



HAL
open science

Fecal microbiota transplantation in inflammatory bowel disease: the quest for the holy grail

B Pigneur, Harry Sokol

► **To cite this version:**

B Pigneur, Harry Sokol. Fecal microbiota transplantation in inflammatory bowel disease: the quest for the holy grail. *Mucosal Immunology*, 2016, 9 (6), pp.1360-1365. 10.1038/mi.2016.67 . hal-01404075

HAL Id: hal-01404075

<https://hal.sorbonne-universite.fr/hal-01404075>

Submitted on 28 Nov 2016

HAL is a multi-disciplinary open access archive for the deposit and dissemination of scientific research documents, whether they are published or not. The documents may come from teaching and research institutions in France or abroad, or from public or private research centers.

L'archive ouverte pluridisciplinaire **HAL**, est destinée au dépôt et à la diffusion de documents scientifiques de niveau recherche, publiés ou non, émanant des établissements d'enseignement et de recherche français ou étrangers, des laboratoires publics ou privés.

Fecal Microbiota Transplantation in Inflammatory bowel disease: the quest for the Holy Grail

Bénédicte Pigneur^{1,2,3} and Harry Sokol^{3,4,5,6*}

¹Institut National de la Santé et de la Recherche Médicale INSERM UMR1163, Laboratory of Intestinal Immunity, Institut Imagine, Paris, France

²Department of Pediatric Gastroenterology, Hepatology and Nutrition, Necker Enfants Malades Hospital, Assistance Publique – Hôpitaux de Paris, Paris Descartes-Sorbonne Paris Cite, Paris, France.

³French Group of Faecal Microbiota Transplantation (GFTF)

⁴Sorbonne University - UPMC Paris 06, INSERM ERL 1157, Avenir Team Gut Microbiota and Immunity, UMR 7203, Saint-Antoine Hospital, Paris, France

⁵INRA, UMR1319 Micalis & AgroParisTech, Jouy-en-Josas, France

⁶Department of Gastroenterology, Saint Antoine Hospital, AP-HP, UPMC Univ Paris 06, Paris, France

*Corresponding author:

Harry Sokol, Service de Gastroentérologie et Nutrition, Hôpital Saint-Antoine, 184 rue du faubourg St Antoine, 75571 Paris cedex 12, France.

E-mail: harry.sokol@aphp.fr Phone: +33 149 283 162. Fax: +33 149 283 188

Abstract:

Inflammatory Bowel Disease is due to an aberrant immune response towards luminal antigens, probably commensal bacteria, in genetically susceptible subjects and is also influenced by environmental factors. An imbalanced intestinal microbiota known as “dysbiosis”, characterized by an increased proportion of pro-inflammatory microorganisms and a decreased proportion of anti-inflammatory microorganisms, has been repeatedly observed in IBD and is now recognised as a key factor in the gut inflammatory process. Fecal Microbiota Transplantation (FMT) has gained interest as a novel treatment option in IBD. The goal of FMT in IBD is not only to correct the dysbiosis, but also to restore a normal dialogue between the host immune system and the microbiota. Data are still scarce but the results of the first studies suggest that FMT could be a promising therapy in IBD. More studies are needed to define the best indications, optimal timing, frequency, mode of delivery and the optimal donor for each patient.

Introduction

Although major progress has been achieved in recent years, the pathogenesis of Inflammatory Bowel Disease (IBD) has not yet been fully elucidated. It is generally accepted that IBD is caused by an aberrant immune response towards luminal antigens, most likely commensal bacteria, in genetically susceptible subjects and is also under the influence of environmental factors¹. Moreover, an imbalanced intestinal microbiota known as “dysbiosis” has been repeatedly observed in IBD and is now recognised as a key factor in the gut inflammatory process^{2, 3, 4, 5, 6, 7}. This dysbiosis is notably characterized by an increased proportion of pro-inflammatory microorganisms and a decreased proportion of anti-inflammatory microorganisms. From a therapeutic point of view, the correction of this dysbiosis is thus an attractive approach. Until now, efficacy of microbiome based therapies such as probiotics or antibiotics has been disappointing in IBD. Fecal Microbiota Transplantation (FMT) consists of the administration of fecal material from a donor into the intestinal tract of a recipient in order to change their microbiota composition and restore healthy conditions. FMT has been used for several years for the treatment of recurrent *Clostridium difficile* infection (CDI), but this treatment in CDI has only recently been proven to be efficient in a randomized control trial⁸. Following this study, FMT is now being evaluated in several other microbiota-driven diseases and particularly in IBD. CDI is a pure ecological problem characterized by a defect in gut microbiota barrier properties^{9, 10}. IBD, however, is far more complex and involves a deregulation of the host-microbes crosstalk. The goal of FMT in IBD is thus not only to correct the dysbiosis, but also to restore a normal dialogue between the host immune system and the microbiota. Data are still scarce but the results of the first studies suggest a complex effect of FMT in IBD.

