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Measuring pH variability using an experimental sensor on an underwater glider

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Abstract. Autonomous underwater gliders offer the capability of measuring oceanic parameters continuously at high resolution in both vertical and horizontal planes, with timescales that can extend to many months. An experimental ion-sensitive field-effect transistor (ISFET) sensor measuring pH on the total scale was attached to a glider during the REP14-MED experiment in June 2014 in the Sardinian Sea in the northwestern Mediterranean. During the deployment, pH was sampled at depths of up to 1000 m along an 80 km transect over a period of 12 days. Water samples were collected from a nearby ship and analysed for dissolved inorganic carbon concentration and total alkalinity to derive the pH for validating the ISFET sensor measurements. The vertical resolution of the pH sensor was good (1 to 2 m), but stability was poor and the sensor drifted in a non-monotonous fashion. In order to remove the sensor drift, a depth-constant time-varying offset was applied throughout the water column for each dive, reducing the spread of the data by approximately two-thirds. Furthermore, the ISFET sensor required temperature- and pressure-based corrections, which were achieved using linear regression. Correcting for this decreased the apparent sensor pH variability by a further 13 to 31 %. Sunlight caused an apparent sensor pH decrease of up to 0.1 in surface waters around local noon, highlighting the importance of shielding the sensor from light in future deployments. The corrected pH from the ISFET sensor is presented along with potential temperature, salinity, potential density anomalies (σ_θ), and dissolved oxygen con-

centrations ($c(\text{O}_2)$) measured by the glider, providing insights into the physical and biogeochemical variability in the Sardinian Sea. The pH maxima were identified close to the depth of the summer chlorophyll maximum, where high $c(\text{O}_2)$ values were also found. Longitudinal pH variations at depth ($\sigma_\theta > 28.8 \text{ kg m}^{-3}$) highlighted the variability of water masses in the Sardinian Sea. Higher pH was observed where salinity was > 38.65 , and lower pH was found where salinity ranged between 38.3 and 38.65. The higher pH was associated with saltier Levantine Intermediate Water, and it is possible that the lower pH was related to the remineralisation of organic matter. Furthermore, shoaling isopycnals closer to shore coinciding with low pH and $c(\text{O}_2)$, high salinity, alkalinity, dissolved inorganic carbon concentrations, and chlorophyll fluorescence waters may be indicative of upwelling.

1 Introduction

It is estimated that one-third of the anthropogenic carbon dioxide emitted between 2004 to 2013 was absorbed by the oceans (Le Quéré et al., 2015). Normally unreactive in the atmosphere, carbon dioxide dissolved in seawater ($\text{CO}_2(\text{aq})$) takes part in a number of chemical reactions. In particular, carbonic acid (H_2CO_3) forms as a result of ($\text{CO}_2(\text{aq})$) reacting with water, which dissociates into bicarbonate (HCO_3^-) and carbonate (CO_3^{2-}). These seawater carbonate species are referred to as dissolved inorganic carbon concentrations

($c(\text{DIC})$), with “ c ” representing a concentration) with HCO_3^- accounting for 90 % of $c(\text{DIC})$. During the dissociation of carbonate species, hydrogen ions (H^+) are released. The pH is calculated as the negative decadal logarithm of the activity (commonly referred to as a concentration) of these H^+ ions. Thus the pH in the ocean is directly related to the activity of H^+ ions in the water (Zeebe and Wolf-Gladrow, 2001).

The pH varies on timescales spanning less than 1 day (Hofmann et al., 2011) to many years (Rhein et al., 2013). Since before the industrial revolution (year 1760), the global surface ocean pH has fallen from 8.21 to 8.10 (corresponding to a 30 % increase in H^+ ion activity) as a result of the atmospheric CO_2 mole fraction increasing by more than $100 \mu\text{mol mol}^{-1}$ (Doney et al., 2009; Fabry et al., 2008). Future projections of anthropogenic carbon dioxide emissions suggest that ocean uptake of CO_2 will continue for many decades, thus contributing to long-term ocean acidification (Rhein et al., 2013). This may have a significant effect on marine organisms, such as calcifying phytoplankton (e.g. coccolithophores) and corals (e.g. scleractinian), dependent on the solubility state of calcium carbonate (Doney et al., 2009).

