# Species of Pseudorhabdosynochus (Monogenea, Diplectanidae) from Groupers (Mycteroperca spp., Epinephelidae) in the Mediterranean and Eastern Atlantic Ocean, with Special Reference to the 'Beverleyburtonae Group' and Description of Two New Species 

Amira Chaabane, Lassad Neifar, Delphine Gey, Jean-Lou Justine

## To cite this version:

Amira Chaabane, Lassad Neifar, Delphine Gey, Jean-Lou Justine. Species of Pseudorhabdosynochus (Monogenea, Diplectanidae) from Groupers (Mycteroperca spp., Epinephelidae) in the Mediterranean and Eastern Atlantic Ocean, with Special Reference to the 'Beverleyburtonae Group' and Description of Two New Species. PLoS ONE, 2016, 11 (8), pp.e0159886. 10.1371/journal.pone. 0159886 . hal01375682

HAL Id: hal-01375682<br>https://hal.sorbonne-universite.fr/hal-01375682

Submitted on 3 Oct 2016

HAL is a multi-disciplinary open access archive for the deposit and dissemination of scientific research documents, whether they are published or not. The documents may come from teaching and research institutions in France or abroad, or from public or private research centers. publics ou privés.

Citation: Chaabane A, Neifar L, Gey D, Justine J-L (2016) Species of Pseudorhabdosynochus (Monogenea, Diplectanidae) from Groupers (Mycteroperca spp., Epinephelidae) in the Mediterranean and Eastern Atlantic Ocean, with Special Reference to the 'Beverleyburtonae Group' and Description of Two New Species. PLoS ONE 11 (8): e0159886. doi:10.1371/journal.pone. 0159886

Editor: Gordon Langsley, Institut national de la santé et de la recherche médicale - Institut Cochin, FRANCE

Received: April 28, 2016
Accepted: July 8, 2016
Published: August 17, 2016
Copyright: © 2016 Chaabane et al. This is an open access article distributed under the terms of the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

Data Availability Statement: All relevant data are within the paper. There are no Supporting Information files.

Funding: This work was supported by Ministère des Affaires Etrangères, France: grant BIOPARMED-
ENVIMED, http://www.mistrals-home.org/spip.php? rubrique82; and Muséum National d'Histoire Naturelle, France: Grant ATM Barcode, Grant PARSUDMED, https://www.mnhn.fr/fr. The funders

# Species of Pseudorhabdosynochus (Monogenea, Diplectanidae) from Groupers (Mycteroperca spp., Epinephelidae) in the Mediterranean and Eastern Atlantic Ocean, with Special Reference to the ‘Beverleyburtonae Group’ and Description of Two New Species 

Amira Chaabane ${ }^{1 \text { * }}$, Lassad Neifar ${ }^{1}$, Delphine Gey ${ }^{2}$, Jean-Lou Justine ${ }^{3}$<br>1 Laboratoire de Biodiversité et Écosystèmes Aquatiques, Faculté des Sciences de Sfax, Université de Sfax, Sfax, Tunisia, 2 UMS 2700 Service de Systématique moléculaire, Muséum National d'Histoire Naturelle, Sorbonne Universités, Paris, France, 3 ISYEB, Institut Systématique, Évolution, Biodiversité, UMR7205 (CNRS, EPHE, MNHN, UPMC), Muséum National d'Histoire Naturelle, Sorbonne Universités, Paris, France<br>* amirachaabene@ hotmail.fr


#### Abstract

Pseudorhabdosynochus Yamaguti, 1958 is a species-rich diplectanid genus, mainly restricted to the gills of groupers (Epinephelidae) and especially abundant in warm seas. Species from the Mediterranean are not fully documented. Two new and two previously known species from the gills of Mycteroperca spp. (M. costae, M. rubra, and M. marginata) in the Mediterranean and Eastern Atlantic Ocean are described here from new material and slides kept in collections. Identifications of newly collected fish were ascertained by barcoding of cytochrome c oxidase subunit I (COI) sequences. Pseudorhabdosynochus beverleyburtonae (Oliver, 1984) Kritsky \& Beverley-Burton, 1986 and P. sosia Neifar \& Euzet 2007 are redescribed from type-specimens and new specimens collected off Tunisia and Libya from M. marginata and $M$. costae, respectively. Pseudorhabdosynochus oliveri n. sp., from M. marginata (type-host) off the Mediterranean coast of France (type-locality), is described from specimens found among voucher specimens of $P$. beverleyburtonae deposited by Guy Oliver in the collection of the Muséum National d'Histoire Naturelle, Paris. Pseudorhabdosynochus oliveri is distinguished by the shape of its sclerotised vagina; it was not found in the other localities investigated. Pseudorhabdosynochus hayet n . sp . is described from $M$. rubra (type host) off Senegal (type-locality) and Tunisia. Pseudorhabdosynochus hayet is morphologically similar to $P$. sosia (type-host: $M$. costae) but was distinguished by differences in measurements of the vagina and male copulatory organ, different host, and divergent COI sequences. The four species ( $P$. beverleyburtonae, $P$. sosia, $P$. oliveri, and $P$. hayet) share common characteristics such as squamodiscs with 2 innermost circular rows


had no role in study design, data collection and analysis, decision to publish, or preparation of the manuscript.

Competing Interests: The authors have declared that no competing interests exist.
of rodlets and a similar general structure of the sclerotised vagina; we propose to group them into a 'beverleyburtonae group' within Pseudorhabdosynochus.

## Introduction

Pseudorhabdosynochus Yamaguti, 1958 is a species-rich diplectanid genus [1-4]; its members are mainly restricted to the gills of groupers (Epinephelidae), with a few exceptions, and, since most groupers inhabit warm seas, they are especially numerous in tropical seas [5]. Species from the Mediterranean are not fully documented, although eleven nominal species have been listed [6, 7].

Pseudorhabdosynochus beverleyburtonae (Oliver, 1984) Kritsky \& Beverley-Burton, 1986 is a gill parasite of the dusky grouper Mycteroperca marginata in the Mediterranean Sea and both sides of the Atlantic [2, 8-11]. We found that three species of Pseudorhabdosynochus from groupers assigned to Mycteroperca Gill in the Mediterranean Sea and Eastern Atlantic Ocean were morphologically very close to $P$. beverleyburtonae. Among these species, one, namely $P$. sosia Neifar \& Euzet, 2007, was already known, and two are new and are described in this paper. Based on the sclerotised vagina, the primary character for species diagnosis within Pseudorhabdosynochus, we propose the 'beverleyburtonae group' to accommodate them. Monogenean COI sequences were used to complement the morphological analysis of parasites. Fish COI sequences were used to confirm the morphological identification of hosts [12, 13].

## Materials and Methods

## Fish sampling and identification

Fish were purchased at the fish markets in Sfax and Tunis, Tunisia and in Tripoli, Libya. These were previously caught by fishermen in the nearby coastal waters of the Mediterranean Sea. In all cases, the fish were dead when available for parasitological studies. No permits were required for the described study. Fish were identified morphologically according to keys [14] and books [15], and these identifications were challenged by analyses of COI sequences of individual fish (Table 1). Fish nomenclature follows [16] and [17].

## COI sequence of fish host

We used the QIAamp DNA Mini Kit (Qiagen), as per the manufacturer's instructions, to perform DNA extraction. The $5^{\prime}$ region of the cytochrome oxidase I (COI) mitochondrial gene was amplified with the primers FishF1 ( $5^{\prime}$-TCAACCAACCACAAAGACATTGGCAC- $3^{\prime}$ ) and FishR1 ( $5^{\prime}$-TAGACTTCTGGGTGGCCAAAGAATCA-3') [12]. PCR reactions were performed in $20 \mu \mathrm{l}$, containing 1 ng of DNA, $1 \times$ CoralLoad PCR buffer, $3 \mathrm{mM} \mathrm{MgCl} 2,66 \mu \mathrm{M}$ of each dNTP, $0.15 \mu \mathrm{M}$ of each primer, and 0.5 units of Taq DNA polymerase (Qiagen). The amplification protocol was 4 min at $94^{\circ} \mathrm{C}$, followed by 40 cycles at $94^{\circ} \mathrm{C}$ for $30 \mathrm{sec}, 48^{\circ} \mathrm{C}$ for 40 sec , and $72^{\circ} \mathrm{C}$ for 50 sec , with a final extension at $72^{\circ} \mathrm{C}$ for 7 min . PCR products were purified and sequenced in both directions on a 3730xl DNA Analyzer 96-capillary sequencer (Applied Biosystems). We used CodonCode Aligner software (CodonCode Corporation, Dedham, MA, USA) to edit sequences, compared them to the GenBank database content with BLAST, and deposited them in GenBank under accession numbers KX255747 -KX255751 (Table 1). Species identification was confirmed with the BOLD identification engine [18].

Table 1. Host fish examined, their monogeneans and their COI sequences.

| Host species | Fish specimen | Locality | Date | Fish COI, GenBank | Monogenean collected | Monogenean COI, GenBank |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| M. marginata | Mmargi3 | Tunisia | 25/09/2014 | KX255749 * | P. beverleyburtonae |  |
| M. costae | Mcostae1 | Tunisia | 13/06/2014 | KX255750* | P. sosia | KX255742 * |
| M. costae | Mcostae2 | Tunisia | 13/06/2014 | KX255751 * | P. sosia |  |
| M. costae | Mcostae3 | Tunisia | 15/04/2014 | KT805240 | P. sosia |  |
| M. costae | Mcostae 4 | Libya | 2013 | - | P. sosia |  |
| M. costae | Myco6 | Tunisia | 17/09/2015 | KX255747 * | P. sosia | KX255741 * KX255743 * KX255744 * |
| M. rubra | Myru01 | Libya | 06/2013 | KX255748 * | P. regius |  |
| M. rubra | Myru02 | Tunisia | 10/09/2015 | KU739518 | $P$. regius $P$. hayet n . sp . | KX255745 * KX255746 * |

doi:10.1371/journal.pone.0159886.t001

Details about molecular identification of host fish are provided briefly in the description of the monogenean species.

## Monogenean morphology

Diplectanids collected from fish gills were prepared by three methods: a) mounted in ammonium picrate-glycerine [19] (designated as 'p'); b) mounted in Berlese fluid (designated as 'b'); and c) dehydrated in an ethanol series, stained with carmine and permanently mounted in Canada balsam (designated as 'c') [20]. Specimens were drawn using an Olympus BH2 microscope equipped with drawing apparatus and DIC optics. The terminology for the sclerotised parts, i.e. the male quadriloculate organ and the vagina follows Justine (2007) [1]. Measurements, in micrometres, were taken with the help of a custom-made transparent rule and are expressed as the mean followed in parentheses by the range, the standard deviation when $\mathrm{n} \geq 30$, and ( n ) the number of observations; measurements were taken as in Fig 1 in Chaabane et al. (2015) [6]. The measurements of the right-hand haptoral hard-parts and left-hand equivalents were pooled. The measurements of the holotype are separated and indicated by ' h '. Drawings were scanned and redrawn on a computer using Adobe Illustrator. The museum abbreviation used is as follows: MNHN, Muséum National d'Histoire Naturelle, Paris.

