

# Variations in reproductive strategy of the silver Arowana, Osteoglossum bicirrhosum Cuvier, 1829 from four sub-basins of the Peruvian Amazon

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1	Variations in reproductive strategy of the silver Arowana,
2	Osteoglossum bicirrhosum Cuvier, 1829 from four sub-basins of the
3	Peruvian Amazon
4	
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24	Summary
25	
26	The aim of this study was to assess the variability of reproductive characteristics of
27	the silver Arowana, Osteoglossum bicirrhosum, in four sub-basins of the Peruvian
28	Amazon: the Amazonas, Ucayali, Napo and Putumayo rivers. An illustrated, detailed
29	gonadal maturation scale is proposed for a clearer determination of the reproductive
30	status of male and female Arowana. The breeding season occurred during the dry
31	season and early rising waters in the Amazonas and Ucayali, whereas it was
32	restricted to the dry season in the Napo and Putumayo. A same female can reproduce
33	more than once during the breeding season. Age at maturity was reached during the

34 second year, at sizes varying between 43-45cm for males and 46-50 cm for females. 35 Important variations in fecundity were observed between fish of the four sub-basins, 36 whereas mean oocytes diameter and mass did not vary significantly, suggesting 37 differences in reproductive investment. Together with previously reported variations 38 of growth patterns in the same sub-basins, these results suggest the existence of 39 distinct populations in the Peruvian Amazon, with important consequences for 40 management and conservation, but also for future aquaculture development 41 programs.

42

#### 43 Introduction

44

45 The silver Arowana, Osteoglossum bicirrhosum, is a relatively large (>1 m length), 46 laterally compressed fish of the archaic bony-tongued fish Order Osteoglossiforms 47 (Goulding, 1980), comprising, in South America, the blue Arowana, Osteoglossum 48 ferreirai Kanazawa 1966, and the species of the genus Arapaima spp., known as 49 pirarucu in Brazil and paiche in the other Amazonian countries. The silver Arowana 50 preferentially inhabits lentic waters of the Amazon, Rupununi and Oyapock rivers 51 (Goulding, 1980; Bayley and Petrere, 1989; Reis et al., 2003; Saint-Paul et al., 2000). 52 This species occupies a particular place in Amazonian fisheries as it is heavily 53 harvested both as an adult for human consumption and as a small juvenile (usually 54 still with its yolk sac) for the ornamental trade (Moreau and Coomes, 2006; 55 Alcantara et al., 2007). The Asian Arowanas (Sleropages spp.) or dragon fish, 56 closely resemble the Amazonian silver Arowana. These are very popular and sought 57 for pet fish in Asian markets, which has lead natural populations to be so 58 dramatically depleted that they have been listed under the Convention on

59	International Trade in Endangered Species (CITES) Appendix I in 1975 (Moreau and
60	Coomes, 2006). Since then, the Amazonian cousin, O. bicirrhosum, has replaced
61	them in Asian markets and its exportation has increased alarmingly since the 1990s
62	(Tello and Canepa, 1991; Moreau and Coomes, 2006, 2007). Owing to its increasing
63	exploitation in Amazonia for ornamental purposes, the species is considered
64	threatened in Peru (Moreau and Coomes, 2006), it has already been placed on the
65	Red Book of Colombia (Mojica et al., 2002) and its trade for ornamental purposes is
66	prohibited in Brazil (Lima and Prang, 2008; Prang, 2008).
67	The reproductive biology of the silver Arowana is relatively well described since the
68	early studies of Goulding (1980, 1990) and Aragao (1989). Recently, patterns of
69	reproductive investment were further investigated in the Mamirauá Reserve
70	(Queiroz, 2008) and the potential role of male aggregations was explored (Verba et
71	al., 2014). In Peru, its reproductive characteristics were studied in one lake of the
72	Putumayo River (Cortegano et al., 2014) and further information also exists from
73	unpublished dissertations (Ayala, 2001; Tang and Gomez, 2005). Despite extensive
74	ornamental fisheries in almost all the distribution area of the species and growing
75	concern about its conservation, however, no assessment of the geographic variability
76	of reproductive traits has yet been attempted. Besides the obvious interest for
77	conservation and fisheries management, the knowledge of whether geographic
78	population may display significant variations in specific reproductive traits such as
79	the timing of maturation and egg production would be determinant for future
80	aquaculture development. Indeed, the production of Arowana juveniles in controlled
81	conditions (aquaculture) appears to be the most immediate alternative to extraction
82	of natural populations for ornamental purposes (Argumedo, 2005; Moreau and
83	Coomes, 2006). Choosing breeders from a population with higher relative fecundity

(egg production per unit body mass), for example, might increase chances of success.
A recent study in the Peruvian Amazon demonstrated important variations in the
growth patterns of the silver Arowana in four sub-basins, suggesting the existence of
distinct populations, which might require independent management (Duponchelle et
al., 2012). The aim of the present study is to assess whether reproductive
characteristics also vary, and to which extent, between these four sub-basins of the
Peruvian Amazon.

