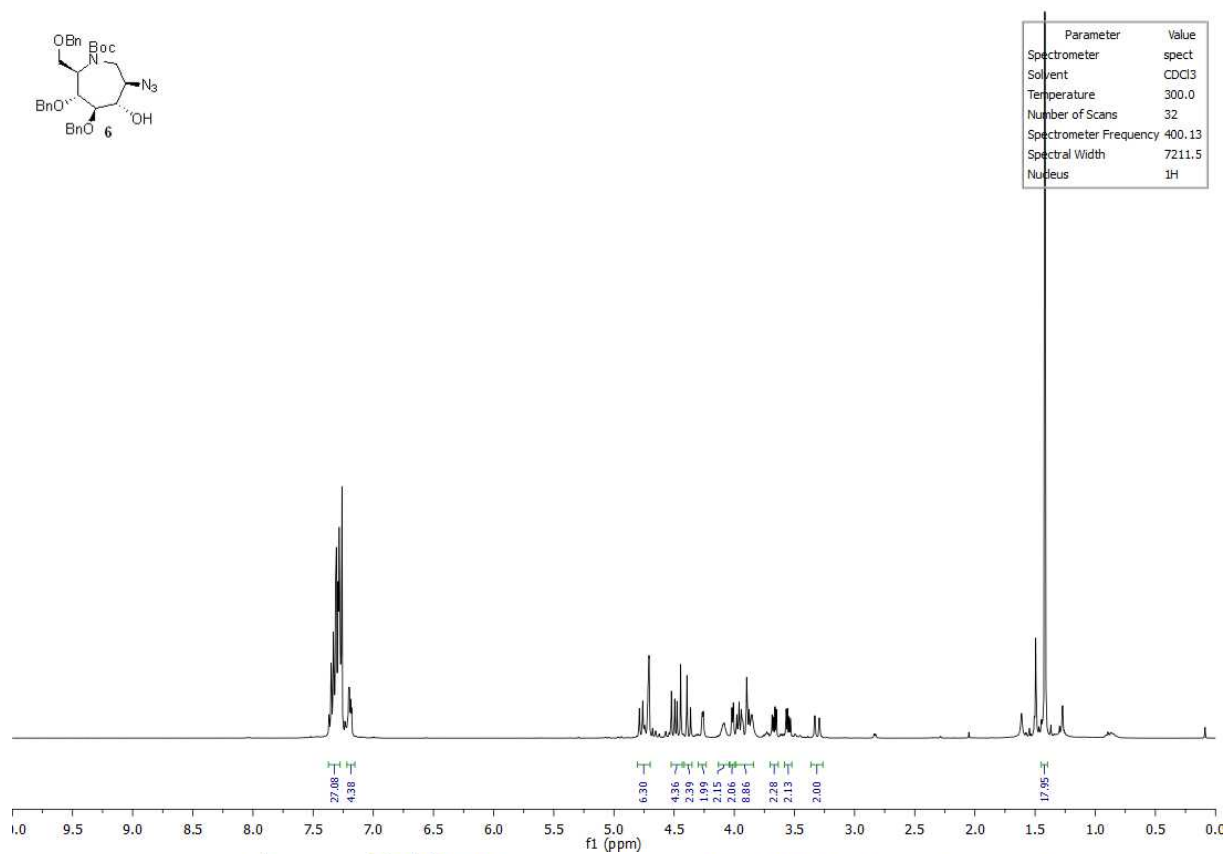
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Spectrometer Frequency	100.61
Spectral Width	29761.9
Nucleus	¹³ C

