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# A cell penetrant manganese SOD-mimic is able to complement MnSOD and exerts an anti-inflammatory effect on cellular and animal models of inflammatory bowel diseases

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Additional material available

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## Supporting Information Placeholder

**ABSTRACT:** Inorganic complexes are increasingly used for biological and medicinal applications and the question of the cell-penetration and of the cell-distribution of metal-lodrugs is key to understand their biological activity. Oxidative stress is known to be involved in inflammation and in Inflammatory Bowel Diseases for which antioxidative defenses are weakened. We report here the study of a Mn-complex **Mn1** mimicking superoxide dismutase, a protein involved in the cell protection against oxidative stress, using an approach in *inorganic cellular chemistry* combining investigation of **Mn1** intracellular speciation using mass spectrometry, of its quantification and distribution using electron paramagnetic resonance and spatially-resolved X-ray fluorescence with evaluation of its biological activity. More precisely, we have looked for and find the MS-signature of **Mn1** in cell lysates and quantified the overall Mn-content. Intestinal epithelial cells activated by bacterial lipopolysaccharide were taken as a cellular model of oxidative stress and inflammation. **Mn1** exerts an intracellular anti-inflammatory activity, remains at least partially coordinated, with a diffuse distribution over the whole cell and functionally complements mitochondrial MnSOD.

## INTRODUCTION

Exploring inorganic compounds in biological or cellular environments requires translating our knowledge from the round-bottom flasks to the cells. Cellular uptake, location inside cells, stability in intricate biological environments, are key features determining bio-activity that have to be studied.<sup>1, 2</sup> Here, we apply a general approach combining evaluations of the biological activity in a cellular model,

with the exploration of the speciation, quantification of the intracellular content and cellular location in the case of the study of a Mn-complex mimicking the activity of the superoxide dismutase (SOD).<sup>2</sup> Correlating the intrinsic activity and the bio-activity in a cellular context, with the actual intracellular concentration, distribution and speciation, is important to decipher the relevant parameters controlling the activity in a biological context. This is surely one of the bases for an inorganic cellular chemistry approach, also more generally called inorganic chemical biology.<sup>1-3</sup> Bio-activities are usually reported against incubation concentrations, but they should also be analyzed as a function of the actual intracellular parameters.<sup>4</sup> Indeed, an intrinsic activity may remain silent if the compound does not enter the cell, or can be finely modulated by the concentration and also by the location inside cells.<sup>5</sup> This is why we clearly need to go beyond the mere observation of macroscopic effects on cell-cultures or biological tissues. To that extent, intracellular quantification and imaging —at the single cell level or within tissues substructures— are key experiments to provide a full understanding of bio-activity.<sup>6</sup>

Superoxide dismutases (SODs) are metalloenzymes that catalyse the dismutation of superoxide ( $O_2^{\cdot-}$ ), a byproduct of respiratory metabolism in living aerobic systems. These proteins are part of the cellular antioxidant protection controlling reactive oxygen species (ROS), including superoxide. Oxidative stress is observed when the balance between ROS production and protective pathways is broken and occurs in a wide range of pathophysiological disorders — aging, neuro-degenerative diseases, chronic inflammation, etc.<sup>7</sup> Oxidative stress and inflammatory bowel diseases

(IBD) have been linked, oxidative stress being thought to sustain and amplify the inflammation in a vicious circle.<sup>8-10</sup> Moreover, a weakening of the anti-oxidant defenses regulating the flow of ROS has been observed in IBD:<sup>11</sup> in epithelial cells from patients with IBD, the mitochondrial manganese SOD (MnSOD) is over-expressed but in an enzymatically inactive form and the cytosolic copper SOD (CuSOD) content decreases with inflammation.<sup>12</sup> Convergent data support anti-oxidant therapies for IBD management.<sup>8, 13-16</sup> For instance, 5-aminosalicylic acid (5-ASA), currently used as the first therapeutic line in ulcerative colitis, is thought to act in part via its anti-oxidant effect,<sup>11, 17, 18</sup> and purified CuSOD has been efficiently tested in murine colitis models.<sup>19</sup> However, the use of proteins for therapeutics is generally associated with major drawbacks, including low cellular penetration, short half-life, and immunogenicity.<sup>20</sup> These shortcomings can be overcome by using low molecular weight redox active complexes mimicking SOD activity.<sup>21-25</sup> Manganese complexes are particularly valuable in this context since manganese ion is known to be less toxic than copper and iron ions,<sup>7, 26</sup> the other cations encountered at the active site of SODs but which can be a source of additional oxidative stress if released. Indeed, to date, a large variety of Mn-derivatives have been reported for their ability to react with superoxide and mimic SOD activity:<sup>21, 23, 24, 27</sup> free Mn<sup>II</sup><sup>28, 29</sup> but also Mn coordinated to salen derivatives,<sup>30-32</sup> cyclic polyamines,<sup>20, 33-38</sup> tri- or dipode N-centered ligands,<sup>39-46</sup> 1,2-ethanediamine centered ligands,<sup>47-49</sup> desferrioxamine derivatives<sup>33, 50, 51</sup> polyaminocarboxylato<sup>52-54</sup> or polycarboxylato ligands,<sup>55</sup> peptides,<sup>56-58</sup> as well as Mn<sup>III</sup>-porphyrins,<sup>59-64</sup> phthalocyanines,<sup>65</sup> or biliverdine Mn-dinuclear complexes.<sup>66</sup> *In vivo* models have shown unambiguously the beneficial effect of these SOD mimics for the protection against radiation injuries,<sup>67-70</sup> and improvement of the therapeutic index in chemotherapy,<sup>71</sup> protection against chemical stress,<sup>71, 72</sup> ischemia reperfusion injury,<sup>73-76</sup> neuronal oxidative stress,<sup>77, 78</sup> endotoxic shock,<sup>79</sup> diabete,<sup>80</sup> or inflammation.<sup>81</sup> SOD-mimics have proven their efficiency to rescue SOD deficient cells,<sup>66, 80, 82-84</sup> cells under irradiation,<sup>85-87</sup> or chemical stress or infection,<sup>71, 72</sup> and in cells activated for ROS production such as macrophages<sup>88, 89</sup> or human leukemia cells.<sup>65</sup> SOD mimics represent also an appealing and promising approach in the field of IBD.<sup>15, 16</sup> Complex **Mn1** is a manganese SODm (Fig. 1A), bio-inspired from the active site of MnSOD and reproducing the coordination sphere of the metal center of this metalloenzyme,<sup>23</sup> that was previously described by our group. We reported its good intrinsic catalytic anti-superoxide activity which is associated with a cycling between two redox states Mn<sup>III</sup>/Mn<sup>II</sup>,<sup>90</sup> as well as its intracellular anti-superoxide effect in activated murine macrophages demonstrating its ability to efficiently reduce the superoxide flow in this cellular model.<sup>89</sup> We aim to go further and investigate the potential role of the SOD mimic **Mn1** in controlling gut inflammation. For this purpose, we have used an intestinal epithelial cell line able to activate inflammation pathways after lipopolysaccharide (LPS) challenge, known to induce oxidative stress,<sup>8, 13, 14, 91, 92</sup> developed in our laboratory and labelled HT29-MD2. Briefly, HT29-MD2 is a HT29 intestinal epithelial cell-line

stably transfected to over-express the protein MD2 conferring sensitivity to LPS, which is associated with an inflammatory response.<sup>93, 94</sup>

