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1 AERYN: a simple standalone application for visualizing and enhancing elemental maps

2

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12

13 Abstract

14 Interpretation of high spatial resolution elemental mineral maps can be hindered by high frequency

15 fluctuations, as well as by strong naturally-occurring or analytically-induced variations. We have

16 developed a new standalone program named AERYN (Aspect Enhancement by Removing Yielded

17 Noise) to produce more reliable element distribution maps from previously reduced geochemical data.

18 The program is Matlab-based, designed with a graphic user interface and is capable of rapidly

19 generating elemental maps from data acquired by a range of analytical techniques. A visual interface

20 aids selection of appropriate outlier rejection and drift-correction parameters, thereby facilitating

21 recognition of subtle elemental fluctuations which may otherwise be obscured. Examples of use are

22 provided for quantitative trace element maps acquired using both laser ablation (LA-) ICP-MS and

23 electron probe microanalysis (EPMA) of the cold-water coral *Lophelia pertusa*. We demonstrate how

24 AERYN allows recognition of high frequency elemental fluctuations, including those which occur

25 perpendicular to the maximum concentration gradient. Such data treatment compliments commonly

26 used processing methods to provide greater flexibility and control in producing elemental maps from

27 micro-analytical techniques.

28

29 Keywords: LA-ICP-MS; EDS; WDS; element distribution maps; processing; Matlab; biominerals.

30

31 1. Introduction

32 Element distribution maps or images acquired using in-situ analytical techniques are widely used in
33 Geology and Earth Sciences (e.g. Cusack et al., 2008; Rittner & Müller, 2012; McGowan et al., 2014).

34 Several software packages exist to process element distribution maps (Paul et al., 2012; Rittner &
35 Müller, 2012; Ortolano et al., 2014), with some recent studies having investigated multi-trace element
36 maps processed from data obtained by laser ablation inductively coupled plasma mass spectrometry
37 (LA-ICP-MS) (e.g. Petrus & Kamber, 2013; Paul et al., 2014; Ubide et al., 2015).

38 Common issues in micro-analytical techniques linked to data acquisition, such as analytical drift and
39 outliers, can greatly influence the quality of resultant elemental maps. We propose a user-friendly
40 method created using the programming software Matlab (MathWorks;
41 <http://www.mathworks.com/products/matlab>) to deconvolute complex data and produce more
42 readable elemental maps. LA-ICP-MS data must have been previously reduced to account for baseline
43 correction and reference material calibration, e.g. by using common data reduction software. Our
44 program, which we term AERYN (Aspect Enhancement by Removing Yielded Noise), allows the
45 filtering of outliers and smoothing of values included in a data matrix, resulting in an increased
46 readability and importantly permitting more representative and accurate interpretation of
47 heterogeneity in elemental distribution. In addition, we propose a de-trending option to highlight
48 subtle geochemical features which may be obscured by strong signal fluctuations, either natural in
49 origin or caused by instrumental drift. AERYN has been compiled and exported as a standalone
50 application for Windows, allowing its use on a PC without the requirement of a Matlab licence
51 (<http://www.tcd.ie/Geology/staff/crowleyq/AERYN/>). The only requirement for this program to
52 perform correctly is to download and install the free Matlab Runtime R2015a (8.5) from the
53 MathWorks website (<http://www.mathworks.com/products/compiler/mcr>).

54 To illustrate how AERYN operates, we present elemental maps acquired by LA-ICP-MS and electron
55 probe microanalysis (EPMA) from the wall of the skeleton of the cold-water coral *Lophelia pertusa*.

56 Virtually any type of numeric compositional map can use part or all of the protocol described here, as

57 long as the data fit to a square or rectangular shape (in laser-based systems, this can be achieved by
58 analysing either parallel equivalent line scans or a grid of regularly-spaced spots). We provide the
59 corresponding Matlab code and the standalone application as well as a "Read Me" file with some
60 specifications as supplementary material, allowing the software to be adapted by other users.

61

62 2. AERYN description

63 2.1 User interface

64 AERYN is presented with a control panel on the left-hand side and three graphic areas on the right-
65 hand side (Fig. 1). The upper graphic displays an elemental map (using Matlab's "imagesc" function)
66 whereas the lower-left graphic displays a 3-dimensional view of the data (using Matlab's "surf"
67 function). The lower-right graph represents a histogram of the data (number of bins calculated using
68 Yule's rule) to graphically illustrate the distribution of elemental concentration represented in the
69 other graphic displays.

70

71 2.2 Importing data

72 AERYN uses text files (.txt, .tsv) exported from a variety of elemental map processing software or
73 from Microsoft Excel. We tested exports from LA-ICP-MS, Scanning Electron Microscopy with
74 electron-dispersive spectroscopy (SEM-EDS) and EPMA maps, but text files from any software
75 should be compatible. If not, we advise the user to follow instructions provided in the "Read Me" file.

76 AERYN does not process raw data, and a preceding data reduction step is still required. We used
77 Iolite (Paton et al., 2011; v. 2.5 and 3.0) for data reduction prior to importing in AERYN. To import
78 the numeric matrix corresponding to the map file, AERYN passes any number of lines composed of
79 characters and reads only the part of the file containing the concentration values. The size of the
80 imported matrix is only restricted by computer performance for very large datasets.

81 Once a numeric matrix has been generated, it is automatically displayed as a map on the upper graph
82 and as a 3D surface plot of the 2D data on the lower-left graph (Fig. 1). It is therefore possible to
83 rotate the lower-left graph or to automatically orient the view to a side section, in order to have better
84 visibility of outliers and fluctuations. Moreover, if required for better display, it is possible to

85 transpose the matrix by clicking on "Transpose" in order to invert lines with columns.
86 From this point and at any stage, the user can cancel any number of operations made on the data by
87 clicking on the "Undo" button.

88

89 2.3 Outlier removal

90 Outliers can be either extremely high or low values in comparison to the rest of the observed data and
91 introduce bias in interpretations. AERYN allows the user to locate and remove all outliers at once. All
92 removed outliers are replaced by empty spaces in the matrix (NaN). Processing of these spaces is
93 explained in the following section. A statistical test is proposed to the user to define automatic
94 thresholds of outliers based on the following equations (Tukey, 1977; Carling, 2000):

95 (1) $T_{up} = P90 + 1.5 * IQR$

96 (2) $T_{down} = P10 - 1.5 * IQR$

97 With P90 the percentile at 0.90 for the dataset, P10 the percentile at 0.10 and $IQR = P90 - P10$. All
98 values not occurring within the interval between T_{down} and T_{up} are considered as dataset outliers.
99 Such a statistical treatment will likely be sufficient for most datasets, though for the data presented
100 here the cut is too drastic and the upper end of some high value areas is lost, preventing an accurate
101 reading of the corresponding concentrations. For this reason, a second technique, based on iterative
102 visual definition of outliers, is made available for the user.

103 Firstly, for the high value outliers, it is necessary to estimate the upper limit of data to be retained. For
104 this, the "surf" function of Matlab is extremely powerful. By displaying the elemental map in 3D on
105 the lower-left graph (with the concentrations in the z axis) and with the possibility of rotating the
106 image, one can clearly (i) identify variations in concentration, (ii) assess the potential presence of
107 trends that would require a correction at a later stage, and (iii) identify obvious outliers and their
108 approximate values. At this stage, successive outlier removal steps can be achieved by searching for
109 values in the dataset lower or higher than a chosen specific value (Fig. 2). Moreover, the histogram on
110 the third graph (lower-right area) allows a better estimate of the minimum and maximum values to be
111 chosen for the colour palette in order to highlight most of the compositional information.

112 Spatially-restricted high concentration areas are maintained as a whole with this method, though more

113 subjectively compared to the automatic definition of outliers as presented above. It should be noted
114 that a possibility of inadvertently “over-trimming” the dataset exists, especially when controlled by
115 the user, and therefore outlier removal should be done with caution (e.g., Fig. 2c).

116 For the second phase of outlier removal, a lower minimum limit value expected for quantitative or
117 semi-quantitative data is zero. This minimum value can be higher if the concentrations of the studied
118 elements are high, but in the present example the concentrations are low and can easily be cut to
119 positive values without further consideration. If the mapped area is not entirely filled with sample
120 material (i.e., if a sample was mounted in epoxy resin and this resin was intentionally ablated in order
121 to obtain a rectangular map), the user must ensure that the resin value, which is generally constant, is
122 still included in the data to prevent incorrect resampling of these areas by the outlier removal step.

