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1 Diacylglycerol kinases activate tobacco NADPH oxidase-dependent oxidative burst in  
2 response to cryptogein

3

4 Running head: Control of RBOHD activity by DGK

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27 Summary statement

28 Cryptogein is a protein secreted by the oomycete *Phytophthora cryptogea* that activates  
29 defence mechanisms in tobacco. We show here in BY-2 tobacco suspension cells that  
30 phosphatidic acid rapidly accumulates in response to cryptogein due to the coordinated onset  
31 of phosphoinositide-dependent phospholipase C and diacylglycerol kinase activities. Both  
32 enzyme specific inhibitors and silencing of the phylogenetic cluster III of the tobacco DGK  
33 family were found to reduce PA production upon elicitation and to strongly decrease the  
34 RBOHD-mediated oxidative burst. This establishes that phosphatidic acid production by  
35 diacylglycerol kinases is upstream of the oxidative burst in response to cryptogein.

36

37

38 **ABSTRACT**

39 Cryptogein is a 10 kDa-protein secreted by the oomycete *Phytophthora cryptogea* that  
40 activates defence mechanisms in tobacco plants. Among early signalling events triggered by  
41 this microbial-associated molecular pattern is a transient apoplastic oxidative burst which is  
42 dependent on the NADPH oxidase activity of the RESPIRATORY BURST OXIDASE  
43 HOMOLOG (RBOH) isoform D. Using radioactive [<sup>33</sup>P]-orthophosphate labelling of tobacco  
44 Bright Yellow-2 suspension cells, we here provide *in vivo* evidence for a rapid accumulation  
45 of phosphatidic acid (PA) in response to cryptogein due to the coordinated onset of  
46 phosphoinositide-dependent phospholipase C and diacylglycerol kinase (DGK) activities.  
47 Both enzyme specific inhibitors and silencing of the phylogenetic cluster III of the tobacco  
48 DGK family were found to reduce PA production upon elicitation and to strongly decrease the  
49 RBOHD-mediated oxidative burst. Therefore, it appears that PA originating from DGK  
50 controls NADPH-oxidase activity. Amongst cluster III DGKs, the expression of *DGK5-like*  
51 was up-regulated in response to cryptogein. Besides *DGK5-like* is likely to be the main cluster  
52 III DGK isoform silenced in one of our mutant line, making it a strong candidate for the  
53 observed response to cryptogein. The relevance of these results is discussed with regard to  
54 early signalling lipid-mediated events in plant immunity.

55

56 **KEYWORDS:** cryptogein, microbial-associated molecular pattern, diacylglycerol kinase;  
57 phospholipase C, phosphatidic acid, reactive oxygen species, RBOHD, NADPH oxidase

58

59

## 60 INTRODUCTION

61 Many studies have suggested that membrane lipids act as reservoirs for signalling molecules  
62 in response to developmental and environmental cues in both animal and plant cells. Indeed,  
63 under biotic or abiotic stresses, a regulated production of phospholipid-derived molecules was  
64 reported to occur, and to launch intracellular signalling cascades. Phospholipases, lipid  
65 kinases and lipid-phosphate phosphatases are considered as the main activities involved in  
66 such early signalling events (Janda et al. 2013, Sheng et al. 2015, Hawkins & Stephens 2015,  
67 Ruelland et al. 2015).

68 Phosphatidic acid (PA) is the most basic phosphoglycerolipids, being composed of a  
69 diacylglycerol with a phosphoryl group esterified on the *sn*-3 hydroxyl of the glycerol  
70 backbone. Besides being an intermediate of phospholipid synthesis (Petroustos et al. 2014),  
71 PA also transduces stress signals in eukaryotic cells. In serum-withdrawn fibroblast cultures  
72 for example, reverse genetic approaches have established a marked correlation between  
73 elevated endogenous PA levels and apoptosis inhibition; the latter probably being partially  
74 responsible for cancer establishment in tissues (Bruntz et al. 2014). In plants, upon pathogen  
75 attack and situations mimicking infection by the addition of elicitors, as well as under many  
76 abiotic (including drought, cold, osmotic) stresses, PA accumulation was either inferred from  
77 pharmacological strategies or directly shown by biochemical methods (Pinosa et al. 2013,  
78 Ruelland et al. 2015). In Eukaryotic cells, the synthesis of PA as a signalling molecule relies  
79 on two distinct enzymatic systems. Phospholipases D (PLD) can hydrolyse structural  
80 phospholipids within membranes, directly releasing PA (Wang et al. 2012) while,  
81 diacylglycerol kinases (DGK) can catalyse the phosphorylation of diacylglycerol (DAG) to  
82 produce PA (Arisz et al. 2009). DAG can be provided to DGK either by phosphoinositide-  
83 dependent phospholipases C (PI-PLCs) that act on phosphorylated phosphoinositides or by  
84 non-specific phospholipases C (NPCs) that use structural phospholipids (Pokotylo et al. 2013,

85 2014). In contrast to animal systems where both DAG and PA have been shown to act as  
86 secondary messengers, it is commonly assumed that PA plays the central role in plant cells.  
87 Indeed, no plant proteins orthologous to mammalian protein kinases C (which are activated  
88 through a direct interaction with DAG) have been identified from plant genomes (Dong et al.  
89 2012), even though some proteins bearing the C1 domain responsible for DAG binding to  
90 PKCs have been found (Janda et al. 2013). On the other hand, up to 35 plant proteins targeted  
91 by PA have been identified on the basis of their direct interaction with the lipid (Hou et al.  
92 2015). However, no consensus sequence to define a canonical PA binding domain was  
93 identified from these proteins.

94 Upon pathogen attack, two lines of defence can successively occur. The first relies on the  
95 recognition of common pathogen motifs, named *microbial-associated molecular patterns*  
96 (MAMPs), at the plasma membrane and this can result in basal resistance to a broad range of  
97 pathogens. The second depends on the specific perception of pathogen effectors within host  
98 cells by resistance proteins (Dangl & Jones 2001, Jones & Dangl 2006, Stotz et al. 2014,  
99 Bigeard et al. 2015). This latter defence response often leads to a rapid and highly localized  
100 hypersensitive cell death that prevents biotrophic pathogens from propagating (Cacas 2015).  
101 Interestingly, PA has been shown to accumulate in response to many MAMPs (xylanase,  
102 flagellin, N-acetylchitooligosaccharide, chitosan) in several independent plant models (van  
103 der Luit et al. 2000, den Hartog 2003, Bargmann et al. 2006). PLD-, PI-PLC-, NPC- and  
104 DGK-encoding genes and/or corresponding enzymatic activities were also reported to be  
105 rapidly up-regulated in numerous studies dealing with either MAMP- or Effector-Triggered  
106 Immunity (MTI or ETI, respectively), suggesting the involvement of these lipid-modifying  
107 enzymes in early defence events (Zhang & Xiao 2015, Ruelland et al. 2015). With the notable  
108 exception of DGK, this assumption was corroborated by several works using loss-of-function  
109 mutants. While *Arabidopsis* PLD $\delta$  isoform was found to be involved in MTI, multiple PLD

110 isoforms were proposed to act redundantly in ETI (Pinosa et al. 2013, Johansson et al. 2014).  
111 In tomato (*Solanum lycopersicum*) cell suspensions, MTI involved PLDB1 (Bargmann et al.  
112 2006) whereas full establishment of ETI required the PI-PLC-encoding gene *sIPLC4* (Vossen  
113 et al. 2010).

114 The small, basic and hydrophilic 10-kDa protein named cryptogein is a MAMP secreted by  
115 the oomycete *Phytophthora cryptogea* (Ricci et al. 1989). More than two decades after its  
116 discovery, extensive investigations have provided a broad picture of cryptogein-induced  
117 physiological responses. Although this protein exhibits both cell death-promoting and  
118 systemic acquired resistance-inducing activities in tobacco (*Nicotiana tabacum*) plants (Keller  
119 et al. 1996, Rustérucchi et al. 1999, Cacas et al. 2005), it is also able to activate many  
120 characteristic hallmarks of MTI. The interaction of cryptogein with a yet-to-be-identified  
121 protein receptor at the plasma membrane results in calcium influx (Tavernier et al. 1995,  
122 Pugin et al. 1997), potassium and chloride effluxes associated with apoplasm alkalization  
123 (Blein et al. 1991), NADPH oxidase-dependent oxidative burst (Simon-Plas et al. 2002),  
124 modification of plasma membrane order (Gerbeau-Pissot et al. 2014), activation of a MAPK  
125 cascade (Dahan et al. 2009) and clathrin-dependent endocytosis (Leborgne-Castel et al. 2008).

126 Reactive oxygen species (ROS) are assumed to be the main downstream actors to PA in  
127 pathogen-induced signalling cascades (Zhang & Xiao 2015). Although it was demonstrated  
128 that PLD-produced PA directly regulates the NADPH oxidase-dependent oxidative burst  
129 during a drought stress (Zhang *et al.* 2009), this link has not been clearly established with  
130 respect to pathogen-response. Firstly, is PA production a common signature of MTI,  
131 irrespective of the MAMP and the cell model? Secondly, can PA produced by DGK activities  
132 act upstream of the NADPH oxidase during plant-pathogen interactions? In this present work,  
133 we report on the regulatory function of DGK in controlling the cryptogein-induced oxidative  
134 burst in Bright Yellow-2 (BY-2) cell suspensions. Using biochemical, pharmacological and



135 reverse genetic approaches, we show that PA, originating from a DGK coupled to PI-PLC, is  
136 an upstream positive regulator of the plasma membrane-localized NADPH oxidase isoform  
137 RBOHD in response to cryptogein.

138

## 139 MATERIALS & METHODS

### 140 Culture conditions and cryptogein elicitation of tobacco cells

141 Bright Yellow-2 (*Nicotiana tabacum*) cells were grown in modified liquid Murashige and  
142 Skoog (MS, M0221, Duchefa-Kalys, Bernin, France) medium at pH 5.6 (M0221, Duchefa-  
143 Kalys, Bernin, France) supplemented with 1 mg.L<sup>-1</sup> thiamine-HCl, 0.2 mg.L<sup>-1</sup> 2,4  
144 dichlorophenylacetic acid, 100 mg.L<sup>-1</sup> myo-inositol, 30 g.L<sup>-1</sup> sucrose, 200 mg.L<sup>-1</sup> KH<sub>2</sub>PO<sub>4</sub> and  
145 2 g.L<sup>-1</sup> MES. Cells were maintained under continuous light (200 μE m<sup>-2</sup> s<sup>-1</sup>) on a rotary shaker  
146 (140 rpm) and sub-cultured to fresh medium (4:80, (v/v)) on a weekly basis.

147 For radiolabeling experiments, ROS quantification and pharmacological treatments, cells  
148 were harvested from 7-day-old suspensions using a vacuum-system and resuspended at  
149 0.1g/mL in either equilibration medium I2 (2 mM MES, 175 mM mannitol, 0.5 mM CaCl<sub>2</sub>,  
150 0.5 mM K<sub>2</sub>SO<sub>4</sub>, pH 5.9) or I20 (20 mM MES, 175 mM mannitol, 0.5 mM CaCl<sub>2</sub>, 0.5 mM  
151 K<sub>2</sub>SO<sub>4</sub>, pH 5.9). After a 3 h incubation period under mild shaking at 24°C, cells were elicited  
152 by adding 50 nM cryptogein (final concentration) from a solution prepared according to  
153 (Ricci et al. 1989).

### 154 Radiolabelling experiments

155 BY-2 cells were aliquoted (7 mL) in wide neck flasks (capacity 50 mL) for equilibration.  
156 Cells were then labelled by the addition of 53 MBq.L<sup>-1</sup> [<sup>33</sup>P]-orthophosphate. Total lipids  
157 were extracted according to the procedure previously described by (Krinke et al. 2009). Lipids  
158 were separated by thin layer chromatography (TLC) and developed either in an acidic solvent  
159 system composed of chloroform:acetone:acetic acid:methanol:water (10:4:2:2:1, (v/v),  
160 Lepage 1967) or in a solvent system composed of an upper phase of ethyl  
161 acetate:isooctane:acetic acid: water (12:2:3:10 (v/v), Munnik et al. 1995). Radiolabelled spots

162 were quantified by autoradiography using a Storm phosphorimager (Amersham Biosciences,  
163 Buckinghamshire, UK). Separated phospholipids were identified by co-migration with  
164 authentic non-labelled standards visualized by primuline staining (under UV light) or by  
165 phosphate staining.

#### 166 **Quantification of reactive oxygen species**

167 Reactive oxygen species were measured using a luminol-based method as described  
168 previously (Simon-Plas et al. 2002) but with slight modifications. Briefly, after a 3 h-  
169 equilibration period in I20 medium, cells were treated with cryptogein. Before elicitation,  
170 samples corresponding to zero time were harvested and the luminescence background of the  
171 assay was measured. Samples (250  $\mu$ L) were regularly harvested over the first 90 min for  
172 ROS determination using a luminometer (Lumat LB9507, Berthold Technologies, Bad  
173 Wilbad, Germany). Fifty microliters of a 0.3 mM luminol solution and 300  $\mu$ L of the assay  
174 buffer (50 mM MES, 175 mM mannitol, 0.5 mM CaCl<sub>2</sub>, 0.5 mM K<sub>2</sub>SO<sub>4</sub>, pH 6.5) were  
175 automatically added to cell aliquots before luminescence was measured (delay in injection of  
176 1.2 s, data acquisition time of 10 s). For each biological and technical replicates, 2 h after  
177 elicitation (when the oxidative burst was over), a calibration curve was made in order to  
178 convert relative luminescence units (RLU) into H<sub>2</sub>O<sub>2</sub> equivalents by recording cell  
179 luminescence upon the addition of increasing H<sub>2</sub>O<sub>2</sub> concentrations during the assay. Results  
180 were normalized to BY-2-cell weight and expressed as nmoles H<sub>2</sub>O<sub>2</sub> equivalents per gram of  
181 cells.

