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Layal Fahed, Marc El Beyrouthy, Naïm Ouaini, Véronique Eparvier, Didier Stien. Isolation and characterization of santolinoidol, a bisabolene sesquiterpene from *Achillea santolinoides* subsp *wilhelmsii* (K. Koch) Greuter. *Tetrahedron Letters*, 2016, 57 (17), pp.1892-1894. 10.1016/j.tetlet.2016.03.059 . hal-01299147

HAL Id: hal-01299147

<https://hal.sorbonne-universite.fr/hal-01299147>

Submitted on 7 Apr 2016

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Isolation and characterization of santolinoidol, a bisabolene sesquiterpene from *Achillea santolinoides* subsp *wilhelmsii* (K. Koch) Greuter

Layal Fahed^{a,b}, Marc El Beyrouthy^b, Naïm Ouaini^b, Véronique Eparvier^a, Didier Stien^{a,c,*}

^a) CNRS, Institut de Chimie des Substances Naturelles, UPR 2301, 1 Avenue de la Terrasse, 91198 Gif-sur-Yvette, France

^b) Department of Agricultural Sciences, Holy Spirit University of Kaslik, Kaslik, B.P. 446, Jounieh, Lebanon

^c) Sorbonne Universités, UPMC Univ Paris 06, CNRS, Laboratoire de Biodiversité et Biotechnologies Microbiennes (LBBM), Observatoire Océanologique, Banyuls-sur-Mer, France

* Corresponding author. Tel.: +33 4 30 19 24 76. E-mail address: didier.stien@cnrs.fr (D. Stien).

Abstract

The essential oil (EO) obtained by hydrodistillation of aerial parts of Lebanese *Achillea santolinoides* subsp *wilhelmsii* (K. Koch) Greuter was analysed by GC and GC-MS. The identity of the second major compound was uncertain. Its structure was elucidated using 1D and 2D NMR spectroscopy as well as MS data. It appeared to be a new sesquiterpene with a 1-oxa-bicyclo[5,4,0]-undecane skeleton which was named santolinoidol. The EO revealed interesting antimicrobial potential particularly against *Trichophyton* species, *Staphylococcus aureus* and Methicillin-resistant *Staphylococcus aureus* (MRSA) with MIC values ranging from 16 to 128 µg/ml.

Keywords:

Achillea santolinoides subsp *wilhelmsii*; Antimicrobial activity; Santolinoidol; Structural determination; Essential oil

Antimicrobial resistance is a global problem and a major public health threat. This applies to both developed and developing countries. In fact, according to the United States Centers for Disease Control and prevention (CDC), there are more than 23000 deaths per year in the US as a direct result of antibiotic-resistant infections.¹ Finding alternative complementary antibiotics to be used for non-lethal conditions can contribute to drive back infectious diseases and pathogen resistance.

The genus *Achillea* comprises many species known for their therapeutic benefits.² It was named after Achilles the hero of Iliad who, according to the legend, used the leaves of *Achillea millefolium* known commonly as Yarrow to treat his wounded soldiers.³ *Achillea santolinoides* subsp *wilhelmsii* is used in Pakistan as blood purifier and as cure for stomach-ache, gastric troubles and fever.^{4,5} It is also used in Iran and Turkey for bleeding and wound healing.^{6,7} The main purpose of this study was to characterize the essential oil (EO) of Lebanese *A. santolinoides* subsp *wilhelmsii* and assess its antibacterial and antifungal activities. Although the EO of *A. santolinoides* subsp *wilhelmsii* has been described elsewhere,⁸⁻¹¹ no new metabolites have ever been detected in these oils. In the literature, the major compounds were camphor (48.2 %),⁸ linalool (24.2 %) and 1,8-cineole (15.5 %),⁹ (3Z)-cemberene A (22.6 %) and linalool (14.8 %),⁹ carvacrol (25.1%) and linalool (11.0%),¹⁰ or carvacrol (22.5 %), dihydrocarvone (13.2 %), and linalool (12 %).¹¹

A. santolinoides subsp *wilhelmsii* aerial part EO was light green and was obtained with a 0.58% yield (v/w). GC and GC-MS analyses led to the identification and quantification of 35 components accounting for 84.3% of the EO (Table 1). However, the comparison with MS databases and kovats index (KI) wasn't enough for the identification of the second most abundant compound (**1**) which accounted for 7.1% of the EO and was eventually characterized by purification and NMR. The other main compounds were eucalyptol (8.0%), terpinen-4-ol (6.9%), α -pinene (6.0%), and *trans*-pinocarveol (3.5%).

Table 1

Partial chemical composition of *Achillea santolinoides* subsp *wilhelmsii* essential oil, including compounds accounting for more than 6 % of the oil^a

RI ^b	RI ^c	Compounds	(%)
938	1076	α -Pinene	6.0
1034	1213	Eucalyptol	8.0
1176	1611	Terpinen-4-ol	6.9
1640	2188	<i>T</i> -Cadinol	2.6
1651	2160	Santolinoidol (1)	7.1

^a) Full table is given in supplementary material.

^b) Retention index on a DB-5MS column.

^c) Retention index on an HP Innnowax column.