91

#### 92 Material and Methods

93

#### 94 Fish collection

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96 Fish were collected monthly between November 2006 and April 2009 at the 97 Iquitos and Requena markets for the Amazonas, Ucayali and Napo sub-basins 98 (Figure 1). Requena is an important market located on the lower Ucavali River and 99 Iquitos is the main landing market of the Peruvian Amazon, concentrating fishes 100 from the Amazonas, Ucavali, Marañon, Napo, Huallaga, Tigre, Putumayo, Nanay, 101 Yavari and Morona sub-basins (Garcia-Vasquez et al., 2009). For a better accuracy 102 on specimen provenance, fish (of the largest possible size range) were bought from 103 specific providers from each river basin for the Amazonas, Napo and Ucavali rivers. 104 For the Putumayo River, fish were bought monthly, from March 2008 to April 2009, 105 from local fishermen in El Estrecho and Huapapa communities (Figure 1). The sub-106 basins were chosen on the basis of their hydrological cycle: the Amazonas and 107 Ucavali have very similar hydrological cycles; the Putumayo and Napo also have 108 similar hydrological cycles, but lag from about three months from that of the Amazonas-Ucayali. The availability and reliability of specimen's provenance was
also taken into account. Monthly means of water levels were provided by the SOHYBAM (Observation Service - Geodynamical, hydrological and biogeochemical
control of erosion/alteration and material transport in the Amazon basin) and
SENAMHI (Servicio Nacional de Meteoloría e Hidrología), Peru.

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#### 115 **Biological sample analysis**

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133

117 A total of 951 specimens of O. bicirrhosum were analysed. On each fish, total length 118  $(T_L)$ , standard length  $(S_L)$  and body mass were measured to the nearest mm and g, 119 respectively. Gonads were photographed and macroscopically checked for maturity 120 stage. On each fish, a gonad sample was preserved in Bouin's fluid for histological 121 analyses. Fixed gonads were further processed under classical histological methods 122 with Haematoxylin and Eosin staining for confirmation of macroscopic maturity 123 stages. A gonadal maturation scale was then established, based on the macroscopic 124 criteria detailed in Nuñez and Duponchelle (2009). Nevertheless, the proposed scale 125 also provides the equivalence with Brown-Petersen et al. (2011)'s terminology, 126 which was developed in an effort to standardize the confusing number of terms 127 referring to key phases of the reproductive cycle in fisheries science' literature. 128 The breeding season was estimated from the monthly proportions of females' 129 gonadal maturation stage "spawning capable" (Brown-Petersen et al., 2011). To 130 obtain a better representation of the annual cycle, data collected over the study period 131 were pooled for each river basin. 132 The average size at first sexual maturity  $(L_m)$  is defined as the standard length

at which 50% of the individuals are at an advanced stage of the first sexual cycle

during the breeding season (Legendre and Ecoutin, 1989). Practically, this is the size at which 50% of the fish have reached stage 2 of the maturity scale for females and males (Nuñez and Duponchelle, 2009). The mean SL at first maturity ( $L_m$ ) was estimated by fitting the fraction of mature individuals per 50 mm S<sub>L</sub> intervals to a logistic regression function weighted by the total number of individuals in each size class (Barbieri et al., 1994; Duponchelle and Panfili, 1998):

140 
$$\%M = \frac{100}{1 + e^{(-a \times (L - Lm))}} \quad (1)$$

141 where %M = percentage of mature individual by size class, L = central value of 142 each size class, and a and L<sub>m</sub> = constants of the model.

143 The age at first sexual maturity (A<sub>m</sub>) was calculated from the von Bertalanffy growth
144 function (VBGF) as follows:

145 
$$A_{m} = (-\ln(1 - (L_{m}/L_{\infty})) / K) + t_{0}$$
(2)

where  $L_m$  is the size at first sexual maturity previously calculated and t,  $L_{\infty}$ , K and t<sub>0</sub> the parameters of the VBGF obtained from Duponchelle et al. (2012) on the same samples.

149 Batch fecundity (number of eggs to be potentially laid per spawning event, 150 resulting from the maturation of fully developed intra-follicular oocytes) and oocyte 151 diameter were estimated by image analysis (using NIH Image J software) from 152 whole gonads in final maturation stage (stage 3). As the oocytes were not exactly 153 circular in shape, the mean arithmetic diameter was used:  $\sqrt{d.D}$ , where d is the 154 minimum diameter and D the maximum diameter of each oocyte. Oocyte mean mass 155 was calculated from each stage 3 gonads, by weighing 30 oocytes to the nearest 156 0.001 g.

