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# 1 **The interplay of stiffness and force anisotropies drive embryo elongation**

2

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15

## 16 **Abstract**

17

18 The morphogenesis of tissues, like the deformation of an object, results from the interplay between  
19 their material properties and the mechanical forces exerted on them. Whereas the importance of  
20 mechanical forces in influencing cell behaviour is widely recognized, the importance of tissue material  
21 properties, in particular stiffness, has received much less attention. Using *C. elegans* as a model, we  
22 examine how both aspects contribute to embryonic elongation. Measuring the opening shape of the  
23 epidermal actin cortex after laser nano-ablation, we assess the spatiotemporal changes of  
24 actomyosin-dependent force and stiffness along the antero-posterior and dorso-ventral axis.

25 Experimental data and analytical modelling show that myosin II-dependent force anisotropy within the  
26 lateral epidermis, and stiffness anisotropy within the fiber-reinforced dorso-ventral epidermis are  
27 critical to drive embryonic elongation. Together, our results establish a quantitative link between  
28 cortical tension, material properties and morphogenesis of an entire embryo.

## 29 Introduction

30

31 Morphogenesis and organ formation rely on force distribution and tissue material properties, which  
32 are often heterogeneous and evolve over time. Forces are generated through a group of relatively  
33 well-conserved molecular motors associated with the cytoskeleton, among which myosin II linked to  
34 actin filaments is the most prevalent during epithelial morphogenesis (Vicente-Manzanares, 2009).  
35 Myosin II spatial distribution and dynamics greatly influence morphogenetic processes (Levayer and  
36 Lecuit, 2012). In particular, the asymmetric distribution of the actomyosin network and its pulsatile  
37 behaviour define the direction of extension during *Drosophila* germband elongation (Bertet, 2004,  
38 Blankenship, 2006), *Drosophila* renal tubule formation (Saxena, 2014) or *Xenopus* mesoderm  
39 convergent extension (Shindo and Wallingford, 2014). Whereas the implication of mechanical forces  
40 has been intensively investigated (Zhang and Labouesse, 2012, Heisenberg and Bellaiche, 2013),  
41 much fewer studies have considered the impact of tissue material properties *in vivo*, except for their  
42 influence on cell behaviour *in vitro* (Kasza, 2007).

43 *C. elegans* embryonic elongation represents an attractive model for studying morphogenesis, as it  
44 offers single cell resolution and powerful genetic analysis. During its elongation, the embryo evolves  
45 from a lima-bean to a typical cylindrical shape with a four-fold increase in length, without cell  
46 migration, cell division, or a notable change in embryonic volume (Sulston, 1983, Priess and Hirsh,  
47 1986) (figure 1a). This process requires the epidermal actomyosin cytoskeleton, which acts mostly in  
48 the lateral epidermis (also called seam cells), while the dorso-ventral (DV) epidermal cells may remain  
49 passive (Appendix 1)(Wissmann, 1997, Wissmann, 1999, Shelton, 1999, Piekny, 2003, Diogon, 2007,  
50 Gally, 2009, Chan, 2015, Vuong-Brender, 2016). Indeed, the non-muscle myosin II is concentrated in  
51 seam cells; in addition short disorganized actin filaments, which favour actomyosin contractility, are  
52 present in seam cells, but not in the DV epidermis where they instead form parallel circumferential  
53 bundles (Figure 1b-d)(Gally, 2009, Priess and Hirsh, 1986). The actomyosin forces are thought to

54 squeeze the embryo circumferentially, to thereby increase the hydrostatic pressure and promote  
55 embryo elongation in the antero-posterior (AP) direction (Priess and Hirsh, 1986) (figure 1e).  
56 Although the published data clearly imply myosin II in driving elongation, they raise a number of  
57 issues. First, myosin II does not show a polarized distribution (figure 1f-g) nor does it display dynamic  
58 pulsatile foci at this stage; hence, it is difficult to account for the circumferential squeezing. Moreover,  
59 force measurements are lacking to establish that the actomyosin network does squeeze the embryo  
60 circumferentially. Second, a mechanical continuum model is needed to understand how the embryo  
61 extends preferentially in the AP direction.

62 To address those issues, we used laser ablation to map the distribution of mechanical stress (i.e the  
63 force per unit area) and assess tissue stiffness (i.e. the extent to which it resists deformation) in the  
64 embryonic epidermis. We then correlated the global embryonic morphological changes with these  
65 physical parameters. Finally, we developed continuum mechanical models to account for the  
66 morphological changes. Altogether, our data and modelling highlight that the distribution of forces in  
67 the seam cells and the stiffness in the DV epidermis must be polarized along the circumferential axis  
68 (or DV axis) to drive elongation.

69

70

## 71 **Results**

72

### 73 **Measuring the mechanical stress on the actin cortex through laser ablation**

74 To measure the stress distribution on the actin cortex, we used laser nano-ablation, which has now  
75 become a standard method to assess forces exerted in cells, to sever the actin cytoskeleton and  
76 observe the shape of the opening hole (figure 2a). We visualized actin with a GFP- or mCherry-  
77 labelled actin-binding-domain protein (ABD) expressed in the epidermis (Gally, 2009) (figure 1b-d).

78 We adjusted the region of interest to cut within one cell, restricting our analysis to the early phase of  
79 elongation ( $\leq 1.7F$ ; for staging, see figure 1 legend).

80 We observed two types of ablation responses (see Methods). In the first (accounting for > 80% of the  
81 cases), the opening hole within the actin cytoskeleton reached equilibrium in less than 10 s, and  
82 resealed within less than 2 min (Figure 2 -Figure supplement 1, supplementary movie 1). In such  
83 embryos, actin occasionally accumulated around the cut borders but not around cell borders  
84 (supplementary movie 1). Imaging calcium levels, which can rise after laser wounding (Xu and  
85 Chisholm, 2011, Razzell, 2013, Antunes, 2013), showed either no change or a localized increase  
86 (supplementary movie 2, Figure 2 – Figure supplement 2a-b). In the second, an actin ring  
87 accumulated around the cell borders during the repair process (supplementary movie 3) and a  
88 calcium wave propagated to nearby epidermal cells (supplementary movie 4; Figure 2 –Figure  
89 supplement 2c). Whereas embryos showing the first response continued to develop and hatched,  
90 those showing the second response arrested their development and eventually died. In all  
91 subsequent studies, we only took into account the first type of response, which should correspond to  
92 a local cortex disruption.

93 To compare the response between different conditions, we detected the cut opening shape, which we  
94 fitted with an ellipse to derive the shape parameters (see Methods). The laser setup we used did not  
95 enable us to image the recoil dynamics within the first second after the cut, which other investigators  
96 previously used to assess the extent of mechanical stress (Rauzi and Lenne, 2015, Smutny, 2015,  
97 Saha, 2016). To circumvent this issue, we developed a novel analysis method to derive mechanical  
98 stress, based on the equilibrium shape of a thin cut in an infinite elastic isotropic plane, subjected to  
99 biaxial loading (stress applied in two perpendicular directions)(Theocaris, 1986). The rationale for  
100 approximating the epidermis to such a plane is further outlined in the Appendix 2-3. In these  
101 conditions, a thin cut will open to form an elliptical hole at equilibrium (figure 2a). The opening of the  
102 cut reflects mechanical stress in the direction perpendicular to the cut direction.

103 We cut the epidermal actin specifically in the AP and DV directions, which we found to correspond to  
104 the stress loading directions (figure 2b; Appendix 2). For a cut in the AP direction, the minor axis of  
105 the ellipse at equilibrium,  $b_{DV}$ , will be proportional to the cut length,  $l$ , and to the ratio of stress in the  
106 DV direction,  $\sigma_{DV}$ , over the Young modulus  $E$  of the plane (Theocaris, 1986) (figure 2b):

$$107 \quad b_{DV} = 2 \frac{\sigma_{DV}}{E} l \quad (1)$$

108 We will call the ratio  $\frac{b_{DV}}{l}$  the opening in the DV direction of a cut made along the AP direction (figure  
109 2b), and similarly  $\frac{b_{AP}}{l}$  the opening in the AP direction of a cut made in the DV direction. Thus, we  
110 used the opening of the hole in a given direction to derive the stress in that direction.

111 We compared the conclusions drawn from this method with methods relying on the recoil dynamics  
112 (Rauzi and Lenne, 2015, Smutny, 2015, Saha, 2016) (Appendix 3). The half-time of the cut border  
113 relaxation, which depends on the ratio of viscosity over stiffness, was similar in the AP and DV  
114 directions (Appendix 3), supporting the hypothesis that the seam cell cortex is isotropic. We found an  
115 agreement between both methods for the AP versus DV stress ratio, and similar trends for the stress  
116 magnitude.

117 To further examine the validity of this method, we did two tests. First, the aforementioned theory  
118 (Theocaris, 1986) predicts that the minor to major axis ratio of the opening ellipse is independent of  
119 the initial cut length. We found that it is the case when the cut length varied from 3  $\mu\text{m}$  to 6  $\mu\text{m}$   
120 (Appendix 4). Second, to prove that the opening observed after laser cutting depends on myosin II  
121 activity, we performed cuts in embryos defective for the main myosin II regulator, LET-502/Rho-kinase  
122 (Gally, 2009). As shown in figure 2c, the opening in the seam cell H1 at the 1.5F stage in *let-*  
123 *502(sb118ts)* embryos changed very little and was significantly smaller than in WT embryos,  
124 consistent with a decrease of mechanical stress.

125 Thus, we feel confident that the method based on the opening shape measures actomyosin-  
126 dependent stress and can reliably report on the differences of stress along the DV and AP directions.

127

## 128 **Stress anisotropy in seam cells correlates with embryonic morphological changes**

129 We applied the method described above on three seam cells (head H1, body V3, tail V6; figure 3a),  
130 since myosin II acts mainly in seam cells (Gally, 2009), and compared the response with embryonic  
131 morphological changes. We focused on the anisotropy of stress between the DV and AP directions  
132 (difference of stress along both directions) in a given cell (figure 3b). Indeed, in other systems, such  
133 as *Drosophila* embryos (Rauzi, 2008) and *C. elegans* zygotes (Mayer, 2010), this parameter is critical.  
134 At the 1.3F stage in H1, there was no significant stress anisotropy; however, as the embryo elongated  
135 to the 1.5F and 1.7F stages, the stress became anisotropic (figure 3b). In V3, the anisotropy of stress  
136 evolved in the opposite direction, with higher stress anisotropy at the 1.3F compared to the 1.5F stage  
137 (figure 3b). In V6, the stress was slightly anisotropic at both the 1.3F and 1.5F stages (figure 3b). In all  
138 cells, whenever the stress became anisotropic it was higher in the DV direction. Overall, the opening  
139 increased as the embryo elongated from the 1.3F to the 1.5F stage and from the 1.5F to the 1.7F  
140 stage for H1.

141 To correlate the stress anisotropy with the morphological changes of the embryo, we used markers  
142 labelling cortical actin (an ABD) and junctions (HMR-1/E-cadherin). We observed that the head, body  
143 and tail diameter (at the level of H1, V3 and V6, respectively) decreased at different rates over time  
144 (figure 3c), as also observed by Martin and colleagues (Martin, 2014). The head diameter did not  
145 diminish between the 1.3F and 1.5F stages when the stress was nearly isotropic, but decreased  
146 significantly between the 1.5F and 1.7F stages as the stress anisotropy increased. Conversely, the  
147 body diameter decreased the fastest between the 1.3F and 1.5F stages when the stress was highly  
148 anisotropic, then changed at a lower pace beyond the 1.5F stage when the stress became less  
149 anisotropic. Finally, the tail diameter decreased nearly linearly between the 1.3F and 1.7F stages, at a  
150 lower rate than the body diameter, coinciding with a smaller anisotropic stress in V6. Thus, the local  
151 morphological changes within the embryo correlate with locally higher stress in the DV compared to  
152 the AP direction.



153 To define whether all cells equally contribute to the diameter change, we quantified the circumferential  
154 width of the epidermal cells H1, V3 and their adjacent DV cells (figure 3d-e). At the level of V3, the  
155 decrease in body diameter came from both seam (V3) and DV cells, whereas in the head it came  
156 mainly from DV cells (figure 3e). Collectively, our results strongly suggest that the stress anisotropy  
157 correlates with morphological changes. Furthermore, we found that both seam and DV epidermal  
158 cells contribute to the changes in embryo diameter, irrespective of their level of active myosin II.

159

### 160 **The establishment of stress anisotropy depends in part on the spectrin cytoskeleton**

161 Taking the H1 cell as an example, we considered some cellular factors that could contribute to the  
162 stress anisotropy in seam cells: (i) actin-anchoring proteins, (ii) muscle-induced tension. To ease  
163 comparisons, we defined the anisotropy of stress (AS) as

$$164 \quad AS = \frac{DV \text{ stress}}{AP \text{ stress}} = \frac{\sigma_{DV}}{\sigma_{AP}} \quad (2)$$

165 which can be derived from the ratio of the opening along the DV and AP directions, see equation (1).

166 First, we examined the actin-anchoring spectrin cytoskeleton, which is essential for embryonic  
167 elongation (Moorthy, 2000, Norman and Moerman, 2002, Praitis, 2005). In *spc-1(RNAi)* embryos, at a  
168 developmental timing equivalent to the 1.7F stage in control embryos, we found a smaller opening in  
169 both AP and DV directions and a decrease of AS compared to WT (figure 3f-g). This may account for  
170 the slower elongation rate of *spc-1(RNAi)* embryos and their arrest at the 2F stage (Figure 3 –Figure  
171 supplement 1). Thus, spectrin partially contributes to the AS at the 1.7F stage.

172 Second, we wondered whether muscle contractions, which start after the 1.5F stage, could account  
173 for AS changes (figure 3b). Compared to controls, embryos depleted of UNC-112/Kindlin, which  
174 mediates sarcomere assembly (Rogalski, 2000), showed a significantly higher opening in both the AP  
175 and DV directions at the 1.7F stage, but no change in stress anisotropy (figure 3f-g). This is  
176 consistent with their wild-type elongation rate up to the 2F stage (Figure 3 –Figure supplement 1).  
177 Thus, AS establishment in the H1 cell after the 1.5F stage is independent of muscle contractions.

**179 A mechanical model for seam cell elongation depending on stress anisotropy**

180 To define the possible causal relationship between the AS and embryonic shape changes, we aimed  
181 at simplifying the shape of the embryo to apply classical physical laws such as the Young-Laplace  
182 equation, which predicts the relationship between surface tension and the surface curvature. As  
183 illustrated in figure 1, the embryo has a circular section and a cylindrical or conical shape depending  
184 on the stage, in which the epidermis is thin (100 nm to 2  $\mu\text{m}$ , depending on areas;  
185 [www.wormatlas.org](http://www.wormatlas.org)) compared to the embryo diameter (25  $\mu\text{m}$ ). Within the embryo, the epidermis is  
186 subjected to hydrostatic pressure when the section decreases (Priess and Hirsh, 1986). We can thus  
187 model the *C. elegans* embryo as an isotropic thin-wall (the epidermis) vessel with capped ends under  
188 hydrostatic pressure, and determine the relationship between the mechanical stress on the epidermis  
189 and the embryo shape.

190 First, we calculated the anisotropy of stress on the wall of such a vessel. For an axisymmetric vessel,  
191 the AS on the wall depends on the surface curvature and the radius (Appendix 5), which for simple  
192 geometrical configurations can be written as shown in figures 4a-c. Typically, the AS factor, or the DV  
193 to AP stress ratio, is equal to 1 for a sphere, equal to 2 for a cylinder and takes an intermediate value  
194 between 1 and 2 for an ellipsoid. We can simplify the geometry of *C. elegans* embryos as a curved  
195 cylinder (body), attached to a sphere (head) between the 1.3F and 1.5F stages (figure 4d-e). The  
196 head evolves into an ellipsoid between the 1.5F and 1.7F stages (figure 4f). Thus, the AS of the head  
197 can be determined easily. We previously observed that the AP stress among the seam cells at a  
198 given stage differs by 20% (figure 3b). Thus, if we approximate the AP stress as a constant at a given  
199 stage, the AS in the body will depend on the ratio of the body to head radius (figure 4d-e, Appendix  
200 5). Given the head and body diameter of the embryo (figure 3c), we can compare the AS predicted by  
201 the thin-wall vessel model with those derived experimentally with laser ablation (figure 4g). Both  
202 values are nearly identical, showing that the AS can be predicted based on the embryonic geometry.

203 To examine whether the AS can dictate embryonic morphological changes, we related the  
204 deformation of the vessel wall with the forces applied using the Hooke's law (figure 4h, Appendix 6A)  
205 – for instance Hooke's law states that the one-dimensional deformation of a spring equals to the ratio  
206 of the applied force to the spring stiffness. Similarly in a two-dimensional system and for an isotropic  
207 material, the deformation is proportional to the mechanical stress (forces) and inversely proportional  
208 to the Young modulus (stiffness) along the different loading directions (figure 4h). The resulting  
209 equations, which assume that seam cells have an isotropic cortex and are subjected to contractile  
210 stress, correctly predict that the seam cell dimension increases along the AP axis ( $\epsilon_{AP}$ ) with the AS  
211 (figure 4h-j), and decreases along the DV axis ( $\epsilon_{DV}$ ). Indeed, consistent with the equations, the head  
212 evolves from a sphere to an ellipsoid between the 1.5F to the 1.7F stages as the AS becomes greater  
213 than 1 (figure 3b-c).

214 In conclusion, our experimental and modelling data show that the AS induces morphological changes  
215 occurring in embryonic seam cells and provide a basis to understand how the embryo elongates from  
216 a mechanical standpoint.

217

### 218 **Stiffness anisotropy-based elongation of the DV epidermis**

219 As shown in figure 3e, the head diameter reduction primarily involves changes of the circumferential  
220 width in the DV epidermis. Since the RhoGAP RGA-2 maintains myosin II activation in these cells at a  
221 low level (Diogon, 2007), actomyosin contractility in DV cells cannot account for such changes.

222 However, DV epidermal cells have circumferentially-oriented actin bundles, in contrast to seam cells  
223 (figure 1b-d), which based on recent observation could affect cell stiffness (Calzado-Martin, 2016,  
224 Salker, 2016). We thus hypothesized that the circumferential polarized actin distribution in DV  
225 epidermal cells could induce higher stiffness in that direction and thereby influences their deformation.

226 To establish whether it is the case, we investigated both stress and stiffness distribution in the  
227 epidermal cells dorsal and ventral to the H1 seam cell using laser nano-ablation (figure 5a). Since  
228 these cells are the precursors of the HYP7 syncytium, we will denote them HYP7 henceforth.

229 We found that, in the HYP7 cell, the opening in the DV direction was larger than in the AP direction  
230 (figure 5b; dorsal and ventral cells behaved similarly after laser cutting), at the 1.5F and 1.7F stages,  
231 similarly to the H1 cell (figure 3b). However, the ratio of DV/AP opening in HYP7 was more important  
232 than in H1 (figure 5c). Assuming that the HYP7 cell cortex has isotropic material properties like that of  
233 H1, our model (figure 4; Appendix 5) would predict that the DV/AP opening ratio in HYP7 depends  
234 only on the head axisymmetric shape and is equal to that of H1, and would thus contradict our  
235 observations. Hence, this *reductio ad absurdum* argument suggests that the HYP7 cell has  
236 anisotropic cortical material properties indeed.

237 To model the DV epidermal cell deformation, we examined two classes of anisotropic stiffness  
238 material previously described: orthotropic materials such as bones (Miller, 2002, Helwig, 2009), and  
239 fiber-reinforced materials such as arteries (Gasser, 2006), articular cartilage (Federico and Gasser,  
240 2010) or fibrous connective tissues (Ben Amar, 2015). Orthotropic materials have different stiffnesses  
241 along orthogonal directions, and thus respond differently to the same stress magnitude along these  
242 directions. Fiber-reinforced materials also have different stiffnesses in the directions along and  
243 transverse to the fibers; in addition, such materials can respond differently to extensive or  
244 compressive stress (Bert, 1977). To define which model best applies to the DV epidermis, we used  
245 continuum linear elastic analysis (Muskhelishvili, 1975, Suo, 1990, Theocaris, 1986, Yoffe, 1951)  
246 (Appendix 7) to interpret the laser cutting data on the DV epidermis. We discarded the orthotropic  
247 model, as it did not adequately describe our data (Appendix 8), and focused on the fiber-reinforced  
248 plane model, which better accounts for the presence of well aligned actin fibers in DV cells.

249 In a fiber-reinforced material composed of a matrix superimposed with fibers, the contribution of the  
250 fibers to the stiffness of the material depends on their orientation. In the direction parallel to the fibers,  
251 the Young modulus is much increased, whereas in the direction perpendicular to the fibers this  
252 contribution is small. According to our modelling, the Young modulus along the fiber direction,  
253 increases linearly with a factor  $K$  related to the fiber stiffness and density; whereas the stiffness along  
254 the direction transverse to the fibers varies as a hyperbolic function of  $K$  and reaches a plateau  
255 (Appendix 7). For fiber-reinforcement in the DV direction, the change in Young modulus along the DV

256 and AP directions predicted by modelling is given in figure 6ab. Cuts perpendicular to the fibers  
257 opened similarly to an isotropic material with the matrix Young modulus, because they locally  
258 destroyed the fibers (figure 6c; see equation 1 above). By contrast, cuts along the fibers opened with  
259 an equilibrium value that depends on the fiber stiffness and distribution through the factor  $K$  defined  
260 above (figure 6d, Appendix 7).

261 Since the H1 seam and the head HYP7 cells are adjacent along the circumference (figure 3d), they  
262 should be under the same DV stress due to tension continuity across cell-cell junctions. According to  
263 equation (1), if the stress in two cells is the same their opening should vary inversely with their  
264 respective Young moduli. Since the DV opening of HYP7 was about 1.5 times smaller than that of H1  
265 (figure 7a), we infer that the Young modulus of the HYP7 matrix without fibers was about 1.5 times  
266 stiffer than that of H1 (Appendix 9), suggesting that these cells have distinct material properties.  
267 Comparing the DV and AP opening for the HYP7 cell, we found that the factor  $K$  increased during  
268 early elongation (figure 7b; Appendix 10). More importantly, the calculated ratio of DV/AP Young  
269 moduli also increased, and was greater than the DV/AP stress anisotropy (figure 7c).

270 To understand how a change in stiffness affects head HYP7 deformation, we wrote again Hooke's law  
271 for these cells (Appendix 6B; figure 7d). Since myosin II activity in DV cells is low, their cortex should  
272 be exposed to tensile stress induced by actomyosin contractility in the seam cells. The cell length  
273 along the AP direction increased when the stiffness anisotropy (DV/AP stiffness ratio) increased  
274 (figure 7d-e), whereas the trend was opposite in the DV direction (figure 7d,f). Thus the stiffness  
275 anisotropy helps the HYP7 cell extend along the AP direction and shrink along the DV direction.  
276 Interestingly, the equations predict that increasing stress anisotropy has an opposite effect on HYP7  
277 cell deformation, since it prevents them from extending antero-posteriorly (figure 7e). Altogether, our  
278 model strongly suggests that when the DV/AP stiffness anisotropy increases and is higher than the  
279 DV/AP stress anisotropy, as observed in the head HYP7 (figure 7c), elongation along the AP direction  
280 is favoured. Furthermore, our data highlight that the distinct mechanical properties of cells composing  
281 a complex tissue enables its morphogenesis and does not require all cells to be contractile.

## 282 **The stress and stiffness anisotropies correlate with actin arrangement**

283 Since myosin II is not polarized (figure 1e-f), to find out if actin distribution accounts for the stress and  
284 stiffness anisotropies, we carried out an analysis of actin filament alignment in the seam cells H1 and  
285 V3, as well as in the head HYP7 cell. We found that the polarization of actin filaments in seam cells  
286 correlated with the observed pattern of stress anisotropy. Indeed, in H1 at the 1.3F stage, actin  
287 filaments had a nearly isotropic angular distribution correlating with the isotropic stress (figure 8a,  
288 figure 3b), whereas they became increasingly aligned along the DV direction from the 1.3F to 1.7F  
289 stages (figure 8a) which mirrors the increase of stress anisotropy (figure 3b). Likewise, actin  
290 alignment decreased along the DV direction in V3 from the 1.3F to the 1.5F stage, in parallel to the  
291 decrease of stress anisotropy between those stages (figures 8b and 3b). The changes in H1 (figure  
292 8c) were statistically significant, whereas it is not the case in V3 (figure 8d). In the HYP7 cell, actin  
293 filaments already acquired a preferential DV alignment at the 1.3F stage (figure 8c), but became  
294 increasingly organized along the DV direction as the embryo elongated to the 2F stage, with a highly  
295 significant difference between the 1.5F and 1.7F stages (figure 8c and 8f). These changes correlated  
296 with the increase of stiffness anisotropy observed in the HYP7 cell (figure 7c).

297 We have attempted to functionally test how actin organization could affect stress and stiffness by  
298 manipulating actin polymerization through two different strategies to express cofilin during early  
299 elongation. However, we could not obtain meaningful results. Altogether, we conclude that the pattern  
300 of actin distribution showed a good correlation with the observed stress and stiffness anisotropy. It will  
301 remain important to define the mechanisms bringing changes in actin distribution, and ultimately  
302 whether it is a cause or a consequence of anisotropy.

303

304

305

## 306 **Discussion**

307 Classical experiments in embryology have outlined how the juxtaposition of cells with different  
308 properties is critical to power important morphogenetic movements, such as *Xenopus* gastrulation  
309 (Hardin and Keller, 1988, Keller and Winklbauer, 1992). In this work, we have dissected the  
310 mechanical contributions of the different epidermal cells driving *C. elegans* embryonic morphogenesis  
311 at the single cell resolution, highlighting the importance of juxtaposition of cells with different  
312 properties. Combining laser nano-ablation and continuum mechanics modelling, we first highlight the  
313 importance of stress anisotropy in the seam cells. Second, we emphasize that stiffness anisotropy is  
314 equally important for embryonic elongation but matters in another epidermal cell type, the DV cells.  
315 Thereby, we reveal the critical role of tissue material properties in morphogenesis.

316 Many studies analysing morphogenetic processes have focused on 2D epithelial sheets such as the  
317 *Drosophila* mesoderm (Martin, 2009), germband (Rauzi, 2008, Rauzi, 2010, Blankenship, 2006,  
318 Fernandez-Gonzalez and Zallen, 2011), amnioserosa (Solon, 2009, Gorfinkiel, 2009), wing and  
319 thorax (Aigouy, 2010, Bosveld, 2012), or the zebrafish enveloping cell layer (Behrndt, 2012) during  
320 embryonic development. They have revealed the role of contractile actomyosin pulses and planar  
321 polarity in coordinating events over long distances. The *C. elegans* embryonic elongation is distinct  
322 from those situations since it does not involve myosin II polarized distribution nor actomyosin pulses.  
323 Interestingly, this process still requires stress anisotropy, outlining that it can be reached by different  
324 means. We suggest that several factors contribute to establish stress anisotropy in *C. elegans*. First,  
325 the actin network displayed a more polarized dorso-ventral distribution in seam cells when the stress  
326 anisotropy was higher, which should increase the stress in that direction. Second, akin to a planar  
327 polarized distribution, myosin II activity displays an asymmetric distribution along the embryo  
328 circumference in cells with different material properties (figure 8g). Intriguingly, tissue culture cells can  
329 sense the spatial stiffness distribution (Walcott and Sun, 2010, Fouchard, 2011, Trichet, 2012, Lange  
330 and Fabry, 2013), raising the possibility that seam cells sense the higher circumferential stiffness and  
331 respond with higher DV-oriented stress through the mechanosensitive adherens junctions (le Duc,  
332 2010, Yonemura, 2010). Third, we found that the spectrin cytoskeleton matters to reach normal levels  
333 of stress magnitude and anisotropy. Spectrin is known to impinge on actin filament alignment and

334 continuity in DV cells (Praitis, 2005, Norman and Moerman, 2002) and could thus affect DV stiffness  
335 anisotropy by reducing the level of actin fiber alignment. Finally, although myosin II activity is low in  
336 DV cells (Diogon, 2007), the remaining activity might create some DV-oriented stress feeding back on  
337 seam cells.