Since 1989, it has been possible to measure pH to an accuracy of 10^{-3} using a spectrophotometric approach (Byrne and Breland, 1989). Although there have been some advances in adapting this approach to measure pH autonomously in situ (Martz et al., 2003; Aßmann et al., 2011), spectrophotometry is largely used for shipboard measurements, as it requires the use of indicator dye and a means to measure spectrophotometric blanks, which is challenging outside of a laboratory (Martz et al., 2010).

A limited number of hydrographic surveys have been undertaken, and stations offering long-term time series of pH are available (Rhein et al., 2013). However, there is a drive to improve spatial and temporal data coverage via autonomous means, similar to what was experienced for temperature and salinity with Argo floats 16 years ago (Roemmich et al., 2003). There is demand to develop a reliable autonomous sensor with a precision and accuracy of 10^{-3} that is affordable to the scientific community (Johnson et al., 2016).

The Mediterranean Sea comprises just 0.8 % of the global oceanic surface, but it is regarded as an important sink for anthropogenic carbon due to its physical and biogeochemical characteristics (Álvarez et al., 2014). Between 1995 and 2012, surface $c(\text{DIC})$ increased by $3 \mu\text{mol kg}^{-1} \text{a}^{-1}$ in the northwest Mediterranean Sea, which is consistent with a rise in temperature of $0.06 \text{ }^\circ\text{C a}^{-1}$ and a decrease in pH of 0.003 a^{-1} (Yao et al., 2016). In contrast, the pH in the neighbouring North Atlantic Ocean decreased by just 0.0017 a^{-1} associated with an increase in $c(\text{DIC})$ of around $1.4 \mu\text{mol kg}^{-1} \text{a}^{-1}$ and a temperature rise of $0.01 \text{ }^\circ\text{C a}^{-1}$ (Bates et al., 2012). The greater potential of the Mediterranean Sea to store anthropogenic carbon can be explained by its higher alkalinity, warmer temperatures, and thus lower Revelle factor (Álvarez et al., 2014; Touratier and Goyet,

2011) when compared with other oceans, such as the North Atlantic.

The pH in the Mediterranean Sea is typically higher than most other oceanic regions (Álvarez et al., 2014). The pH on the total scale normalised to $25 \text{ }^\circ\text{C}$ ($\text{pH}_{T,25}$) collected by ship between 1998 and 1999 within the northwestern Mediterranean Sea varied between 7.92 and 8.04 at the surface and between 7.9 and 7.93 at depths greater than 100 m (Copin-Montégut and Bégovic, 2002). When considering the Mediterranean Sea as a whole, $\text{pH}_{T,25}$ obtained by ship in April 2011 varied between 7.98 and 8.02 at the surface and between 7.88 and 7.96 at greater depths (Álvarez et al., 2014). The peak-to-peak amplitude of the annual pH cycle in the northwest Mediterranean Sea is typically 0.1 with maxima and minima in spring and summer, respectively (Yao et al., 2016).

Measurements of pH with higher temporal resolution, such as those measured by in situ sensors, can vary greatly depending on their location and depth. Hofmann et al. (2011) presented the results of 15 deployments using SeaFET pH sensors (Sea-Bird Scientific) close to the surface at a number of locations worldwide. They found that pH could vary by as much as 1.1 in extreme environments, such as those obtained close to volcanic CO_2 vents off the coast of Italy, or as little as 0.02 in open ocean areas, such as in the temperate eastern Pacific Ocean, over a time period of 30 days. Hofmann et al. (2011) were able to capture diel cycles in pH, with the most consistent variations found in coral reef locations. The pH was at a maximum in the early evening and at a minimum in the morning with amplitudes between 0.1 and 0.25; this is similar in range to other studies based in subtropical estuaries (Yates et al., 2007).