In the following, we combine in an inorganic cellular chemistry approach, characterization of **Mn1** — including metal-ligand stability constants, stability in growth medium, quantification of the intracellular content upon incubation, characterization in cell-lysates by mass spectrometry and imaging in cells using micro-X-fluorescence — with evaluations of the biological activity in cellular and animal models of IBD. We investigated the effect of **Mn1** on both the expression and activity of anti-oxidant protective enzymes (MnSOD and CuSOD) and inflammatory markers (IL8 and COX2). Additionally, an *in vivo* assay using a murine colitis model was set up to assess the efficiency of **Mn1**.

## EXPERIMENTAL SECTION

### Material

Information about chemicals is provided in the SI.

### Preparation of the complexes

Complex **Mn1** (**Zn1**) was prepared by addition of 1 eq. of MnCl<sub>2</sub> (ZnBr<sub>2</sub>) to a solution of **L**<sup>90</sup> in HEPES buffer (100 mM, pH 7.4) at 5 mM and then diluted in culture medium at the desired incubation concentration. See the structures in Fig. 1.

### Isothermal Calorimetric titration (ITC)

Dissociation constant of the manganese complex was determined by recording the heat of complexation with a Nano Isothermal Titration Calorimeter from TA Instruments. In a typical experiment, a ligand **L** solution (500 μM) in HEPES buffer (pH 7.4, 100 mM) containing NaCl (150 mM) in the calorimetric cell was titrated by a MnCl<sub>2</sub> solution (5 mM) (25 injections of 10 μL every 300 s under stirring at 25°C). Dilution heat was obtained in a separate measurement by injection of MnCl<sub>2</sub> into the buffer and was subtracted from the titration experiment. Thermograms were used to obtain a binding isotherm which was fitted by a one binding site L:M model using NanoAnalyze program provided by TA Instruments, to obtain the dissociation constant  $K_d$  (Fig. S1 A and B).

### UV-vis and electrochemistry

**Mn1** stability over time in culture medium was assessed by UV-visible spectroscopy and cyclic voltammetry. UV-visible spectra were recorded on a CARY-5 UV-vis spectrophotometer using a double beam mode with media as the reference. Spectra of **Mn1** (500 μM) in complete medium (DMEM, 10% FBS, 0.1% blasticidin). were recorded at 37 °C for 8 h in semi-micro quartz cuvette (0.2 cm path length). Cyclic voltammograms were recorded at 37 °C under argon atmosphere on an Autolab μAUTOLABIII/FRA2 (Metrohm) at 0.5 V.s<sup>-1</sup>. The auxiliary electrode was a Pt wire and the working electrode was a glassy carbon disk carefully polished before each voltammogram (diameter 3 mm). The reference electrode was a

SCE electrode. Cyclic voltammogram of **Zn1** was also recorded (see Fig. S1C).

### Cell culture

HT29-MD2 intestinal epithelial cells were used for all experiments. HT29 human colon adenocarcinoma were obtained from the European Collection of Cell Cultures (ECCC, Wiltshire, UK) and stably transfected to over-express MD2 as previously described.<sup>93</sup> Cells were cultured in DMEM supplemented with 10% of heat inactivated foetal bovine serum (FBS), 1% of penicillin-streptomycin and 0.1% of blasticidin (10 µg/mL) at 37 °C in a 5% CO<sub>2</sub>/air atmosphere.

### Intracellular quantification of Mn1 by EPR

**Mn1** was quantified by determining Mn<sup>2+</sup> total content in HT29-MD2 cells lysates using Electron Paramagnetic Resonance (EPR). Cells were cultured in a 75 cm<sup>2</sup> flask to reach 90% of confluency. They were incubated with media only, **Mn1** or MnCl<sub>2</sub> (100 µM) for 0.5 to 7 hours, at 37 °C. After a washing with NaCl 0.9%, a chaotropic shock was performed by adding a solution of NaCl 1 M. Cells were washed (EDTA 50 mM, 2 washings NaCl 0.9%). They were harvested by scraping and centrifuged at 4°C during 10 min at 900 rpm. Supernatant was removed, 100 µL of milliQ water was added and two freezing/thawing cycles in liquid nitrogen were performed. Protein content was determined for each sample (See SI). Cell lysates were acidified with HClO<sub>4</sub> 70 % (w/w). A calibration curve was established using MnCl<sub>2</sub> in water acidified with HClO<sub>4</sub> and the quantification of the Mn-content was performed using the two first lines in the six-line Mn<sup>II</sup> X-band EPR-spectrum and reported *per* mg of proteins (Fig. 2). EPR spectra were recorded on a Elexsys 500 spectrometer from Bruker in a glass capillary tube: frequency: 9.82 GHz, microwave power: 32 mW, amplitude of the modulation: 2G, 9 scans, room temperature. The same procedure was used to determine Mn-content in mitochondria enriched-fractions (see SI and Fig. S2).

