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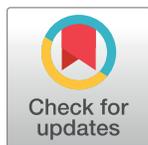
RESEARCH ARTICLE

Physiologic Determinants of Exercise Capacity in Pulmonary Langerhans Cell Histiocytosis: A Multidimensional Analysis

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Abstract

Background

Reduced exercise capacity severely impacts quality of life in pulmonary Langerhans cell histiocytosis. Ascertaining mechanisms that impair exercise capacity is necessary to identify targets for symptomatic treatments.

Methods

Dyspnea, pulmonary function tests and cardiopulmonary exercise test were analysed in 62 study participants. Data were compared between subjects with impaired and normal aerobic capacity ($V'O_2$ peak less than 84% versus 84% predicted or more). Data were reduced using a principal component analysis. Multivariate analysis included $V'O_2$ peak as the dependent variable and principal components as covariates.

Results

$V'O_2$ peak was reduced in 44 subjects (71%). Subjects with impaired aerobic capacity presented: (i) decreased FEV₁, FVC, FEV₁/FVC, DL_{CO} and DL_{CO}/VA and increased AaDO₂, (ii) increased ventilatory equivalents at ventilatory threshold, V_D/V_T peak, AaDO₂ peak and PaCO₂ peak and decreased ventilatory reserve and PaO₂ peak. There was no difference

between groups in dyspnea scores. Principal component analysis extracted 4 principal components interpreted as follows: PC1: gas exchange; PC2: "pseudorestriction"; PC3: exercise-induced hyperpnea; PC4: air trapping. Multivariate analysis explained 65% of $\dot{V}O_2$ peak. The 4 principal components were independently associated with $\dot{V}O_2$ peak (β coefficients: PC1: 9.3 [4.6; 14], PC2: 7.5 [3; 11.9], PC3: -5.3 [-9.6; -1.], PC4: -9.8 [-14.9; -4.7]).

Conclusion

Impaired exercise capacity is frequent in pulmonary Langerhans cell histiocytosis. It is mainly caused by pulmonary changes but is not associated with increased dyspnea intensity. Therefore, treating the lung represents a relevant approach for improving exercise capacity, even in patients experiencing mild dyspnea.

Introduction

Langerhans cell histiocytosis is a rare systemic disease characterized by proliferation of Langerhans cells that infiltrate the tissues. In adults, this disorder affects a limited number of organs, predominantly the lungs, skin and bone [1]. Pulmonary Langerhans cell histiocytosis (PLCH) is mostly observed in genetically predisposed young adult smokers [2]. The most common symptoms are cough, breathlessness and fatigue. Chest computed tomography (CT) scan typically shows bilateral micronodular interstitial syndrome, excavated nodules and cysts predominantly located in the upper and middle areas of the lung [3]. Pulmonary hypertension is present in 10% of cases [3] due to infiltration of pulmonary capillaries by Langerhans cells [4] or complicating the interstitial lung disease. Pulmonary function tests (PFT) show various abnormalities including reduction of the diffusing capacity of the lung for carbon monoxide (DL_{CO}), decreased vital capacity (VC), increased residual volume (RV) and airway obstruction [5].

PLCH impairs daily life activities and deteriorates quality of life [6,7]. Accordingly, it is associated with decreased maximum oxygen consumption ($\dot{V}O_2$ peak) and reduced distance walked during a 6-minute walk test [8,9]. The pathophysiologic mechanisms that limit exercise capacity have been poorly investigated and consequently, the effect of symptomatic treatments is unknown. One study reported correlations between $\dot{V}O_2$ peak and the resting dead space/tidal volume ratio (V_D/V_T), RV, resting alveolar-to-arterial oxygen tension difference ($AaDO_2$) and DL_{CO} [8]. However, in that study the relationship between $\dot{V}O_2$ peak and variables at exercise, which are rather impaired and provide a better understanding of exercise capacity alteration, were not studied.

Cardiopulmonary exercise testing (CPET) is a crucial tool for determining the respective roles of cardiovascular, respiratory or peripheral responses in exercise limitation [10]. For instance, in patients with sarcoidosis, CPET revealed that exercise capacity is essentially limited by the cardiocirculatory response in mild-to-moderate stages of the disease, while it is mainly limited by gas exchange impairment in severe stages [11]. Moreover, CPET can reveal exercise-induced physiologic dysfunction able to explain impaired exercise capacity in patients with normal PFT [10]. In PLCH, $AaDO_2$ at peak exercise can be increased whereas DL_{CO} is normal [8]. However, no data are available on the relationship between $\dot{V}O_2$ peak and physiologic variables measured during exercise. Hence, this study was designed to investigate the physiologic determinants of reduced exercise capacity in patients with PLCH.

Methods

Design

We conducted a multicenter retrospective study in 8 hospitals. All patients referred between 1993 and 2013 at the time of diagnosis or during follow-up of PLCH were screened for inclusion. Patients who had performed several CPET were included only once. In this condition, only the first available CPET was selected. Inclusion criteria were: (i) a diagnosis of PLCH confirmed by histologic examination or a combination of typical clinical history and chest CT scan features, (ii) a maximal CPET including arterial blood gases at peak exercise.

Ethics

The study was approved by the Institutional Review Board of the French Learned Society for Pulmonology (CEPRO 2012 039).