123 This step can have multiple iterations as successive outlier removals allow an improved visualisation
124 of geochemical fluctuations. The automatic Side View is extremely valuable at this stage as the outlier
125 values as well as the signal to be retained are indicated by the vertical axis. During the whole process
126 however it is necessary to keep in mind that some valuable geochemical information can reside in
127 high-frequency variability. It is thus unwise to crop the highest or lowest values too close to the main
128 visible fluctuations (see Fig. 2a and caption). Moreover, one must be aware that the larger the holes
129 caused by outlier removal, the less accurate the following steps will become. We suggest that the
130 number of outliers compared to the size of the data must be reasonably low. As an indication, the
131 number of points removed in Figure 2 represent approximately 0.5 % of the dataset (see Fig. 2c).

132 Each time the user filters the dataset, a temporary message is displayed in the history box to indicate
133 the percentage of data cropped compared from the entire matrix. The cut values are displayed on the
134 map as white areas. Once outlier removal has been validated, the side view and the histogram can
135 provide preliminary boundaries for the colour palette (Fig. 2b). The filters can be cancelled to retain
136 the entire initial dataset while maintaining the palette range values.

137

138 2.4 Filling gaps following outlier removal

139 In the outlier removal step, the outliers have been automatically replaced by the specific passive value
140 NaN. The "inpaint_nans" function (D'Errico, 2004) allows replacement of NaNs by analyzing

141 surrounding values for interpolation and extrapolation. With this step completed, the maps are
142 efficiently “cleaned” of outliers.

143 This gap-filling step should not be performed after further steps such as smoothing, as the method
144 used here substantially increases the number of NaN values in direct connection to pre-existing NaN
145 values. For this reason, we chose to automatically fill the NaNs at the validation of the outlier
146 removal.

147

148 2.5 Smoothing

149 An option to perform median filtering is available at this stage of data correction. Numerous
150 smoothing functions and methods are available in the Matlab environment. In order to take into
151 account the possible variability present on both x and y axes, the "medfilt2" function was chosen. This
152 function calculates the mean of all values included in a moving square centred on a value to smooth,
153 in order to replace that value with the mean, before moving the mask to the adjacent value. A larger
154 mask results in a stronger applied smoothing function. If the dataset presents a strong background
155 noise, it may be valid to repeat this step a second time, but it should be noted that if this option is
156 performed, absolute values of concentration will not be accurate. The mean will be identical, but the
157 absolute values of the extremes will no longer correspond to those given by initial data reduction and
158 the map will therefore be semiquantitative. Relative fluctuations, however, will remain the same.

159

160 2.6 Detrending

161 The detrending option is useful in two different scenarios. Firstly, if a strong naturally-occurring trend
162 is measured in the sample, the colour palette (commonly used to present fluctuation patterns) will
163 only show this particular trend. Any higher frequency fluctuations along this trend or variations
164 orthogonal to this direction (commonly observed in biominerals due to non-linear growth) and/or
165 fluctuations of lower amplitude are thus invisible. Removing this main trend will allow qualitative
166 study of such fluctuations. Additionally, if the background noise or sensitivity gradient is skewed as a
167 result of experimental drift, a correction must be applied to stabilize the response across the elemental
168 image. The technique presented here makes it possible to use data, even when a dramatic instrumental

169 drift occurs during an analytical session.

170 As is the case for smoothing, the process of detrending may change absolute values, so any
171 interpretations can only be proposed for elemental distribution and relative fluctuations in
172 concentration, in a semiquantitative fashion. Even in the case of naturally occurring trends, this option
173 allows observation of lateral variations in concentrations that would otherwise be concealed.

174 The particular data set presented here shows a general trend of a supposedly decrease in Sr all along
175 acquisition time, with data acquisition progressing in successive lines from left to right then top to
176 bottom.

177 A first option to deal with this trend is to execute an automatic detrending. By clicking the
178 corresponding button, AERYN calculates the best fitting linear plane to the data and applies a
179 detrending corresponding to this plane.

180 Alternatively, to improve visibility in this case we can select a transect representative of the main
181 fluctuation. By calculating the equation of the linear trend of this transect, it is possible to apply this
182 equation as a correction to all lines (or columns) and reveal geochemical fluctuations across the entire
183 map (Fig. 3). With this simple operation and even in the case of instrumental drift, it is possible to
184 work on the map without necessarily having to return to data acquisition.

185 When selecting a line or a column and clicking on "Display", two new windows present the selected
186 transect and its calculated linear trend on one panel (Fig. 3a), and the same transect along with its
187 detrended transect on the other (Fig. 3b). No operations to the map have been done at this stage. If
188 satisfied with these detrending parameters, the user can close both computer windows and Validate.

189 This will detrend the matrix and display the detrended map on all graphs (Fig. 3c). If the calculated
190 transects are not satisfactory, they can be cancelled by resetting the detrending operation.

191 The corrected map now presents fluctuations in Sr which are visible as coherent bands both at the top
192 and bottom of the map, showing continuity of these features. The map can be smoothed to clean the
193 high-frequency fluctuations using a 5x5 mask (Fig. 4).

194

195 2.7 Export map

196 At any stage, it is possible to export the matrix. In order to maintain optimum picture quality, the

197 maps are exported as TIFF files without compression with a uniform size of both dimensions of the
198 map. The user is asked to enter the path and file names. The matrix itself and the historic record of
199 operations made are saved as tab delimited text files, and the data are subsequently importable in
200 AERYN for further modification if required.

201 Figure 5 presents the Sr map used as an example here as well as a Li map from the same acquisition
202 session after data reduction (Iolite v. 3.0) and after post-processing using AERYN. Whereas maps
203 processed only with data reduction software are noisy and hamper visibility of concentration bands,
204 these bands are clearly visible after processing with AERYN. If required, in order to counteract the
205 standardization of the dimension of the map after saving with AERYN, it is possible to import the
206 TIFF file in another image software (e.g., Adobe Photoshop) to resize the image to the proper aspect
207 ratio.

208

209 3. Example materials

210 Specimens of the cold water coral *L. pertusa* were prepared at Trinity College Dublin using the
211 protocols described in Mouchi et al. (2014). Cleaned aragonitic skeletons were embedded in epoxy
212 resin and cut longitudinally with a Buehler Isomet Low Speed Saw. Sections obtained were polished
213 and subsequently etched with 2 % formic acid for 50 s. The etching protocol allowed direct
214 acquisition of measurements on LA-ICP-MS without pre-ablation.

215

216 3.1 LA-ICP-MS

217 Geochemical data were collected using an Analyte Excite 193 nm excimer ArF* laser ablation system
218 (Photon-Machines Inc.) coupled to an iCAP-Q ICP-MS unit (Thermo Scientific) at the Trinity
219 College Dublin geochemistry facility. Details on the instrument setup for elemental mapping can be
220 found in Ubide et al. (2015). Parallel and slightly overlapping (1 μm) line scans were acquired as
221 suggested by Ubide et al. (2015) to avoid gaps during data acquisition without notable contamination
222 from adjacent lines. A square beam of 12 μm was selected with 10 Hz repetition rate at 8 $\mu\text{m}\cdot\text{s}^{-1}$ scan
223 speed and a laser fluence of 3.9 $\text{J}\cdot\text{cm}^{-2}$. Ion intensities of ^7Li , ^{43}Ca , ^{86}Sr and other isotopes not
224 discussed here were measured for a total dwell time of 146 milliseconds. Individual dwell times were

225 70 ms for ^7Li , 5 ms for ^{86}Sr and 10 ms for ^{43}Ca . Data acquisition procedures ran for a total of 2.5 h.
226 Either NIST SRM 610 or NIST SRM 612 was ablated for approximately 120 s using the same
227 ablation parameters as the unknown line scans. Two calibration lines were used at the beginning and
228 end of each experiment and one additional line every 10 unknown sample lines which equates to 120 s
229 of data acquisition on the reference material every approximately 25 minutes. Collected data were
230 then processed for quantitative values with the Iolite software (Paton et al., 2011; v. 2.5 and 3.0) using
231 the Trace_Elements data reduction scheme (included in the Iolite program package) and the recently
232 accepted values of NIST SRM 610 and NIST SRM 612 (Jochum et al., 2011). ^{43}Ca was used as the
233 internal standard, assuming a stoichiometric Ca content. Reduced data were then exported to be
234 readable and further treated by AERYN; instructions to export text files readable by AERYN from
235 Iolite are in the "Read Me" file provided as supplementary material.
236 Depending on the size of the acquisition area for each specimen, typical maps have between 950 and
237 1950 columns for 30 to 60 lines, corresponding to approximately 1-2 mm by 330-660 μm ,
238 respectively.
239 The description of AERYN presented above (Fig. 1-5) used the map of distribution of Sr in *L. pertusa*
240 as example. The map of distribution of Li (Fig. 6a) does not require the outlier removal step as the
241 Side View indicates that the majority of measured fluctuations are below the resin value (see the
242 arrows on Fig. 6a) and visible. Indeed, though some positive outliers are present on the right-hand
243 side, they do not prevent reading of the minimum and maximum values on the histogram for palette
244 definition. The map is however difficult to interpret in terms of the location of fluctuations in Li
245 concentrations. The top part of the map is depleted in Li, as indicated by blue areas in opposition to
246 red and yellow areas on the lower part of the map. An automatic detrending applied to reduce the
247 amplitude of the fluctuations from top to bottom (Fig. 6b) aids in the visualization of several areas of
248 higher and lower Li concentration oriented N-S in this view.
249 From this step, a light smoothing using a 3 by 3 points mask further highlights the fluctuations. A
250 wider mask was not chosen due to the reduced number of lines in this map (only 30). Except for the
251 high value areas (in red) on the rightmost part of the map, both the map and the Side View show
252 higher amplitudes of fluctuations on the left-hand part compared to the right-hand part of the map

253 (Fig. 7). As this side of the coral skeleton displaying elevated elemental concentrations corresponds to
254 the initial stage of skeletal growth, this map can be interpreted in terms of environmental forcing or
255 kinetic incorporation, as suggested by Mouchi et al. (2014) from a simple transect (i.e. the elemental
256 variation of Li shown here is real and not an analytical or data processing artefact).