#### 182 **Reverse transcription-quantitative polymerase chain reaction**

183 RNA extraction, reverse transcription and quantitative measurement of gene expression by Q-  
184 PCR were carried out as described by (Anca et al. 2014). Two reference genes were used for  
185 expression normalization: *ELONGATION FACTOR 1 $\alpha$*  (EF-1 $\alpha$ , AF120093), forward, 5'-

186 TGAGATGCACCACGAAGCTC-3', and reverse, 5'-CCAACATTGTCACCAGGAAGTG-  
187 3'; and *L25* gene coding for a ribosomal subunit (L18908), forward, 5'-  
188 CCCCTCACCACAGAGTCTGC-3', and reverse, 5'-  
189 AAGGGTGTGTTGTTGTCCTCAATCTT-3'. Primer sequences for *NtDGK5-like* gene are as  
190 follows: *NtDGK5-like* specific couple: forward (F85sq), 5'-  
191 AGTCCGAGCTCAATGACAACA-3'; and reverse (R85sq2), 5'-  
192 GTACGAAGAAGTTCTCCTCCAAGTT-3'; Cluster III gene-targeting couple: forward  
193 (ClusterIII-F), 5'-TTCAGCATGGGGATGGATGCA-3'; and reverse (ClusterIII-R), 5'-  
194 AACCATCCTTGYGTACATCC-3'

#### 195 **Cloning of *NtDGK5-like* coding sequence**

196 *NtDGK5-like* coding sequence was amplified by PCR from cDNA obtained from untreated 7-  
197 day-old BY-2 cells using high fidelity DNA polymerase (Phire Hot Start, ThermoFisher  
198 SCIENTIFIC) and specific primers: forward (F1c85), 5'-ATGGCGAATTCTGAGTCCGA-  
199 3'; and reverse (R1c85), 5'-CTAACTGAGACGAGAAACATCGA-3'. Primers were  
200 designed according to the sequence of SGN-U439985 (Solanacea Genomic Network  
201 database). Resulting PCR products were cloned into pSC-B Amp/Kan vector using the  
202 topoisomerase-based StrataClone Blunt PCR cloning kit (Agilent). Seven clones were  
203 sequenced among which one was selected for further work. The sequence was deposited in  
204 GenBank (accession number KU934207). This cDNA was named *NtDGK5-like* because its  
205 closest *Arabidopsis thaliana* ortholog is AtDGK5.

#### 206 **Production of overexpressing *NtDGK5-like* transgenic lines**

207 The *NtDGK5-like* coding sequence was first amplified by PCR using high fidelity DNA  
208 polymerase from *pSC-B::DGK5-like* using the following primer pairs: couple 1, forward  
209 (CACC-F85), 5'-CACCATGGCGAATTCTGAGTCCGA-3'; and the reverse R1c85; couple

210 2: the forward (CACC-F85) and reverse (R85-c-myc), 5'-  
211 CTAGAGGTCTTCTTCGCTGATGAGCTTTTGTTCCTGAGACGAGAAACATCGAC-  
212 3'. Two independent intermediate constructs to express a C-terminally c-myc tagged protein  
213 and an untagged version were generated by cloning the PCR products into pENTR/D-TOPO  
214 (ThermoFisher SCIENTIFIC). Sequencing of both inserts revealed no mutations. Using  
215 Gateway LR Clonase II enzyme mix (ThermoFisher SCIENTIFIC), MluI-digested  
216 *pENTR/D::DGK5-like-c-myc* and MluI-digested *pENTR/D::DGK5-like* were then  
217 independently recombined with pMDC32 (Karimi et al. 2007) to generate plant-expressing  
218 vectors that were checked for correct recombination by restriction digestions before being  
219 used to transform WT BY-2 cell suspensions following a previously described procedure  
220 (Simon-Plas *et al.* 2002). Among the 6 hygromycin-resistant calli isolated, only one  
221 overexpressed the untagged version of *NtDGK5-like* (named *line OE.NtDGK5-A*) while five  
222 out of twenty hygromycin-resistant calli overexpressed the C-terminally tagged version of  
223 *NtDGK5-like*. The two lines, named *OE.NtDGK5myc-A* and *OE.NtDGK5myc-B*, which  
224 exhibited the strongest levels of transgene expression, were selected for further  
225 characterization.

### 226 **Generation of cluster III DGK-silenced cell lines**

227 A conserved region of cluster III DGKs of the *NtDGK5-like* coding sequence, from  
228 nucleotides 511 to 692 (see supporting information Fig. S2) was chosen to be inserted into the  
229 post-transcriptional gene silencing vector (pH7GWIWG2(II), Karimi et al. 2007). This  
230 fragment was PCR-amplified using high fidelity DNA polymerase and the following specific  
231 primers: forward (Fc-ClusterIII), 5'-CACCTCATTTTTGAAGCAAGTAATGAATGCA-3',  
232 and reverse (Rc-ClusterIII), 5'-TGGAACCTTCCACATTCAGTTCA-3'. The resulting PCR  
233 product was directionally cloned into pENTR/D-TOPO and the *pENTR::DGK5-like* fragment  
234 was sequenced before the plasmid was recombined with pH7GWIWG2(II). The final

235 construct was verified by restriction digestion and the fragment-intron loop-fragment was also  
236 sequenced. To check that the construct was functional, a transient tobacco expression system  
237 was designed (explained hereafter). Wild-type (WT) BY-2 cells were transformed as  
238 previously described (Simon-Plas et al. 2002). Among the 30 hygromycin-resistant calli  
239 isolated, 5 were screened and 2 independent transgenic lines (named *hp-clusterIII-A* and *hp-*  
240 *clusterIII-B* lines) silenced for *NtDGK5-like* were identified and selected for further  
241 characterization.

#### 242 **Transient expression system in tobacco plants**

243 To test for functional miRNA constructs, *Agrobacterium tumefaciens* (GV3101) were  
244 transformed with the Cluster III-targeting vector and selected clones were grown overnight  
245 under mild shaking at 28°C in regular Luria-Bertani medium containing 50 µg.mL<sup>-1</sup>  
246 rifampicin, 50 µg.mL<sup>-1</sup> gentamycin and 50 µg.mL<sup>-1</sup> spectinomycin. Bacteria were collected  
247 the following day by centrifugation and the pellets were resuspended in induction buffer (20  
248 mM MES pH 5.5, 10 mM MgSO<sub>4</sub>, 200 µM acetosyringone) so that the OD<sub>600 nm</sub> was 0.5-0.6.  
249 After incubation at room temperature for 3 h, the bacterial suspension was infiltrated into the  
250 abaxial side of detached tobacco leaves (*Nicotiana tabaccum*, 4 to 6 week-old plants) using a  
251 needleless syringe. Samples for RT-PCR were harvested 2 days-post inoculations. Each  
252 experiment included non-infiltrated and infiltrated leaf material with bacterial clones  
253 containing the empty vector as controls. Stem-loop reverse transcription reactions to show  
254 specific pri-miRNA was performed on 2 independent biological replicates according to (Chen  
255 2005) using the following primer: 5'-  
256 GTCGTATCCAGTGCAGGGTCCGAGGTATTCGCACTGGATACGACTggaaac-3'; the  
257 underscored sequence corresponding to the gene specific part of the primer whereas the other  
258 part corresponds to the stem-loop backbone of the microRNA *miR172* (Martin et al. 2009).  
259 Stem-loop RT controls were carried out using a 5S-rRNA-specific oligonucleotide 5'-

260 GTCGTATCCAGTGCAGGGTCCGAGGTATTCGCACTGGATACGACgcaaca-3'. The  
261 generated cDNA matrix was amplified by regular PCR with a gene specific-forward primer  
262 and a *miR172* reverse primer (5'-GGGTCGACATCAAGTCATCAATTTGCCA-3'). For the  
263 5S-rRNA template, the forward primer was as follows: 5S-rRNA-F, 5'-  
264 GGATGCGATCATAACCAGCACT-3'; for the Cluster III-targeting miRNA, the primer used  
265 for generating the construct (Fc-ClusterIII, described above) was used. PCR products were  
266 then sequenced and proved to correspond to Cluster III pri-miRNA.

### 267 **Western blot analysis**

268 Seven-day old cells from the *OE.NtDGK5myc-A* and *OE.NtDGK5myc-B* lines and the  
269 corresponding empty vector line were harvested by filtration and protein extracts (cytosol,  
270 microsomes and plasma membrane) were prepared as described by (Mongrand et al. 2004).  
271 Proteins were solubilized in a buffer containing 40 mM Tris-HCl pH 6.8, 5%  $\beta$ -  
272 mercaptoethanol, 1.5% SDS, 1 mM EDTA, 2M urea, 1M thiourea, 1% n-octyl-glucoside,  
273 10% glycerol and bromophenol blue for 2h at room temperature, separated by SDS-PAGE  
274 (8% (w/v) acrylamide-bisacrylamide) and transferred onto nitrocellulose membrane (Hybond  
275 ECL; GE Healthcare, Chalfont St Giles, UK). The membrane was blocked overnight in a 5%  
276 (w/v) milk solution and probed with an anti-c-myc antibody directly coupled to horseradish  
277 peroxidase (GenScript, Piscataway, New Jersey, USA). Specific signals were revealed using  
278 an ECL kit (Perkin Elmer, Waltham, MA, USA) according to the manufacturer's instructions.

### 279 **Assessment of cell suspension density**

280 The packed cell volume (PCV) and dry weight (DW) were used as proxies of cell density.  
281 PCV was determined upon centrifugation of 15mL cell suspension samples (700 x g, 3 min).  
282 It represents the ratio of pelleted cell volume to the total sample volume, expressed in percent  
283 (Cacas et al. 2005). Dry weight was measured upon drying 12mL cell suspension aliquots at

284 55°C for 3 days; a condition that allows the culture medium to fully evaporate. It is expressed  
285 in percent with reference to WT BY-2 conditions set to 100.

286 **Phylogenetic analysis**

287 Sequence alignment and phylogenetic tree building were carried out as described previously  
288 (Cacas et al. 2011).

289



## 290 RESULTS

### 291 The MAMP cryptogein triggers rapid accumulation of phosphatidic acid in tobacco BY- 292 2 cell suspensions

293 The potential production of PA following elicitation of BY-2 tobacco cell suspensions with  
294 cryptogein was investigated by treating cells with the peptide and extracting lipids for up to  
295 one hour after elicitor addition. Cells were incubated with [<sup>33</sup>Pi]-orthophosphate for 20 min  
296 before lipid extraction and [<sup>33</sup>P]-labelled PA levels were measured after lipid separation by  
297 thin layer chromatography (TLC). Marked changes in labelled PA levels were observed after  
298 the addition of cryptogein (Fig. 1a) and quantification of the signal showed that after a 3 min  
299 lag, radiolabelled PA increased and reached a maximum within the first 10 min and then  
300 plateaued (Fig. 1b).

### 301 PLC and DGK inhibitors reduce the accumulation of phosphatidic acid in response to 302 cryptogein

303 Under our experimental conditions, i.e. short-time radioisotopic labelling, the main pool of  
304 radiolabel is incorporated into ATP whereas structural phospholipids are only weakly  
305 labelled. This experimental design is commonly used to detect *in vivo* activity of lipid kinases,  
306 including DGKs, because these enzymes use the  $\gamma$ -phosphate group of ATP as their substrate.  
307 Hence, it could be postulated from our data that, in response to cryptogein, a phospholipase C  
308 activity produced DAG that was then processed by DGK into PA. These two hypotheses were  
309 tested by a pharmacological approach relying on the use of edelfosine, an inhibitor of  
310 phosphoinositide-dependent PLC (PI-PLC; Wong *et al.* 2007; Kelm *et al.* 2010; Djafi *et al.*  
311 2013) and R59022, an inhibitor of DGK (Gómez-Merino *et al.* 2004). Wild-type cell  
312 suspensions were preincubated with inhibitors and radioactivity incorporated into PA was  
313 quantified 10 min after cryptogein addition, i.e. when PA levels are maximal. Edelfosine was

314 found to significantly alter cryptogein-induced PA accumulation in a dose-dependent manner,  
315 leading to a maximum inhibition of radioactive PA production of around 70% (Fig. 2a).  
316 Furthermore, 75  $\mu$ M R59022 provoked a similar decrease of radioactive PA accumulation  
317 (Fig. 2b).