## COI sequence of monogeneans

We used a QIAmp DNA Micro Kit (Qiagen) to extract DNA from whole monogenean specimens. The specific primers COI-ASmit1 (forward $5^{\prime}$-TTTTTTGGGCATCCTGAGGTTTAT$3^{\prime}$ ) and COI-ASmit2 (reverse $5^{\prime}$-TAAAGAAAGAACATAATGAAAATG- $3^{\prime}$ ) were used to amplify a fragment of 424 bp of the COI gene [21]. The PCR reaction was performed in $20 \mu \mathrm{l}$, containing 1 ng of DNA, $1 \times$ CoralLoad PCR buffer, $3 \mathrm{mM} \mathrm{MgCl}_{2}, 0.25 \mathrm{mM}$ dNTP, $0.15 \mu \mathrm{M}$ of each primer, and 0.5 units of Taq DNA polymerase (Qiagen). Thermocycles consisted of an initial denaturation step at $94^{\circ} \mathrm{C}$ for 2 min , followed by 37 cycles of denaturation at $94^{\circ} \mathrm{C}$ for 30 sec , annealing at $48^{\circ} \mathrm{C}$ for 40 sec , and extension at $72^{\circ} \mathrm{C}$ for 50 sec . The final extension was conducted at $72^{\circ} \mathrm{C}$ for 5 min . Sequences were edited with CodonCode Aligner software (CodonCode Corporation, Dedham, MA, USA), compared to the GenBank database content with BLAST, and deposited in GenBank under accession numbers KX255741 -KX255746 (Table 1).

Pairwise nucleotide distances were assessed using the Kimura 2-parameter (K2P) model [22] in MEGA 7. The phylogenetic tree was constructed using the Neighbour Joining (NJ) method based on the Kimura 2-parameter (K2P) model in MEGA 7 [23]; all codon positions were used.


Fig 1. Pseudorhabdosynochus beverleyburtonae from Mycteroperca marginata in the Mediterranean Sea and Atlantic Ocean, various shapes of sclerotised vagina according to specimens, orientation and preparation. A-F, H, new specimens from Tunisia. G, MNHN HEL466, J, MNHN HEL465, voucher specimens from Brazil. I, MNHN 249H-Tc167, type specimen. A-F, H, Berlese. G, J, Gray and Wess medium. I, carmine.
doi:10.1371/journal.pone.0159886.g001

## Nomenclatural acts

The electronic edition of this article conforms to the requirements of the amended International Code of Zoological Nomenclature, and hence the new names contained herein are available under that Code from the electronic edition of this article. This published work and the nomenclatural acts it contains have been registered in ZooBank, the online registration system for the ICZN. The ZooBank LSIDs (Life Science Identifiers) can be resolved and the associated information viewed through any standard web browser by appending the LSID to the prefix "http://zoobank.org/". The LSID for this publication is: urn:lsid:zoobank.org:pub:CB13D383-7994-4BCE-BB8F-A46E874E26D3. The electronic edition of this work was published in a journal with an ISSN, and has been archived and is available from the following digital repositories: PubMed Central, LOCKSS.

## Results

## Pseudorhabdosynochus beverleyburtonae (Oliver, 1984) Kritsky \& Beverley-Burton, 1986

Synonyms: Diplectanum americanum of Euzet \& Oliver, 1965, nec Price, 1937; Cycloplectanum americanum (Price, 1937) Oliver, 1968 (pro parte); Cycloplectanum beverleyburtonae Oliver, 1984.

Type-host: Dusky grouper, Mycteroperca marginata (Lowe) (Perciformes, Epinephelidae) sometimes designated as Epinephelus guaza (Linnaeus) or E. marginatus (Lowe).

Molecular identification of fish via DNA barcoding: The COI sequence of our specimen (KX255749) (Table 1) was identical to three sequences (KU739519-521) previously obtained from the same fish species in the same locality (Tunisia) [24]. We conclude that our specimen belongs to M. marginata.

Site of infection: Gills
Type-locality: Off France, Mediterranean Sea, as "Côte Vermeille, Golfe du Lion, Méditerranée occidentale" [10].

Other localities: Off Rosas, Mediterranean Sea, France [25]; off Banyuls-sur-Mer, France [8, 9]; off Ilhas Cagarras, Rio de Janeiro, Brazil [11]; off Ubatuba, coast of São Paulo, south-eastern Brazil [26]; off Barra Velha, State of Santa Catarina, Brazil [2]; Sfax (fish market), Tunisia (present study). The Bay of Naples, Italy is sometimes indicated as a locality based on Ulmer \& James (1981) [27] but this paper does not describe the sclerotised vagina so it could be, in our opinion, any species of Pseudorhabdosynochus.

Material examined: 2 type-specimens from off France (MNHN 249H-Tc167, $249 \mathrm{H}-\mathrm{Tc} 167 \mathrm{bis}$ ) collected by Guy Oliver; voucher specimens from off Brazil (MNHN HEL465, HEL466, HEL470, HEL471) collected 3 February 2014 and deposited by Kritsky et al. [2] in the MNHN collection; new specimens collected off Tunisia (MNHN HEL560) (see Table 2).

Prevalence: In our newly examined fish specimen from Tunisia, $1 / 1$.
Redescription (Figs 1 and 2). Measurements of 17 specimens in Berlese from Tunisia; for other specimens see Table 2. Body length b 648 ( $570-750, \mathrm{n}=6$ ), including haptor; maximum width b $145(120-180, n=6)$ at level of ovary. Tegument smooth. Anterior region with 3 pairs of head organs and 2 pairs of dorsal eye-spots, distance between outer margins of anterior eyespots $b 25(14-40, n=16)$, of posterior eye-spots $b 21(14-37, n=16)$. Pharynx median, subspherical, length b $36(29-40, n=5)$, width b $32(25-39, n=5)$. Haptor bearing 2 similar squamodiscs, 2 pairs of lateral anchors, 1 ventral bar and 2 lateral (dorsal) bars (Fig 2C-2E) and 14 marginal hooklets. Squamodiscs with 9-12 concentric rows of rodlets; two innermost rows forming circles (Fig 2C and 2D). Rodlets sometimes with visible spurs ('éperons'). Ventral

ONE

Table 2. Measurements of $P$. beverleyburtonae from various sources.

| Source | Santos, Buchmann \& Gibson, 2000 | Kritsky, Bakenhaster \& Adams, 2015 | Type-specimens, MNHN 249H-Tc167, 249H-Tc167 bis | Slides deposited in MNHN, HEL 465, HEL 466, HEL 470, HEL 471 |  | MNHN HEL560, Present study |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| Hosts | M. marginata | M. marginata | M. marginata | M. marginata |  | M. marginata |
| Locality | Off llhas Cagarras, Rio de Janeiro, Brazil | Off Barra Velha, State of Santa Catarina, Brazil | Off Cap Béar, France | Off Barra Velha, State of Santa Catarina, Brazil |  | Off Sfax, Tunisia |
| Method | Gomori's trichrome, Mayer's paracarmine | Gomori's trichrome, Gray and Wess medium | Carmine | Gomori's trichrome | Gray and Wess medium | Berlese |
| n | 26 | - | 2 | 2 | 2 | 17 |
| Body Length | 492-617 | 741 (569-974, n=24) | 510 (490-530, n=2) | $\begin{aligned} & 755(710- \\ & 800, n=2) \end{aligned}$ | $\begin{aligned} & 780(750- \\ & 810, n=2) \end{aligned}$ | $\begin{gathered} 648(570-750, \\ n=6) \end{gathered}$ |
| Body Width | 114-152 | 170 (143-235, n=27) | 185 (180-190, $\mathrm{n}=2)$ | $\begin{aligned} & 165(130- \\ & 200, n=2) \end{aligned}$ | $\begin{aligned} & 165(150- \\ & 180, n=2) \end{aligned}$ | $\begin{gathered} 145(120-180, \\ n=6) \end{gathered}$ |
| Haptor length | 61-95 | - | - | - | - | - |
| Haptor Width | 73-164 | 168 (141-197, n=22) | 150 (150-150, $\mathrm{n}=2)$ | - | $\begin{aligned} & 165(150- \\ & 180, n=2) \end{aligned}$ | - |
| Pharynx Length | 34-48 | - | $44(40-48, \mathrm{n}=2)$ | $\begin{gathered} 43(34-52, \\ n=2) \end{gathered}$ | $\begin{gathered} 44(38-50, \\ n=2) \end{gathered}$ | $36(29-40, n=5)$ |
| Pharynx Width | 32-41 | 47 (39-60, n=31) | $44(40-48, \mathrm{n}=2)$ | $\begin{gathered} 45(37-52, \\ n=2) \end{gathered}$ | $\begin{gathered} 44(38-50, \\ n=2) \\ \hline \end{gathered}$ | $32(25-39, n=5)$ |
| Penis Internal Length | 46-82 | - | $62(61-62, \mathrm{n}=2)$ | $\begin{gathered} 69(67-70, \\ n=2) \end{gathered}$ | $\begin{gathered} 68(63-73, \\ n=2) \end{gathered}$ | $\begin{gathered} 89(82-100, \\ n=17) \end{gathered}$ |
| Penis Cone Length | - | - | $10(9-11, \mathrm{n}=2)$ | $\begin{gathered} 12(10-13 \\ n=2) \end{gathered}$ | $\begin{gathered} 10(8-11, \\ n=2) \end{gathered}$ | $11(7-12, n=17)$ |
| Penis Tube Length | 27-55 | - | - | - | $33(\mathrm{n}=2)$ | $\begin{gathered} 35(29-44, \\ n=17) \end{gathered}$ |
| Penis Tube Diameter | - | - | $3(\mathrm{n}=2)$ | - | $4(\mathrm{n}=2)$ | $4(4-4.5, \mathrm{n}=16)$ |
| Penis (chamber + cone) Length | - | $84(75-100, \mathrm{n}=28)$ | - | - | - | - |
| Penis Filament Length | - | - | ${ }^{-}$ | ${ }^{-}$ | $\begin{gathered} 34(28-40, \\ n=2) \\ \hline \end{gathered}$ | - |
| Sclerotised Vagina Total Length | 34-41 | - | $39(38-39, \mathrm{n}=2)$ | $\begin{gathered} 44(43-44, \\ n=2) \\ \hline \end{gathered}$ | $\begin{gathered} 40(36-43, \\ n=2) \\ \hline \end{gathered}$ | $\begin{gathered} 50(45-54, \\ n=17) \end{gathered}$ |
| Primary Chamber External Diameter | - | - | $9(8-9, \mathrm{n}=2)$ | $7(\mathrm{n}=2)$ | $7(\mathrm{n}=2)$ | $9(7-10, \mathrm{n}=17)$ |
| Squamodisc Length | - | 56 (47-64, n=32) | $53(50-60, n=4)$ | $\begin{gathered} 54(53-55, \\ n=2) \\ \hline \end{gathered}$ | $\begin{gathered} 55(54-55, \\ n=2) \\ \hline \end{gathered}$ | - |
| Squamodisc Width | 41-62 | 55 (42-67, n=34) | $51(47-58, \mathrm{n}=4)$ | $\begin{gathered} 52(51-53, \\ n=2) \end{gathered}$ | $\begin{gathered} 51(48-54, \\ n=2) \\ \hline \end{gathered}$ | - |
| Squamodisc, Number of Rows | 10-12 | 10-12 (usually 12) | $10(9-12, \mathrm{n}=4)$ | - | - | - |
| Squamodisc, Number of Closed Rows | 2 | 2-3 | $2(\mathrm{n}=4)$ | $2(\mathrm{n}=2)$ | - | - |
| Ventral Anchor Outer Length | 39-55 | 56 (50-61, n=15) | $55(53-58, \mathrm{n}=4)$ | $\begin{gathered} 55(53-56, \\ n=2) \end{gathered}$ | $50(\mathrm{n}=2)$ | $\begin{gathered} 60(55-63, \\ n=15) \\ \hline \end{gathered}$ |
| Ventral Anchor Inner Length | - | ${ }^{-}$ | $55(53-57, \mathrm{n}=3)$ | $\begin{gathered} 55(54-55, \\ n=2) \end{gathered}$ | $\begin{gathered} 53(52-53 \\ n=2) \end{gathered}$ | $\begin{gathered} 58(53-62, \\ n=15) \end{gathered}$ |
| Dorsal Anchor Outer Length | 34-52 | $49(44-53, \mathrm{n}=16)$ | $50(50-51, \mathrm{n}=4)$ | $\begin{gathered} 49(43-59 \\ n=3) \\ \hline \end{gathered}$ | $45(\mathrm{n}=2)$ | $\begin{gathered} 50(47-53, \\ n=14) \\ \hline \end{gathered}$ |
| Dorsal Anchor Inner Length | - | - | $33(29-37, n=3)$ | $\begin{gathered} 29(20-33, \\ n=3) \end{gathered}$ | $\begin{gathered} 31(30-32, \\ n=2) \end{gathered}$ | $\begin{gathered} 33(31-39, \\ n=14) \\ \hline \end{gathered}$ |
| Ventral Bar Length | 50-78 | $77(72-84, \mathrm{n}=13)$ | $72(68-75, \mathrm{n}=2)$ | - | $\begin{gathered} 72(64-80 \\ n=2) \end{gathered}$ | $\begin{gathered} 100(93-106, \\ n=8) \end{gathered}$ |