257

258 3.2 Electron microprobe

259 Another *L. pertusa* specimen was mapped with a Cameca SX-*Five* electron microprobe at the
260 Camparis Service, Paris, France. Acquisition parameters were 15 keV, 200 nA, 500 ms dwell time per
261 point with a 4 μm beam diameter and step size on a surface of 580 by 1352 μm . The map presented
262 here corresponds to Mg wt% (Fig. 8) after calibration using internal reference materials for each
263 measurement. The analytical uncertainty for this element using this protocol is approximately 100
264 $\mu\text{g}\cdot\text{g}^{-1}$ (2 SD).

265 Although a dwell time of 500 ms was used for this analytical protocol, which is high compared to
266 common trace element mapping protocols (usually set at around 300 ms), fluctuations in Mg
267 concentrations are still difficult to observe (Fig. 8a) and seem to be lower than the 100 $\mu\text{g}\cdot\text{g}^{-1}$
268 uncertainty. The visibility of these subtle fluctuations in Mg concentration can be improved using
269 AERYN (Fig. 8b).

270 The corresponding export files were written with transposed matrices compared to the ones generated
271 by Iolite. The "Transpose" button in AERYN was used to get the same orientation as for the LA-ICP-
272 MS maps. After filtering the data to cut the outliers, the map remains very noisy due to low
273 concentration values. We applied a 7x7 mask to strongly reduce the high-frequency fluctuations and
274 applied a weak detrending as the first lines presented lower values. The resulting map is shown in
275 Figure 8b.

276

277 The limits of the bands of high and low concentrations remain difficult to establish, but AERYN
278 permits areas of lower concentrations within high concentration bands to be more clearly visualized,
279 in particular around columns number 40, 80, 170 and 225. The map is therefore ready for
280 interpretation. High concentration bands are wider on the left-hand side (corresponding to the initial

281 stages of skeletal growth) than on the right-hand side of the map. Whereas a detailed interpretation of
282 the geochemical variations presented in these maps is beyond the scope of this paper, if Mg
283 incorporation is, as Sr incorporation, governed by kinetics in the skeleton of *L. pertusa*, this
284 observation would indicate in the present dataset that early stages of skeletal growth may feature
285 enhanced growth fluctuations. The large bands of high Mg concentration also tend to indicate that
286 most of these early stages present high growth rates and that the low growth rate periods are
287 temporally extremely restricted. These observations are in accordance of those of Mouchi et al. (2014)
288 and Rollion-Bard & Blamart (2015).

289

290 4. Conclusion

291 AERYN uses data generated from micro-analytical techniques and processed by data reduction
292 software (such as Iolite) to improve the interpretation of elemental distribution maps. This paper has
293 presented an alternative way to the trial-and-error routine of modification of palette thresholds by
294 allowing the user to display the geochemical dataset in 3D and observe data extremes systematically.
295 Moreover, AERYN allows the user to counteract any apparent changes due to instrumental drift,
296 which might not be possible to correct with traditional data reduction schemes. AERYN makes it
297 possible to observe discrete areas of high or low concentration which may be hidden by natural strong
298 fluctuations or analytical artefacts.

299 This program can highlight subtle variations in elemental maps in just a couple of minutes, thereby
300 enhancing the interpretation of elemental maps obtained with a variety of analytical techniques such
301 as LA-ICP-MS, EPMA and SEM-EDS.

302

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311

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350

Figure captions

Figure 1: Main window of AERYN showing a Sr distribution map measured by LA-ICP-MS on a *Lophelia pertusa* specimen. Data were reduced in terms of baseline correction and standard calibration prior importation in AERYN. At this stage, no corrections have been applied.

Figure 2: Coral skeleton Sr content measured by LA-ICP-MS as line scans at the outlier removal step (colour bar represents concentration in $\mu\text{g.g}^{-1}$) using Matlab's "surf" function. a: Map after filtering data out below 0 and above $100,000 \mu\text{g.g}^{-1}$. b: Side view rotated to expose the surface, showing the areas with the cut values in white (located by the arrows). c: Side view once the filtering has been validated. In order to retain the representative variability in the dataset, we advise not to cut the outliers below the values indicated here by the embedded red dashed line below which fluctuations do not appear anymore as discrete outliers. d: Final map and side view after removing outliers above $20,000 \mu\text{g.g}^{-1}$ (corresponding to the red dashed line in c). At this stage, the histogram distribution can help to define the range for the colour palette (red dashed lines). The high count bin around $0.6 \cdot 10^4 \mu\text{g.g}^{-1}$ corresponds to the resin value (i.e. not the biogenic carbonate) and is represented in dark blue on both sides of the map.

Figure 3: Detrending applied to the Sr distribution map. a: Selected transect from the map (in black) and calculated trend (in red). b: The original transect (black) and the detrended transect (red). c: Map after outlier removal and prior detrending is applied (compare to Fig. 2). d: Graphs obtained after detrending and rescaling palette range.

Figure 4: Sr distribution map after automatic detrending and smoothing with a 5 by 5 mask.

Figure 5: Element distribution maps resulting from data reduction using Iolite v. 3.0 (left) and after processing by AERYN (right) for Sr (a) and Li (b). Concentration bands are more clearly visible after processing with AERYN.

Figure 6: First steps for the Li map. a: Initial data after import of the Iolite export file. The resin is indicated by the arrows on the map and the Side View. b: Map after automatic detrending.

Figure 7: Smoothed map of Li concentration, using a 3x3 mask (each value replaced by the mean of all surrounding values in a 3 by 3 points area). Skeletal growth direction is right to left on this map.

Figure 8: Mg (wt.%) map of a *L. pertusa* specimen acquired by electron microprobe, as obtained by the SX acquisition software (a) and after processing with AERYN by smoothing using a 7x7 mask and detrending (b). Skeletal growth direction is left to right on this map.