Autonomous underwater gliders offer the possibility to observe the oceanic system with a greater level of detail on both temporal and spatial scales when compared with ship measurements (Eriksen et al., 2001). A low consumption of battery power and a great degree of manoeuvrability enable such vehicles to cover large areas and profile depths of up to 1000 m during missions that can last from weeks to months. They are suitable platforms for a range of sensors, measuring both physical and biogeochemical parameters (Piterberg et al., 2014; Queste et al., 2012).

This paper is a contribution to the special issue “REP14-MED: A Glider Fleet Experiment in a Limited Marine Area”. The main goal of this paper is to describe the trial of a novel ion-sensitive field-effect transistor (ISFET) pH sensor, which was attached to an autonomous underwater glider in the northwest Mediterranean Sea during the REP14-MED sea experiment. The secondary objective is to provide a method of correcting the pH measured by this sensor and to discuss the spatial and temporal variability observed. The experiment, the glider sensors including the ISFET sensor, and the method of validation are described in Sect. 2. The ship-based data are presented in Sect. 3.1, and a comparison between ship and glider measurements is made in Sect. 3.2.1.

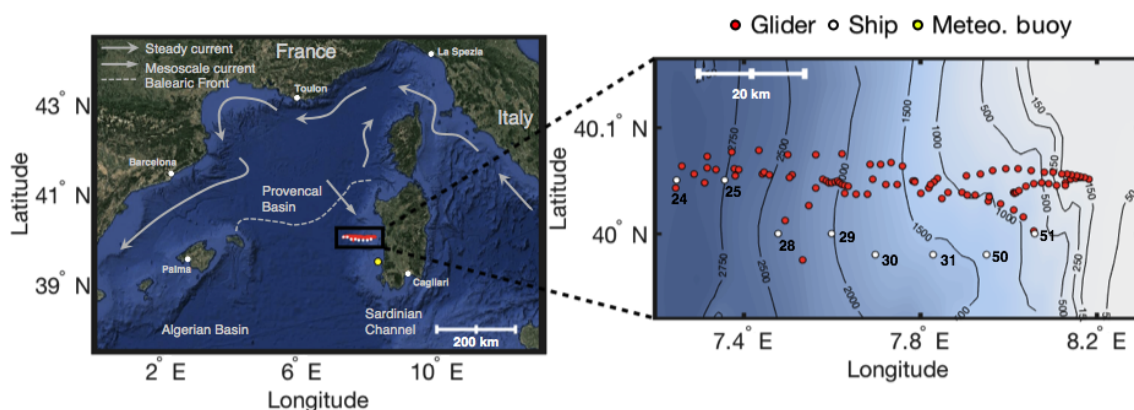


Figure 1. The locations of the 93 dives undertaken by the Seaglider (red markers), the eight ship CTD casts in which water samples were obtained (white markers), and meteorological buoy M1 (yellow marker) within the REP14-MED observational domain off the coast of Sardinia, Italy between 11 and 23 June 2014. GEBCO 1 min resolution bathymetry data (metres) were used (<http://www.bodc.ac.uk/projects/international/gebco/>), and surface circulation patterns were adapted from figures presented by Millot (1999).

The initial pH results and validation, the method of further correcting pH, and an artefactual light-induced effect are described in Sect. 3.2.2. Corrected pH measurements are analysed alongside other collected parameters in Sect. 3.2.3, and the conclusions are presented in Sect. 4.

