

Two gonad-infecting species of Philometra (Nematoda: Philometridae) from groupers (Serranidae) off Tunisia, with a key to Philometra species infecting serranid gonads

František Moravec, Amira Chaabane, Jean-Lou Justine, Lassad Neifar

▶ To cite this version:

František Moravec, Amira Chaabane, Jean-Lou Justine, Lassad Neifar. Two gonad-infecting species of Philometra (Nematoda: Philometridae) from groupers (Serranidae) off Tunisia, with a key to Philometra species infecting serranid gonads. Parasite, 2016, 23, pp.8. 10.1051/parasite/2016008 . hal-01304063

HAL Id: hal-01304063 https://hal.sorbonne-universite.fr/hal-01304063

Submitted on 19 Apr 2016

HAL is a multi-disciplinary open access archive for the deposit and dissemination of scientific research documents, whether they are published or not. The documents may come from teaching and research institutions in France or abroad, or from public or private research centers. L'archive ouverte pluridisciplinaire **HAL**, est destinée au dépôt et à la diffusion de documents scientifiques de niveau recherche, publiés ou non, émanant des établissements d'enseignement et de recherche français ou étrangers, des laboratoires publics ou privés.



Distributed under a Creative Commons Attribution 4.0 International License

Research Article



OPEN ∂ ACCESS

Two gonad-infecting species of *Philometra* (Nematoda: Philometridae) from groupers (Serranidae) off Tunisia, with a key to *Philometra* species infecting serranid gonads

František Moravec^{1,*}, Amira Chaabane², Jean-Lou Justine³, and Lassad Neifar²

¹ Institute of Parasitology, Biology Centre of the Czech Academy of Sciences, Branišovská 31, 370 05 České Budějovice, Czech Republic

² Laboratoire de Biodiversité et Écosystèmes Aquatiques, Faculté des Sciences de Sfax (FSS), Université de Sfax, BP 1171, 3038 Sfax, Tunisia

³ ISYEB, Institut Systématique, Évolution, Biodiversité, UMR7205 CNRS, EPHE, MNHN, UPMC, Muséum National d'Histoire Naturelle, Sorbonne Universités, CP51, 55 Rue Buffon, 75231 Paris cedex 05, France

Received 8 January 2016, Accepted 9 February 2016, Published online 8 March 2016

Abstract – Based on light and scanning electron microscopical studies of nematode specimens (males and mature females) collected from the ovary of groupers (Serranidae, Perciformes) in the Mediterranean Sea off Tunisia (near Tunis and Sfax), two gonad-infecting species of *Philometra* Costa, 1845 (Nematoda, Philometridae) are reported: *Philometra inexpectata* n. sp. from the mottled grouper *Mycteroperca rubra* and *P. jordanoi* (López-Neyra, 1951) from the dusky grouper *Epinephelus marginatus*. Identification of both fish species was confirmed by molecular barcoding. The new species is mainly characterized by the length of equally long spicules (147–165 µm), the gubernaculum (63–93 µm long) bearing at the tip two dorsolateral lamellar parts separated from each other by a smooth median field, a V-shaped mound on the male caudal extremity, the presence of a pair of large caudal papillae located posterior to the cloaca and by the body length of the males (1.97–2.43 mm). *Philometra inexpectata* n. sp. is the fifth known gonad-infecting philometrid species parasitizing serranid fishes in the Mediterranean region. The males of *P. jordanoi* were examined by scanning electron microscopy for the first time; this detailed study revealed some new taxonomically important morphological features, such as the number and arrangement of cephalic and caudal papillae, presence of amphids and phasmids and mainly the lamellate structures at the posterior end of the gubernaculum. A key to gonad-infecting species of *Philometra* parasitic in serranid fishes is provided.

Key words: Philometridae, Fish ovary, *Philometra inexpectata* n. sp., *Philometra jordanoi*, Tunisia, Groupers, Mediterranean Sea.

Résumé – Deux espèces de *Philometra* (Nematoda : Philometridae) parasites des gonades de mérous (Serranidae) de Tunisie, avec une clé des espèces de *Philometra* infectant les gonades des Serranidae. Sur la base d'études, au microscope photonique et électronique, de spécimens de nématodes (mâles et femelles mûres) recueillis dans les ovaires de mérous (Serranidae, Perciformes) de la mer Méditerranée au large de la Tunisie (près de Tunis et Sfax), deux espèces de *Philometra* Costa, 1845 (Nematoda, Philometridae) parasites de gonades sont signalées : *Philometra inexpectata* n. sp. du mérou royal *Mycteroperca rubra* et *P. jordanoi* (López-Neyra, 1951) du mérou noir *Epinephelus marginatus*. L'identification des deux espèces de poissons a été confirmée par les codes-barres moléculaires. La nouvelle espèce est caractérisée principalement par la longueur des spicules (147-165 µm), de tailles égales, le gubernaculum (63-93 µm de long) portant à sa pointe deux parties lamellaires dorsolatérales séparées l'une de l'autre par un champ médian lisse, une bosse en forme de V sur l'extrémité caudale du mâle, la présence d'une paire de grandes papilles caudales situées en arrière du cloaque et par la longueur du corps des mâles (1.97-2.43 mm). *Philometra inexpectata* n. sp. est la cinquième espèce de Philometridae des gonades de poissons Serranidae dans la région méditerranéenne. Les mâles de *P. jordanoi* ont