338 By modelling the DV cells as a fiber-reinforced material, we reveal how the polarized cytoskeleton in  
339 DV cells increases their stiffness to orient the extension in the AP direction, acting like a ‘molecular  
340 corset’. Related ‘molecular corsets’ have been described and proposed to drive axis elongation in  
341 other systems (Wainwright, 1988). In *Drosophila*, a network of extracellular matrix fibrils was  
342 proposed to help elongate developing eggs (Haigo and Bilder, 2011). In plant cells, the orientation of  
343 cellulose microfibrils determines the axis of maximal expansion. In the latter, stiffness anisotropy also  
344 helps overcome stress anisotropy (Green, 1962, Baskin, 2005). Importantly, *C. elegans* embryos  
345 reduce their circumference during elongation, while *Drosophila* eggs and plants increase it. It  
346 suggests that to conserve the actin reinforcement properties when the diameter decreases, *C.*  
347 *elegans* DV epidermal cells should have a mechanism to actively shorten the actin bundles, as  
348 observed in a biomimetic *in vitro* system (Murrell and Gardel, 2012).

349 Our experimental data were consistent with the predictions from Hooke’s law. They prove that the  
350 actomyosin cortex preferentially squeezes the embryo circumferentially, and that the stress anisotropy  
351 is tightly linked to the geometry of the embryo. By quantitatively assessing the contribution of stiffness  
352 anisotropy in tissue elongation, we have emphasized its importance relative to the more established  
353 role of stress anisotropy. The precise relationship between both anisotropies remains to be  
354 investigated. Thus, the juxtaposition of cells with different “physical phenotypes”, seam epidermis  
355 expressing stress anisotropy and DV epidermal cell showing stiffness anisotropy, powers *C. elegans*  
356 elongation, as previously suggested in chicken limb bud outgrowth (Damon, 2008) or chick intestinal  
357 looping (Savin, 2011). We did not mention other potential stress bearing components, like  
358 microtubules and the embryonic sheath (Priess and Hirsh, 1986), since the former mainly serves to  
359 enable protein transport (Quintin, 2016) whereas the function of the later will be the focus of an  
360 upcoming work.



361 In conclusion, our work highlights that tissue elongation relies on two fundamental physical quantities  
362 (mechanical stress and tissue stiffness), and provide the most advanced mesoscopic understanding  
363 to date of the mechanics at work during the first steps of *C. elegans* embryonic elongation.

364

## 365 **Methods**

366

### 367 ***C. elegans* alleles and strains**

368 Bristol N2 was used as the wild-type (WT) strain and animals were maintained as described in  
369 Brenner (Brenner, 1974). The strain ML1540: *mcls50[lin-26p::vab-10(abd)::gfp; myo-2p::gfp]* LGI  
370 carrying the actin-binding domain (ABD) of the protein VAB-10 under the epidermal promoter *lin-26*  
371 was described elsewhere (Gally, 2009). The endogenous NMY-1::GFP reporter strain was built by  
372 CRISPR knock-in (ML2540: *nmy-1(mc82)[nmy-1::gfp]* LGX; the NMY-2::GFP reporter strain LP162  
373 *nmy-2(cp13)[nmy-2::gfp + LoxP]* LGI was a generous gift from Daniel Dickinson.

374 For parallel calcium and actin imaging during ablation, we used the strain ML2142: *mcls43 [lin26p::*  
375 *vab-10::mCherry; myo-2p::gfp]; juls307[dpy-7p::GCaMP3]* carrying a calcium sensor under the  
376 epidermal promoter *dpy-7p* and mCherry-labeled VAB-10(ABD) under the *lin-26* promoter. The  
377 thermosensitive Rho kinase mutation *let-502(sb118ts)* was crossed with ML1540 to give the strain  
378 ML2216: *let-502(sb118ts); mcls50[lin-26p::vab-10(abd)::gfp; myo-2p::gfp]* LGI.

379 For determining morphological changes, we used the strain ML2386: *mcls50[lin-26p::vab-*  
380 *10(abd)::gfp; myo-2p::gfp] I; xnls97[hmr-1::gfp] III* expressing both a junctional marker (HMR-1/E-  
381 cadherin) and an actin marker (VAB-10(ABD)).

382 For actin alignment analysis we used the ML1966 *unc-119(ed3) mcls67 [dpy7p::LifeAct::GFP; unc-*  
383 *119(+)]* strain, expressing the actin reporter LIFEACT under the *dpy-7* epidermal promoter.

384

### 385 **RNA interference**

386 RNAi experiments were done using injection of double-stranded RNA synthesized from PCR-  
387 amplified genomic fragments using a T3 or T7 mMESSAGE mMACHINE Kit (Ambion, Austin, TX,  
388 USA). The embryos were analyzed from 24h to 48h post-injection.

389

### 390 **Time-lapse analysis and morphological change quantification**

391 Freshly laid embryos or embryos from dissected hermaphrodites were mounted on 5% agarose pads  
392 in M9 buffer and the coverslip was sealed with paraffin oil. DIC time-lapse movies were recorded at  
393 20°C using a Leica DM6000 upright microscope with a 40X oil immersion objective. For each embryo,  
394 a Z-stack of 7-8 focal planes with 4  $\mu\text{m}$  step size was acquired. The length of embryos was estimated  
395 by tracing the embryo body axis (through the middle of the embryo). Fluorescence time-lapse movies  
396 were recorded at 20°C using a spinning-disk Zeiss microscope Axio Observer.Z1 using a 63X oil  
397 immersion objective. Other fluorescence images were acquired with the same microscope using a  
398 100X oil immersion objective. To determine the morphological changes of the embryo, sections of the  
399 embryo imaged with junctional and actin markers at the level of H1, V3 and V6 were reconstructed to  
400 determine the radius, seam and DV cell width along the circumferential direction. All images were  
401 analysed using the ImageJ (FiJi) software (NIH, Bethesda, Maryland, USA; <http://rsb.info.nih.gov/ij/>)  
402 and MATLAB R2014b (The MathWorks Inc., Natick, MA).

403

### 404 **Actin alignment analysis**

405 Z-stack images of LIFEACT::GFP fluorescence expression in the epidermis were acquired using a  
406 confocal Leica SP5 microscope with a 63X oil immersion objective and zoom factor 8. We used a step  
407 size of 0.08  $\mu\text{m}$ , a pinhole opening of 0.6 Airy Unit and projected 2  $\mu\text{m}$  around the actin cortex. The  
408 embryos were rotated on the scan field to have the same antero-posterior orientation. The acquired  
409 images were deconvoluted using the Huygens Essential software from Scientific Volume Imaging  
410 (Hilversum, Netherlands). We chose a region of interest (ROI) of 4x4  $\mu\text{m}^2$  within the seam cell H1 or  
411 dorso-ventral epidermal cell HYP7, and of 3x3  $\mu\text{m}^2$  within the seam cell V3 to perform Fast Fourier  
412 Transform (FFT). We used a high-pass filter to remove the low frequencies then did inverse FFT. We  
413 found that the high pass filter removed changes in intensity due to unequal labelling or out of focus  
414 signals but retain the actin texture. Finally we use an ImageJ plugin "Spectral Texture Analysis"

415 written by Julien Pontabry to derive the angle distribution of actin texture. This plugin performed the  
416 Fast Fourier Transform (FFT) of the given ROI and computing coefficients in Fourier space, such as  
417 the angle distribution of the given structure, as detailed in Gonzalez, R.C. & Woods, R.E. (2008).  
418 Digital image processing, Nueva Jersey, chapter 11, section 3.3 (Gonzalez, 2008).

### 419 **Embryo staging for ablation**

420 For ablations, we compared embryos of the same developmental timing. To do so, we recorded the  
421 elongation curve of different genetics background (Figure 3 –Figure supplement 1) and took embryos  
422 at the corresponding developmental time from the beginning of elongation. Thus, *unc-112(RNAi)*  
423 embryos elongating up to 1.7F similarly to WT have the same length as WT at 1.7F stage. In contrast,  
424 *spc-1(RNAi)* embryos elongated slower than WT (Figure 3 –Figure supplement 1) and thus at a time  
425 corresponding to 1.7F in a control embryos were shorter than wild-type embryos 1.7F stage. For  
426 comparison between *let-502(sb118ts)* embryos, measurements were carried out at 25.5°C and the  
427 embryos were taken when muscles start to twitch (around 1.5F in control embryos).

428

### 429 **Laser ablation**

430 Laser ablation was performed using a Leica TCS SP8 Confocal Laser Scanning microscope, with a  
431 femtosecond near-infrared Coherent Chameleon Vision II, Ti:Sapphire 680-1080 nm laser, 80 MHz.  
432 To make a line cut, a region of interest with a length varying from 3 to 6  $\mu\text{m}$  and a width of 0.08  $\mu\text{m}$  (1  
433 pixel width) was drawn. We used a laser wavelength varying from 800 nm to 900 nm, which gave  
434 similar ablation responses. The laser power was tuned before each imaging section to obtain local  
435 disruption of the cortex response (>80% of the cases, visible opening, no actin accumulation around  
436 cell borders in the repair process and the ablated embryos developed normally). Typically the power  
437 of the laser was 2000 mW, and we used 50% power at 100% gain. Wounding response (actin  
438 accumulation around cell borders in the repair process, embryo died afterwards) was rarely observed  
439 at the power used for local disruption, but more often when the power was increased to 60-65%. The  
440 first time point was recorded 1.44 s after cutting, which corresponded to the time needed to reset the

441 microscope from a two-photon to a regular imaging configuration. The image scanning time recorded  
442 by the software was usually less than 400 ms, so the total exposure time of the chosen ROI to  
443 multiphoton laser was less than 1 ms. The cuts were oriented either in the antero-posterior (AP) or  
444 dorso-ventral (DV) directions relative to the global orientation of the embryos. After ablation, the  
445 embryos were monitored to see if they continued to develop normally or they expressed the desired  
446 phenotype. More precisely, we verified whether embryos ablated at 1.3F and 1.5F developed past the  
447 2F stage, embryos ablated at 1.7F developed past the 2.5F stage, *unc-112(RNAi)* embryos and *spc-*  
448 *1(RNAi)* embryos arrested at 2F stage.

449

#### 450 **Laser ablation image processing and data analysis**

451 The shape of the cut opening was detected using the Active Contour plugin ABsnake (Boudier, 1997).  
452 A starting ROI was drawn around the opening as the initiation ROI for ABsnake. After running the  
453 plugin, the results were checked and corrected for detection errors. The detected shape was fitted  
454 with an ellipse to derive the minor axis, major axis and the angle formed by the major axis with the  
455 initial cut direction. The average opening of the five last time points before the repair process began  
456 (Figure 2 –Figure supplement 1, from around 8 to 10 s after cutting) was taken as the opening at  
457 equilibrium. The standard error of the mean was shown.

458 The curve fit was performed on the average value of the cut opening (defined as the minor axis/initial  
459 cut length) using GraphPad Prism 5.00 (San Diego, California, USA) and the equation of one-phase  
460 association:

$$y = y_0 + (Plateau - y_0) * (1 - e^{-\gamma t})$$

461 where  $y_0$  is the initial width of the cut opening, *Plateau* is the minor axis of the opening at equilibrium  
462 and  $\gamma$  is the relaxation rate. The standard error of the mean given by the software was shown.

463

#### 464 **Statistical analysis**

465 The two-tailed t-test was performed on the average of the last five time points (from about 8 s to 10 s)  
466 of the cut opening using MATLAB R2014b (The MathWorks Inc., Natick, MA). Z-test was performed  
467 using QuickCalcs of GraphPad Prism (San Diego, California, USA) to compare the anisotropy of  
468 stress (AS), the relaxation half-time and the initial recoil speed of the cut opening.

469

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## 486 **Competing financial interests**

487 The authors declare no competing financial interests.

489 **References**

- 490 AIGOUY, B., FARHADIFAR, R., STAPLE, D. B., SAGNER, A., ROPER, J. C., JULICHER, F. &  
 491 EATON, S. 2010. Cell flow reorients the axis of planar polarity in the wing epithelium of  
 492 *Drosophila*. *Cell*, 142, 773-86, [doi:10.1016/j.cell.2010.07.042](https://doi.org/10.1016/j.cell.2010.07.042).
- 493 ANTUNES, M., PEREIRA, T., CORDEIRO, J. V., ALMEIDA, L. & JACINTO, A. 2013. Coordinated  
 494 waves of actomyosin flow and apical cell constriction immediately after wounding. *J Cell Biol*,  
 495 202, 365-79, [doi:10.1083/jcb.201211039](https://doi.org/10.1083/jcb.201211039).
- 496 BASKIN, T. I. 2005. Anisotropic expansion of the plant cell wall. *Annu Rev Cell Dev Biol*, 21, 203-22,  
 497 [doi:10.1146/annurev.cellbio.20.082503.103053](https://doi.org/10.1146/annurev.cellbio.20.082503.103053).
- 498 BEHRNDT, M., SALBREUX, G., CAMPINHO, P., HAUSCHILD, R., OSWALD, F., ROENSCH, J.,  
 499 GRILL, S. W. & HEISENBERG, C. P. 2012. Forces driving epithelial spreading in zebrafish  
 500 gastrulation. *Science*, 338, 257-60, [doi:10.1126/science.1224143](https://doi.org/10.1126/science.1224143).
- 501 BEN AMAR, M., WU, M., TREJO, M. & ATLAN, M. 2015. Morpho-elasticity of inflammatory fibrosis:  
 502 the case of capsular contracture. *J R Soc Interface*, 12, 20150343, [doi:10.1098/rsif.2015.0343](https://doi.org/10.1098/rsif.2015.0343).
- 503 BERT, C. W. 1977. Models for Fibrous Composites with Different Properties in Tension and  
 504 Compression. *Journal of Engineering Materials and Technology-Transactions of the Asme*, 99,  
 505 344-349,
- 506 BERTET, C., SULAK, L. & LECUIT, T. 2004. Myosin-dependent junction remodelling controls planar  
 507 cell intercalation and axis elongation. *Nature*, 429, 667-71, [doi:10.1038/nature02590](https://doi.org/10.1038/nature02590).
- 508 BLANKENSHIP, J. T., BACKOVIC, S. T., SANNY, J. S., WEITZ, O. & ZALLEN, J. A. 2006.  
 509 Multicellular rosette formation links planar cell polarity to tissue morphogenesis. *Dev Cell*, 11,  
 510 459-70, [doi:10.1016/j.devcel.2006.09.007](https://doi.org/10.1016/j.devcel.2006.09.007).
- 511 BOSVELD, F., BONNET, I., GUIRAO, B., TLILI, S., WANG, Z., PETITALOT, A., MARCHAND, R.,  
 512 BARDET, P. L., MARCQ, P., GRANER, F. & BELLAICHE, Y. 2012. Mechanical control of  
 513 morphogenesis by Fat/Dachsous/Four-jointed planar cell polarity pathway. *Science*, 336, 724-  
 514 7, [doi:10.1126/science.1221071](https://doi.org/10.1126/science.1221071).
- 515 BOUDIER, T. 1997. Elaboration d'un modèle de déformation pour la détection de contours aux  
 516 formes complexes. *In: MED., I. T. B. (ed.)*.
- 517 BRENNER, S. 1974. The genetics of *Caenorhabditis elegans*. *Genetics*, 77, 71-94,
- 518 CALZADO-MARTIN, A., ENCINAR, M., TAMAYO, J., CALLEJA, M. & SAN PAULO, A. 2016. Effect of  
 519 Actin Organization on the Stiffness of Living Breast Cancer Cells Revealed by Peak-Force  
 520 Modulation Atomic Force Microscopy. *ACS Nano*, 10, 3365-74,  
 521 [doi:10.1021/acsnano.5b07162](https://doi.org/10.1021/acsnano.5b07162).
- 522 CHAN, B. G., ROCHELEAU, S. K., SMIT, R. B. & MAINS, P. E. 2015. The Rho guanine exchange  
 523 factor RHGF-2 acts through the Rho-binding kinase LET-502 to mediate embryonic elongation  
 524 in *C. elegans*. *Dev Biol*, [doi:10.1016/j.ydbio.2015.07.010](https://doi.org/10.1016/j.ydbio.2015.07.010).
- 525 DAMON, B. J., MEZENTSEVA, N. V., KUMARATILAKE, J. S., FORGACS, G. & NEWMAN, S. A.  
 526 2008. Limb bud and flank mesoderm have distinct "physical phenotypes" that may contribute  
 527 to limb budding. *Dev Biol*, 321, 319-30, [doi:10.1016/j.ydbio.2008.06.018](https://doi.org/10.1016/j.ydbio.2008.06.018).
- 528 DIOGON, M., WISSLER, F., QUINTIN, S., NAGAMATSU, Y., SOOKHAREEA, S., LANDMANN, F.,  
 529 HUTTER, H., VITALE, N. & LABOUESSE, M. 2007. The RhoGAP RGA-2 and LET-502/ROCK  
 530 achieve a balance of actomyosin-dependent forces in *C. elegans* epidermis to control  
 531 morphogenesis. *Development*, 134, 2469-79, [doi:10.1242/dev.005074](https://doi.org/10.1242/dev.005074).
- 532 FEDERICO, S. & GASSER, T. C. 2010. Nonlinear elasticity of biological tissues with statistical fibre  
 533 orientation. *Journal of the Royal Society Interface*, 7, 955-966, [doi:10.1098/rsif.2009.0502](https://doi.org/10.1098/rsif.2009.0502).
- 534 FERNANDEZ-GONZALEZ, R. & ZALLEN, J. A. 2011. Oscillatory behaviors and hierarchical  
 535 assembly of contractile structures in intercalating cells. *Physical Biology*, 8, [doi:Artn 045005](https://doi.org/10.1088/1478-3975/8/4/045005)  
 536 [10.1088/1478-3975/8/4/045005](https://doi.org/10.1088/1478-3975/8/4/045005).



537 FOUCHARD, J., MITROSSILIS, D. & ASNACIOS, A. 2011. Acto-myosin based response to stiffness  
538 and rigidity sensing. *Cell Adh Migr*, 5, 16-9, doi:10.4161/cam.5.1.13281.

539 GALLY, C., WISSLER, F., ZAHREDDINE, H., QUINTIN, S., LANDMANN, F. & LABOUESSE, M.  
540 2009. Myosin II regulation during *C. elegans* embryonic elongation: LET-502/ROCK, MRCK-1  
541 and PAK-1, three kinases with different roles. *Development*, 136, 3109-19,  
542 doi:10.1242/dev.039412.

543 GASSER, T. C., OGDEN, R. W. & HOLZAPFEL, G. A. 2006. Hyperelastic modelling of arterial layers  
544 with distributed collagen fibre orientations. *J R Soc Interface*, 3, 15-35,  
545 doi:10.1098/rsif.2005.0073.

546 GONZALEZ, R. W., R.E 2008. *Digital image processing*.

547 GORFINKIEL, N., BLANCHARD, G. B., ADAMS, R. J. & MARTINEZ ARIAS, A. 2009. Mechanical  
548 control of global cell behaviour during dorsal closure in *Drosophila*. *Development*, 136, 1889-  
549 98, doi:10.1242/dev.030866.

550 GREEN, P. B. 1962. Mechanism for Plant Cellular Morphogenesis. *Science*, 138, 1404-5,  
551 doi:10.1126/science.138.3548.1404.

552 HAIGO, S. L. & BILDER, D. 2011. Global tissue revolutions in a morphogenetic movement controlling  
553 elongation. *Science*, 331, 1071-4, doi:10.1126/science.1199424.

554 HARDIN, J. & KELLER, R. 1988. The behaviour and function of bottle cells during gastrulation of  
555 *Xenopus laevis*. *Development*, 103, 211-30,

556 HEISENBERG, C. P. & BELLAICHE, Y. 2013. Forces in tissue morphogenesis and patterning. *Cell*,  
557 153, 948-62, doi:10.1016/j.cell.2013.05.008.

558 HELWIG, P., FAUST, G., HINDENLANG, U., HIRSCHMULLER, A., KONSTANTINIDIS, L., BAHRS,  
559 C., SUDKAMP, N. & SCHNEIDER, R. 2009. Finite element analysis of four different implants  
560 inserted in different positions to stabilize an idealized trochanteric femoral fracture. *Injury*, 40,  
561 288-95, doi:10.1016/j.injury.2008.08.016.

562 KASZA, K. E., ROWAT, A. C., LIU, J., ANGELINI, T. E., BRANGWYNNE, C. P., KOENDERINK, G. H.  
563 & WEITZ, D. A. 2007. The cell as a material. *Curr Opin Cell Biol*, 19, 101-7,  
564 doi:10.1016/j.ceb.2006.12.002.

565 KELLER, R. & WINKLBAUER, R. 1992. Cellular basis of amphibian gastrulation. *Curr Top Dev Biol*,  
566 27, 39-89,

567 LANGE, J. R. & FABRY, B. 2013. Cell and tissue mechanics in cell migration. *Exp Cell Res*, 319,  
568 2418-23, doi:10.1016/j.yexcr.2013.04.023.

569 LE DUC, Q., SHI, Q., BLONK, I., SONNENBERG, A., WANG, N., LECKBAND, D. & DE ROOIJ, J.  
570 2010. Vinculin potentiates E-cadherin mechanosensing and is recruited to actin-anchored  
571 sites within adherens junctions in a myosin II-dependent manner. *J Cell Biol*, 189, 1107-15,  
572 doi:10.1083/jcb.201001149.

573 LEVAYER, R. & LECUIT, T. 2012. Biomechanical regulation of contractility: spatial control and  
574 dynamics. *Trends Cell Biol*, 22, 61-81, doi:10.1016/j.tcb.2011.10.001.

575 MARTIN, A. C., KASCHUBE, M. & WIESCHAUS, E. F. 2009. Pulsed contractions of an actin-myosin  
576 network drive apical constriction. *Nature*, 457, 495-9, doi:10.1038/nature07522.

577 MARTIN, E., HAREL, S., NKENGFAC, B., HAMICHE, K., NEAULT, M. & JENNA, S. 2014. pix-1  
578 controls early elongation in parallel with mel-11 and let-502 in *Caenorhabditis elegans*. *PLoS*  
579 *one*, 9, e94684, doi:10.1371/journal.pone.0094684.

580 MAYER, M., DEPKEN, M., BOIS, J. S., JULICHER, F. & GRILL, S. W. 2010. Anisotropies in cortical  
581 tension reveal the physical basis of polarizing cortical flows. *Nature*, 467, 617-21,  
582 doi:10.1038/nature09376.

583 MILLER, Z., FUCHS, M. B. & ARCAN, M. 2002. Trabecular bone adaptation with an orthotropic  
584 material model. *J Biomech*, 35, 247-56,

585 MOORTHY, S., CHEN, L. & BENNETT, V. 2000. *Caenorhabditis elegans* beta-G spectrin is  
586 dispensable for establishment of epithelial polarity, but essential for muscular and neuronal  
587 function. *J Cell Biol*, 149, 915-30,

588 MURRELL, M. P. & GARDEL, M. L. 2012. F-actin buckling coordinates contractility and severing in a  
589 biomimetic actomyosin cortex. *Proc Natl Acad Sci U S A*, 109, 20820-5,  
590 doi:10.1073/pnas.1214753109.

591 MUSKKHELISHVILI, N. I. (ed.) 1975. *Some basic Problems of the Mathematical Theory of Elasticity*,  
592 Leyden: Noordhoff International Publishing.

593 NORMAN, K. R. & MOERMAN, D. G. 2002. Alpha spectrin is essential for morphogenesis and body  
594 wall muscle formation in *Caenorhabditis elegans*. *J Cell Biol*, 157, 665-77,  
595 [doi:10.1083/jcb.200111051](https://doi.org/10.1083/jcb.200111051).

596 PIEKNY, A. J., JOHNSON, J. L., CHAM, G. D. & MAINS, P. E. 2003. The *Caenorhabditis elegans*  
597 nonmuscle myosin genes *nmy-1* and *nmy-2* function as redundant components of the let-  
598 502/Rho-binding kinase and *mel-11*/myosin phosphatase pathway during embryonic  
599 morphogenesis. *Development*, 130, 5695-704, [doi:10.1242/dev.00807](https://doi.org/10.1242/dev.00807).

600 PRAITIS, V., CICCONE, E. & AUSTIN, J. 2005. SMA-1 spectrin has essential roles in epithelial cell  
601 sheet morphogenesis in *C. elegans*. *Dev Biol*, 283, 157-70, [doi:10.1016/j.ydbio.2005.04.002](https://doi.org/10.1016/j.ydbio.2005.04.002).

602 PRIESS, J. R. & HIRSH, D. I. 1986. *Caenorhabditis elegans* morphogenesis: the role of the  
603 cytoskeleton in elongation of the embryo. *Dev Biol*, 117, 156-73,

604 QUINTIN, S., WANG, S., PONTABRY, J., BENDER, A., ROBIN, F., HYENNE, V., LANDMANN, F.,  
605 GALLY, C., OEGEMA, K. & LABOUESSE, M. 2016. Non-centrosomal epidermal microtubules  
606 act in parallel to LET-502/ROCK to promote *C. elegans* elongation. *Development*, 143, 160-  
607 73, [doi:10.1242/dev.126615](https://doi.org/10.1242/dev.126615).

608 RAUZI, M. & LENNE, P. F. 2015. Probing cell mechanics with subcellular laser dissection of  
609 actomyosin networks in the early developing *Drosophila* embryo. *Methods Mol Biol*, 1189,  
610 209-18, [doi:10.1007/978-1-4939-1164-6\\_14](https://doi.org/10.1007/978-1-4939-1164-6_14).

611 RAUZI, M., LENNE, P. F. & LECUIT, T. 2010. Planar polarized actomyosin contractile flows control  
612 epithelial junction remodelling. *Nature*, 468, 1110-4, [doi:10.1038/nature09566](https://doi.org/10.1038/nature09566).

613 RAUZI, M., VERANT, P., LECUIT, T. & LENNE, P. F. 2008. Nature and anisotropy of cortical forces  
614 orienting *Drosophila* tissue morphogenesis. *Nat Cell Biol*, 10, 1401-10, [doi:10.1038/ncb1798](https://doi.org/10.1038/ncb1798).