(Continued)

Table 2. (Continued)

| Source | Santos, Buchmann \& Gibson, 2000 | Kritsky, Bakenhaster \& Adams, 2015 | Type-specimens, MNHN 249H-Tc167, 249H-Tc167 bis | Slides deposited in MNHN, HEL 465, HEL 466, HEL 470, HEL 471 |  | MNHN HEL560, Present study |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| Ventral Bar Width | - | - | 15 (14-15, $\mathrm{n}=2$ ) | - | 12 ( $\mathrm{n}=2$ ) | $18(14-20, n=8)$ |
| Lateral (dorsal) Bar Length | 50-69 | $67(61-72, \mathrm{n}=15)$ | $64(63-66, \mathrm{n}=4)$ | $\begin{gathered} 61(54-74, \\ n=4) \end{gathered}$ | $\begin{gathered} 59(56-63, \\ n=4) \end{gathered}$ | $\begin{gathered} 90(84-96, \\ n=15) \end{gathered}$ |
| Lateral (dorsal) Bar Width | - | - | $20(18-20, \mathrm{n}=4)$ | $\begin{gathered} 17(14-22, \\ n=4) \end{gathered}$ | $\begin{gathered} 16(15-18, \\ n=4) \end{gathered}$ | $\begin{gathered} 22(17-26, \\ n=15) \end{gathered}$ |

doi:10.1371/journal.pone.0159886.t002
anchors with distinct handle and guard (Fig 2E), outer length b 60 (55-63, $n=15$ ), inner length b 58 (53-62, $n=15$ ). Dorsal anchors with indistinct guard (Fig 2E), outer length b 50 (47-53, $\mathrm{n}=14$ ), inner length b $33(31-39, \mathrm{n}=14)$. Lateral (dorsal) bars, with flattened medial end (Fig 2E), length $b 90(84-96, n=15)$, maximum width b $22(17-26, n=15)$. Ventral bar (Fig 2E), length $b 100(93-106, n=8)$, maximum width, b18 $(14-20, n=8)$. Male copulatory organ a quadriloculate organ, first (anterior) chamber almost as sclerotised as the 3 others; fourth chamber forming short cone, prolonged by thin sclerotised tube; inner length b 89 (82-100, $\mathrm{n}=17$ ); cone length b 11 ( $7-12, \mathrm{n}=17$ ); tube length b 35 (29-44, $\mathrm{n}=17$ ); tube diameter b 4 (4-4.5, $\mathrm{n}=16$ ). Filament of variable length (Fig 2A and 2B).

Sclerotised vagina comprises sclerotised trumpet, primary canal, primary chamber, secondary canal and secondary chamber. Trumpet anterior, ring-shaped; primary canal with regular diameter and medium-thick wall, curved in its anterior part, sometimes looped; connection between primary canal and primary chamber posterior; primary chamber elongate, roughly parallel to primary canal but reversely oriented, its wall slightly thicker than primary canal; secondary canal well visible, often curved, its lumen thin, its wall slightly less sclerotised than chambers; secondary chamber roughly spherical, its wall thick, smaller than primary chamber; secondary chamber located just posterior to trumpet, often at the level of curve of primary canal (Fig 1A-1J). Total length of sclerotised vagina b $50(45-54, n=17)$. External diameter of primary chamber b 9 ( $7-10, \mathrm{n}=17$ ).

Comments. Pseudorhabdosynochus beverleyburtonae was originally described by Oliver as Cycloplectanum beverleyburtonae, from M. marginata (as Epinephelus guaza) in the Western Mediterranean Sea off France [10]. Kritsky \& Beverley-Burton (1986) regarded Cycloplectanum Oliver, 1968 a junior synonym of Pseudorhabdosynochus Yamaguti, 1958 and placed several species, including C. beverleyburtonae, in the latter genus [28]. Most subsequent authors have accepted Pseudorhabdosynochus as the valid genus; however, Oliver continued to designate the species as C. beverleyburtonae [25].

Pseudorhabdosynochus beverleyburtonae parasitises M. marginata, which is a trans-Atlantic grouper fish. The original description does not include any measurements [10]. Measurements in specimens from various origins show differences (Table 2) but these should be attributed to various methods of fixation and degree of flattening, as is usual in diplectanids [20]. The morphology of the sclerotised vagina is similar in all specimens, including our newly examined specimens from Tunisia. We conclude, as previous authors [2, 11], that the populations assigned to M. marginata from the Mediterranean Sea and both sides of the Atlantic Ocean are conspecific. Tunisia is a new geographical record for the species, and is the first record on the southern shore of the Mediterranean Sea.

Differential diagnosis. Pseudorhabdosynochus beverleyburtonae is close to P. oliveri n. sp., $P$. sosia and $P$. hayet n. sp. in terms of general morphology of body, presence of two circular innermost rows of rodlets in squamodiscs and general morphology of the sclerotised vagina. It


Fig 2. Pseudorhabdosynochus beverleyburtonae from Mycteroperca marginata in the Mediterranean Sea and Atlantic Ocean, quadriloculate organ, squamodiscs and haptoral hard parts. A, quadriloculate organ, MNHN 249H-Tc167, typespecimen. B, quadriloculate organ, voucher MNHN HEL466. C, ventral squamodisc, D, dorsal squamodisc, MNHN $249 \mathrm{H}-\mathrm{Tc} 167$ bis, type-specimen. E, haptoral hard parts, new specimen from Tunisia. A, C, D, carmine. B, Gray and Wess medium. E, Berlese.
doi:10.1371/journal.pone.0159886.g002
is separated from P. oliveri, which also occurs on M. marginata, by the shape of its primary canal (narrow in P. beverleyburtonae vs. wide in $P$. oliveri), and from $P$. sosia and $P$. hayet by the shape of its primary chamber, wider and more sclerotised in the two latter species.

## Pseudorhabdosynochus oliveri n. sp.

urn:lsid:zoobank.org:act:8BDBF329-0700-44A1-B720-6C172ECE0701
Type-host: Mycteroperca marginata (Lowe) (Perciformes, Epinephelidae)
Site of infection: Gills
Type-locality: Off Cap Béar, Mediterranean Sea, France; date 4 February 1965.
Material examined: 4 voucher specimens MNHN HEL68 OLI 8-1 to 8-4 on slides, labelled as Cycloplectanum beverleyburtonae by Guy Oliver.

Holotype: MNHN HEL68 OLI 8-2
Paratypes: MNHN HEL68 OLI 8-1, HEL68 OLI 8-3, HEL68 OLI 8-4
Etymology: This species is named for Guy Oliver, French parasitologist, in recognition of his research on fish monogeneans.

Description (Figs 3 and 4). Measurements of 4 specimens in carmine. Body length $h$ 1100, c 1050 (1000-1100, $n=2$ ), including haptor; maximum width h 250, c 235 (220-250, $\mathrm{n}=2$ ) at level of ovary (Fig 3A). Tegument smooth. Anterior region with 3 pairs of head organs and 2 pairs of dorsal eye-spots, distance between outer margins of anterior eye-spots h 50, c 42 ( $35-50, \mathrm{n}=3$ ), of posterior eye-spots h 40, c $36(30-40, \mathrm{n}=3)$. Pharynx median, ovate, length c $56(50-62, \mathrm{n}=2)$, width $\mathrm{c} 59(56-62, \mathrm{n}=2)$. Oesophagus very short or absent. Two simple lateral intestinal caeca not united posteriorly. Haptor bearing 2 squamodiscs, 2 pairs of lateral anchors, 1 ventral bar and 2 lateral (dorsal) bars, and 14 marginal hooklets. Squamodiscs with 10-11 concentric rows of rodlets; the 2 innermost rows form circles. Rodlets with visible spurs ('éperons') (Fig 3H). Squamodisc length c $72(\mathrm{n}=2)$, width c $85(81-88, n=2)$. Ventral anchors with distinct handle and guard (Fig 3E), outer length h 70, c $72(67-82, \mathrm{n}=8$ ), inner length h 73, c $73(70-78, n=8)$. Dorsal anchors with indistinct guard (Fig 3G), outer length c $66(63-73, \mathrm{n}=6)$, inner length $\mathrm{c} 47(43-50, \mathrm{n}=5)$. Lateral (dorsal) bars, with flattened medial end (Fig 3F), length h 85, c $87(84-91, n=8)$, maximum width $h 24, \mathrm{c} 24(23-25, \mathrm{n}=8)$. Ventral bar with constricted median portion (Fig 3D), length h 104, c $102(97-105, n=4)$, maximum width, h 20, c $20(16-22$, $\mathrm{n}=4)$. Testis subspherical, posterior, intercaecal. Male copulatory organ a quadriloculate organ, first (anterior) chamber as sclerotised as the 3 others; fourth chamber forming short cone, prolonged by thin sclerotised tube. Inner length c 133 (128-138, $\mathrm{n}=2$ ); cone length $\mathrm{c} 9(8-10, \mathrm{n}=2)$; tube length $\mathrm{c} 35(33-36, \mathrm{n}=2)$; tube diameter c $6(5-6, n=2)$. Filament not seen (Fig 3B).