In this paper we will review the data currently available on FMT and IBD and discuss the various and unresolved issues.

Fecal Microbiota Transplantation and Inflammatory Bowel Disease: state of the art

1. FMT in Ulcerative Colitis

There are several studies and case reports on the use of FMT in ulcerative colitis (UC) but only 2 randomized control studies. Angelberger reported the results of FMT in a small group of 5 patients with moderate to severe UC who received FMT via nasojejunal tube and enema¹¹. None of the patients achieved remission by week 12 but a positive response was observed in one patient who showed an improvement in Mayo score. In a small pilot study in 4 pediatric UC patients, Suskind observed no clinical or biological improvement after a single FMT via nasogastric tube¹². Cui et al. reported the results of a prospective observational study in 15 patients with moderate to severe steroid-dependent UC¹³. FMT was administered by upper endoscopy. They developed an original step-up FMT strategy where patients who failed to benefit from the initial FMT received a second treatment after one week. Eight patients (57%) responded to FMT with clinical improvement and discontinuation of steroids, 5 of whom received only 1 FMT. Four of the 8 responders maintained long term remission. Kunde et al. performed FMT in a small group of paediatric UC patients receiving enemas once a day for 5 days. Seven of the 9 patients showed clinical remission at 1 week and 6 of them maintained clinical remission at 1 month¹⁴. A recent meta-analysis of 9 studies (including 122 patients, 79 with UC, 39 CD and 4 IBD unclassified) described a pooled proportion of patients who achieved clinical remission of 36.2%¹⁵. Among studies that included only UC patients the proportion of patients achieving remission was 22%. There are only very limited data on FMT in pouchitis, with very few patients and disappointing results for most of them^{16, 17}.

There are only 2 randomized controlled studies evaluating the efficacy of FMT in UC. Moayyedi *et al.* reported the results of a randomized, placebo-controlled trial using FMT to induce remission in patients with mild to moderate UC¹⁸. Patients were randomized to receive FMT, or placebo, via enema once a week for 6 weeks. Results showed that subjects receiving FMT achieved remission significantly more often than those receiving placebo (9/38 versus 2/37,

p=0.03). The second trial was conducted by Rossen *et al.* in 50 patients with mild-to-severe UC¹⁹. Fecal transplant was administered via nasoduodenal tube at week 0 and 3 with feces from healthy donors or autologous fecal microbiota. There was no statistically significant difference between the 2 groups regarding clinical and endoscopic remission. The results of these two randomized control trials may be influenced by inconsistent study designs, including differences in the control groups, dose and preparation of donor feces, delivery method and frequency of FMT.

Compared to the impressive results of FMT in recurrent CDI (between 80 and 95% efficacy), these results in UC are disappointing. However, studies are still sparse with few patients and heterogeneous design, making comparison and conclusion difficult. Nevertheless, these data highlight the fact that the mechanism of action of FMT in IBD is radically different from that in CDI and suggest that results obtained in CDI will not be directly transposable in IBD.

2. FMT in Crohn's disease

Less data is available for FMT in Crohn's disease (CD) and, to our knowledge, no randomized controlled trial results have been published to date. Results reported in isolated cases or small series are heterogeneous but suggest a positive effect of FMT in CD in some conditions^{15, 20}. In a meta-analysis, subgroup analysis demonstrated a pooled estimate of clinical remission of 60.5% in CD patients¹⁵. In an open label uncontrolled study, Suskind *et al.* reported the effect of FMT in 9 paediatric patients (12–19 years) with mild to moderate active CD²¹. Patients received FMT prepared from their parents stool by nasogastric tube with follow-up evaluations at 2, 6, and 12 weeks. Seven out of the 9 patients were in clinical remission at week 2 with a decrease in disease activity score (PCDAI), CRP level and calprotectin. However, the effect was transient as only 5 patients remained in remission at week 12. A prospective pilot study by Cui *et al.* enrolled 30 patients with active CD who received FMT through mid-gut administration in conjunction with treatment with Mesalazine²². After FMT, the overall clinical improvement and clinical remission

rates at 1 month were 86.7% and 76.7% respectively. The follow up (after 15 months) showed sustained clinical remission with improvement in biological markers. Although these studies were uncontrolled, the results are impressive. Randomized placebo controlled trials are needed to draw a clear picture of the potential effect of FMT in CD.