František Moravec – urn:lsid:zoobank.org:author:DD65585B-7274-4A7B-B7F7-36D20D623633 Amira Chaabane – urn:lsid:zoobank.org:author:8B2085BC-83F8-4631-B439-119DE1A65549 Jean-Lou Justine – urn:lsid:zoobank.org:author:17643DCB-2C9D-4386-BB94-D2F04966B0E9 Lassad Neifar – urn:lsid:zoobank.org:author:5548804C-824C-4193-8913-237041B12E70

This is an Open Access article distributed under the terms of the Creative Commons Attribution License (http://creativecommons.org/licenses/by/4.0), which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

^{*}Corresponding author: moravec@paru.cas.cz

été examinés par microscopie électronique à balayage pour la première fois ; cette étude détaillée a révélé quelques nouvelles caractéristiques morphologiques taxinomiquement importantes, telles que le nombre et la disposition des papilles céphaliques et caudales, la présence d'amphides et de phasmides et principalement les structures lamellaires à l'extrémité postérieure du gubernaculum. Une clé des espèces de *Philometra* parasites des poissons Serranidae est fournie.

Introduction

Gonad-infecting species of philometrid nematodes (Philometridae) are widely distributed in marine fishes of the Atlantic, Indian and Pacific Oceans, and sometimes occur in brackish-water environments [21, 26]. These parasites may be severely pathogenic in fish ovaries and can affect reproduction [15].

The species identification of these parasites, previously mostly based on the morphology of large-sized females, was rather problematic. However, scanning electron microscopical (SEM) examinations of minute philometrid males made the identification more reliable and indicated considerable species diversity in these nematodes. To date, many gonad-infecting species of *Philometra* Costa, 1845 have been described from a variety of marine fishes belonging to different families and their number is quickly increasing [13, 14, 16, 17, 19, 20, 22, 23, 27–29].

During recent helminthological investigations of some marine fishes in the Mediterranean Sea off the Tunisian coast near Tunis and Sfax [2, 28], males and mature females of philometrid nematodes were collected from the ovary of two species of serranid fishes, the mottled grouper *Mycteroperca rubra* (Bloch) and the dusky grouper *Epinephelus marginatus* (Lowe) (both Serranidae, Perciformes). A close examination revealed that they represent one new and one known insufficiently studied species. Both host species are subtropical marine fishes, which are distributed in the Mediterranean Sea and the eastern Atlantic (*M. rubra*) or in the eastern and southern Atlantic and western Indian Oceans (*E. marginatus*) and are targeted by commercial and recreational fisheries [3].

Materials and methods

Fish and their identification

Fish were purchased at the fish market in Tunis and Sfax, Tunisia; these were previously caught by fishermen in the nearby coastal waters of the Mediterranean Sea. Fish DNA was extracted from tissue samples using the NucleoSpin 96 tissue kit (Macherey-Nagel, Düren, Germany) following the manufacturer's instructions. Sequences were obtained by amplification and sequencing of a region of the cytochrome oxidase subunit I (COI) mitochondrial gene using the primers FishF1 (5'-TCAACYAATCAYAAAATYGGCAC-3') and (5'-TGATTYTTYGGYCACCCRGAAGT-3') [34]. FishR1 Standard PCRs were carried out in 20 µL total volume, containing about 30 ng of DNA, $1 \times 10 \times$ PCR buffer, 2 mM MgCl₂, 200 µM mix dNTPs, 150 nM of each primer and 1 unit of Taq polymerase (Qiagen, Hilden, Germany). After an initial denaturation of 3 min at 95 °C, the mitochondrial DNA was amplified through 39 cycles of 15 s at 95 °C, 20 s at 48 °C and 40 s at 72 °C, with a terminal elongation for 5 min at 72 °C. PCR products were purified and sequenced in both directions on 3730xl DNA Analyser 96-capillary sequencer (Applied Biosystems, Waltham, MA, USA). Sequences were edited using CodonCode Aligner software (CodonCode Corporation, Dedham, MA, USA), compared with the GenBank database content using BLAST, and deposited in GenBank under Accession Numbers KU739518– KU739521. Species identification was confirmed using the BOLD identification engine [32]. Since BOLD does not include all sequences available in GenBank but includes others, comments are added for similarities with other sequences. The fish nomenclature adopted follows FishBase [3].

Nematodes

Philometrid specimens were collected from frozen-thawed fish gonads under the dissecting microscope. They were fixed in hot 70% ethanol and cleared with glycerine for light microscopical (LM) examination. Drawings were made with the aid of a Zeiss drawing attachment. Specimens used for scanning electron microscopy were postfixed in 1% osmium tetroxide (in phosphate buffer), dehydrated through a graded acetone series, critical-point-dried and sputter-coated with gold; they were examined using a JEOL JSM-7401F scanning electron microscope at an accelerating voltage of 4 kV (GB low mode). All measurements are in micrometres unless otherwise indicated.

Results and discussion

Philometra inexpectata n. sp. (Figs. 1, 2)

urn:lsid:zoobank.org:act:FF5B31D3-A815-42FB-B64A-FC445FF77824

Type-host: Mottled grouper, *Mycteroperca rubra* (Bloch) (Serranidae, Perciformes). After identifying the fish based upon morphological characteristics, identification was confirmed via barcoding. The COI sequence obtained for our specimen (GenBank Accession Number KU739518) was identical (100% similarity, 94% cover) to a sequence identified as *M. rubra* from off Israel (KF564307; unpublished). BOLD provided similar results. *Mycteroperca rubra* could be confused with the Island grouper *M. fusca* (Lowe), a species from the eastern Atlantic Ocean. Heemstra et al. (2010) [5] reported the presence of *M. fusca* for the first time in the Mediterranean Sea, off Israel. They wrote: "A reasonable possibility is that



Figure 1. *Philometra inexpectata* n. sp. A: Anterior end of male, lateral view. B: Cephalic end of male, apical view. C, D: Distal end of gubernaculum, dorsal and lateral views, respectively. E: Anterior end of mature female, lateral view. F: Vulva of mature female, lateral view. G, H: Posterior end of male, lateral and ventral views, respectively. I: Posterior end of mature female, lateral view. J: Caudal end of male, apical view.