157

#### 158 Statistical analyses

160	Differences in mean standard length and mass between sexes within each sub-basin
161	were tested with a t-test. Differences of mean standard length or mass between sub-
162	basins were tested with Kruskal-Wallis One Way Anova on ranks and Dunn's all
163	pairwise post-hoc tests.
164	For each sub-basin, logistic regression models provided an estimate of the $L_{\rm m}\pm$
165	standard deviation (equation 1). Therefore, sizes at maturity between sexes or
166	geographic locations were compared using a classical Student t-test (degrees of
167	freedom calculated from the number of size classes).
168	Analysis of covariance (ANCOVA) was used to test for differences in the fecundity-
169	body mass relationships between geographic populations, using fecundity as the
170	dependent variable, body mass as the covariate, and geographic population as
171	categorical variable.
172	Comparisons of mean oocyte diameter and mass between geographic populations
173	were carried out using one-way ANOVA.
174	For all comparisons, when more than two pairwise-tests were performed, the
175	Bonferroni correction was applied to keep the experiment-wide error rate at the
176	desired $\alpha$ (0.05, 0.01 or 0.001).
177	
178	Results

179

180 A total of 290 specimens of *O. bicirrhosum* were analysed for the Amazonas, 276 for

181 the Ucayali, 145 for the Napo and 240 for the Putumayo (Figure 2a). Mean standard

- 182 lengths  $(S_L)$  and masses did not differ among sexes in any population. There were,
- 183 however, significant differences in mean length (Kruskal-Wallis One Way ANOVA

184	on Ranks, $H = 83.4$ , P<0.001) and mass ( $H = 79.3$ , P<0.001) between populations
185	when females and males were pooled: the Ucayali population differed from all the
186	others in both mean length (Q = 8.6, P<0.05 with the Putumayo, Q = 6.7, P<0.05
187	with the Amazonas and Q = 4.8, P<0.05 with the Napo) and mass (Q = 8.3, P<0.05
188	with the Putumayo, $Q = 6.8$ , P<0.05 with the Amazonas and $Q = 4.5$ , P<0.05 with
189	the Napo), whereas the other populations did not differ between each other in mean
190	length nor in mean mass. Maximum lengths and even more masses, however, did
191	vary considerably between populations, reaching barely 80 cm and about 3.8 kg in
192	the Putumayo and more than 90 cm and 8 kg in the Amazonas (Figure 2a). In the
193	Amazonas, maximum observed body mass for the silver Arowana was 49, 75 and
194	113% heavier than in the Napo, Ucayali and Putumayo, respectively. These
195	important differences in body mass, however, are not a mere reflection of the larger
196	maximum lengths in the Amazonas. Indeed, although mass-at-length is relatively
197	similar between populations until about 50 cm (~ size at first sexual maturity, see
198	below), afterwards fish start to become progressively heavier at a given length in the
199	Amazonas than in the Ucayali, Napo and Putumayo, in order of increasing difference
200	(Fig. 2b). These differences reached $\sim 500$ g and $\sim 1000$ g at 70 and 80 cm,
201	respectively, between the Amazonas and Putumayo sub-basins (Fig. 2b).
202	For comparison purposes with other studies, a relationship between standard length
203	(SL) and total length (TL) is provided, calculated from all individuals in Figure 2a,
204	giving: $SL = 0.9345 TL + 0.102$ .
205	
206	Gonadal maturity scale
207	

Females and males silver Arowana only have one functional gonad, the left one. The right gonad is atrophied. An illustrated, detailed gonadal maturity scale is presented in Figure 3 for both females and males. It allows for a clear determination of the breeding season, size / age at maturity and fecundity.

212 The precise description of the gonadal cycle also allowed identifying that the silver

213 Arowana is a multiple-spawner (sensu Nuñez and Duponchelle 2009). Indeed, the

214 presence of small, vetelogenic oocytes in between large oocytes in "spawning

capable" ovaries, both in stage 3 or 4 (Figure 4a) and in stage 5-2 (recently spawned)

216 ovary initiating another vitellogenic cycle (Figure 4b), observed in all four sub-

217 basins, proves that a same female can reproduce at least twice during a breeding

season. Further evidences come from the size frequency distribution of oocytes in

219 "spawning capable" ovaries from the Putumayo sub-basin, showing two clearly

separated modes of vitellogenic oocytes: the largest (of 244 oocytes) that will be

spawned shortly and a smaller one (of 221 oocytes) that will likely be released in

another spawning bout during the same breeding season (Fig. 5).