615 RAZZELL, W., EVANS, I. R., MARTIN, P. & WOOD, W. 2013. Calcium flashes orchestrate the wound  
616 inflammatory response through DUOX activation and hydrogen peroxide release. *Curr Biol*,  
617 23, 424-9, [doi:10.1016/j.cub.2013.01.058](https://doi.org/10.1016/j.cub.2013.01.058).

618 ROGALSKI, T. M., MULLEN, G. P., GILBERT, M. M., WILLIAMS, B. D. & MOERMAN, D. G. 2000.  
619 The UNC-112 gene in *Caenorhabditis elegans* encodes a novel component of cell-matrix  
620 adhesion structures required for integrin localization in the muscle cell membrane. *J Cell Biol*,  
621 150, 253-64,

622 SAHA, A., NISHIKAWA, M., BEHRNDT, M., HEISENBERG, C. P., JULICHER, F. & GRILL, S. W.  
623 2016. Determining Physical Properties of the Cell Cortex. *Biophys J*, 110, 1421-9,  
624 [doi:10.1016/j.bpj.2016.02.013](https://doi.org/10.1016/j.bpj.2016.02.013).

625 SALKER, M. S., SCHIERBAUM, N., ALOWAYED, N., SINGH, Y., MACK, A. F., STOURNARAS, C.,  
626 SCHAFFER, T. E. & LANG, F. 2016. LeftyA decreases Actin Polymerization and Stiffness in  
627 Human Endometrial Cancer Cells. *Sci Rep*, 6, 29370, [doi:10.1038/srep29370](https://doi.org/10.1038/srep29370).

628 SAVIN, T., KURPIOS, N. A., SHYER, A. E., FLORESCU, P., LIANG, H., MAHADEVAN, L. & TABIN,  
629 C. J. 2011. On the growth and form of the gut. *Nature*, 476, 57-62, [doi:10.1038/nature10277](https://doi.org/10.1038/nature10277).

630 SAXENA, A., DENHOLM, B., BUNT, S., BISCHOFF, M., VIJAYRAGHAVAN, K. & SKAER, H. 2014.  
631 Epidermal growth factor signalling controls myosin II planar polarity to orchestrate convergent  
632 extension movements during *Drosophila* tubulogenesis. *PLoS Biol*, 12, e1002013,  
633 [doi:10.1371/journal.pbio.1002013](https://doi.org/10.1371/journal.pbio.1002013).

634 SHELTON, C. A., CARTER, J. C., ELLIS, G. C. & BOWERMAN, B. 1999. The nonmuscle myosin  
635 regulatory light chain gene *mlc-4* is required for cytokinesis, anterior-posterior polarity, and  
636 body morphology during *Caenorhabditis elegans* embryogenesis. *J Cell Biol*, 146, 439-51,  
637 SHINDO, A. & WALLINGFORD, J. B. 2014. PCP and septins compartmentalize cortical actomyosin to  
638 direct collective cell movement. *Science*, 343, 649-52, [doi:10.1126/science.1243126](https://doi.org/10.1126/science.1243126).

639 SMUTNY, M., BEHRNDT, M., CAMPINHO, P., RUPRECHT, V. & HEISENBERG, C. P. 2015. UV  
640 laser ablation to measure cell and tissue-generated forces in the zebrafish embryo in vivo and  
641 ex vivo. *Methods Mol Biol*, 1189, 219-35, [doi:10.1007/978-1-4939-1164-6\\_15](https://doi.org/10.1007/978-1-4939-1164-6_15).

642 SOLON, J., KAYA-COPUR, A., COLOMBELLI, J. & BRUNNER, D. 2009. Pulsed forces timed by a  
643 ratchet-like mechanism drive directed tissue movement during dorsal closure. *Cell*, 137, 1331-  
644 42, [doi:10.1016/j.cell.2009.03.050](https://doi.org/10.1016/j.cell.2009.03.050).

645 SULSTON, J. E., SCHIERENBERG, E., WHITE, J. G. & THOMSON, J. N. 1983. The embryonic cell  
646 lineage of the nematode *Caenorhabditis elegans*. *Dev Biol*, 100, 64-119,  
647 SUO, Z. 1990. Singularities, interfaces and cracks. *Proc. R. Soc. London, Ser. A*, 427, 331-358,  
648 THEOCARIS, P. S., PAZIS, D. & KONSTANTELLOS, B. D. 1986. The Exact Shape of a Deformed  
649 Internal Slant Crack under Biaxial Loading. *International Journal of Fracture*, 30, 135-153,  
650 TRICHET, L., LE DIGABEL, J., HAWKINS, R. J., VEDULA, S. R., GUPTA, M., RIBRAULT, C.,  
651 HERSEN, P., VOITURIEZ, R. & LADOUX, B. 2012. Evidence of a large-scale  
652 mechanosensing mechanism for cellular adaptation to substrate stiffness. *Proc Natl Acad Sci*  
653 *U S A*, 109, 6933-8, [doi:10.1073/pnas.1117810109](https://doi.org/10.1073/pnas.1117810109).  
654 VICENTE-MANZANARES, M., MA, X., ADELSTEIN, R. S. & HORWITZ, A. R. 2009. Non-muscle  
655 myosin II takes centre stage in cell adhesion and migration. *Nat Rev Mol Cell Biol*, 10, 778-90,  
656 [doi:10.1038/nrm2786](https://doi.org/10.1038/nrm2786).  
657 VUONG-BRENDER, T. T., YANG, X. & LABOUESSE, M. 2016. *C. elegans* Embryonic  
658 Morphogenesis. *Curr Top Dev Biol*, 116, 597-616, [doi:10.1016/bs.ctdb.2015.11.012](https://doi.org/10.1016/bs.ctdb.2015.11.012).  
659 WAINWRIGHT, S. A. 1988. *Axis and Circumference*, Harvard University Press.  
660 WALCOTT, S. & SUN, S. X. 2010. A mechanical model of actin stress fiber formation and substrate  
661 elasticity sensing in adherent cells. *Proc Natl Acad Sci U S A*, 107, 7757-62,  
662 [doi:10.1073/pnas.0912739107](https://doi.org/10.1073/pnas.0912739107).  
663 WISSMANN, A., INGLES, J. & MAINS, P. E. 1999. The *Caenorhabditis elegans mel-11* myosin  
664 phosphatase regulatory subunit affects tissue contraction in the somatic gonad and the  
665 embryonic epidermis and genetically interacts with the Rac signaling pathway. *Dev Biol*, 209,  
666 111-27,  
667 WISSMANN, A., INGLES, J., MCGHEE, J. D. & MAINS, P. E. 1997. *Caenorhabditis elegans* LET-502  
668 is related to Rho-binding kinases and human myotonic dystrophy kinase and interacts  
669 genetically with a homolog of the regulatory subunit of smooth muscle myosin phosphatase to  
670 affect cell shape. *Genes Dev*, 11, 409-22,  
671 XU, S. & CHISHOLM, A. D. 2011. A Galphaq-Ca(2)(+) signaling pathway promotes actin-mediated  
672 epidermal wound closure in *C. elegans*. *Curr Biol*, 21, 1960-7, [doi:10.1016/j.cub.2011.10.050](https://doi.org/10.1016/j.cub.2011.10.050).  
673 YOFFE, E. H. 1951. The moving Gri

674 ffith crack. *Philo. Mag*, 42, 739-750,  
675 YONEMURA, S., WADA, Y., WATANABE, T., NAGAFUCHI, A. & SHIBATA, M. 2010. alpha-Catenin  
676 as a tension transducer that induces adherens junction development. *Nat Cell Biol*, 12, 533-  
677 42, [doi:10.1038/ncb2055](https://doi.org/10.1038/ncb2055).  
678 ZHANG, H. & LABOUESSE, M. 2012. Signalling through mechanical inputs: a coordinated process. *J*  
679 *Cell Sci*, 125, 3039-49, [doi:10.1242/jcs.093666](https://doi.org/10.1242/jcs.093666).  
680

## 681 **Figure legends**

682 Figure 1: **Overview of *C. elegans* embryonic elongation.** (a) *C. elegans* embryonic elongation is  
683 driven in part by epidermal actomyosin contractility and in part by muscle contractions. The length of  
684 the embryo is used for staging: 2-fold (2F) stage means roughly 2-fold increase in length from the  
685 beginning of elongation. Representative stages are shown; anterior to the left, dorsal up. (b, c, d)  
686 Actin filament organization at the 1.3F, 1.5F and 1.7F stages, respectively, visualized with an  
687 ABD::GFP marker. Actin filaments progressively organize into circumferential parallel bundles in DV  
688 cells (arrows), arrowheads point to seam cells. Note that the integrated ABD::GFP marker shows  
689 some cell to cell variation of expression. (b', b'', c', c'', d', d'') Zoom in of actin pattern in DV cells  
690 (rectangle above) and seam cells (rectangle below), respectively, of the images in (b), (c) and (d)  
691 respectively. (e) Actomyosin forces squeeze the embryo circumferentially to make it elongate in the  
692 antero-posterior direction. (f, g) Endogenous distribution of the two non-muscle myosin II isoforms  
693 visualized with CRISPR GFP-labelled myosin heavy chains NMY-1 and NMY-2, respectively.  
694 Arrowheads point to seam cells, which are delineated by the junctional marker DLG-1::RFP.

695

696 Figure 2: **Physical model using the shape of the cut opening at equilibrium to measure the ratio**  
697 **of stress to Young modulus.** (a) The GFP-labelled actin cortex of HYP7 dorsal epidermal cell at the  
698 1.7F stage before (0 s), 1.4 s and 10 s after laser severing; cut along the AP direction with a 5  $\mu$ m cut  
699 length. Double arrowheads, distance between cut borders, which increases with time. (b) Model of  
700 epidermal cells as an infinite elastic plane under biaxial stress in the AP and DV directions after an  
701 incision of length  $l$ . The final shape of the cut opening is an ellipse. The cut opening in the DV  
702 direction (after an incision along the AP direction) depends on the stress along the DV direction and  
703 the Young modulus (see text). (c) The opening depends on myosin II activity: comparison of the cut  
704 response in the seam cell H1 between wild-type (WT) and *let-502(sb118)/Rho kinase* mutant embryos  
705 at a stage when muscles start to twitch (around 1.5F). The average value and standard error are  
706 reported. Time zero, moment of the cut; DV and AP show direction of opening. Two-tailed t-test, \*\*\*:

707  $p=4*10^{-7}$  between WT DV and *let-502(sb118)* DV,  $p=4*10^{-6}$  between WT AP and *let-502(sb118)* AP;  
708 N, number of embryos examined.

709

710 Figure 2 -Figure supplement 1: **Evolution of the distance between the cut borders (the minor axis**  
711 **of the cut opening) versus time.** The average value and standard error are reported. Time 0 is the  
712 moment of the cut. Solid lines show the single exponential fit with an initial width of cut opening of 0.6  
713  $\mu\text{m}$  (see Methods).

714

715 Figure 2 -Figure supplement 2: **Calcium imaging of ablated embryos.** (a) Scheme for measuring  
716 the calcium sensor GCaMP3 mean fluorescence intensity at positions 9  $\mu\text{m}$  and 17  $\mu\text{m}$  away from the  
717 cut site (red bar). The intensity was averaged over a region of interest (ROI) of  $1.5 \times 1.5 \mu\text{m}^2$ . A,  
718 anterior; P, posterior; D, dorsal; V, ventral. (b, c) Calcium level over time for a representative embryo  
719 of the majority of ablated embryos likely corresponding to a local disruption of the cortex (b), or of the  
720 minority of ablated embryos likely corresponding to a more severe wounding response (c).

721

722 Figure 3: **Stress anisotropy in seam cells correlates with morphological changes and partially**  
723 **depends on the spectrin cytoskeleton.** (a) Scheme showing laser ablation experiments in the AP  
724 and DV directions for H1, V3 and V6 seam cells at different stages. A, anterior; P, posterior; D, dorsal;  
725 V, ventral. (b) Cut opening in H1, V3 and V6 from the 1.3F to the 1.7F stages (see figure 2b). p-value  
726 of two-tailed t-test is reported. (c) Changes in embryo length, head diameter at the level of H1, body  
727 diameter at the level of V3 and tail diameter at the level of V6, between the 1.3F and 1.7F stages.  
728 N=10. (d) Scheme showing the measurement of circumferential cell width in the head (above) and  
729 corresponding section (below). (e) The circumferential width of H1, V3, head and body DV cells  
730 (averaged for dorsal and ventral cells) is reported. N=10. (f) Measures of the cut opening in H1 for  
731 WT, *spc-1(RNAi)* treated and *unc-112(RNAi)* muscle defective embryos at a stage equivalent to the  
732 1.7F stage. p-value of two-tailed t-test is reported. (g) Comparison of the stress anisotropy in H1,

733 defined by DV/AP stress, between WT 1.7F stage, *spc-1(RNAi)* embryos and *unc-112(RNAi)* embryos  
734 at the equivalent 1.7F stage. p-value of Z-test is reported. The number of embryos used for ablation is  
735 given in supplementary table 1. For **(b,c,e-g)**, the average (or calculated) value and standard error  
736 are reported.

737

738 Figure 3 -Figure supplement 1: **Elongation curves normalized to the initial embryo length for**  
739 **different genetic backgrounds.** ML1540 is the strain carrying the actin-binding domain fused to GFP  
740 (see Methods). The average value and standard error are reported.  $N \geq 9$  for each genotype.

741

742 Figure 4: **Stress anisotropy induces embryonic morphological changes.** **(a, b, c)** Anisotropy of  
743 stress (AS) for a sphere, an ellipsoid and a cylinder with DV and AP axis defined in the schemes; **(a,**  
744 **b)** show the middle plane; the major and minor axis of the ellipsoid are called  $a_1$  and  $a_2$ . **(d, e, f)** The  
745 embryo is schematized with a spherical (1.3F and 1.5F stages) or ellipsoidal (1.7F stage) head, and a  
746 curved cylindrical body. The AS in the head evolves from 1 (sphere) to that of an ellipsoid, whereas  
747 the body AS depends on the ratio of body to head radius ( $R_2/R_1$ ). **(g)** Comparison of the predicted AS  
748 based on embryo diameter measurements (see figure 3c) and the measured AS obtained from laser  
749 ablation experiments (figure 3b). **(h)** Hooke's law written for an isotropic material like seam cells  
750 (Appendix 6A);  $\epsilon_{AP}^S$  and  $\epsilon_{DV}^S$  are the relative length changes along the AP and DV directions,  
751 respectively. The stress  $\sigma_{AP}^S$  and  $\sigma_{DV}^S$  along the AP and DV directions are supposed to be contractile  
752 (so negative).  $E$ , seam cell Young modulus; A, anterior; P, posterior; D, dorsal; V, ventral. **(i, j)**  
753 Dependence of the AP and DV relative length change on the anisotropy of stress for different values  
754 of  $\sigma_{AP}^S/E$ .

755

756 Figure 5: **The dorso-ventral epidermis behaves differently than the H1 seam cell in ablation**  
757 **experiments.** **(a)** Scheme showing laser ablation experiments in the epithelial cell HYP7 dorsal and  
758 ventral to H1, yellow crosses show cut directions. **(b)** Cut opening in the DV and AP directions

759 measured in HYP7 between the 1.3F and 1.7F stages (see figure 2b). p-value of two-tailed t-test is  
760 reported. (c) Comparison of the DV/AP opening ratio in the seam cell H1 and head HYP7 cell. The  
761 data was derived from 3b and 5b. To simplify, we will call the cells participating in the future HYP7  
762 syncytium the HYP7 cells. The average (or computed) value and standard error are reported. The  
763 number of embryos used for ablation is given in supplementary table 1.

764

765 Figure 6: **Model of cut opening for a fiber-reinforced material.** (a) Considering a composite  
766 material with fiber reinforcement along the DV direction, the ratio of the Young modulus of the  
767 composite material along the DV direction,  $E_{DV}$ , to the Young modulus of the material without fibers  
768 (matrix),  $E_0$ , depends linearly on a factor  $K$  (see Appendix 7);  $K$  is related to fiber density and  
769 stiffness. (b) The ratio of the Young modulus along the AP direction,  $E_{AP}$ , to the matrix Young  
770 modulus,  $E_0$ , varies little with  $K$ . (c) The opening to the cuts perpendicular to the fibers is similar to an  
771 isotropic plane response, and depends on the ratio of DV stress to the matrix Young modulus  $\sigma_{DV}/E_0$ .  
772 (d) The opening to the cuts parallel to the fibers depends on the factor  $K$ , the ratio of AP/DV stiffness,  
773  $E_{AP}/E_{DV}$ , and the ratio of stress over Young modulus in the AP direction,  $\sigma_{AP}/E_{AP}$ .

774

775 Figure 7: **The anisotropy of stiffness in the HYP7 cell helps the embryo to elongate.** (a) The cut  
776 opening in HYP7 is linearly related on the cut opening in H1. The slope of the linear regression gives  
777 the ratio of the HYP7 matrix without fibers to H1 Young moduli. (b) The factor  $K$  increases during  
778 elongation from the 1.3F to 1.7F stages. (c). The ratio of DV/AP stiffness increases and is greater  
779 than the AS during early elongation. The data was derived from 3c and SI10. (d) Hooke's law written  
780 for a fiber-reinforced material such as DV cells (Appendix 6B).  $\epsilon_{AP}$  and  $\epsilon_{DV}$  are the relative length  
781 change along the AP and DV directions, respectively. The stress  $\sigma_{AP}$  and  $\sigma_{DV}$  along the AP and DV  
782 directions are supposed to be tensile (so positive).  $E_{AP}$  and  $E_{DV}$  are the Young moduli in the DV cells  
783 along the AP and DV direction, respectively.  $\nu_1$  and  $\nu_2$  are Poisson's ratios in DV cells;  $\omega$  is the  
784  $E_{DV}/E_{AP}$  ratio. A, anterior; P, posterior; D, dorsal; V, ventral. (e) Dependence of the AP relative length



785 change  $\epsilon_{AP}$  on the ratio of DV/AP stiffness  $\omega$  for different value of AS and  $\sigma_{AP}/E_{AP}$ ; the  $\nu_1$  value is  
786 taken to be 1. (f) Dependence of the DV relative length change  $\epsilon_{DV}$  on the ratio of DV/AP stiffness  $\omega$   
787 for different value of AS and  $\sigma_{DV}/E_{DV}$ ; the  $\nu_2$  value is taken to be 1. For (a-c), the average (or  
788 computed) value and standard error are reported.

789

790 Figure 8: **Actin filament organization correlated with stress and stiffness anisotropy pattern.** (a-  
791 c) Angle distribution of actin filaments in the seam cell H1 (a), seam V3 (b) and the HYP7 cell (c), at  
792 different elongation stages. D, dorsal; V, ventral; A, anterior; P, posterior. 90° correspond to DV  
793 direction. (d, e, f) Comparison of the peak values at  $90^\circ \pm 8^\circ$  (DV direction) of angular distribution  
794 showed in (a, b, c) respectively. p-value of two-tailed t-test is reported. (g) (left) The anisotropy of  
795 mechanical stress generated by the polarized actomyosin network and medial myosin pulses promote  
796 *Drosophila* germband extension; (right) The interplay of stress anisotropy (generated in seam cells -  
797 red) and stiffness anisotropy (DV cells – white) promote *C. elegans* embryo elongation. Note that,  
798 while myosin II does not display a polarized distribution within individual *C. elegans* epidermal cells  
799 like in *Drosophila* germband epithelial cells, its enrichment in seam cells along the circumference is  
800 reminiscent of the localized myosin II enrichment at vertical junctions in *Drosophila*. A, anterior; P,  
801 posterior. For (a-f), the average value and standard error are reported.

802

## 803 **Supplementary movie legends**

804 **Supplementary movie 1:** local disruption of actin cortex with laser ablation, visualized with the actin  
805 marker (ABD::mCHERRY) expressed under the epidermal promoter *lin-26*. 0 s time corresponds to  
806 first picture after laser cut. Yellow line shows the cut region.

807 **Supplementary movie 2:** local disruption of actin cortex with laser ablation does not induce  
808 noticeable change in calcium level. The calcium sensor GCaMP3 was expressed under the  
809 epidermal promoter *dpy-7* (strong expression in the dorso-ventral cells). 0 s time corresponds to first  
810 picture after laser cut. Yellow line shows the cut region. It is the same embryo as shown in movie 1.

811 **Supplementary movie 3:** wound healing response after laser ablation visualized with the actin  
812 marker ABD::mCHERRY expressed under the epidermal promoter *lin-26*. 0 s time corresponds to first  
813 picture after laser cut. Yellow line shows the cut region.

814 **Supplementary movie 4:** calcium wave propagation in wound healing response after laser ablation.  
815 The calcium sensor GCaMP3 was expressed under the epidermal *dpy-7* promoter (strong expression  
816 in the dorso-ventral cells). 0 s time corresponds to first picture after laser cut. Yellow line shows the  
817 cut region. It is the same embryo as shown in movie 3.

818

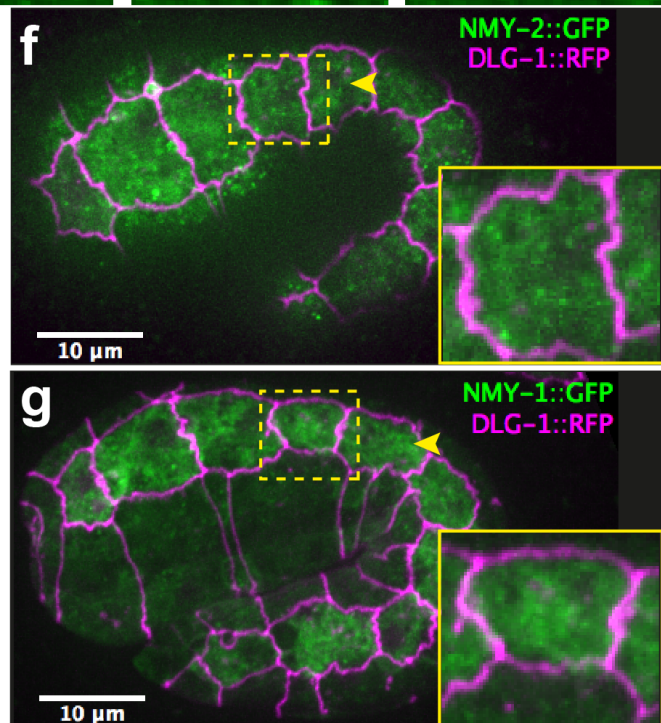
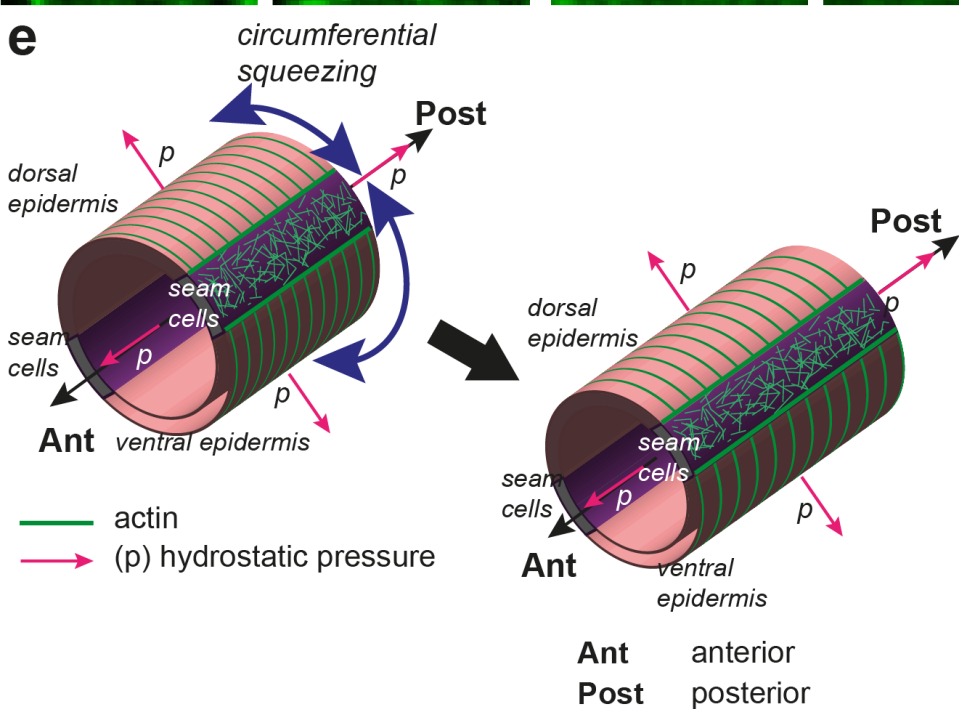
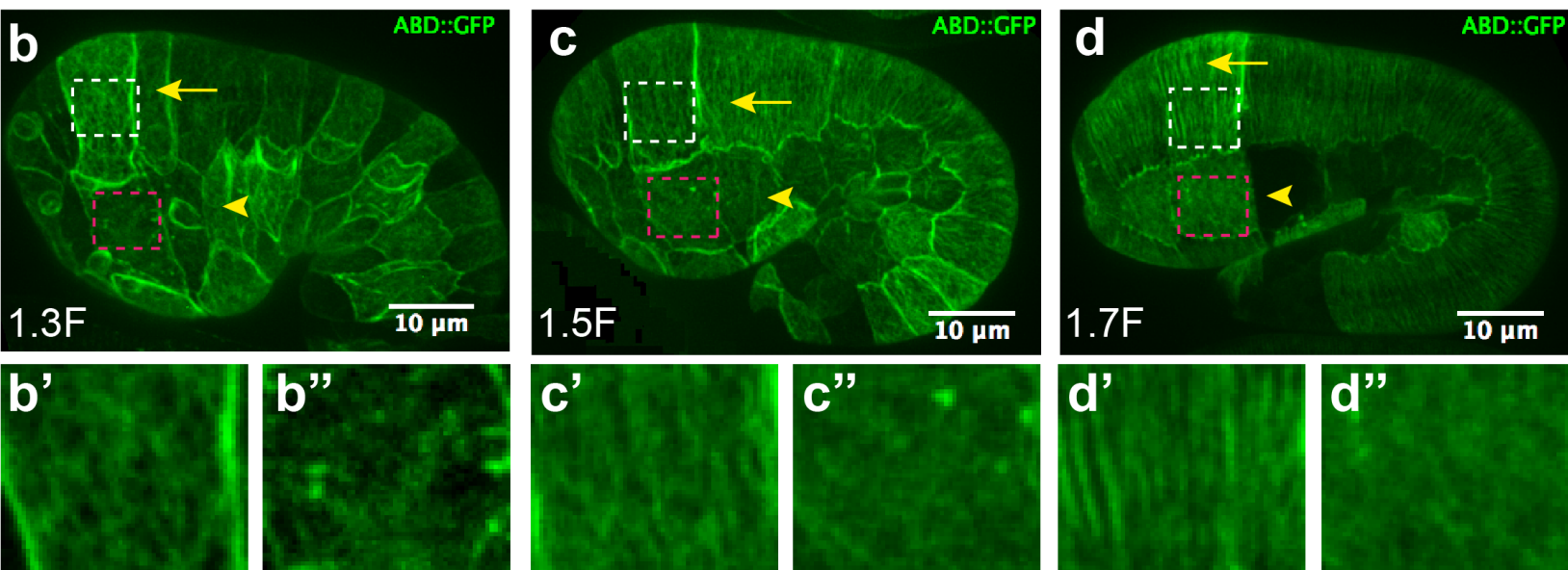
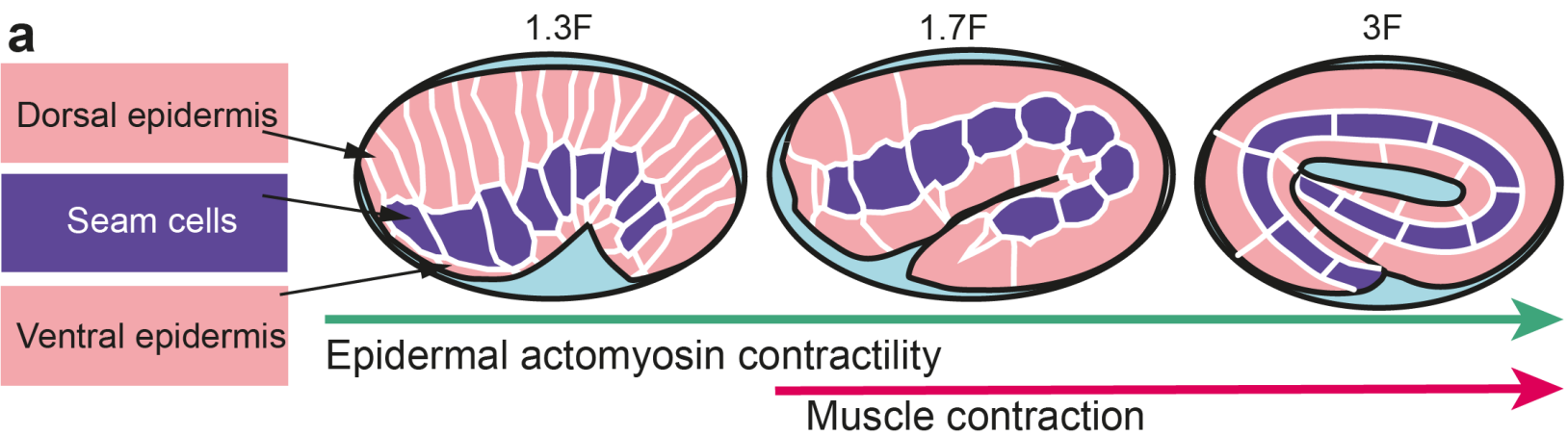
819 **Supplementary File legends**