318 It was shown in *Arabidopsis thaliana* that phosphoinositides, substrates of PI-PLCs, are  
319 provided by type III-phosphatidylinositol-4 kinases (PI4K) that are sensitive to micromolar  
320 concentrations of wortmannin (Delage et al. 2012, Djafi et al. 2013). Pre-treatment of BY-2  
321 cells with 30  $\mu$ M wortmannin resulted in a 46% inhibition of cryptogein-induced PA  
322 accumulation whereas 1  $\mu$ M of the inhibitor, a concentration that represses  
323 phosphatidylinositol-3 kinase activity (Krinke et al. 2007) but not that of PI4K, did not exhibit  
324 any effect (Fig. 2c). Furthermore, since PI-PLC activity is highly dependent on calcium (Hunt  
325 et al. 2004, Pokotylo et al. 2014), lanthanides that can act as inhibitors of plasma membrane  
326 calcium channels (Tester 1990, Knight et al. 1997) were tested for their ability to impact PA  
327 production upon elicitation. As expected, increasing concentrations of  $\text{La}^{3+}$  led to decreasing  
328 PA accumulation in response to cryptogein (Fig. 2d).

329 Taken together, these findings clearly point to a role for a PI-PLC/DGK pathway in  
330 cryptogein-induced PA production in BY-2 tobacco cell cultures.

### 331 **PLC and DGK inhibitors diminish RBOHD-mediated oxidative burst intensity in** 332 **response to cryptogein**

333 Among early signalling events triggered by cryptogein in BY-2 cells is the RBOHD-mediated  
334 oxidative burst which takes place within the first 10-15 min following elicitation and is,  
335 therefore, concomitant to the onset of PA production. To examine the hypothesis that a PI-  
336 PLC/DGK pathway could control the oxidative burst, a pharmacological approach was  
337 undertaken. The exogenous application of the PI-PLC inhibitor edelfosine was found to

338 drastically alter the cryptogein-induced oxidative burst (Fig. 3a), resulting in a 70% decrease  
339 of the total amount of ROS when compared to untreated WT cells (Fig. 3b). Similarly,  
340 treatment with the DGK inhibitor R59022 also promoted a significant decrease in ROS levels  
341 (Fig. 3c,d). Interestingly, neither PI-PLC nor DGK inhibitors led to the inhibition of  
342 extracellular pH alkalization, another early signalling event associated with cryptogein  
343 elicitation (Supporting information Fig. S3). As the results of both PA production and ROS  
344 burst supported a working scenario where a PI-PLC/DGK pathway controls the cryptogein-  
345 induced signalling cascade, a reverse genetic strategy was developed to identify DGK  
346 candidates involved in the regulation of RBOHD.

#### 347 **Phylogenetic clustering of the tobacco diacylglycerol kinase family**

348 Tobacco DGK nucleotide sequences were retrieved by both a keywords and a sequence  
349 homology search with coding sequences of the seven identified *Arabidopsis thaliana* DGK  
350 genes and those published for rice (*Oryza sativa*) and tomato (*Solanum lycopersicum*)  
351 (Snedden & Blumwald 2000, Zhang et al. 2008) using the BLAST algorithm of the *Solanacea*  
352 *Genomics Network* (SGN) server. No unigene could be identified, but 16 partial contigs were  
353 isolated (Supporting information Fig. S4). Manual EST and contig assembly led to the  
354 identification of two potential full-length mRNAs corresponding to the accessions SGN-  
355 U440091 and SGN-U439985. A phylogenetic tree was built using the longest *in silico*  
356 translated tobacco primary amino acid sequences, including SGN-U440091, SGN-U439985  
357 and 7 additional partial coding sequences, in addition to Arabidopsis, rice and tomato  
358 sequences (Fig. 4a).

359 DGKs are bipartite proteins composed of one catalytic domain and one accessory domain,  
360 assumed to be necessary for the kinase activity. While the animal DGK family can be  
361 subdivided into 5 classes (Goto & Kondo 2004), plant DGKs only form three phylogenetic

362 clusters characterized by distinct protein structures (Fig. 4a). Cluster I encompasses the  
363 closest sequence orthologous of metazoan enzymes with proteins that display in their N-  
364 terminus one or two DAG-binding domain(s) (noted C1) and either one or no transmembrane  
365 helix (Vaultier et al. 2008). Contrary to Cluster II DGKs that have a minimal DGK structure,  
366 cluster III DGKs can exhibit a C-terminal calmodulin binding region (referred to as CBD)  
367 depending on splicing variants (Arisz et al. 2009). SGN-U440091 and SGN-U439985 appear  
368 to belong to clusters II and III, respectively. Furthermore, amongst the 7 partial tobacco DGK  
369 coding sequences that were retrieved, four are present in cluster I, one in cluster II and two in  
370 cluster III.

371 **Overexpressing a member of the phylogenetic cluster III, *NtDGK5-like*, does not result**  
372 **in a gain-of-function**

373 Among tobacco cluster III members, the full-length cDNA corresponding to the unigene  
374 SGN-U439985 (hereafter referred to as *NtDGK5-like* after its closest *Arabidopsis thaliana*  
375 ortholog) was cloned and deposited in GenBank (Accession number KU934207). The isolated  
376 *NtDGK5-like* cDNA corresponds to a 1482 bp-long coding sequence that encodes a 493-  
377 amino acid protein displaying a molecular weight of 55.24 kDa and a theoretical pI of 6.75,  
378 with no predicted signal peptide. The *NtDGK5-like* gene is constitutively expressed in BY-2  
379 cells but its expression is significantly up-regulated in response to cryptogein (Fig. 4b). Using  
380 transgenic lines overexpressing a C-terminal c-myc-tagged NtDGK5-like protein (named  
381 *OE.NtDGK5myc-A* and *OE.NtDGK5myc-B* lines), a cell fractioning approach was developed  
382 to identify where the protein was located. Cytosolic, microsomal and plasma membrane  
383 fractions were isolated and their proteins analysed by western blotting. The tagged protein  
384 was found in both the endomembrane and plasma membrane fractions (Fig. 4c).

385 To study the role of *NtDGK5-like*, the two *OE.NtDGK5myc-A* and B lines were analysed  
386 further along with a third line (named *OE.NtDGK5-A*) allowing the expression of an untagged  
387 *NtDGK5-like* protein under the control of the CaMV35S promoter. Despite a constitutive 10-  
388 to-15-fold increase in steady-state levels of *NtDGK5-like* transcripts with respect to  
389 endogenous gene expression (Fig. 5a), all three overexpressor lines accumulated PA levels  
390 comparable to those of WT BY-2 and an empty vector-containing cell line in response to  
391 cryptogein (Fig. 5b). Accordingly, no differences in ROS production could be seen between  
392 overexpressing lines and WT or empty vector-control genotypes (Fig. 5c), with the notable  
393 exception of the *OE.NtDGK5myc-B* line. This line displays a significant difference with the  
394 WT line, but not with the empty vector control line. Because cryptogein-induced oxidative  
395 burst intensity was assessed on 7-day-old cell suspensions in our experimental system, the  
396 impact of overexpressing *NtDGK5-like* on cell density was estimated at this time-point.  
397 Packed cell volume (PCV) and dry weight (DW) of overexpressing lines were measured as  
398 two independent proxies of biomass and compared to those of WT cell culture. Only the  
399 *OE.NtDGK5myc-B* line had a slight defect in cell size (Fig. 5d) and this could explain the  
400 small but significant decrease in ROS levels produced upon elicitation.

#### 401 **Silencing the phylogenetic cluster III DGK family of tobacco strongly inhibits** 402 **cryptogein-induced oxidative burst**

403 In an attempt to specifically silence *NtDGK5-like* gene expression, a miRNA transgenic cell  
404 line strategy was developed but it was unsuccessful due to the early death of transformed  
405 calli. However, to get insights into the function of cluster III DGKs, a construct targeting a  
406 conserved DNA region of cluster III members was engineered. A 180 bp-long portion of the  
407 *NtDGK5-like* coding sequence was cloned into the hairpin vector pH7GWIWIG2(II) (Fig. 6a).  
408 Using an *Agrobacterium*-mediated transient expression system, this construct was shown to  
409 produce DGK specific miRNA *in planta* (Fig. 6b) and it was therefore used to generate stable

410 transgenic cell lines in the WT BY-2 genetic background. Amongst the 30 transformed clones  
411 obtained, two independent lines (named *hp-clusterIII-A* and *hp-clusterIII-B* lines) were  
412 identified and selected on the basis of their reduced constitutive expression of *NtDGK5-like*.  
413 While an empty vector-containing line displayed slightly higher levels of expression  
414 compared to the WT cell line, miRNA cell lines exhibited similar lower steady state-levels of  
415 *NtDGK5-like* transcripts, averaging 30 to 40% of that of WT cells (Fig. 6c). To test whether  
416 the expression of other cluster III DGK genes were affected in the two transgenic lines, Q-  
417 PCR was carried out using primers that allowed to simultaneously amplify the coding  
418 sequences of each cluster III genes. In this way, it was found that the bulk expression of  
419 cluster III genes was not altered in *hp-clusterIII-A* line whereas it was silenced approximately  
420 3-fold in line *hp-clusterIII-B* line (Fig. 6c). A 3-fold down-regulation of *NtDGK5-like* is  
421 probably not enough to induce changes in the bulk expression of cluster III genes (see *hp-*  
422 *clusterIII-A* cell line). This implies that that *NtDGK5-like* is not the only cluster III isoform to  
423 be silenced in the *hp-clusterIII-B* line. On the contrary, this gene is likely to be the main  
424 cluster III-DGK isoform to be silenced in the *hp-clusterIII-A* cell line *NtDGK5-like*.

425 The impact of silencing DGK cluster III on cell density was estimated. No evidence for  
426 statistical differences between the four analysed genotypes (BY-2, empty vector line, *hp-*  
427 *clusterIII-A* and *hp-clusterIII-B* lines) on PCV and DW at day 7 after sub-culturing was found  
428 (Fig. 6d). Hence, potential effects of cluster III silencing on both PA and ROS production was  
429 determined upon cryptogin elicitation. The *DGK*-silenced lines were shown to no longer  
430 over-accumulate PA, 10 min after cryptogin addition (Fig. 7a). Remarkably, the absence of  
431 early PA accumulation in these elicited transgenics could be correlated with a marked  
432 decrease in oxidative burst intensity. Wild-type and empty-vector lines showed a similar  
433 stereotyped behaviour characterized by a rapid and transient peak of ROS, being maximum  
434 within 10-15 min post-treatment before decreasing and stabilizing for the last 45min, whereas

435 ROS profiles of the two *hp-clusterIII*- lines were devoid of a well-defined peak, exhibiting  
436 rather weak and stable ROS levels over 90 min (Fig. 7b). Indeed, total levels of extracellular  
437 ROS generated by the miRNA lines accounted for approximately one third to one half of  
438 those recorded in WT and empty vector-containing lines upon cryptogein application (Fig.  
439 7c).

## 440 DISCUSSION

441 In the present work, radio-labelling experiments clearly showed that *in vivo* levels of PA  
442 rapidly increased when BY-2 cell suspensions were challenged with cryptogein, reaching a  
443 maximum after 10 min and then staying stable during 1 h. Under our experimental conditions,  
444 [<sup>33</sup>Pi]-orthophosphate was added to cell cultures 20 min before lipid extraction. In contrast to  
445 long time periods of isotope incubation that lead to labelled structural phospholipids, short  
446 time labelling results in the incorporation of isotopic phosphorus into ATP. Hence, this setup  
447 is optimized to detect products of lipid kinases using ATP as substrate (Vaultier et al. 2006)  
448 and being active within the duration of labelling. These kinase activities include DGK and  
449 others, such as PI4-kinase and PI4P-5-kinase. Indeed, the radioactivity incorporated into PI4P  
450 is typically 4- to 5-fold that of PI, in control cells, although PIP (not labelled) is much less  
451 abundant than PI (Furt et al. 2010). However, this experimental design is not optimal for  
452 detecting the possible occurrence of PLD activity. The radioactivity associated with PA upon  
453 cryptogein treatment represented 3- to 4-fold of that monitored in phosphatidylcholine,  
454 phosphatidylethanolamine and phosphatidylglycerol, putative substrates of PLDs. This  
455 strongly supports the idea that radiolabelled PA found after a 20 min labelling cannot arise  
456 from PLD action. Yet, this does not mean a PLD is not activated upon cryptogein addition. A  
457 long time labelling with [<sup>33</sup>Pi]-orthophosphate (16 hours) designed to reveal PLD activity was  
458 also carried out. In the presence of primary alcohol *n*-butanol, PLD catalyses a trans-  
459 phosphatidyl transfer reaction transferring the phosphatidyl moiety from structural lipids onto the  
460 hydroxyl group of the primary alcohol. In our hands, no radiolabeled phosphatidylbutanol  
461 accumulated in response to cryptogein (data not shown). However, since the signal-to-noise  
462 ratio was low in these experiments, we cannot rule out the possible activation of PLD by  
463 cryptogein.



464 The fact that the increase in radiolabelled PA in our experimental design was due to DGK was  
465 indicated by the pharmacological approach where DGK inhibitors led to an inhibition of  
466 radiolabelled PA accumulation. Moreover, the inhibiting effects of wortmannin and  
467 edelfosine strongly suggest that the DAG substrate of DGK was produced by a PI-PLC  
468 activity, the substrate of which is provided by a type III PI4K. In *Arabidopsis thaliana*, it has  
469 been shown that PI-PLC substrates are provided by such PI4Ks, either for basal PI-PLC  
470 activity (i.e. active in control cell, Djafi et al. 2013) or during PI-PLC activation by chilling  
471 (Delage et al. 2012). Therefore it seems that the role of type III PI4Ks to supply substrates to  
472 PI-PLC is likely to be the case for other plant species, including tobacco. Besides, PI-PLCs  
473 are strictly dependent on calcium and therefore it was not surprising that a blocker of  
474 plasmalemma calcium channels, lanthanides, inhibited radioactive PA accumulation.