Vitelline follicles lateral, coextensive with intestinal caeca and contiguous posterior to testis. Ovary on right side, looping dorsoventrally around right intestinal caecum.

Sclerotised vagina comprises sclerotised trumpet, primary canal, primary chamber, secondary canal and secondary chamber. Trumpet in anterior part; primary canal with thick wall, curved in its anterior part, enlarged in its posterior part with wide lumen; connection between primary canal and primary chamber posterior; primary chamber elongate, roughly parallel to primary canal but reversely oriented, its wall the same thickness as primary canal; secondary canal short, its lumen thin; secondary chamber located just posterior to trumpet, roughly spherical, its wall thick, smaller than primary chamber (Figs 3C and 4A-4C). Total length of sclerotised vagina c $63(59-65, n=4)$. Diameter of primary canal h 11, c $14(11-16, n=4)$.

Comments. This new species was found among voucher specimens labelled as Cycloplectanum beverleyburtonae by Guy Oliver (now P. beverleyburtonae). These specimens were deposited by Guy Oliver in the MNHN collections after his retirement and were not mentioned


Fig 3. Pseudorhabdosynochus oliveri n. sp. from Mycteroperca marginata off France. A, composite, dorsal view. B, male quadriloculate organ. C, vagina. D, ventral bar. E, ventral anchor. F, lateral (dorsal) bar. G, dorsal anchor. H, dorsal squamodisc. All in carmine.
doi:10.1371/journal.pone.0159886.g003


Fig 4. Pseudorhabdosynochus oliveri n. sp. from Mycteroperca marginata off France, different shapes of sclerotised vagina according to specimens and orientation. A, MNHN HEL68 OLI 8-1. B, MNHN HEL68 OLI 8-3. C, MNHN HEL68 OLI 8-4. All in carmine.
doi:10.1371/journal.pone.0159886.g004
in the original description. This is not the first time that new species of Pseudorhabdosynochus are found among museum specimens labelled as other species; previous cases include $P$. satyui Justine, 2009, found among specimens of P. epinepheli Yamaguti, 1958 [29], P. williamsi Kritsky et al., 2015 found among specimens of $P$. monaensis Dyer et al., 1994, and P. justinella Kritsky et al., 2015 found among specimens of P. yucatanensis Vidal-Martinez et al., 1997. In the latter case, Kritsky et al. (2015) demonstrated that the original description of P. yucatanensis [30] contained drawings of specimens from both species; in the case of the $P$. beverleyburtonae/P. oliveri couple, we did not find any indication of $P$. oliveri being included in the original description of $P$. beverleyburtonae.

Although P. oliveri and P. beverleyburtonae apparently co-occurred on dusky groupers from off France examined by Oliver, we did not find the former in our fish specimen from Tunisia.

Differential diagnosis. Pseudorhabdosynochus oliveri is close to $P$. beverleyburtonae, $P$. sosia and $P$. hayet n. sp. in terms of general morphology of body, presence of two circular innermost rows of rodlets in squamodiscs and general morphology of the sclerotised vagina. It is separated from the three others by the development of its primary canal, which is wide, vs. thin, and the small primary chamber, vs. large in the other three species.

## Pseudorhabdosynochus sosia Neifar \& Euzet, 2007

Type-host: Goldblotch grouper, Mycteroperca costae (Steindachner) (Perciformes, Epinephelidae); synonyms: Epinephelus alexandrinus (Valenciennes), Epinephelus costae.

Molecular identification of fish via DNA barcoding: Four specimens from Tunisia were sequenced (Table 1), including three new and one already sequenced as KT805240 [31]. All sequences had $99-100 \%$ similarity with sequences already identified as M. costae in GenBank, confirming the morphological identification of the host.

Site of infection: Gills
Type-locality: Off Sfax, Tunisia.
Other localities: Off Zarzis (Tunisia) [3]; off Dakar (Senegal) [3]; Tripoli (fish market), Libya (present study).

Material examined: Holotype, MNHN HEL11-Th84 (darkened picrate slide); paratypes, MNHN HEL12-Th85, MNHN HEL13-Th86 (paratype specimens marked on slides); voucher specimens collected by Neifar and Euzet (MNHN HEL561) from off Tunisia; new specimens collected from off Tunisia (MNHN HEL562) (Table 3).

Prevalence: In our specimens from Tunisia, $4 / 4$ (100\%); from Libya, $1 / 1$ ( $100 \%$ ).
Molecular characterisation: COI sequences were obtained for 4 specimens from Tunisia (Table 1).

Redescription (Fig 5). Measurements of 30 specimens in Berlese. Body length b 1,039 (800-1,200, $\mathrm{n}=12$ ), including haptor; maximum width b $219(150-330, \mathrm{n}=11)$ at level of ovary. Tegument smooth. Anterior region with 3 pairs of head organs and 2 pairs of dorsal eye-spots, distance between outer margins of anterior eye-spots $b 39(27-63, n=22)$, of posterior eye-spots b $30(15-38, n=26)$. Pharynx median, subspherical. Oesophagus very short or absent. Two simple lateral intestinal caeca not united posteriorly. Haptor bearing 2 squamodiscs, 2 pairs of lateral anchors, 1 ventral bar and 2 lateral (dorsal) bars and 14 marginal hooklets; width b $198(180-210, n=4)$. Squamodiscs with $10-12$ concentric rows of rodlets; 2 innermost rows forming circles. Rodlets with visible spurs ('éperons') (Fig 5A and 5B). Squamodisc length $b 63(52-80, n=23)$, width $b 62(53-73, n=23)$. Ventral anchors with distinct handle and guard (Fig 5C), outer length b $48 \pm 3(42-53, n=33)$, inner length b $47 \pm 5.2$ ( $40-74$, $\mathrm{n}=35$ ). Dorsal anchors with indistinct guard (Fig 5C), outer length b $40 \pm 2.1$ (33-43, $\mathrm{n}=38$ ), inner length $\mathrm{b} 25 \pm 2.4(21-32, \mathrm{n}=36)$. Lateral (dorsal) bars with wide flattened medial extremity and cylindrical lateral extremity (Fig 5C), length b $72 \pm 4.7$ ( $60-80, \mathrm{n}=40$ ), width b $21 \pm 1.9$ (17-25, $n=40$ ). Ventral bar (Fig 5C), length b $88(68-100, n=20)$, width b $16(13-20, n=20)$. Testis subspherical, posterior, intercaecal. Male copulatory organ a quadriloculate organ, length $b 92(63-120, \mathrm{n}=23)$; divided into four chambers, fourth chamber more sclerotised than 3 anterior chambers; fourth chamber ends in short sclerotised cone, length b 8 (5-10, $\mathrm{n}=26)$, prolonged by sclerotised tube, length b $30(24-39, \mathrm{n}=27)$, diameter b $3(3-4, \mathrm{n}=27)$; end of tube prolonged by thin unsclerotised filament, length b 28 (20-33, $n=3$ ) (Fig 5D).

Sclerotised vagina comprises sclerotised trumpet, primary canal, primary chamber, secondary canal and secondary chamber. Trumpet anterior, ring-shaped; primary canal with regular diameter and medium-thick wall, curved or S-shaped; connection between primary canal and primary chamber posterior; primary chamber ovoid or pear-shaped, roughly parallel to primary canal but reversely oriented, its wall thick; secondary canal well visible, its lumen thin, its wall less sclerotised than chambers; secondary chamber spherical, its wall thick, smaller than primary chamber; secondary chamber located just posterior to trumpet (Fig 5E-5H). Sclerotised vagina length $b 44 \pm 4.3(35-58, n=30)$, external diameter of primary chamber $b 15$ (11$17, \mathrm{n}=28$ ).

Table 3. Measurements of $P$. sosia and $P$. hayet n . sp.