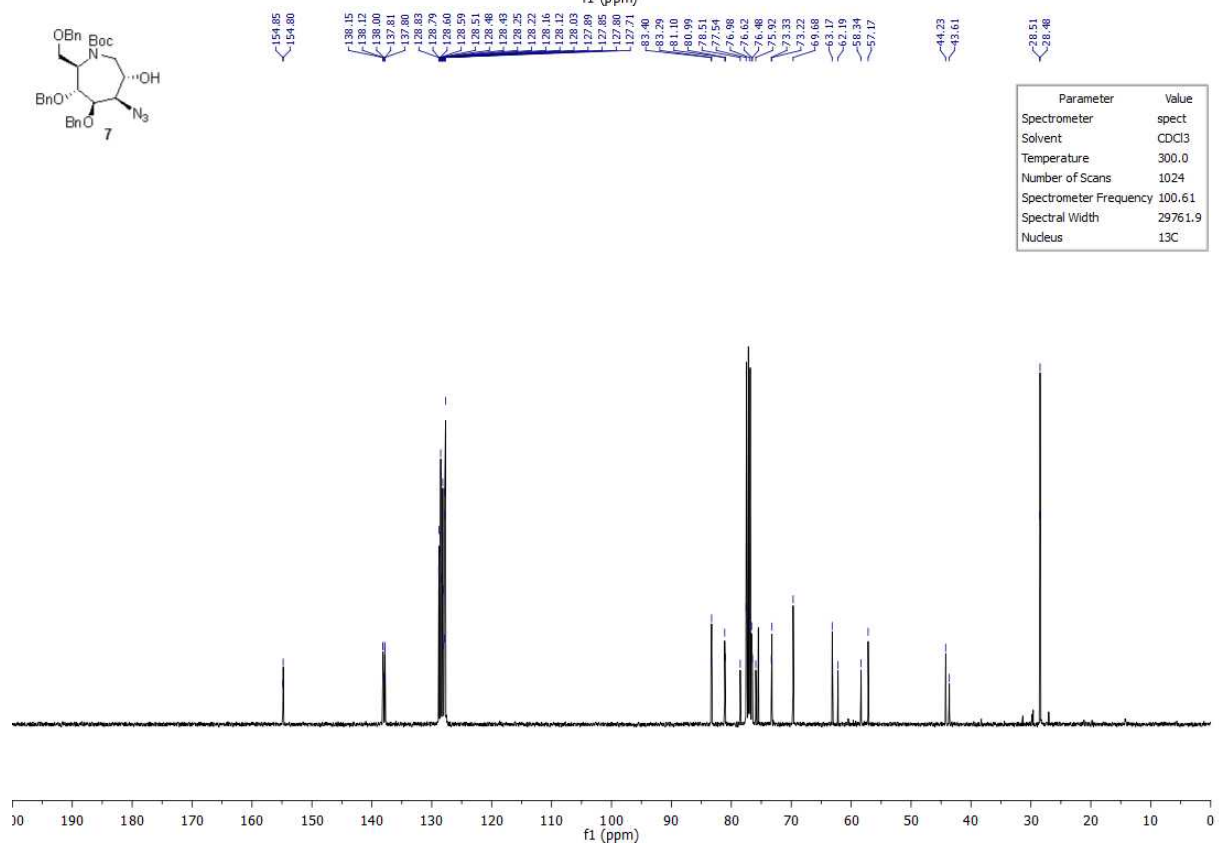