223

#### 224 Breeding season

225

There exists a relationship, albeit not a direct one, between the reproductive cycle of the silver Arowana (estimated from the monthly proportions of females in "spawning capable" phase of the gonadal maturation scale) and the hydrological cycle (Figure 6). In the Amazonas and Ucayali rivers, which have comparable hydrological cycles, females have similar breeding seasons. Although sampling could not be completed at some months, it can nevertheless be observed that the breeding season started during the dry season (August-September) and ended before the water level reaches its

233	maximum in both rivers. Some residual reproductive activity could be observed in
234	the Ucayali at almost every month outside the main breeding season. Again, in the
235	Napo and Putumayo rivers, which have comparable hydrological periodicity, the
236	breeding season of the silver Arowana also tended to be similar (Figure 6). Although
237	sampled could not be obtained in December and January in the Putumayo, it is likely
238	that, as in the Napo, the breeding season initiated in December to peak in February-
239	March. The breeding season in the Napo and Putumayo, however, did not last as
240	long as in the Amazonas and Ucayali and was restricted to the low water period.
241	
242	Size and age at sexual maturity
243	
244	Females tended to have slightly larger size at first sexual maturity (Figure 7) than
245	males in every sub-basin, but these differences were not significant (t-test, t=0.292,
246	df=24, p=0.773 for the Amazonas; t=0.226, df=24, p=0.823 for the Ucayali; t=0.630,
247	df=18, p=0.536 for the Napo; and t=0.327, df=24, p=0.746 for the Putumayo).
248	The size at maturity did not differ significantly between sub-basins for females nor
249	males.
250	Age at first sexual maturity was reached during the second year for both males and
251	females in all four sub-basins (Figure 7).
252	In all four sub-basins, the size at which 100% of both males and females were
253	sexually mature was slightly before 60 cm $S_L$ (Figure 7).
254	
255	Fecundity
256	

The number of oocytes per spawning event (batch fecundity) significantly varied (ANCOVA,  $D_{3,41}$ = 6.610, P<0.001) between fish of the four sub-basins (Figure 8, Table 1). Batch fecundity ranged from 93 to 256 oocytes for females of 1300 and 4960 g, respectively in the Amazonas, from 94 to 311 oocytes for females of 2080 and 3480 g, respectively in the Ucayali, from 92 to 197 oocytes for females of 1460 and 3220 g, respectively in the Napo and from 125 to 360 oocytes for females of 1770 and 2980 g, respectively in the Putumayo.

264 The Putumayo population differed significantly from the Amazonas (ANCOVA,

265  $D_{1,32}=25.9$ , P<0.0001), Napo ( $D_{1,30}=9.7$ , P<0.05 with Napo) and Ucayali ( $D_{1,33}=$ 

266 53.9, P<0.001) populations, whereas the Amazonas, Ucayali and Napo populations

did not significantly differ between each other ( $D_{2,12}$ = 1.4, P>0.05).

268 Relative fecundity (batch fecundity per unit body mass), oocyte diameter and oocyte

269 mass are presented in Table 2. Relative fecundity differed significantly between fish

of the four sub-basins (One-way ANOVA, F3 = 11.6, P < 0.001). Holm-Sidak post-

271 hoc test indicated that fish from the Putumayo significantly differed from the

272 Amazonas (t = 4.812, P < 0.001), the Ucayali (t = 3.614, P = .0.004) and the Napo (t

273 = 2.732, P = 0.037), whereas fish from the Amazonas, Ucayali and Napo did not

274 differ significantly between each other.

There was no significant difference in mean oocytes diameter or mass between fishof the four sub-basins (Table 2).

277

#### 278 Discussion

279

280 Important length and even more important body mass differences were observed

between the four sub-basins, as already pointed out while studying age and growth

282 patterns of these same populations (Duponchelle et al., 2012). After reaching sexual 283 maturity, fishes in the Amazonas become progressively heavier than those from the 284 Ucayali, Napo and Putumayo, in order of increasing difference. Beside inter-basin 285 differences, silver Arowana from the Peruvian Amazon grow relatively heavier than 286 fish from the Mamirauá Reserve in Brazil. An individual of 80 cm  $T_L$  (~ 75 cm  $S_L$ ) 287 weighs ~ 3000 g in the Mamirauá Reserve (Cavalcante, 2008), whereas it weighs in 288 average 3161 g in the Putumayo, 3369 g in the Napo, 3552 g in the Ucayali and 3702 289 g in the Amazonas. The better body condition of Arowana in the Peruvian Amazon 290 might result from better trophic conditions than in the Mamirauá Reserve, which 291 would have to be tested.

- 292
- 293 Gonadal maturity scale
- 294

The description of the different maturity stages were consistent with Aragao' (1989) descriptions, for both females and males, although the number of stages was reduced to fit Nuñez and Duponchelle' (2009) scale. This illustrated gonadal maturity scale provides a clear reference for comparative works on the reproductive biology of this species.

300 Previous studies had suggested, from the observation of different sizes of oocytes in

301 ripe gonads (but without providing evidence) that a same female could reproduce

302 more than once during the breeding season (Ayala, 2001; Tang and Gomez, 2005). In

303 the present study, the presence of small, vitellogenic oocytes in all stages of

- 304 "spawning capable" ovaries and the size frequency distribution of oocyte in
- 305 "spawning capable" ovaries provide strong evidence that a same individual female
- 306 can reproduce more than once during the breeding season in the Peruvian Amazon.