475 Since inhibitors of different chemical nature acting on different enzymes (PI4K, PI-PLC,  
476 DGK) of a given pathway led to a similar inhibition of cryptogein-induced accumulation of  
477 radiolabelled PA, it is unlikely that this is due to hypothetical side effects of each inhibitor  
478 (Djafi et al. 2013, Ruelland et al. 2014). Moreover, silencing the phylogenetic cluster III of  
479 tobacco DGKs resulted in the loss of the cryptogein-induced PA increase thus demonstrating  
480 unambiguously the involvement of such enzymes in this response to cryptogein. Our  
481 knowledge concerning plant DGKs is still limited. Prokaryotic DGKs exhibit a peculiar  
482 multimeric membrane-embedded structure with three transmembrane spanning-regions per  
483 subunit (Van Horn & Sanders 2012). Metazoan counterparts display a conserved bidomain  
484 architecture associated with catalytic activity and decorated by a huge diversity of additional  
485 functional domains that are responsible for membrane or protein interactions and subcellular  
486 localization (Goto & Kondo 2004). Predictions of plant DGK structures using the Conserved  
487 Domain Architecture Retrieval Tool (Geer et al. 2002) reveal a simpler organization that falls  
488 into three evolutionary phyla (Vaultier et al. 2008). In this work, we focused our efforts on

489 DGKs of phylogenetic cluster III. Two members of this cluster have been either  
490 biochemically or functionally characterized. The rice DGK1 was stably expressed in tobacco  
491 plants and found to confer enhanced disease resistance to tobacco mosaic virus (TMV) and to  
492 the oomycete *Phytophthora infestans* (Zhang et al. 2008). Two distinct spliced mRNA  
493 variants were reported for the tomato LeDGK1/CBK1-encoded gene and this transcriptional  
494 regulatory mechanism was demonstrated to regulate protein docking at the plasma membrane  
495 (Snedden & Blumwald 2000) In addition, the *AtDGK5* gene expression was found to be  
496 upregulated in response to flagellin 22, a bacterial MAMP, as well as several avirulent  
497 pathogens (GENEVESTIGATOR database, Hruz et al. 2008).

498 In this work, one member of cluster III was cloned and named *NtDGK5-like* after its closest  
499 Arabidopsis ortholog. Independent transgenic cell lines were generated that allowed  
500 constitutive overexpression of either native *NtDGK5-like* (*OE.NtDGK5-A*) or a C-terminally  
501 c-myc tagged version of the protein (*OE.NtDGK5myc-A* and *OE.NtDGK5myc-B*). Despite the  
502 absence of any predicted signal peptide and transmembrane domain, western-blotting  
503 established that the protein was localized to the plasma membrane, as already reported for a  
504 *LeDGK1/CBK1* variant from tomato (Snedden & Blumwald 2000). In all *NtDGK5-like*  
505 overexpressing lines the accumulation of radioactive PA after 10 min of cryptogein addition  
506 was similar to that of control and WT lines. Thus it is tempting to speculate that DGK is not a  
507 limiting step in the coupled reaction leading to PA in response to cryptogein.

508 Previously, we had established that early cryptogein-induced ROS synthesis was fully  
509 dependent on the isoform D of the RBOH family in BY-2 cell suspensions (Simon-Plas et al.  
510 2002). In addition, Zhang et al. (2009) demonstrated that PA could directly interact with the  
511 Arabidopsis RBOHD isoform *via* an N-terminally-localized PA-binding domain and,  
512 subsequently, stimulate its activity. Alignment of the highly homologous tobacco RBOHD  
513 isoform with its Arabidopsis counterpart pinpointed that the location and sequence of the PA-

514 binding domain, as well as the basic nature of the two amino acid residues involved in the  
515 interaction with PA, are conserved between the two plant species (Supporting information Fig  
516 S5). Both PI-PLC and DGK inhibitors reduced the RBOHD-dependent oxidative burst.  
517 Furthermore, transgenic lines deficient for the tobacco cluster III DGKs displayed a  
518 significantly reduced oxidative burst upon cryptogein addition. Such findings strongly point  
519 to a direct regulation of RBOHD activity by PA, and place the PI-PLC/DGK pathway  
520 upstream from the enzyme in a signalling cascade induced by the MAMP cryptogein. It can  
521 be postulated that at least one DGK member of cluster III is responsible for the required DGK  
522 activity. Even though our genetic strategy concerning *NtDGK5-like* was partly unsuccessful,  
523 this gene still remains a good candidate since it was the only cluster III DGK gene suppressed  
524 in the *hp-clusterIII-A* cell line.

525 Distinct regulatory mechanisms have been described for plant RBOH proteins. These include  
526 post-translational modifications like phosphorylation (Kobayashi et al. 2007, Ogasawara et al.  
527 2008) and S-nitrosylation (Yun et al. 2011), association with RacGTPase (Wong et al. 2007b,  
528 Oda et al. 2010) as well as regulation by extracellular ATP level and calcium influx (Song et  
529 al. 2006, Demidchik et al. 2009). Zhang *et al.* (2009) also elegantly demonstrated that direct  
530 binding of PA activates AtRBOHD, *in fine* controlling stomatal aperture upon drought stress.  
531 Contrary to our model, where a PI-PLC/DGK pathway is involved, the authors showed that  
532 PLD $\alpha$ 1 was the main source of PA under their experimental conditions. In the context of plant  
533 defence, whether it is MTI or ETI, to date there is no genetic evidence for a role of DGKs in  
534 the positive regulation of RBOH enzymes, even though it has been suggested previously by  
535 several works (de Jong et al. 2004, Andersson et al. 2006, Vossen et al. 2010). Instead, many  
536 reports indicate PLDs as the main suppliers of PA in response to MAMP/pathogen effectors,  
537 irrespective of the downstream protein target (Young et al. 1996, van der Luit et al. 2000, de  
538 Torres Zabela et al. 2002, Bargmann et al. 2006, Laxalt et al. 2007, Krinke et al. 2009,

539 Yamaguchi et al. 2009, Kalachova et al. 2013, Zhao et al. 2013, Pinoso et al. 2013). Thus, our  
540 work sheds light on new insights into regulatory mechanisms that control early signalling  
541 events associated with NADPH oxidase since PA produced by a PI-PLC/DGK pathway can  
542 also control NADH oxidase activity. As underlined by (Zhang & Xiao 2015), this raises  
543 interesting questions as to how the spatiotemporal regulation of PA neo-synthesis and the  
544 engaged molecular species of PA could influence the specificity of downstream responses.

545

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555 experiments; FSP, SM, EJ, PGP, JLC & ER designed experiments and discussed results; JLC,  
556 PGP & ER wrote the manuscript; SM & FSP coordinated the project and retrieved ANR  
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559       **REFERENCES**

- 560    Anca I.A., Fromentin J., Bui Q.T., Mhiri C., Grandbastien M.A. & Simon-Plas F. (2014)  
561        Different tobacco retrotransposons are specifically modulated by the elicitor cryptogein  
562        and reactive oxygen species. *Journal of Plant Physiology* 171, 1533–1540.
- 563    Andersson M.X., Kourtchenko O., Dangl J.L., Mackey D. & Ellerström M. (2006)  
564        Phospholipase-dependent signalling during the AvrRpm1- and AvrRpt2-induced disease  
565        resistance responses in *Arabidopsis thaliana*. *The Plant Journal* 47, 947–959.
- 566    Arisz S.A., Testerink C. & Munnik T. (2009) Plant PA signaling via diacylglycerol kinase.  
567        *Biochimica et biophysica acta* 1791: 869–875.
- 568    Bargmann B.O.R., Laxalt A.M., Riet B.T., Schouten E., van Leeuwen W., Dekker H.L., ...,  
569        Munnik T. (2006) LePLDbeta1 activation and relocalization in suspension-cultured  
570        tomato cells treated with xylanase. *The Plant Journal* 45, 358–368.
- 571    Bigeard J., Colcombet J. & Hirt H. (2015) Signaling Mechanisms in Pattern-Triggered  
572        Immunity (PTI). *Molecular Plant* 8, 521–539.
- 573    Blein J.P., Milat M.L. & Ricci P. (1991) Responses of Cultured Tobacco Cells to Cryptogein,  
574        a Proteinaceous Elicitor from *Phytophthora cryptogea*: Possible Plasmalemma  
575        Involvement. *Plant Physiology* 95, 486–491.
- 576    Bruntz R.C., Taylor H.E., Lindsley C.W. & Brown H.A. (2014) Phospholipase D2 mediates  
577        survival signaling through direct regulation of Akt in glioblastoma cells. *The Journal of*  
578        *Biological Chemistry* 289, 600–616.
- 579    Cacas J.-L. (2015) Out for a Walk Along the Secretory Pathway During Programmed Cell  
580        Death. In: Gunawardena AN, McCabe PF (eds) *Plant Programmed Cell Death* 123–161.

581 Springer International Publishing, Cham.

582 Cacas J.-L., Petitot A.-S., Bernier L., Estevan J., Conejero G., Mongrand S. & Fernandez D.  
583 (2011) Identification and characterization of the Non-race specific Disease Resistance 1  
584 (NDR1) orthologous protein in coffee. *BMC Plant Biology* 11, 144.

585 Cacas J.-L., Vailleau F., Davoine C., Ennar N., Agnel J.-P., Tronchet M., ..., Montillet J.-L.  
586 (2005) The combined action of 9 lipoxygenase and galactolipase is sufficient to bring  
587 about programmed cell death during tobacco hypersensitive response. *Plant, Cell and*  
588 *Environment* 28,1367–1378.

589 Canton J. & Grinstein S. (2014) Priming and activation of NADPH oxidases in plants and  
590 animals. *Trends in Immunology* 35, 405–407.

591 Chen C. (2005) Real-time quantification of microRNAs by stem-loop RT-PCR. *Nucleic Acids*  
592 *Research* 33, e179–e179.

593 Dahan J., Pichereaux C., Rossignol M., Blanc S., Wendehenne D., Pugin A. & Bourque S.  
594 (2009) Activation of a nuclear-localized SIPK in tobacco cells challenged by cryptogein,  
595 an elicitor of plant defence reactions. *Biochemical Journal* 418, 191–200.

596 Dangl J.L. & Jones J.D. (2001) Plant pathogens and integrated defence responses to infection.  
597 *Nature* 411, 826–833.

598 Delage E., Ruelland E., Guillas I., Zachowski A. & Puyaubert J. (2012) Arabidopsis Type-III  
599 Phosphatidylinositol 4-Kinases  $\beta$ 1 and  $\beta$ 2 are Upstream of the Phospholipase C Pathway  
600 Triggered by Cold Exposure. *Plant & Cell Physiology* 53, 565–576.

601 Demidchik V., Shang Z., Shin R., Thompson E., Rubio L., Laohavisit A., ..., Davies J.M..  
602 (2009) Plant extracellular ATP signalling by plasma membrane NADPH oxidase and Ca

603 <sup>2+</sup> channels. *The Plant Journal* 58, 903–913.

604 Djafi N., Vergnolle C., Cantrel C., Wietrzynski W., Delage E., Cochet F., ..., Ruelland E.  
605 (2013) The Arabidopsis DREB2 genetic pathway is constitutively repressed by basal  
606 phosphoinositide-dependent phospholipase C coupled to diacylglycerol kinase. *Frontiers*  
607 *in Plant Science* 4: 307.

608 Dong W., Lv H., Xia G. & Wang M. (2012) Does diacylglycerol serve as a signaling  
609 molecule in plants? *Plant Signaling & Behavior* 7, 472–475.

610 Furt F., König S., Bessoule J.-J., Sargueil F., Zallot R., Stanislas T., ..., Mongrand S. (2010)  
611 Polyphosphoinositides are enriched in plant membrane rafts and form microdomains in  
612 the plasma membrane. *Plant Physiology* 152, 2173–2187.

613 Gajate C. & Mollinedo F. (2015) Lipid rafts and raft-mediated supramolecular entities in the  
614 regulation of CD95 death receptor apoptotic signaling. *Apoptosis* 20, 584–606.

615 Geer L.Y., Domrachev M., Lipman D.J. & Bryant S.H. (2002) CDART: protein homology by  
616 domain architecture. *Genome Research* 12, 1619–1623.

617 Gerbeau-Pissot P., Der C., Thomas D., Anca I.-A., Grosjean K., Roche Y., ..., Simon-Plas F  
618 (2014) Modification of Plasma Membrane Organization in Tobacco Cells Elicited by  
619 Cryptogein. *Plant Physiology* 164, 273–286.

620 Gómez-Merino F.C., Brearley C.A., Ornatowska M., Abdel-Haliem M.E.F., Zanon M.-I. &  
621 Mueller-Roeber B. (2004) AtDGK2, a novel diacylglycerol kinase from Arabidopsis  
622 thaliana, phosphorylates 1-stearoyl-2-arachidonoyl-sn-glycerol and 1,2-dioleoyl-sn-  
623 glycerol and exhibits cold-inducible gene expression. *The Journal of Biological*  
624 *Chemistry* 279, 8230–8241.



625 Goto K. & Kondo H. (2004) Functional implications of the diacylglycerol kinase family.  
626 *Advances in Enzyme Regulation* 44, 187–199.