The first data on FMT in IBD suggest a positive effect. However, its magnitude and the way to optimize it remain to be clarified. Only larger randomized controlled studies will be able to address these questions.

How to optimize FMT in IBD?

The efficacy of FMT in a patient with IBD is likely to be affected by many factors: patient status and preparation, donor selection, stool processing, and transplant delivery method²³. Additionally, many questions regarding safety remain unanswered (Figure 1).

1. Selection of the donor, safety and efficacy

- Safety of FMT

Although FMT is perceived as “natural” or even “organic” by many patients and physicians, it has potential side effects. Using feces from a healthy donor presents potential risk of contracting a disease that can be spread through fecal material. Transmission of enteric pathogens via FMT is an important concern but appears to be rare due to the current screening procedure of donors.

Diverse adverse effects have been reported with FMT specifically in IBD, most of them being mild fever and mild gastrointestinal symptoms. IBD flares following FMT have been described both in UC and CD^{24, 25}. The route of FMT administration seems to modify the adverse effect profile. Fever and C-reactive protein (CRP) rise have been described with nasojejunal route whereas administration via the rectal route appears to be safer. Rare cases of aspiration

pneumonia has been described in FMT via the nasojejunal route^{26, 27}. Mortality reported in the literature was not attributed to the FMT procedure but to patients' comorbidities^{20, 23}. Adverse metabolic response with important weight gain was reported in a woman who received FMT from an overweight donor. This case must however be interpreted with caution as the recipient was also overweight (BMI 26) at the time of FMT²⁸.

Screening of fecal donors before FMT may be just as necessary as screening organ donors to ensure that unwanted traits are not transferred with the transplant. In the literature, donors were either close relatives, household members or unrelated (defined as healthy voluntary donors not related to the patient). Donors should be screened for social behaviours that could increase risk of transmitting infection and should be free of diseases that may be transmissible by stool²³.

- Efficacy of the FMT depends on the donor microbiota composition

Each individual has a distinct fecal microbiota composition, representing major difficulties in obtaining homogenous FMT treatment to test. Indeed, each FMT can be considered as donor-specific leading to donor-dependant effects. This microbiota diversity may account for some of the variability in the outcomes of different studies explaining the heterogeneity of the results.

Intrinsic characteristics of the donor's microbiota could play a major role in the outcome of FMT.

This is highlighted in the study by Moayyedi *et al.*¹⁸. Patients who received donor B's feces were seen to achieve remission more often than others. Kelly *et al.* suggested that the efficacy of FMT may be related to the richness of a donor's stool, and that one donor's stool may be more "rich" than another²³. Indeed, the microbiota of donor B was enriched in members of the Lachnospiraceae family and *Ruminococcus* genus²⁹. Vermeire *et al.*, investigated FMT as a therapeutic option in 14 patients with IBD²⁶. They observed that stools from donors which resulted in a successful FMT were characterised by a significantly higher bacterial richness³⁰.

Richness of the donor microbiota is thus an important parameter to take into consideration.

Another potentially important aspect to investigate in FMT is the amount of anti-inflammatory

bacteria such as *Faecalibacterum prausnitzii* or bacteria producing Short Chain Fatty Acid (SCFA)^{31, 32}. SCFA represent the major carbon flux from the diet through the gut microbiota to the host and evidence is emerging for a regulatory role of SCFA in local, intermediary and peripheral metabolism³². FMT enriched in SFCA producers might be of importance for the recipient and the sustained and durable implantation of bacteria given by FMT.

Beyond characteristics of donor microbiota, it is possible that combination of donor and recipient features might influence FMT efficacy in IBD, as HLA system does in tissue or bone marrow transplantation. These factors could be host- or microbiota-driven and could include genetics, microbiota and diet. For example, the transfer of microbiota across gender has been identified as a possible issue. Studies have demonstrated a significant relationship between gut microbiome composition and sex, which may explain in part the sex-specific disparities in a variety of diseases such as autoimmune diseases³³. Despite these concerns, no published FMT study to date has age- or sex-matched donors with recipients, factors that may not affect short-term outcomes, but again may have consequence in the long term follow up especially in paediatric patients.