Dyspnea score

Limitation of activity related to dyspnea was evaluated using the modified Medical Research Council (mMRC) scale.

Chest CT scans

Chest CT scans performed up to six months after CPET were analysed. Chest CT scans performed more than 6 months after CPET were not considered for analysis. The prevalence of nodular and cystic abnormalities was analysed by two blinded radiologists (K.Y. and N.B.). Chest CT changes were classified as previously described [12]. The extent of nodular lesions was categorised as follows: no nodules, mild, moderate, or diffuse nodules. The extent of cystic lesions was classified by the percentages of the lung surface analysed: no cyst, <25%, 25–49%, 50–75%, or >75%.

Echocardiography

Echocardiography performed up to 6 months after CPET were analysed. Pulmonary artery pressure was collected.

Pulmonary function tests

Forced vital capacity (FVC), forced expiratory volume in 1 s (FEV_1), FEV_1 to FVC ratio, total lung capacity (TLC), functional residual capacity (FRC) and RV were evaluated by plethysmography. DLco was measured using the single-breath method after adjusting for hemoglobin concentration and according to Cotes' equation. Predicted normal values were derived from standard equations [13]. Values were expressed as percentages of the predicted normal values calculated according to gender, weight, and age. Reference equations were taken from ERS [14].

CardioPulmonary Exercise Testing

CPET was performed according to a similar standardized protocol in the 8 centers on the same day as PFT. Short and long acting bronchodilators were withheld for at least 8 and 24 hours before the exam, respectively. The CPET protocol consisted in a symptom-limited incremental exercise test on an ergometric bicycle including a warm-up period of 3 min at 20 W followed by a progressively increasing work rate in a ramp fashion (8 to 30 W/min) and then 3 min recovery. It is described in details in previous publications [15]. Immediately after

exercise, subjects were asked to score breathlessness and muscle fatigue at the end of exercise using Borg scales. Limitation of aerobic capacity was defined as $\dot{V}O_2$ peak < 84% of predicted [10].

Statistical analyses

Continuous variables were expressed as mean \pm standard deviation or median and interquartile range according to their normal or non-normal distribution. Qualitative variables were expressed as sample size and percentage of the population. Continuous variables were compared between groups using a Student's T test or a Mann-Whitney test. Qualitative variables were compared using Chi-square or Fischer's test. Correlation between $\dot{V}O_2$ peak, CPET and PFT variables were evaluated using a Pearson's coefficient or a Spearman's coefficient. Principal component analysis (PCA) was performed in order to resume the data in principal components. When a variable was expressed both in absolute value and percentage of predicted, only the variable expressed as percentage of predicted was selected. We retained factors with eigenvalue greater than 1. Rotation was performed by the varimax method. Linear regression was performed using $\dot{V}O_2$ peak as the dependent variable and PCA-extracted factors as covariates. Linear regression was adjusted for age, gender, weight, height, active smoking (yes/no) and duration of disease. The normality of the distribution of residuals and the diagnosis of homoscedasticity were evaluated graphically. Statistical analyses were performed on SPSS (IBM SPSS Statistics 22); the threshold for statistical significance was set to $p < 0.05$.

Results

Sociodemographic and clinical characteristics

A total of 62 subjects (27 men, 35 women) with a mean age of 37 ± 10 years were included. The diagnosis of PLCH was confirmed by suggestive clinical features associated with typical CT lesions in 63% of cases, surgical lung biopsy in 31% of cases and extrathoracic biopsy in 6% of cases. Subjects were Caucasians in 94% of cases. The study population comprised 66% of smokers, 31% of ex-smokers and 3% of non-smokers. The median interval between the date of diagnosis and the date of CPET was 3 years. No statistically significant difference was observed for sociodemographic data and dyspnea severity between subjects with or without reduced $\dot{V}O_2$ peak (Table 1).

CT scan

A CT scan was available in 37 cases (Table 1). CT scans revealed varying degrees of nodular and cystic abnormalities: 30/37 patients had both nodules and cysts; 3 patients had an isolated cystic pattern, and 4 patients had an isolated nodular pattern. The mean CT nodular score was 7.7 ± 5.6 and the mean CT cystic score was 9.1 ± 6.3 . The mean cystic score was statistically higher in subjects with reduced $\dot{V}O_2$ peak.

Echocardiography

An echocardiography was available in 21 cases (median sPAP 29 mmHg (23–37)). Of those, 14 had reduced $\dot{V}O_2$ peak and 7 had normal $\dot{V}O_2$ peak (sPAP 28 (22–37) versus 29 (28–37); $p = 0.65$). Pulmonary hypertension was suspected (sPAPs > 35mmHg) in 6 patients. No right catheterization was available. No correlation was observed between sPAP and $\dot{V}O_2$ peak; $r = 0.14$, $p = 0.54$).

Table 1. Sociodemographic and clinical characteristics.