### LC-MS/MS: Intracellular stability of Mn1

Intracellular stability of **Mn1** in cells was investigated using LC-MS/MS. LC-MS of pure compound **Mn1** (1 µM) in water showed two peaks corresponding to the free ligand and the expected complex (m/z 355.2 and 408.2 respectively), meaning that partial decoordination occurs under the LC-MS conditions. By MS-MS analysis, the ligand L and **Mn1** exhibited different fragmentation patterns (m/z 249.2, 167.2 and 302.2, 248.2 respectively) (Fig. S3), from which a clear identification of **Mn1** inside cells can be obtained. A lysate of cells incubated with **Mn1** (100 µM, 3 h) was prepared as explained for EPR experiments, without addition of perchloric acid. The lysate was diluted with a maximum volume of 0.5 mL of ultrapure water and filtered over membrane with a 3 kDa cutoff. The resulting filtrate and a solution of complex **Mn1** in water were analyzed by a QTRAP LC-MS/MS system Applied Biosystem (injection of 10 µL). The LC system consisted of a Dionex-LC Packings Ultimate Plus integrated micro-HPLC system. A Dionex Acclaim Pepmap100 column (150 mm Å~ 1 mm

i.d., C18, 3 µm particle size, 100 Å pore size) was used for the separation, with a gradient of 0–60% B in 75 min (solvent A = water–acetonitrile, 98:2, v/v, with 0.1% formic acid, solvent, B = acetonitrile–water, 98:2,v/v, with 0.1% formic acid) and a flow rate of 10 µL min<sup>-1</sup>. The column output was connected to an Applied Biosystems Qtrap LC-MS/MS system mass spectrometer through a NanoSpray ion-source interface (Spray voltage set to 2.4 kV, desolvating potential (DP) set to 40 V, collision energy (CE) set to 30 V). The LC-MS/MS system was controlled by Analyst 1.4.2 software (Applied Biosystems), allowing a 2.7 s cycle of 4 experiments for mass spectra acquisition: one full single MS by scanning the linear trap followed by three MS/MS experiments on the (pseudo-) molecular ions of the compounds. MS/MS optimization experiments on standard compounds were performed by direct injection on a 1 µL loop Rheodyne LC injection valve connected to a Turbolonspray source, at a 20 µL.min<sup>-1</sup> flow rate of solvent A (Ionspray voltage set to 5.5 kV, DP = 40 V, CE = 30 V).

### Synchrotron radiation X-ray fluorescence microscopy: Intracellular distribution of Mn1

The intracellular distribution of Mn in control cells and cells incubated with **Mn1** or MnCl<sub>2</sub> was determined using synchrotron radiation X-ray fluorescence microscopy. HT29-MD2 cells were seeded on silicon nitride windows (size: 1 mm x 1 mm, thickness: 500 nm) in 24-wells plate (75000 cells/well). After 36 h, cells were incubated with medium only (control), **Mn1** or MnCl<sub>2</sub> (100 µM) for 2 h. Cells were washed with NaCl 0.9% and a chaotropic shock was performed by adding a solution of NaCl (1 M). They were then washed once with EDTA (50 mM), and twice with NaCl 0.9%. Cells were cryofixed in liquid ethane and freeze-dried. Mappings of intracellular manganese (Mn), potassium (K), phosphate (P), and sulfur (S) (Fig. 3 and S4) were performed on 2-ID-D beamline of Advanced Photon Source synchrotron (Argonne National Laboratory, Chicago, USA). Cells were located using a phase-contrast optical microscope. All measurements were conducted at room temperature, under a He atmosphere using a 6.8 keV monochromatic X-ray incident beam focused to 0.2 µm diameter. The fluorescence signal was detected with an integration time of 2 s per pixel, with a 200 nm pixel size, at 90° to the incident beam using a Vortex EM single element silicon drift detector. Images analyses were performed by using MAPS software from APS. Due to the last washing of cells with NaCl 0.9 % before cryofixation and freeze-drying, some NaCl crystals were present on silicon nitride membranes. The strong signal of Cl was then subtracted from other element signals.

### Cell assays

Cells were seeded in 12 or 24 well-plates at 100 000 cells/well to reach 90% confluence after 3 or 4 days. Cells were incubated with **Mn1** and controls for 1 h (**Zn1**, MnCl<sub>2</sub>: 100 µM; bovine CuSOD 100 U/mL; 5-ASA: 10 mM, 1 mM, 100 µM). Then, LPS was added (0.1 µg/mL, 6 hours). Supernatants were collected, and stored at -20 °C

before ELISA and LDH assay. Cells were washed with NaCl 0.9%, lysed in PBS containing 1% triton X-100 and protease inhibitors cocktail. They were harvested by scraping, and stored at -20°C before western blot, SOD activity on gels, cytotoxicity and protein quantification experiments.

#### *Cytotoxicity assay*

Cytotoxicity of **Mn1** and controls, with and without LPS, was assessed using lactate dehydrogenase (LDH) release assay with a limit for non-cytotoxicity chosen at 10% (see SI).

#### *Protein Assay*

Protein concentrations were determined in cell lysates using BCA protein assay reagents and bovine serum albumin (BSA) as standard according to the manufacturer's instructions (see SI).

#### *Cell activation with LPS and incubation with the agents to be tested*

Cells were seeded in 12-well culture plates to reach 90% of confluency after 3 or 4 days. They were then incubated with the agents to be assayed (typically, depending on the experiment: complex **Mn1** (100 µM), its redox silent analogue (Zn<sup>II</sup> complex shown in Fig. 1A) **Zn1** (100 µM), MnCl<sub>2</sub> (100 µM) or bovine CuSOD (100 U/mL)) for 1 h and further incubated with LPS (0.1 µg/mL) for 6 more hours. Cell culture supernatants were then collected and stored at -20 °C before ELISA and LDH assay. Cells were washed with 0.9 % NaCl, lysed in PBS containing 1% triton X-100 and protease inhibitors cocktail. They were harvested by scraping, sonicated and stored at -20°C for western blot, SOD activity on gels, cytotoxicity and protein quantification experiments.

#### *SOD activity on gel*

SOD activity was determined by using the nitroblue tetrazolium (NBT) method.<sup>95</sup> A loading buffer containing 50 % of glycerol and 1 % of bromophenol blue solution in TRIS buffer (0.5 M, pH 6.8) was added to cell lysates (1/1 mixture). A non-denatured 10 % acrylamide gel was used and pre-electrophoresis (40 mA, 1 h, 4°C) was performed as described.<sup>95</sup> 100 µg of proteins were then loaded on gel and subjected to electrophoresis under non-denaturing conditions (40 mA, 4 h, 4°C) (NATIVE-PAGE). Gel was soaked and shaken in a staining solution containing NBT (2.43 mM), TEMED (28 mM) and riboflavin (0.14 M) in PBS for 20 min at room temperature in the dark. After washing with water, the gel was placed under UV light. Under illumination, the gel became uniformly purple except at locations containing SOD. Illumination was stopped when maximum contrast between the clear and the purple zones was achieved.

#### *IL8 quantification*

Levels of IL8 produced by cells were determined in cell supernatants using a commercially available ELISA kit according to the instructions of the manufacturer. IL8 lev-

els were normalized by the protein content determined in the corresponding cell lysates.