M. fusca entered the Mediterranean through the Strait of Gibraltar, as many Atlantic species do, then expanded its distribution along the North African coast and was overlooked or confused with *M. rubra*." Unfortunately, no COI sequence of *M. fusca* is available for comparison. Incidentally, we also noted that the sequence we obtained from our specimen was very close (99% similarity, 94% cover) to several other sequences labelled as *M. acutirostris* from Brazil (e.g., KF836485; unpublished); *M. acutirostris* also resembles *M. rubra* and *M. fusca* morphologically, but it has never been reported in the Mediterranean Sea [5]. We thus conclude that the species studied herein from Tunisia is *M. rubra*, but we note that barcoding of more species is needed.

Site of infection: Ovary.

Type-locality: Tunis (fish market), Tunisia (collected 10 September 2015).

Prevalence and intensity: 1 fish infected/1 fish examined; 16 philometrid specimens.

Type-specimens: Holotype, allotype and 39 paratypes, Muséum National d'Histoire Naturelle, Paris, MNHN HEL553-554; 3 male paratypes in the Helminthological Collection of the Institute of Parasitology, Biology Centre of the Czech Academy of Sciences, České Budějovice (Cat. No. N–1109).

Etymology: The specific name *inexpectata* (= unexpected) is a Latin adjective and relates to the fact that gonad-infecting philometrids from M. *rubra* were not previously considered to represent a new species.



Figure 2. *Philometra inexpectata* n. sp., scanning electron micrographs of male. A: Cephalic end, apical view (arrows indicate amphids). B: Caudal end, apical view. C: Region of cloaca, ventral view. D: Caudal end, dorsal view. E, F: Distal end of gubernaculum, dorsal and lateral views, respectively. G: Caudal end, lateral view. *Abbreviations*: a, submedian pair of external cephalic papillae; b, submedian cephalic papilla of internal circle; c, lateral cephalic papilla of internal circle; d, group of four adanal caudal papillae; e, large papilla posterior to cloaca; f, caudal mound; g, gubernaculum; o, oral aperture; s, spicule.

Description

Male (13 specimens; measurements of holotype in parentheses): Body filiform, whitish, 1.97-2.43 (2.43) mm long, maximum width at middle of body 54-69 (57); anterior part of body not narrowed just posterior to cephalic end (Fig. 1A). Maximum width/body length ratio 1:33-43 (1:43). Cuticle smooth. Cephalic end rounded, 27-36 (27) wide. Oral aperture small, triangular, surrounded by small circular elevation. Fourteen minute cephalic papillae arranged in 2 circles present: external circle formed by 4 submedian pairs of papillae; internal circle by 4 submedian and 2 lateral papillae. Small lateral amphids just posterior to lateral papillae of internal circle, followed by fairly large lateral crescent-shaped formations of slightly elevated cuticle (Figs. 1B, 2A). Oesophagus 246-360 (249) long, comprising 10-17% (10%) of body length, with inflation at anterior end measuring 27- $33 \times 18-24$ (27 × 18); posterior part of muscular oesophagus overlapped by well-developed oesophageal gland with large cell nucleus; maximum width of gland 15-21 (18). Nerve ring and oesophageal nucleus 105-135 (120) and 171-225 (171) from anterior extremity, respectively. Excretory pore 135-177 (165) from anterior end. Testis extending anteriorly to level of nerve ring (Fig. 1A), usually overlapping posterior portion of oesophagus. Posterior end of body blunt, 27-33 (30) wide, provided with broad V-shaped mound situated laterally and dorsally to cloacal opening. Four adanal pairs of very flat, hardly visible caudal papillae present on anterior parts of caudal mound; additional pair of large subdorsal papillae situated posterior to cloacal aperture (Figs. 1J, 2B-D, G). Phasmids not observed. Spicules slender, needlelike, equally long, with somewhat expanded proximal and sharply pointed distal tips (Figs. 1G, H, 2B-G); length of spicules 147-165 (153), representing 6-8% (7%) of body length. Gubernaculum 63-93 (72) long, with anterior portion somewhat dorsally bent; length of anterior bent part 27-33 (33), representing 31-50% (46%) of entire gubernaculum length (Figs. 1G, H); distal end of gubernaculum with numerous dorsal transverse lamella-like structures demarcating depressed smooth field between them and with two ventral longitudinal grooves (Figs. 1C, D, 2B-G). Length ratio of gubernaculum and spicules 1:1.72-2.38 (1:2.13). Spicules and gubernaculum well sclerotized, yellowish, anterior part of gubernaculum colourless.

Nongravid female (3 mature specimens; measurements of allotype in parentheses): Length of body 1.65–1.73 (1.73) mm, maximum width 45–57 (57); maximum width/body length ratio 1:30-38 (1:30). Width of anterior end 30–33 (33). Cephalic structures not studied. Entire oesophagus 270–459 (310) long and 30 (30) wide. Anterior oesophageal bulb 30–36 (30) long, 21–34 (24) wide. Nerve ring and oesophageal nucleus 99–129 (99) and 216–300 (219), respectively, from anterior extremity (Fig. 1E). Vulva and incompletely developed vagina present (Fig. 1F); former situated 1.13–1.21 (1.21) mm from anterior extremity, at 68–70% (70%) of body length. Uterus empty. Posterior end rounded, without caudal projections (Fig. 1I).