2 Methodology

2.1 REP14-MED sea trial

This trial took place between 6 and 25 June 2014 in the northwest Mediterranean Sea off the coast of Sardinia, Italy (Fig. 1). This was part of the Environmental Knowledge and Operational Effectiveness (EKOE) research programme led by the North Atlantic Treaty Organisation (NATO) Centre for Maritime Research and Experimentation (CMRE) based in La Spezia, Italy. This was the fifth Recognised Environmental Picture (REP) trial, which was jointly conducted by two research vessels: the NRV *Alliance* and the RV *Planet*.

Eleven gliders with varying pressure tolerances were deployed during the trial, each making repeated west–east transects separated by roughly 0.13° of latitude from one another within the REP14-MED observational domain. One of these gliders was operated by the University of East Anglia (UEA): an iRobot Seaglider model 1KA (SN 537) with an ogive fairing. All gliders were deployed to meet the objectives of the trial, such as to improve ocean forecasting techniques (e.g. model validation and evaluation of forecasting skill), conduct a cost–benefit analysis of autonomous gliders, analyse mesoscale and sub-mesoscale features, and test new glider payloads. The latter objective was perhaps the most relevant to the deployment of the UEA glider. A more in-depth overview of the REP14-MED trial, its objectives, and the collected observational data is described by Onken et al. (2016).

The UEA glider completed a total of 126 dives between 11 and 23 June 2014. The first 24 dives did not record pH and the last 9 dives were very shallow, leaving 93 usable dives. Successive dives were approximately 2 to 4 km apart, descending to depths of up to 1000 m.

2.2 Glider sensors

Conductivity, pressure, and in situ temperature measurements were obtained by the glider using a Sea-Bird Scientific glider payload CTD sensor (Bellevue, WA, USA; Fig. 2). These measurements were then used to obtain potential temperature (θ) and practical salinity.

Dissolved oxygen concentrations ($c(\text{O}_2)$, where “c” refers to a concentration) were measured using an Aanderaa 4330 oxygen optode sensor (Aanderaa Data Instruments, Bergen, Norway) positioned towards the rear of the glider fairing (Fig. 2). The method of calibrating $c(\text{O}_2)$ closely followed that described by Binetti (2016), using the oxygen sensor engineering parameters TCPhase and CalPhase, which will be summarised here. The first step involved correcting $c(\text{O}_2)$ to account for the response time (τ) of the sensor, as the diffusion of oxygen across the silicon foil of the sensor is not an instantaneous process. Each oxygen sensor has a different τ , which depends on the structure, thickness, age, and usage of the foil (McNeil and D’Asaro, 2014), as well as the external environmental conditions such as temperature. An average τ of 17 s was obtained using the method outlined by Binetti (2016). After correcting the TCPhase for τ , glider TCPhase profiles were matched in time and space with pseudo-CalPhase profiles back-calculated from the measurements of $c(\text{O}_2)$ obtained by the ship Sea-Bird Scientific SBE 43 sensor (CTD package) using the manufacturer’s set of optode calibration equations. The relationship between the glider TCPhase and the ship pseudo-CalPhase was established, and the calculated slope and offset coefficients were