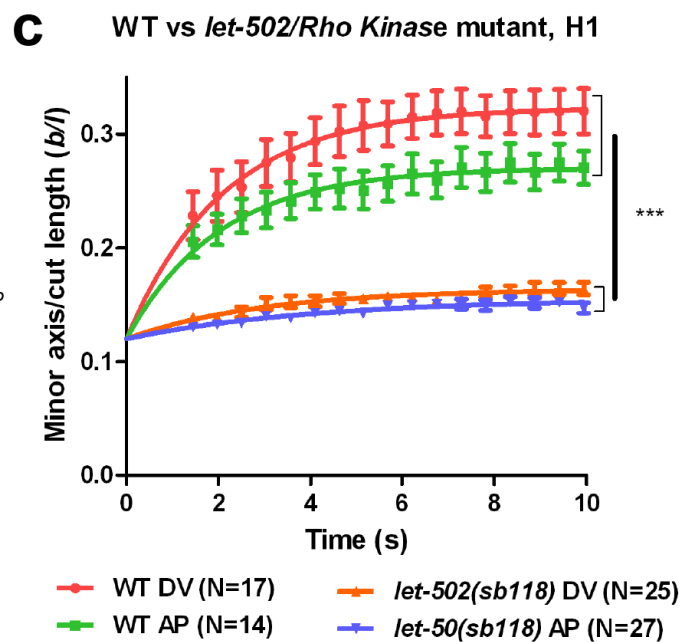
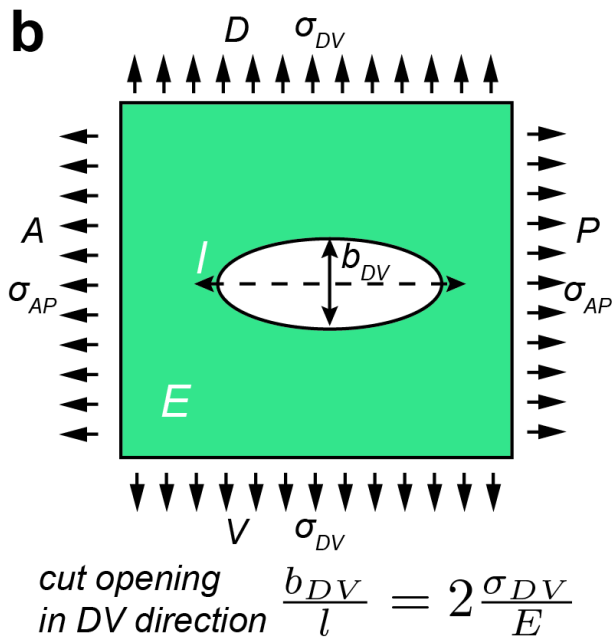
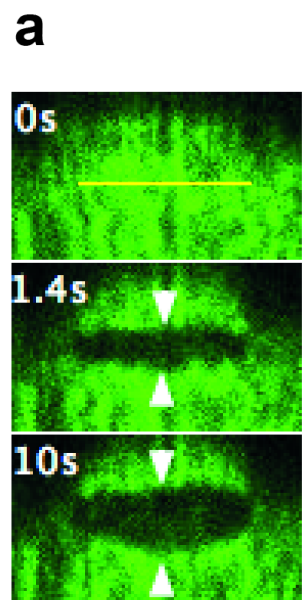
820 **Supplementary File 1:** Number of embryos used for laser ablation.

821 **Supplementary File 2:** Goodness of fit ( $R^2$ ) of cut border relaxation with different initial width of the  
822 cut openings.

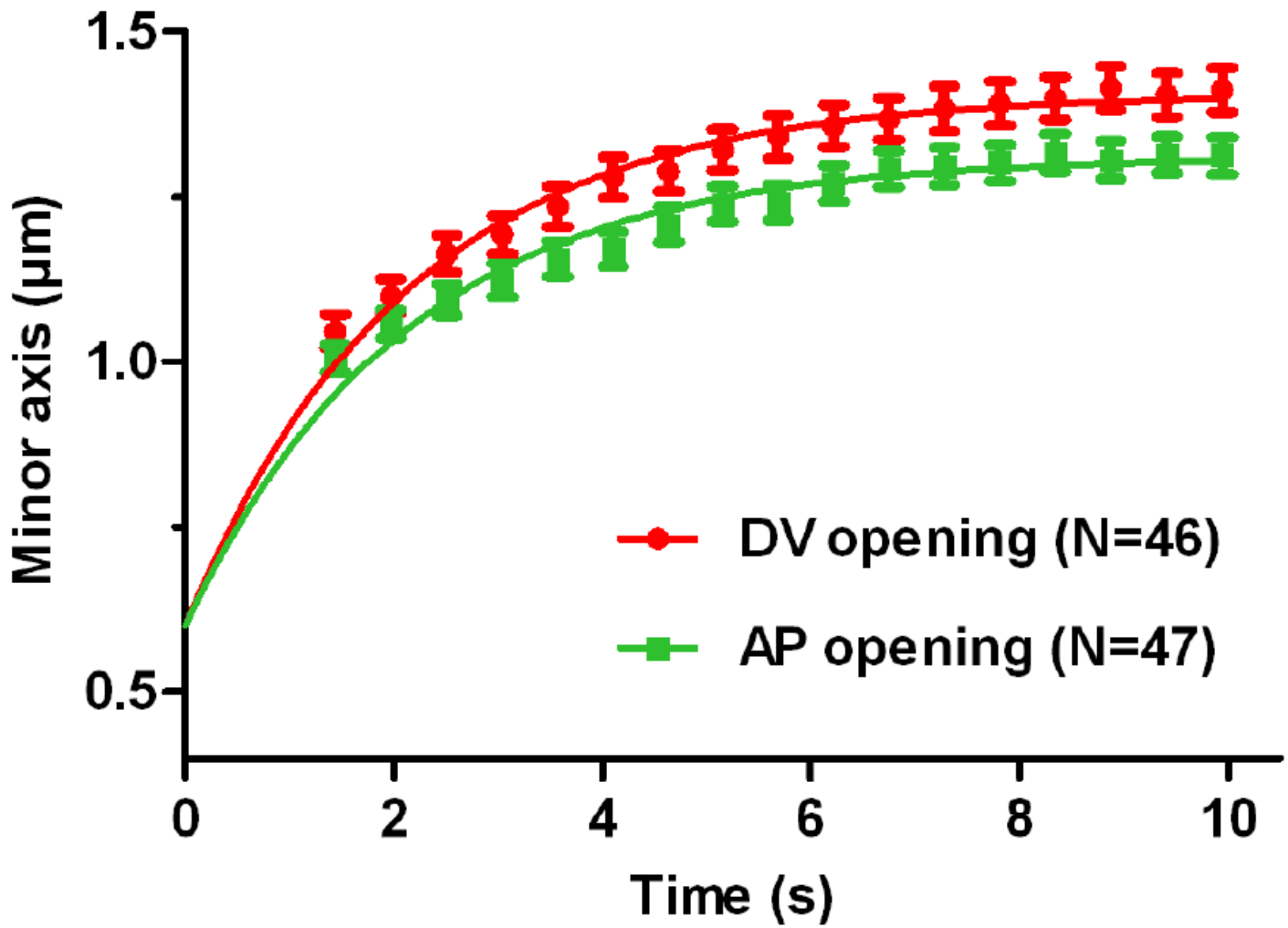
823 **Supplementary File 3:** Parameters derived from single exponential fit of the cut relaxation with an  
824 initial width of cut openings of 0.6  $\mu\text{m}$ . (see Methods).

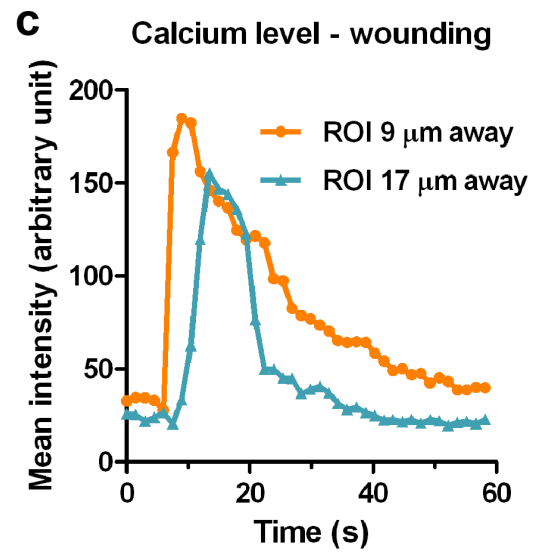
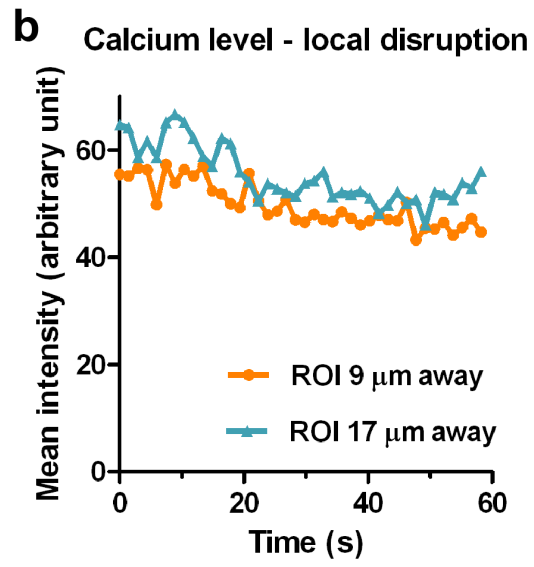
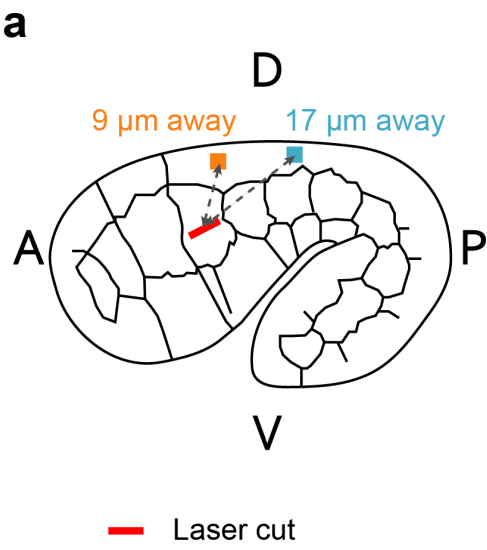
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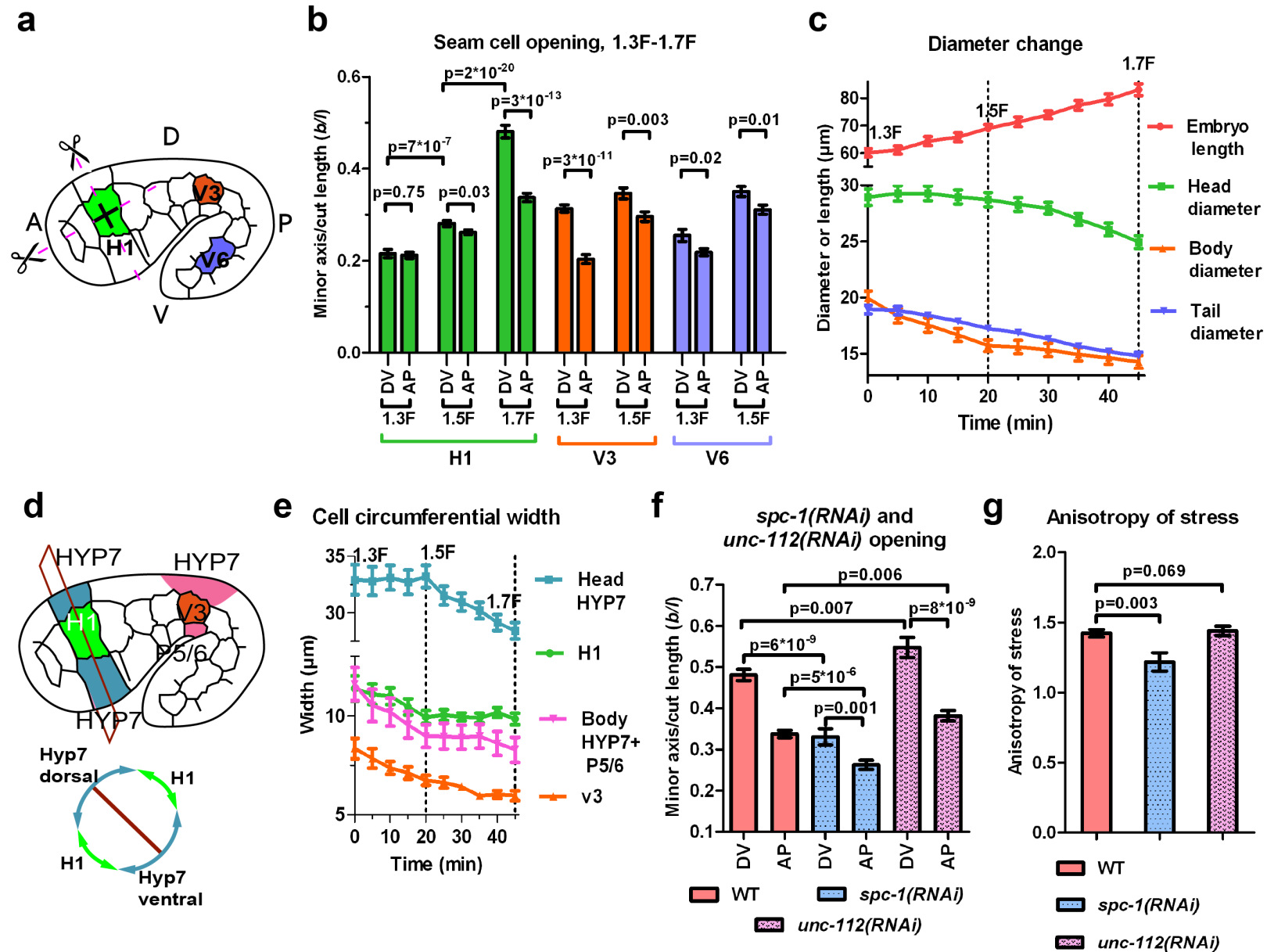




# 1.5F H1

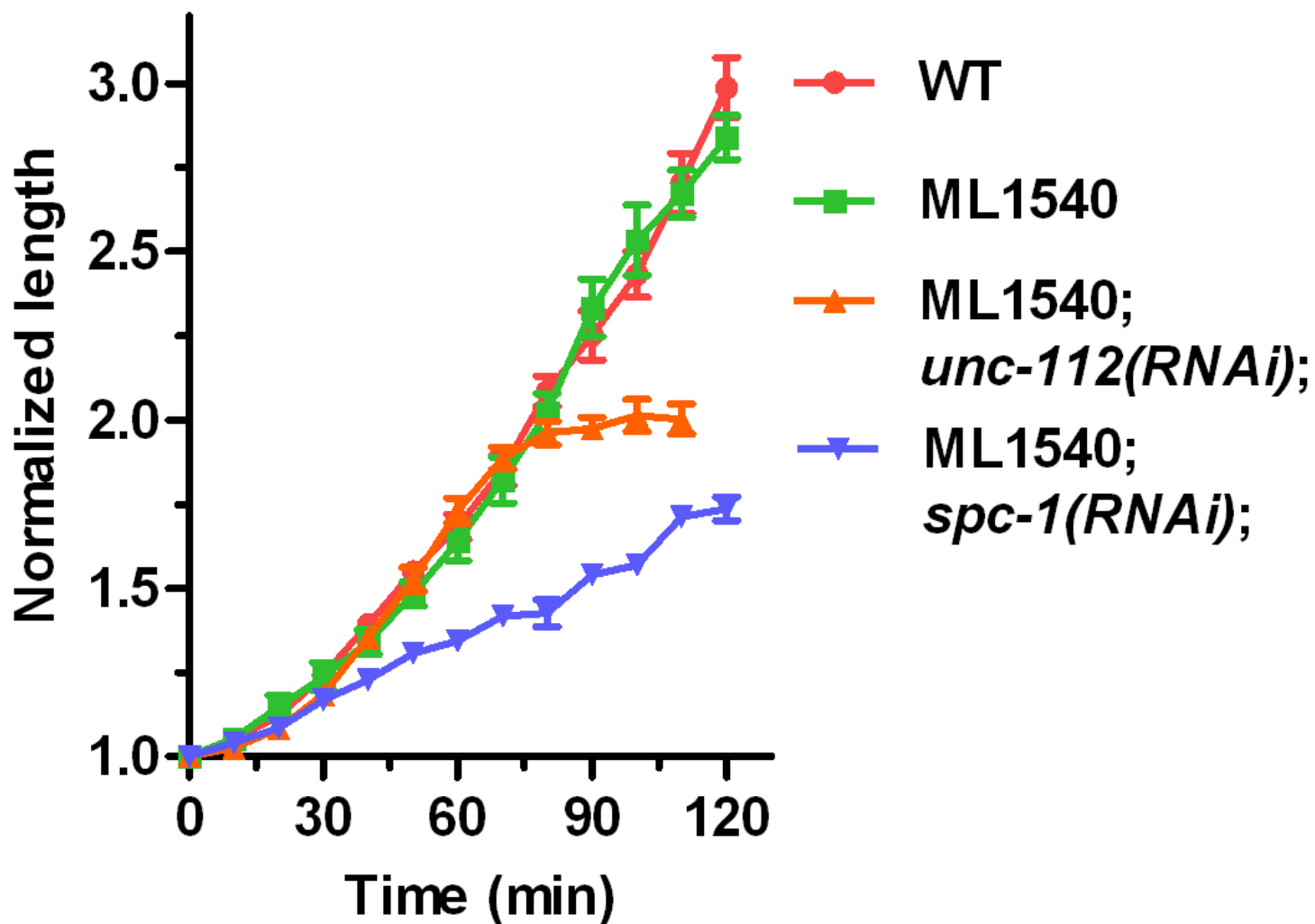


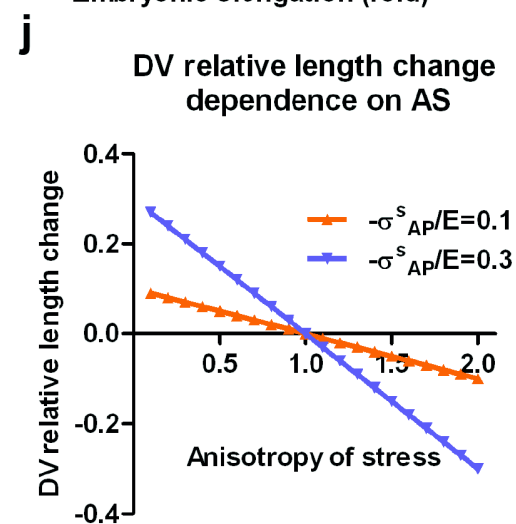
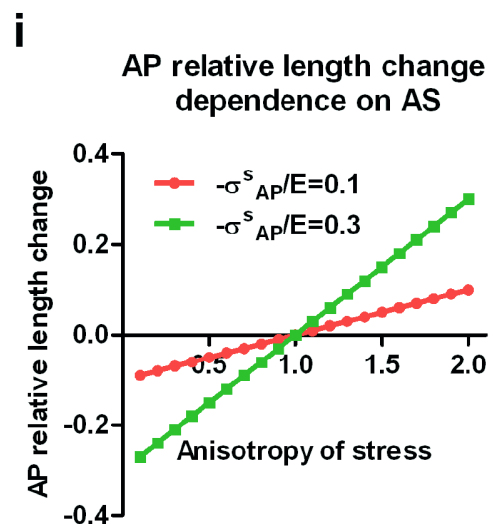
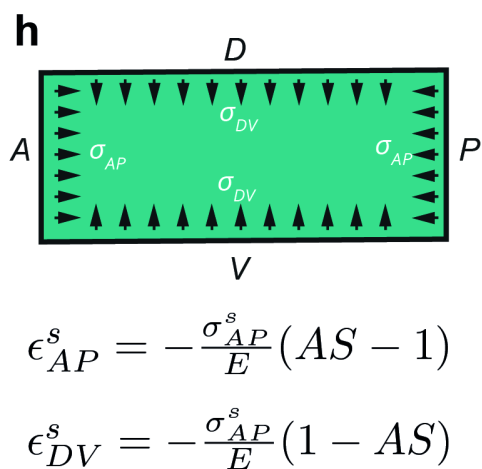
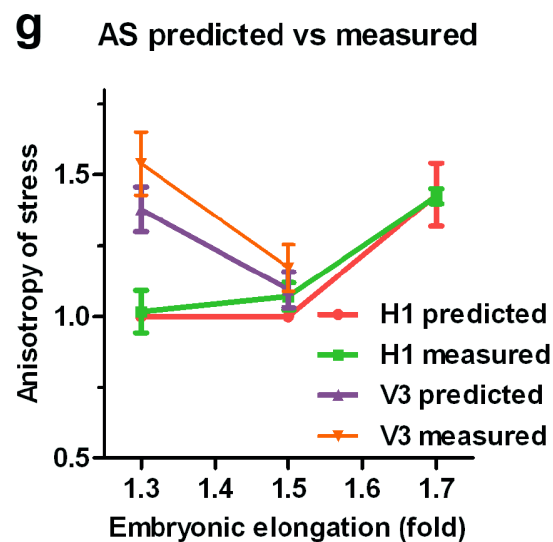
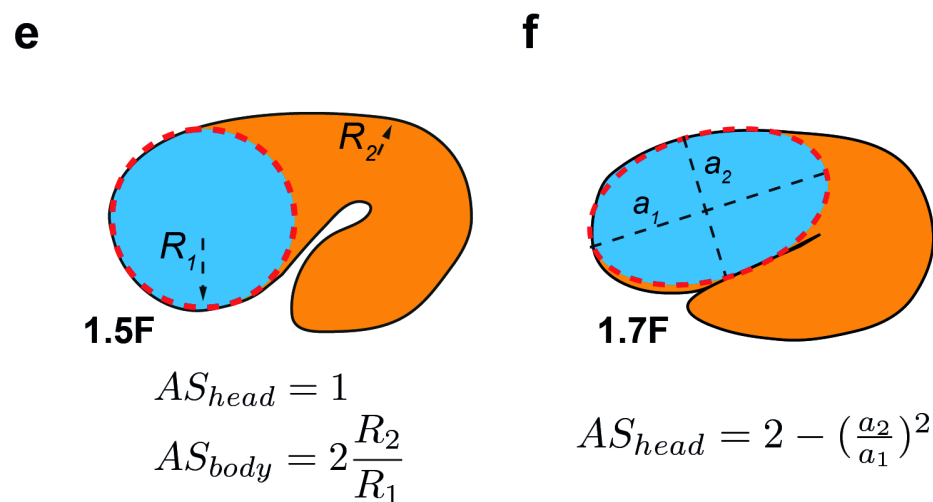
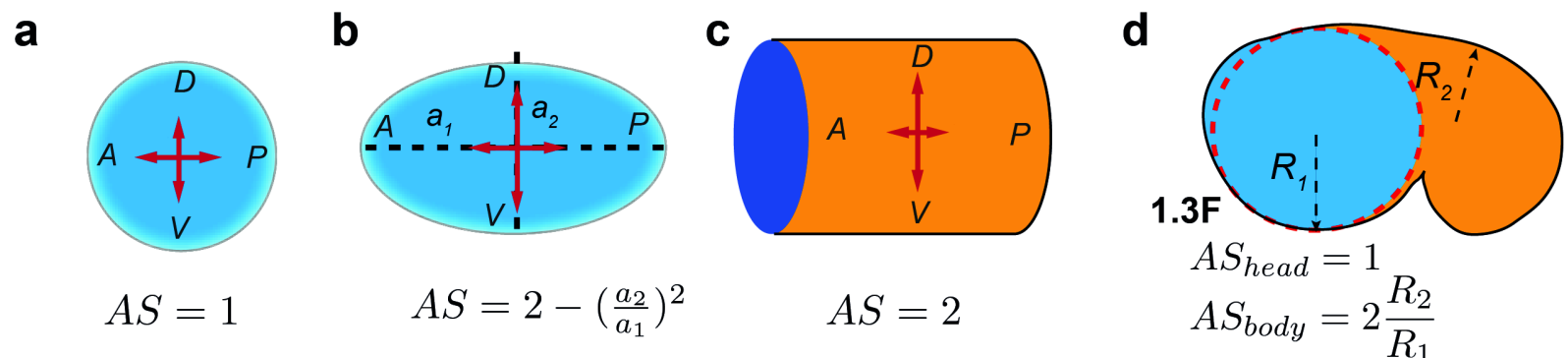