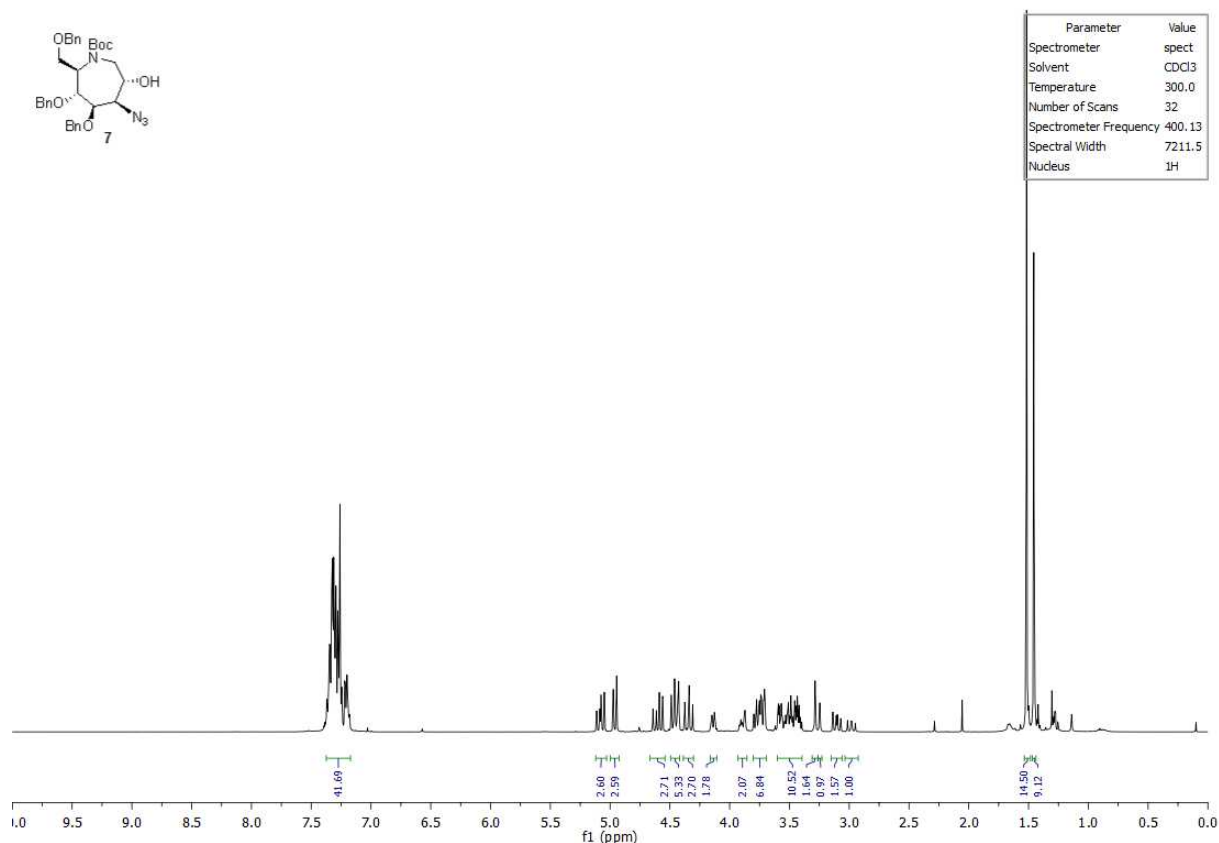