Remarks

In view of a high degree of host specificity in gonad-infecting species of Philometra [22, 23, 27, 28], P. inexpectata n. sp. is compared with 14 other gonad-infecting nominal species of this genus described from fishes of the perciform family Serranidae: P. aenei Moravec, Chaabane, Neifar, Gey & Justine, 2016; P. cephalopholidis Moravec & Justine, 2015; P. charlestonensis Moravec, de Buron, Baker & González-Solís, 2008; P. cyanopodi Moravec & Justine, 2008; P. fasciati Moravec & Justine, 2008; P. hyporthodi Moravec & Bakenhaster, 2013; P. indica Moravec & Manoharan, 2014; P. jordanoi (López-Neyra, 1951); P. margolisi Moravec, Vidal-Martínez & Aguirre-Macedo, 1995; P. mexicana Moravec & Salgado-Maldonado, 2007; P. piscaria Moravec & Justine, 2014; P. serranellicabrillae Janiszewska, 1949; P. tropica Moravec & Manoharan, 2014; and P. tunisiensis Moravec, Chaabane, Neifar, Gey & Justine, 2016. Four of them, P. aenei, P. jordanoi, P. serranellicabrillae and P. tunisiensis, occur in the Mediterranean region [6, 11, 28].

Morphological and biometrical differences of P. inexpectata n. sp. from all the above-mentioned species are apparent from the identification key at the end of this paper. To date, only two gonad-infecting species of Philometra have been reported to parasitize hosts of the genus Mycteroperca Gill: P. charlestonensis, a parasite of M. phenax in the North American Atlantic region (USA) [25], and P. lateolabracis (Yamaguti, 1935) from M. rubra in the Mediterranean Sea off Turkey (Iskenderun Bay) [18]. However, with respect to the redescription of *P. lateolabracis* by Quiazon et al. [30], the nematodes from M. rubra off Turkey, studied only by LM, were evidently misidentified [12]. Since their morphology and measurements agree with those of the newly described species and because the host species (M. rubra), localization in the host and the geographical region (Mediterranean Sea) are identical, they evidently belong to P. inexpectata n. sp.

Philometra charlestonensis distinctly differs from *P. inexpectata* n. sp. in having shorter spicules $(123-141 \ \mu m \ vs. 147-165 \ \mu m)$ and mainly in the male caudal mound consisting of two lateral parts widely separated from each other dorsally (vs. caudal mound V-shaped, not interrupted dorsally).

Philometra jordanoi (López-Neyra, 1951) Yamaguti, 1961 (Figs. 3, 4)

Syn.: Sanguinofilaria jordanoi López-Neyra, 1951.

Type-host and type-locality: Dusky grouper, *Epinephelus marginatus* (Lowe) (as *E. guaza* (Linnaeus)) (Serranidae, Perciformes), Tétouan (fish market), Morocco. Other localities: see text.

Material from Tunisia.

Host: *Epinephelus marginatus*. After identifying the fish based upon morphological characteristics, identification was confirmed via barcoding. We obtained sequences (GenBank KU739519–KU739521) from three specimens; only one



Figure 3. *Philometra jordanoi* (López-Neyra, 1951). A: Anterior end of male, lateral view. B, C: Posterior end of male, lateral and ventral views, respectively. D: Cephalic end of male, apical view. E: Anterior end of mature female, lateral view. F: Caudal end of male, apical view. G: Posterior end of mature female, lateral view. H, I: Posterior end of gubernaculum, dorsal and lateral views, respectively.

(corresponding sequence: KU739521) had mature gonads and was used in this study. The three sequences were almost identical, with 1–2 bp differences; BLAST results show that our sequences were identical or almost identical (99–100% similarity) to several sequences all identified as *E. marginatus*, such as KC500679 (from Iskenderun Bay, Turkey [7]), JF493449 and JF493450 (from South Africa, unpublished), KM077929 (Senegal [33]) and sequences from Brazil such as KF836469 (unpublished). Our sequences were also identical or almost identical (99–100% similarity) to shorter sequences identified as *E. marginatus* such as FN688939 (from France [8]). BOLD gave similar results, but the automatic identification was disrupted by the presence in the database of an "early sequence" labelled as *Epinephelus fasciatus* (Forsskål) (clearly a misidentification). We consider that barcoding definitely demonstrates that our specimens belong to *E. marginatus*.

Site of infection: Ovary.

Locality: Sfax (fish market), Tunisia, collected 22 October 2015.

Prevalence and intensity: 1 fish infected/1 fish examined; 7 philometrid specimens.

Voucher specimens: 1 male and 1 female nematode, Muséum National d'Histoire Naturelle, Paris, MNHN HEL555; 1 male and 1 female specimen in the Helminthological



Figure 4. *Philometra jordanoi* (López-Neyra, 1951), scanning electron micrographs of male. A: Cephalic end, subapical view. B: Caudal end, apical view (arrows indicate phasmids). C: Same, sublateral view. D: Same, dorsal view (arrows indicate phasmids). E, F: Distal end of gubernaculum, dorsal and lateral views, respectively. G, H: Region of cloaca, ventral and dorsal views (distal ends of gubernaculum and one spicule broken-away). *Abbreviations*: a, submedian pair of external cephalic papillae; b, submedian cephalic papilla of internal circle; c, lateral cephalic papilla of internal circle; d, group of four adanal caudal papillae; e, large papilla posterior to cloaca; f, caudal mound; g, gubernaculum; o, oral aperture; s, spicule.