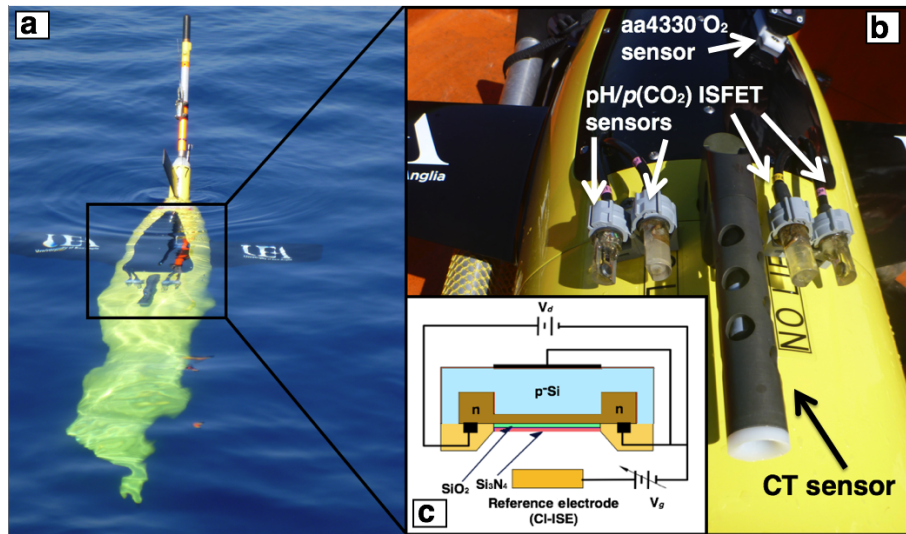


Figure 2. (a) Seaglider SN 537 during the deployment; (b) a close-up of the sensors and (c) a schematic diagram of the ISFET sensor adapted from Shitashima (2010).

used to correct the glider CalPhase, which is required for calibrating $c(\text{O}_2)$ measurements. A comparison between the ship $c(\text{O}_2)$ measurements and the calibrated glider $c(\text{O}_2)$ measurements is made in Sect. 3.2.1.

Glider variables have been processed using an open-source MATLAB-based toolbox (<https://bitbucket.org/bastienqueste/uea-seaglider-toolbox/>) in order to correct for differing timestamp allocations, sensor lags (Garau et al., 2011; Bittig et al., 2014), and to tune the hydrodynamical flight model (Frajka-Williams et al., 2011). Outliers outside of a specified range (e.g. 6 standard deviations) were flagged and not used for analysis, and glider profiles were smoothed using a lowess low-pass filter with a span of 5 data points (< 4 m range), which implements a local regression using weighted linear least squares and a first-order polynomial linear model. Individual profiles were inspected afterwards to ensure that potentially correct data points were not removed.

The ISFET pH sensor used in this study (Fig. 2) was custom built by a working group led by Kiminori Shitashima at the Tokyo University of Marine Science and Technology (previously the University of Kyushu), and it is not commercially available. The ISFET unit was housed in acrylic resin material. The ISFET unit and the reference chlorine ion-selective electrode (Cl-ISE) were moulded with epoxy resin in the custom-built housing. The ISFET pH unit was stand-alone, meaning that the sensor was not integrated into any of the onboard glider electronics. The power source of the sensor was 10.5 V supplied by three 3.5 V Li-ion batteries.

The glider also carried another ISFET pH sensor that was integrated into the glider electronics (Fig. 2) and two $p(\text{CO}_2)$ sensors (Shitashima, 2010), one stand-alone and one inte-

grated. The data retrieved from the integrated pH sensor and the $p(\text{CO}_2)$ sensors could not be used due to quality issues. We think the regular on/off cycling of power to the integrated dual pH– $p(\text{CO}_2)$ sensor between sampling did not allow it to function properly. In future, we would suggest the addition of backup batteries to supply power to the sensor between sampling. The cause of the problem with the stand-alone $p(\text{CO}_2)$ unit is unclear.

To measure pH, the activity of H^+ ions is determined using the interface potential between the Cl-ISE and the semi-conducting ion-sensing transistor coated with silicon dioxide (SiO_2) and silicon nitride (Si_3N_4). The ISFET pH sensor was previously found to have a response time of a few seconds with an accuracy of 0.005 pH and suitable temperature and pressure sensitivities (Shitashima et al., 2002, 2013; Shitashima, 2010). Before deploying the sensor, the ISFET and Cl-ISE were conditioned (as recommended by Bresnahan et al., 2014 and Takeshita et al., 2014) in a bucket of local sea surface water with a salinity of 38.05. However, due to time constraints, conditioning took place over just 1 hour rather than weeks as specified by Bresnahan et al. (2014) and Takeshita et al. (2014). During the deployment, pH measurements were obtained vertically every 1 to 2 m.