627 den Hartog M. (2003) Nod Factor and Elicitors Activate Different Phospholipid Signaling  
628 Pathways in Suspension-Cultured Alfalfa Cells. *Plant Physiology* 132, 311–317.

629 Hawkins P.T. & Stephens L.R. (2015) PI3K signalling in inflammation. *Biochimica et*  
630 *Biophysica Acta* 1851, 882–897.

631 Hou Q., Ufer G. & Bartels D. (2015) Lipid signalling in plant responses to abiotic stress.  
632 *Plant, Cell and Environment* 39,1029-1048.

633 Hruz T., Laule O., Szabo G., Wessendorp F., Bleuler S., Oertle L., ..., Zimmermann P (2008)  
634 Genevestigator v3: a reference expression database for the meta-analysis of  
635 transcriptomes. *Advances in bioinformatics* 2008, 420747.

636 Hunt L., Otterhag L., Lee J.C., Lasheen T., Hunt J., Seki M., ..., Gray J.E. (2004) Gene-  
637 specific expression and calcium activation of *Arabidopsis thaliana* phospholipase C  
638 isoforms. *New Phytologist* 162, 643–654.

639 Janda M., Planchais S., Djafi N., Martinec J., Burketova L., Valentova O., ..., Ruelland E.  
640 (2013) Phosphoglycerolipids are master players in plant hormone signal transduction.  
641 *Plant Cell Reports* 32, 839–851.

642 Johansson O.N., Fahlberg P., Karimi E., Nilsson A.K., Ellerström M. & Andersson M.X.  
643 (2014) Redundancy among phospholipase D isoforms in resistance triggered by  
644 recognition of the *Pseudomonas syringae* effector AvrRpm1 in *Arabidopsis thaliana*.  
645 *Frontiers in Plant Science* 5, 639.

646 Jones J.D.G. & Dangl J.L. (2006) The plant immune system. *Nature* 444, 323–329.

647 de Jong C.F., Laxalt A.M., Bargmann B.O.R., de Wit P.J.G.M., Joosten M.H.A.J. & Munnik  
648 T. (2004) Phosphatidic acid accumulation is an early response in the Cf-4/Avr4  
649 interaction. *The Plant Journal* 39, 1–12.

650 Kalachova T., Iakovenko O., Kretinin S. & Kravets V. (2013) Involvement of phospholipase  
651 D and NADPH-oxidase in salicylic acid signaling cascade. *Plant Physiology and*  
652 *Biochemistry* 66, 127–133.

653 Karimi M., Depicker A. & Hilson P. (2007) Recombinational Cloning with Plant Gateway  
654 Vectors. *Plant Physiology* 145, 1144–1154.

655 Keller H., Blein J.P., Bonnet P. & Ricci P. (1996) Physiological and Molecular  
656 Characteristics of Elicitin-Induced Systemic Acquired Resistance in Tobacco. *Plant*  
657 *Physiology* 110, 365–376.

658 Kelm M.K., Weinberg R.J., Criswell H.E. & Breese G.R. (2010) The PLC/IP3R/PKC  
659 Pathway is Required for Ethanol-enhanced GABA Release. *Neuropharmacology* 58,  
660 1179–1186.

661 Knight H., Trewavas A.J. & Knight M.R. (1997) Calcium signalling in *Arabidopsis thaliana*  
662 responding to drought and salinity. *The Plant Journal* 12, 1067–1078.

663 Kobayashi M., Ohura I., Kawakita K., Yokota N., Fujiwara M., Shimamoto K., ..., Yoshioka  
664 H. (2007) Calcium-dependent protein kinases regulate the production of reactive oxygen  
665 species by potato NADPH oxidase. *The Plant Cell* 19, 1065–1080.

666 Krinke O., Flemr M., Vergnolle C., Collin S., Renou J.-P., Taconnat L., ..., Ruelland E..  
667 (2009) Phospholipase D activation is an early component of the salicylic acid signaling  
668 pathway in *Arabidopsis* cell suspensions. *Plant Physiology* 150, 424–436.

669 Krinke O., Ruelland E., Valentová O., Vergnolle C., Renou J.-P., Taconnat L., ..., Zachowski  
670 A. (2007) Phosphatidylinositol 4-kinase activation is an early response to salicylic acid in  
671 Arabidopsis suspension cells. *Plant Physiology* 144, 1347–1359.

672 Larkin M.A., Blackshields G., Brown N.P., Chenna R., McGettigan P.A., McWilliam H., ...,  
673 Higgins D.G. (2007) Clustal W and Clustal X version 2.0. *Bioinformatics* 23, 2947–2948.

674 Laxalt A.M., Raho N., Have A.T. & Lamattina L. (2007) Nitric Oxide Is Critical for Inducing  
675 Phosphatidic Acid Accumulation in Xylanase-Elicited Tomato Cells. *Journal of*  
676 *Biological Chemistry* 282, 21160–21168.

677 Leborgne-Castel N., Lherminier J., Der C., Fromentin J., Houot V. & Simon-Plas F. (2008)  
678 The Plant Defense Elicitor Cryptogein Stimulates Clathrin-Mediated Endocytosis  
679 Correlated with Reactive Oxygen Species Production in Bright Yellow-2 Tobacco Cells.  
680 *Plant Physiology* 146, 1255–1266.

681 Lepage M. (1967) Identification and composition of turnip root lipids. *Lipids* 2, 244–250.

682 van der Luit A.H., Piatti T., van Doorn A., Musgrave A., Felix G., Boller T. & Munnik T.  
683 (2000) Elicitation of suspension-cultured tomato cells triggers the formation of  
684 phosphatidic acid and diacylglycerol pyrophosphate. *Plant physiology* 123, 1507–1516.

685 Martin A., Adam H., Diaz-Mendoza M., Zurczak M., Gonzalez-Schain N.D. & Suarez-Lopez  
686 P. (2009) Graft-transmissible induction of potato tuberization by the microRNA miR172.  
687 *Development* 136, 2873–2881.

688 Mongrand S., Morel J., Laroche J., Claverol S., Carde J.-P., Hartmann M.-A., ..., Bessoule  
689 J.-J. (2004) Lipid rafts in higher plant cells: purification and characterization of Triton X-  
690 100-insoluble microdomains from tobacco plasma membrane. *The Journal of Biological*  
691 *Chemistry* 279, 36277–36286.

692 Munnik T., Arisz S.A., De Vrije T. & Musgrave A. (1995) G Protein Activation Stimulates  
693 Phospholipase D Signaling in Plants. *The Plant cell* 7, 2197–2210.

694 Oda T., Hashimoto H., Kuwabara N., Akashi S., Hayashi K., Kojima C., ..., Shimizu T.  
695 (2010) structure of the n-terminal regulatory domain of a plant NADPH Oxidase and its  
696 functional implications. *Journal of Biological Chemistry* 285, 1435–1445.

697 Ogasawara Y., Kaya H., Hiraoka G., Yumoto F., Kimura S., Kadota Y., ..., Kuchitsu K.  
698 (2008) Synergistic activation of the Arabidopsis NADPH oxidase AtrbohD by Ca<sup>2+</sup> and  
699 phosphorylation. *The Journal of Biological Chemistry* 283, 8885–8892.

700 Petroustos D., Amiar S., Abida H., Dolch L.-J., Bastien O., Rébeillé F., ..., Maréchal E.  
701 (2014) Evolution of galactoglycerolipid biosynthetic pathways--from cyanobacteria to  
702 primary plastids and from primary to secondary plastids. *Progress in Lipid Research* 54,  
703 68–85.

704 Pinosa F., Buhot N., Kwaaitaal M., Fahlberg P., Thordal-Christensen H., Ellerström M. &  
705 Andersson M.X. (2013) Arabidopsis phospholipase dδ is involved in basal defense and  
706 nonhost resistance to powdery mildew fungi. *Plant Physiology* 163, 896–906.

707 Pokotylo I., Kolesnikov Y., Kravets V., Zachowski A. & Ruelland E (2014) Plant  
708 phosphoinositide-dependent phospholipases C: Variations around a canonical theme.  
709 *Biochimie* 96, 144–157.

710 Pokotylo I., Pejchar P., Potocký M., Kocourková D., Krčková Z., Ruelland E., ..., Martinec J.  
711 (2013) The plant non-specific phospholipase C gene family. Novel competitors in lipid  
712 signalling. *Progress in Lipid Research* 52, 62–79.

713 Pugin A., Frachisse J.M., Tavernier E., Bligny R., Gout E., Douce R. & Guern J. (1997) Early  
714 events induced by the elicitor cryptogein in tobacco cells: involvement of a plasma

715 membrane NADPH Oxidase and activation of glycolysis and the pentose phosphate  
716 pathway. *The Plant Cell* 9, 2077–2091.

717 Ricci P., Bonnet P., Huet J.-C., Sallantin M., Beauvais-Cante F., Bruneteau M., ..., Pernollet  
718 J.-C. (1989) Structure and activity of proteins from pathogenic fungi *Phytophthora*  
719 eliciting necrosis and acquired resistance in tobacco. *European Journal of Biochemistry*  
720 183, 555–563.

721 Ruelland E., Kravets V., Derevyanchuk M., Martinec J., Zachowski A. & Pokotylo I. (2015)  
722 Role of phospholipid signalling in plant environmental responses. *Environmental and*  
723 *Experimental Botany* 114, 129–143.

724 Ruelland E., Pokotylo I., Djafi N., Cantrel C., Repellin A. & Zachowski A. (2014) Salicylic  
725 acid modulates levels of phosphoinositide dependent-phospholipase C substrates and  
726 products to remodel the Arabidopsis suspension cell transcriptome. *Frontiers in Plant*  
727 *Physiology* 5, 608.

728 Rustérucci C., Montillet J.L., Agnel J.P., Battesti C., Alonso B., Knoll A., ...,  
729 Triantaphylidès C. (1999) Involvement of lipoxygenase-dependent production of fatty  
730 acid hydroperoxides in the development of the hypersensitive cell death induced by  
731 cryptogein on tobacco leaves. *The Journal of Biological Chemistry* 274, 36446–36455.

732 Sheng X., Yung Y.C., Chen A. & Chun J. (2015) Lysophosphatidic acid signalling in  
733 development. *Development* 142, 1390–1395.

734 Simon-Plas F., Elmayan T. & Blein J.-P. (2002) The plasma membrane oxidase NtrbohD is  
735 responsible for AOS production in elicited tobacco cells. *The Plant Journal* 31, 137–147.

736 Snedden W.A. & Blumwald E. (2000) Alternative splicing of a novel diacylglycerol kinase in  
737 tomato leads to a calmodulin-binding isoform. *The Plant Journal* 24, 317–326.

738 Song C.J., Steinebrunner I., Wang X., Stout S.C. & Roux S.J. (2006) Extracellular ATP  
739 induces the accumulation of superoxide via NADPH oxidases in Arabidopsis. *Plant*  
740 *Physiology* 140, 1222–1232.

741 Stotz H.U., Mitrousis G.K., de Wit P.J.G.M. & Fitt B.D.L. (2014) Effector-triggered defence  
742 against apoplastic fungal pathogens. *Trends in Plant Science* 19, 491–500.

743 Suzuki K.G.N. (2015) New insights into the organization of plasma membrane and its role in  
744 signal transduction. *International Review of Cell and Molecular Biology* 317; 67–96.

745 Tavernier E., Wendehenne D., Blein J.-P. & Pugin A. (1995) Involvement of free calcium in  
746 action of cryptogein, a proteinaceous elicitor of hypersensitive reaction in tobacco cells.  
747 *Plant Physiology* 109, 1025–1031.

748 Tester M. (1990) Plant ion channels: whole-cell and single channel studies. *New Phytologist*  
749 114, 305–340.

750 de Torres Zabela M., Fernandez-Delmond I., Niittyla T., Sanchez P. & Grant M. (2002)  
751 Differential expression of genes encoding Arabidopsis phospholipases after challenge  
752 with virulent or avirulent *Pseudomonas* isolates. *Molecular plant-microbe interactions*  
753 15, 808–816.

754 Van Horn W.D. & Sanders C.R. (2012) Prokaryotic diacylglycerol kinase and undecaprenol  
755 kinase. *Annual Review of Biophysics* 41, 81–101.

756 Vaultier M.-N., Cantrel C., Guerbette F., Boutté Y., Vergnolle C., Çiçek D., ..., Ruelland E.  
757 (2008) The hydrophobic segment of Arabidopsis thaliana cluster I diacylglycerol kinases  
758 is sufficient to target the proteins to cell membranes. *FEBS Letters* 582, 1743–1748.

759 Vaultier M.-N., Cantrel C., Vergnolle C., Justin A.-M., Demandre C., Benhassaine-Kesri G.,

760 ..., Ruelland E (2006) Desaturase mutants reveal that membrane rigidification acts as a  
761 cold perception mechanism upstream of the diacylglycerol kinase pathway in Arabidopsis  
762 cells. *FEBS Letters* 580, 4218–4223.

763 Vossen J.H., Abd-El-Haliem A., Fradin E.F., van den Berg G.C.M., Ekengren S.K., Meijer  
764 H.J.G., ..., [Joosten M.H.](#) (2010) Identification of tomato phosphatidylinositol-specific  
765 phospholipase-C (PI-PLC) family members and the role of PLC4 and PLC6 in HR and  
766 disease resistance. *The Plant Journal* 62, 224–239.