Collection of the Institute of Parasitology, Biology Centre of the Czech Academy of Sciences, České Budějovice (Cat. No. N-1110).

Description

Male (5 specimens): Body whitish, filiform, tapering to both ends, 2.45-2.91 mm long, maximum width at middle 51-60; anterior part of body not narrower just posterior to cephalic end (Fig. 3A). Maximum width/body length ratio 1:48-57; width of cephalic end 24-27, that of posterior end 27. Cuticle smooth. Cephalic end rounded. Oral aperture small, with lobular rim, surrounded by 14 cephalic papillae arranged in 2 circles: external circle formed by 4 submedian pairs of papillae; internal circle formed by 4 submedian and 2 lateral papillae. Small lateral amphids just posterior to lateral papillae of internal circle (Figs. 3D, 4A). Oesophagus 501-564 long, maximum width 18-30, slightly inflated at anterior end; posterior part of muscular oesophagus overlapped by well-developed oesophageal gland with large cell nucleus in middle (Fig. 3A); anterior oesophageal inflation 27-30 long and 18-21 wide. Nerve ring, excretory pore and oesophageal nucleus 144-159, 195-225 and 312-357, respectively, from anterior extremity. Testis reaching anteriorly to level of nerve ring (Fig. 3A). Posterior end of body blunt, with broad, V-shaped mound extending laterally and dorsally (Figs. 3B, C, F, 4B-D). Four pairs of very flat, hardly visible caudal papillae close to each other situated on sides of cloacal aperture on mound (Figs. 3F, 4B–D, G, H). Pair of small phasmids present at about middle of each mound arm (Figs. 3F, 4B, D). Spicules equally long; proximal ends somewhat expanded, distal tips sharply pointed (Fig. 3B, C); length of spicules 213–252, comprising 8% of body length. Gubernaculum 81-84 long, with anterior portion somewhat dorsally bent; length of anterior bent part 24-27, representing 30-32% of entire gubernaculum length; posterior end of gubernaculum with 2 dorsolateral longitudinal parts bearing numerous transverse lamella-like structures demarcating smooth field between them and with two ventral longitudinal grooves (Figs. 3B, C, H, I, 4E-G). Length ratio of gubernaculum and spicules 1:2.61-3.11. Spicules and gubernaculum well sclerotized, yellowish, anterior part of gubernaculum colourless.

Nongravid female (2 specimens): Body whitish, filiform, tapering to both ends, 2.48-2.94 mm long, maximum width at middle 48-51. Maximum width/body length ratio 1:52-58; width of cephalic end 24-27, that of posterior end 21. Cuticle smooth. Cephalic end rounded (Fig. 3E), its detailed structure not studied by SEM. Oesophagus provided with large oesophageal gland at its posterior portion, extending from level of nerve ring to end of oesophagus; length of entire oesophagus 600-642 (comprising 22-24% of body length), maximum width including gland 27-30; anterior end of oesophagus slightly inflated at anterior end (Fig. 3E); anterior inflation 27-30 long and 18-21 wide. Nerve ring 147 from anterior extremity; excretory pore indistinct in largest specimen, in smaller specimen situated at 231 from anterior end of body. Small ventriculus 6 long and 9 wide. Intestine narrow, ending blindly, attached by short ligament to body wall near caudal

extremity. Vulva absent. Uterus empty. Caudal end rounded (Fig. 3G).

Remarks

This species was originally described by López-Neyra [9] as Sanguinofilaria jordanoi, based solely on female specimens collected in the ovary of E. marginatus [syn. E. gigas (Brünnich)] obtained from the market in Tétouan, Morocco (probably caught in the Mediterranean Sea). Later the species was transferred to Philometra as P. jordanoi [36]. The genus Sanguinofilaria Yamaguti, 1941 was subsequently synonymized with Philometra [31]. The original description of P. jordanoi [9] was inadequate. Later Moravec et al. [24], based on available nematode specimens from the ovary of wild E. marginatus collected in the Mediterranean Sea near the Balearic Islands, Spain and those from wild and cultured E. marginatus in the Tyrrhenian Sea off Sicily, Italy, provided a somewhat more detailed description of philometrid gravid females, which was, more or less, in agreement with that of P. jordanoi. Nevertheless, the authors identified this material as P. lateolabracis, a species described from females collected in three species of perciform fishes off Japan [35], and designated P. jordanoi to be its junior synonym.

Subsequently, Moravec and Genc [18], based on available body fragments of nematode gravid females from the ovary of *E. marginatus* in the Mediterranean Sea off Turkey (Iskenderun Bay), described the female anterior end including cephalic structures of these nematodes and the first-stage larva from the uterus. They again identified these nematodes as *P. lateolabracis*. From the same locality (Iskenderun Bay off Turkey) and the same host species, *P. lateolabracis* was also reported by Genc et al. [4].

Merella et al. [10] were the first to provide the description of the male (based on a single available specimen studied by LM) of a philometrid collected from the ovary of *E. marginatus* in waters near Majorca, Spain, which was identified as *P. lateolabracis*. But in their subsequent paper [11], they re-erected *P. jordanoi*, to which they assigned the abovementioned male specimen.

However, Quiazon et al. [30] were the only ones to discover the males of *P. lateolabracis* from the type-host in Japan and provided their detailed description based on LM and SEM examinations, which enabled a comparison with other gonadinfecting *Philometra* spp. with described males. Their study showed that *P. lateolabracis* is a specific parasite of *Lateolabrax japonicus* (Cuvier) (Lateolabracidae) and that all previous records of this parasite from many other fish species were apparently based on misidentifications. Comparison of the males of *P. lateolabracis* and those of *P. jordanoi* confirmed the validity of the latter species [12].