The measurements obtained by the ISFET pH sensor were converted from raw output counts to pH on a total scale using a two-point calibration with 2-aminopyridine (AMP) and a 2-amino-2-hydroxymethyl-1, 3-propanediol (TRIS) buffer solution before and after the deployment of the glider. The same buffer solutions (Wako Pure Chemical Industries, Ltd., Osaka, Japan) created in synthetic seawater ($S = 35$, ionic strength of around 0.7 M) were used before and after deployment. These buffer solutions had a pH of 6.79 ± 0.03 (AMP) and 8.09 ± 0.03 (TRIS). The pH uncertainty of the

buffer solutions takes into account the effect of changing air temperature, ranging between 30.5 and 33.3 °C during pre-calibration and between 27.5 and 28 °C during post-calibration. A linear fit using the raw output measured from these buffer solutions was used to convert the raw counts to pH (Shitashima et al., 2002). A drift was observed between the pH of these buffer solutions before and after the deployment, which was corrected for. As the ISFET sensor was previously described as having a pressure-resistant performance and good temperature characteristics for oceanographic use (Shitashima et al., 2002; Shitashima, 2010), no compensations for temperature and pressure were performed on the ISFET measurements at this stage. The ISFET pH sensor has a salinity sensitivity of $\partial\text{pH} / \partial S = 0.011$, which was taken into account. The effect of biofouling on the ISFET pH measurements, as well as on all other glider measurements, was ruled out after a post-deployment inspection of the sensors indicated no problems.

2.3 Ship-based measurements

As the in situ ISFET pH sensor was under trial, some form of validation of the results was required. In total, 124 water samples were collected from Niskin bottles sampled at 12 depths (down to 1000 m) using a CTD rosette platform at eight locations (eight casts, numbered 24–51) close to the path of the glider (Fig. 1). Water samples were collected between 05:19 local time (LT, UTC+2) on 9 June and 16:58 LT on 11 June. The glider ISFET pH sensor started operating at 16:36 LT on 11 June. Overall, the measurements obtained by the glider and the CTD overlapped better in space than in time (Fig. 3).

When collecting carbon samples, water was drawn into 250 mL borosilicate glass bottles from Niskin bottles on the CTD rosette using tygon tubing. Bottles were rinsed twice before filling and were overflowed for 20 s, allowing the bottle volume to be flushed twice. Each sample was poisoned with 50 μL of saturated mercuric chloride and then sealed using greased stoppers, secured with elastic bands, and stored in the dark (Dickson et al., 2007). The total alkalinity (A_T) and the $c(\text{DIC})$ of each water sample was measured in the laboratory using a Marianda Versatile INstrument for the Determination of Titration Alkalinity (VINDTA 3C; www.marianda.com). The $c(\text{DIC})$ was measured by coulometry (Johnson et al., 1985) following standard operating procedure SOP 2, and A_T was measured by potentiometric titration (Mintrop et al., 2000) following SOP 3b, both described by Dickson et al. (2007). During the analytical process, 21 bottles of certified reference material (CRM; batch 107) supplied by the Scripps Institution of Oceanography (San Diego, CA, USA) were run through the instrument to keep track of stability and to calibrate the instrument. For each day in the lab, one CRM was used before and after the samples were processed. A total of 19 concurrent replicate-depth water samples were collected with around 2 to 3 replicates per CTD cast. Calculating the mean stan-

dard deviation of these replicate samples enabled a measurement of the instrument precision. The mean standard deviations of the $c(\text{DIC})$ and A_T replicates were 1.7 $\mu\text{mol kg}^{-1}$ and 1.4 $\mu\text{mol kg}^{-1}$, respectively. This corresponds to a pH uncertainty of 0.003 for $c(\text{DIC})$ and A_T , resulting in a combined uncertainty of 0.009.