| Species | P. sosia Neifar \& Euzet, 2007 |  |  |  | P. hayet n . sp. |  |  |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| Hosts | M. costae |  |  |  | M. rubra |  |  |
| Source | Neifar and <br> Euzet, 2007 | Paratypes, MNHN HEL12-Th 85, MNHN HEL13-Th 86 | MNHN HEL561, <br> Vouchers collected by Neifar \& Euzet | MNHN HEL562, <br> Vouchers, newly collected specimens | MNHN HEL563- <br> 564, Specimens collected by Neifar \& Euzet | MNHN HEL565, <br> Specimens collected by Neifar | MNHN 306HG, Specimens from Euzet collection |
| Locality | Off Sfax, Tunisia | Off Sfax, Tunisia | Off Sfax and Zarzis, Tunisia | Off Sfax, Tunisia | Off Dakar, Senegal | Off Sfax, Tunisia | Off Dakar, Senegal |
| Methods | Picrateglycerine, carmine, Berlese | Picrate-glycerine | Picrate-glycerine | Berlese | Carmine | Picrate-glycerine | Berlese |
| n | 34 | 9 | 19 | 30 | 4 | 3 | 5 |
| Body Length | $\begin{gathered} 1,100(800- \\ 1,300, n=24) \end{gathered}$ | $\begin{gathered} 398(347-438, \\ n=9) \end{gathered}$ | $\begin{gathered} 733(550-950, \\ n=17) \end{gathered}$ | $\begin{gathered} 1,039(800- \\ 1,200, n=12) \end{gathered}$ | $\begin{gathered} 795(710-920, \\ n=4) \end{gathered}$ | $\begin{gathered} 970(830-1,110, \\ n=2) \end{gathered}$ | $\begin{gathered} 1,150(1,100- \\ 1,200, n=2) \end{gathered}$ |
| Body Width | $\begin{gathered} 270(170- \\ 350, \mathrm{n}=24) \end{gathered}$ | $79(75-91, \mathrm{n}=9)$ | $\begin{gathered} 150(120-180, \\ n=16) \end{gathered}$ | $\begin{gathered} 219(150-330, \\ n=11) \end{gathered}$ | $\begin{gathered} 215(180-260, \\ n=4) \end{gathered}$ | $\begin{gathered} 180(130-230, \\ n=2) \end{gathered}$ | $\begin{gathered} 245(240-250, \\ n=2) \end{gathered}$ |
| Haptor Width | - | $85(75-98, \mathrm{n}=9)$ | $\begin{gathered} 147(130-166, \\ n=13) \end{gathered}$ | $\begin{gathered} 198(180-210, \\ n=4) \end{gathered}$ | $\begin{gathered} 165(140-180, \\ n=4) \end{gathered}$ | - | - |
| Pharynx Length | $\begin{gathered} 66(50-80, \\ n=14) \end{gathered}$ | $28(18-42, \mathrm{n}=9)$ | $\begin{gathered} 38(25-50, \\ n=18) \end{gathered}$ | - | $46(42-48, n=4)$ | - | - |
| Pharynx Width | $\begin{gathered} 57(40-70, \\ n=14) \\ \hline \end{gathered}$ | 24 (18-30, n = 9) | $\begin{gathered} 34(25-40, \\ n=18) \end{gathered}$ | - | $49(45-52, \mathrm{n}=4)$ | - | - |
| Penis Internal Length | $\begin{aligned} & 110 \pm 3(90- \\ & 130, n=29) \end{aligned}$ | $84(75-91, \mathrm{n}=8)$ | $\begin{gathered} 81(65-94, \\ n=15) \end{gathered}$ | $\begin{gathered} 92(63-120, \\ n=23) \end{gathered}$ | $85(83-92, \mathrm{n}=4)$ | $89(86-94, n=3)$ | $\begin{gathered} 150(140-165, \\ n=5) \end{gathered}$ |
| Penis Cone Length | $\begin{gathered} 13 \pm 1(10-17 \\ n=29) \end{gathered}$ | $11(8-15, \mathrm{n}=8)$ | $8(6-10, n=14)$ | $8(5-10, \mathrm{n}=26)$ | $7(5-9, n=4)$ | $7(6-7, n=2)$ | 10 ( $\mathrm{n}=5$ ) |
| Penis Tube Length | $\begin{gathered} 38 \pm 2(25-45, \\ n=29) \\ \hline \end{gathered}$ | $29(24-42, n=6)$ | $23(10-27, n=9)$ | $\begin{gathered} 30(24-39, \\ n=27) \\ \hline \end{gathered}$ | $18(15-20, \mathrm{n}=2)$ |  | $26(5-33, \mathrm{n}=5)$ |
| Penis Tube Diameter | - | $4(4-4, n=8)$ | $4(3-4, n=13)$ | 3 (3-4, $n=27)$ | $4(3-5, n=4)$ | $4(\mathrm{n}=2)$ | $5(4-5, n=3)$ |
| Penis Filament Length | - | - | - | $28(20-33, n=3)$ | $30(\mathrm{n}=2)$ | - | - |
| Sclerotised Vagina Total Length | $\begin{gathered} 48 \pm 1(40-55, \\ n=34) \end{gathered}$ | $35(32-38, \mathrm{n}=9)$ | $\begin{gathered} 36(26-40, \\ n=19) \end{gathered}$ | $\begin{gathered} 44 \pm 4.3(35-58 \\ n=30) \end{gathered}$ | $46(45-49, n=4)$ | $43(40-45, n=3)$ | $61(54-70, n=5)$ |
| Primary Chamber External Diameter | - | $12(11-14, \mathrm{n}=9)$ | $\begin{gathered} 13(10-15, \\ n=19) \end{gathered}$ | $\begin{gathered} 15(11-17, \\ \mathrm{n}=28) \end{gathered}$ | 10 ( $\mathrm{n}=4$ ) | $9(7-12, n=3)$ | $15(14-16, n=5)$ |
| Squamodisc Length |  | 46 (44-49, n = 7) | $\begin{gathered} 51(40-62, \\ n=15) \end{gathered}$ | $\begin{gathered} 63(52-80, \\ \mathrm{n}=23) \\ \hline \end{gathered}$ | $53(40-62, \mathrm{n}=8)$ | $49(48-50, n=2)$ | - |
| Squamodisc Width | $\begin{gathered} 67(60-80, \\ n=12) \end{gathered}$ | $44(39-45, \mathrm{n}=7)$ | $\begin{gathered} 45(29-51, \\ n=14) \end{gathered}$ | $\begin{gathered} 62(53-73, \\ n=23) \end{gathered}$ | $52(38-58, \mathrm{n}=8)$ | $63(62-63, n=2)$ | - |
| Squamodisc, Number of Rows | 10-11 | $11(10-11, \mathrm{n}=5)$ |  | $\begin{gathered} 11(10-12, \\ \mathrm{n}=22) \end{gathered}$ | $10(10-11, \mathrm{n}=4)$ | 10 ( $\mathrm{n}=1$ ) | - |
| Squamodisc, Number of Closed Rows | 1-2 | $2(\mathrm{n}=5)$ | $2(\mathrm{n}=11)$ | $2(\mathrm{n}=22)$ | $2(\mathrm{n}=2)$ | - | - |
| Ventral Anchor Outer Length | $\begin{gathered} 60 \pm 1(55-67, \\ n=34) \\ \hline \end{gathered}$ | $41(36-45, n=14)$ | $\begin{gathered} 44(41-48, \\ n=27) \end{gathered}$ | $\begin{gathered} 48 \pm 3(42-53, \\ n=33) \end{gathered}$ | $49(45-52, \mathrm{n}=2)$ | $46(43-48, n=4)$ | $51(48-55, n=8)$ |
| Ventral Anchor Inner Length | - | 41 (39-45, n= 13) | $\begin{gathered} 44(38-50, \\ n=28) \end{gathered}$ | $\begin{gathered} 47 \pm 5.2(40-74, \\ n=35) \end{gathered}$ | $48(45-50, \mathrm{n}=6)$ | $45(40-48, n=4)$ | $48(38-52, n=8)$ |
| Dorsal Anchor Outer Length | $\begin{gathered} 52(50-57, \\ \mathrm{n}=28) \\ \hline \end{gathered}$ | 37 (33-41, n= 15) | $\begin{gathered} 38 \pm 2(35-41, \\ \mathrm{n}=29) \end{gathered}$ | $\begin{gathered} 40 \pm 2.1(3-43, \\ n=38) \\ \hline \end{gathered}$ | $40(38-42, n=7)$ | $42(40-43, n=4)$ | $43(41-45, n=7)$ |

(Continued)

ONE

Table 3. (Continued)

| Species | P. sosia Neifar \& Euzet, 2007 |  |  |  | P. hayet n. sp. |  |  |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| Hosts | M. costae |  |  |  | M. rubra |  |  |
| Dorsal Anchor Inner Length | - | $24(23-27, n=13)$ | $\begin{gathered} 24 \pm 1.4(22-28 \\ n=29) \end{gathered}$ | $\begin{gathered} 25 \pm 2.4(21-32, \\ n=36) \end{gathered}$ | 25 (24-26, n=7) | $24(23-26, \mathrm{n}=4)$ | $26(25-30, n=7)$ |
| Ventral Bar Length | $\begin{gathered} 83 \pm 3(70- \\ 105, n=34) \end{gathered}$ | 70 (57-128, $\mathrm{n}=7$ ) | $\begin{gathered} 61(55-73, \\ n=15) \end{gathered}$ | $\begin{gathered} 88(68-100, \\ n=20) \end{gathered}$ | 69 (63-73, $\mathrm{n}=4)$ | $77(68-86, \mathrm{n}=2)$ | $\begin{gathered} 93(85-100, \\ n=3) \end{gathered}$ |
| Ventral Bar Width | $\begin{gathered} 12 \pm 1(7-17 \\ \mathrm{n}=34) \end{gathered}$ | $8(7-10, \mathrm{n}=7)$ | $10(7-13, n=15)$ | $\begin{gathered} 16(13-20, \\ \mathrm{n}=20) \\ \hline \end{gathered}$ | $13(\mathrm{n}=4)$ | $11(\mathrm{n}=2)$ | $18(17-20, n=3)$ |
| Lateral Bar Length | $\begin{gathered} 66 \pm 2(58-73 \\ n=36) \end{gathered}$ | $47(45-49, n=18)$ | $\begin{gathered} 50 \pm 4.2(42-63 \\ n=33) \end{gathered}$ | $\begin{gathered} 72 \pm 4.7(60-80 \\ n=40) \end{gathered}$ | $53(47-55, \mathrm{n}=8)$ | $54(53-56, \mathrm{n}=4)$ | $75(72-78, n=8)$ |
| Lateral Bar Width | $\begin{gathered} 14 \pm 2(12-22 \\ n=36) \end{gathered}$ | $12(9-15, \mathrm{n}=18)$ | $\begin{gathered} 13 \pm 1(10-15 \\ n=33) \end{gathered}$ | $\begin{gathered} 21 \pm 1.9(17-25 \\ n=40) \end{gathered}$ | $16(13-18, n=8)$ | $15(14-16, n=4)$ | $26(17-30, n=8)$ |

Bold: important differences for species differentiation.
doi:10.1371/journal.pone.0159886.t003

Differential diagnosis. By the morphology of its sclerotised vagina, $P$. sosia is close to $P$. beverleyburtonae, P. oliveri n. sp. and $P$. hayet n . sp. Indeed, Neifar \& Euzet named the species for its resemblance with P. beverleyburtonae [3]. Pseudorhabdosynochus sosia and P. beverleyburtonae have similar vaginal lengths but can be differentiated, as indicated by Neifar \& Euzet, 2007, by the external diameter of the primary chamber (larger in $P$. sosia) and its shape (pearshaped in P. sosia vs elongated in P. beverleyburtonae), and by the shape and length of the primary canal (longer and more coiled in P. beverleyburtonae). Pseudorhabdosynochus sosia is readily distinguished from $P$. oliveri by the shape of its primary canal (thin vs. wide in the latter).

The morphology of the vagina is very similar in $P$. sosia and $P$. hayet, but $P$. sosia differs by smaller male copulatory organ (see details under P. hayet, and Table 3), different host (M. costae for $P$. sosia vs M. rubra for $P$. hayet) (Table 1), and divergent COI sequences (Table 4).

## Pseudorhabdosynochus hayet n . sp.

urn:lsid:zoobank.org:act:10C2CE08-E772-4817-A4BC-EF7CF91ED44A
Synonym: Pseudorhabdosynochus sp. of Chaabane et al., 2015 (Table 1 in [6])
Type-host: Mottled grouper Mycteroperca rubra (Bloch) (Perciformes, Epinephelidae)
Molecular identification of fish via DNA barcoding: Two fish were sequenced, one from Libya (new sequence KX255748) and one from Tunisia (sequence already published as KU739518 [24] (Table 1). The sequences were similar (99\% identity) to sequences of the same fish species, therefore confirming the identity of the host fish.

Site of infection: Gills.
Type-locality: Off Dakar, Senegal.
Other localities: Sfax and Tunis (fish markets), Tunisia (present study)
Material examined: Specimens off Sfax and Dakar (Table 3).
Specimens deposited: Holotype, MNHN HEL563 (carmine), from off Senegal, material collected by Neifar \& Euzet, precise date unknown (2002-2004); paratypes MNHN HEL564 from off Senegal (carmine), material collected by Neifar \& Euzet, precise date unknown (20022004); paratypes MNHN 306HG from off Senegal (Berlese), material collected by Neifar \& Euzet (2004) in Euzet's collection; paratypes MNHN HEL565 from off Tunisia collected by Neifar (picrate). The holotype was chosen among carmine slides because slides in Canada balsam are permanent. One specimen collected off Tunis, Tunisia, used for molecular analysis and destroyed.