A recent study of infants and children with healthy growth phenotypes living in Mirpur, Bangladesh, reported the 16S ribosomal RNA (rRNA) analysis of monthly fecal collection from birth up to the end of the second year of life³⁴. In this study, the rRNA analysis of bacterial components in the gut microbiota revealed 24 “age-discriminatory” taxa, whose changes in relative abundance over time define a program of normal maturation of the microbiota across biologically unrelated individuals. The authors developed two related metrics, relative microbiota maturity and microbiota-for-age Z-score (MAZ) that significantly correlated with the chronological age of children with healthy growth phenotypes. Then applying their metrics on malnourished children in the same region, they showed that children with malnutrition have “immature” gut microbiota³⁴. The authors suggested that in healthy children, microbiota development is optimized to satisfy the different growth needs of the host at different ages. Hence, age-matched

donors are more likely to be at a comparable level of microbiological and immunological maturity and thus represent a more relevant source of material for pediatric populations.

Colonization success seems also to be different from one bacterium to another. Indeed, a donor strain belonging to a species already present in the recipient microbiota is more likely to establish in the donor gut³⁵. There might thus be some donor-recipient microbiota compatibility factors to take into account to maximize the FMT effect.

The recent development of stool banks in several countries could help to facilitate FMT activities³⁶. The Microbiome Health Research Institute, d.b.a.OpenBiome, is a nonprofit organization dedicated to expanding safe access to fecal microbiota transplants, and to catalysing research into the human microbiome. OpenBiome has built an international public stool bank which has two main objectives. Firstly; donor stools are screened and processed in a standardized manner with the goal of facilitating safe FMTs, and secondly; they offer a platform for investigating other microbiome associated disease.

Donor selection is a major challenge for FMT in IBD and being able to identify a “good” donor is of high importance. There is a large volume of research available on the gut microbiota which serves as useful to narrow down donor selection, however, only large trial results will allow us to identify relevant criteria to select donors.

2. Preparation of the Fecal Microbiota Transplant

Prior to FMT, fecal material has to be diluted and homogenized so that it can be easily administered. According to many protocols, freshly produced donor stool (50–300 g) is dissolved in sterile saline water and used preferably within 6 hours after emission. Water and other diluents (eg yogurt or milk) have also been described as a vehicle, with a trend towards improved outcome using larger volumes of prepared solution³⁷.

Almost all fecal preparations are processed in an aerobic environment. In most publications feces are infused as quickly as possible following production by the donor. The exposure of fecal microbiota to aerobic conditions, even briefly, could be detrimental to anaerobes and conversely favour aerobes, with potential consequences to FMT outcome. The fact that autologous FMT led to a change in microbiota in Rossen's trial suggests that the microbiota was altered by fecal processing and thus supports this hypothesis¹⁹. The majority of the beneficial bacteria in the gut are strict anaerobes, thus it is possible that preparing the transplant under anaerobic conditions could lead to better results.

For many techniques, logistical issues associated with preparation and use of fresh material may represent a limitation. To combat this FMT using frozen material has recently been developed, thus reducing the barriers presented by the number and frequency of donors, screening and the cost of such techniques. Frozen material is easier to use for both patients and physicians and, as there is a larger pool of samples to choose from, it allows the selection of the best matched donor. The standardization of donor material preparation using frozen feces will significantly simplify the clinical practice of FMT³⁸. Metagenomics studies highlight the role of storage conditions in maintaining intestinal microbiota integrity³⁹. The storage temperature (-20°C or -80°C) interferes in the conservation of DNA and freeze-thaw is known to impact microbiota. Lee C, *et al* conducted the first randomized controlled trial comparing fresh versus frozen FMT in recurrent CDI⁴⁰. FMT was given by enema and findings for frozen FMT and fresh FMT were similar in terms of efficacy and safety. A non-randomized study using oral administration of capsules containing frozen FMT also showed good results in terms of efficacy in CDI⁴¹. However, the expected effects of FMT in IBD are very different from those seen in recurrent CDI and therefore it is probable that the results seen in CDI cannot be extrapolated to IBD.

As well as differences in fecal microbiota preparation techniques, the procedure for preparing the recipient for transplant of the new microbiota also varies. Most of the authors use a classical colon cleansing with polyethylene glycol¹⁹, but others use large spectrum antibiotics¹¹ or no

preparation at all¹⁸. Although it seems logical to make some space for the new microbiota before administering it, the best way to do so has not been directly studied and thus remains an open question.

The optimum preparation of donor stool has not yet been determined and there is currently high heterogeneity from one study to another: fresh versus frozen stools; storage at -20°C versus -80°C; use of cryopreservative or not etc. Further studies are needed to identify the best approach and to set up a standard. Development of stool banks will help to reinforce homogeneity³⁶.

3. Route of administration

The route of delivery represents another tricky matter. As with donor stool preparation, there is no clear consensus on the best method of instillation.