	Total	V'O ₂ peak ≥ 84% pred	V'O ₂ peak < 84% pred	p
Sociodemographic data	n = 62	n = 18	n = 44	
Gender, male	27 (43%)	5 (28%)	22 (50%)	0.109
Age, years	37 ± 10	39 ± 10	36 ± 11	0.323
BMI, kg/m ²	21.8 (6.4)	21.9 (6.8)	21.5 (6.4)	0.625
Ethnic group, Caucasian	52 (94%)	16 (89%)	36 (82%)	0.254
History of Smoking				
Smoker	41 (66%)	13 (72%)	28 (64%)	0.598
Ex-smoker	19 (31%)	5 (28%)	14 (32%)	
Non-smoker	2 (3%)	0 (0%)	2 (4.5%)	
Clinical data	n = 62	n = 18	n = 44	
Interval between diagnosis and evaluation, years	3 (6)	1 (6)	4 (7)	0.262
mMRC scale	1 (1)	0.8 (1)	0.9 (2)	0.8
CT scan data	n = 37	13	24	
Nodular score	6 (3.5–11)	6 (3.5–9)	7 (3.25–14)	0.59
Low	20 (54%)	8 (62%)	12 (50%)	
Intermediate	9 (24%)	4 (31%)	5 (21%)	
High	8 (22%)	1 (7%)	7 (29%)	
Cystic score	7 (6–13.5)	6 (5–6)	9.5 (6–14)	0.024
Low	19 (51%)	11 (85%)	8 (33%)	
Intermediate	9 (24%)	0 (0%)	9 (37%)	
High	5 (14%)	2 (15%)	3 (13%)	
Very high	4 (11%)	0 (0%)	4 (17%)	

BMI: body mass index, CT: computed tomography, mMRC: modified Medical Research Council, V'O₂ peak: maximal oxygen consumption. Results are expressed as n(%), mean ± SD or median (Q1-Q3)

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Pulmonary function tests

PFT results are summarised in Table 2. In subjects with reduced V'O₂ peak, FEV₁, FVC, FEV₁/FVC, DL_{CO} and DL_{CO}/VA were significantly lower and AaDO₂ was significantly higher compared with subjects with normal V'O₂ peak. No statistically significant difference was observed between groups in RV, FRC, TLC, RV/TLC, PaO₂ and PaCO₂.

CardioPulmonary Exercise Testing

A maximal symptom-limited exercise was obtained in all cases judged on symptoms and/or a functional limitation as all patients had at least one of the following: breathing reserve (BR) < 15%, peak heart rate (HR) > 90% of predicted, peak lactate > 7 mEq/L, peak PaO₂ < 55 mmHg and peak ventilator equivalent for O₂ (VE/VO₂) > 35 or peak Respiratory Exchange Ratio > 1.15. (10) Subjects attributed discontinuation of exercise to breathlessness in 1/3 of cases (n = 14), muscle fatigue in 1/3 of cases (n = 12) and both in 1/3 of cases (n = 17). The CPET results are summarized in Table 2. The median dyspnoea Borg intensity at the end of maximum incremental exercise was scored as severe with no statistically significant difference between the 2 groups (5(7) versus 5.5(4), p = 0.74). V'O₂ peak was reduced in 44 (71%) subjects. No HR, blood pressure or electrocardiographic abnormalities were observed. Oxygen pulse was greater than 70% of predicted in 54 of the 62 subjects and displayed normal kinetics in all subjects. It is of note that 6 of the 8 subjects with a lower than 70% predicted oxygen

Table 2. Pulmonary function tests and incremental cycle exercise results.

	Total	V'O ₂ peak ≥ 84% pred	V'O ₂ peak < 84% pred	p
Pulmonary Function Test	n = 62	n = 18	n = 44	
FEV ₁ , % pred	74 ± 25	95 ± 16	66 ± 23	< 0.001
FVC, % pred	91 ± 22	106 ± 17	84 ± 20	< 0.001
FEV ₁ /FVC, %	68 ± 14	75 ± 7	65 ± 15	< 0.001
RV, % pred	129 ± 50	117 ± 25	115 (81)	0.821
FRC, % pred	115 ± 30	119 ± 23	113 ± 32	0.508
TLC, % pred	102 ± 19	108 ± 14	99 ± 21	0.112
RV/TLC, %	36 (17)	33 ± 8	37 (20)	0.352
DL _{CO} , % pred	61 ± 19	75 ± 19	55 ± 15	< 0.001
PaO ₂ , mmHg	87 ± 13	91 ± 15	86 ± 11	0.118
PaCO ₂ , mmHg	37 ± 4	37 ± 5	37 ± 4	0.656
AaDO ₂ , mmHg	18 (23)	10 (15)	22 ± 13	0.049
Incremental cycle exercise	n = 62	n = 18	n = 44	
Workload peak, % pred	71 ± 19	85 ± 16	65 ± 17	< 0.001
V'O ₂ peak, % pred	73.7 ± 17.9	95 ± 9	66 (15)	< 0.001
V'E peak, L/min	67 ± 22	72 ± 14	65 ± 25	0.181
V'E/V'CO ₂ peak	38 ± 8	36 ± 7	38 (12)	0.113
V'E/V'O ₂ VT	33 (12)	30 (6)	36 ± 7	0.023
V'E/V'CO ₂ VT	34 (12)	33 ± 7	38 ± 8	0.047
BR, %	31 (28)	37 ± 12	25 ± 21	0.005
V _D /V _T peak	0.33 ± 0.11	0.27 ± 0.11	0.36 ± 0.11	0.006
V _D /V _T rest	0.38 ± 0.13	0.35 ± 0.09	0.39 ± 0.14	0.337
PaO ₂ peak, mmHg	79 ± 18	94 (24)	74 ± 17	0.001
PaCO ₂ peak, mmHg	38 ± 5	35 ± 4	39 ± 5	0.005
AaDO ₂ peak, mmHg	35 (26)	22 (11)	38 ± 15	0.004