#### *COX2 and SODs analysis*

Western blot analysis was performed to examine expression of COX2, MnSOD and CuSOD versus actin in cell lysates. Laemmli buffer was added to cell lysates (1/10 mixture) and the solution was heated at 100 °C for 3 min. 25 µg of denatured proteins were then loaded on a 10% acrylamide gel and subjected to electrophoresis (150 V at room temperature for 1.5 h) (SDS-PAGE). Proteins were transferred onto a nitrocellulose membrane (30 V, overnight at room temperature). The membrane was cut at 55 and 35 kDa to be revealed by the three antibodies because of their different sensitivity. It was first incubated for 1 h at room temperature in a solution containing non-fat dry milk (5 %) and Tween-20 (0.5 %) in PBS. After washing, membrane was incubated for 1 h at room temperature with primary antibodies diluted to 1/500 in milk solution. After washing, membrane was incubated for 1 h at room temperature with horseradish-peroxidase-conjugated secondary antibodies diluted to 1/5000 in 0.5 % of Tween-20 in PBS. Protein bands were visualized by chemiluminescence using ECL reagents. Intensity bands analysis was performed using ImageJ Software.

#### *In vivo experiment*

Male C57BL/6 mice (6-8 weeks old) were purchased from Janvier Labs (St Berthevin, France) and maintained at the animal care facilities of the National Institute of Agricultural Research (IERP, INRA, Jouy-en-Josas, France) under specific pathogen-free conditions. Mice were housed under standard conditions for 1 week before experimentation start (Light/Dark 12h/12h, 20-22°C, 45-55% humidity, wood shaving as bedding and cellulose sheets as enrichment). They also had free access to water and food (R/M-H, Ssniff, Soest, Germany). All procedures were approved by the French ministry of Research and recorded under the project number 3445-2016010615159974.

Colitis was induced by intrarectal injection of 150 mg/kg of 2,4-dinitrobenzenesulfonic acid (DNBS) solution (Sigma, Saint-Quentin Fallavier) in 30% ethanol (EtOH). Final volume for this injection was 50 µl. Mice also received daily, starting the day of the DNBS injection, 100 µl of Phosphate Buffer Saline (PBS) containing either NaHCO<sub>3</sub> 4.2% (m/V, pH 8), NaHCO<sub>3</sub> 4.2% + MnCl<sub>2</sub> (8 mM) or NaHCO<sub>3</sub> + **Mn1** (4 mM) by oral gavage.

Mice were sacrificed by cervical dislocation. The colon was removed and opened longitudinally to assess macroscopic damages. A score was given to each mouse. Briefly, the macroscopic criteria include macroscopic mucosal damage (such as hyperemia, ulcers, thickening of the colon wall measured with an electronic ruler, the presence of adhesions between the colon and other intra-abdominal organs) and the consistency of fecal material as diarrhea indicator.

#### *Statistical analysis*

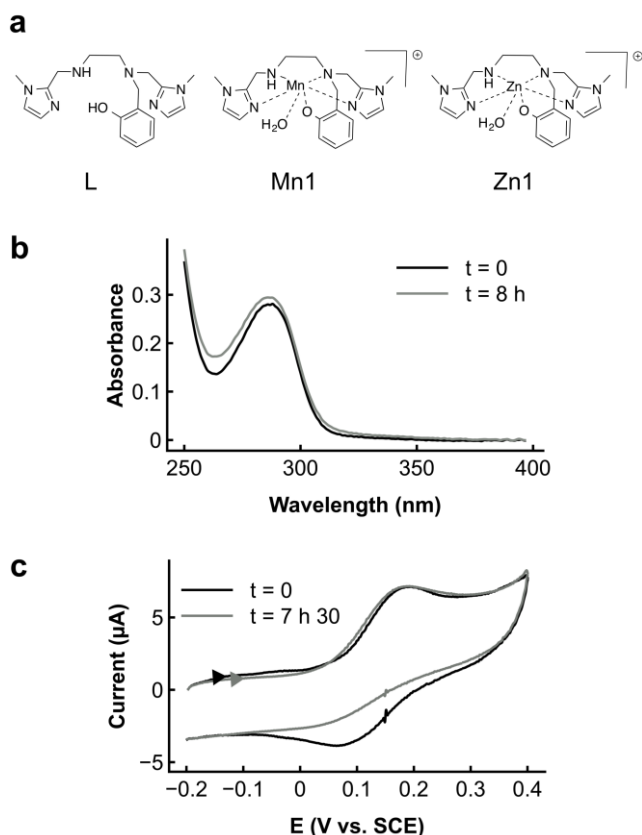
All data are shown as mean ± SEM and were tested for statistical significance using the Mann-Whitney test or

Student's *t* test. Differences were considered significant when  $p < 0.05$ .

## RESULTS

### Stability of Mn1 in solution

The stability of **Mn1** was characterized in HEPES buffer (0.1 M, pH 7.4) and in cell culture medium. The apparent dissociation constant ( $K_d$ ) in HEPES buffer was found to be  $0.76 \pm 0.12 \cdot 10^{-6}$  M at pH 7.4, as measured by Isothermal Titration Calorimetry (ITC) (Fig. S1 A and B). The stability of **Mn1** in cell culture medium was studied using both its UV-visible signature and redox properties. The UV-vis and the redox signature were recorded for several hours, with no modification (see Fig. 1B and C). The redox potential of the  $Mn^{III}/Mn^{II}$  couple in complex **Mn1** is *ca.* 0.15 V/SCE, which is close to that found for metallic center in all SODs and is the optimal redox potential for superoxide dismutation (*ca.* 0.12 V/SCE or 0.36 V/NHE).<sup>23, 46, 96, 97</sup> As expected, the  $Zn^{II}$  complex (**Zn1**) was found redox inactive (see Fig. S1C).