This result, however, differs from other studies in the Brazilian Amazon, Aragao
(1989) in Lake Janavaca and more recently Queiroz (2008) in the Mamirauá
Reserve, who reported that this species probably spawns only once during the
breeding season, but without providing demonstrating evidence.

311

#### 312 Breeding season

313

314 In Central Amazonia, Brasil, where most published studies about the silver 315 Arowana's breeding cycle were carried out, the species was reported to reproduce 316 during the dry season and early rising waters (Goulding, 1980; Aragao, 1989; 317 Queiroz, 2008; Cavalcante, 2008). This is consistent with the observations from the 318 present study in the Amazonas and Ucayali sub-basins, or with previous observations 319 in Peru, in the Pacaya Samiria Reserve (Tang and Gomez, 2005) and in the Ucayali 320 River (Ayala, 2001). This pattern, however, does not seem to hold for all silver 321 Arowana of the Peruvian Amazon, as the breeding season seemed restricted to the 322 dry season in the Napo and Putumayo sub-basins. This shorter breeding season might 323 be an adaptation to the much shorter flooding season in the Napo and Putumayo 324 (Figure 5). Once females have spawned, males guard the eggs and then the fry in 325 their mouth for five to seven weeks (Goulding, 1990; Argumedo, 2005; Queiroz, 326 2008), after which free swimming juveniles of over 6 cm are released from parental 327 care and have to fend for themselves. It is well known that in Amazonian 328 environments, the high water season with its extended floodplains and inundated 329 vegetation, providing food and shelter from predators is particularly advantageous 330 for the development and survival of fish's young stages (Lowe-McConnell, 1964, 331 1987; Goulding, 1980). In order for males to release their free-swimming young in

332 time to make the most of these advantageous environmental conditions - which 333 barely last three months in the Napo and Putumayo sub-basins (Figure 5) -, while 334 allowing for the long incubation, the spawning period might have to be finished 335 when the water starts to rise. Nevertheless, size frequency distribution of oocytes in 336 ripe gonads (Figure 4c) and the presence of small vitellogenic oocytes in all stages of 337 "spawning capable" ovaries support the idea that individual females may have time 338 to produce a second brood during the spawning period, despite its shorter duration in 339 the Putumayo. This suggests that the time needed for the small vitellogenic oocytes 340 (Fig. 4a-b) to complete their growth is relatively short (a few weeks). These results 341 also imply polyandry in the species: indeed, the short breeding season in the 342 Putumayo and Napo and the long paternal mouthbrooding period do not allow for a 343 same male to carry successively two broods of a same female. On the other hand, in 344 the Amazonas and Ucayali sub-basins, where the high water season lasts nearly 345 twice as much (December to May), females that spawn when the water has already 346 started to rise may still ensure their progeny will find favourable environmental 347 conditions long enough to survive, hence ensure reproductive success. This 348 hypothesis could be tested by additional studies on these two specific sub-basins and 349 others with similarly short high water seasons. As Arowana do not live in rivers, but 350 instead in adjacent lakes and lagoons, testing this hypothesis would also require 351 having water level data in specific lakes where Arowana concentrations are high. 352 353 Age and size at maturity

354

355 In the Mamirauá Reserve, the size at first sexual maturity was about 52 cm  $T_L$  (~ 49 356 cm  $S_L$ ) for both sexes and was reached during the second year (Cavalcante, 2008).

357	These values were relatively similar to those observed in the Peruvian Amazon,
358	where silver Arowana also reached maturity during their second year at sizes varying
359	between 43-45 cm for males and between 46-50 cm for females. These values were
360	relatively smaller than the $\sim 60$ cm $T_L$ ( $\sim 56$ cm $S_L)$ reported in the El Dorado lake in
361	the Pacaya Samiria Reserve, Peru (Tang and Gomez, 2005). In a preliminary study in
362	the Ucayali River, Peru, Ayala (2001) reported sizes at first maturity of 65.9 and 73.9
363	cm $T_L~(\sim 62~and~69~cm~S_L)$ for females and males, respectively. These obviously
364	over-estimated values, however, were likely due to the fact that the author
365	erroneously considered individuals in vitellogenesis (stage II of his gonadal
366	maturation scale) as still immature. Cortegano et al. (2014) reported sizes at maturity
367	of $\sim$ 58 and 62 cm for females and males, respectively in lake Grande, close to
368	Huapapa in the Putumayo. These large differences with our results from the
369	Putumayo (47 and 43 cm for females and males, respectively) are particularly
370	surprising, as about a third of our own sampling also came from lake Grande. Most
371	of the observed differences might result from the interpretation of what is considered
372	as "sexual maturity" in Cortegano et al.' study, which was not properly defined. It is
373	likely, when looking at their logistic curves that reach 100% only close to the
374	maximum observed sizes, that they considered as "sexually mature" female at the
375	end of stage 2 rather than at the beginning of stage 2 (Fig. 3a). Likewise, for males,
376	they likely considered males in late stage 2 or stage 3 rather than in stage 2 (Fig. 3b).
377 378	It is worth noting that while the other reproductive traits (breeding season and
379	fecundity) largely varied between sub-basins, size at maturity was relatively
380	conserved and that the slight differences observed were not significant. This is
381	particularly surprising as size at maturity is a plastic trait that readily responds to
382	environmental variation (Stearns and Crandall, 1984; Stearns and Koella, 1986;

383 Duponchelle and Panfili, 1998; Duponchelle et al., 2007) or fishing pressure (Olsen
384 et al., 2004). This observation will have to be confirmed on other geographic
385 populations.