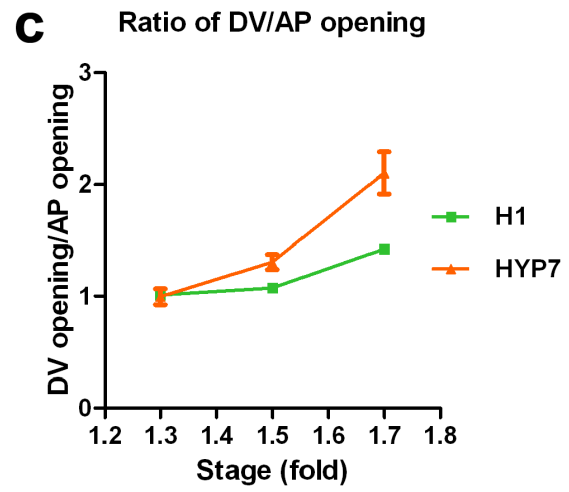
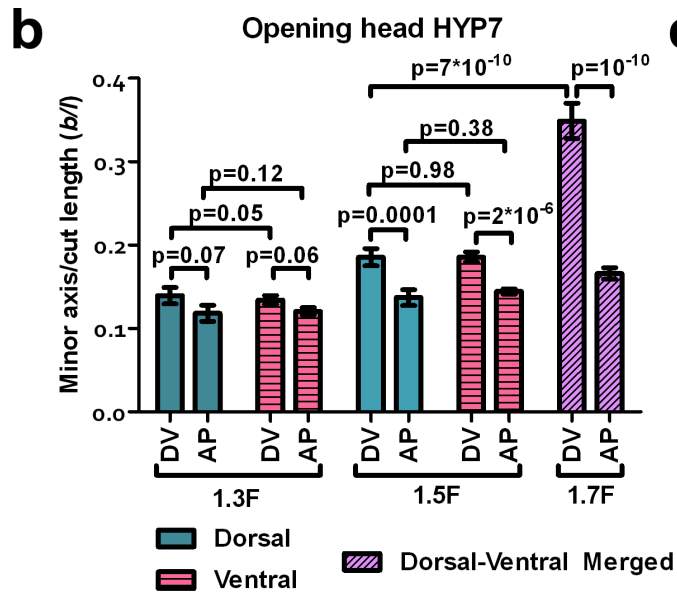
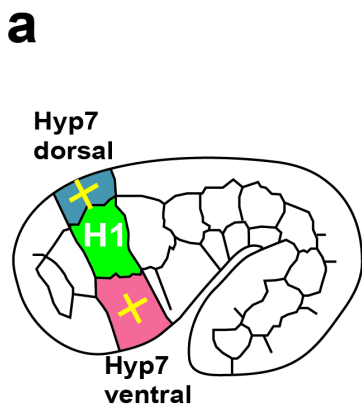


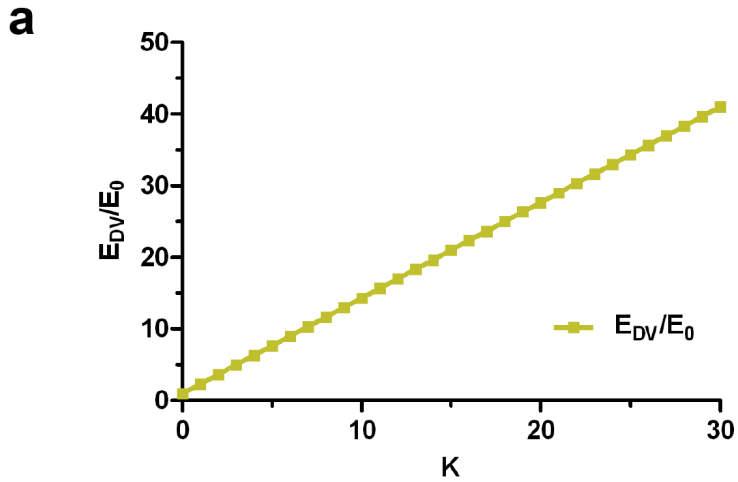


## Elongation in different backgrounds

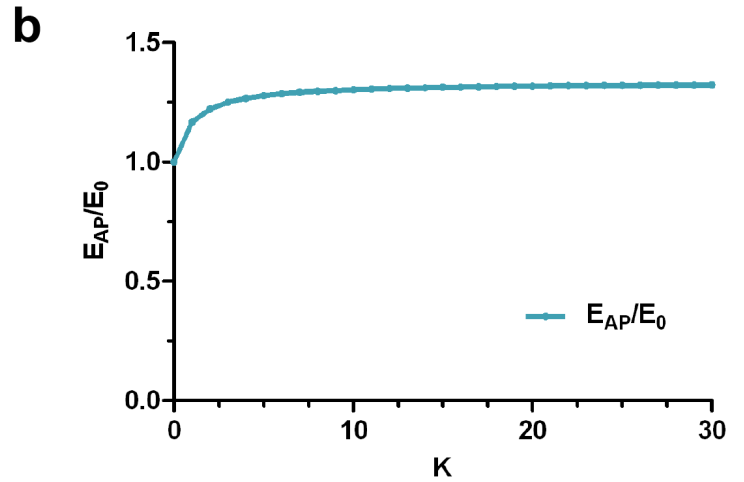




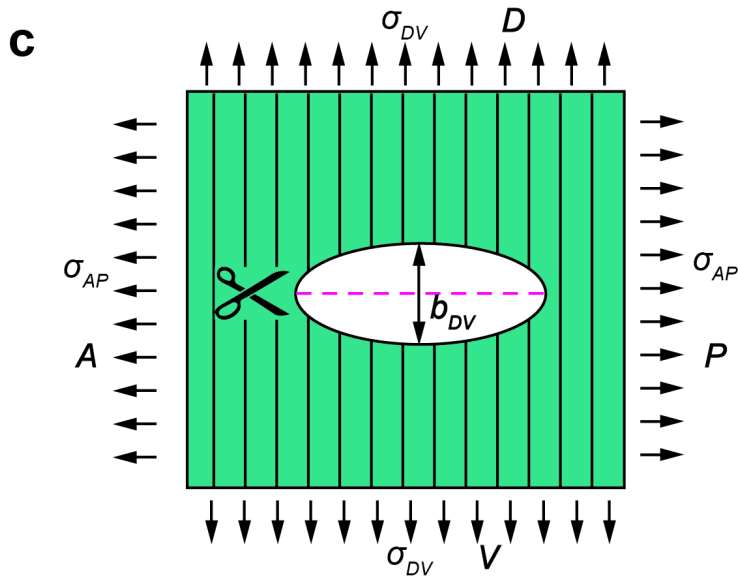




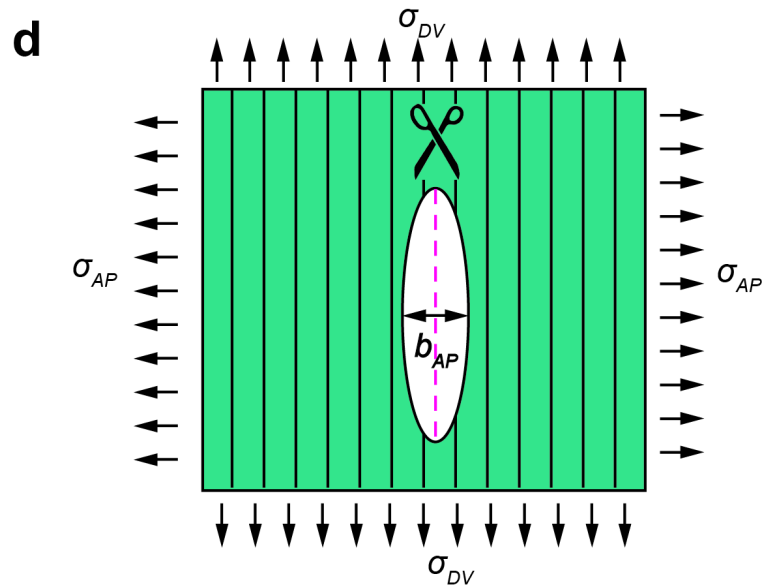
$$E_{DV} = \frac{E_0}{3}(3 + 4K)$$



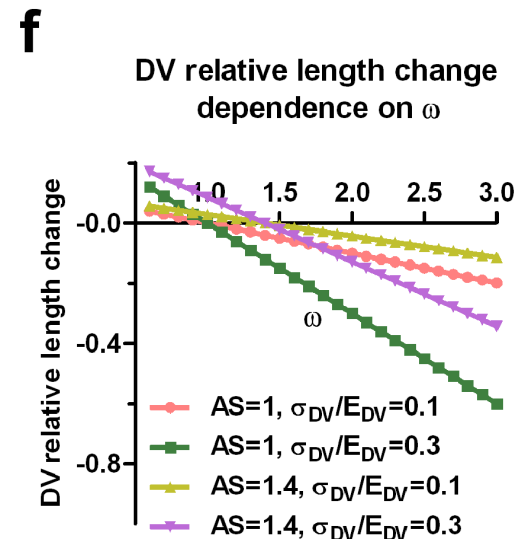
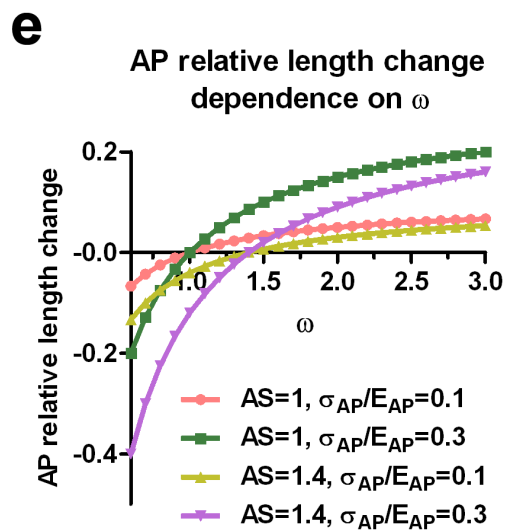
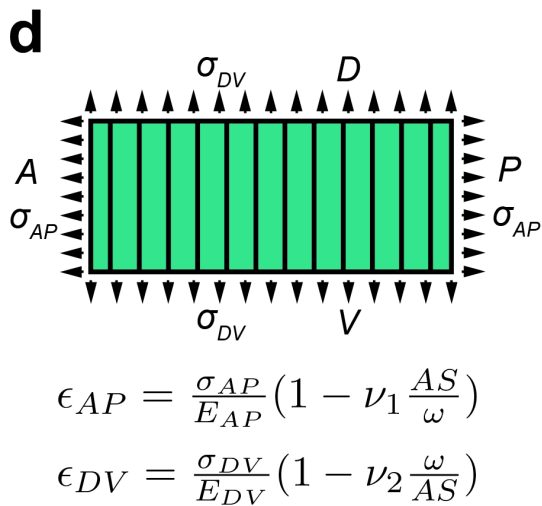
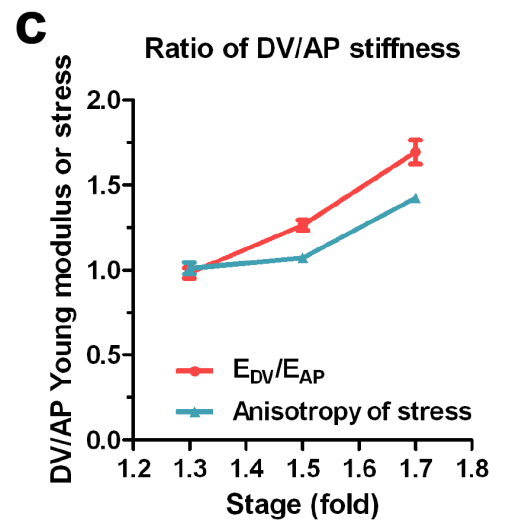
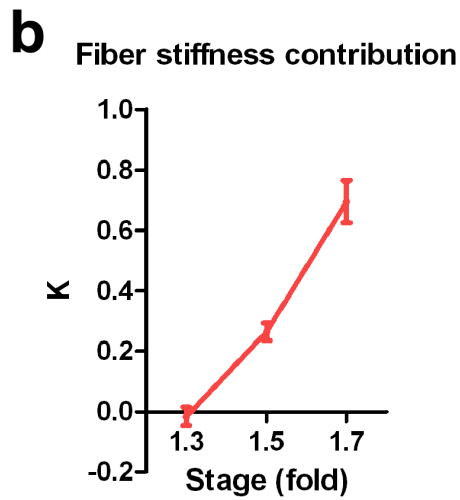
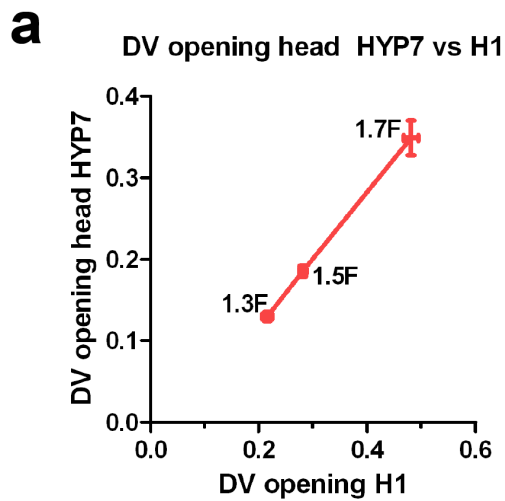
$$E_{AP} = \frac{E_0}{3} \frac{3+4K}{1+K}$$

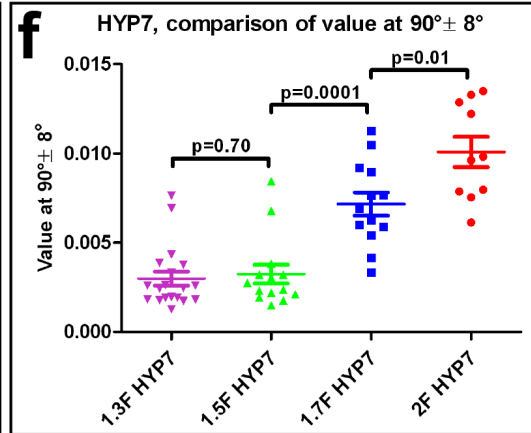
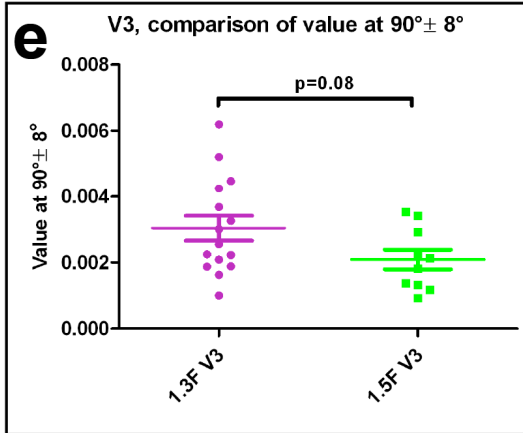
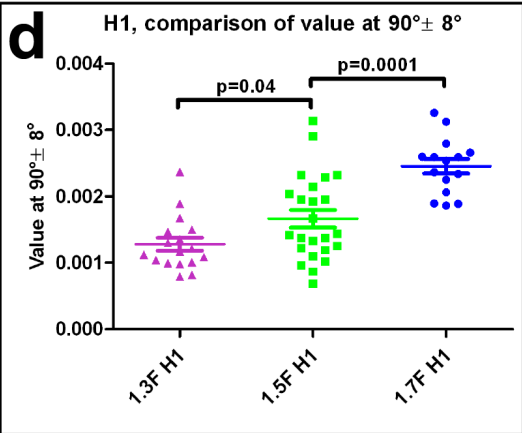
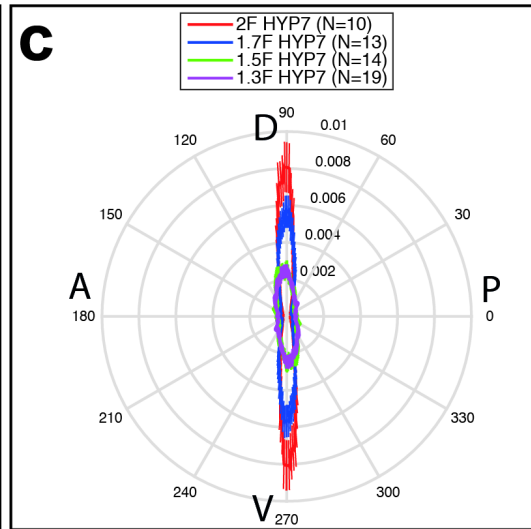
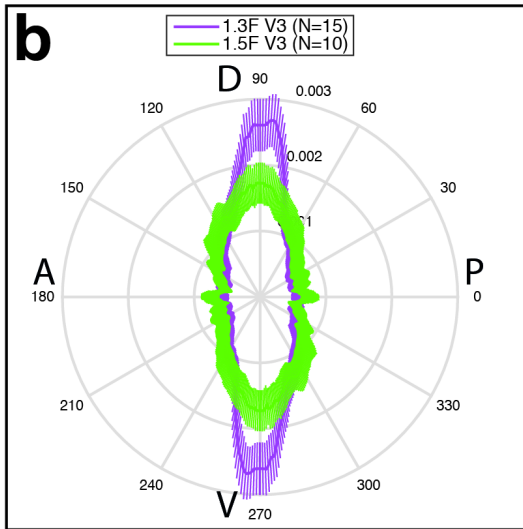
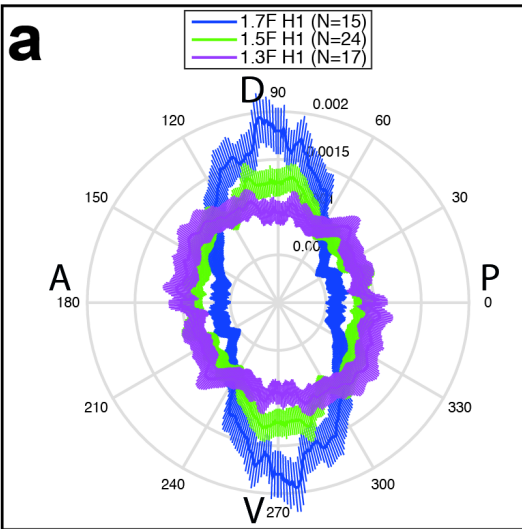


$$DV \text{ opening} = \frac{b_{DV}}{l} = 2 \frac{\sigma_{DV}}{E_0}$$

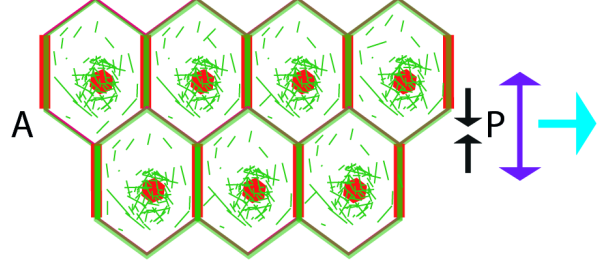


$$AP \text{ opening} = \frac{b_{AP}}{l} = \sqrt{2} \left(1 + \frac{\sqrt{1+K}}{1+2K}\right)^{\frac{1}{2}} \left(\frac{E_{AP}}{E_{DV}}\right)^{\frac{1}{4}} \frac{\sigma_{AP}}{E_{AP}}$$

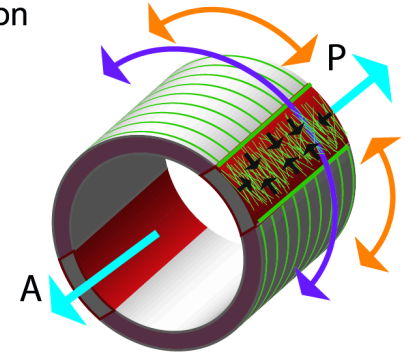
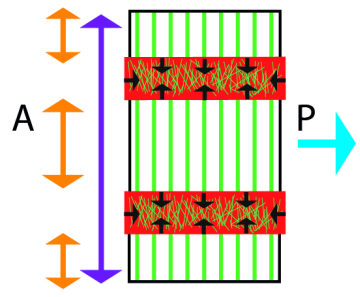




**g** *Drosophila* germband extension



*C. elegans* embryo elongation



- Myosin II      — Mechanical stress
- Actin            — Elongation direction
- ↔ Stress anisotropy
- ↔ Stiffness anisotropy

# Appendices: The interplay of stiffness and force anisotropies drive embryo elongation

Thanh TK Vuong-Brender, Martine Ben Amar, Julien Pontabry and Michel Labouesse

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## 1 Appendix 1: An overview of the early phase of *C. elegans* embryonic elongation

Soon after the ventral enclosure has completed, *C. elegans* embryos elongate from a lima-bean shape to the characteristic cylindrical shape, resulting in a 4-fold increase in length and approximately a 2-fold decrease in diameter<sup>1</sup> (figure 1a). The elongation is thought to be driven by cell shape changes, as can be most easily observed among seam cells (figure 1a). Muscle contractions, starting about midway through the process, are essential, since muscle-defective embryos are paralyzed and arrest at the 2F stage. In the following paragraphs, we discuss mostly the early phase of elongation which occurs prior to muscle contraction onset.

Pharmacological and genetic studies have proved the critical role of actomyosin contractility during the early elongation. Inhibition of actin polymerization with cytochalasin-D prior to the 1.5F stage blocks elongation, whereas application at later stage causes embryos to retract to their pre-elongation state<sup>1</sup>. Non-muscle myosin II activation is regulated through phosphorylation and dephosphorylation of the regulatory light chain MLC-4 by the LET-502/Rho-binding kinase and MEL-11/Myosin phosphatase, respectively<sup>2,3,4</sup>. LET-502, the effector of the Rho GTPase RHO-1, can be activated and inactivated by the *C. elegans* RhoGEF (Guanine Exchange Factors) RHGF-2, and RhoGAP (GTPase Activating Protein) RGA-2<sup>5,6</sup>, respectively. Mutations affecting myosin II or its activation, such as MLC-4 and NMY-1/Non-muscle Myosin II heavy chain, RHGF-2/RhoGEF, LET-502/ROCK, lead to hypo-elongation, whereby embryos arrest earlier than or at the 2F stage<sup>2,4,5,7,8</sup>. By contrast, mutations affecting negative regulators of myosin II, like *mel-11* or *rga-2*, cause embryos to burst during elongation due to increased tension exerted on adherens junctions<sup>3,6</sup>. The two myosin heavy chains, NMY-1 and NMY-2, work redundantly to regulate actomyosin contractility<sup>8</sup>. Although essential during early embryonic development, our data shows that NMY-2 is not required during elongation. Indeed, embryos homozygous for the strong thermosensitive mutant *nmy-2(ne3409)*<sup>9</sup> still elongated normally after shifting to the restrictive temperature (25.5° C, data not show).

Several lines of evidence suggest that seam cells generate most of the actomyosin forces, while the DV cells may remain passive. First, the myosin II regulatory light chain MLC-4 is mainly required in seam cells<sup>4</sup>. Second, MLC-4, MLC-5/myosin essential light chain and NMY-1 are higher expressed in seam cells<sup>4,7,8</sup>. Third, rescue experiments have shown that the positive regulator of contractility RHGF-2/RhoGEF is required only in seam cells, whereas the negative regulator RGA-2/RhoGAP acts specifically in DV cells<sup>5,6</sup>. Thus, all the players of acto-myosin regulation pathway promote a high contractility in seam cells and keep a low contractility in DV cells.

Although the myosin II activity is crucial in seam cells, the phenotypes of several mutants affecting junctional proteins, which are thought to anchor actin bundles<sup>10</sup>, show an important role of actin bundles in DV epidermal cells during elongation. In particular, zygotic *hmp-1/α-catenin* mutants, in which actin bundles detach from the junctional belt, show bulges and cannot elongate. Similarly, loss of ZOO-1/ZO-1 and VAB-9/Claudin homologues affects actin bundle organization in DV cells, leading to deformities and an incomplete elongation<sup>11,12</sup>.

In summary, the epidermal actomyosin network is essential for the early elongation phase of *C. elegans* embryo.

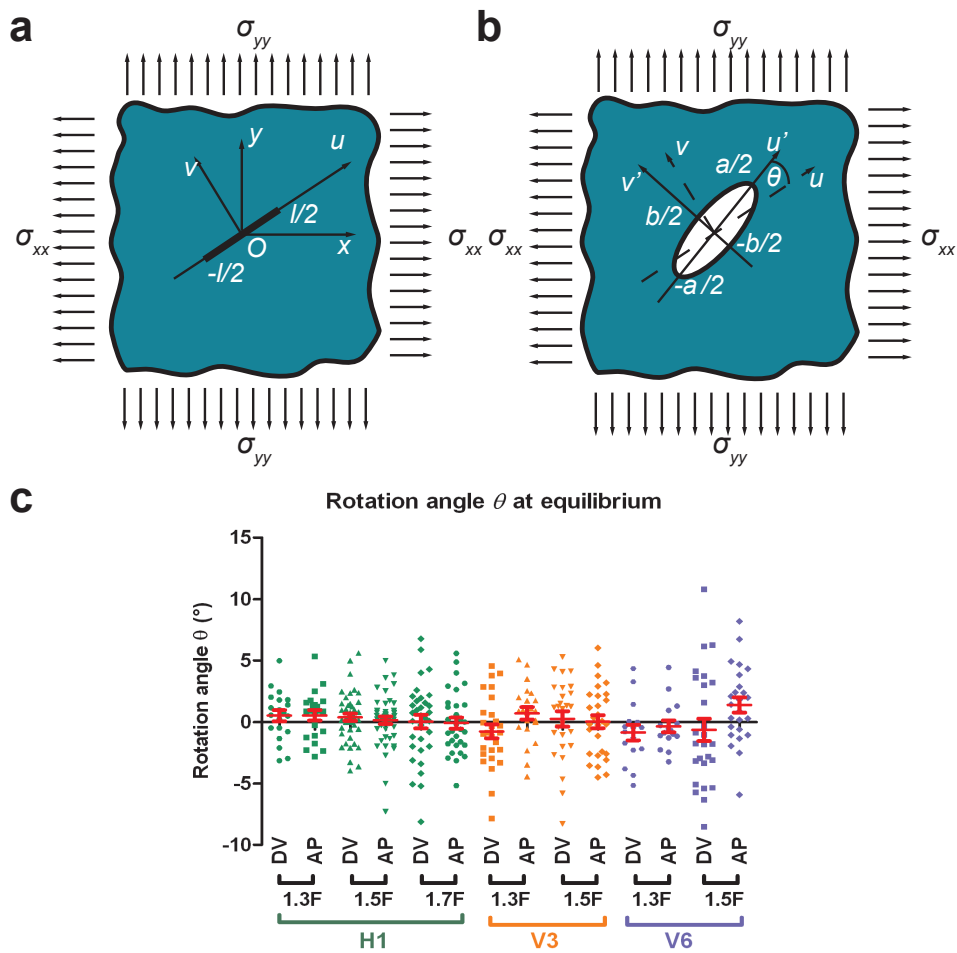
## 2 Appendix 2: The *C. elegans* embryonic epidermis is under biaxial stress loading along the AP and DV directions

Despite the fact that a biological material is in general viscoelastic<sup>13</sup>, the elastic aspect of *C. elegans* embryos seems to be more important. Indeed, inhibition of actin polymerization with cytochalasin-D induces a retraction of the embryo to nearly its original length<sup>1</sup>, like a spring after force release. Thus, we used an elastic model to describe *C. elegans* embryonic deformation. In particular, we considered the epidermal cell cortex as an elastic plane. In this section, we examine, for the analysis of laser ablation responses, whether the epidermal cell cortex is subjected to biaxial stress loading (stress along two orthogonal directions) along the AP and DV directions.