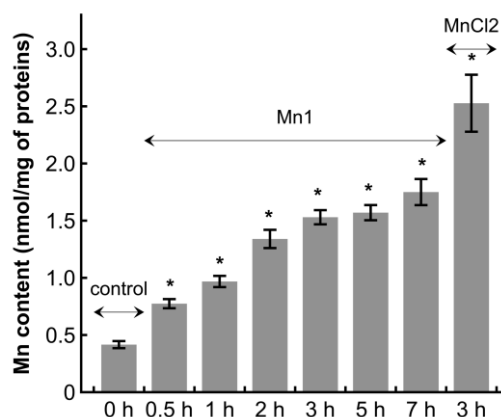
**Figure 1: Structure of ligand L,  $Mn^{II}$ -complex Mn1, and  $Zn^{II}$ -complex Zn1 (a). Stability of Mn1 in solution (see also Fig. S1)** Mn1 stability in cell culture medium was assessed by UV-visible spectroscopy (b) and cyclic voltammetry (CV, c), as the specific signature of the complex was observed after at least 7 hours. b: UV-visible spectra of **Mn1** recorded in complete media (DMEM, 10% FBS, 0.1% blasticidin) at 37 °C in a semi-micro quartz cuvette ( $l = 0.2$  cm) using a double beam mode with media as a reference. c: Cyclic voltammogram of Mn1 in DMEM at 37 °C

under argon atmosphere at a glassy carbon electrode, scan rate  $0.5 \text{ V}\cdot\text{s}^{-1}$ . The forward peak corresponds to the  $Mn^{II}$  oxidation into  $Mn^{III}$  at  $E_{pa} = 0.19 \text{ V/SCE}$ . The fact that it does not decrease indicates that the complex is stable. The diminution of the current intensity of the return peak corresponds to the formation of an hydroxo-Mn complex in solution due to the increase in pH over time.<sup>98</sup>

### Quantification and identification in cell-lysates

Electron paramagnetic resonance (EPR) can be used to quantify  $Mn^{2+}$ , which shows six-sharp lines in its hexa-aqua form. However, in conventional X-band EPR, the EPR spectrum of **Mn1** is broad because of the geometry of the  $Mn^{II}$  coordination sphere being distorted from the regular octahedron.<sup>90</sup> Hence, for quantification purpose,  $Mn^{2+}$  was released from the coordination with **L** by acidification of cell-lysates,<sup>99</sup> which had the additional advantage to favor the EPR responsive  $Mn^{II}$  redox state.

Total  $Mn^{2+}$  was titrated in acidified cell lysates using its six-sharp-line signature in X-band electron paramagnetic resonance (EPR) (Fig.2 and S2) and reported *per* mg of proteins (see protein assay in the SI). Note that this protocol frees manganese from all sites and leads to the evaluation of a total-Mn content.



**Figure 2: Titration of  $Mn^{II}$  in acid-digested lysates of cells HT29 MD2 by EPR.** Cells were incubated at 37°C with **Mn1** (100  $\mu\text{M}$ ) from 0.5 to 7 h or with  $MnCl_2$  (100  $\mu\text{M}$ ) for 3 h. Data represent means  $\pm$  SEM for 5 independent experiments, \*  $p < 0.05$  versus control (see spectra in Fig. S2).

The cellular accumulation of **Mn1** upon incubation at 100  $\mu\text{M}$  was observed with a stabilization of the cellular content after a 3h-incubation. The total  $Mn^{II}$ -content increased from 0.4 nmol/mg of proteins in the control cells to 1.75 nmol/mg of proteins after a 7h-incubation with **Mn1** at 100  $\mu\text{M}$ .  $MnCl_2$  (100  $\mu\text{M}$ ) showed a more efficient uptake.

The detection of a metal complex in biological environments is not straightforward, and LC-MS/MS is well-adapted for that purpose. **Mn1** exhibits a specific signature (see Fig. S3 A, B and C) that was clearly identified in cell-lysates (Fig. S3 D and E) using this method, showing **Mn1** was present in cells.

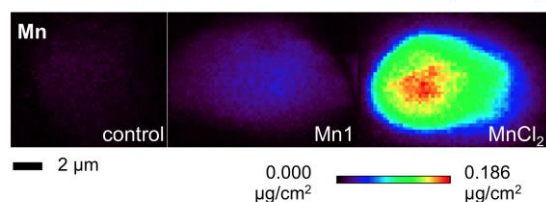
### Cell distribution of Mn



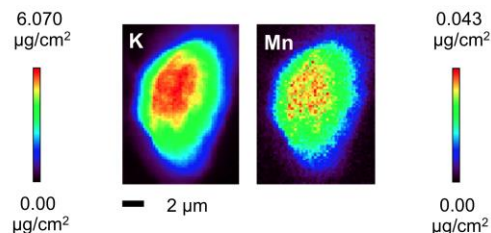
Manganese was mapped on cryofixed freeze-dried cells using the highly spatially resolved synchrotron based X-fluorescence (resolution: 200 nm) with an excitation energy of 6.8 keV, which is above at the K-edge of Mn. The cellular manganese content in a single cell was higher after incubation with **Mn1** than in control cell and MnCl<sub>2</sub> was more efficiently uptaken (Fig. 3a), confirming EPR results. Mappings show a diffuse distribution of Mn, close to that of K (Fig. 3b) — known to be homogeneously distributed in cells<sup>100</sup>—with higher amounts where the cell is the thickest. The distribution of other elements, such as P and S, are shown in Fig. S4.

To confirm the diffuse distribution in the overall cell, Mn was quantified using EPR in mitochondria-enriched cell fractions after 6h-incubation with MnCl<sub>2</sub> or **Mn1**, which shows that Mn is present in the mitochondria (see Fig. S2).

#### a. Mn distribution in cells incubated with Mn1, or MnCl<sub>2</sub>



#### b. K and Mn distribution in cells incubated with Mn1



**Figure 3: X-fluorescence mappings on cryo-fixed cells.** a: Manganese detection by X fluorescence on cryofixed and freeze-dried cells HT29 MD2 incubated 2 h with medium only (control), **Mn1** (100 µM) or MnCl<sub>2</sub> (100 µM) shown with a single scale for the intensity to point at the difference in quantity. b: Potassium and manganese detection by X fluorescence on cryofixed and freeze-dried cell HT29 MD2 incubated with **Mn1** (100 µM). Images recorded on 2-ID-D beamline of APS synchrotron, excitation at 6.8 keV, integration time 2 s / pixel, pixel size 200 nm. Other maps (P and S) are given in Fig. S4.