386 It is important to keep in mind that the size at first sexual maturity  $(L_m)$  is the size at 387 which 50% of the individuals have reached maturity during the breeding season, 388 which means the other 50% have not. Considering the size at first sexual maturity as 389 the legal minimum size at capture, as is customary in Amazonian fisheries 390 regulations, we implicitly allow that half the fish be caught as immature, hence 391 before they could reproduce (growth-overfishing). In order to ensure a healthy 392 spawning stock, however, fisheries should ensure that all captured individuals have 393 had a chance to reproduce at least once (Myers and Mertz, 1998; Froese, 2004). This 394 goal can only be attained by fixing the minimum size at capture not at the size at 395 which 50% of the individuals in a population are mature  $(L_m)$ , but at the size at 396 which 100% are mature. The size at which 100% of the fish are mature, considering 397 all four sub-basins of the Peruvian Amazon, was slightly below 60 cm S<sub>1</sub> (Fig. 6). 398 Therefore, considering the approximately 5 cm difference between total and standard 399 lengths, we recommend that the minimum size at capture should be fixed at 60 cm 400 T<sub>L</sub> in the Peruvian Amazon.

401

#### 402 Fecundity

403

The silver Arowana reputedly produces very few eggs for its size (Goulding, 1980;
Aragao, 1989; Queiroz, 2008). Fecundity variations between geographical samples,
however, have never been reported so far. In the present study, important differences
in batch fecundity were observed between the silver Arowana of the four sub-basins.

408 The Putumayo population significantly differed from the other three populations, 409 which in turn did not differed between each other's. The absence of significant 410 difference between the Amazonas, Napo and Ucayali populations, however, was 411 likely due to the low number of gonads analysed and to the large variance observed 412 in the Ucayali population (Fig. 7). Such a large variance could result from the mixing 413 of individuals from distinct populations within the Ucayali sub-basin. Indeed, fish 414 were sampled from fishermen operating in large portions of the Ucayali basin and 415 which could hold populations with distinct reproductive characteristics. This 416 indicates that further studies on this species will need to be carried out with on-site 417 sampling rather than market sampling. Nevertheless, as an illustration of the 418 important fecundity variation between fish of the four sub-basins, despite the absence 419 of statistical significance, the batch fecundity at two representative female body 420 masses (1500 and 3500 g) was calculated for each sub-basin from the regressions 421 among batch fecundity and body mass (Table 1). The differences of batch fecundity 422 between populations ranged from 33 to 104 % for females weighing 1500 and 3500 423 g, respectively. Given the energetic cost of gamete production, food is considered 424 one of the most important environmental factors involved in the regulation of 425 fecundity (Wooton, 1979; Wooton, 1998). One would thus expect the fish with the 426 better body condition (in decreasing order those of the Amazonas, then Ucavali, 427 Napo and Putumayo, see Fig. 2b), which usually reflects better trophic conditions, to 428 also display the highest relative fecundity. Yet, relative fecundity was inversely 429 proportional to body condition (Table 2), suggesting contrasted energy allocation 430 (towards body growth vs reproduction) strategies between fish of the four sub-basins. 431 Additionally, these large variations in fecundity were not compensated by variations 432 of oocyte size or mass, suggesting differences in reproductive effort between fish of

the four the sub-basins. These hypotheses, however, will need to be further validated
with an increased number of ripe gonads analysed in the Amazonas, Napo and
Ucayali sub-basins.

436 As a comparison, in the Mamirauá Reserve females of 65 cm  $T_L$  (~ 61 cm  $S_L$  and 437 1500 g) and 75 cm  $T_L$  (~ 70 cm  $S_L$  and 2500 g) produce ~ 200 and 230 oocytes, 438 respectively (Queiroz, 2008, using the length-mass relationship provided in 439 Cavalcante, 2008). This would be approximately twice as much as the fecundity of a 440 1500 g female in the Peruvian Amazon and in the range of the Putumayo fecundity 441 for a 2500 g female (Table 1), suggesting a higher relative fecundity in small females 442 progressively decreasing with length in the Mamirauá Reserve. Again, fish from the 443 Mamirauá, which have lower body condition than fish from the Peruvian Amazon, 444 also have higher relative fecundity, contrary to expectations. A surprising 445 information comes from the mean diameter (9.75 mm) and mass (1.8 g) of oocytes in 446 the Mamirauá Reserve, which are 25% smaller but a 100% heavier than the largest 447 and heaviest oocytes in the Peruvian Amazon (12.2 mm and 0.98 g in the Putumayo), resulting in a density over 3.5 g/cm<sup>3</sup>, (vs 1.03 g/cm<sup>3</sup> in the Putumayo), which appears 448 449 much too dense for an egg. 450 In the Pacaya-Samiria Reserve, Peru, Tang and Gomez (2005) observed that females 451 of 1500 and 2500 g would produce in average 215 and 266 oocytes, respectively, 452 which is above the mean values observed in the present study. A recent study in lake 453 Grande on the Putumayo River, reported batch fecundities ranging from 115 oocytes 454 for a female weighing 1100 g to ~240 oocytes for a female of ~3000 g (Cartegano et 455 al., 2014), within the range of the values observed for the Putumayo in the present 456 study.