It is apparent that all the previous records of *P. lateolabracis* in *E. marginatus* in the Mediterranean region [4, 18, 24] concerned, in fact, *P. jordanoi*. Also, the nematodes designated as *Philometra* sp. from the ovary of *E. marginatus* in Iskenderun Bay off Turkey [1] should be assigned to this species.

The present detailed study of the males of *P. jordanoi*, including the first use of SEM, made it possible to describe

some new taxonomically important morphological features in this species, such as the presence of lamellate structures on the distal end of the gubernaculum, the number and distribution of cephalic and caudal papillae, the character of the male caudal mound, the structure of the male oesophagus, the location of the nerve ring and excretory pore, and the morphology of the mature female. As compared with other gonad-infecting congeneric species parasitizing serranid fishes, *P. jordanoi* is remarkable for its rather long spicules, which may attain up to 265 µm [11]. The comparison of *P. jordanoi* with other species from the gonads of serranids is more apparent from the following key.

Key to gonad-infecting species of *Philometra* parasitizing fishes of the family Serranidae

1.	Males unknown. Body length of gravid female 40–60 mm.
	In Serranus cabrilla (Linnaeus); Mediterranean Sea
	regionP. serranellicabrillae
_	Both males and females known
2.	Spicules 432–468 µm long. Gravid female 65–85 mm
	long. In <i>Epinephelus morio</i> (Valenciennes): Gulf of
	Mexico
_	Spicules shorter than 300 µm
3.	Spicules longer than 160 µm
_	Spicules shorter than 160 µm
4.	Spicules conspicuously distended between second and
	fourth quarter of their length: spicules 168–186 µm long.
	length of gubernaculum 120–138 um. In <i>Epinephelus</i>
	<i>bleekeri</i> (Vaillant): Indian Ocean (Bay of Bengal)
	P. tronica
_	Spicules slender, not markedly distended
5	Length of gubernaculum at most 87 µm 6
_	Gubernaculum longer than 90 µm
6.	Pair of large caudal papillae posterior to cloaca absent.
	Length of spicules 201–219 µm, gubernaculum 78–
	87 um long: length ratio of gubernaculum and spicules
	1:2.52–2.77. In Epinephelus costae (Steindachner): Medi-
	terranean Sea
_	Pair of large caudal papillae posterior to cloaca
	present
7.	Length of spicules 192–195 µm, gubernaculum 84 µm
	long; length ratio of gubernaculum and spicules 1:2.32.
	In Epinephelus merra Bloch; Indian Ocean (Bay of
	Bengal)P. indica
—	Length of spicules 213–265 µm, gubernaculum 81–84 µm
	long; length ratio of gubernaculum and spicules
	1:2.61-3.15. In Epinephelus marginatus; Mediterranean
	SeaP. jordanoi
8.	Length of spicules 171-180 µm, representing 4-5% of
	male body length; gubernaculum 126–144 µm long. Male
	caudal mound V-shaped. Body length of male 3.67-
	4.19 mm. In Epinephelus coioides (Hamilton); South
	Pacific (off New Caledonia) and Indian Ocean (Persian
	Gulf)P. piscaria
—	Length of spicules 183-228 µm, representing 6-8% of
	male body length; gubernaculum 129–162 µm long.
	Male caudal mound U-shaped. Body length of male

2.72–3.59 mm. In *Epinephelus cyanopodus* (Richardson); South Pacific (off New Caledonia)......P. cyanopodi

- Gubernaculum with distinct dorsal barb situated postequatorially and conspicuous dorsal protuberance at posterior end in lateral view. Length of spicules 108–123 μm, gubernaculum 96–108 μm long. In *Epinephelus aeneus* (Geoffroy Saint-Hilaire); Mediterranean Sea....... *P. aenei*

- Spicules 147–156 μm long, length of gubernaculum 69– 84 μm. Testis reaching anteriorly only to posterior end of oesophagus. Body length of males 2.75–3.32 mm, of gravid female 387 mm. In *Epinephelus fasciatus* (Forsskål); South Pacific (off New Caledonia).......*P. fasciati*

- Length of spicules 129–147 μm, gubernaculum 96– 111 μm long. Testis reaching anteriorly to region of oesophageal nucleus. Body length of male 2.53– 2.91 mm. In *Cephalopholis sonnerati* (Valenciennes); South Pacific (off New Caledonia)......*P. cephalopholidis*
- Length of spicules 147–165 μm, gubernaculum 63–93 μm long. Testis reaching anteriorly to level of nerve ring. Body length of male 1.97–2.43 mm. In *Mycteroperca rubra*; Mediterranean Sea*P. inexpectata* n. sp.
- Body length of male 2.01–3.14 mm, of gravid females 178–230 mm. Maximum body width/length ratio of gravid females 1:131–163. Length of spicules 123–141 μm, gubernaculum 54–93 μm long. Gravid and subgravid females with pair of small papilla-like caudal projections. Larvae from uterus 544–648 μm long. In *Mycteroperca phenax*; western Atlantic Ocean (off USA) including Gulf of Mexico......*P. charlestonensis*

Conflict of interest

The Editor-in-Chief of Parasite is one of the authors of this manuscript. COPE (Committee on Publication Ethics, http://publicationethics.org), to which Parasite adheres, advises special treatment in these cases. In this case, the peer-review process was handled by an Invited Editor, Dominique Vuitton.