Fig 5. Pseudorhabdosynochus sosia from Mycteroperca costae in the Mediterranean Sea and Eastern Atlantic Ocean, sclerotised parts. A, ventral squamodisc. B, dorsal squamodisc. C, haptor hard parts. D, quadriloculate organ. E, F, vaginae. A-F, newly collected specimens from Tunisia, MNHN HEL562. G, H, vaginae, voucher specimens MNHN HEL561 from Tunisia. A-F, Berlese. G, H, picrate-glycerine.
doi:10.1371/journal.pone.0159886.g005

ONE

Table 4. Estimates of evolutionary divergence between COI sequences of Pseudorhabdosynochus spp.

| Species | Accession number |  | [1] | [2] | [3] | [4] | [5] | [6] | [7] | [8] |
| :--- | :--- | :--- | :--- | :--- | :--- | :--- | :--- | :--- | :--- | :--- |
| Pseudorhabdosynochus hayet n. sp | KX255746 | $[1]$ |  |  |  |  |  |  |  |  |
| Pseudorhabdosynochus sosia | KX255744 | $[2]$ | $\mathbf{0 . 1 3}$ |  |  |  |  |  |  |  |
| Pseudorhabdosynochus sosia | KX255743 | $[3]$ | $\mathbf{0 . 1 4}$ | 0.00 |  |  |  |  |  |  |
| Pseudorhabdosynochus sosia | KX255741 | $[4]$ | $\mathbf{0 . 1 4}$ | 0.00 | 0.00 |  |  |  |  |  |
| Pseudorhabdosynochus sosia | KX255742 | $[5]$ | $\mathbf{0 . 1 3}$ | 0.00 | 0.00 | 0.00 |  |  |  |  |
| Pseudorhabdosynochus regius | KX255745 | $[6]$ | 0.26 | 0.21 | 0.21 | 0.21 | 0.21 |  |  |  |
| Pseudorhabdosynochus sulamericanus | KT023569 | $[7]$ | 0.21 | 0.19 | 0.20 | 0.20 | 0.19 | 0.22 |  |  |
| Pseudorhabdosynochus cyanopodus | JQ400135 | $[8]$ | 0.26 | 0.20 | 0.20 | 0.20 | 0.20 | 0.23 | 0.19 |  |
| Lamellodiscus ignoratus | JF427655 | $[9]$ | 0.39 | 0.34 | 0.34 | 0.34 | 0.34 | 0.31 | 0.38 | 0.38 |

The analysis involved 9 nucleotide sequences, and there were a total of 290 positions in the final dataset. Bold: differences between $P$. hayet n . sp . and $P$. sosia.
doi:10.1371/journal.pone.0159886.t004

Prevalence: In Tunisia, 1/1, in Libya, 0/1.
Molecular characterisation: A COI sequence was obtained for a specimen from off Tunis, Tunisia (Table 1).

Etymology: The specific name, hayet, is from Arabic (حياة). The species is named after the mother of the first author. Name in apposition, invariable.

Description (Figs 6 and 7). Measurements based on 12 specimens in picrate, Berlese and carmine; holotype in carmine. Body length h 730, c 795 (710-920, $n=4$ ), p 970 (830-1,110, $\mathrm{n}=2), \mathrm{b} 1,150(1,100-1,200, \mathrm{n}=2)$ including haptor; maximum width h 200, c 215 (180-260, $\mathrm{n}=4)$, b 245 ( $240-250, \mathrm{n}=2$ ), at level of ovary (Fig 6A). Tegument smooth. Anterior region with 3 pairs of head organs and 2 pairs of dorsal eye-spots, distance between outer margins of anterior eye-spots h 42, c $39(32-43, n=4)$, b $51(30-62, n=3)$, of posterior eye-spots h 28 , c $28(28-29, n=4)$, b $32(20-40, n=3)$. Pharynx median, subspherical length h 42, c $46(42-48$, $\mathrm{n}=4)$, width c $49(45-52, \mathrm{n}=4)$. Oesophagus very short or absent. Two simple lateral intestinal caeca not united posteriorly. Haptor bearing 2 squamodiscs, 2 pairs of lateral anchors, 1 ventral bar and 2 lateral (dorsal) bars and 14 marginal hooklets; width h 170, c $165(140-180, \mathrm{n}=4)$. Squamodiscs with $10-11$ concentric rows of rodlets; 2 innermost rows forming circle. Rodlets with visible spurs ('éperons') (Fig 6H). Squamodisc length h 50, p 49 (48-50, n = 2), c 53 (40$62, \mathrm{n}=8)$, width $\mathrm{h} 51, \mathrm{p} 63(62-63, \mathrm{n}=2)$, $\mathrm{c} 52(38-58, \mathrm{n}=8)$. Ventral anchors with distinct handle and guard (Fig 6E), outer length c $49(45-52, n=2)$, p $46(43-48, n=4)$, b $51(48-55$, $\mathrm{n}=8)$, inner length $\mathrm{h} 46, \mathrm{c} 48(45-50, \mathrm{n}=6), \mathrm{p} 45(40-48, \mathrm{n}=4)$, $\mathrm{b} 48(38-52, \mathrm{n}=8)$. Dorsal anchors with indistinct guard (Fig 6F), outer length h 39, c $40(38-42, \mathrm{n}=7)$, p 42 (40-43, $\mathrm{n}=4)$, b $43(41-45, \mathrm{n}=7)$, inner length h 26 , c $25(24-26, \mathrm{n}=7)$, p $24(23-26, \mathrm{n}=4)$, b 26 ( $25-$ $30, \mathrm{n}=7$ ). Lateral (dorsal) bars with wide flattened medial extremity and cylindrical lateral extremity (Fig 6G), length h 55, c $53(47-55, \mathrm{n}=8)$, p $54(53-56, \mathrm{n}=4)$, b $75(72-78, \mathrm{n}=8)$, width h 18, c16 $(1-18, \mathrm{n}=8)$, p $15(14-16, \mathrm{n}=4)$, b $26(17-30, \mathrm{n}=8)$. Ventral bar with blunt extremities (Fig 6D), length h 73, c $69(63-73, \mathrm{n}=4)$, b $93(85-100, \mathrm{n}=3)$, width h 13, c 13 ( $\mathrm{n}=4$ ), b $18(17-20, \mathrm{n}=3)$. Testis subspherical, posterior, intercaecal. Male copulatory organ a quadriloculate organ, length h 92, c $85(83-92, n=4)$, p $89(86-94, n=3)$, b $150(140-165$, $\mathrm{n}=5$ ); divided into four chambers, fourth (posterior) chamber more sclerotised than 3 anterior chambers, finishes with short sclerotised cone, cone length h 7, c $7(5-9, n=4), p 7(6-7$, $\mathrm{n}=2)$, b $10(\mathrm{n}=5)$ prolonged by sclerotised tube, tube length c $18(15-20, \mathrm{n}=2), \mathrm{b} 26(5-33$, $\mathrm{n}=5)$, tube diameter $\mathrm{h} 4.5, \mathrm{c} 4(3-5, \mathrm{n}=4), \mathrm{p} 4(\mathrm{n}=2)$, $\mathrm{b} 5(4-5, \mathrm{n}=3)$; end of tube sometimes prolonged by thin unsclerotised filament, length c $30(\mathrm{n}=2)$ (Fig 6B). Ovary subequatorial,


Fig 6. Pseudorhabdosynochus hayet n. sp. from Mycteroperca rubra off Senegal. A, holotype, dorsal view. B, male quadriloculate organ. C, vagina. D, ventral bar. E, ventral anchor. F, dorsal anchor. G, lateral (dorsal) bar. H, ventral squamodisc. All in carmine.
doi:10.1371/journal.pone.0159886.g006
intercaecal, pre-testicular, encircles right caecum. Vitelline follicles present from pre-pharynx level to haptoral peduncle.

Sclerotised vagina comprises sclerotised trumpet, primary canal, primary chamber, secondary canal and secondary chamber. Trumpet anterior, ring-shaped; primary canal with regular diameter and medium-thick wall, curved or S-shaped; connection between primary canal and primary chamber posterior; primary chamber pear-shaped or elongate, roughly parallel to primary canal but reversely oriented, its wall thick; secondary canal well visible, its lumen narrow; secondary chamber spherical, smaller than primary chamber; wall of secondary chamber thick; secondary chamber located just posterior to trumpet (Figs 6C and 7A-7D). Sclerotised vagina length $h 45$, c $46(45-49, n=4), p 43(40-45, n=3)$, b $61(54-70, n=5)$, diameter of primary chamber external h 10, c $10(\mathrm{n}=4)$, p $9(7-12, \mathrm{n}=3)$, b $15(14-16, \mathrm{n}=5)$.

Differential diagnosis. Pseudorhabdosynochus hayet n . sp. can be distinguished from $P$. oliveri n . sp. by the shape of its primary canal (thin vs. wide in the latter), and from P. beverleyburtonae by the shape of its primary chamber (wide vs. thin in the latter). The morphology of the sclerotised vagina and other organs is very similar to that of $P$. sosia. The two species can, however, be distinguished by larger dimensions of the vagina in $P$. hayet. It is well known that measurements of sclerotised parts change with degree of flattening, i.e. the method for preparing slides [20]. Measurements between the two species are different when compared between specimens prepared with the same methods (Table 2): picrate (43 in P. hayet vs 35-36 in P. sosia) and Berlese ( 61 vs 44 ). The male copulatory organ is longer in $P$. hayet: picrate ( 89 vs $81-84$ in P. sosia), and Berlese ( 150 vs 92 ). In addition, the hosts are different (M. rubra for $P$. hayet vs M. costae for P. sosia) (Table 1). The COI sequences are divergent (Table 4).


Fig 7. Pseudorhabdosynochus hayet $\mathbf{n}$. sp. from Mycteroperca rubra, various shapes of sclerotised vagina according to specimens, orientation and preparation. A-D, different forms of vagina. A-C, Berlese. D, carmine.
doi:10.1371/journal.pone.0159886.g007

## The 'beverleyburtonae group'

The four species assigned to Pseudorhabdosynochus described and/or redescribed herein, namely P. beverleyburtonae and P. oliveri n. sp. from M. marginata, P. sosia from M. costae, and $P$. hayet n . sp. from M. rubra, exhibit great similarity in the morphology of the sclerotised vagina, the main character on which Pseudorhabdosynochus species are distinguished $[3,4,6$, 32, 33]. Fig 8 shows a comparison of the sclerotised vagina in all four species; colours were used to show the various parts of the vaginae and the homologies between different species. In addition, all these four species are characterised by a similar structure of the squamodiscs, with


General diagram
Fig 8. Homologies of the various parts of the sclerotised vaginae illustrated by coloured diagrams. The same colours are used in each diagram for the same parts, to show homologies between species. The nomenclature of vaginal parts and the general diagram are from Justine (2007) [1]. All vaginae drawn with same sizes-magnifications vary.
doi:10.1371/journal.pone.0159886.g008
two innermost rows as closed circles; this character was used as a primary characteristic to erect Cycloplectanum Oliver, 1968, which was later considered a junior synonym of Pseudorhabdosynochus [9] [28]. Squamodiscs with two innermost rows as closed circles are found in many Pseudorhabdosynochus species and even in species now placed in other genera such as Echinoplectanum Justine \& Euzet, 2006 and thus cannot be used for defining a genus [34].