A. santolinoides subsp *wilhelmsii* EO was fractionated by HPLC. The fractions were searched by GC-MS in order to point out the one containing the unknown compound **1**, which according to GC accounted for 77% of fraction 20. This fraction was analyzed by high resolution MS and NMR. Compound **1** had a pseudomolecular ion peak at m/z 239.2011 in the positive ionization mode corresponding to the molecular formula $C_{15}H_{27}O_2$. Its planar structure was deduced by analysis of 1H , ^{13}C , COSY, HSQC and HMBC spectral data (Table 2). COSY spectrum made it possible to follow the sequence of protons H-2 to H-13, and a group of protons between H-5a and H-9a forming a cyclohexene ring. Chemical shifts and COSY correlation between H-14 and H-9 indicated that H-14 was linked to a double bond bearing the vinyl proton H-9. 1H and ^{13}C chemical shifts indicated that carbon 9a was linked to an oxygen. Two singlet methyl groups at δ_H 1.13 and 1.18, and δ_C 23.7 and 26.3 were not correlated in COSY. HMBC correlations indicated that these two methyl groups were linked to the same quaternary carbon at δ 72.7. This carbon was linked to an oxygen and HMBC correlations between both H-11 and H-12 with C-2 ascertained the position of this isopropenol moiety which was linked to position 2. Long range proton carbon correlations between H-5 and C-5a and between H-13 and C-5a confirmed the existence of the C-

Fig. 2. Selected NOESY correlations in **1**

Table 2

Spectral data for santolinoidol (**1**) in CDCl₃

Position	δ_C , type	δ_H , (J in Hz)	COSY	HMBC	NOESY
2	90.5, CH	3.24, dd (11.7, 2.9)	3x, 3y	3, 9a, 10, 11	3x, 9a, 12
3	32.0, CH ₂	x: 1.78, m y: 1.45, ddd (16.1, 12.5, 2.9)	2, 3y, 4 2, 3x, 4		2, 3y, 12 3x, 4
4	32.3, CH ₂	1.54-1.64, m	3x, 3y, 5	2, 3, 5, 5a, 13	3y, 9a, 13
5	38.3, CH	1.37, m	4, 5a, 13	5a	
5a	46.3, CH	1.15 ddd (16.9, 12.5, 4.0)	5, 6y, 9a		9a
6	25.1, CH ₂	x: 1.68, m y: 1.38, m	6y, 7x, 7y 5a, 6x, 7x, 7y		6y, 13 6x
7	31.0, CH ₂	x: 1.96, br dd (17.6, 4.4) y: 1.87, m	6x, 6y, 7y 6x, 6y, 7x, 8	5a, 6, 8, 9 5a, 9	9
8	139.0				
9	121.5, CH	5.49, br d (3.7)	7y, 9a, 14	7	7x, 9a, 14
9a	76.4, CH	3.93, br t (4.0)	5a, 8, 14	2, 6, 8, 9	2, 4, 5a, 9
10	72.1				
11	23.7, CH ₃	1.13, s		2, 10, 12	
12	26.3, CH ₃	1.18, s		2, 10, 11	2, 3x
13	22.6, CH ₃	1.03, d (6.6)	5	4, 5, 5a	4, 6x
14	23.6, CH ₃	1.72, br s	8, 9a	7, 8, 9	9

Minimum inhibitory concentrations (MICs) of *A. santolinoides* subsp *wilhelmsii* EO are presented in

Table 3. EOs with MICs of 128 μ g/ml and below are generally considered useful as antimicrobial

agents.^{12,13}

Table 3Antimicrobial activity (MIC in µg/ml) of *A. santolinoides* subsp *wilhelmsii* EO

Strain tested ^a	<i>A. santolinoides</i> EO	Reference compound
<i>S. aureus</i> ATCC 29213	128	1 ^b
<i>S. aureus</i> ATCC 33591	128	1 ^c
<i>C. albicans</i> ATCC 10231	256	1 ^d
<i>P. aeruginosa</i> CIP 82118	>512	0.25 ^e
<i>T. rubrum</i> SNB-TR1	32	2 ^d
<i>T. mentagrophytes</i> SNB-TM1	32	1 ^f
<i>T. soudanense</i> SNB-TS1	16	2 ^d
<i>T. violaceum</i> SNB-TV1	16	2 ^d
<i>T. tonsurans</i> SNB-TT1	16	4 ^d

^a *Staphylococcus aureus* ATCC 29213; Methicillin-resistant *Staphylococcus aureus* ATCC 33591; *Candida albicans* ATCC 10231; *Pseudomonas aeruginosa* CIP 82118; *Trichophyton rubrum* SNB-TR1; *Trichophyton mentagrophytes* SNB-TM1; *Trichophyton soudanense* SNB-TS1; *Trichophyton violaceum* SNB-TV1; *Trichophyton tonsurans* SNB-TT1

^b Oxacillin; ^c Vancomycin; ^d Fluconazole; ^e Gentamicin; ^f Itraconazole

A. santolinoides EO was most active on dermatophytes (MIC values 16-32 µg/ml). It also exhibited a significant activity against both strains of *Staphylococcus aureus* (MIC 128 µg/ml). *Candida albicans* was moderately sensitive (MIC 256 µg/ml), whereas *Pseudomonas aeruginosa* was resistant (MIC > 512 µg/ml). The origin of the antimicrobial activity remains uncertain. The oil is composed of many compounds and none of them really dominates. However, the antimicrobial activity of *Melaleuca alternifolia* (tea tree) oil is principally attributed to its terpinen-4-ol content.¹⁴ Hence, this compound may have had an important contribution to the whole activity. Unfortunately, the fraction enriched in santolinoidol was obtained in very small amount and could not be tested alone.

In conclusion, this letter reports the first description of santolinoidol, an unusual 1-oxa-bicyclo[5,4,0]-undecane sesquiterpene isolated in the essential oil of *Achillea santolinoides* subsp *wilhelmsii*.

Acknowledgements

This work has benefited from an "Investissement d'Avenir" grant managed by Agence Nationale de la Recherche (CEBA, ref. ANR-10-LABX-25-01) and from the support of CNRS Lebanon.

Supplementary data

Supplementary data associated with this article can be found, in the online version, at <http://dx.doi.org/....> These data include experimental procedure and all NMR data of compound **1**.

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