Many different ways of administration of the new microbiota have been described and tested. FMT can be delivered via the upper or the lower gastrointestinal tract. By the upper digestive tract, suspension of healthy donor stool is administered using a tube via the stomach, duodenum or jejunum, or with oral ingestion of gelatin coated capsules or frozen capsules. By the lower digestive tract, FMT is given directly via the endoscope channel into the terminal ileum, caecum or sigmoid or using rectal enema.

The upper GI route might render the active constituent of FMT ineffective by the time it reaches the diseased colon. Conversely, in patients with extended colitis rectal enema might be insufficient to induce a reset of microbiota in the entire colon. Gastric juice can damage Bacteroidetes, whereas some Firmicutes need to pass through the upper gastrointestinal tract to be activated. A lower gastrointestinal route of administration appears to achieve better outcomes than upper in CDI⁴². It is likely that the choice of route of administration will depend on the microbes deemed important to be infused and on the disease being treated.

The number of infusions may be also critical. A single administration may be adequate for CDI, but not for chronic diseases such as IBD. It seems logical that, in IBD, the dysbiosis cannot be definitively corrected after only one FMT. It is possible that there is a dose response in FMT and the scheme and number of infusions might be important. Although the microbiota has an effect on the host, it is also known that the host shapes the microbiota. Indeed, it is clear that host genes and diet are able to profoundly modify the microbiota^{42, 43, 44}, suggesting that FMT could only have a transient effect in IBD.

4. Timing of FMT

One important question is the timing of FMT administration during the course of the disease. Firstly, at what point during the natural disease history should FMT be proposed? Data from Moayyedi *et al.* suggest that newly diagnosed UC patients may have the best outcome with FMT¹⁸, suggesting a potential window of opportunity to treat patients with FMT early after diagnosis. The perturbation in the intestinal homeostasis might be more easily restored early in the course of the disease.

It is also important to consider whether or not flare is the best time to perform FMT. It is known that inflammation by itself is a major driver of dysbiosis⁴⁵, therefore, performing FMT during active gut inflammation might result in only a transient effect, as the infused microbiota will be immediately altered by the recipient's inflammatory status. Moreover, the administration of a massive amount of microbial antigens on an inflamed and permeable mucosa may have, like adding fuel on the fire, some detrimental effect on the inflammatory process itself and could cause potential side effects such as microbial translocation. Therefore, a good treatment strategy might be to perform FMT in a patient who has achieved remission through conventional treatment.

5. Long-term safety of FMT

The greater concern about FMT relates to long term safety. Risks include possible transmission of infectious agents (including unknown ones) or the development of diseases related to changes in the gut microbiota. The gut microbiota is a complex consortium with many poorly characterized components. The impact of transferring these complex communities from one person to another is not known. Studies in mice indicate that the composition of gut microbiota can affect host susceptibility to disease. Some groups have reported transferring colitis phenotypes from different knockout mouse models to wild-type-mice^{46, 47}. In humans also, transfer of complex microbiota can modify the phenotypic expression of disease⁴⁸. Nevertheless, transmission of microbiota-driven disease in mice has been mostly shown in germ-free recipients and the effect of FMT in humans has been shown to be transient.

Long term follow-up of patients receiving FMT is mandatory to answer the questions concerning safety and future adverse events.

Conclusion

Fecal microbiota transplantation is a promising therapy in IBD but more studies are needed to define the best indications, optimal timing, frequency, mode of delivery and the optimal donor for each patient. Moreover, large studies will present the opportunity to identify active components of the gut microbiota that could then be isolated and used as potential therapeutic agents. Using this type of clinical-based rational selection process, we identified *Faecalibacterium prausnitzii* as a key player in intestinal homeostasis with therapeutic potential in IBD³¹. Anti-inflammatory molecules produced by this bacterium have been recently identified^{49, 50}. In the future, gut-derived bacteria, used either alone as next generation probiotics or in combination as artificial microbiota, could be used to counterbalance dysbiosis in a more controlled and standardized way compared to FMT.

Although IBD pathogenesis is related to an abnormal host-microbiota crosstalk, the only therapeutic strategy envisaged until recently was to inhibit the over activated immune system.

With the current rise of microbiota-derived therapeutic approaches, a similar mistake should not be done. The host immune response and the microbiota are two players of the disease pathogenesis which should be taken into account together and probably targeted simultaneously to achieve optimal results.