AaDO₂: Alveolar-to-arterial oxygen tension difference; BR: Breathing Reserve; DL_{CO}: Diffusing Capacity of the Lung for carbon monoxide; FEV₁: Forced Expiratory Volume in 1 s; FRC: Functional Residual Capacity; FVC: Forced Vital Capacity; HR: Heart Rate; PaCO₂: partial pressure of carbon dioxide in arterial blood; PaO₂: Partial pressure of oxygen in arterial blood; RR: Respiratory Rate; RV: Residual Volume; TLC: Total Lung Capacity; VA: Alveolar Volume; V_D/V_T: dead space/tidal volume ratio; V'E: minute ventilation; V'E/V'O₂: ventilatory equivalent for O₂; V'E/V'CO₂: ventilatory equivalent for CO₂; V_T: Tidal Volume; VT: ventilatory threshold; V'O₂: oxygen consumption.

Results are expressed as mean ± SD or median (Q1-Q3)

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pulse had performed echocardiography showing no sign of pulmonary hypertension (pulmonary artery systolic pressure lower than 40 mmHg) or left ventricular dysfunction (data not shown). Subjects with decreased V'O₂ peak had significantly higher ventilatory equivalents at ventilator threshold (VT), V_D/V_T peak, AaDO₂ peak and PaCO₂ peak and lower BR, PaO₂ peak and oxygen pulse. In 21 patients with a recent echocardiography, pulmonary artery pressure did not correlate with VD/VT at rest and peak (data not shown).

Correlations between V'O₂ peak and CPET/PFT variables

V'O₂ peak was not correlated with either mMRC score (r = -0.21, p = 0.11) or Borg score (r = 0.023; p = 0.89). Nineteen of the 40 PFT and CPET variables recorded were statistically correlated with V'O₂ peak (Table 3). We did not consider the correlation between VO₂ peak and pulse oxygen, which is evidently high but not informative, as the calculation of pulse oxygen includes VO₂ peak.

Table 3. Variables statistically correlated with V'O₂ peak.

	n	r	p
FEV ₁ , % pred	62	0.709	< 0.001
FVC, % pred	62	0.543	< 0.001
FEV ₁ /FVC, %	62	0.582	< 0.001
RV, % pred	57	-0.286	0.031
RV/TLC, %	58	-0.291	0.027
DL _{CO} , % pred	57	0.662	< 0.001
DL _{CO} /VA, % pred	53	0.533	< 0.001
PaO ₂ rest, mmHg	59	0.409	0.001
AaDO ₂ rest, mmHg	54	-0.484	< 0.001
V'E peak, L/min	62	0.349	0.005
V'E/V'CO ₂ peak	62	-0.297	0.019
V'E/V'O ₂ VT	58	-0.436	0.001
V'E/V'CO ₂ VT	58	-0.390	0.002
BR, %	62	0.410	0.001
V _D /V _T peak	58	-0.475	< 0.001
V _D /V _T rest	59	-0.265	0.043
PaO ₂ peak, mmHg	58	0.553	< 0.001
PaCO ₂ peak, mmHg	58	-0.445	< 0.001
AaDO ₂ peak, mmHg	54	-0.504	< 0.001

AaDO₂: alveolar-to-arterial oxygen tension difference; BR: breathing reserve; DL_{CO}: Diffusing capacity of the lung for carbon monoxide; FEV₁: forced expiratory volume in 1s; FRC: functional residual capacity; FVC: Forced vital capacity; PaCO₂: partial pressure of carbon dioxide in arterial blood; PaO₂: partial pressure of oxygen in arterial blood; RV: residual volume; TLC: total lung capacity; VA: alveolar volume; V_D/V_T: dead space/tidal volume ratio; V'E: minute ventilation; V'E/V'CO₂: ventilatory equivalent for CO₂; V'E/V'O₂: ventilatory equivalent for O₂; V'O₂: maximal oxygen consumption

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Principal component analysis

After exclusion of the subjects with any missing data, 45 subjects were included in the principal component analysis. Subjects with and without missing data showed similar sociodemographic, PFT and CPET characteristics apart from higher AaDO₂ at rest and peak and lower PaO₂ peak in the missing data group (S1 Table).

Four factors had eigenvalues greater than 1. The first factor alone explained 48% of total variance. The 4 factors explained 79% of total variance. The component matrix after rotation is shown in Table 4.

Principal component 1 (PC1) was associated with variables measuring gas exchange, as it was negatively correlated with AaDO₂ at rest (r = -0.79) and peak (r = -0.804) and positively correlated with PaO₂ at rest (r = 0.775) and peak (r = 0.743) and DL_{CO}/VA (r = 0.683). PC1 was also associated with FEV₁/FVC (r = 0.551). Principal component 2 (PC2) was associated with lung volumes, as it was positively correlated with FVC (r = 0.872) and FEV₁ (r = 0.803) and with variables including lung volumes in their calculation: DL_{CO} (r = 0.567), BR (r = 0.595), V_D/V_T peak (r = -0.619) and V_D/V_T at rest (r = -0.571). PC2 was also negatively correlated with PaCO₂ peak (r = -0.735). Principal component 3 (PC3) was correlated with ventilatory equivalents: peak V'E/V'CO₂ (r = 0.910), VT V'E/V'CO₂ (r = 0.726) and VT V'E/V'O₂ (r = 0.762). Principal component 4 (PC4) was positively correlated with measurements of air trapping including RV (r = 0.818) and RV/TLC (r = 0.845) and negatively correlated with V'E peak (r = -0.718).