Once A_T and $c(\text{DIC})$ were known, pH could be derived using the CO2SYS programme (Van Heuven et al., 2011). This calculation has an estimated pH probable error of around 0.006 due to uncertainty in the dissociation constants pK_1 and pK_2 (Millero, 1995). Temperature and salinity were obtained from the Sea-Bird CTD sensor on the ship rosette sampler, and the seawater equilibrium constants presented by Mehrbach et al. (1973) were used as refitted by Dickson and Millero (1987), which has been recommended by previous studies (e.g. the CARINA data synthesis project) for the Mediterranean Sea (Álvarez et al., 2014; Key et al., 2010). The sulfate constant described by Dickson (1990) and the parameterisation of total borate presented by Uppström (1974) were used. More information on the equilibrium constants used in CO2SYS and other available carbonate system packages is described by Orr et al. (2015). The pH derived from the water samples collected by ship and the glider-retrieved ISFET pH are both on the total pH scale (as described by Dickson, 1984) at in situ temperature and will from now on be referred to as pH_s and pH_g , respectively.

Standard deviation ranges will from this point be listed when referring to variability in measurements, such as pH_s and pH_g . To obtain these standard deviation ranges, data points for a given variable were sorted into 10 m depth bins down to a maximum depth of 1000 m. The standard deviation was then calculated for each bin.

3 Results and discussion

3.1 Ship-based data

Measurements obtained by the ship CTD package provide an overview of the temporal and spatial variability for the time at which the water samples used to derive pH_s were collected (Fig. 4). The θ gradient was strong in the top 100 m of the water column due to limited vertical mixing with a maximum between 19 and 23 °C found in the upper 10 m of the water column, decreasing to between 13 and 14 °C at depths greater than 100 m. The salinity was low in the top 100 m, increasing to a maximum at around 400 to 600 m. These fresher waters in the top 100 m are likely modified Atlantic water (MAW), typically having a salinity between 38 and 38.3 in the northwest Mediterranean Sea (Millot, 1999). These waters enter from the Atlantic Ocean through the Strait of Gibraltar, flowing along the North African coast. Some water makes its way northward and follows the shelf back west towards the Atlantic Ocean (Rivaro et al., 2010; Millot, 1999). At deeper depths, warmer saltier waters were found

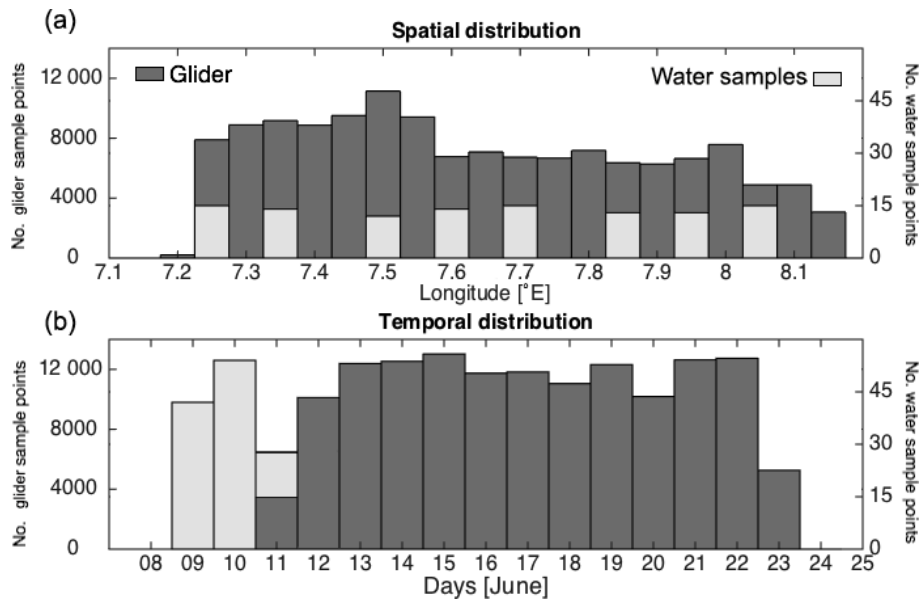


Figure 3. Histograms showing the (a) spatial and (b) temporal distribution of samples collected by the glider (dark grey) and by CTD water bottle sampling (light grey). The y axis on the left is for the sum of the glider samples, whilst the y axis on the right is for the sum of the water samples.