References

1. Khor, B., Gardet, A. & Xavier, R.J. Genetics and pathogenesis of inflammatory bowel disease. *Nature* **474**, 307-317 (2011).
2. Sokol, H. & Seksik, P. The intestinal microbiota in inflammatory bowel diseases: time to connect with the host. *Curr Opin Gastroenterol* **26**, 327-331 (2010).
3. Sokol, H. *et al.* Low counts of *Faecalibacterium prausnitzii* in colitis microbiota. *Inflamm Bowel Dis* **15**, 1183-1189 (2009).
4. Manichanh, C. *et al.* Reduced diversity of faecal microbiota in Crohn's disease revealed by a metagenomic approach. *Gut* **55**, 205-211 (2006).
5. Mangin, I. *et al.* Molecular inventory of faecal microflora in patients with Crohn's disease. *FEMS Microbiol Ecol* **50**, 25-36 (2004).
6. Gevers, D. *et al.* The treatment-naive microbiome in new-onset Crohn's disease. *Cell Host Microbe* **15**, 382-392 (2014).
7. Morgan, X.C. *et al.* Dysfunction of the intestinal microbiome in inflammatory bowel disease and treatment. *Genome Biol* **13**, R79 (2012).
8. van Nood, E. *et al.* Duodenal infusion of donor feces for recurrent *Clostridium difficile*. *N Engl J Med* **368**, 407-415 (2013).
9. Leffler, D.A. & Lamont, J.T. *Clostridium difficile* Infection. *N Engl J Med* **373**, 287-288 (2015).
10. Gupta, S., Allen-Vercoe, E. & Petrof, E.O. Fecal microbiota transplantation: in perspective. *Therap Adv Gastroenterol* **9**, 229-239 (2016).
11. Angelberger, S. *et al.* Temporal bacterial community dynamics vary among ulcerative colitis patients after fecal microbiota transplantation. *Am J Gastroenterol* **108**, 1620-1630 (2013).
12. Suskind, D.L., Singh, N., Nielson, H. & Wahbeh, G. Fecal microbial transplant via nasogastric tube for active pediatric ulcerative colitis. *J Pediatr Gastroenterol Nutr* **60**, 27-29 (2015).
13. Cui, B. *et al.* Step-up fecal microbiota transplantation strategy: a pilot study for steroid-dependent ulcerative colitis. *J Transl Med* **13**, 298 (2015).

14. Kunde, S. *et al.* Safety, tolerability, and clinical response after fecal transplantation in children and young adults with ulcerative colitis. *J Pediatr Gastroenterol Nutr* **56**, 597-601 (2013).
15. Colman, R.J. & Rubin, D.T. Fecal microbiota transplantation as therapy for inflammatory bowel disease: a systematic review and meta-analysis. *J Crohns Colitis* **8**, 1569-1581 (2014).
16. Landy, J. *et al.* Variable alterations of the microbiota, without metabolic or immunological change, following faecal microbiota transplantation in patients with chronic pouchitis. *Sci Rep* **5**, 12955 (2015).
17. Fang, S. *et al.* Successful treatment of chronic Pouchitis utilizing fecal microbiota transplantation (FMT): a case report. *Int J Colorectal Dis* **31**, 1093-1094 (2016).
18. Moayyedi, P. *et al.* Fecal Microbiota Transplantation Induces Remission in Patients With Active Ulcerative Colitis in a Randomized Controlled Trial. *Gastroenterology* **149**, 102-109 e106 (2015).
19. Rossen, N.G. *et al.* Findings From a Randomized Controlled Trial of Fecal Transplantation for Patients With Ulcerative Colitis. *Gastroenterology* **149**, 110-118 e114 (2015).
20. Sha, S. *et al.* Systematic review: faecal microbiota transplantation therapy for digestive and nondigestive disorders in adults and children. *Aliment Pharmacol Ther* **39**, 1003-1032 (2014).
21. Suskind, D.L. *et al.* Fecal microbial transplant effect on clinical outcomes and fecal microbiome in active Crohn's disease. *Inflamm Bowel Dis* **21**, 556-563 (2015).
22. Cui, B. *et al.* Fecal microbiota transplantation through mid-gut for refractory Crohn's disease: safety, feasibility, and efficacy trial results. *J Gastroenterol Hepatol* **30**, 51-58 (2015).
23. Kelly, C.R. *et al.* Update on Fecal Microbiota Transplantation 2015: Indications, Methodologies, Mechanisms, and Outlook. *Gastroenterology* **149**, 223-237 (2015).
24. De Leon, L.M., Watson, J.B. & Kelly, C.R. Transient flare of ulcerative colitis after fecal microbiota transplantation for recurrent *Clostridium difficile* infection. *Clin Gastroenterol Hepatol* **11**, 1036-1038 (2013).
25. Khoruts, A. *et al.* Inflammatory Bowel Disease Affects the Outcome of Fecal Microbiota Transplantation for Recurrent *Clostridium difficile* Infection. *Clin Gastroenterol Hepatol* (2016).