Figure 1
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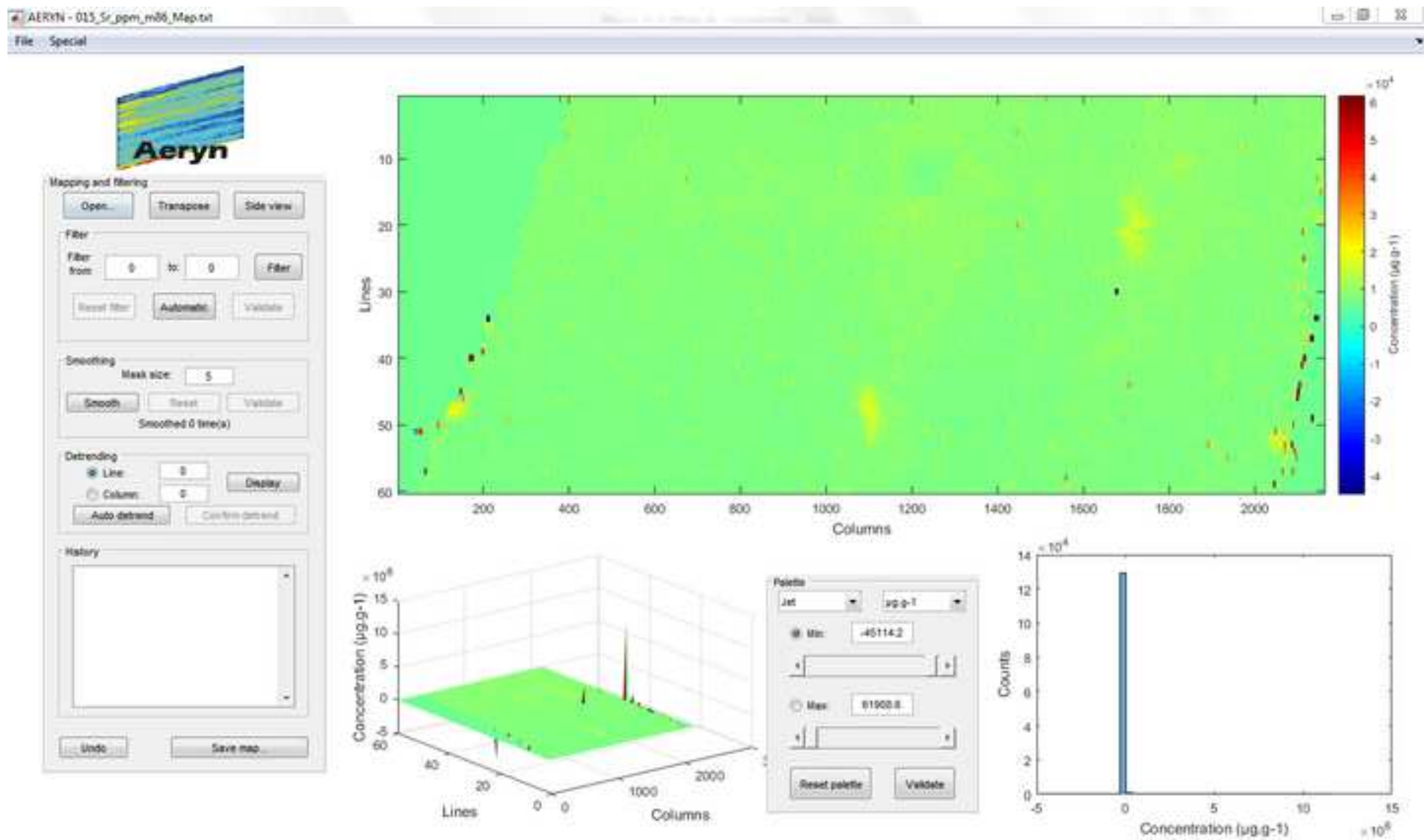


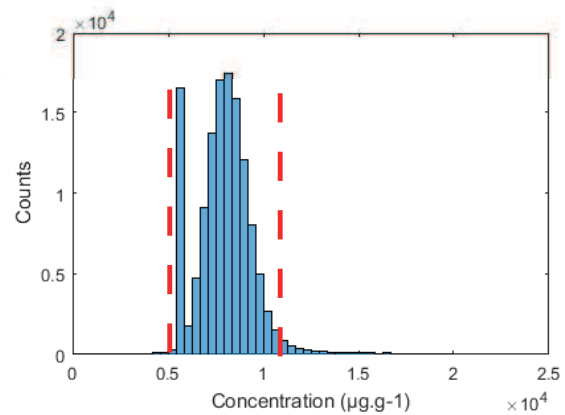
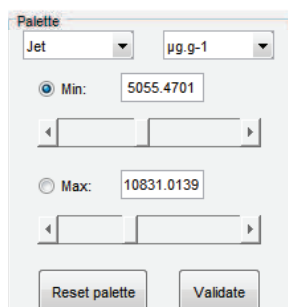
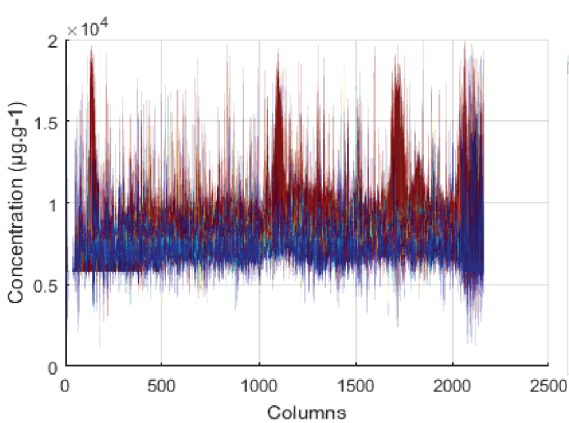
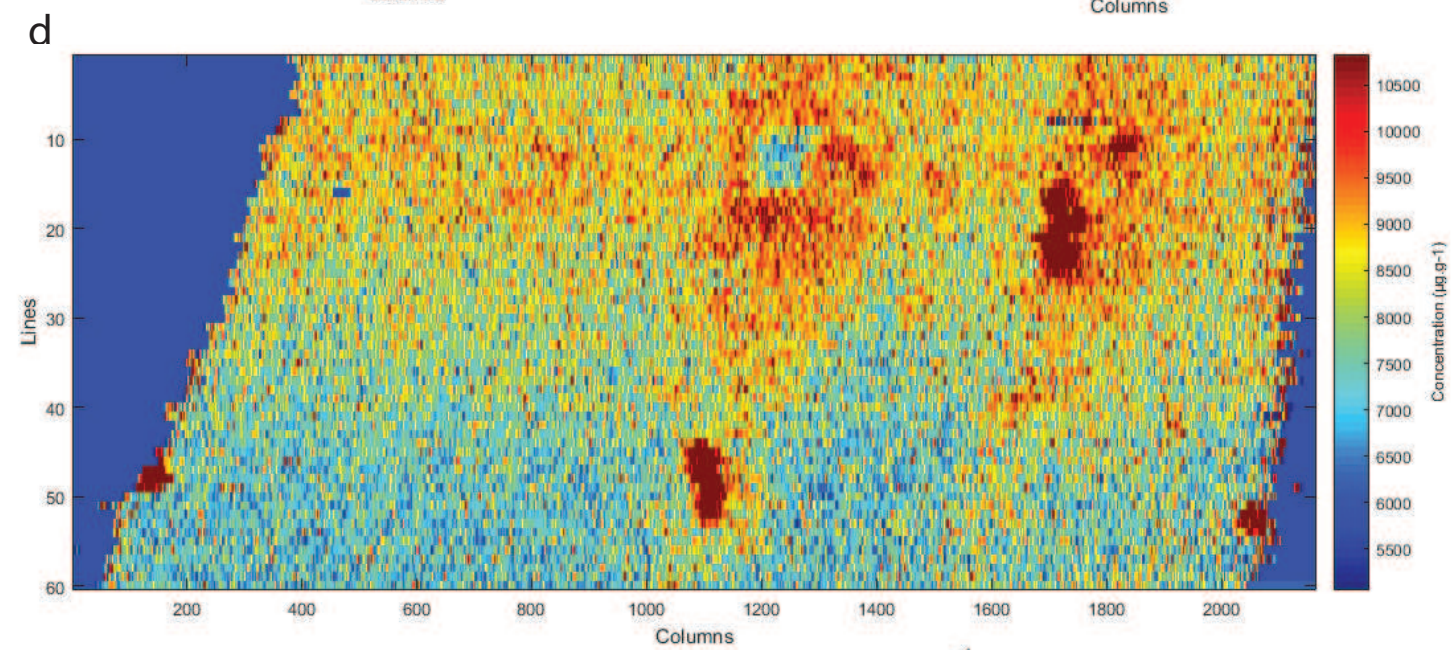
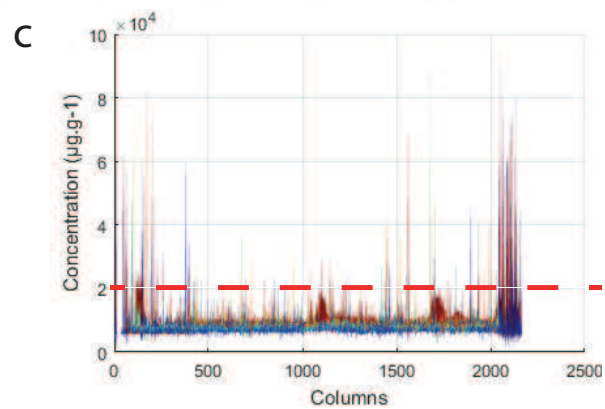
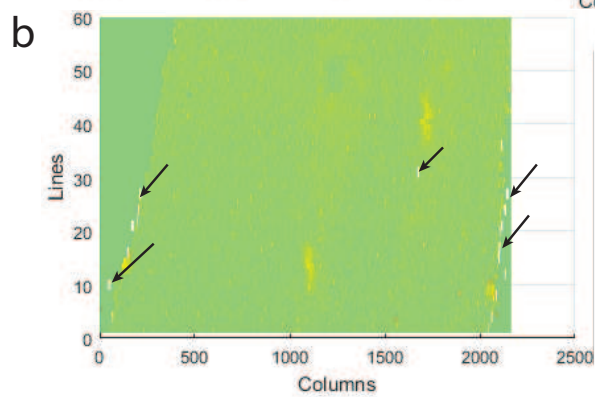
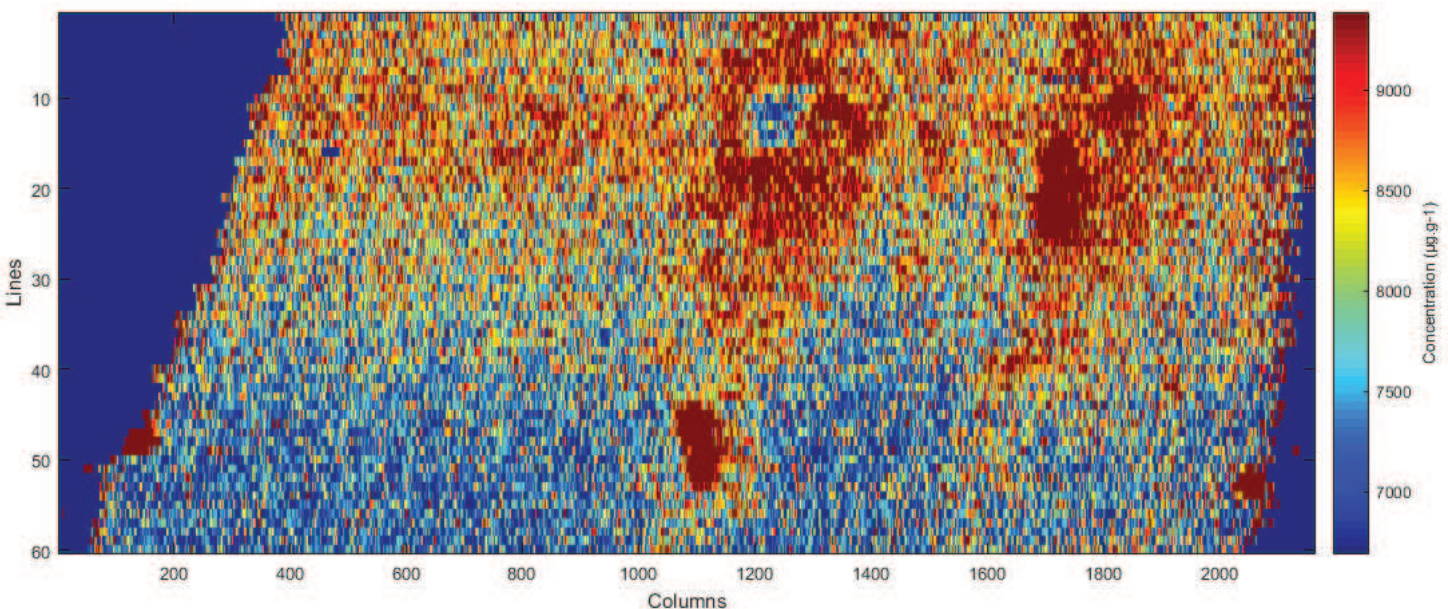
Figure 2