If a thin cut is introduced in an infinite isotropic elastic plane under biaxial stress loading (Appendix 2-figure1a), Theocaris *et al.*<sup>14</sup> have shown that the opening is an ellipse. The authors have shown that, the rotation angle  $\theta$  between the direction of the cut and the major axis of the opening ellipse, is in general different from zero (Appendix 2-figure1b).  $\theta$  is equal to zero when the cuts are parallel to the directions of stress loading or in case of equal tension-tension loading ( $\sigma_{xx} = \sigma_{yy} > 0$ )<sup>14</sup>.

To test if the AP and DV directions are indeed the directions of stress loading for different seam cells (H1, V3, V6, figure 3a) during early elongation, we performed laser cuts in the AP and DV directions and measured the rotation angle  $\theta$  at equilibrium from the 1.3F to the 1.7F stages. Appendix 2-figure 1c shows that  $\theta$  was not significantly different from zero for different magnitudes of stress (figure 3b), consistent with the hypothesis that AP and DV are the principal directions of stress loading. The difference of  $\theta$  compared to 0 was more important for V6, as we had difficulties to unambiguously determine the AP and DV direction for V6. In conclusion, the epidermal cell cortex can be considered as an elastic plane under biaxial stress loading along the AP and DV directions.



Appendix 2-figure 1: (a) An incision of length  $l$  was introduced in an elastic plane under biaxial stress along the  $x$  and  $y$  directions. The cut was along the  $u$  axis with the  $uv$  coordinates. (b) The shape of the opening at equilibrium was an ellipse<sup>14</sup>. We called  $u'$  (of  $u'v'$  coordinates) the axis parallel to the major axis of the opening ellipse, which formed an angle  $\theta$  to the direction of the initial cut. (c) The rotation angle  $\theta$  at equilibrium (measured around 9.5 s after cut) for H1 from the 1.3F to the 1.7F stages and V3 and V6 from the 1.3F to the 1.5F stages. 'DV' and 'AP' mean DV and AP opening, respectively. Red bars show the mean and standard error.

### 3 Appendix 3: Comparison of two methods to analyze laser ablation responses using the recoil dynamics and the cut opening at equilibrium

In order to compare the two analysis methods of laser ablation using the equilibrium shape of the cut opening or the recoil dynamics<sup>15;16</sup>, we estimated the initial recoil speed and the relaxation half-time by fitting the relaxation of the cut borders (the minor axis of the cut opening) with the equation:

$$y = y_0 + (Plateau - y_0)(1 - e^{-\gamma t}) \quad (1)$$

where  $y_0$  is the initial width of the cut opening,  $Plateau$  is the minor axis of the cut opening at equilibrium and  $\gamma$  is the relaxation rate. The recoil speed can be obtained by taking the derivation of the previous equation versus time:

$$v = \frac{dy}{dt} = (Plateau - y_0)\gamma e^{-\gamma t} \quad (2)$$

Thus, the initial recoil speed is:

$$v_0 = (Plateau - y_0)\gamma \quad (3)$$

The half-time, defined as the time interval needed to reach half of the distance between the initial opening and the  $Plateau$ , is given by:

$$\tau_{\frac{1}{2}} = \frac{\ln(2)}{\gamma} \quad (4)$$

According to the model described in Rauzi *et al.*, Smutny *et al.* and Mayer *et al.*<sup>15;16;17</sup>, the initial recoil speed is proportional to the ratio of the cortical stress  $\sigma$  to the viscosity of the cellular medium  $\eta$ :

$$v_0 \sim \frac{\sigma}{\eta} \quad (5)$$

and the relaxation half-time depends on the ratio of  $\eta$  to the cortex stiffness  $k$ :

$$\tau_{\frac{1}{2}} \sim \frac{\eta}{k} \quad (6)$$

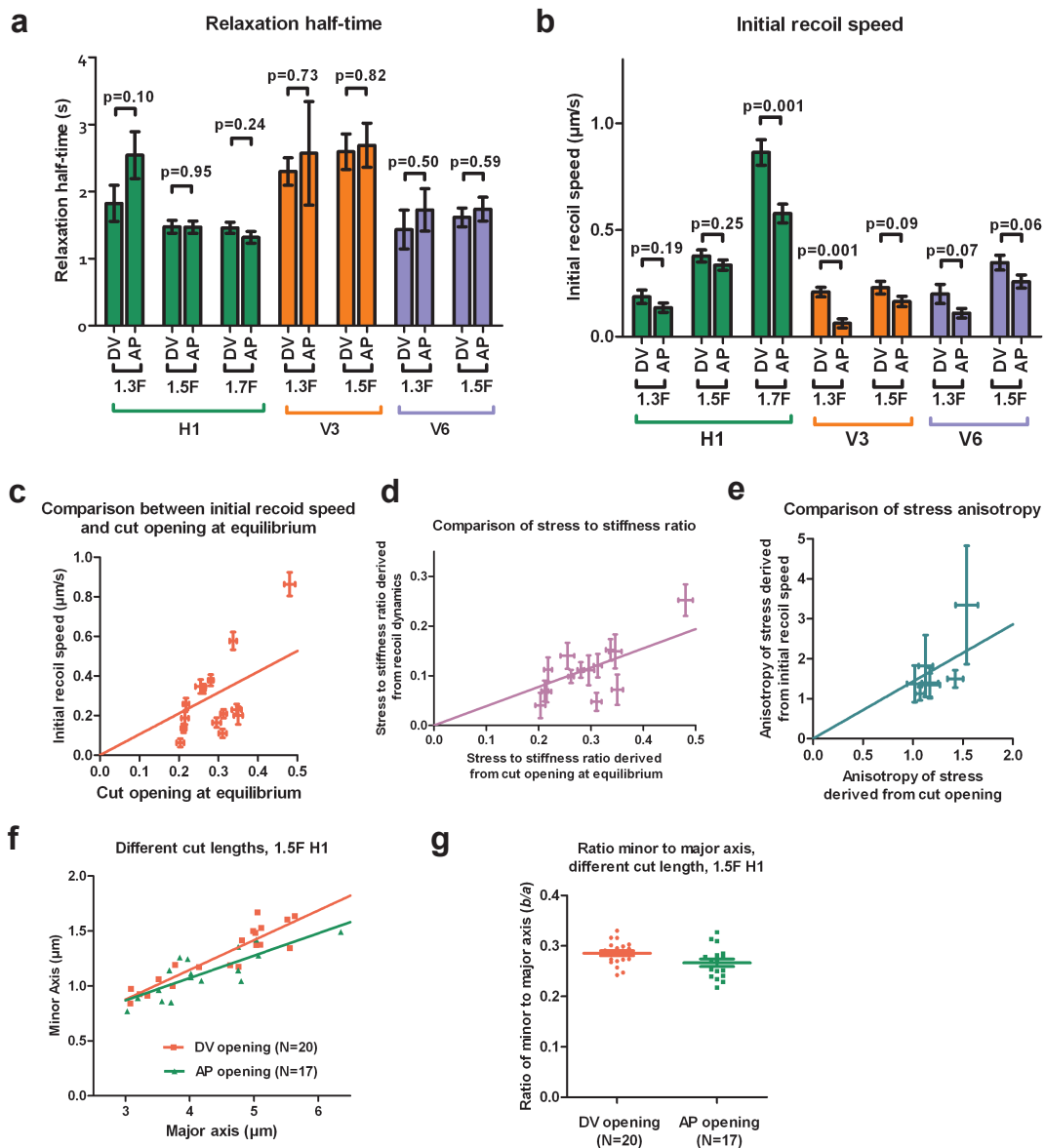
The fitting of the equation (1) depends on the initial width of the cut opening  $y_0$ . We used here three methods to estimate  $y_0$ . First, since the cut opening depends on the actomyosin contractility (figure 2c), we reasoned that  $y_0$  should be close to the opening observed in a mutant where actomyosin contractility is strongly inhibited. Indeed, in *let-502(sb118ts)* mutant, the opening changed very little (figure 2c), which corresponded to a nearly complete inhibition of the early elongation. The smallest opening in *let-502(sb118ts)* mutant was around  $0.6 \mu m$  (figure 2c). Second, our evaluation of the initial width of the opening using the same setup to photobleach a thin fluorescent layer also gave around  $0.6 \mu m$ . Finally, we tried to fit the recoil of cut borders using the equation (1) with values of  $y_0$  decreasing from  $0.6 \mu m$  to  $0.2 \mu m$  (with  $0.1 \mu m$  step) and found a decrease of goodness of fit  $R^2$  (supplementary table 2). The goodness of fit  $R^2$  is a fraction between 0 and 1 and higher values indicate better fits (GraphPad Prism 5, *Goodness of fit of nonlinear regression*). Thus, all three approaches indicated that  $0.6 \mu m$  was a good estimation of the initial width of the cut opening (an example of fit is shown in Figure 2-figure supplement 1). Subsequently, we used  $y_0 = 0.6 \mu m$  to derive the initial recoil speed and the relaxation half-time, and the results are shown in the supplementary table 3.

We found that the relaxation half-time in the AP direction was similar with the one in the DV direction in most of the seam cells examined at different stages (Appendix 3-figure 1a). Since the relaxation time is proportional to the ratio of viscosity over the stiffness of the cortex, and the cytoplasmic viscosity is likely homogeneous within a given cell, the cortex in seam cells is likely isotropic. However it can vary from one cell to another as indicated in the Appendix 3-figure 1a.

Next, we wanted to know if the two methods to analyze the ablation response (based on the cut recoil dynamics and the cut opening at equilibrium) gave consistent results about the stress

magnitude and anisotropy. Since we compared different seam cells (H1, V3, V6) with potentially different material properties (viscosity and stiffness), we multiplied the initial recoil speed by the relaxation half-time to have the stress to stiffness ratio (equations (5,6)). We normalized the previous ratio to the different cut lengths used in the different seam cells ( $5 \mu m$  in H1,  $4 \mu m$  in V3 and V6), then plotted the resulting values against the cut opening at equilibrium, which also reports the stress over stiffness ratio (figure 2b, Appendix 3-figure 1a,b,d). The linear regression strongly suggests that the two methods are in good agreement on the stress magnitude. Moreover, the anisotropy of stress obtained by the two methods showed a linear correlation (Appendix 3-figure 1e).

In summary, the two methods to analyze laser ablation responses gave results consistent with each other on the magnitude and the anisotropy of stress. The relaxation half-time obtained from recoil dynamics analysis indicated that the seam cell cortex is isotropic.



Appendix 3-figure 1: **(a)** Relaxation half-time and **(b)** initial recoil speed derived from fitting the cut border relaxation using an initial width of cut opening of  $0.6 \mu\text{m}$ , in H1, V3 and V6 from the 1.3F to the 1.7F stages. Z-test, ns,  $p > 0.05$ ; \*\*,  $0.001 < p < 0.01$ ; \*\*\*,  $p < 0.001$ . DV and AP indicate the directions of opening. **(c)** Comparison between the ratio of stress to stiffness derived using the cut recoil dynamics (after normalization to the cut length) and the one derived from the cut opening at equilibrium. **(d)** Comparison of stress to stiffness ratio measured from the recoil dynamics versus the one using cut opening at equilibrium. **(e)** Comparison of the anisotropy of stress (defined by DV/AP stress) derived from the recoil dynamics and from the cut opening at equilibrium. **(f)** The minor and major axes of cut opening at equilibrium show a linear relationship when the cut length varied from  $3 \mu\text{m}$  to  $6 \mu\text{m}$ . The ablations were performed in H1 at the 1.5F stage. Solid lines show a linear fit.  $R^2 > 0.65$ . **(g)** The ratio of the minor to major axis of the cut opening at equilibrium calculated from data shown in **(f)**.

## 4 Appendix 4: The ratio of the minor to major axis of the cut opening at equilibrium does not depend on the cut length

As predicted by Theocaris *et al.*<sup>14</sup>, the minor to major axis ratio of the cut opening should be independent of the cut length. Indeed, the equation for the major axis  $a$  over the cut length  $l$  of the opening at equilibrium is given as:

$$\frac{a}{l} = \frac{E + \sigma_{xx} - \sigma_{yy}}{E} \quad (7)$$

where  $\sigma_{xx}$  and  $\sigma_{yy}$  are the stresses in the principal loading directions  $x$  and  $y$ , respectively, which are also the laser cut directions.  $E$  is the Young modulus of the plane (Appendix 2-figure 1a-b). Thus, according to the ratio of minor axis to cut length given in figure 2b, the minor to major axis ratio of the ellipse at equilibrium is:

$$\frac{b}{a} = \frac{2\sigma_{yy}}{E + \sigma_{xx} - \sigma_{yy}} \quad (8)$$

and does not depend on the cut length  $l$ . Our experimental data fitted well with this prediction as shown in Appendix 3-figure 1f-g. Indeed, when we plotted the minor versus major axes of the opening at equilibrium, we observed a linear relationship (Appendix 3-figure 1f), when the cut length varied from 3 to 6  $\mu m$  in the seam cell H1, for cuts in both the AP and DV directions. Thus, the minor to major axis ratio is nearly a constant and independent of the cut length (Appendix 3-figure 1g), consistent with the theory of Theocaris *et al.*<sup>14</sup>.

## 5 Appendix 5: Modeling of the *C. elegans* embryo as a capped thin-wall pressured vessel and calculation of the anisotropy of stress

The rationale for modeling the *C. elegans* embryo as a thin-wall pressured vessel is given in the main text. To do so, first, we calculated the stress anisotropy for an axisymmetric vessel. We derived the stress anisotropy for the head from this calculation since the head was considered axisymmetric. Second, we calculated the stress anisotropy in the embryo body at the position of the seam cell V3. The body was not axisymmetric due to the important folding of the embryo in the eggshell, but the stress anisotropy can be obtained using a similar method.

### 5.1 Anisotropy of stress for an axisymmetric thin-wall pressured vessel

We consider an axisymmetric thin-wall pressured vessel with two ends capped. To make a parallel with *C. elegans* embryos, we call the axis of the vessel AP and the circumferential axis DV (Appendix 5-figure 1a). We calculate the anisotropy of stress  $\frac{\sigma_{DV}}{\sigma_{AP}}$ , where  $\sigma_{AP}$  is the longitudinal (AP) stress and  $\sigma_{DV}$  is the circumferential stress (DV) on the wall.

Let us consider a point Q on the wall for which we calculate  $\frac{\sigma_{DV}}{\sigma_{AP}}$  at this point. The tangent at Q in the plane going through Q and the AP axis makes an angle  $\alpha$  with AP.

Imagine that we cut the vessel in two parts by a plane going through Q and perpendicular to the AP axis (Appendix 5-figure 1a). The vessel is divided into  $\Omega_1$  and  $\Omega_2$ . The forces applied by  $\Omega_2$  on  $\Omega_1$  have two components  $F_1$  and  $F_2$  (Appendix 5-figure1b):  $F_1$  is the force applied by the wall of  $\Omega_2$  on  $\Omega_1$ , whereas  $F_2$  is the force exerted by the hydrostatic pressure from  $\Omega_2$ . We have

$$F_1 = \sigma_{AP}2\pi Rh \quad (9)$$

where  $R$  is the radius of the vessel at Q;  $h$  is the thickness of the wall (epidermis), considered as a constant.  $F_{1AP}$  is the component in the AP direction of  $F_1$  and is written as:

$$F_{1AP} = F_1 \cos\alpha = \sigma_{AP}2\pi Rh \cos\alpha \quad (10)$$

$F_{1AP}$  is balanced by the hydrostatic force  $F_2$  because  $\Omega_1$  is at equilibrium

$$F_2 = p\pi R^2 \quad (11)$$

where  $p$  is the hydrostatic pressure. By combining the equations (10,11), we have:

$$\sigma_{AP}2\pi Rh\cos\alpha = p\pi R^2 \quad (12)$$

$$\Rightarrow \sigma_{AP} = \frac{pR}{2h\cos\alpha} \quad (13)$$

Now let's consider a volume element  $\Omega_4$  of length  $\Delta l$  with two limiting sections perpendicular to the AP axis, so that  $\Omega_4$  is between  $\Omega_3$  and  $\Omega_5$  (Appendix 5-figure 1c). Imagine that we cut  $\Omega_4$  into two halves  $\Omega_{41}$  and  $\Omega_{42}$  (Appendix 5-figure 1d-e).

Let's examine an element  $\Delta s$  on the wall of  $\Omega_{41}$ , at the interface between  $\Omega_3$  and  $\Omega_{41}$  (Appendix 5-figure 1f). The force applied by the wall of  $\Omega_3$  to this element is:

$$F_3 = \sigma_{AP}(R_3)h\Delta s = \sigma_{AP}(R_3)hR_3d\beta \quad (14)$$

where  $\sigma_{AP}(R_3)$  means that  $\sigma_{AP}$  is a function of  $R_3$ ,  $\beta$  is the angle formed by the position of  $\Delta s$  with the  $\Omega_{41}$ - $\Omega_{42}$  interface as shown in Appendix 5-figure 1f. The force applied by  $\Omega_3$  on  $\Omega_{41}$  in the radial direction (Appendix 5-figure 1e-f) is:

$$F_{3R} = F_3\sin\alpha(R_3) = \sigma_{AP}(R_3)hR_3d\beta\sin\alpha(R_3) \quad (15)$$

where  $\alpha(R_3)$  means that  $\alpha$  is a function of  $R_3$ . The force applied by  $\Omega_3$  in the direction  $\vec{n}$  perpendicular to the  $\Omega_{41}$ - $\Omega_{42}$  interface (Appendix 5-figure 1f) is:

$$F_{3n} = \int_0^\pi F_{3R}\sin\beta d\beta = \int_0^\pi \sigma_{AP}(R_3)hR_3\sin\alpha(R_3)\sin\beta d\beta \quad (16)$$

$$= 2\sigma_{AP}(R_3)hR_3\sin\alpha(R_3) \quad (17)$$

If we replace  $\sigma_{AP}$  obtained from the equation (13), we have:

$$F_{3n} = 2\sigma_{AP}(R_3)hR_3\sin\alpha(R_3) = \frac{pR_3^2\sin\alpha(R_3)}{\cos\alpha(R_3)} \quad (18)$$

The force  $F_{5n}$  exerted by  $\Omega_5$  to  $\Omega_3$  can be calculated in the same manner. We obtain:

$$F_{5n} = -\frac{pR_5^2\sin\alpha(R_5)}{\cos\alpha(R_5)} \quad (19)$$

The resulting force applied by  $\Omega_3$  and  $\Omega_5$  to  $\Omega_{41}$  in the  $\vec{n}$  direction can be expressed as :

$$F_{35n} = F_{3n} + F_{5n} = \frac{d}{dR}\left(\frac{pR^2\sin\alpha(R)}{\cos\alpha(R)}\right)\Delta R = 2pR\tan\alpha(R)\Delta R + \frac{pR^2}{\cos^2\alpha(R)}\frac{d\alpha}{dR}\Delta R \quad (20)$$

The force applied by  $\Omega_{42}$  to  $\Omega_{41}$  (Appendix 5-figure 1e-f) in the  $\vec{n}$  direction is :

$$F_{12} = p\Delta l\cos\alpha(R)2R - \sigma_{DV}2\Delta lh \quad (21)$$

Since  $\Omega_4$  is at equilibrium, we have :

$$F_{35n} + F_{12} = 0 \quad (22)$$

Note that

$$\Delta R = \Delta l\sin\alpha \quad (23)$$

From the equations (20, 21, 22, 23) we have

$$2pR\tan\alpha\Delta l\sin\alpha + \frac{pR^2}{\cos^2\alpha}\frac{d\alpha}{dR}\Delta l\sin\alpha + p\Delta l\cos\alpha 2R - \sigma_{DV}2\Delta lh = 0 \quad (24)$$

$$\Rightarrow \sigma_{DV} = \frac{pR \sin^2 \alpha}{h \cos \alpha} + \frac{pR \cos \alpha}{h} + \frac{pR^2 \sin \alpha}{2h \cos^2 \alpha} \frac{d\alpha}{dR} = \frac{pR}{h \cos \alpha} + \frac{pR^2 \sin \alpha}{2h \cos^2 \alpha} \frac{d\alpha}{dR} \quad (25)$$

Thus we obtain the anisotropy of stress on the wall:

$$AS = \frac{\sigma_{DV}}{\sigma_{AP}} = \frac{\frac{1}{\cos \alpha} + \frac{R \sin \alpha}{2 \cos^2 \alpha} \frac{d\alpha}{dR}}{\frac{1}{2 \cos \alpha}} = 2 + R \frac{d\alpha}{dR} \tan \alpha \quad (26)$$

We can now calculate the  $AS$  for vessels with a particular shape : a sphere, a cylinder and an ellipsoid.

- For a sphere of radius  $R_0$  :

$$R = R_0 \cos \alpha \quad (27)$$

According to the equation (13), the AP stress is:

$$\sigma_{AP} = \frac{pR_0}{2h} \quad (28)$$

If we take the derivation of the equation (27) with respect to  $R$ , we have

$$-R_0 \sin \alpha \frac{d\alpha}{dR} = 1 \quad (29)$$

$$\Rightarrow R_0 \cos \alpha \tan \alpha \frac{d\alpha}{dR} = -1 \quad (30)$$

$$\Rightarrow R \tan \alpha \frac{d\alpha}{dR} = -1 \quad (31)$$

Combing the previous equation with the equation (26), we have:

$$\Rightarrow AS = \frac{\sigma_{DV}}{\sigma_{AP}} = 1 \quad (32)$$

- For a cylinder :  $\alpha = \text{const} = 0$

$$AS = \frac{\sigma_{DV}}{\sigma_{AP}} = 2 \quad (33)$$

According to the equation (13), the AP stress is:

$$\sigma_{AP} = \frac{pR}{2h} \quad (34)$$

where  $R$  is the radius of the cylinder.

- For an ellipsoid with the major axis  $a_1$  and minor axis  $a_2$  (Appendix 5-figure 1g), we can write the coordinates of point Q in the plane going through Q and AP axis as:

$$\Rightarrow \begin{cases} x = a_1 \cos t \\ y = a_2 \sin t \end{cases} \quad (35)$$

thus

$$\tan \alpha = \frac{dy}{dx} = \frac{a_2 \cos t}{a_1 (-\sin t)} = -\frac{a_2}{a_1} \cot \alpha \quad (36)$$

$$\frac{d(\tan \alpha)}{dt} = (1 + \tan^2 \alpha) \frac{d\alpha}{dt} = \frac{a_2}{a_1} (1 + \cot^2 \alpha) \quad (37)$$

Thus

$$\frac{d\alpha}{dt} = \frac{\frac{a_2}{a_1}(1 + \cotan^2 t)}{1 + \frac{a_2^2}{a_1^2} \cotan^2 t} = \frac{a_1 a_2}{a_1^2 \sin^2 t + a_2^2 \cos^2 t} \quad (38)$$

Due to the symmetry of the system, we examine only  $t \in [0, \pi]$ . We have:

$$R = y = a_2 \sin t \quad (39)$$

$$\frac{d\alpha}{dR} = \frac{d\alpha}{dt} \frac{dt}{dR} = \frac{a_1 a_2}{(a_1^2 \sin^2 t + a_2^2 \cos^2 t) a_2 \cos t} = \frac{a_1}{(a_1^2 \sin^2 t + a_2^2 \cos^2 t) \cos t} \quad (40)$$

From equations (36,39,40) we have

$$R \frac{d\alpha}{dR} \tan \alpha = \frac{-a_2^2}{a_1^2 \sin^2 t + a_2^2 \cos^2 t} \quad (41)$$

Thus:

$$AS = \frac{\sigma_{DV}}{\sigma_{AP}} = 2 - \frac{a_2^2}{a_1^2 \sin^2 t + a_2^2 \cos^2 t} \quad (42)$$

For the middle of the ellipsoid,  $y = a_2$  and  $x = 0$ ,  $t = \frac{\pi}{2}$ , thus

$$AS = \frac{\sigma_{DV}}{\sigma_{AP}} = 2 - \left( \frac{a_2}{a_1} \right)^2 \quad (43)$$

According to the equation (13), the AP stress at the middle of the ellipsoid is:

$$\sigma_{AP} = \frac{pa_2}{2h} \quad (44)$$

Note that the radial stress on the wall is  $-p$ . Since the wall is thin, i.e  $h \ll R$ , we expect that the radial stress is much smaller than the AP and DV stress on the wall for a sphere, an ellipsoid or a cylinder.

## 5.2 Anisotropy of stress for the body seam cell V3 of *C. elegans* embryo

For the seam cell  $V_3$  at the 1.3F and 1.5F stages, there is an important curvature of the embryo in the ventral part (Appendix 5-figure 1h). We cut a part of the embryo going through  $V_3$  and the dorsal part of  $V_3$  (dash line, Appendix 5-figure 1h) and approximate that the resulting half-section as half a cylinder (Appendix 5-figure 1i).