### Activity of Mn1 in HT29-MD2 epithelial cells

#### Cytotoxicity

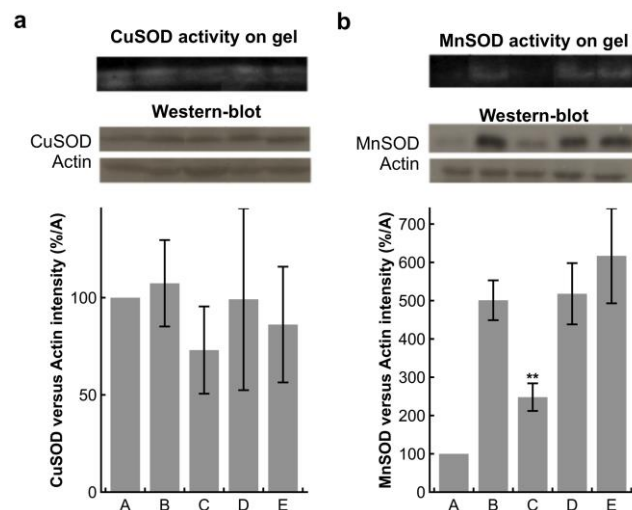
Cytotoxicity was evaluated by LDH release assay. No cytotoxicity (LDH released <10%) was observed for **Mn1** at 100 µM or controls (MnCl<sub>2</sub>, SOD, **Zn1**, ZnBr<sub>2</sub>) in HT29-MD2 cells in steady state or after LPS challenge, whereas the non-coordinated ligand **L** was found cytotoxic in the same conditions and was then not further assayed. The Zn<sup>II</sup>-complex (**Zn1**) constitutes a relevant control, as it is not able to catalyze the dismutation of superoxide but displays the same overall chemical structure and charge as **Mn1**, being thus its redox-silent analogue.

### Anti-superoxide activity of Mn1 in HT29-MD2

Hydroethidine (HE) is a fluorescent probe used for detection of intracellular superoxide in cells.<sup>101, 102</sup> In HT29-MD2 steady state cells HE assay showed that **Mn1** was able to reduce oxidation of HE (Fig. S5), which was not the case for MnCl<sub>2</sub>. This observation confirmed the intracellular anti-superoxide activity of **Mn1** as previously described by our group in macrophages.<sup>89</sup>

### Effect of Mn1 on SODs expressions

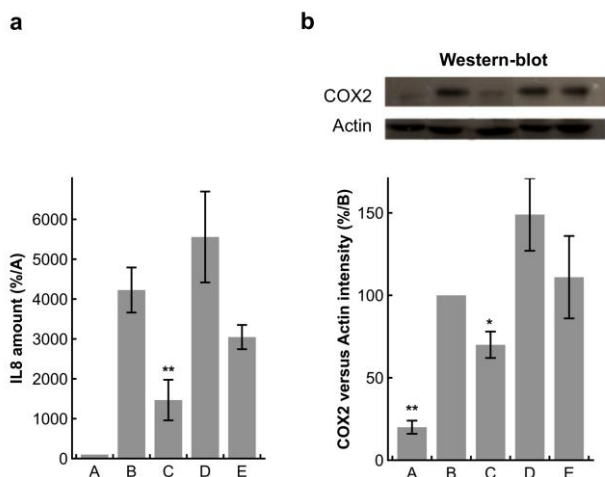
Endogenous mitochondrial MnSOD and cytosolic CuSOD expression and activity were investigated in cell lysates by western blot and colorimetric assay on gel respectively, as shown on Fig. 4. As previously published on other cell lines,<sup>8, 91</sup> LPS challenge induced an over-expression of MnSOD, indicative of an oxidative stress mediated by LPS.<sup>92</sup> Very interestingly, **Mn1** was able to blunt the LPS-induced MnSOD up-regulation. In contrast, cytosolic CuSOD expression and activity were not significantly modified after LPS challenge, and incubation with **Mn1** induced only a weak, non-significant decrease (Fig. 4A). Neither MnCl<sub>2</sub> or purified SOD (Fig. 4), nor **Zn1** (Fig. S6) affected MnSOD over-expression after LPS challenge. In the absence of LPS, incubation of HT29-MD2 cells with **Mn1**, **Zn1** or MnCl<sub>2</sub> induced no significant modification of the expression of MnSOD or activity (Fig. S6).



**Figure 4: CuSOD (a) and MnSOD (b) expression and activity in intestinal epithelial cells activated with LPS.** Representative experiment of CuSOD, and MnSOD expression and activity measured by western blot (WB) and colorimetric assay on gel respectively, after a 7 h incubation under several conditions: (A) control, (B) LPS, (C) **Mn1** (100 µM) + LPS, (D) MnCl<sub>2</sub> (100 µM) + LPS, (E) bovine CuSOD (100 U/mL) + LPS. LPS (0.1 µg/mL) was added after 1 h of incubation. *Top*: NBT assay on non-denatured gel for the measurement of the SODs activities, and WB to evaluate the expression of SODs in comparison with actin. *Bottom*: Quantification of MnSOD or CuSOD versus actin expression obtained by western blot analysis. Data represent means ± SEM for at least 4 (MnSOD) or 3 (CuSOD) independent experiments respectively, \*\* p < 0.01 versus B. Expression of MnSOD in controls without LPS and with LPS and HEPES are shown in Fig. S6. Full blots are shown in Figure S7.

### Anti-inflammatory activity of **Mn1**

The anti-inflammatory activity of the complex **Mn1** was evaluated by quantifying two markers of inflammation — the pro-inflammatory chemokine interleukine 8 (IL8) and cyclo-oxygenase 2 (COX2).<sup>93</sup> **Mn1** was shown to be more efficient against inflammation than 5-ASA, which is an anti-inflammatory agent currently used as the first therapeutic line in IBD<sup>11</sup> (see Fig. S10). LPS-induced IL8 as well as COX2 levels were significantly decreased in presence of **Mn1**, with *ca.* 65 % of inhibition for IL8 secretion (Fig. 5). Upon incubation with MnCl<sub>2</sub>, or purified bovine CuSOD, no significant decrease of IL8 or COX2 was observed. Similarly, **Zn1**, the redox-inactive analogue of **Mn1**, did not decrease IL8 secretion (Fig. S11).