767 Wang G., Ryu S. & Wang X. (2012) Plant phospholipases: an overview. *Methods in*  
768 *molecular biology* 861, 123–137.

769 Wong R., Fabian L., Forer A. & Brill J.A. (2007a) Phospholipase C and myosin light chain  
770 kinase inhibition define a common step in actin regulation during cytokinesis. *BMC Cell*  
771 *Biology* 8, 15.

772 Wong H.L., Pinontoan R., Hayashi K., Tabata R., Yaeno T., Hasegawa K., ..., [Shimamoto K.](#)  
773 (2007b) Regulation of rice NADPH Oxidase by binding of Rac GTPase to Its N-terminal  
774 extension. *The Plant Cell* 19, 4022–4034.

775 Yamaguchi T., Kuroda M., Yamakawa H., Ashizawa T., Hirayae K., Kurimoto L., ...,  
776 Shibuya N (2009) Suppression of a phospholipase D gene, OsPLDbeta1, activates  
777 defense responses and increases disease resistance in rice. *Plant Physiology* 150, 308–  
778 319.

779 Young S.A., Wang X. & Leach J.E. (1996) Changes in the Plasma Membrane Distribution of  
780 Rice Phospholipase D during Resistant Interactions with *Xanthomonas oryzae* pv *oryzae*.  
781 *The Plant Cell* 8, 1079.

782 Yun B.-W., Feechan A., Yin M., Saidi N.B.B., Le Bihan T., Yu M., ..., Loake G.J. (2011) S-

783 nitrosylation of NADPH oxidase regulates cell death in plant immunity. *Nature* 478,  
784 264–268.

785 Zhang W., Chen J., Zhang H. & Song F. (2008) Overexpression of a rice diacylglycerol  
786 kinase gene OsBIDK1 enhances disease resistance in transgenic tobacco. *Molecules and*  
787 *cells* 26, 258–264.

788 Zhang Q. & Xiao S. (2015) Lipids in salicylic acid-mediated defense in plants: focusing on  
789 the roles of phosphatidic acid and phosphatidylinositol 4-phosphate. *Frontiers in Plant*  
790 *Science* 6, 387.

791 Zhang Y., Zhu H., Zhang Q., Li M., Yan M., Wang R., ..., Wang X. (2009) Phospholipase  
792  $\alpha 1$  and phosphatidic acid regulate NADPH oxidase activity and production of  
793 reactive oxygen species in ABA-mediated stomatal closure in Arabidopsis. *The Plant cell*  
794 21, 2357–2377.

795 Zhao J., Devaiah S.P., Wang C., Li M., Welti R. & Wang X. (2013) Arabidopsis  
796 phospholipase D $\beta 1$  modulates defense responses to bacterial and fungal pathogens. *The*  
797 *New Phytologist* 199, 228–240.

798



799 **FIGURE LEGENDS**

800

801 **Figure 1: Early accumulation of phosphatidic acid in response to cryptogein.** Lipids were  
802 extracted at different time points after cryptogein addition. Cells were radiolabeled for 20 min  
803 before lipid extraction. (a) TLC plates were developed in an isoctane based solvent system.  
804 (b) The radioactivity associated with PA was quantified relative to the radioactivity associated  
805 with all others phospholipids (PL) and then expressed as % of the highest value. For each  
806 point of the kinetics, 3 flasks of tobacco cells were treated independently with cryptogein.  
807 Data are mean  $\pm$  SD,  $n=3$ . Note that the control error bars are within the symbol.

808

809 **Figure 2: Decrease in cryptogein-induced phosphatidic acid accumulation by PLC and**  
810 **DGK inhibitors.** Edelfosine (a) and R50022 (b) were used as PI-PLC and DGK inhibitors,  
811 respectively. Wortmannin (c) and lanthanum (d) were used as type III-PI4K and calcium  
812 channel inhibitors, respectively. Cells were radiolabeled for 10 min and then cryptogein was  
813 added. Lipids were extracted at 10 min after cryptogein addition. When necessary, inhibitors  
814 were added 15 min before the cryptogein. Labelling was carried out during 20 min for each  
815 point. Extracted lipids were separated using TLC plates and developed in an acidic solvent  
816 system. Radioactivity in PA spots was quantified relative to PE and results are expressed as %  
817 of that in cryptogein treated cells. For each treatment, 3 flasks of tobacco cells were used to be  
818 elicited or not with cryptogein in presence or not of inhibitors. The data are mean  $\pm$  SD,  $n=3$ .  
819 Results were analysed using a one-way ANOVA, with a Tukey honestly significant difference  
820 (HSD) multiple mean comparison *post hoc* test. Different letters indicate a significant  
821 difference (Tukey HSD,  $P<0.05$ ).

822

823 **Figure 3: Inhibition of RBOHD-dependent oxidative burst by both DGK and PLC**  
824 **inhibitors.** Time course of ROS production of WT BY-2 cells pretreated with either R59022,  
825 an inhibitor of DGK activity (a), or edelfosine, an inhibitor of PLC activity (c). Total amounts  
826 of ROS produced over a 90 min period following elicitation were calculated for cell  
827 suspensions treated with R59022 (b) or edelfosine (d). Cells were preincubated in I20 medium  
828 for 30 min with the inhibitor at the indicated concentrations before cryptogein was added.  
829 Extracellular ROS synthesis was regularly quantified using a luminol-based method. Data are  
830 expressed as nmoles of H<sub>2</sub>O<sub>2</sub> equivalents per gram of cells. Means and SD for 3 independent  
831 replicates. Statistical difference for the total ROS level was assessed by a Student T-test,  
832 where \* indicates P<0.05 and \*\* refers to P<0.01.

833

834 **Figure 4: Phylogenetic clustering of tobacco DGKs and characterization of one member**  
835 **of cluster III.** (a) Phylogenetic cluster of plant DGKs. Nucleic acid sequences were retrieved  
836 for DGKs from the *Solanacea Genomics Network* (SGN) database (<https://solgenomics.net/>)  
837 using the BLAST algorithm and keyword search tool. Known Arabidopsis, rice and tomato  
838 DGKs were used as nucleic acid sequence queries. The phylogenetic analysis was carried out  
839 using identified nucleic acid sequences (indicated by their SGN references) as previously  
840 described (Cacas et al. 2011). Plant DGKs were grouped into 3 phyla, each showing slightly  
841 different protein domain structures. Abbreviations: C1, C1 DAG binding domain; CBD,  
842 calmodulin-binding domain; DGKcat, DGK catalytic domain; DGKacc, DGK accessory  
843 domain. For plant species, At refers to *Arabidopsis thaliana*, Le to *Solanum esculentum*  
844 (previously *Lycopersicon esculentum*), Nt to *Nicotiana tabacum* and Os to *Oryza sativa* (b)  
845 Time course of *NtDGK5-like* gene expression in response to cryptogein. Gene expression was  
846 determined by Q-PCR as described in the *Material and Methods*. Data are means and SD for  
847 3 independent biological replicates (n=3 samples per cell line). (c) Subcellular localization of  
848 NtDGK5-like protein. Cells from overexpressing *NtDGK5-like-c-myc* lines  
849 (*OE.NtDGK5myc-A* and *OE.NtDGK5myc-B*) and the corresponding empty vector-containing  
850 line (*i.e.* line transformed with an empty pMDC32) were harvested 7 days after subculture.  
851 Proteins were extracted from cytosolic (lanes 1), microsomal (lanes 2) and plasma membrane  
852 (lanes 3) fractions. After western blotting using anti-c-myc antibodies, a specific signal was  
853 only found in membrane fractions. Two independent biological replicates were carried out  
854 (n=2 samples per cell line).

855

856 **Figure 5: Characterization of *NtDGK5-like* overexpressing transgenic lines.** (a)  
857 Expression levels of *NtDGK5-like* overexpressing lines. Gene expression was determined by  
858 Q-PCR using primers that amplify both the endogenous and transgene-encoding transcripts.  
859 Data are means and SD for 4 independent experiments. These experiments were carried out  
860 on material obtained from untreated 7-day-old cells. BY-2, WT cells; the empty vector-  
861 containing line is a BY-2 line transformed with an empty pMDC32; *OE.NtDGK5-A*, line  
862 overexpressing an untagged version of NtDGK5-like protein; *OE.NtDGK5myc-A* and  
863 *OE.NtDGK5myc-B* are independent lines overexpressing a C-terminally tagged version of  
864 NtDGK5-like protein. (b) Radioactivity associated with PA in cryptogein-elicited  
865 overexpressors. Cells were radiolabeled for 20 min before lipid extraction. Lipids were  
866 extracted 10 min after cryptogein addition. TLC plates were developed in an acid solvent  
867 system. The radioactivity associated with PA was normalized to that of PE and PC. The data  
868 are expressed as % of the value in the empty vector control line elicited with cryptogein. Data  
869 are means and SD,  $n=3$ . (c) Total ROS levels produced during a 90 min period (same  
870 experimental design as that described in the legend of figure 4). Data are expressed as percent  
871 compared to the ROS level of BY-2 cells being set to 100 (corresponding to 500 nmoles  $H_2O_2$   
872 eq./g of cells). Means and SD for 3 independent experiments. Statistical differences for the  
873 total ROS level was assessed using a one-way ANOVA, with a Tukey honestly significant  
874 difference (HSD) multiple mean comparison *post hoc* test. Different letters indicate a  
875 significant difference, Tukey HSD,  $P<0.05$ . (d) Cell density of untreated transgenic lines.  
876 Packed cell volume (PCV) and dry weight (DW), were expressed in percent. Presented data  
877 are means and SD for 6 independent experiments ( $n=6$  samples for each cell line). Control  
878 (empty vector) and overexpressors were compared to WT BY-2 suspensions using a one-way  
879 ANOVA, with a Tukey honestly significant difference (HSD) multiple mean comparison *post*  
880 *hoc* test. For PCV, different letters indicate a significant difference, Tukey HSD,  $P<0.05$ . No

881 significant difference could be detected for DW (Tukey HSD,  $P < 0.05$ .) significant differences  
882 (Tukey HSD,  $P < 0.01$ ) in PCV and DW between genotypes were detected. Letters marked  
883 with an apostrophe (') are used for DW data.

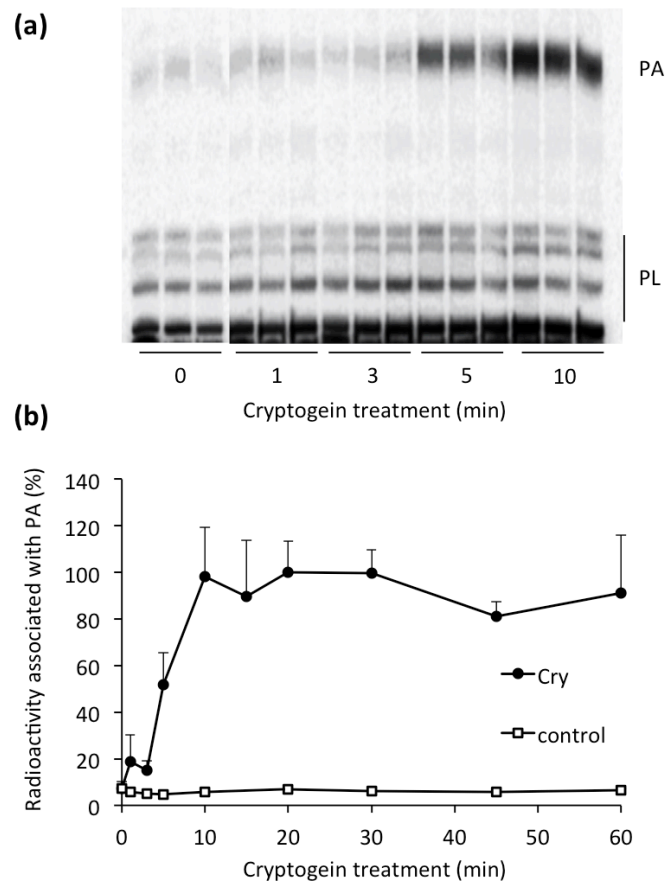
884

885 **Figure 6: Characterization of DGK Cluster III-targeted miRNA transgenic cell lines.** (a)  
886 Scheme of the nucleic region targeted by the miRNA construct (see also Supporting  
887 information Fig. S2). (b) Evidence for miRNA production *in planta*. The cluster III-specific  
888 miRNA construct was expressed in tobacco leaves using *Agrobacterium tumefaciens*  
889 (GV3101). Foliar samples were harvested 2 day-post inoculation. Total RNAs were extracted,  
890 the cluster III pri-miRNA was specifically reverse transcribed using a stem-loop system and  
891 generated cDNA used as template for PCR amplification (expected size of PCR  
892 product=181bp). Sequencing showed PCR products to be the hairpin construct. (c) Expression  
893 levels of cluster III and *NtDGK5-like* genes in miRNA cell lines. Expression levels were  
894 quantified by Q-PCR on samples obtained from untreated 7-day-old cell suspensions. Means  
895 and SD are from 4 independent biological replicates (n=4 samples for each cell line). The  
896 empty vector line corresponds to a line transformed with an empty pH7GWIWG2(II). (d) Cell  
897 suspension density of 7 day-old transgenic lines. Packed cell volume (PCV) and dry weight  
898 (DW), are expressed as a %. Presented data are means and SD for 6 independent biological  
899 replicates (n=6 samples for each cell line). The empty vector line and the *hp-clusterIII-A* and  
900 *hp-clusterIII-B* cell lines were compared to WT BY-2 suspensions using a one-way ANOVA,  
901 with a Tukey honestly significant difference (HSD) multiple mean comparison *post hoc* test.  
902 No significant differences (Tukey HSD, P<0.01) in PCV and DW between genotypes were  
903 detected. Letters marked with an apostrophe (') are used for DW data.