The only other available information about the silver Arowana's fecundity comes
from Aragao (1989) in the Solimões River basin (lake Janauacá), but the range of
female body masses he worked with (551 – 1750 g, with only 2 females above 1500
g) barely overlap with ours, hampering a direct comparison. Nevertheless, a 1500 g
female would produce approximately 182 oocytes, which is also above the mean
values observed in the present study.

463

464 Conclusions

465 The important variability observed in reproductive characteristics and body condition 466 of the silver Arowana from the four sub-basins is consistent with the important 467 growth variations previously reported in these same localities (Duponchelle et al., 468 2012). This variability might result either from the phenotypic plasticity of the 469 species due to environmental heterogeneity between the sub-basins or from the 470 existence of several distinct genetic populations within Peru, that may need to be 471 further characterized and managed independently. The important differences in 472 breeding season, condition, growth and fecundity, however, strongly suggest that fish 473 from the different sub-basins are distinct populations. The different breeding season 474 and sedentary nature of the species should result in reduced gene flow between sub-475 basins, hence inducing genetic structuring. Although microsatellites markers are 476 available for the silver Arowana (Silva et al., 2009), they have only been used to test 477 for multiple paternity in the species (Verba et al., 2014) and no population genetic 478 study has been carried out so far. Yet, as previously pointed out (Duponchelle et al., 479 2012), the growing concerns about O. bicirrhosum conservation (Moreau and 480 Coomes, 2006, 2007) emphasize the need for comparative life history and genetics 481 studies at the inter- and intra-basin levels (with sampling carried out on-site) in order

482 to understand the population structuring of this species and to define appropriate483 management strategies.

484

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- 494

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620			
621			
622	Figure legends		
623			
624	Fig. 1. Map of the collection sites of Osteoglossum bicirrhosum in the Peruvian		
625	Amazon (black circles). A total of 711 specimens were collected between November		
626	2006 and April 2009 at the Iquitos and Requena markets for the Amazonas, Ucayali		
627	and Napo sub-basins. For the Putumayo, 240 fish were bought between March 2008		
628	and April 2009 from local fishermen at El Estrecho and Huapapa communities.		
629			
630	Fig. 2. Relationships between standard length (SL) and total body mass (M) of		
631	Osteoglossum bicirrhosum caught in four sub-basins of the Peruvian Amazon		

between 2006 and 2009 (a). Modelled regressions (M as a function of SL) between

633 60 and 90 cm SL, to illustrate inter-basin differences (b).

- 634
- Fig. 3. Macroscopic gonadal maturity scale for females (a) and males (b)
- 636 Osteoglossum bicirrhosum, following Nuñez and Duponchelle (2009). Equivalence
- 637 with Brown-Petersen et al. (2011)'s terminology is provided.
- 638
- 639 Fig. 4. Small vitellogenic oocytes (svo) in "spawning capable" ovaries of female
- 640 Osteoglossum bicirrhosum: (a) in between large vitellogenic oocytes (lvo) in a ripe -
- 641 stage 4 gonad and (b) in a post-spawning gonad reinitiating another vitellogenic
- 642 cycle stage 5-2.
- 643
- 644 Fig. 5. Size frequency distribution of oocytes in a "spawning capable" ovary of
- 645 *Osteoglossum bicirrhosum* from the Putumayo River showing two clearly separated
  646 modes of vitellogenic oocytes.
- 647
- Fig. 6. Proportions of breeding *Osteoglossum bicirrhosum* females in relation to themean water level in four sub-basins of the Peruvian Amazon.
- 650
- Fig. 7. Age (Am) and size (Lm) at first sexual maturity of both males (black circles
- and solid lines) and females (white triangles and dashed lines) Osteoglossum

653 *bicirrhosum* in four sub-basins of the Peruvian Amazon.