We propose, for these four species, a group which is referred to here as the 'beverleyburtonae group'. The four species are parasitic on groupers of the genus Mycteroperca in the Mediterranean Sea and eastern Atlantic (with P. beverleyburtonae also found in the western Atlantic).

Three species of Pseudorhabdosynochus from groupers in the Mediterranean Sea share vaginal characters with species of the beverleyburtonae group but do not belong to it. These species are:

- Pseudorhabdosynochus regius Chaabane, Neifar \& Justine, 2015, from M. rubra [6]. This species possesses the same general vaginal structure but it has a straight primary canal (vs curved or S-shaped in species of beverleyburtonae group), a less differentiated trumpet, and the primary and secondary chambers are grouped in a heavily sclerotised structure.
- Pseudorhabdosynochus dolicocolpos Neifar \& Euzet, 2007, from M. costae [3]. This species has a well-defined trumpet but the primary canal is extremely long and thin, the primary chamber is very small and smaller than the secondary chamber.
- Pseudorhabdosynochus sinediscus Neifar \& Euzet, 2007, from M. costae [3]. This species has a vaginal structure superficially resembling species of the beverleyburtonae group but there is no secondary chamber. In addition, it is distinguished by the lack of squamodiscs.


## Analysis of COI sequences of monogeneans

We obtained COI sequences of $P$. hayet n. sp. (1 specimen), P. sosia Neifar \& Euzet, 2007 (4 specimens) and $P$. regius Chaabane et al., 2015 ( 1 specimen). The phylogenetic analysis was performed with these 6 new sequences and available sequences of Pseudorhabdosynochus, i.e. 1 sequence of $P$. sulamericanus Santos et al., 2000, 7 sequences of $P$. cyanopodus Sigura \& Justine, 2008, and, as an outgroup, Lamellodiscus ignoratus Palombi, 1943, a member of the Diplectanidae family to which Pseudorhabdosynochus belongs. In the tree obtained by the NJ method (Fig 9), the two species $P$. hayet and $P$. sosia were sister-groups. The genetic distance between species of Pseudorhabdosynochus varied from 13 to $26 \%$, and the distance between P. hayet and P. sosia was 13-14\% (Table 4).

## Discussion

Gills of groupers (Epinephelidae) harbour numerous diplectanid monogeneans assigned to Pseudorhabdosynochus [1-5, 20, 29, 35-38]. This genus currently contains more than 80 valid species [39] which are generally specialists. Based on the great similarity in the shape of the sclerotised vagina, the primary character for Pseudorhabdosynochus species diagnosis, the 'beverleyburtonae group' is proposed herein to include two new and two previously described species on Mycteroperca spp. in the Mediterranean and Eastern Atlantic Ocean. Several groups of Pseudorhabdosynochus species have already been suggested such as those on Epinephelus spp. in the southern Pacific, namely the 'P. cupatus group' [35] (including P. cupatus (Young, 1969), P. cyathus Hinsinger \& Justine 2006, P. calathus Hinsinger \& Justine 2006, and three unnamed species) [20, 40], the 'huitoe complex' (including P. huitoe Justine, 2007, P. manifestus Justine \& Sigura, 2007, and P. crassus Schoelinck \& Justine, 2011) [1, 37, 41], and a small group that includes only P. exoticus Sigura \& Justine, 2008 and P. exoticoides Justine \& Henry, 2010 [38, 42]. Kritsky et al. 2015 also proposed three groups of Pseudorhabdosynochus species


Fig 9. Evolutionary relationships of taxa. The matrix includes new sequences of Pseudorhabdosynochus spp. and available sequences from GenBank. The analysis, using the Neighbour-Joining method, involved 15 nucleotide sequences, and there were a total of 290 positions in the final dataset. Bootstrap test results ( 500 replicates) are shown next to the branches.

## doi:10.1371/journal.pone.0159886.g009

on Epinephelus spp., Mycteroperca spp., and Hyporthodus spp., respectively, in the western Atlantic Ocean [2]. These groups do not have formal systematic value but are useful for distinguishing species among the numerous valid Pseudorhabdosynochus species.

In this paper, we used mainly the morphology of the sclerotised vaginae to differentiate the diplectanid species. However, we added molecular information on two species for which the differentiation was mainly based on differences of measurements, but not on differences of morphology, namely $P$. sosia and the new species $P$. hayet. Using multiple and complementary sources of data, i.e. 'integrative taxonomy' [43], is important for better species characterisation and delimitation. In the case of $P$. sosia and $P$. hayet, the fact that the hosts are different is additional information advocating different parasite species, but is certainly not sufficient because it has been demonstrated that, in some cases, the same Pseudorhabdosynochus species can parasitize different host fish [44]; in that particular case, the genetic differences between specimens of P. cyanopodus Sigura \& Justine, 2008 from two hosts were very low ( $0-1.5 \%$ ). The genetic distance between $P$. sosia and $P$. hayet was 13-14\%, thus lower than the differences observed between other Pseudorhabdosynochus species in our dataset (19-26\%), but much higher than the differences between specimens of P. cyanopodus found on two different hosts [44]. The literature on monogenean COI sequences and their interspecific distances is scarce. Vanhove et al. (2013) have emphasized the difficulties encountered in using COI for the differentiation of species in various groups of flatworms, including monogeneans; however, they cite a case in which two valid diplectanid species were divergent by only $3.2 \%$ [45]. In other flatworms (geoplanid triclads), such as Microplana spp., intraspecific variation was up to $4 \%$, and interspecific variation was $19 \%$ [46], and a difference of $4.8 \%$ between Platydemus manokwari haplotypes was considered intraspecific [47]. New species are hypotheses that should be tested [48]. We conclude that the $13-14 \%$ difference of COI sequences between $P$. sosia and $P$. hayet is indicative of two different species, in addition to morphological differences and different hosts, and that $P$. hayet should be erected as a new species.

A list of 10 grouper-diplectanid species in the Mediterranean Sea has been provided (see Table 1 in [6]), and another species, $P$. sulamericanus, was recently added [6, 7]. The two new species of Pseudorhabdosynochus described here bring the total of species known in the Mediterranean to thirteen; however, the biodiversity of diplectanids on Mediterranean groupers is not yet fully documented.

## Author Contributions

Conceived and designed the experiments: AC LN DG JLJ. Performed the experiments: AC LN DG JLJ. Analyzed the data: AC LN DG JLJ. Contributed reagents/materials/analysis tools: AC LN DG JLJ. Wrote the paper: AC LN DG JLJ. Made the drawings: AC JLJ. Obtained sequences: DG. Performed phylogenetic analysis: AC JLJ.