904

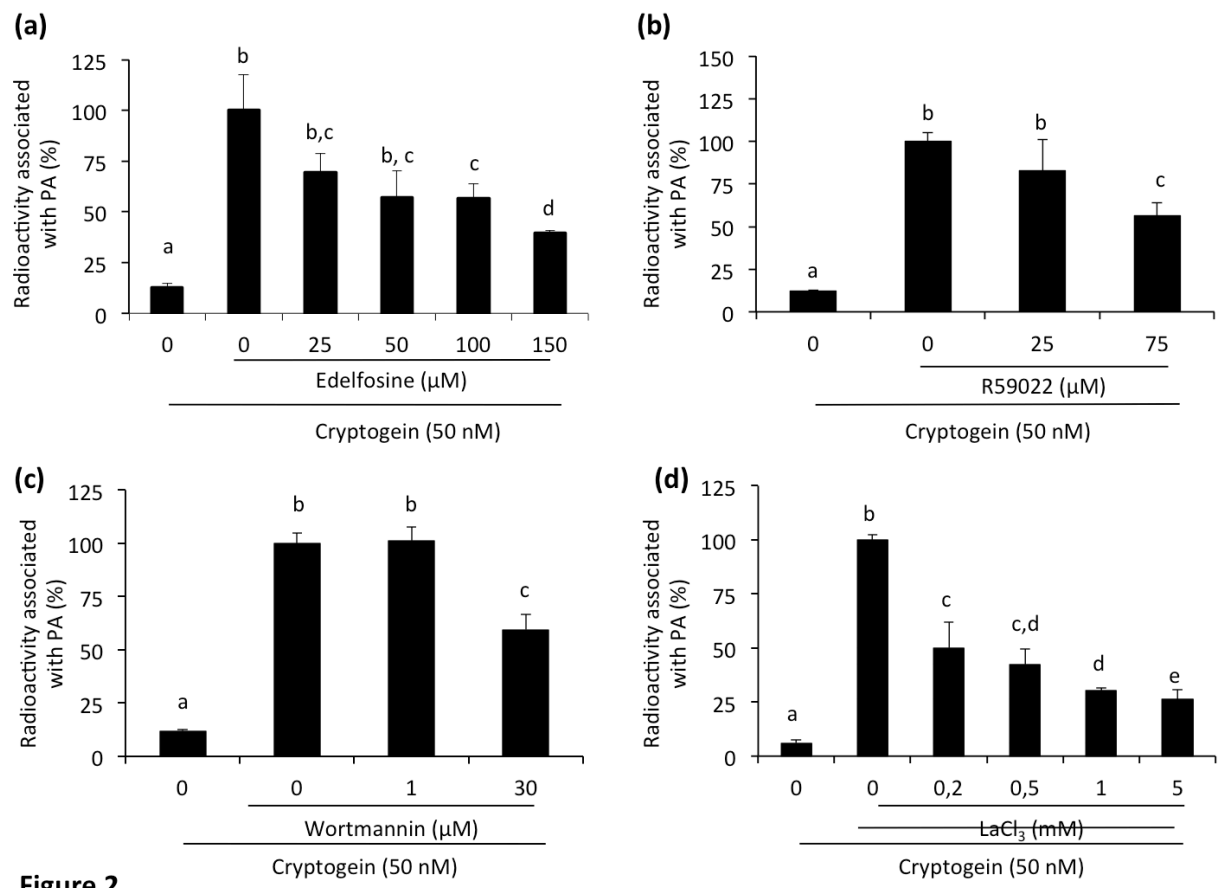


905 **Figure 7: Silencing DGK Cluster III of tobacco abolishes PA production and strongly**  
906 **inhibits the oxidative burst in response to cryptogein.** (a) Levels of PA in response to  
907 cryptogein. Cells were radiolabeled for 20 min before lipid extraction. Lipids were extracted  
908 10 min after cryptogein addition. TLC plates were developed using an acid solvent system.  
909 The radioactivity associated with PA was normalized to that of PE and PC. For each line, the  
910 data are expressed as % of the value of the non-elicited cells. For each treatment, 3 flasks of  
911 cells were used and the data are mean  $\pm$  SD,  $n=3$ . Statistical difference between treated and  
912 non-treated conditions was assessed by a Student T-test, where \* indicates  $P<0.05$ . (b) Time-  
913 course of ROS levels following elicitation with cryptogein. Data are means and SD for 8  
914 independent biological replicates ( $n=8$  samples per kinetic point for each cell line). (c) Total  
915 amounts of ROS produced over a 90 min period after cryptogein addition (same data as in  
916 panel b). Statistical differences was assessed using a one-way ANOVA, with a Tukey  
917 honestly significant difference (HSD) multiple mean comparison *post hoc* test. Different  
918 letters indicate a significant difference, Tukey HSD,  $P<0.01$ .  
919



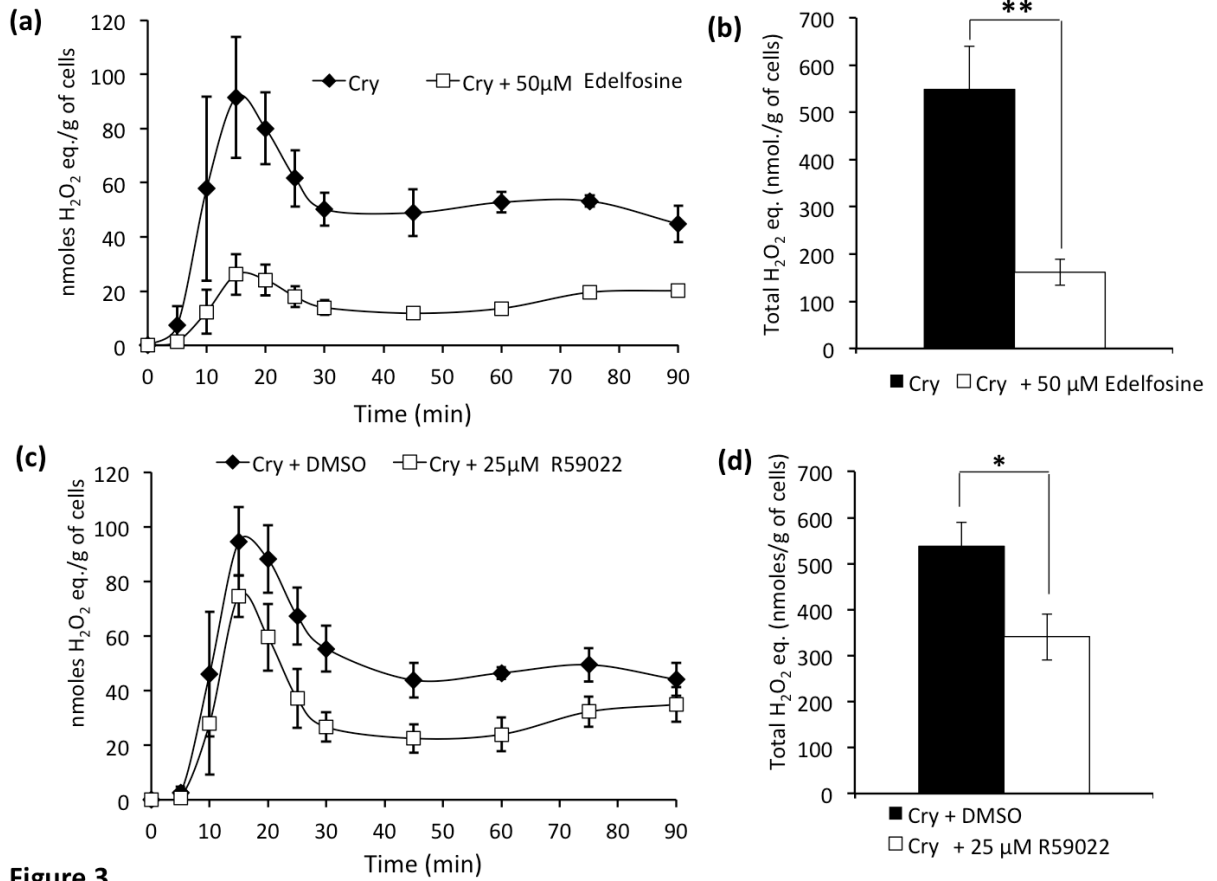
921 Figure 1

922



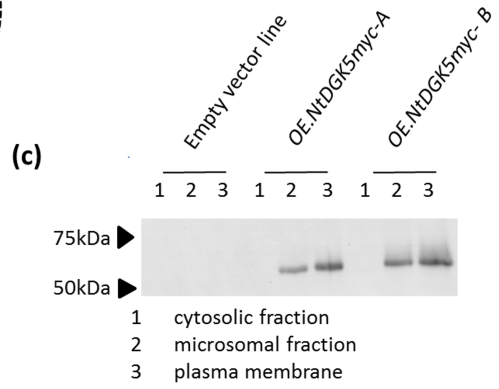
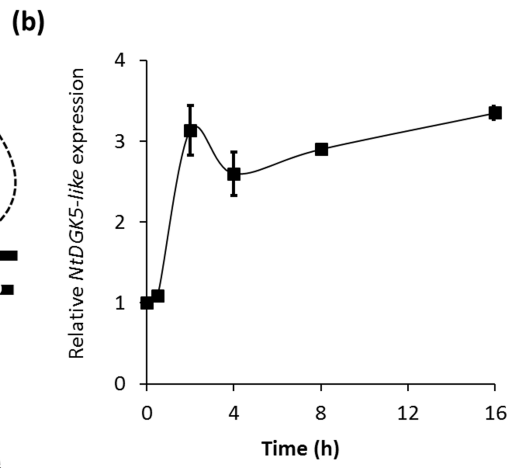
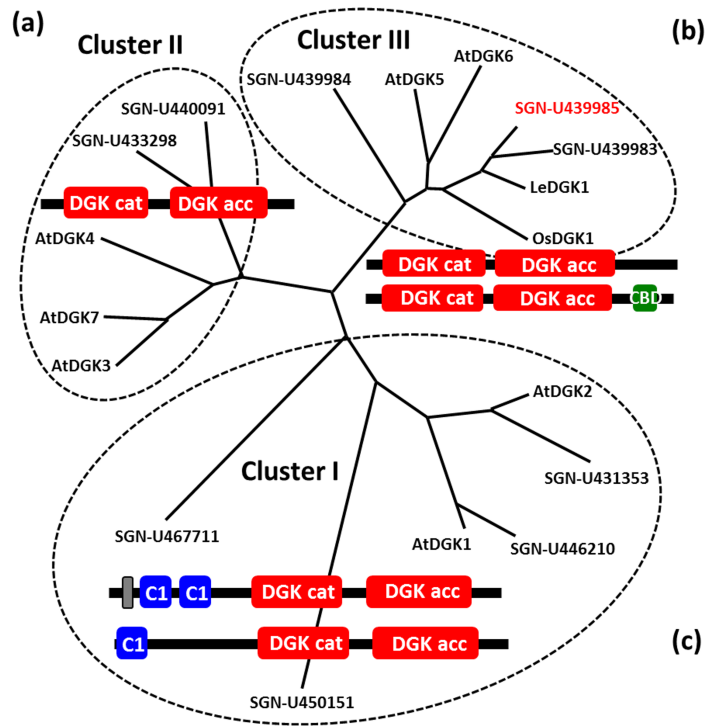
923 **Figure 2**