26. Vermeire, S. *et al.* Donor Species Richness Determines Faecal Microbiota Transplantation Success in Inflammatory Bowel Disease. *J Crohns Colitis* **10**, 387-394 (2016).
27. Baxter, M., Ahmad, T., Colville, A. & Sheridan, R. Fatal Aspiration Pneumonia as a Complication of Fecal Microbiota Transplant. *Clin Infect Dis* **61**, 136-137 (2015).
28. Alang, N. & Kelly, C.R. Weight gain after fecal microbiota transplantation. *Open Forum Infect Dis* **2**, ofv004 (2015).
29. Grinspan, A.M. & Kelly, C.R. Fecal Microbiota Transplantation for Ulcerative Colitis: Not Just Yet. *Gastroenterology* **149**, 15-18 (2015).
30. Sokol, H. Toward Rational Donor Selection in Faecal Microbiota Transplantation for IBD. *J Crohns Colitis* **10**, 375-376 (2016).
31. Sokol, H. *et al.* Faecalibacterium prausnitzii is an anti-inflammatory commensal bacterium identified by gut microbiota analysis of Crohn disease patients. *Proc Natl Acad Sci U S A* **105**, 16731-16736 (2008).
32. Morrison, D.J. & Preston, T. Formation of short chain fatty acids by the gut microbiota and their impact on human metabolism. *Gut Microbes*, 1-12 (2016).
33. Ding, T. & Schloss, P.D. Dynamics and associations of microbial community types across the human body. *Nature* **509**, 357-360 (2014).
34. Subramanian, S. *et al.* Persistent gut microbiota immaturity in malnourished Bangladeshi children. *Nature* **510**, 417-421 (2014).
35. Li, S.S. *et al.* Durable coexistence of donor and recipient strains after fecal microbiota transplantation. *Science* **352**, 586-589 (2016).
36. Kazerouni, A., Burgess, J., Burns, L.J. & Wein, L.M. Optimal screening and donor management in a public stool bank. *Microbiome* **3**, 75 (2015).
37. Gough, E., Shaikh, H. & Manges, A.R. Systematic review of intestinal microbiota transplantation (fecal bacteriotherapy) for recurrent *Clostridium difficile* infection. *Clin Infect Dis* **53**, 994-1002 (2011).
38. Hamilton, M.J., Weingarden, A.R., Sadowsky, M.J. & Khoruts, A. Standardized frozen preparation for transplantation of fecal microbiota for recurrent *Clostridium difficile* infection. *Am J Gastroenterol* **107**, 761-767 (2012).
39. Cardona, S. *et al.* Storage conditions of intestinal microbiota matter in metagenomic analysis. *BMC Microbiol* **12**, 158 (2012).

40. Lee, C.H. *et al.* Frozen vs Fresh Fecal Microbiota Transplantation and Clinical Resolution of Diarrhea in Patients With Recurrent *Clostridium difficile* Infection: A Randomized Clinical Trial. *JAMA* **315**, 142-149 (2016).
41. Youngster, I. *et al.* Oral, capsulized, frozen fecal microbiota transplantation for relapsing *Clostridium difficile* infection. *JAMA* **312**, 1772-1778 (2014).
42. Kassam, Z., Lee, C.H., Yuan, Y. & Hunt, R.H. Fecal microbiota transplantation for *Clostridium difficile* infection: systematic review and meta-analysis. *Am J Gastroenterol* **108**, 500-508 (2013).
43. David, L.A. *et al.* Diet rapidly and reproducibly alters the human gut microbiome. *Nature* **505**, 559-563 (2014).
44. Rawls, J.F., Mahowald, M.A., Ley, R.E. & Gordon, J.I. Reciprocal gut microbiota transplants from zebrafish and mice to germ-free recipients reveal host habitat selection. *Cell* **127**, 423-433 (2006).
45. Winter, S.E. *et al.* Host-derived nitrate boosts growth of *E. coli* in the inflamed gut. *Science* **339**, 708-711 (2013).
46. Garrett, W.S. *et al.* Communicable ulcerative colitis induced by T-bet deficiency in the innate immune system. *Cell* **131**, 33-45 (2007).
47. Elinav, E. *et al.* NLRP6 inflammasome regulates colonic microbial ecology and risk for colitis. *Cell* **145**, 745-757 (2011).
48. Vrieze, A. *et al.* Transfer of intestinal microbiota from lean donors increases insulin sensitivity in individuals with metabolic syndrome. *Gastroenterology* **143**, 913-916 e917 (2012).
49. Quevrain, E. *et al.* Identification of an anti-inflammatory protein from *Faecalibacterium prausnitzii*, a commensal bacterium deficient in Crohn's disease. *Gut* **65**, 415-425 (2016).
50. Miquel, S. *et al.* Identification of metabolic signatures linked to anti-inflammatory effects of *Faecalibacterium prausnitzii*. *MBio* **6** (2015).