The force equilibrium for this part of the embryo in the circumferential direction (Appendix 5-figure 1i) is written as:

$$\sigma_{DV}^{V3} 2hL = p2R_2L \quad (45)$$

(force generated by circumferential stress on the wall = force due to the hydrostatic pressure), where  $\sigma_{DV}^{V3}$  is the DV stress at  $V_3$ ,  $h$  is the thickness of the epidermis,  $R_2$  is the radius at  $V_3$ ,  $L$  is the length of the region considered. Thus:

$$\sigma_{DV}^{V3} = \frac{pR_2}{h} \quad (46)$$

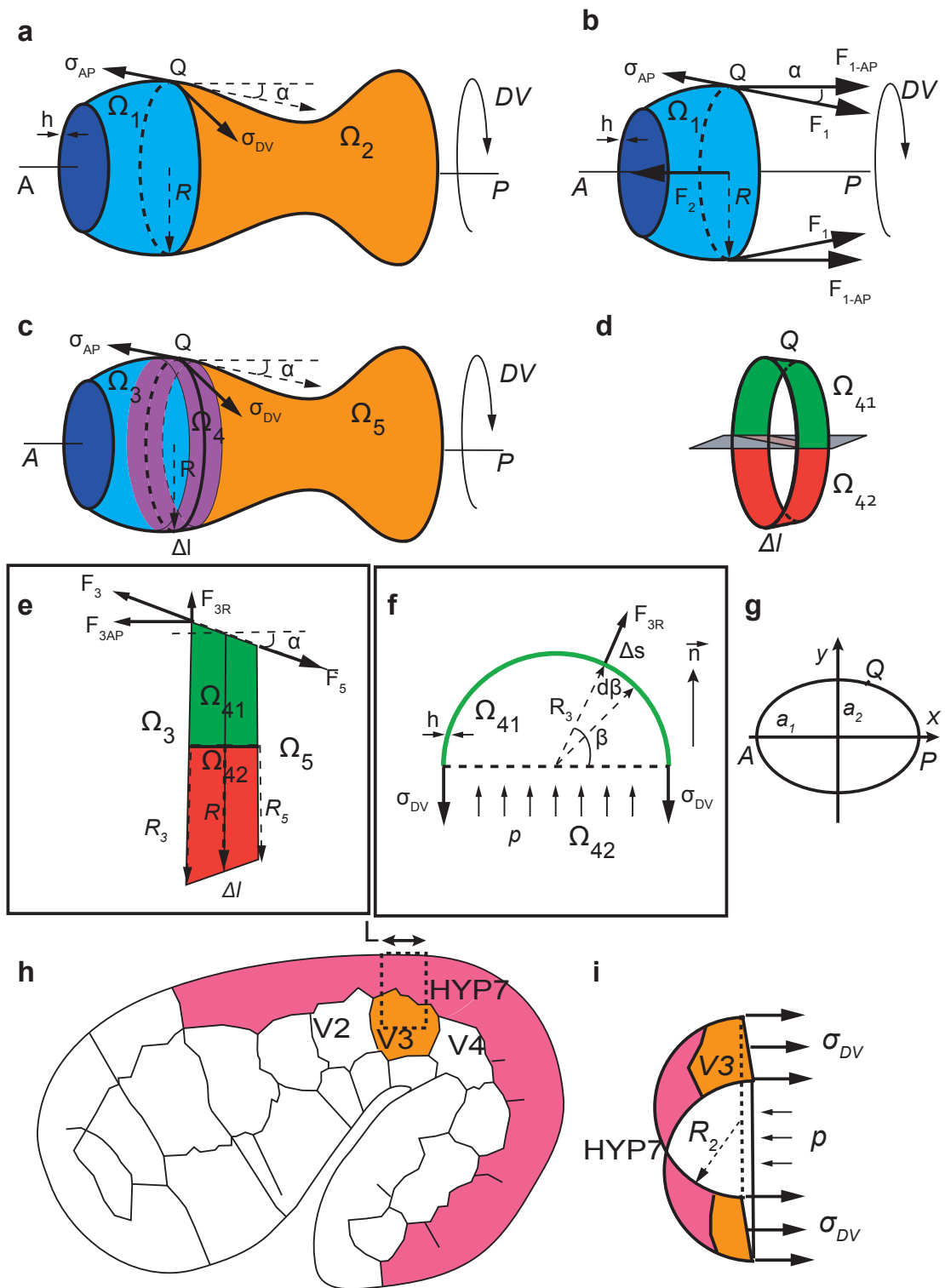
For H1 in the head, if we considered the head as a sphere, the DV and the AP stresses are the same and is given as (equations 28, 32):

$$\sigma_{DV}^{H1} = \sigma_{AP}^{H1} = \frac{pR_1}{2h} \quad (47)$$

where  $R_1$  is the head radius. If the AP stress is the same for H1 and  $V_3$ , then the anisotropy of stress at  $V_3$  is

$$AS = \frac{\sigma_{DV}^{V3}}{\sigma_{AP}^{H1}} = \frac{\frac{pR_2}{h}}{\frac{pR_1}{2h}} = 2 \frac{R_2}{R_1} \quad (48)$$





Appendix 5-figure 1

## 6 Appendix 6: The Hooke's law written for seam and DV epidermal cells

In this section, we further detail how we used the Hooke's law to describe the deformation of the seam and DV epidermal cells.

### 6.1 Appendix 6A: The Hooke's law written for seam cells

The Hooke's law for the deformation of seam cells is given by:

$$\epsilon_{AP}^s = \frac{\Delta L_{AP}^s}{L_{0AP}^s} = \frac{\sigma_{AP}^s}{E} - \nu \frac{\sigma_{DV}^s}{E} = -\frac{\sigma_{AP}^s}{E}(\nu AS - 1) \quad (49)$$

$$\epsilon_{DV}^s = \frac{\Delta L_{DV}^s}{L_{0DV}^s} = \frac{\sigma_{DV}^s}{E} - \nu \frac{\sigma_{AP}^s}{E} = -\frac{\sigma_{AP}^s}{E}(\nu - AS) \quad (50)$$

Here we supposed that the embryonic cortex material property is isotropic with a Young modulus  $E$ .  $\epsilon_{AP}^s$  and  $\epsilon_{DV}^s$  are the strain (which is equal to the relative length change) along the AP and DV directions, respectively;  $\Delta L_{AP}^s$  and  $\Delta L_{DV}^s$  are the length change,  $L_{0AP}^s$  and  $L_{0DV}^s$  are the initial length along the AP and DV directions, respectively. Positive values of  $\epsilon_{AP}^s$  or  $\epsilon_{DV}^s$  correspond to an increase in length (or extension), whereas negative values correspond to a decrease in length (shrinking).  $\sigma_{AP}^s$  and  $\sigma_{DV}^s$  are the stress along the AP and DV directions, respectively. Positive values of  $\sigma_{AP}^s$  or  $\sigma_{DV}^s$  correspond to tensile stress, whereas negative values correspond to contractile stress.  $\nu$  is the Poisson's ratio describing the shrinking in the AP direction when tensile stress is applied in the DV direction, and vice-versa. Here we omit the stress along the radial direction, since it is much smaller than the AP and DV stress for a thin-wall vessel (supplementary S15).  $AS$  is the stress anisotropy which equals to the DV to AP stress ratio.

$$AS = \frac{\sigma_{DV}^s}{\sigma_{AP}^s} \quad (51)$$

If we have an isotropic spherical embryo covered with contractile seam cells ( $AS = 1$ ), the embryo would not deform due to its incompressibility and symmetry:  $\epsilon_{AP}^s = 0$  and  $\epsilon_{DV}^s = 0$ . From this we derived that  $\nu = 1$ . We can thus rewrite the previous equations (49, 50) as:

$$\epsilon_{AP}^s = -\frac{\sigma_{AP}^s}{E}(AS - 1) \quad (52)$$

$$\epsilon_{DV}^s = -\frac{\sigma_{AP}^s}{E}(1 - AS) \quad (53)$$

### 6.2 Appendix 6B: The Hooke's law written for DV epidermal cells

As the DV epidermal cells have different stiffnesses along the AP and DV directions, the stress-strain relationship along the AP and DV axes can be written:

$$\epsilon_{AP} = \frac{\Delta L_{AP}}{L_{0AP}} = \frac{\sigma_{AP}}{E_{AP}} - \nu_1 \frac{\sigma_{DV}}{E_{DV}} \quad (54)$$

$$\epsilon_{DV} = \frac{\Delta L_{DV}}{L_{0DV}} = \frac{\sigma_{DV}}{E_{DV}} - \nu_2 \frac{\sigma_{AP}}{E_{AP}} \quad (55)$$

where  $\epsilon_{AP}$  and  $\epsilon_{DV}$  are the strain (which is equal to the relative length change) along the AP and DV axes, respectively;  $\Delta L_{AP}$  and  $\Delta L_{DV}$  are the length change,  $L_{0AP}$  and  $L_{0DV}$  are the initial length of the cell in the the AP and DV directions, respectively. Positive values of  $\epsilon_{AP}$  or  $\epsilon_{DV}$  correspond to an increase in length (or extension), whereas negative values correspond to a decrease in length (shrinking) of the cells.  $\sigma_{AP}$  and  $\sigma_{DV}$  are the stress along the AP and DV directions, respectively. Positive values of  $\sigma_{AP}$  or  $\sigma_{DV}$  correspond to tensile stress, whereas negative values correspond to contractile stress.  $\nu_1$  and  $\nu_2$  are Poisson's ratios.

Since the head is considered axisymmetric, we have:

$$AS = \frac{\sigma_{DV}^s}{\sigma_{AP}^s} = \frac{\sigma_{DV}}{\sigma_{AP}} \quad (56)$$

and replace

$$\omega = \frac{E_{DV}}{E_{AP}} \quad (57)$$

we have

$$\epsilon_{AP} = \frac{\sigma_{AP}}{E_{AP}} \left( 1 - \nu_1 \frac{AS}{\omega} \right) \quad (58)$$

$$\epsilon_{DV} = \frac{\sigma_{DV}}{E_{DV}} \left( 1 - \nu_2 \frac{\omega}{AS} \right) \quad (59)$$

While the *CB* model (see supplementary SI7, section 8.4) seems to be appropriate to describe fiber-reinforced material in extension, its application is questionable to describe the shrinking in the fiber direction. It is known for many fiber-reinforced composites to exhibit different stiffnesses in response to extension and compression<sup>18;19</sup>. It is also known for biological polymers, such as actin filaments, that they can have different mechanical properties under tensile or compressive stress, as actin filaments buckle under compression<sup>20</sup>. The use of Poisson's ratios for fiber-reinforced material in compression is arguable, since the symmetry of stiffness (or compliance) matrix may not be satisfied<sup>18</sup>. Given that the DV cells decrease their length in the fiber (DV) direction (figure 3e), the fibers should be under compression. For this reason, we suggest that the DV epidermal cells must have an active mechanism to adjust the actin bundle length to the cell shrinkage along the DV direction, in order to maintain the reinforcement properties.

## 7 Appendix 7: Cracks opening in orthotropic and fiber-reinforced planes

### 7.1 Introduction

This supplementary gives the proofs of the various relations used in the main paper to extract the residual stresses in the epithelial cells. Exhibiting oriented actin cables, the epithelium can be considered as a thin anisotropic soft layer. The correct description of the epithelium behaviour under strong deformations is achieved via hyper-elasticity. However, the determination of a crack-shape in nonlinear elastostatic remains challenging and is not achieved to the best of our knowledge. It is why we assume first a linear orthotropic planar material and solve the geometry of the crack by potential functions<sup>21;22</sup>, following the pioneering contributions of Muskhelishvili<sup>23</sup>, Suo<sup>24</sup>, Theocaris *et al.*<sup>14</sup> and Yoffe<sup>25</sup>. In the following, the shape of the crack is given, under simultaneous tension imposed far away along  $Ox$  and  $Oy$ , including also shear stresses (combined Mode *I* and *II* of fracture). Then, we present the model for fiber soft material, which is a better representation for living matter. Aiming to estimate residual stresses from the shape aperture, we identify the correspondence between the linear elastic coefficients of anisotropic elasticity and the parameters of a fiber model, at low strains.

### 7.2 Fracture in orthotropic linear elasticity

In material sciences, a common choice of elastic coefficients for orthotropic samples consists in the definition of Young moduli affected to each axis and Poisson ratios defined for each pair of orientation, in addition to shear moduli (equivalent to the second Lamé coefficients<sup>21</sup>  $\mu$ ). In case of plane stress elasticity, the equivalent Hooke's law is reduced to six independent coefficients:

$$\begin{cases} u_{xx} = \frac{1}{E_x}(\sigma_{xx} - \nu_{xy}\sigma_{yy}); & u_{yy} = -\frac{\nu_{xy}}{E_x}\sigma_{xx} + \frac{\sigma_{yy}}{E_y} \\ u_{zz} = -\frac{\nu_{xz}}{E_x}\sigma_{xx} + \frac{\nu_{yz}}{E_y}\sigma_{yy}; & u_{xy} = \frac{1}{2\mu_{xy}}\sigma_{xy} \end{cases} \quad (60)$$

The incompressibility condition:  $\Sigma_i u_{ii} = 0$  involves the third components of the deformation  $u_{zz}$ , leading to:  $\nu_{xz} = \nu_{yz} = 1$ , so finally 4 elastic independent coefficients are required. When the loads are applied at the border of the sample, the two in-plane components of the equilibrium condition  $\text{Div } \sigma = 0$ :

$$\frac{\partial \sigma_{xx}}{\partial x} + \frac{\partial \sigma_{xy}}{\partial y} = 0 \quad \text{and} \quad \frac{\partial \sigma_{xy}}{\partial x} + \frac{\partial \sigma_{yy}}{\partial y} = 0, \quad (61)$$

are automatically satisfied by the definition of the Airy potential<sup>21</sup>:  $U(x, y)$ :

$$\sigma_{xx} = \frac{\partial^2 U}{\partial y^2}; \quad \sigma_{xy} = -\frac{\partial^2 U}{\partial x \partial y}; \quad \sigma_{yy} = \frac{\partial^2 U}{\partial x^2} \quad (62)$$

Taking into account the Hooke's law, Eq.(60), one recovers the usual fourth order partial differential equation for  $U$ :

$$\frac{\partial^4 U}{\partial x^4} + 2\rho\Lambda^{1/2} \frac{\partial^4 U}{\partial x^2 \partial y^2} + \Lambda \frac{\partial^4 U}{\partial y^4} = 0 \quad (63)$$

where  $\Lambda = E_y/E_x$  and  $\rho = \frac{1}{2}\sqrt{\Lambda}(E_x/\mu_{xy} - 2\nu_{xy})$ . For isotropic materials,  $\Lambda = \rho = 1$ , since the second Lamé coefficient (related to the Young modulus  $E$  and to the Poisson ratio  $\nu$ ) reads:  $\mu = E/(2(1 + \nu))$ . In the isotropic case and in the context of fracture, Eq.(63) has been solved<sup>23</sup> with the help of holomorphic functions and complex analysis, in the case of plane-stress or plain strain elasticity and in Mode  $I$  (uniaxial loading, perpendicular to the crack direction). The method has been extended to bi-axial loading and arbitrary crack orientation by Theocaris *et al*<sup>14</sup> using the same strategy. Coming back to the anisotropic case and as pointed by Suo<sup>24</sup>, the theoretical analysis differs according to the  $\rho$  value. However, to the best of our knowledge, the shape of a crack of finite length has not been determined before, for an orthotropic material. Since we are concerned with  $\rho < 1$ , proofs will be given for  $-1 < \rho < 1$  and the results for the shape crack will be simply mentioned without demonstration for arbitrary  $\rho$ .

### 7.3 Definition of two complex functions

Eq.(63) is an even quartic partial differential equation which can be solved by 2 holomorphic functions  $\tilde{F}(z_1)$  and  $\tilde{G}(z_2)$  where:

$$z_1 = x + \Lambda^{-1/4}(m + In)y; \quad z_2 = \Lambda^{-1/4}(-m + In)y; \quad n = \sqrt{(1 + \rho)/2} \quad \text{and} \quad m = \sqrt{(1 - \rho)/2}. \quad (64)$$

This treatment is inspired from the work of Yoffe<sup>25</sup> for elasto-dynamic cracks in mode  $I$  and differs slightly from the work of Muskhelishvili<sup>23</sup> and Lekhnitskii<sup>26</sup>, more fancy but much less intuitive. In the following, the formulation via complex potentials can be checked at each step by elementary calculations. Each stress component  $\sigma_{ij}$  also verifies Eq.(63) and can be written as:

$$\sigma_{yy} = 2\text{Re}[F'(z_1) + G'(z_2)]; \quad \sigma_{xy} = -2\text{Re}[\mu_1 F'(z_1) + \mu_2 G'(z_2)]; \quad \sigma_{xx} = 2\text{Re}[\mu_1^2 F'(z_1) + \mu_2^2 G'(z_2)] \quad (65)$$

where  $F(z) = d\tilde{F}(z)/dz$ . The reader can check easily that the two components of the equilibrium equation (61) are verified. A standard choice for  $F'(z_1)$  and  $G'(z_2)$  for a crack lying on the x-axis between  $-a < x < a$  is:

$$F'(z_1) = (A_1 + iA_2) \frac{z_1}{\sqrt{z_1^2 - (l/2)^2}} + B_1 + iB_2 \quad \text{and} \quad G'(z_2) = (C_1 + iC_2) \frac{z_2}{\sqrt{z_2^2 - (l/2)^2}} + D_1 + iD_2 \quad (66)$$

where the 8 constants are real and will be determined by the boundary conditions on the crack lips and the loads far from the crack. On the lips, we have the cancellation of  $\sigma_{xy}$  and  $\sigma_{yy}$  and both square-roots in Eq.(66) are imaginary, so it reads:

$$\begin{cases} \sigma_{yy} = 0 \implies B_1 = -D_1 \quad \text{and} \quad C_2 = -A_2, \\ \sigma_{xy} = 0 \implies C_1 = (C_2 - A_2)m/n - A_1 \quad \text{and} \quad D_2 = (B_1 - D_1)m/n - B_2 \end{cases} \quad (67)$$

Eliminating the solid rotation at infinity leads to  $B_1 = -n(mA_2 + nA_1)$ , thus giving a fifth relationship. It remains to evaluate the 3 stress components at infinity  $\sigma_{yy}^\infty, \sigma_{xy}^\infty, \sigma_{xx}^\infty$  and we obtain:

$$A_1 = \frac{1}{4}(\sigma_{yy}^\infty - \Lambda^{1/4}\sigma_{xy}^\infty/m), \quad A_2 = -\frac{n}{4m}\sigma_{yy}^\infty, \quad B_2 = \frac{1}{8mn}(-\sqrt{\Lambda}\sigma_{xx}^\infty + \sigma_{yy}^\infty + 2mn^2\Lambda^{1/4}\sigma_{xy}^\infty) \quad (68)$$

Finally, taking into account the first relationship of the Hooke's law, Eq.(60), knowing that  $u_{xx} = \partial u/\partial x$ , one can find the horizontal displacement on the lips by quadrature :

$$u_{\pm} = \frac{2}{\sqrt{E_x E_y}} \left\{ \frac{1}{2}(\sigma_{xx}^\infty \Lambda^{1/2} - \sigma_{yy}^\infty)x \pm n\Lambda^{1/4}\sigma_{xy}^\infty \sqrt{(l/2)^2 - x^2} \right\} \quad (69)$$

The determination of the vertical displacement is more subtle since  $v$  comes from the shear relation:  $\partial v/\partial x = -\partial u/\partial y + \sigma_{xy}/\mu_{xy}$ , and we obtain after elimination of solid rotation,

$$v_{\pm} = \frac{2n}{E_x^{1/4} E_y^{3/4}} \left\{ n\Lambda^{1/4}\sigma_{xy}^\infty x \pm \sigma_{yy}^\infty \sqrt{(l/2)^2 - x^2} \right\} \quad (70)$$

In pure tensional loading,  $\sigma_{xy}^\infty = 0$ , calling  $\beta$  the ratio between the imposed vertical tension and the horizontal one:  $\beta = \sigma_{xx}/\sigma_{yy}$ , we obtain:

$$u_{\pm} = \Lambda^{1/2} \frac{\sigma_{yy}^\infty}{E_2} (\beta\Lambda^{1/2} - 1)x \quad \text{and} \quad v_{\pm} = \pm 2n\Lambda^{1/4} \frac{\sigma_{yy}^\infty}{E_y} \sqrt{(l/2)^2 - x^2} \quad (71)$$

Even if the demonstration given here is for  $\rho < 1$ , a slightly different treatment can be achieved for  $\rho = 1$  and  $\rho > 1$  but Eq.(69) and (70) and Eq.(71) remain valid with the same definition of  $n = \sqrt{(1+\rho)/2}$  and  $\Lambda$ .

Linear elasticity allows to solve exactly problems of interest in two dimensions, but it is not fully adapted to living matter which responds differently to low and high stresses. Indeed, in linear elasticity, the material answers linearly to the forcing while for living tissues, we know that large strains resist to the forcing mostly because of the fibers present in the tissue. Finite elasticity for soft materials is a fast-developing domain presently but it is technically more difficult. In particular, no exact crack solutions exist. Nevertheless, it presents a better description of the elastic energy: it is the reason why we present hereafter the fiber model in finite elasticity, which is probably more adapted to the epithelium we are considering.

## 7.4 Fiber model in finite elasticity

For a fibrous material, the elastic energy density  $W$  is chosen as the superposition of the energy of a gelatinous matrix  $W_{nH}$  (most often, the neo-Hookean or the Mooney-Rivlin model<sup>29</sup>) and a fiber contribution<sup>27;28;30</sup>. Different models exist, more or less complicated, based on the experimental responses of fibrous samples to stresses: muscles, arteries, and on required mathematical properties. We select a model which has the property to behave likely in tension or compression, that is the  $CB$  model<sup>27;28</sup>, contrary to other models such the  $G - O - H$  model<sup>30</sup> which gives a non symmetric answer in tension and compression. As shown in the Lecture<sup>31</sup>, the  $CB$  model eliminates unexpected singularities such as the one obtained at low strains in dispersion relations. Choosing a neo-Hookean matrix,

$$W_{nH} = \frac{\mu_0}{2} (\lambda_1^2 + \lambda_2^2 + \lambda_3^2 - 3) \quad (72)$$

where  $W_{nH}$  is function only of the first invariant  $I_1 = \text{tr}[\mathbf{F}\mathbf{F}^T]$ , ( $\mathbf{F}$  being the strain tensor, defined by  $F_{ij} = \partial u_i/\partial u_j$ ), contrary to the Mooney-Rivlin model which incorporates also the second invariant  $I_2$ . We choose the fiber contribution<sup>27;28</sup> as:

$$W_{CB} = \frac{\mu_0}{2} K \left\{ 2\kappa(\lambda_1^2 + \lambda_2^2 + \lambda_3^2 - 3) + (1 - 3\kappa)(\lambda_2^2 + \frac{1}{\lambda_2^2}) - 2 \right\} \quad (73)$$

where  $\kappa$  is a dispersion coefficient when the fibers are disordered<sup>28</sup> and  $K$  is directly connected to the elasticity of the fibers compared to the elasticity of the matrix. For simplicity, we put  $\kappa = 0$  hereafter. Since we cannot solve the crack problem for a nonlinear sample, our plan is to consider the low strain limit and relate the coefficients  $\Lambda$  and  $\rho$  which are responsible for the crack shape to the coefficients  $\mu_0$  and  $K$  of nonlinear elasticity. Considering plane-stress elasticity, minimization of the elastic energy concerns the free energy under the constrain of incompressibility and the condition that the Cauchy stress components  $\sigma_{iz}$  cancel.

$$G = \int dX_1 dX_2 dX_3 \{W(\lambda_1, \lambda_2, \lambda_3) - 3\} - P\lambda_3 J_{2D}, \quad (74)$$

where  $J_{2D}$  represents the Jacobian in 2D and  $P$  is a Lagrange multiplier. Minimization with respect to the strain in the third direction gives:

$$P = \frac{1}{J_{2D}} \frac{\partial W}{\partial \lambda_3} = \lambda_3 \frac{\partial W}{\partial \lambda_3} \quad (75)$$

## 7.5 Fiber versus orthotropic elasticity in extension

The correspondence between the anisotropic coefficients of the linear elasticity and the finite fiber elasticity is possible at small values of the strain such that  $\epsilon_i = |\lambda_i - 1| \ll 1$ . For plane-stress elasticity, the Cauchy stress leads to:

$$\sigma_i = \lambda_i \frac{\partial W}{\partial \lambda_i} - P = \lambda_i \frac{\partial W}{\partial \lambda_i} - \lambda_3 \frac{\partial W}{\partial \lambda_3} \quad (76)$$

Expanding all  $\lambda_i$  for weak deformations, we derive without difficulty, for the C-B model:

$$\epsilon_1 = \frac{1}{\mu_0} \frac{1+K}{3+4K} \left( \sigma_1 - \frac{1}{2(1+K)} \sigma_2 \right) \quad \text{and} \quad \epsilon_2 = \frac{1}{\mu_0} \frac{1}{3+4K} \left( \sigma_2 - \frac{1}{2} \sigma_1 \right) \quad (77)$$

Comparison with the orthotropic Hookean law, Eq.(60), in-plane stress elasticity gives:

$$E_x = \mu_0 \left( \frac{3+4K}{1+K} \right); \quad E_y = \mu_0(3+4K); \quad \Lambda = 1+K; \quad \text{and} \quad \nu_{xy} = \frac{1}{2(1+K)} \quad (78)$$

If we call the stiffness of the matrix (without fiber)  $E_0$ , we have

$$E_0 = 3\mu_0 \quad (79)$$

and

$$E_x = \frac{E_0}{3} \left( \frac{3+4K}{1+K} \right); \quad E_y = \frac{E_0}{3} (3+4K) \quad (80)$$

## 7.6 Fiber versus orthotropic elasticity in the case of shear deformation

To complete the set of coefficients, we need the shear coefficient  $\mu_{xy}$  so we treat a pure shear deformation such as: Then, the new coordinates in the current deformation are

$$x = X + \Gamma Y; \quad y = Y \quad z = Z \quad (81)$$

giving the deformation tensor  $\mathbf{F}$ , the left Cauchy -Green tensor<sup>29</sup>  $\mathbf{F}\mathbf{F}^T$  and the Cauchy stress tensor<sup>31</sup>  $\sigma$

$$\mathbf{F} = \begin{bmatrix} 1 & \Gamma & 0 \\ 0 & 1 & 0 \\ 0 & 0 & 1 \end{bmatrix}; \quad \mathbf{F}\mathbf{F}^T = \begin{bmatrix} 1+\Gamma^2 & \Gamma & 0 \\ \Gamma & 1 & 0 \\ 0 & 0 & 1 \end{bmatrix}; \quad \sigma = \begin{bmatrix} 1 & \Gamma\mu_0(1+2K) & 0 \\ \Gamma\mu_0(1+2K) & \Gamma\mu_0(1+2K) & 0 \\ 0 & 0 & 0 \end{bmatrix} \quad (82)$$

where  $\sigma$  is evaluated according to the following expression<sup>31</sup>

$$\sigma = \mu_0(\mathbf{F}\mathbf{F}^T - \mathbf{I}) + \mu_0 K \{ (\mathbf{F} \cdot \mathbf{M}) \otimes (\mathbf{F} \cdot \mathbf{M}) - (\mathbf{F}^{-T} \cdot \mathbf{M}) \otimes (\mathbf{F}^{-T} \cdot \mathbf{M}) \} \quad (83)$$

restricted to the linear approximation for weak value of the shear strain  $\Gamma$ . So the shear modulus  $\mu_{xy}$  of the orthotropic material is then:

$$\mu_{xy} = \mu_0(1 + 2K) = E_x \frac{1 + K}{3 + 4K} (1 + 2K) \quad (84)$$

which allows the calculation of the coefficient  $\rho$  introduced in Eq.(63):

$$\rho = \sqrt{\Lambda} \left( \frac{E_1}{2\mu_{xy}} - \nu_{xy} \right) = \frac{\sqrt{\Lambda}}{2(1 + K)} \left( \frac{3 + 4K}{1 + 2K} - 1 \right) = \frac{\sqrt{1 + K}}{1 + 2K} \quad (85)$$

## 7.7 How to evaluate the residual stress from the crack opening

When we cut a fibrous sample, perpendicularly to the direction of the fibers, we obviously change the structure locally and also the elastic properties along the crack. It is not sure that the *CB* model describes the correct elasticity since the new aperture is free from fibers. Perhaps a better approximation for the shape aperture is an isotropic elasticity. However, far from the crack of length  $l$ , on a distance larger than  $l$ , the stresses reach the value at infinity so  $\sigma_{yy}$  and the sample is fibrous. The *CB* model is an approximation but also the isotropic model and the truth is perhaps between these extremes. In addition, we consider an infinite sample in all directions  $Ox, Oy$  which implicitly assumes that the crack length is small compared to the epithelium size. The the opening ellipse, in the isotropic approximation is then:

$$b_y = 4 \frac{\sigma_{yy}}{3\mu_0} \sqrt{(l/2)^2 - x^2} \quad (86)$$

with a crack on the  $x$  axis having a length  $l$ . The minor axis of the opening is then:

$$b_y = 2 \frac{\sigma_{yy}}{3\mu_0} l = 2 \frac{\sigma_{yy}}{E_0} l \quad (87)$$

Considering now that we cut the sample along the  $Oy$  direction then the cut opening ellipse is

$$b_x = 4n\Lambda^{-1/4} \frac{\sigma_{xx}^\infty (1 + K)}{\mu_0 (3 + 4K)} \sqrt{(l/2)^2 - y^2} \quad (88)$$

where  $n$  is not modified and varies between 1 and 1/2 for increasing stiffness. The minor axis of the opening is:

$$b_x = 2n\Lambda^{-1/4} \frac{\sigma_{xx}^\infty (1 + K)}{\mu_0 (3 + 4K)} l = 2n\Lambda^{-1/4} \frac{\sigma_{xx}^\infty}{E_x} l \quad (89)$$

## 8 Appendix 8: Modeling the DV epidermis as an orthotropic material

As outlined in the main text, we could also have modeled the DV epidermis as an orthotropic material (with different stiffnesses in orthogonal directions). In this section, we argued that doing so is not compatible with the elongation of the embryo.