Acknowledgements. Amira Chaabane thanks the staff of Service de Systématique Moléculaire, MNHN, Paris, especially Régis Debruyne, Josie Lambourdière, Céline Bonillo and Delphine Gey for excellent teaching in molecular techniques. Authors' thanks are also due to the staff of the Laboratory of Electron Microscopy, Institute of Parasitology, Biology Centre of the CAS, České Budějovice for their technical assistance and to Blanka Škoríková of the same Institute for help with illustrations. This study was partly supported by the Czech Science Foundation (Grant No. P505/12/G112) and the Institute of Parasitology, BC AS CR (institutional support RVO: 60077344), by MNHN Grant "ATM barcode" and by the Mistrals ENVI-MED project BIOPARMED.

References

- 1. Cengizler I, Sarihan E. 1995. A case of *Philometra* sp. met in ovarium of *Epinephelus gigas*. Ege University Journal of Fisheries and Aquatic Sciences, 12, 89–92 (In Turkish with English abstract.).
- Chaabane A, Neifar L, Justine J-L. 2015. *Pseudorhabdosynochus regius* n. sp. (Monogenea, Diplectanidae) from the mottled grouper *Mycteroperca rubra* (Teleostei) in the Mediterranean Sea and Eastern Atlantic. Parasite, 22, 9.
- 3. Froese R, Pauly D. 2015. FishBase. World Wide Web electronic publication. Available on http://www.fishbase.org
- Genc E, Genc MA, Genc E, Cengizler I, Can MF. 2005. Seasonal variation and pathology associated with Helminthes infecting two serranids (Teleostei) of Iskenderun Bay (Northeast Mediterranean Sea), Turkey. Turkish Journal of Fisheries and Aquatic Sciences, 5, 29–33.
- 5. Heemstra P, Aronov A, Goren M. 2010. First record of the Atlantic island grouper *Mycteroperca fusca* in the Mediterranean Sea. Marine Biodiversity Records, 3, e92.
- Janiszewska J. 1949. Some fish nematodes from the Adriatic Sea. Zoologica Poloniae, 5, 7–30.
- Keskİn E, Atar HH. 2013. DNA barcoding commercially important fish species of Turkey. Molecular Ecology Resources, 13, 788–797.
- Kochzius M, Seidel C, Antoniou A, Botla SK, Campo D, Cariani A, Vazquez EG, Hauschild J, Hervet C, Hjörleifsdottir S, Hreggvidsson G, Kappel K, Landi M, Magoulas A, Marteinsson V, Nölte M, Planes S, Tinti F, Turan C, Venugopal MN, Weber H, Blohm D. 2010. Identifying fishes through DNA barcodes and microarrays. PLoS One, 5, e12620.
- López-Neyra CR. 1951. Sanguinofilaria jordanoi n. sp. (Nematoda; Filaroidea). Boletin de la Universidad de Granada, Farmacia, 1, 291–293.
- Merella P, Reñones O, Garippa G. 2004. Finding of one male *Philometra lateolabracis* (Nematoda: Philometridae) parasite on the dusky grouper *Epinephelus marginatus* (Osteichthyes: Serranide) in the western Mediterranean. Parassitologia, 46(Supplement 1), 158.
- Merella P, Reñones O, Garippa G. 2005. Reinstatement of *Philometra jordanoi* (López-Neyra, 1951) (Nematoda: Philometridae): a parasite of the Mediterranean dusky grouper *Epinephelus marginatus* (Lowe) (Osteichthyes: Serranidae). Systematic Parasitology, 61, 203–206.
- Moravec F. 2008. Systematic status of *Philometra jordanoi* (López-Neyra, 1951) and some other congeneric species previously identified as *Philometra lateolabracis* (Yamaguti,

1935) (Nematoda: Philometridae). Folia Parasitologica, 55, 159–160.