924



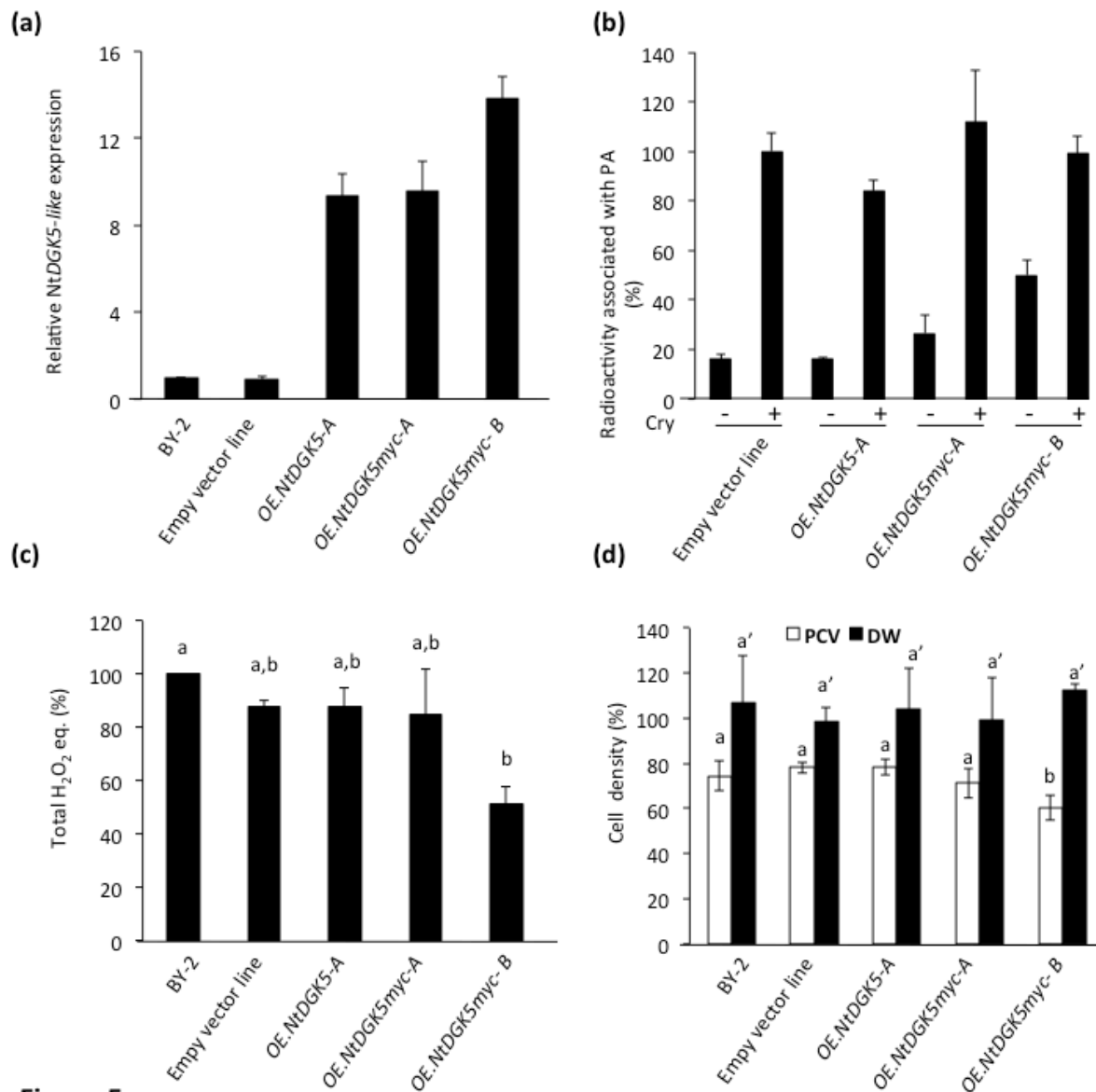
925 **Figure 3**

926



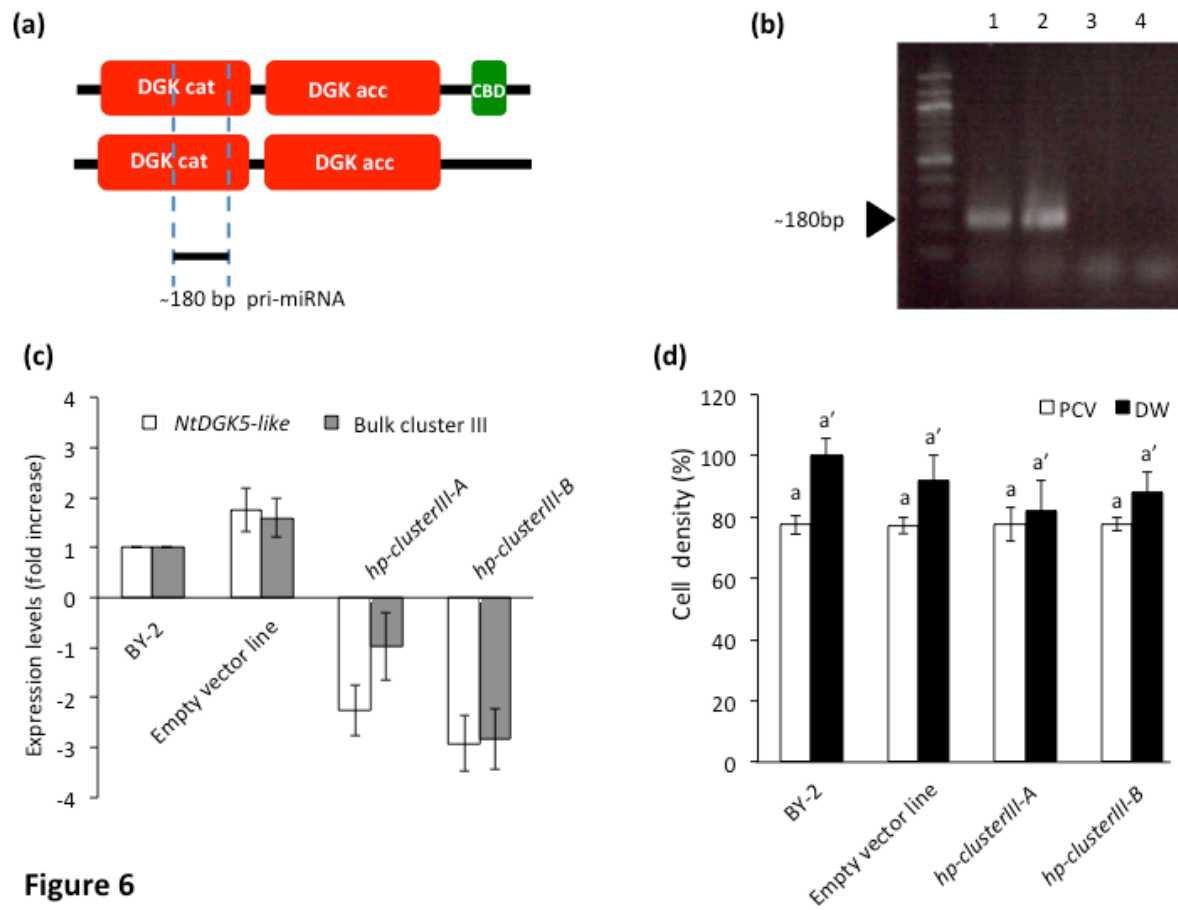
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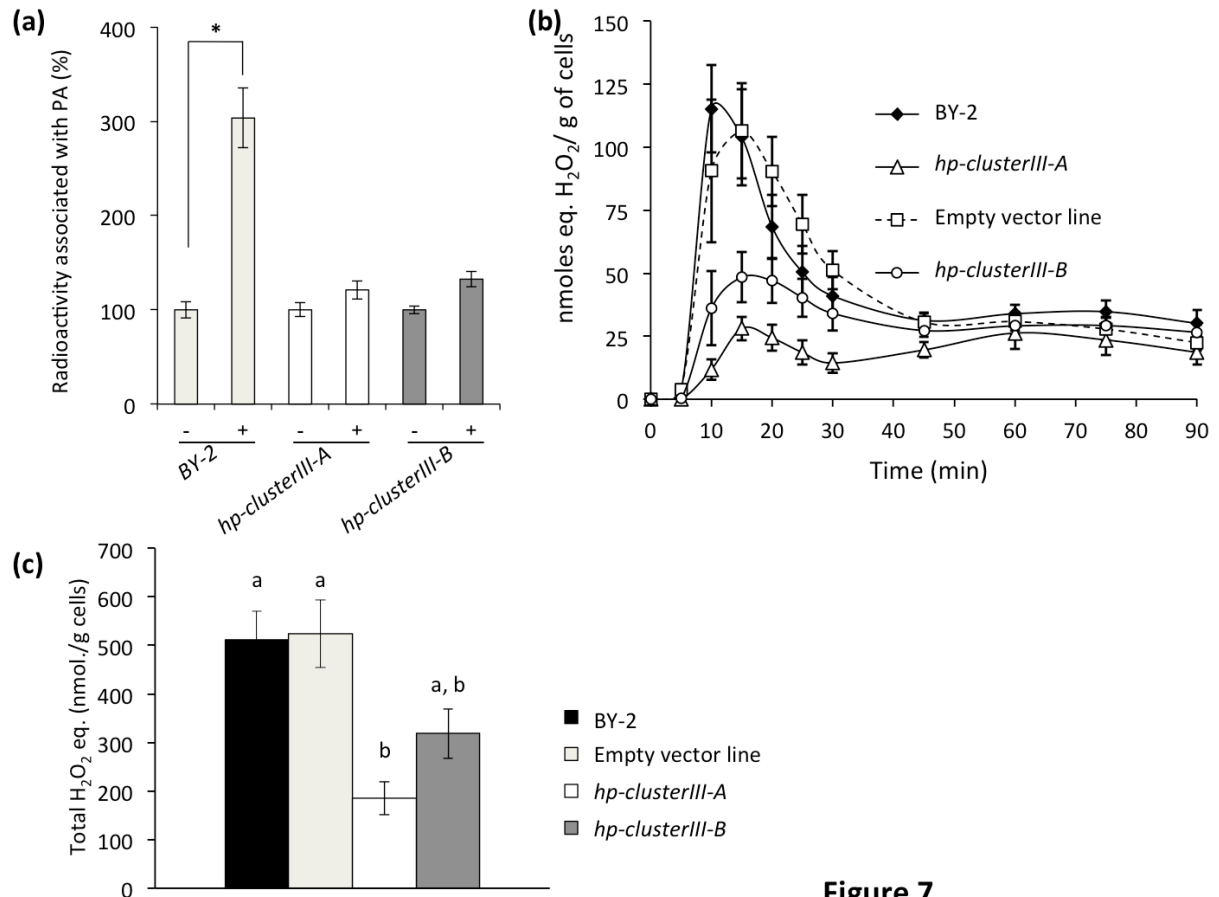
929 **Figure 5**

930



931 **Figure 6**

932



933

934

Figure 7



935 **SUPPORTING INFORMATION**

936 **Supporting information figure S1:** The translated coding sequence of the tobacco *NtDGK5-*  
937 *like* gene. The identified nucleic acid sequence was deposited in GenBank under the reference  
938 KU934207. It codes for a 493-amino acid-long protein (55.24kDa), having a predicted pI of  
939 6.75. *In silico* translation was performed using the freeware ORF Finder available at the  
940 NCBI website (<http://www.ncbi.nlm.nih.gov/gorf/gorf.html>).

941 **Supporting information figure S2:** Partial alignment of plant DGK coding sequences from  
942 clusters II and III. This figure shows the nucleic acid coding region used for miRNA design  
943 (squared in red), which is highly conserved among cluster III members and divergent with  
944 cluster II counterparts. Full-length nucleic acid sequences were aligned using version 2.0.10  
945 of Clustal X (Larkin et al. 2007) and the resulting alignment was processed online at the  
946 BoxShade server ([http://www.ch.embnet.org/software/BOX\\_form.html](http://www.ch.embnet.org/software/BOX_form.html)). Roman numbers (II  
947 and III) on the right of the alignment indicate DGK cluster.

948 **Supporting information figure S3:** Effects of inhibitors on the alkalization triggered by  
949 cryptogein. Cells were preincubated 15 min with inhibitors (or DMSO, solvent of the  
950 inhibitors) before cryptogein (50 nM) elicitation. Final concentrations of edelfosine, R59022  
951 and R59949 were 25  $\mu$ M, 75  $\mu$ M and 75  $\mu$ M, respectively. R59949 is another DGK inhibitor.  
952 Extracellular pH alkalization was measured and reported as the difference between initial  
953 and final (1 hour of cryptogein treatment) pH values. Means +/- SD.

954 **Supporting information figure S4:** Topology of the sixteen identified tobacco DGK  
955 proteins. Protein structure is depicted for the 3 phylogenetic clusters. DGK nucleic acid  
956 sequences were retrieved from the *Solanacea Genomics Network* (SGN) database  
957 (<https://solgenomics.net/>) using the BLAST algorithm and keyword search tool. Known  
958 Arabidopsis, rice and tomato DGKs were used as nucleic acid sequence queries. Upon *in*

959 *silico* translation using ORF Finder (<http://www.ncbi.nlm.nih.gov/gorf/gorf.html>), the  
960 resulting polypeptides were compared to their Arabidopsis counterparts and the Conserved  
961 Domain Architecture Retrieval Tool (CDART, (Geer et al. 2002)) was used to predict domain  
962 location. Within a panel (corresponding to one protein cluster), the closest Arabidopsis  
963 relative is followed by orthologous tobacco translated sequences. The latter are referenced by  
964 their SGN numbers and the identified part of the protein is presented. The length of the  
965 corresponding contigs is also indicated in green. Abbreviations: AtDGK5lv, AtDGK5 long  
966 splice variant; AtDGK5sv, AtDGK5 short splice variant; C1, DAG binding domain; CBD,  
967 calmodulin-binding domain; DGKcat, DGK catalytic domain; DGKacc, DGK accessory  
968 domain. The grey rectangle localized to the N-terminal part of AtDGK1 represents a  
969 transmembrane domain.

970 **Supporting information figure S5:** Identification of the PA binding domain of NtRBOHD  
971 by alignment with the cognate domain of Arabidopsis isoform D. Full length primary amino-  
972 acid sequences of RBOHD from *Arabidopsis thaliana* (SwissProt:Q9FIJ0.1) and *Nicotiana*  
973 *tabacum* (GenBank:ABN58915) were aligned using version 2.0.10 of Clustal X (Larkin et al.  
974 2007) and the resulting alignment was then processed online at the BoxShade server  
975 ([http://www.ch.embnet.org/software/BOX\\_form.html](http://www.ch.embnet.org/software/BOX_form.html)). Stars indicate the two basic amino-  
976 acid residues experimentally proven to be critical for interaction with PA (Zhang *et al.* 2009).  
977 Note the conservation of these residues between the two proteins.