We modeled the DV epidermis as an orthotropic material with two principal Young moduli along the DV and AP directions  $E_{DV}$  and  $E_{AP}$ , respectively. We suppose that the stresses  $\sigma_{DV}$  and  $\sigma_{AP}$  are applied along the DV and AP directions, respectively, and there is no shear stress. The opening in the DV and AP directions is given by the equation (70):

$$\frac{b_{DV}}{l} = 2n \left( \frac{E_{DV}}{E_{AP}} \right)^{\frac{1}{4}} \left( \frac{\sigma_{DV}}{E_{DV}} \right) \quad (90)$$

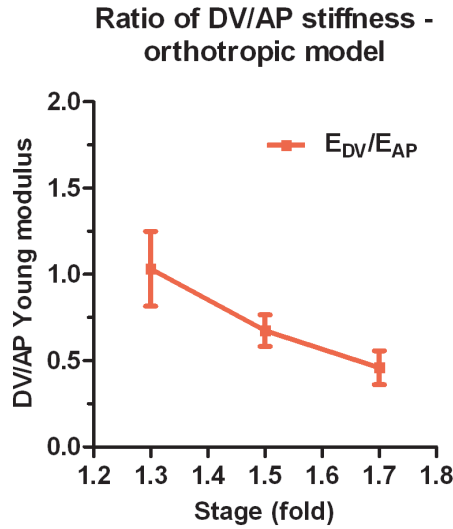
$$\frac{b_{AP}}{l} = 2n \left( \frac{E_{AP}}{E_{DV}} \right)^{\frac{1}{4}} \left( \frac{\sigma_{AP}}{E_{AP}} \right) \quad (91)$$

where  $b_{DV}$  and  $b_{AP}$  are the minor axis of the cut opening in the DV and AP directions, respectively;  $l$  is the initial cut length.  $n$  is the parameter given in the equation (64).

To obtain the DV/AP Young moduli ratio, if we divide the opening in the DV to the AP direction (same cut length), then we have :

$$\frac{b_{DV}}{b_{AP}} = \left( \frac{E_{AP}}{E_{DV}} \right)^{\frac{1}{2}} \left( \frac{\sigma_{DV}}{\sigma_{AP}} \right) = \left( \frac{E_{AP}}{E_{DV}} \right)^{\frac{1}{2}} AS \quad (92)$$

From the measurement of the AP and DV opening and given the same anisotropy of stress  $AS$  in the HYP7 cell as in the H1 cell, we can derive the ratio of DV/AP Young moduli  $\frac{E_{DV}}{E_{AP}}$ .



Appendix 8-figure 1: Ratio of DV/AP Young moduli calculated from the orthotropic model for the DV epidermis.

Appendix 8-figure 1 shows that the ratio of DV/AP Young moduli, calculated from the orthotropic model, decreased as the embryo elongated. This ratio became less than 1 after the 1.3F stage. Since the activity of myosin II in the DV epidermis is low, the DV cells are likely submitted to tensile stress from the seam cells. According to the Hooke's law written for DV cells (Appendix 6B) and given a positive tensile stress on the DV cells, a decrease in the DV/AP Young moduli ratio ( $\omega$ ) should decrease the AP length and increases the DV length. Thus, a decrease in the DV/AP Young moduli ratio as given by the orthotropic model would hinder the elongation of the DV cells in the AP direction and thus of the embryo as a whole, and be inconsistent with the contribution of the DV cells during *C. elegans* embryo elongation (figure 3e).

## 9 Appendix 9: Calculation of the ratio of Young moduli between the seam cell H1 and the head HYP7 cell matrix

To compare the material properties (Young modulus) between the seam cell H1 and the head HYP7 cell matrix (without fibers), we made use of the opening in the DV direction when performing laser cuts in these cells. The DV opening in the seam cell H1 is given as indicated in figure 2b:

$$\frac{b_{DV}^{H1}}{l} = 2 \frac{\sigma_{DV}^{H1}}{E} \quad (93)$$



where  $b_{DV}^{H1}$  is the minor axis of the cut opening at equilibrium,  $l$  is the cut length,  $\sigma_{DV}^{H1}$  is the DV stress in H1 and  $E$  is the Young modulus of H1. The head HYP7 cell behaved like an isotropic medium with a Young modulus  $E_0$  with cuts perpendicular to the actin fibers (DV opening). Thus, the DV opening in the head HYP7 cell is given as:

$$\frac{b_{DV}^{HYP7}}{l} = 2 \frac{\sigma_{DV}^{HYP7}}{E_0} \quad (94)$$

where  $b_{DV}^{HYP7}$  is the minor axis of the cut opening at equilibrium,  $l$  is the cut length,  $\sigma_{DV}^{HYP7}$  is the DV stress in the head HYP7 cell. Given their adjacent position (figure 3d), H1 and head HYP7 should be under the same DV stress:

$$\sigma_{DV}^{H1} = \sigma_{DV}^{HYP7} \quad (95)$$

Thus:

$$\frac{b_{DV}^{HYP7}}{l} = 2 \frac{\sigma_{DV}^{HYP7}}{E_0} = 2 \frac{\sigma_{DV}^{H1}}{E} \frac{E}{E_0} = \frac{b_{DV}^{H1}}{l} \frac{E}{E_0} \quad (96)$$

When we plotted the DV opening in the head HYP7 cell versus the DV opening in H1, the slope gives us the ratio of Young moduli  $\frac{E}{E_0}$ .

## 10 Appendix 10: Calculation of K and DV/AP Young moduli ratio for a fiber-reinforced material

For a fiber-reinforced material, from the equations (78, 85, 87, 89), we have:

$$\frac{b_{DV}}{b_{AP}} = \frac{(3 + 4K)\sqrt{2}}{3(1 + \frac{\sqrt{1+K}}{1+2K})^{\frac{1}{2}}(1+K)^{\frac{3}{4}}} \frac{\sigma_{DV}}{\sigma_{AP}} = \frac{(3 + 4K)\sqrt{2}}{3(1 + \frac{\sqrt{1+K}}{1+2K})^{\frac{1}{2}}(1+K)^{\frac{3}{4}}} AS \quad (97)$$

where  $b_{DV}$  and  $b_{AP}$  are the minor axis of the DV and AP openings (we used the same cut length  $l$ ), respectively; AS is the anisotropy of stress and  $K$  is the fiber contribution factor. Since we can measure the openings, given the anisotropy of stress, we can calculate  $K$ . We can easily derive the DV/AP Young moduli ratio according to the equation (78):

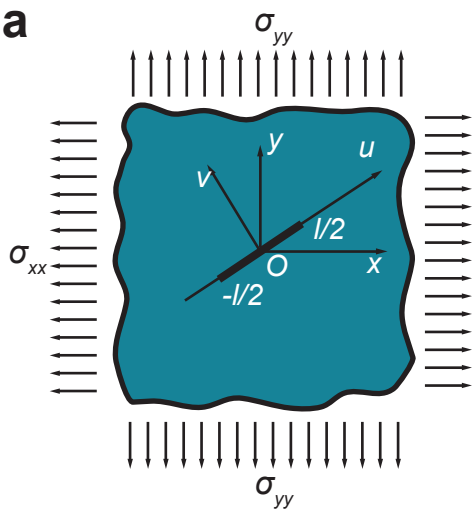
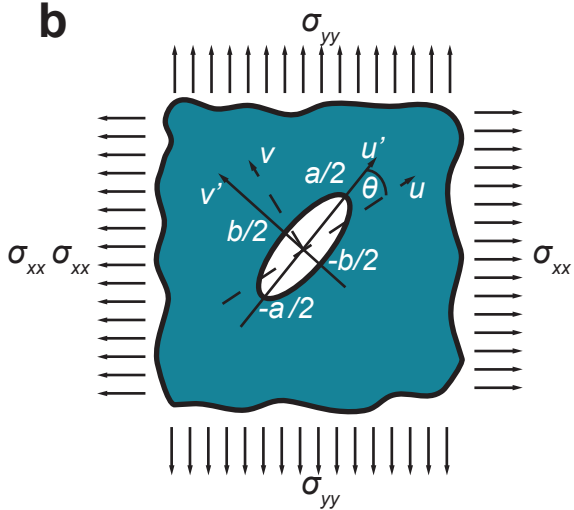
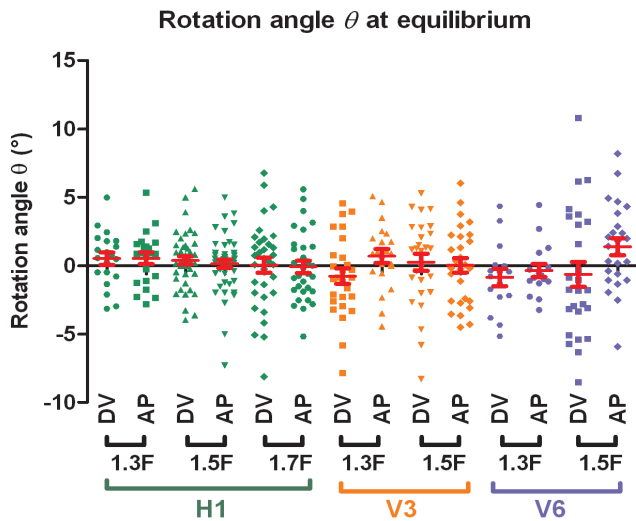
$$\omega = \frac{E_{DV}}{E_{AP}} = 1 + K \quad (98)$$

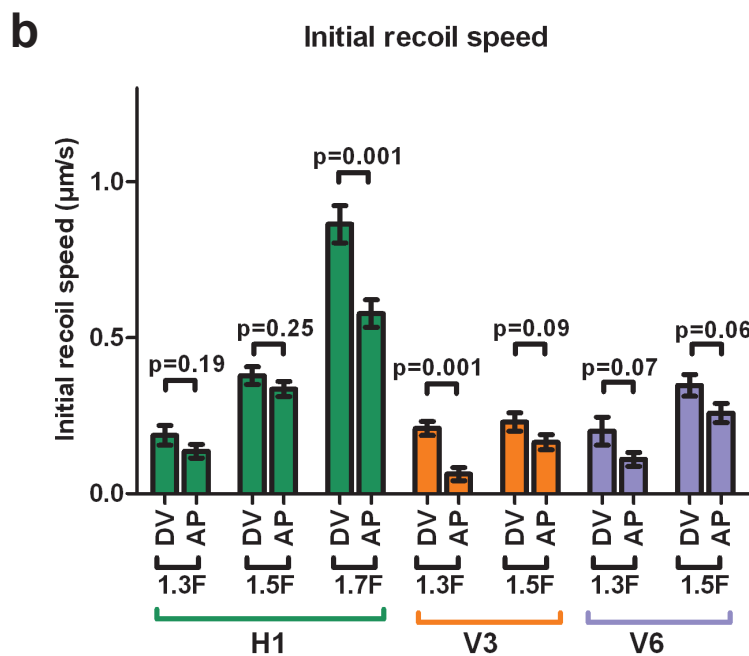
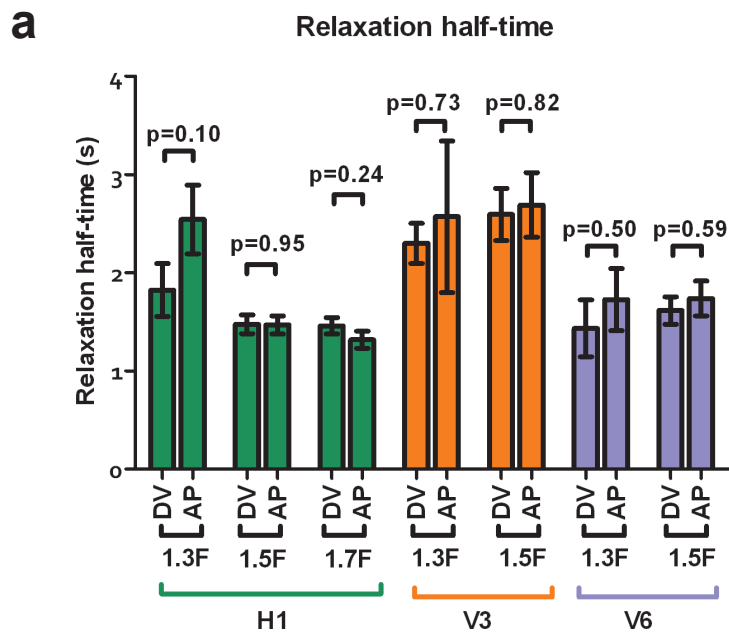
## References

- [1] Priess, J.R. & Hirsh, D.I. *Caenorhabditis elegans* morphogenesis: the role of the cytoskeleton in elongation of the embryo. *Developmental biology* **117**, 156-173 (1986).
- [2] Wissmann, A., Ingles, J., McGhee, J.D. & Mains, P.E. *Caenorhabditis elegans* LET-502 is related to Rho-binding kinases and human myotonic dystrophy kinase and interacts genetically with a homolog of the regulatory subunit of smooth muscle myosin phosphatase to affect cell shape. *Genes Dev* **11**, 409-422 (1997).
- [3] Wissmann, A., Ingles, J. & Mains, P.E. The *Caenorhabditis elegans* mel-11 myosin phosphatase regulatory subunit affects tissue contraction in the somatic gonad and the embryonic epidermis and genetically interacts with the Rac signaling pathway. *Developmental biology* **209**, 111-127 (1999).
- [4] Gally, C. et al. Myosin II regulation during *C. elegans* embryonic elongation: LET-502/ROCK, MRCK-1 and PAK-1, three kinases with different roles. *Development (Cambridge, England)* **136**, 3109-3119 (2009).

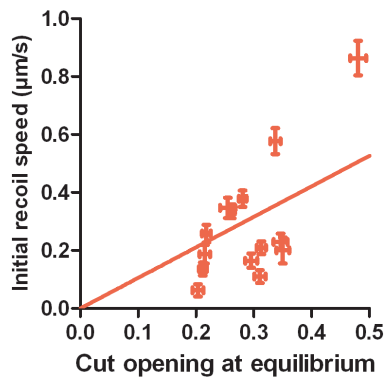
- [5] Chan, B.G., Rocheleau, S.K., Smit, R.B. & Mains, P.E. The Rho guanine exchange factor RHGF-2 acts through the Rho-binding kinase LET-502 to mediate embryonic elongation in *C. elegans*. *Developmental biology* (2015).
- [6] Diogon, M. et al. The RhoGAP RGA-2 and LET-502/ROCK achieve a balance of actomyosin-dependent forces in *C. elegans* epidermis to control morphogenesis. *Development* (Cambridge, England) **134**, 2469-2479 (2007).
- [7] Shelton, C.A., Carter, J.C., Ellis, G.C. & Bowerman, B. The nonmuscle myosin regulatory light chain gene *mlc-4* is required for cytokinesis, anterior-posterior polarity, and body morphology during *Caenorhabditis elegans* embryogenesis. *The Journal of cell biology* **146**, 439-451 (1999).
- [8] Piekny, A.J., Johnson, J.L., Cham, G.D. & Mains, P.E. The *Caenorhabditis elegans* nonmuscle myosin genes *nmy-1* and *nmy-2* function as redundant components of the let-502/Rho-binding kinase and *mel-11*/myosin phosphatase pathway during embryonic morphogenesis. *Development* (Cambridge, England) **130**, 5695-5704 (2003).
- [9] Liu, J., Maduzia, L.L., Shirayama, M. & Mello, C.C. NMY-2 maintains cellular asymmetry and cell boundaries, and promotes a SRC-dependent asymmetric cell division. *Developmental biology* **339**, 366-373 (2010).
- [10] Costa, M. et al. A putative catenin-cadherin system mediates morphogenesis of the *Caenorhabditis elegans* embryo. *The Journal of cell biology* **141**, 297-308 (1998).
- [11] Lockwood, C., Zaidel-Bar, R. & Hardin, J. The *C. elegans* zonula occludens ortholog cooperates with the cadherin complex to recruit actin during morphogenesis. *Current biology : CB* **18**, 1333-1337 (2008).
- [12] Simske, J.S. et al. The cell junction protein VAB-9 regulates adhesion and epidermal morphology in *C. elegans*. *Nature cell biology* **5**, 619-625 (2003).
- [13] Kasza, K.E. et al. The cell as a material. *Curr. Opin. Cell Biol.* **19**, 101-107 (2007).
- [14] Theocaris, P.S., Pazis, D. & Konstantellos, B.D. The Exact Shape of a Deformed Internal Slant Crack under Biaxial Loading. *Int J Fracture* **30**, 135-153 (1986).
- [15] Rauzi, M. & Lenne, P.F. Probing cell mechanics with subcellular laser dissection of actomyosin networks in the early developing *Drosophila* embryo. *Methods in molecular biology* (Clifton, N.J.) **1189**, 209-218 (2015).
- [16] Smutny, M., Behrndt, M., Campinho, P., Ruprecht, V. & Heisenberg, C.P. UV laser ablation to measure cell and tissue-generated forces in the zebrafish embryo in vivo and ex vivo. *Methods in molecular biology* (Clifton, N.J.) **1189**, 219-235 (2015).
- [17] Mayer, M., Depken, M., Bois, J.S., Julicher, F. & Grill, S.W. Anisotropies in cortical tension reveal the physical basis of polarizing cortical flows. *Nature* **467**, 617-621 (2010).
- [18] Bert, C.W. Models for Fibrous Composites with Different Properties in Tension and Compression. *J Eng Mater-T Asme* **99**, 344-349 (1977).
- [19] Jones, R.M. Stress-Strain Relations for Materials with Different Moduli in Tension and Compression. *Aiaa J* **15**, 16-23 (1977).
- [20] Murrell, M.P. & Gardel, M.L. F-actin buckling coordinates contractility and severing in a biomimetic actomyosin cortex. *Proceedings of the National Academy of Sciences of the United States of America* **109**, 20820-20825 (2012).
- [21] Landau, L.D. & Lifshitz, E.M. Theory of Elasticity, (Volume 7 of *A Course of Theoretical Physics*), Pergamon Press (1970).
- [22] Freund, L.B. Dynamic Fracture Mechanics, *Cambridge Monographs on Mechanics*(1990).

- [23] Muskhelishvili, N.I Some basic Problems of the Mathematical Theory of Elasticity. *Noordhoff International Publishing*, Leyden (1975).
- [24] Suo, Z. Singularities, interfaces and cracks. *Proc. R. Soc. London, Ser. A* **427**, 331-358 (1990).
- [25] Yoffe, E.H. The moving Griffith crack. *Philo. Mag.***42**, 739-50(1951).
- [26] Lekhnitskii, S.G. Theory of elasticity of an anisotropic body, San Francisco: Holden day(1963).
- [27] Wu ,M. & Ben Amar, M. Modelling fibers in growing discs of soft tissues. *Journ. Math. And Mech. of solids* **39** (2): 219-224 (2014)
- [28] Ben Amar, M. Wu, M. Trejo, M. & Atlan, M. Morpho-elasticity of inflammatory fibrosis: the case of capsular contracture. *Journ. Roy. Soc. Interface* **12** 20150343 (2015)
- [29] Ogden, R.W. Non-linear elastic deformations, *Dover Publications and Ellis Horwood* (1984)
- [30] Gasser , T.R. Ogden, R.W. & G. Holzapfel, G. Hyperelastic modelling of arterial layers with distributed collagen fiber orientation. *J.R.Soc. Interface* **3**,15-35(2006).
- [31] Destrade, M. Incremental equations for soft fibrous materials, *Nonlinear Mechanics of Soft Fibrous Materials .CISM Lecture Notes*, R.W. Ogden & L. Dorfmann, Editors, Springer, **559** 233-267 (2015)

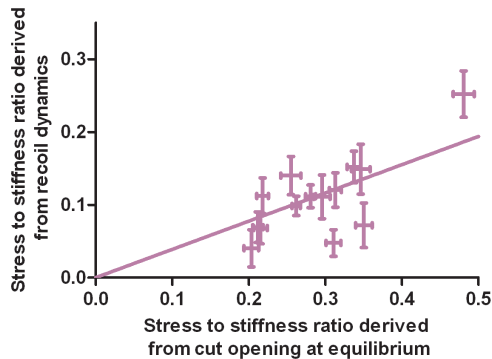
**a****b****c**



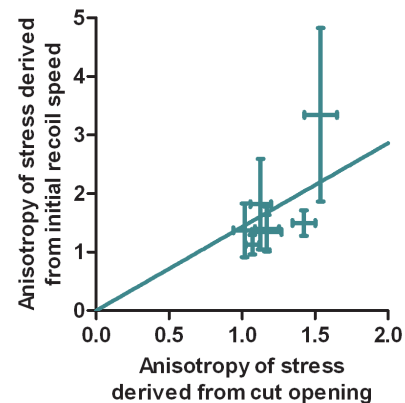
**c** Comparison between initial recoil speed and cut opening at equilibrium



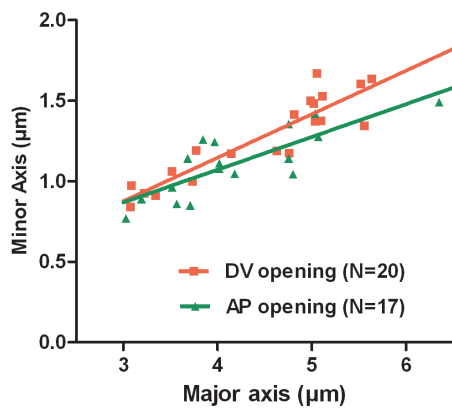
**d** Comparison of stress to stiffness ratio



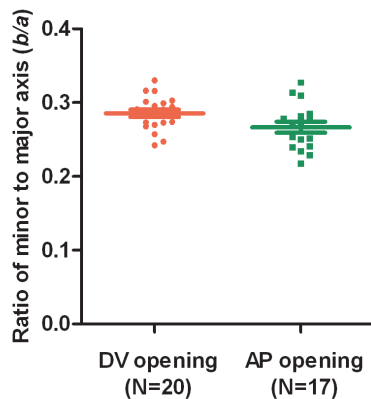
**e** Comparison of stress anisotropy

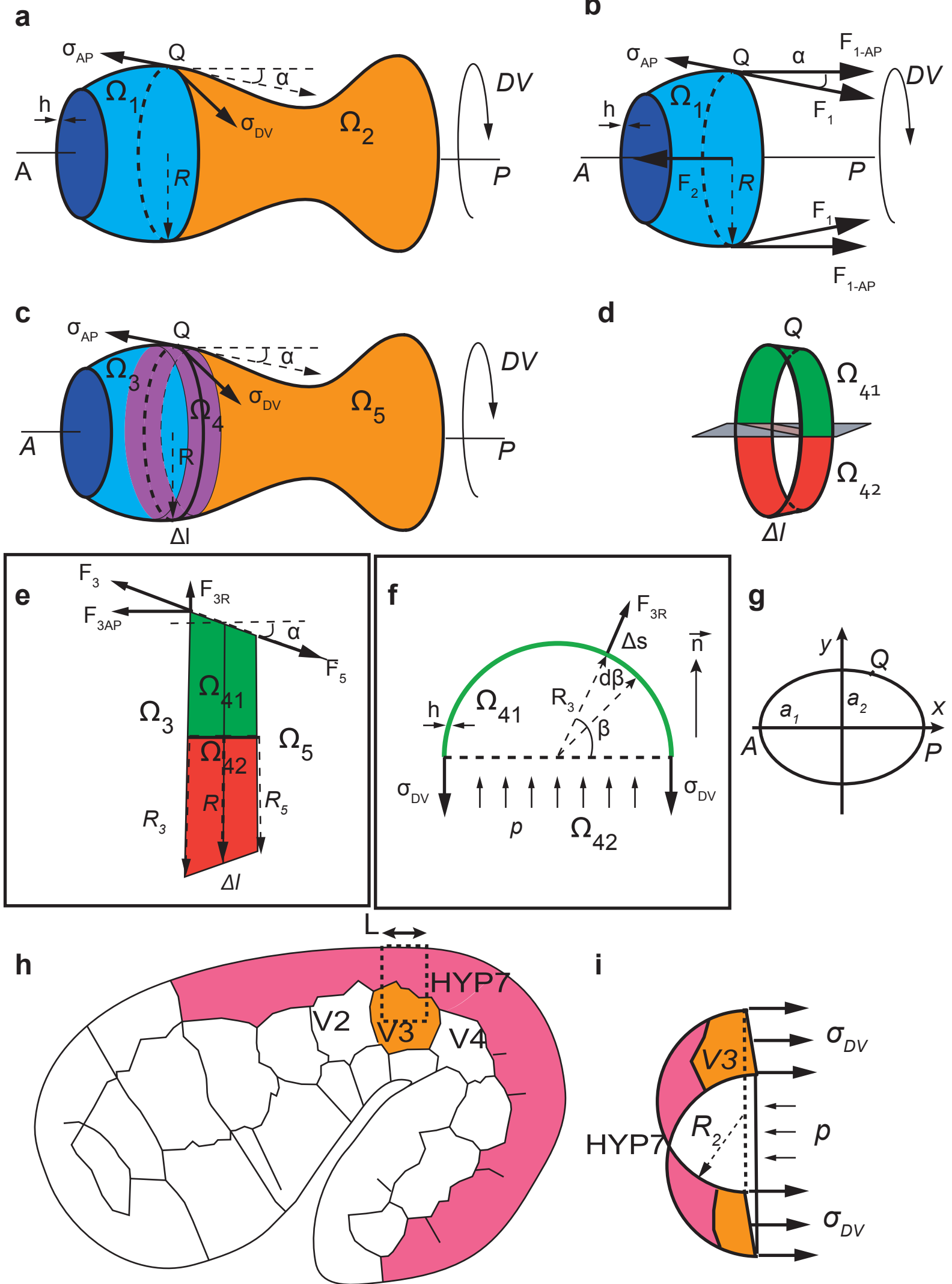


**f** Different cut lengths, 1.5F H1



**g** Ratio minor to major axis, different cut length, 1.5F H1





## Ratio of DV/AP stiffness - orthotropic model