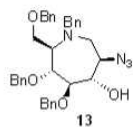




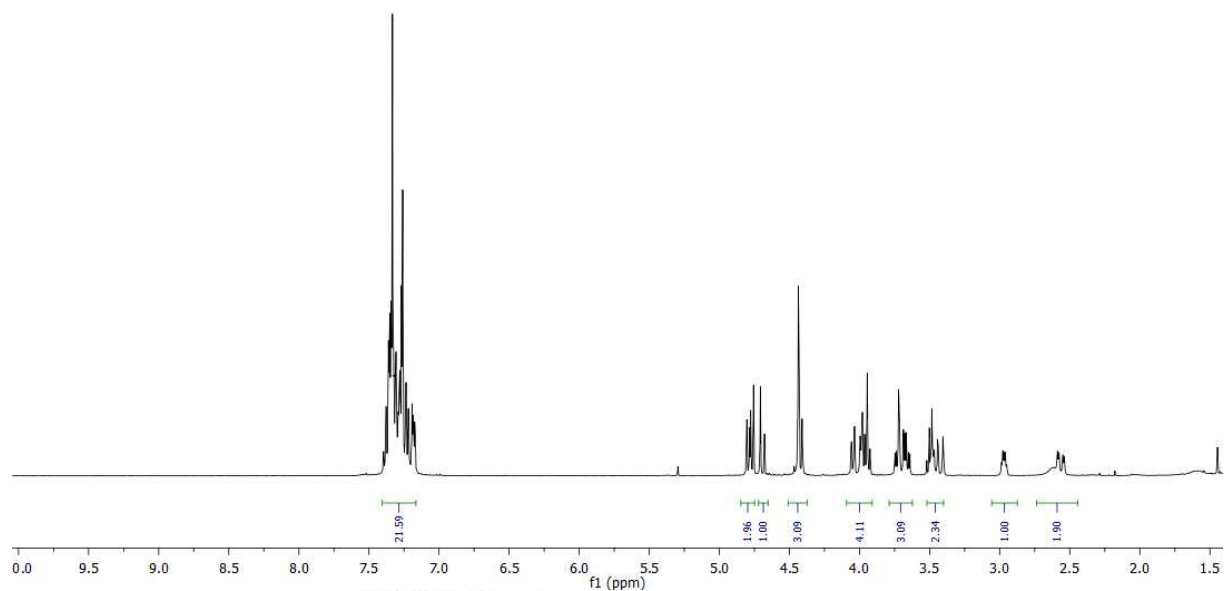




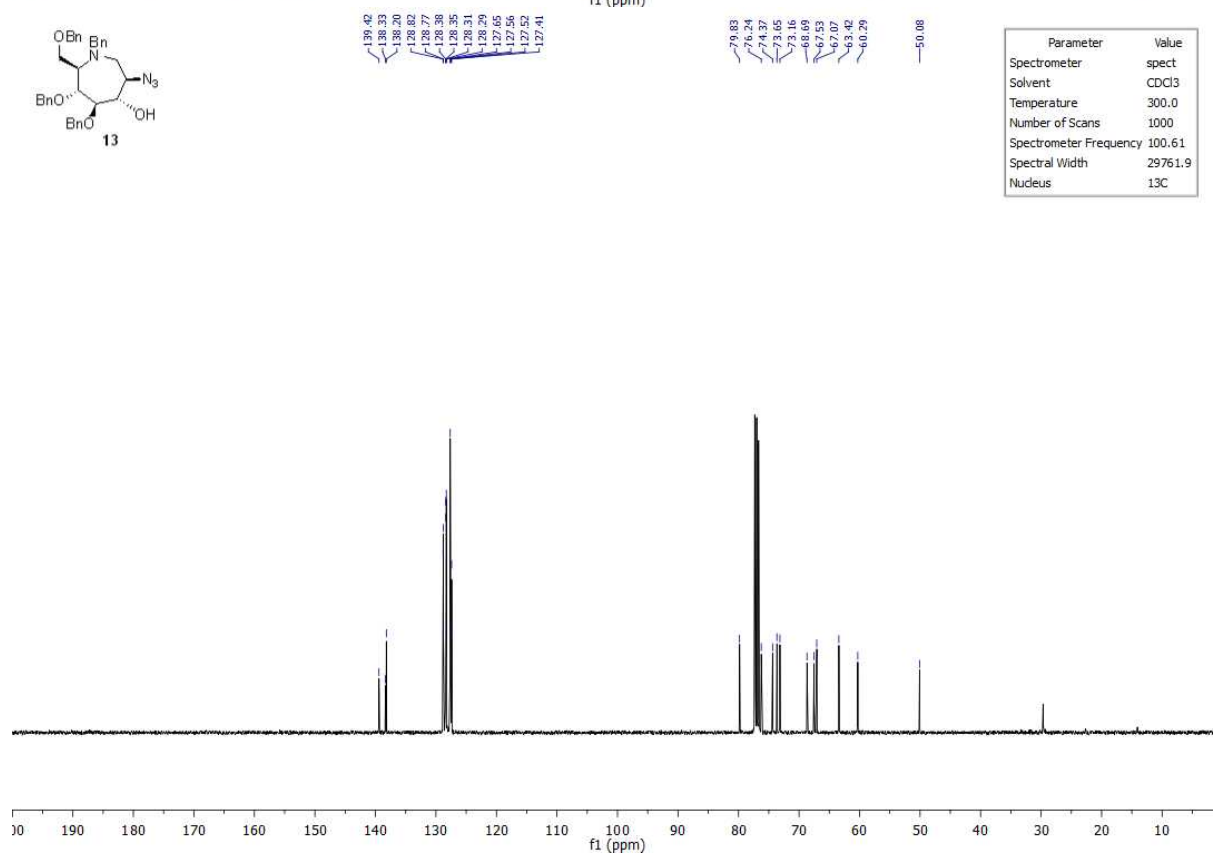


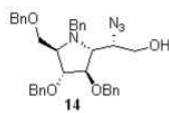


Parameter	Value
Spectrometer	spect
Solvent	CDCl ₃
Temperature	300.0
Number of Scans	8
Spectrometer Frequency	400.13
Spectral Width	7211.5
Nucleus	¹ H

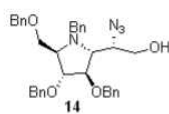
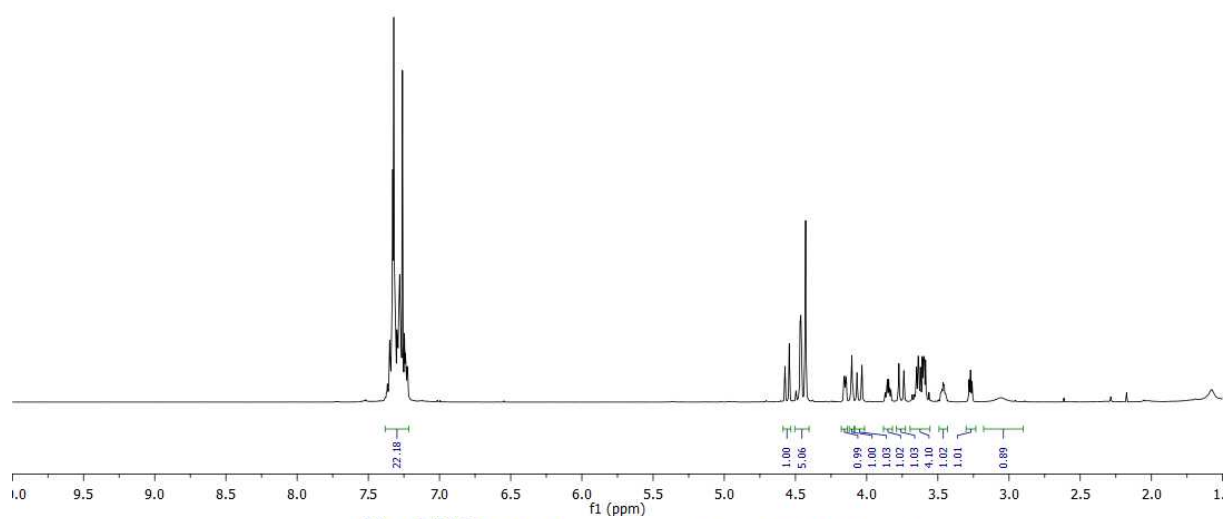


Parameter	Value
Spectrometer	spect
Solvent	CDCl ₃
Temperature	300.0
Number of Scans	1000
Spectrometer Frequency	100.61
Spectral Width	29761.9
Nucleus	¹³ C





Parameter	Value
Spectrometer	spect
Solvent	CDCl ₃
Temperature	300.0
Number of Scans	64
Spectrometer Frequency	400.13
Spectral Width	7211.5
Nucleus	¹ H



138.23
138.02
137.72
137.57
128.47
128.38
128.32
128.26
127.94
127.91
127.73
127.68
127.58
127.10

85.67
83.64

73.20
71.98
68.82
66.60
63.17
62.89
62.04

51.90

Parameter	Value
Spectrometer	spect
Solvent	CDCl ₃
Temperature	300.0
Number of Scans	2048
Spectrometer Frequency	100.61
Spectral Width	29761.9
Nucleus	¹³ C