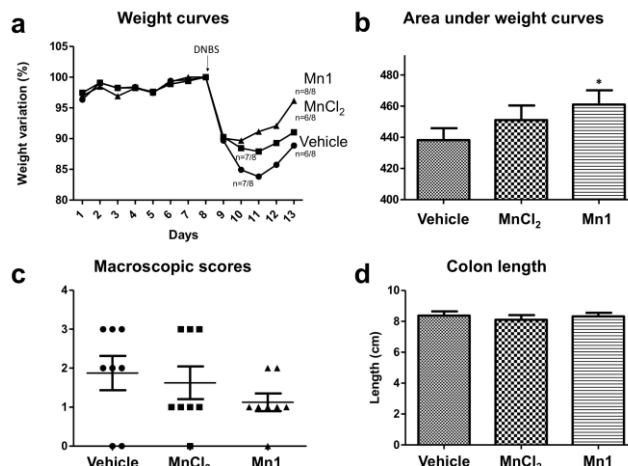


**Figure 5: IL8 and COX2 expression in intestinal epithelial cells activated with LPS.** Anti-inflammatory activity in HT29-MD2 cells after a 7 h incubation under several conditions: (A) control (B) LPS, (C) **Mn1** (100  $\mu$ M) + LPS, (D) MnCl<sub>2</sub> (100  $\mu$ M) + LPS, (E) bovine CuSOD (100 U/mL) + LPS. LPS (0.1  $\mu$ g/mL) was added after 1 h of incubation. **a:** IL8 secretion was measured by ELISA in supernatants. Data represent means  $\pm$  SEM for at least 5 independent experiments, \*\*  $p < 0.01$  versus B. Controls without LPS and with LPS and HEPES are shown in Fig. S11. Note that, without LPS, no significant differences were observed between all these conditions (see Fig. S11, control, **Mn1**, MnCl<sub>2</sub>, HEPES) or upon incubation with **Zn1**. **b:** representative experiment of COX2 and actin expression measured by western blot (top) in cell lysates and quantification of COX2 versus actin expression (bottom). Data represent means  $\pm$  SEM for at least 4 independent experiments, \*  $p < 0.05$  and \*\*  $p < 0.01$  versus D. Full blots are shown in Figure S8.

### Effect of SODm **Mn1** in a chemically induced colitis in mice

In order to assess its anti-inflammatory effect *in vivo*, **Mn1** was administered by oral gavage in mice subjected to DNBS-induced colitis. No change in colon lengths was observed in the three groups (vehicle (NaHCO<sub>3</sub>), MnCl<sub>2</sub>, and **Mn1**, Fig. 6d). Macroscopic score of colitis in mice treated by **Mn1** was lower than in mice treated by vehicle or MnCl<sub>2</sub>, but the difference was weak (Fig. 6b). In this model mortality was observed in the case for mice gavaged with vehicle or MnCl<sub>2</sub>, with 2 deaths out of 8 mice in both

cases. Interestingly, all mice survived with **Mn1**-gavage (Fig. 6a). **Mn1** was also able to reduce weight loss compared to vehicle and MnCl<sub>2</sub> (Fig. 6a and 6b). Moreover, mice treated by **Mn1** started to recover earlier than other mice.



**Figure 6: Effect of **Mn1** in DNBS-induced colitis mice.**

**Mn1** anti-inflammatory effect was investigated in DNBS-induced murine model of colitis by looking at weight variation (**a**, **b**), macroscopic scores (**c**), and colon length (**d**). Colitis was induced by intrarectal injection of DNBS (150 mg/kg, 50  $\mu$ L) in adult male C57BL/6 mice at day 8 after acclimatization start. Mice were treated daily by oral gavage, starting the day of DNBS injection, with 100  $\mu$ L of vehicle (NaHCO<sub>3</sub> 4.2%), MnCl<sub>2</sub> (8 mM), or **Mn1** (4 mM) in NaHCO<sub>3</sub> 4.2%. **a:** Weight variation. **b:** Area under weight curves, \*  $p < 0.05$  versus control. **c:** Macroscopic scores (mucosal damage, consistency of feces, and mortality). **d:** Colon length. The values are expressed as the mean  $\pm$  SEM.

## DISCUSSION

To consider the question of the use of SOD mimics *in vivo*, it is important first to characterize them in cells, and to address questions with regard to their stability, speciation, distribution, as well as bio-activity. Herein, we evaluated, in an inorganic biological chemistry approach, the ability of a manganese complex known for its SOD-like activity (**Mn1**),<sup>89, 90</sup> to control inflammatory response in HT29-MD2 intestinal epithelial cells. This work is in line with our previous studies about the development of SODm inspired from Mn-SOD active site towards an optimization of the SOD-like activity<sup>23, 43, 46, 90, 98</sup> and evaluation of anti-oxidant activity in cells.<sup>89</sup>

Biological media abound with coordinating bio-molecules in competition with **L** for Mn<sup>II</sup>-coordination. Consequently, the question of the speciation of the Mn ion in cells is crucial. The association constants for Mn<sup>II</sup> are known to be in a weak range for any ligand. This is due to Mn<sup>II</sup> d<sup>5</sup> electronic state, with 5 electrons in the d-orbitals and no ligand-field stabilization energy. Hence, the criteria for Mn<sup>II</sup>-complexes stability is far less drastic than with most transition metal ions. The dissociation constant of ligand **L** for Mn<sup>II</sup> was found to be  $0.76 \pm 0.12 \cdot 10^{-6}$  M at pH 7.4. SODm **Mn1** should therefore be stable in presence of biological competitive ligands, such as Human Serum Albumin (HSA) or amino acids,<sup>46</sup> which endow higher dissociation constants for Mn<sup>II</sup> ( $1.2 \cdot 10^{-4}$  M for HSA).<sup>103</sup> The stability of



**Mn1** in the growth medium was checked using UV-visible spectroscopy and cyclic voltammetry. The UV-vis and CV signatures of **Mn1** are unchanged over a seven-hour incubation at 37°C in both DMEM and completed growth medium (see Fig. 1), confirming the stability of **Mn1**. In addition, even if Mn<sup>II</sup> is known to be rather labile,<sup>104, 105</sup> lysate of cells incubated with **Mn1** showed its specific MS-MS signature, indicating that **Mn1** remains at least partially intact in cells (Fig. S3). Moreover, the effect observed on HE conversion attested the anti-superoxide activity of **Mn1** in cells. This experiment is consistent with an intracellular antisuperoxide activity of **Mn1**, as previously shown in the case of macrophages.<sup>89</sup> These results are clear indications that **Mn1** is stable in the experimental conditions of incubation and present in cells, at least for the time-course of the experiments.