Table 4. Correlations between the four principal components and CPET and PFT variables, n = 45.

	Principal component			
	1	2	3	4
AaDO ₂ peak, mmHg	-0.804	-0.354	0.223	-0.108
AaDO ₂ rest, mmHg	-0.790	0.023	0.317	0.272
PaO ₂ rest, mmHg	0.775	0.100	-0.150	-0.267
PaO ₂ peak, mmHg	0.743	0.566	-0.081	-0.054
DL _{co} /VA, % pred	0.683	0.188	-0.398	0.053
FEV ₁ /FVC, %	0.551	0.354	-0.058	-0.493
FVC, %	-0.015	0.872	-0.127	-0.176
FEV ₁ , %	0.302	0.803	-0.118	-0.420
PaCO ₂ peak, mmHg	-0.140	-0.735	-0.506	0.131
V _D /V _T peak	-0.387	-0.619	0.495	0.143
BR, %	0.232	0.595	-0.584	-0.082
V _D /V _T , resting	-0.396	-0.571	0.210	0.183
DL _{co} , % pred	0.504	0.567	-0.416	-0.058
V'E/V'CO ₂ peak	-0.227	-0.118	0.910	-0.007
V'E/V'O ₂ VT	-0.504	-0.070	0.762	0.243
V'E/V'CO ₂ VT	-0.416	-0.138	0.726	0.323
RV/TLC, %	-0.071	-0.316	0.312	0.845
RV, % pred	0.017	-0.110	0.229	0.818
V'E peak, L/min	0.253	0.131	0.283	-0.718

AaDO₂: Alveolar-to-arterial Oxygen tension Difference; BR: Breathing Reserve; DL_{co}: Diffusing capacity of the Lung for Carbon monoxide; FEV₁: Forced Expiratory Volume in 1 s; FVC: Forced Vital Capacity; PaCO₂: Partial pressure of carbon dioxide in arterial blood; PaO₂: Partial pressure of oxygen in arterial blood; RV: Residual Volume; TLC: Total Lung Capacity; VA: Alveolar Volume; V_D/V_T: dead space/tidal volume ratio; V'E: minute ventilation; V'E/V'CO₂: ventilatory equivalent for CO₂; V'E/V'O₂: ventilatory equivalent for O₂

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Association between V'O₂ peak and PFT and CPET variables

V'O₂ peak was independently correlated with the 4 principal components (Table 5). Overall, 65% of the variability of V'O₂ peak was explained by the regression model (R² = 0.65). Graphic analysis showed that the residues of linear regression followed a normal distribution and were

Table 5. Association between V'O₂ peak and the four principal components, n = 45.

	Beta coefficient	95% confidence interval	p
PC1	9.3	[4.6;14]	< 0.001
PC2	7.5	[3;11.9]	0.002
PC3	-5.3	[-9.6;-1]	0.017
PC4	-9.8	[-14.9;-4.7]	< 0.001
Age, year	0.6	[0;1.1]	0.047
Gender, male	9.3	[-2.3;20.9]	0.114
Weight, kg	-0.4	[-0.8;0]	0.316
Height, m	21.5	[-55.6;98.7]	0.575
Disease duration, year	0.8	[-0.2;1.7]	0.103
Active smoking, yes	-3	[-13.2;7.2]	0.552

PC: principal component

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scattered homogeneously over the spectrum of values of $\dot{V}'O_2$ peak, confirming the homoscedasticity of the model.

Discussion

This study provides a revisited analysis of physiologic determinants of exercise capacity in PLCH including the largest population studied until now and a more comprehensive set of measurements on exercise endorsed by a principal component analysis. The main results of this study can be summarized as follows: (i) impaired exercise capacity is common among patients with moderate PLCH, (ii) it is caused by multifactorial alteration of lung function including alteration of gas exchange, air trapping, pseudorestriction and exercise-induced hyperventilation (iii) it is not associated with increased dyspnea intensity.

Crausman et al. reported that patients with PLCH have severe alteration of aerobic capacity (mean $\dot{V}'O_2$ peak: 44% of predicted) [8]. The authors attributed exercise limitation to pulmonary hypertension based on the strong correlation between $\dot{V}'O_2$ peak and the increased V_D/V_T ratio at rest, which is an indirect but nonspecific sign of pulmonary hypertension. This hypothesis was not corroborated by a direct measurement of pulmonary artery pressure. In our study, the maximum exercise capacity was less severely impaired (mean $\dot{V}'O_2$ peak: 74% of predicted). This may be due a less severe disease as suggested by low dyspnea scores (median mMRC: 1 versus 2 in previous studies [7,12,16] and only slightly altered PFT (mean FVC: 91% and mean DL_{CO} : 61%) or a lower prevalence of pulmonary hypertension that was estimated at 29% IC95 [9 to 48%] in a subgroup of 21 with echocardiography measurements in our study. Our results show that the reduction of aerobic capacity in PLCH has a multifactorial origin, involving alteration of gas exchange, air trapping, “pseudorestriction” and exercise-induced hyperpnea. It is of note that although exercise was limited by pulmonary changes there was no difference in dyspnea intensity between patients with reduced and normal exercise capacity. This advocates that respiratory symptoms may not be necessarily warning sign for lung related reduction of exercise capacity in PLCH.