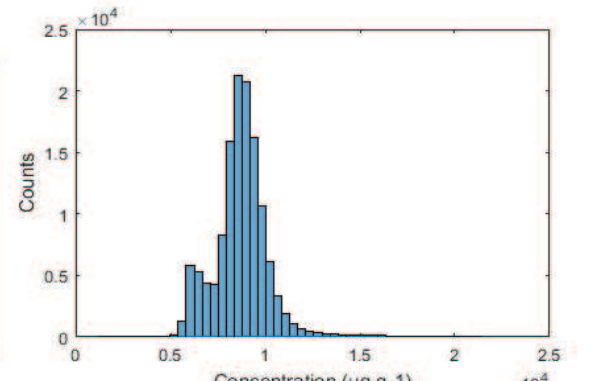
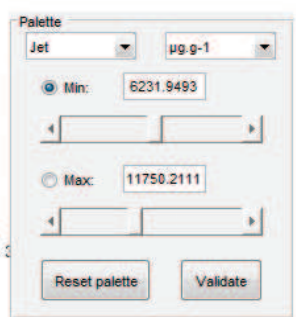
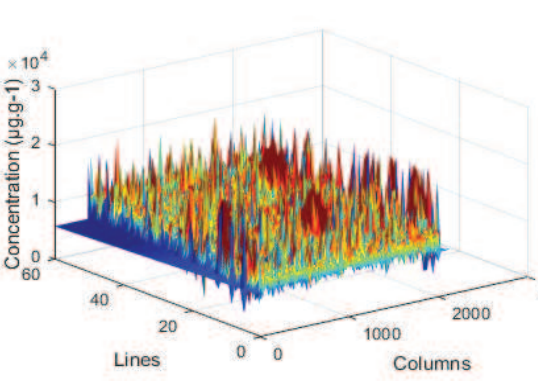
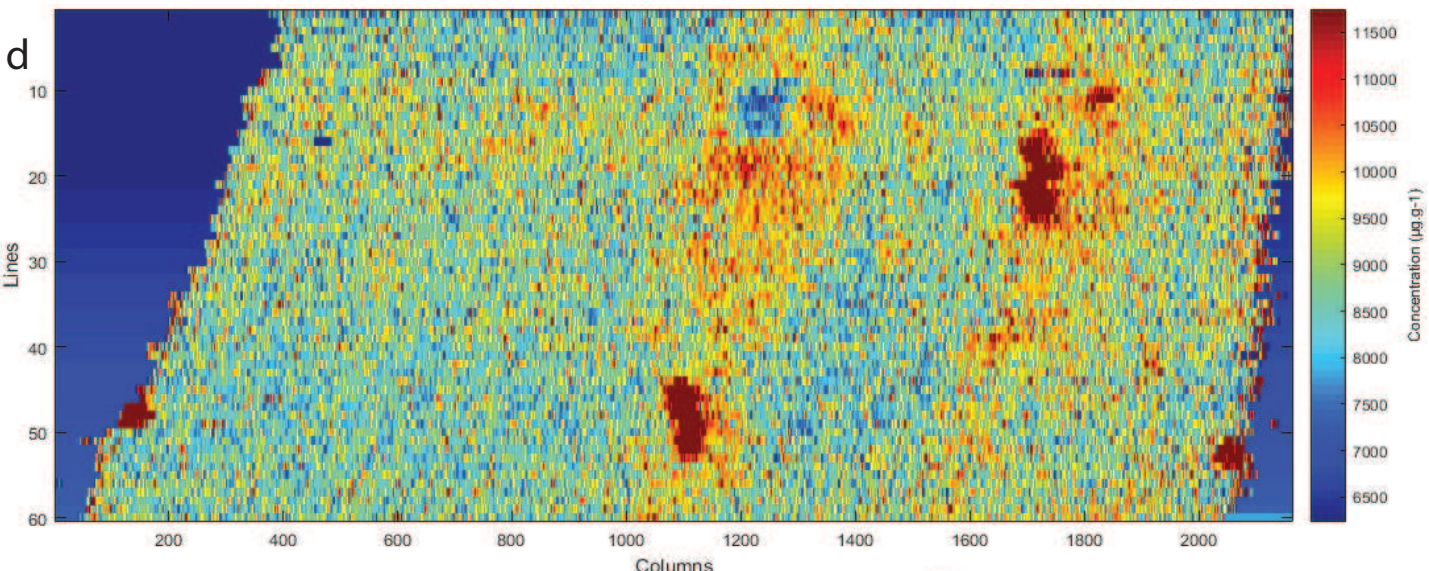
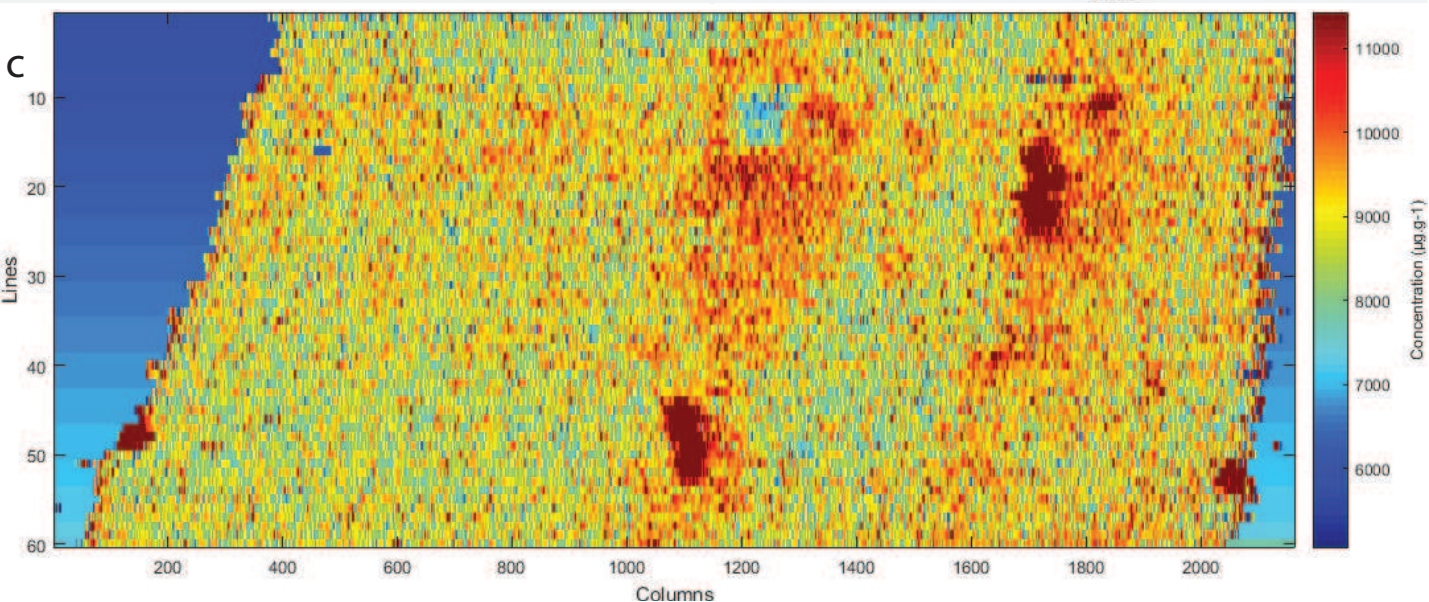
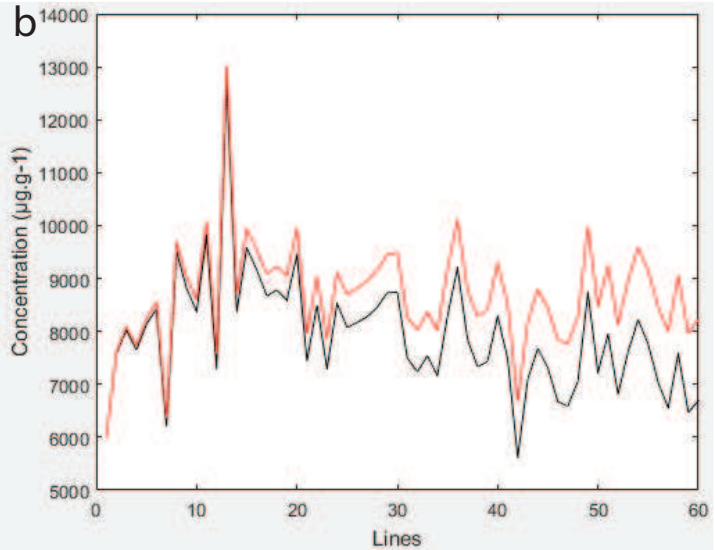
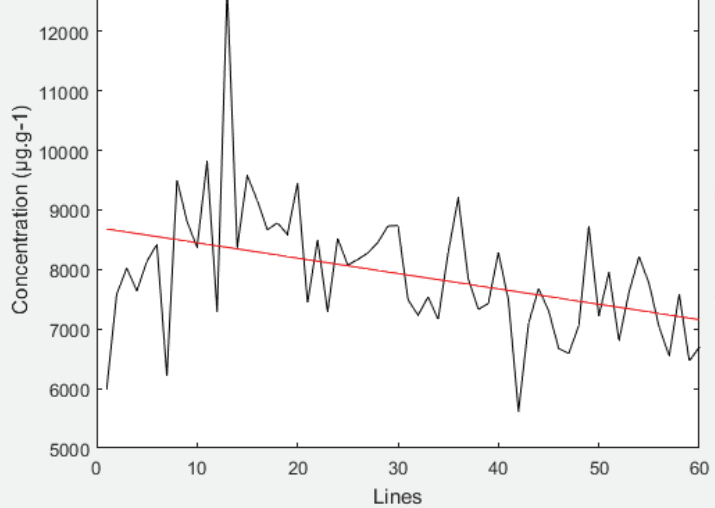
Figure 3