- 654
- Fig. 8. Linear regressions between batch fecundity and total female body mass of
- 656 Osteoglossum bicirrhosum in the Amazonas (N = 5, F = 0.0307\*M + 60.922, R<sup>2</sup> =

657	0.875, P<0.001), Napo (N = 3, F = 0.0578*M + 18.547, R <sup>2</sup> = 0.907, P<0.001),
658	Ucayali (N = 6, F = 0.0708*M – 9.1942, R <sup>2</sup> = 0.537, P<0.001) and Putumayo (N=28,
659	$F = 0.1055*M - 29.253$ , $R^2 = 0.714$ , P<0.001) rivers, Peruvian Amazon.
660	
661	Table 1. Comparison of Osteoglossum bicirrhosum's batch fecundity at a given
662	female body mass in four sub-basins of the Peruvian Amazon (calculated from the
663	regressions between batch fecundity and total body mass presented in Figure 8).
664	
665	Table 2. Mean relative fecundity (oocytes per kg total body mass), oocytes diameter
666	(mm) and oocytes mass (g) $\pm$ SD of <i>Osteoglossum bicirrhosum</i> in four sub-basins of
667	the Peruvian Amazon.
668	



Fig. 1



Fig. 2

Description	Illustration (females)	with Brown- Petersen et al. (2011)'s terminology
Stage 1: immature individuals Ovaries small and firm, opaque and whitish or slightly pinkish. No oocyte can be seen to the naked eye. A typical longitudinal canyon-type depression is clearly visible at the surface of the ovary.		Immature
Stage 2: maturing individuals Ovaries are larger with more important vascularisation and clearly visible oocytes. Oocytes are whitish and sparse at the beginning of stage 2, and the canyon-like depression is still visible. Progressively, the oocytes turn yellow and the canyon disappears. At the end of stage 2, the oocytes are orange and fill the ovary, but are not of a homogeneous size: all intermediaries between small whitish oocytes and large orange ones are visible. This last point is a clear distinction with stage 3.		Developing
<b>Stage 3: individuals in final maturation</b> The ovary now occupies about <sup>3</sup> / <sub>4</sub> of the abdominal cavity. Most oocytes are large, circular, orange, and homogeneous in size. Small oocytes are still visible in between the larger ones, but there is now a very clear size difference between the large and small ones.		
<b>Stage 4: individuals ready to spawn / ripe</b> The ovary occupies almost the whole abdominal cavity and appears even fuller. The ovary's membrane is very thin and transparent. A slight pressure on the abdomen can expulse the ovules, that are now free (out of their follicules) in the ovary.		Spawning capable
Stage 5-2: recently spawned individuals initiating another cycle The appearance is close to that of a stage 2 gonad, but the ovary is still relatively flaccid, larger and more vascularised. Developing oocytes of all colours are filling the ovary and some remaining large atretic oocytes can sometimes be observed. This stage is characteristic of females that have just spawn and that are initiating a new reproductive cycle.		

Stage 5: individuals that have just spawned The ovary is of similar size to a stage 4 ovary, but with an empty, flaccid appearance and well developed vascularisation. Some un-spawned, large oocytes can still be seen together with others already in re-absorption process (atresia) and early vitellogenic ones.	Chi Loug	Regressing
<b>Stage 5-1: resting individuals</b> The ovary looks like a stage 1 ovary (with no visible oocyte), but is larger, with a thicker membrane, a more important vascularisation and a generally redder colour than a stage 1. This stage is characteristic of adult females resting in between two breeding seasons.		Regenerating



Description	Illustration (males)	Equivalence with Brown-Petersen et al. (2011)'s terminology
<b>Stage 1: immature individuals:</b> The testis is a pinkish, translucent curved tube, thinner and longer than stage 1 ovary. It is often bordered by a golden colour towards the genital pore.		Immature
<b>Stage 2: maturing or resting individuals</b> : Testis wider, thicker, of a more opaque pinkish to reddish colour. The golden colour towards the genital pore is often still visible. Resting adults are usually at this stage.		Developing / Regenerating
<b>Stage 3: ripe individuals</b> : Testis even larger, wider and really thick, with dark red opaque appearance. The golden colour is usually not visible at this stage.		Spawning capable
<b>Stage 4: individuals after ejaculation</b> : Testis of a similar size to stage 3, but with a flaccid, more translucent appearance.		Regressing



Fig. 4







Fig. 6



Fig. 7





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Famala hady mass (a)	Batch fecundity			
Female body mass (g)	Amazonas	Napo	Ucayali	Putumayo
1500	107	105	97	129
2500	138	163	168	234
3500	168	221	239	340

Table 2.

	Amazonas	Napo	Ucayali	Putumayo
	(N=5)	(N=3)	(N=6)	(N=28)
Relative fecundity	$56 \pm 15$	$66 \pm 7$	$67 \pm 22$	$91 \pm 14$
Oocyte diameter	$10.8 \pm 0.29$	10.6 ± 0.36	11.8 ± 1.46	$12.2 \pm 0.95$
Oocyte mass	$0.67 \pm 0.05$	0.64 ± 0.01	0.77 ± 0.17	$0.89 \pm 0.24$