In our study, the alteration of gas exchange (PC1) was the main determinant of exercise capacity. All variables measuring gas exchange (DL_{CO} , DL_{CO}/VA and $AaDO_2$) were more severely altered in patients with decreased aerobic capacity and were correlated with PC1 that was strongly associated with $\dot{V}'O_2$ peak. In PLCH, the alteration of gas exchange is mainly due to cystic destruction of lung parenchyma. The correlation between FEV_1/FVC and PC1 is likely due to the strong relationship between bronchial obstruction and the extent of cysts as reported previously [12,16] and also found in our study in the sub-group of 37 patients having a CT scan (data not shown).

Air trapping (PC4) was the second major factor responsible for limitation of exercise capacity. In PLCH, air trapping is caused by obstruction of distal airways due to several mechanisms including infiltration by histiocytes and compression of airways by cysts. RV is increased very early in PLCH(8) and then remains nearly unchanged during the course of the disease [7]. This may explain the absence of any statistically significant difference in RV between patients with and without impaired aerobic capacity. In chronic obstructive pulmonary disease, air trapping impacts exercise capacity by inducing dynamic hyperinflation [17, 18] leading to reduced inspiratory capacity and finally to an inability to increase V_T [19]. We did not have any flow-volume loops available to confirm dynamic hyperinflation. However, the correlation between $\dot{V}'E$ and PC4 is an indirect argument in favour of the development of dynamic hyperinflation, as hyperventilation induces dynamic hyperinflation in obstructive diseases [20].

The third factor limiting exercise in our study was ventilatory abnormalities (PC2). Dynamic volumes (FEV_1 , FVC) were decreased in patients with impaired aerobic capacity,

while static volumes (TLC, FRC) were similar in patients with and without impaired aerobic capacity. This functional profile—commonly called “pseudorestriction”—is typically observed in bronchiolar diseases, in which decreased pulmonary compliance due to cellular infiltration of the lung decreases dynamic volumes, while air trapping increases RV leading to globally normal static volumes [21]. Decreased FEV₁ and FVC were unlikely due to airway obstruction, as PC2 did not correlate with FEV₁/FVC. Dynamic volumes were inversely correlated with PaCO₂ reflecting alveolar hypoventilation during exercise. They were also inversely correlated with V_D/V_T. Although we can not rule out the participation of a vascular dysfunction, V_D/V_T was correlated with “hypoventilation-related” (PC2) variables and not with “vascular-related” variables such as hyperventilation. Furthermore, the absence of severe clinical and functional impairment, the normality of cardiovascular adaptation during exercise and the absence of correlation of systolic pulmonary artery hypertension with V_D/V_T and V'O₂ peak [22] on available echocardiography make the hypothesis of a relationship between pulmonary hypertension and aerobic capacity impairment unlikely [9].

Finally, our study showed that hyperpnea (PC3) independently limited exercise. Increased ventilation was not associated with hypocapnia and thus was appropriate to metabolic requests.

With the exception of specific treatment of pulmonary hypertension, when appropriate, [9,23] no drug treatment has been shown to improve exercise capacity in PLCH. The impact of smoking cessation, systemic steroids or cladribine on exercise capacity has not been evaluated. Pulmonary rehabilitation, which has been shown to improve exercise capacity in several interstitial lung diseases, has not been specifically evaluated in PLCH. Based on the results of the present study, the benefit of pulmonary symptomatic treatments on exercise capacity can be discussed. In non cystic diffuse interstitial lung disease, oxygen supplementation has been showed to improve V'O₂ peak [24] and the six-minute walking test distance [25] in patients with gas exchange impairment. However, the benefit of portable oxygen therapy on physical activities of daily living remains unknown. Bronchodilators decrease dynamic hyperinflation and improve exercise capacity in chronic obstructive pulmonary disease, even when airway obstruction is mild [26]. However, the effect of bronchodilators in PLCH, in which the mechanisms of airway obstruction are partly different from those involved in chronic obstructive pulmonary disease, has not been evaluated.

Strengths and limitations

This study provides new data on exercise capacity in the largest population of PLCH patients published to date. The characteristics of the study population were similar to those reported in the literature, with an equivalent proportion of men and woman, a mean age in the range 20–40 years, predominantly Caucasian ethnic origin and a history of smoking in more than 90% of patients [3,6,7]. Biases related to the retrospective design of the study were limited by standardised recording of objective measurements and by systematic review of all CPET based on international guidelines [10] by a single investigator. However, 17 subjects showing more severe gas exchange impairment could not be included in the multivariate analysis. Thus, we must keep in mind that our results concern mild to moderate stages of PLCH. CT scan were available in only half of the population preventing from inclusion of CT scan changes in the multivariate analysis. This is due to the fact that we only collected CT scans performed close in time to PFT and CPET in the aim to have relevant relationships between CT scan features and functional data. Indeed, PLCH radiological lesions progress over time [27]. For similar reasons, sPAP could not be included in the multivariate analysis. However, among the 21 patients having an echocardiography no correlation was observed between sPAP and V'O₂ peak. If we

extrapolate these results to the whole population, which seems to have similar sociodemographic, PFT and CPET characteristics (S2 Table), sPAP should not be integrated in the multivariate analysis.