Author Contributions

BP and HS wrote the manuscript. They contribute equally to this work

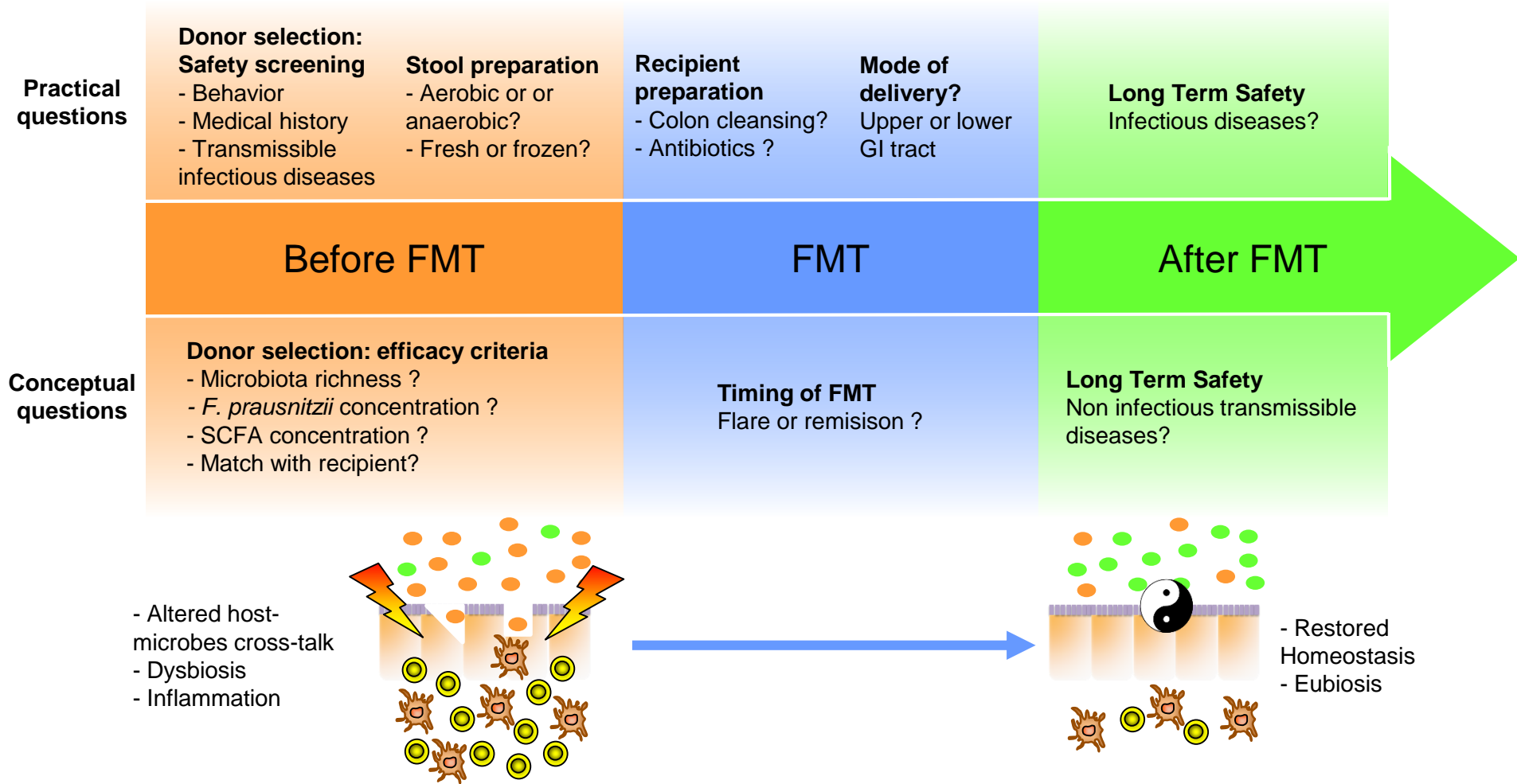


Figure 1: Fecal microbiota transplantation in IBD: unsolved questions.