Figure 4
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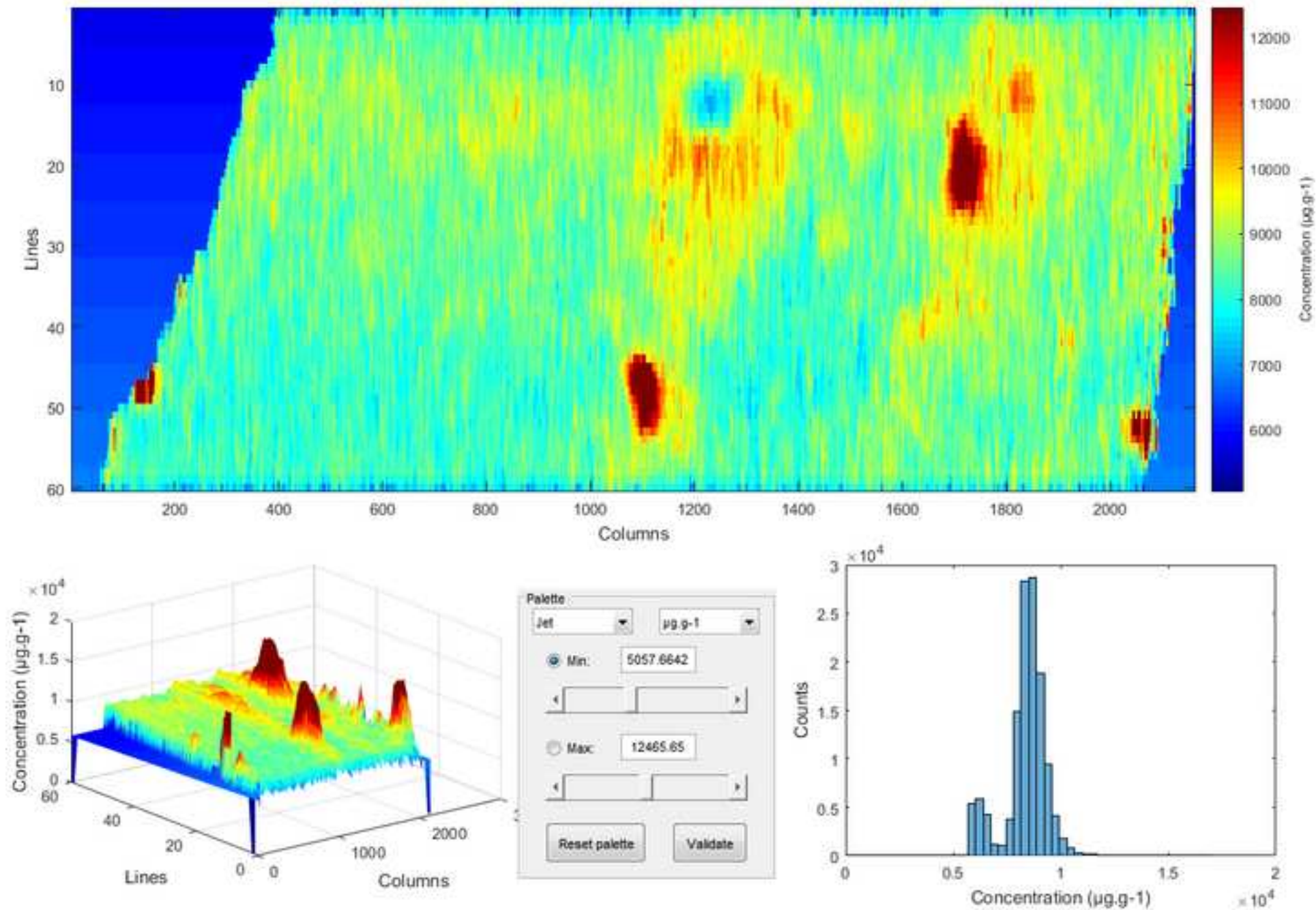