## References

1. Justine J-L. Parasite biodiversity in a coral reef fish: twelve species of monogeneans on the gills of the grouper Epinephelus maculatus (Perciformes: Serranidae) off New Caledonia, with a description of eight new species of Pseudorhabdosynochus (Monogenea: Diplectanidae). Syst Parasitol. 2007; 66 (2):81-129. doi: 10.1007/s11230-006-9057-3 PMID: ISI:000244450600001.
2. Kritsky DC, Bakenhaster M, Adams D. Pseudorhabdosynochus species (Monogenoidea, Diplectanidae) parasitizing groupers (Serranidae, Epinephelinae, Epinephelini) in the western Atlantic Ocean and adjacent waters, with descriptions of 13 new species. Parasite. 2015; 22:24. doi: 10.1051/parasite/ 2015024 PMID: 26272242
3. Neifar L, Euzet L. Five new species of Pseudorhabdosynochus (Monogenea: Diplectanidae) from the gills of Epinephelus costae (Teleostei: Serranidae). Folia Parasitol. 2007; 54:117-28. PMID: 17886741
4. Mendoza-Franco EF, Violante-González J, Rojas Herrera AA. Six new and one previously described species of Pseudorhabdosynochus (Monogenoidea, Diplectanidae) infecting the gills of groupers (Perciformes, Serranidae) from the Pacific coasts of Mexico and Panama. J Parasitol. 2011; 97:20-35. doi: 10.1645/GE-2716.1 PMID: 21348602
5. Justine J-L, Beveridge I, Boxshall GA, Bray RA, Moravec F, Trilles J-P, et al. An annotated list of parasites (Isopoda, Copepoda, Monogenea, Digenea, Cestoda and Nematoda) collected in groupers (Serranidae, Epinephelinae) in New Caledonia emphasizes parasite biodiversity in coral reef fish. Folia Parasitol. 2010; 57:237-62. doi: 10.14411/fp.2010.032 PMID: 21344838
6. Chaabane A, Neifar L, Justine J-L. Pseudorhabdosynochus regius n. sp. (Monogenea, Diplectanidae) from the mottled grouper Mycteroperca rubra (Teleostei) in the Mediterranean Sea and Eastern Atlantic. Parasite. 2015; 22:9. doi: 10.1051/parasite/2015005 PMID: 25674913
7. Chaabane A, Justine J-L, Gey D, Bakenhaster M, Neifar L. Pseudorhabdosynochus sulamericanus (Monogenea, Diplectanidae), a parasite of deep-sea groupers (Serranidae) occurs transatlantically on three congeneric hosts (Hyporthodus spp.), one from the Mediterranean Sea and two from the western Atlantic. PeerJ. 2016; 4:2233. doi: 10.7717/peerj. 2233
8. Euzet L, Oliver G. Diplectanidae (Monogenea) de Téléostéens de la Méditerranée occidentale. II. Parasites d'Epinephelus gigas (Brünnich, 1768). Ann Parasitol Hum Comp. 1965; 40:517-23. PMID: 5884809
9. Oliver G. Recherches sur les Diplectanidae (Monogenea) parasites de téléostéens du Golfe du Lion. I. Diplectaninae Monticelli, 1903. Vie Milieu. 1968;Série A, 19:95-138.
10. Oliver G. Description de deux nouvelles espèces du genre Cycloplectanum Oliver, 1968 (Monogenea, Monopisthocotylea, Diplectanidae). Ann Parasitol Hum Comp. 1984; 59:31-9. PMID: 6721367
11. Santos CP, Buchmann K, Gibson DI. Pseudorhabdosynochus spp. (Monogenea: Diplectanidae) from the gills of Epinephelus spp. in Brazilian waters. Syst Parasitol. 2000; 45(2):145-53. doi: 10.1023/ A:1006232029426 PMID: ISI:000085138500006.
12. Ward RD, Zemlak TS, Innes BH, Last PR, Hebert PD. DNA barcoding Australia's fish species. Philos Trans R Soc Lond B Biol Sci. 2005; 360(1462):1847-57. doi: 10.1098/rstb.2005.1716 PMID: 16214743
13. Schoelinck C, Hinsinger DD, Dettaï A, Cruaud C, Justine J-L. A phylogenetic re-analysis of groupers with applications for ciguatera fish poisoning. PLoS ONE. 2014; 9(8):e98198. doi: 10.1371/journal. pone. 0098198 PMID: 25093850
14. Froese R, Pauly D, editors. FishBase. World Wide Web electronic publication. www.fishbase.org2016.
15. Heemstra PC, Randall JE. FAO Species Catalogue. Vol. 16. Groupers of the world (Family Serranidae, Subfamily Epinephelinae). An annotated and illustrated catalogue of the grouper, rockcod, hind, coral grouper and lyretail species known to date. Rome: FAO; 1993. 382 p.
16. Craig MT, Sadovy de Mitcheson YJ, Heemstra PC. Groupers of the world: a field and market guide. Grahamstown: NISC; 2012. 424 p.
17. Craig MT, Hastings PA. A molecular phylogeny of the groupers of the subfamily Epinephelinae (Serranidae) with a revised classification of the Epinephelini. Ichthyol Res. 2007; 54(1):1-17. doi: 10.1007/ s10228-006-0367-x PMID: ISI:000244724300001.
18. Ratnasingham S, Hebert PDN. BOLD: The Barcode of Life Data System (www. barcodinglife. org). Mol Ecol Notes. 2007; 7(3):355-64. doi: 10.1111/j.1471-8286.2007.01678.x PMID: 18784790
19. Malmberg G. Om förekomsten av Gyrodactylus på svenska fiskar (In Swedish.). Skrifter Utgivna av Södra Sveriges Fiskeriförening, Årsskrift. 1957; 1956:19-76.
20. Justine J-L. Species of Pseudorhabdosynochus Yamaguti, 1958 (Monogenea: Diplectanidae) from Epinephelus fasciatus and E. merra (Perciformes: Serranidae) off New Caledonia and other parts of the Indo-Pacific Ocean, with a comparison of measurements of specimens prepared using different methods, and a description of $P$. caledonicus n. sp. Syst Parasitol. 2005; 62(1):1-37. doi: 10.1007/s11230-005-5480-0 PMID: ISI:000231512700001.
21. Littlewood DTJ, Rohde K, Clough KA. Parasite speciation within or between host species?-Phylogenetic evidence from site-specific polystome monogeneans. Int J Parasitol. 1997; 27:1289-97. doi: 10. 1016/S0020-7519(97)00086-6 PMID: 9421713
22. Kimura M. A simple method for estimating evolutionary rates of base substitutions through comparative studies of nucleotide sequences. J Mol Evol. 1980; 16(2):111-20. Epub 1980/12/01. PMID: 7463489.
23. Kumar S, Stecher G, Tamura K. MEGA7: Molecular Evolutionary Genetics Analysis version 7.0 for bigger datasets. Mol Biol Evol. 2016:in press. doi: 10.1093/molbev/msw054
24. Moravec F, Chaabane A, Justine J-L, Neifar L. Two gonad-infecting species of Philometra (Nematoda: Philometridae) from groupers (Serranidae) off Tunisia, with a key to Philometra species infecting serranid gonads. Parasite. 2016; 23:8. doi: 10.1051/parasite/2016008 PMID: 26956219
25. Oliver G. Les Diplectanidae Bychowsky, 1957 (Monogenea, Monopisthocotylea, Dactylogyridea). Systématique. Biologie. Ontogénie. Écologie. Essai de phylogenèse. [Thèse d'État]: Académie de Montpellier, Université des Sciences et Techniques du Languedoc; 1987.
26. Roumbedakis K, Marchiori NC, Paseto A, Goncalves EL, Luque JL, Cepeda PB, et al. Parasite fauna of wild and cultured dusky-grouper Epinephelus marginatus (Lowe, 1834) from Ubatuba, southeastern Brazil. Braz J Biol. 2013; 73(4):871-8. doi: 10.1590/S1519-69842013000400025 PMID: 24789405.
27. Ulmer MJ, James HA. Monogeneans of marine fishes from the Bay of Naples. Trans Am Microsc Soc. 1981; 100(4):392-409. doi: 10.2307/3226153
28. Kritsky DC, Beverley-Burton M. The status of Pseudorhabdosynochus Yamaguti, 1958, and Cycloplectanum Oliver, 1968 (Monogenea: Diplectanidae). Proc Biol Soc Wash. 1986; 99:17-20.
29. Justine J-L. A redescription of Pseudorhabdosynochus epinepheli (Yamaguti, 1938), the type-species of Pseudorhabdosynochus Yamaguti, 1958 (Monogenea: Diplectanidae), and the description of $P$. satyui n. sp. from Epinephelus akaara off Japan. Syst Parasitol. 2009; 72:27-55. doi: 10.1007/s11230-008-9171-5 PMID: 19048406
30. Vidal-Martínez VM, Aguirre-Macedo L, Mendoza-Franco EF. Pseudorhabdosynochus yucatanensis sp. n. (Monogenea: Diplectanidae) from the gills of the red grouper Epinephelus morio (Pisces: Serranidae) of the Yucatan Peninsula, Mexico. Folia Parasitol. 1997; 44:274-8. PMID: 9437840
31. Moravec F, Chaabane A, Neifar L, Gey D, Justine JL. Descriptions of Philometra aenei n. sp. and P. tunisiensis n. sp. (Nematoda: Philometridae) from Epinephelus spp. off Tunisia confirm a high degree of host specificity of gonad-infecting species of Philometra in groupers (Serranidae). Syst Parasitol. 2016; 93(2):115-28. doi: 10.1007/s11230-015-9610-z PMID: 26790677
32. Justine J-L. Parasites of coral reef fish: how much do we know? With a bibliography of fish parasites in New Caledonia. Belg J Zool. 2010; 140 (Suppl.):155-90. PMID: WOS:000286593900012.
33. Knoff M, Cohen SC, Cárdenas MQ, Cárdenas-Callirgos JM, Gomes DC. A new species of diplectanid (Monogenoidea) from Paranthias colonus (Perciformes, Serranidae) off Peru. Parasite. 2015; 22:11. doi: 10.1051/parasite/2015011 PMID: 25754099
34. Justine J-L, Euzet L. Diplectanids (Monogenea) parasitic on the gills of the coralgroupers Plectropomus laevis and P. leopardus (Perciformes, Serranidae) off New Caledonia, with the description of five new species and the erection of Echinoplectanum n. g. Syst Parasitol. 2006; 64(3):147-72. doi: 10.1007/ s11230-006-9028-8 PMID: ISI:000238446600001.
35. Hinsinger DD, Justine J-L. The 'Pseudorhabdosynochus cupatus group' (Monogenea: Diplectanidae) on Epinephelus fasciatus, E. howlandi, E. rivulatus and E. merra (Perciformes: Serranidae) off New

Caledonia, with descriptions of Pseudorhabdosynochus cyathus n . sp. and P. calathus n. sp. Syst Parasitol. 2006; 64(2):69-90. doi: 10.1007/s11230-005-9018-2 PMID: ISI:000238294400001.
36. Justine J-L. Pseudorhabdosynochus argus n. sp. (Monogenea: Diplectanidae) from Cephalopholis argus, $P$. minutus n . sp . and Diplectanum nanus n . sp . from C . sonnerati and other monogeneans from Cephalopholis spp. (Perciformes: Serranidae) off Australia and New Caledonia. Syst Parasitol. 2007; 68(3):195-215. doi: 10.1007/s11230-007-9096-4 PMID: ISI:000249733100004.
37. Justine J-L, Sigura A. Monogeneans of the malabar grouper Epinephelus malabaricus (Perciformes, Serranidae) off New Caledonia, with a description of six new species of Pseudorhabdosynochus (Monogenea: Diplectanidae). Zootaxa. 2007; 1543:1-44. PMID: ISI:000248581900001.
38. Sigura A, Justine J-L. Monogeneans of the speckled blue grouper, Epinephelus cyanopodus (Perciformes, Serranidae), from off New Caledonia, with a description of four new species of Pseudorhabdosynochus and one new species of Laticola (Monogenea: Diplectanidae), and evidence of monogenean faunal changes according to the size of fish. Zootaxa. 2008; 1695:1-44. PMID: ISI:000252777100001.
39. Gibson DI. Monogenea. Accessed through: World Register of Marine Species at http://www. marinespecies.org/aphia.php?p=taxdetails\&id=798 2016 [2013-05-14].
40. Justine J-L, Vignon M. Monogeneans of the grouper Epinephelus tauvina (Perciformes, Serranidae) off Moorea, French Polynesia, with a description of Pseudorhabdosynochus pai n. sp. (Monogenea: Diplectanidae). Syst Parasitol. 2009; 72:113-25. doi: 10.1007/s11230-008-9159-1 PMID: 19115085
41. Schoelinck C, Justine J-L. Four species of Pseudorhabdosynochus (Monogenea: Diplectanidae) from the camouflage grouper Epinephelus polyphekadion (Perciformes: Serranidae) off New Caledonia. Syst Parasitol. 2011; 79:41-61. doi: 10.1007/s11230-010-9289-0 PMID: 21487947
42. Justine J-L, Henry É. Monogeneans from Epinephelus chlorostigma (Val.) (Perciformes: Serranidae) off New Caledonia, with the description of three new species of diplectanids. Syst Parasitol. 2010; 77:81-105. doi: 10.1007/s11230-010-9263-x PMID: 20852982
43. Dayrat B. Towards integrative taxonomy. Biol J Linn Soc. 2005; 85(3):407-15. doi: 10.1111/j.10958312.2005.00503.x
44. Schoelinck C, Cruaud C, Justine J-L. Are all species of Pseudorhabdosynochus strictly host specific?a molecular study. Parasitol Int. 2012; 61:356-9. doi: 10.1016/j.parint.2012.01.009 PMID: 22326703
45. Vanhove MP, Tessens B, Schoelinck C, Jondelius U, Littlewood DT, Artois T, et al. Problematic barcoding in flatworms: A case-study on monogeneans and rhabdocoels (Platyhelminthes). ZooKeys. 2013; (365: ):355-79. doi: 10.3897/zookeys.365.5776 PMID: 24453567.
46. Álvarez-Presas M, Mateos E, Vila-Farré M, Sluys R, Riutort M. Evidence for the persistence of the land planarian species Microplana terrestris (Müller, 1774) (Platyhelminthes, Tricladida) in microrefugia during the Last Glacial Maximum in the northern section of the Iberian Peninsula. Mol Phylogenet Evol. 2012; 64(3):491-9. doi: 10.1016/j.ympev.2012.05.001 PMID: 22587911
47. Justine J-L, Winsor L, Barrière P, Fanai C, Gey D, Han AWK, et al. The invasive land planarian Platydemus manokwari (Platyhelminthes, Geoplanidae): records from six new localities, including the first in the USA. PeerJ. 2015; 3:e1037. doi: 10.7717/peerj. 1037 PMID: 26131377
48. Sluys R. The unappreciated, fundamentally analytical nature of taxonomy and the implications for the inventory of biodiversity. Biodivers Conserv. 2013; 22(4):1095-105. doi: 10.1007/s10531-013-0472-x