- 13. Moravec F, Ali AH. 2014. Additional observations on *Philometra* spp. (Nematoda: Philometridae) in marine fishes off Iraq, with the description of two new species. Systematic Parasitology, 87, 259–271.
- 14. Moravec F, Barton DP. 2015. Two gonad-infecting species of *Philometra* (Nematoda: Philometridae) from marine fishes off the northern coast of Australia. Parasite, 22, 4.
- 15. Moravec F, de Buron I. 2013. A synthesis of our current knowledge of philometrid nematodes, a group of increasingly important fish parasites. Folia Parasitologica, 60, 81–101.
- Moravec F, Diggles BK. 2014. Two new gonad-infecting species of *Philometra* Costa, 1845 (Nematoda: Philometridae) from marine fishes off the northern coast of Australia. Systematic Parasitology, 89, 33–44.
- 17. Moravec F, Diggles BK. 2015. A new gonad-infecting species of *Philometra*, *P. barnesi* sp. n. (Nematoda: Philometridae), from the marine fish *Pomadasys argenteus* (Haemulidae) off the northern coast of Australia. Parasitology Research, 114, 4121–4126.
- Moravec F, Genc E. 2004. Redescription of three *Philometra* spp. (Nematoda, Philometridae) from the gonads of marine perciform fishes of Iskenderun Bay (North-East Mediterranean), Turkey. Acta Parasitologica, 49, 31–40.
- 19. Moravec F, Justine J-L. 2014. Philometrids (Nematoda: Philometridae) in carangid and serranid fishes off New Caledonia, including three new species. Parasite, 21, 21.
- Moravec F, Justine J-L. 2015. New records of species of *Philometra* (Nematoda: Philometridae) from marine fishes off New Caledonia, including *P. cephalopholidis* sp. n. from *Cephalopholis sonnerati* (Serranidae). Parasitology Research, 114, 3223–3228.
- Moravec F, Manoharan J. 2013. Gonad-infecting philometrids (Nematoda: Philometridae) including four new species from marine fishes off the eastern coast of India. Folia Parasitologica, 60, 105–122.
- Moravec F, Manoharan J. 2014. Gonad-infecting species of *Philometra* (Nematoda: Philometridae) from groupers *Epinephelus* spp. (Osteichthyes: Serranidae) in the Bay of Bengal, India. Acta Parasitologica, 59, 596–605.
- Moravec F, Manoharan J. 2014. Two new gonad-infecting species of *Philometra* (Nematoda: Philometridae) parasitic in *Lutjanus* spp. (Osteichthyes: Lutjanidae) in the Bay of Bengal, India. Parasitology Research, 113, 3299–3307.
- 24. Moravec F, Glamuzina B, Marino G, Merella P, Di Cave D. 2003. Occurrence of *Philometra lateolabracis* (Nematoda: Philometridae) in the gonads of marine perciform fishes in the Mediterranean region. Diseases of Aquatic Organisms, 53, 267–269.
- 25. Moravec F, de Buron I, Baker TG, González-Solís D. 2008. Some gonad-infecting species of *Philometra* (Nematoda, Philometridae) from offshore fishes of South Carolina and Georgia, USA, including *Philometra charlestonensis* sp. nov. from the scamp *Mycteroperca phenax*. Acta Parasitologica, 53, 382–391.
- 26. Moravec F, Gopalakrishnan A, Rajkumar M, Saravanakumar A, Kaliyamoorthy S. 2011. A new gonad-infecting species of *Philometra* Costa, 1845 (Nematoda: Philometridae) from the marine fish *Terapon jarbua* (Forsskål) (Terapontidae) off the eastern coast of India. Systematic Parasitology, 80, 23–33.

- 27. Moravec F, Bakenhaster M, Fajer-Ávila EJ. 2014. Three new gonad-infecting species of *Philometra* (Nematoda: Philometridae) parasitic in *Lutjanus* spp. (Lutjanidae) in the northern Gulf of Mexico off Florida. USA. Folia Parasitologica, 61, 355–369.
- 28. Moravec F, Chaabane A, Neifar L, Gey D, Justine J-L. 2016. Descriptions of *Philometra aenei* n. sp. and *P. tunisiensis* n. sp. (Nematoda: Philometridae) from *Epinephelus* spp. off Tunisia confirm a high degree of host specificity of gonad-infecting species of *Philometra* Costa, 1845 in groupers (Serranidae). Systematic Parasitology, 93, 115–128.
- Quiazon KMA, Yoshinaga T. 2013. Gonad-infecting philometrid *Philometra philippinensis* sp. nov. (Nematoda, Philometridae) from the bigeye barracuda *Sphyraena forsteri* Cuvier (Sphyraenidae) off Mariveles, Bataan Province, Philippine Archipelago. Acta Parasitologica, 58, 504–514.
- Quiazon KMA, Yoshinaga T, Ogawa K. 2008. Taxonomical study into two new species of *Philometra* (Nematoda: Philometridae) previously identified as *Philometra lateolabracis* (Yamaguti, 1935). Folia Parasitologica, 55, 29–41.

- Rasheed S. 1963. A revision of the genus *Philometra* Costa, 1845. Journal of Helminthology, 37, 89–130.
- Ratnasingham S, Hebert PDN. 2007. BOLD: The Barcode of Life Data System (www.barcodinglife.org). Molecular Ecology Notes, 7, 355–364.
- Schoelinck C, Hinsinger DD, Dettaï A, Cruaud C, Justine J-L. 2014. A phylogenetic re-analysis of groupers with applications for ciguatera fish poisoning. PLoS One, 9, e98198.
- Ward RD, Zemlak TS, Innes BH, Last PR, Hebert PD. 2005. DNA barcoding Australia's fish species. Philosophical Transactions of the Royal Society of London B Biological Sciences, 360, 1847–1857.
- Yamaguti S. 1935. Studies on the helminth fauna of Japan. Part
 I. Nematodes of fishes. Japanese Journal of Zoology, 6, 337–386.
- Yamaguti S. 1961. The nematodes of vertebrates. Part 1. Systema helminthum III. Interscience Publishers: New York– London. p. 679.

Cite this article as: Moravec F, Chaabane A, Justine J & Neifar L: Two gonad-infecting species of *Philometra* (Nematoda: Philometridae) from groupers (Serranidae) off Tunisia, with a key to *Philometra* species infecting serranid gonads. Parasite, 2016, 23, 8.

PARASITE

An international open-access, peer-reviewed, online journal publishing high quality papers on all aspects of human and animal parasitology

Reviews, articles and short notes may be submitted. Fields include, but are not limited to: general, medical and veterinary parasitology; morphology, including ultrastructure; parasite systematics, including entomology, acarology, helminthology and protistology, and molecular analyses; molecular biology and biochemistry; immunology of parasitic diseases; host-parasite relationships; ecology and life history of parasites; epidemiology; therapeutics; new diagnostic tools.

All papers in Parasite are published in English. Manuscripts should have a broad interest and must not have been published or submitted elsewhere. No limit is imposed on the length of manuscripts.

Parasite (open-access) continues Parasite (print and online editions, 1994-2012) and Annales de Parasitologie Humaine et Comparée (1923-1993) and is the official journal of the Société Française de Parasitologie.

Editor-in-Chief: Jean-Lou Justine, Paris Submit your manuscript at http://parasite.edmgr.com/