Figure 5

Pre-AERYN

Post-AERYN

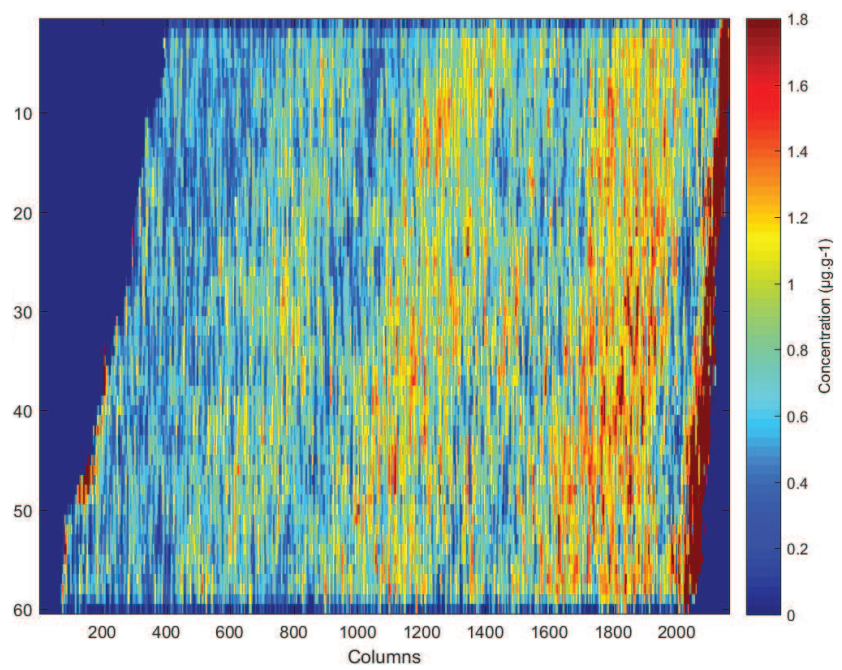
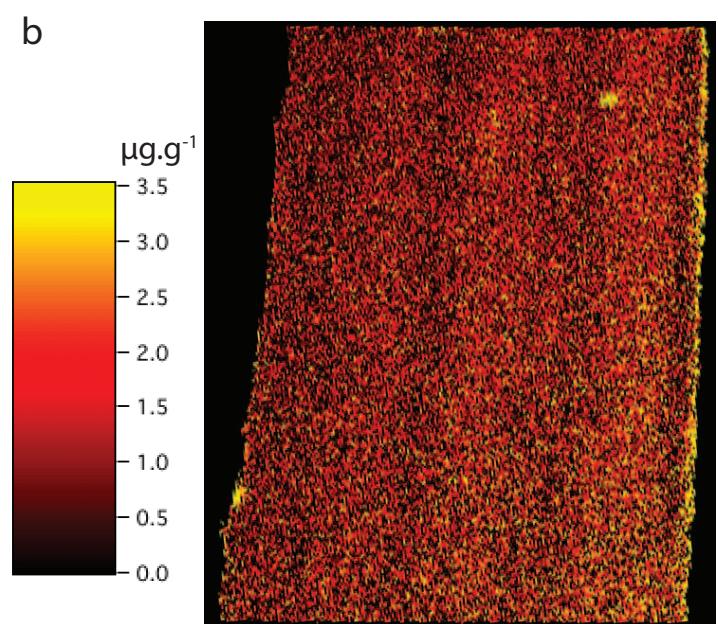
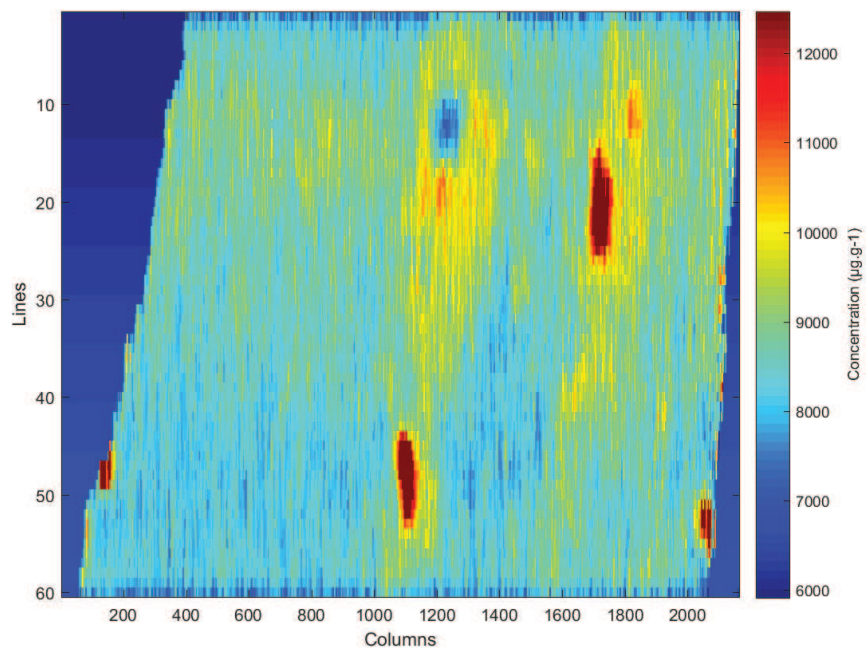
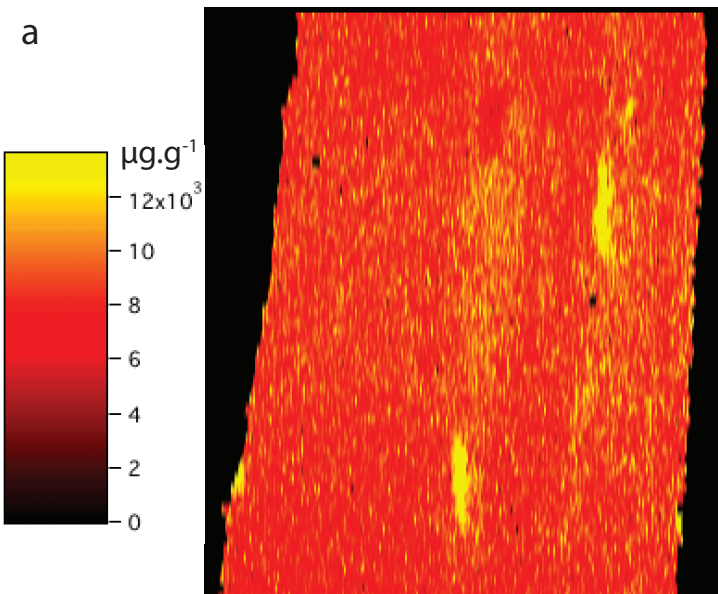
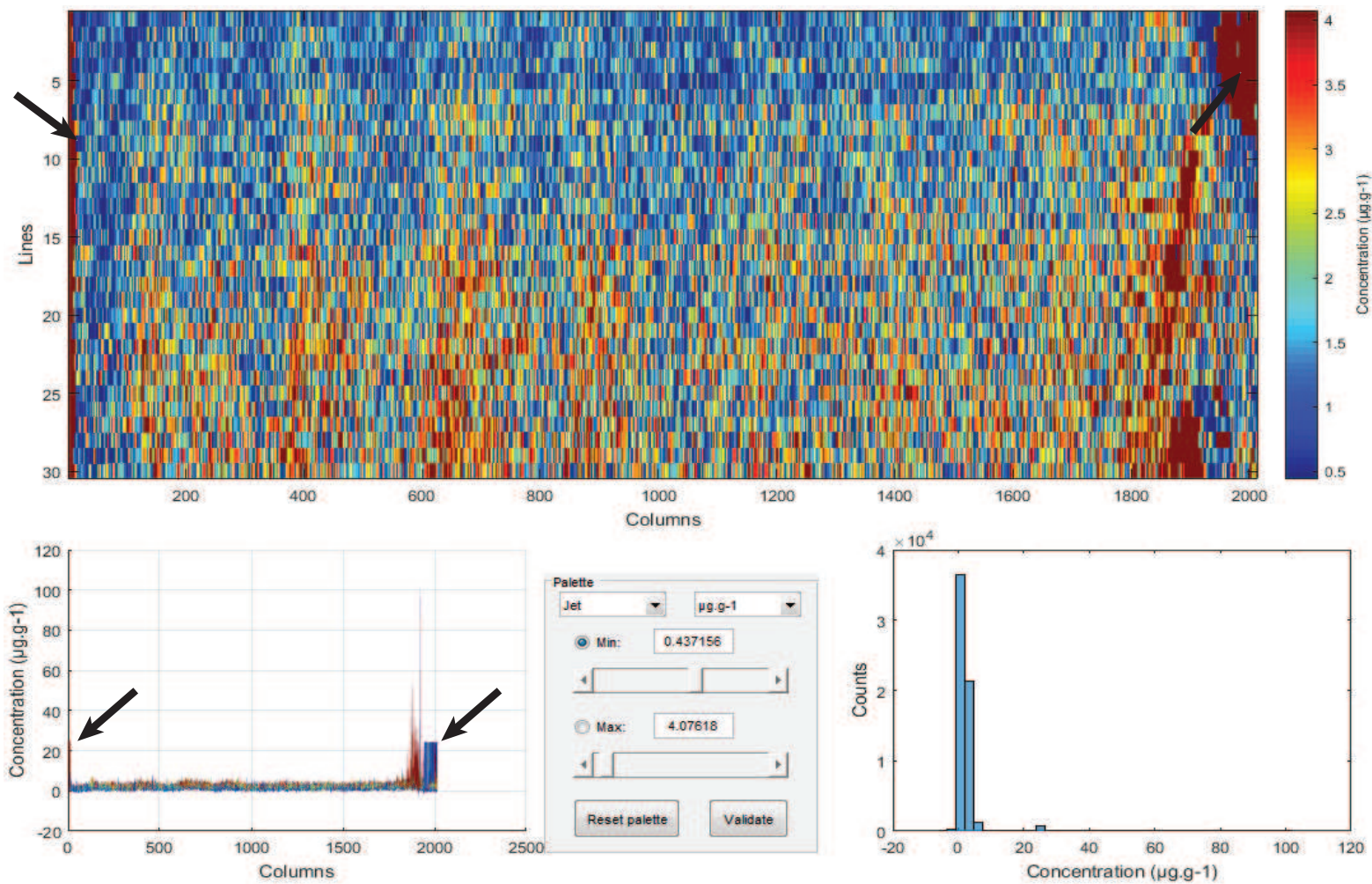