Conclusion

In conclusion, exercise limitation is frequent in PLCH and is mainly caused by multifactorial pulmonary changes including alteration of gas exchange, air trapping, “pseudorestriction” and hyperpnea but does not appear to be associated with increased dyspnea. This suggests that treating the lung represents an approach for improving exercise, even in patients experiencing mild dyspnea. In particular, our results suggest assessing the effect of therapies targeting air trapping such as bronchodilators on aerobic capacity.

Supporting Information

S1 Table. Comparison of sociodemographic, PFT and CPET data between subjects with and without missing data.

(DOCX)

S2 Table. Comparison of sociodemographic, PFT and CPET data between subjects with and without echocardiography.

(DOCX)

S3 Table. Data set as a Supporting Information file.

(XLSX)

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References

1. Favara BE, Feller AC, Pauli M, Jaffe ES, Weiss LM, Arico M et al. Contemporary classification of 2histiocytic disorders. The WHO Committee On Histiocytic/Reticulum Cell Proliferations. Reclassification Working Group of the Histiocyte Society. *Med Pediatr Oncol*. 1997; 29(3):157–66. PMID: [9212839](#)
2. Roden AC, Hu X, Kip S, Parrilla Castellar ER, Rumilla KM, Vrana JA, et al. BRAF V600E expression in Langerhans cell histiocytosis: clinical and immunohistochemical study on 25 pulmonary and 54 extra-pulmonary cases. *Am J Surg Pathol*. 2014; 38(4):548–51. doi: [10.1097/PAS.000000000000129](#) PMID: [24625419](#)
3. Elia D, Torre O, Cassandro R, Caminati A, Harari S. Pulmonary Langerhans cell histiocytosis: a comprehensive analysis of 40 patients and literature review. *Eur J Intern Med*. 2015; 26(5):351–6. doi: [10.1016/j.ejim.2015.04.001](#) PMID: [25899682](#)
4. Raouf S, Bondalapati P, Vydyula R, Ryu J, Gupta N, Raouf S, et al. Cystic Lung Diseases: Algorithmic Approach. *Chest*. In press.
5. Delobbe A, Durieu J, Duhamel A, Wallaert B. Determinants of survival in pulmonary Langerhans' cell granulomatosis (histiocytosis X). Groupe d'Etude en Pathologie Interstitielle de la Société de Pathologie Thoracique du Nord. *Eur Respir J*. 1996; 9(10):2002–6. PMID: [8902457](#)
6. Vassallo R, Ryu JH, Schroeder DR, Decker PA, Limper AH. Clinical outcomes of pulmonary Langerhans'-cell histiocytosis in adults. *N Engl J Med*. 2002; 346(7):484–90. doi: [10.1056/NEJMoa012087](#) PMID: [11844849](#)
7. Tazi A, de Margerie C, Naccache JM, Fry S, Dominique S, Jouneau S, et al. The natural history of adult pulmonary Langerhans cell histiocytosis: a prospective multicentre study. *Orphanet J Rare Dis*. 2015; 10:30. doi: [10.1186/s13023-015-0249-2](#) PMID: [25887097](#)
8. Crausman RS, Jennings CA, Tuder RM, Ackerson LM, Irvin CG, King TE Jr. Pulmonary histiocytosis X: pulmonary function and exercise pathophysiology. *Am J Respir Crit Care Med*. 1996; 153(1):426–35. doi: [10.1164/ajrccm.153.1.8542154](#) PMID: [8542154](#)
9. Le Pavec J, Lorillon G, Jaïs X, Tcherakian C, Feuillet S, Dorfmueller P, et al. Pulmonary Langerhans cell histiocytosis-associated pulmonary hypertension: clinical characteristics and impact of pulmonary arterial hypertension therapies. *Chest*. 2012; 142(5):1150–7. doi: [10.1378/chest.11-2490](#) PMID: [22459770](#)
10. Ross RM. ATS/ACCP statement on cardiopulmonary exercise testing. *Am J Respir Crit Care Med*. 2003; 167(10): 211–77.
11. Wallaert B, Talleu C, Wemeau-Stervinou L, Duhamel A, Robin S, Aguilaniu B. Reduction of maximal oxygen uptake in sarcoidosis: relationship with disease severity. *Respiration*. 2011; 82(6):501–8. doi: [10.1159/000330050](#) PMID: [21934275](#)
12. Tazi A, Marc K, Dominique S, de Bazelaire C, Crestani B, Chinnet T, et al. Serial computed tomography and lung function testing in pulmonary Langerhans' cell histiocytosis. *Eur Respir J*. 2012; 40(4):905–12. doi: [10.1183/09031936.00210711](#) PMID: [22441752](#)
13. Macintyre N, Crapo RO, Viegi G, Johnson DC, van der Grinten CP, Brusasco V, et al. Standardisation of the single-breath determination of carbon monoxide uptake in the lung. *Eur Respir J*. 2005; 26(4):720–35. doi: [10.1183/09031936.05.00034905](#) PMID: [16204605](#)
14. Stocks J, Quanjer PH. Reference values for residual volume, functional residual capacity and total lung capacity. ATS Workshop on Lung Volume Measurements. Official Statement of The European Respiratory Society. *Eur Respir J*. 1995; 8(3):492–506. PMID: [7789503](#)
15. Pastré J, Prévotat A, Tardif C, Langlois C, Duhamel A, Wallaert B. Determinants of exercise capacity in cystic fibrosis patients with mild-to-moderate lung disease. *BMC Pulm Med*. 2014; 14:74. doi: [10.1186/1471-2466-14-74](#) PMID: [24884656](#)
16. Canuet M, Kessler R, Jeung M-Y, Métivier A-C, Chaouat A, Weitzenblum E. Correlation between High-Resolution Computed Tomography Findings and Lung Function in Pulmonary Langerhans Cell Histiocytosis. *Respiration*. 2007; 74(6):640–6. doi: [10.1159/000106843](#) PMID: [17675829](#)
17. Laveneziana P, Parker CM, O'Donnell DE. Ventilatory constraints and dyspnea during exercise in chronic obstructive pulmonary disease. *Appl Physiol Nutr Metab*. 2007; 32(6):1225–38. doi: [10.1139/H07-119](#) PMID: [18059601](#)
18. Laveneziana P, Webb KA, Ora J, Wadell K, O'Donnell DE. Evolution of dyspnea during exercise in chronic obstructive pulmonary disease: impact of critical volume constraints. *Am J Respir Crit Care Med*. 2011; 184(12):1367–73. doi: [10.1164/rccm.201106-1128OC](#) PMID: [21885624](#)