The SODm **Mn1** accumulates in HT29-MD2 cells in a time-dependent manner with a maximum concentration reached after about 3h (see Fig. 2). The difference in the intracellular concentration upon incubation with **Mn1** and MnCl<sub>2</sub> was qualitatively confirmed by the mappings using X-fluorescence above the K-edge of Mn at the single cell level (see Fig. 3). Synchrotron-based X-ray fluorescence microscopy ( $\mu$ -SXRF) is a technique giving access to the distribution of heavy elements at the single cell-level without the need of labels and that can be performed on whole cells.<sup>106, 107</sup> Very few examples of subcellular imaging of Mn-SODm have been published.<sup>108</sup>  $\mu$ -SXRF was performed on cryofixed freeze-dried cells, a procedure that ensures the preservation of the cellular ultrastructure and avoids any artifactual redistribution of diffusible low-molecular weight molecules and ions.<sup>106</sup>

The HT29-MD2 cell line is of interest to evaluate the anti-inflammatory and anti-oxidant activity of **Mn1** in cells as it shows a high sensitivity to LPS, with generation of high levels of inflammatory markers.<sup>93</sup> LPS-induced inflammation is known to be associated with oxidative stress,<sup>8, 13, 14, 91, 92</sup> with overproduction of superoxide in the mitochondria.<sup>8, 91, 92</sup> This is in line with the observation reported here that LPS induces overexpression of active MnSOD.<sup>8, 91</sup> Upon incubation with LPS and **Mn1**, a significant decrease in the MnSOD expression was recorded. No effect of the Zn<sup>II</sup> redox-inactive analogue **Zn1**, MnCl<sub>2</sub> or bovine CuSOD was observed (Fig. 4, S6). The absence of effect of **Zn1**, the Zn<sup>II</sup> redox-inactive analogue of **Mn1**, is a clear evidence that the activity is associated with the redox Mn-based properties. MnCl<sub>2</sub> is shown here to efficiently penetrate inside the HT29-MD2 cells but to be biologically inactive. This supports an activity of the Mn-complex and not of the Mn cation, known to react with superoxide but only slowly and in a non-catalytic manner. Upon incubation with bovine CuSOD, active for the catalysis of the superoxide dismutation but unable to cross cell-membranes,<sup>109</sup> no effect was recorded, highlighting that to be efficient on the inflammation cascade the SOD-activity must be exerted inside cells.

Altogether, these observations on MnSOD expression are of interest as it suggests that SODm **Mn1** efficiently com-

plements the mitochondrial MnSOD, which is consistent with an anti-superoxide intracellular effect of **Mn1**. A similar favorable effect of **Mn1** was recorded on LPS-induced IL8 and COX2 expression, with no effect of **Zn1**, MnCl<sub>2</sub>, or bovine CuSOD (Fig. 5, S11)

Very interestingly, **Mn1** was shown to be more efficient against inflammation than 5-ASA which is currently used as the first therapeutic line in IBD<sup>11</sup> (see Fig. S7).

For further confirmation of the anti-inflammatory properties of SODm **Mn1**, we performed a preliminary *in vivo* assay using a chemically induced colitis. SODm **Mn1** was orally gavaged in a carbonate buffer to reduce the instability caused by the passage through the acidic medium of the gastro-intestinal tract. Although SODm **Mn1** was probably not in the optimal conditions, a limited but significant effect was recorded on inflammation and health status of the mice. This constitutes an additional evidence that the ability of this complex to accelerate the superoxide dismutation into H<sub>2</sub>O<sub>2</sub> is a valuable asset in strategies against inflammatory pathologies.

## Conclusion

Antioxidant strategies have been suggested as appropriate to manage IBD.<sup>8, 13, 14, 17, 18</sup> Some other SOD mimics, but few, have been previously shown promising in IBD context.<sup>15</sup> The SOD mimic **Mn1** is an interesting derivative: indeed, it shows an anti-superoxide activity previously characterized out of any cellular context,<sup>90</sup> and in cells.<sup>89</sup> In addition, **Mn1** is easy to synthesize and easily modified, as recently shown.<sup>98</sup> Overall, the approach presented here combining investigation the penetration, speciation and location, with that of the bio-activity, clearly indicates that the SOD mimic **Mn1** is an efficient intracellular anti-inflammatory agent. One important finding of this study is the effect of **Mn1** on mitochondrial MnSOD: **Mn1** is able to complement MnSOD, which has been described as deficient in IBD.<sup>12</sup> In addition, **Mn1** shows a better activity than 5-ASA, already at use in the treatments of IBD, and the oral administration of **Mn1** to *in vivo* model of IBD have led to encouraging results. This is very promising and suggests that the use of this anti-superoxide Mn<sup>II</sup>-complex as an anti-inflammatory agent could constitute a relevant strategy in the treatment of IBD in particular and more generally of oxidative stress induced diseases.

## ASSOCIATED CONTENT

**Supporting Information.** Chemicals, ITC experiment and **Zn1** CV (Fig S1), EPR spectra (Fig S2); intracellular stability: LC-MS-MS experiment (Fig S3); mapping of P, K, Mn and S in cell (Fig. S4); HE-assay (Fig. S5); complementary control assays (Fig S6, S11), anti-inflammatory activity of 5-ASA (Fig. S10), complementary control assays (Fig S6, S8) and pictures of full blots (Fig. S7-S9); supplementary experimental procedures.

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All authors have given approval to the final version of the manuscript. ‡These authors contributed equally.

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### Abbreviations

5-ASA: 5-aminosalicylic acid, BSA: bovine serum albumin, COX2: cyclooxygenase 2, DMEM: Dulbecco Modified Eagle Medium, ECL: enhanced chemiluminescence, EDTA: ethylenediaminetetraacetic acid, ELISA: enzyme-linked immunosorbent assay, EPR: electron paramagnetic resonance, HSA: human serum albumin, IBD: inflammatory bowel diseases, IL8: Interleukine 8, ITC: isothermal titration calorimetry, LC: liquid chromatography LDH: lactate dehydrogenase, LPS: lipopolysaccharide, MS: mass spectrometry, NADH: Nicotinamide adenine dinucleotide, NBT: nitro blue tetrazolium, PBS: phosphate buffer saline, ROS: reactive oxygen species, SOD: superoxide dismutase, SXRF: Synchrotron-based X-ray fluorescence microscopy, TEMED: tetramethylethylenediamine

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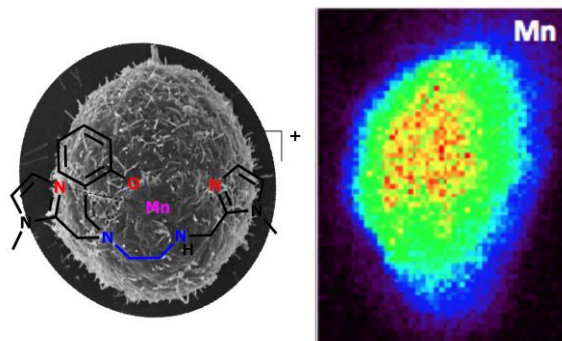
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### Table of contents synopsis

A Mn-complex SOD-mimic was studied in intestinal cell line, HT29-MD2, in which oxidative stress and inflammation was induced using bacterial lipopolysaccharide. This complex was shown to enter cells where it exerts an efficient specific anti-inflammatory activity, to be present in cell lysates, and to have a distribution all over the cell, with the ability to functionally complement the mitochondrial MnSOD.

### Table of contents graphic



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