Figure 6

a



b

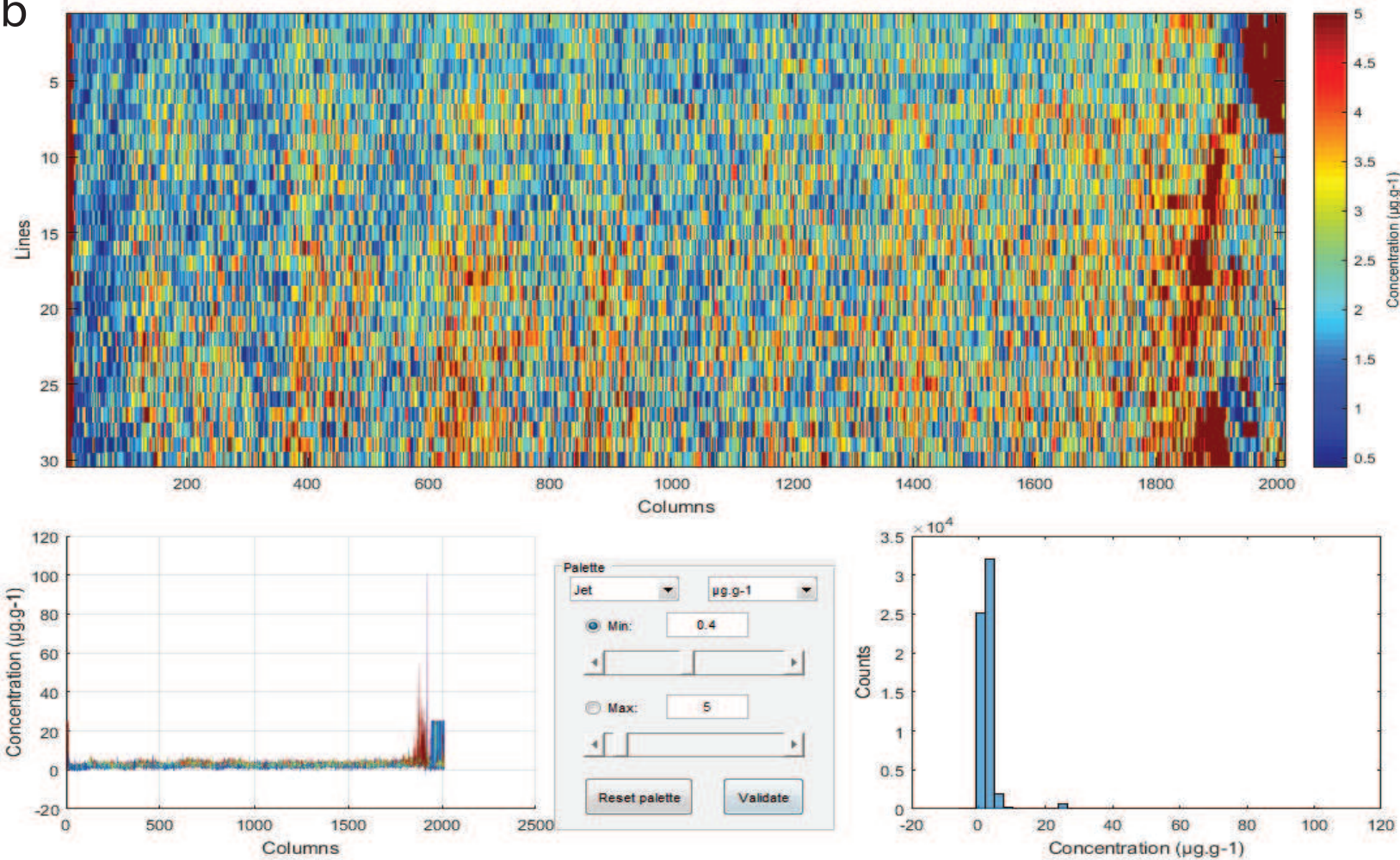


Figure 7

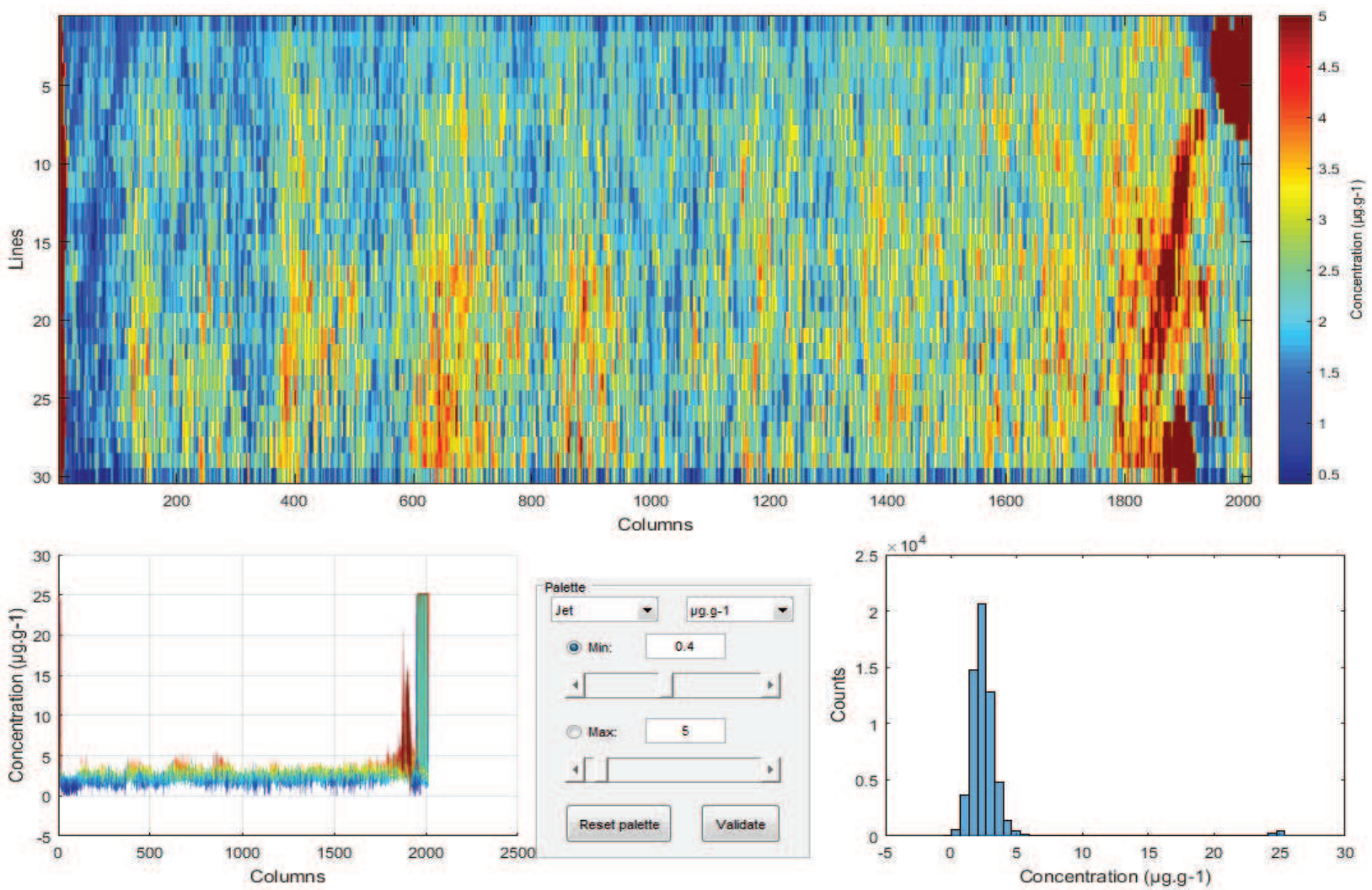


Figure 8