19. Ofir D, Laveneziana P, Webb KA, Lam Y-M, O'Donnell DE. Mechanisms of dyspnea during cycle exercise in symptomatic patients with GOLD stage I chronic obstructive pulmonary disease. *Am J Respir Crit Care Med*. 2008; 177(6):622–9. doi: [10.1164/rccm.200707-1064OC](https://doi.org/10.1164/rccm.200707-1064OC) PMID: [18006885](https://pubmed.ncbi.nlm.nih.gov/18006885/)
20. Hannink J, Lahaije A, Bischoff E, van Helvoort H, Dekhuijzen R, Schermer T, et al. Dynamic hyperinflation after metronome-paced hyperventilation in COPD—a 2 year follow-up. *Respir Med*. 2010; 104(11):1700–5. doi: [10.1016/j.rmed.2010.04.007](https://doi.org/10.1016/j.rmed.2010.04.007) PMID: [20452759](https://pubmed.ncbi.nlm.nih.gov/20452759/)
21. Obert J, Tazi A. Manifestations pulmonaires de l'histiocytose langerhansienne. *Revue des Maladies Respiratoires*. 2015; 32(8):850–66.
22. Chenivresse C, Rachenne V, Fournier C, Leroy S, Nevière R, Le Tourneau T, et al. Cardiopulmonary exercise testing in exercise-induced pulmonary hypertension. *Rev Mal Respir*. 2006; 23, 141–148. PMID: [16788438](https://pubmed.ncbi.nlm.nih.gov/16788438/)
23. Fukuda Y, Miura S, Fujimi K, Yano M, Nishikawa H, Yanagisawa J, et al. Effects of treatment with a combination of cardiac rehabilitation and bosentan in patients with pulmonary Langerhans cell histiocytosis associated with pulmonary hypertension. *Eur J Prev Cardiol*. 2014; 21(12):1481–3. doi: [10.1177/2047487313497603](https://doi.org/10.1177/2047487313497603) PMID: [23897898](https://pubmed.ncbi.nlm.nih.gov/23897898/)
24. Harris-Eze AO, Sridhar G, Clemens RE, Gallagher CG, Marciniuk DD. Oxygen improves maximal exercise performance in interstitial lung disease. *Am J Respir Crit Care Med*. 1994; 150(6):1616–22.
25. Visca D, Montgomery A, de Lauretis A, Sestini P, Soteriou H, Maher TM, et al. Ambulatory oxygen in interstitial lung disease. *Eur Respir J*. 2011; 38(4):987–90. doi: [10.1183/09031936.00190710](https://doi.org/10.1183/09031936.00190710) PMID: [21965506](https://pubmed.ncbi.nlm.nih.gov/21965506/)
26. Elbehairy AF, Webb KA, Neder JA, Alberto Neder J, O'Donnell DE. Should mild COPD be treated? Evidence for early pharmacological intervention. *Drugs*. 2013; 73(18):1991–2001. doi: [10.1007/s40265-013-0145-9](https://doi.org/10.1007/s40265-013-0145-9) PMID: [24214364](https://pubmed.ncbi.nlm.nih.gov/24214364/)
27. Brauner MW, Grenier P, Tijani K, Battesti JP, Valeyre D. Pulmonary Langerhans cell histiocytosis: evolution of lesions on CT scans. *Radiology* 1997; 204:497–502 doi: [10.1148/radiology.204.2.9240543](https://doi.org/10.1148/radiology.204.2.9240543) PMID: [9240543](https://pubmed.ncbi.nlm.nih.gov/9240543/)