east of 7.5° E, which is likely to be Levantine Intermediate Water (LIW) identified in the western Mediterranean Sea by a salinity range of 38.45 to 38.65 and θ between 13.07 and 13.88 °C (Rivaro et al., 2010), typically found at depths between 200 and 800 m close to the shelf slope (Millot, 1999). The $c(\text{O}_2)$ maxima were found at depths between 20 and 90 m. The Mediterranean Sea on the whole is considered to be oligotrophic (Álvarez et al., 2014). However, a deep chlorophyll maximum (DCM) is common at these depths when waters are thermally stratified (Estrada, 1996). There is a build-up of actively growing biomass with greater cell pigment content as a result of photoacclimation due to increased concentrations of nitrate, phosphate, and silicate, as well as sufficient levels of light at these depths (Estrada, 1996). It is likely that this high $c(\text{O}_2)$ was related to the DCM, further evidenced by the high chlorophyll fluorescence layer observed at 60 to 100 m of depth, particularly in the east.

The objective of deriving pH_s using A_T and $c(\text{DIC})$ was to make a comparison with the pH_g measured by the ISFET sensor. The $c(\text{DIC})$ and A_T were greatest at depths below 250 m with lower values seen closer to the surface (Fig. 5a–b), which is typical of the northwest Mediterranean Sea (Copin-Montégut and Bégovic, 2002; Álvarez et al., 2014). The higher values of A_T and $c(\text{DIC})$ at depth and in the east support the notion that this is LIW, as this water mass has previously been identified as having an A_T of around $2590 \mu\text{mol kg}^{-1}$ and a $c(\text{DIC})$ of roughly $2330 \mu\text{mol kg}^{-1}$ (Álvarez et al., 2014), coinciding with the warmer saltier waters. The mean $c(\text{DIC})$ and A_T (averages over eight casts) have standard deviations of 6.1 to 11.9 and 5.9 to

$10.6 \mu\text{mol kg}^{-1}$, respectively, for the top 150 m of the water column and 1.7 to $3.9 \mu\text{mol kg}^{-1}$ and 3.7 to $7.6 \mu\text{mol kg}^{-1}$ for deeper waters. The pH_s had a maximum of 8.14 between 50 and 70 m of depth (Fig. 5c). The mean pH_s values have standard deviations of 0.004 to 0.011 within the top 150 m and 0.006 to 0.017 deeper than this. A proportion of these standard deviations can be explained by the instrumental error associated with the analysis of $c(\text{DIC})$ and A_T discussed in Sect. 2.3.

3.2 Glider data

3.2.1 Temperature, salinity, and oxygen validation

Since the sensors were calibrated before deployment, it was expected that the measurements from the glider would match those from the CTD because any discrepancies between datasets would indicate possible instrumental or methodological issues with the glider measurements. The mean profiles of θ , salinity, and $c(\text{O}_2)$ collected by the glider and by ship (Fig. 6) agreed well. The values obtained by both ship and glider were mostly within 1 standard deviation of one another. The mean θ and salinity retrieved during the eight ship pH_s casts differed from the binned mean calculated using all available REP14-MED ship casts at depths between 100 and 500 m. However, this is likely related to temporal or spatial variability as mean θ and salinity were within the range of all available glider measurements. Furthermore, differences of roughly 0.1 °C, 0.02, and $1.5 \mu\text{mol kg}^{-1}$ can be seen for θ , salinity, and $c(\text{O}_2)$, respectively, between the binned mean profile of CTD measurements and the binned mean profile

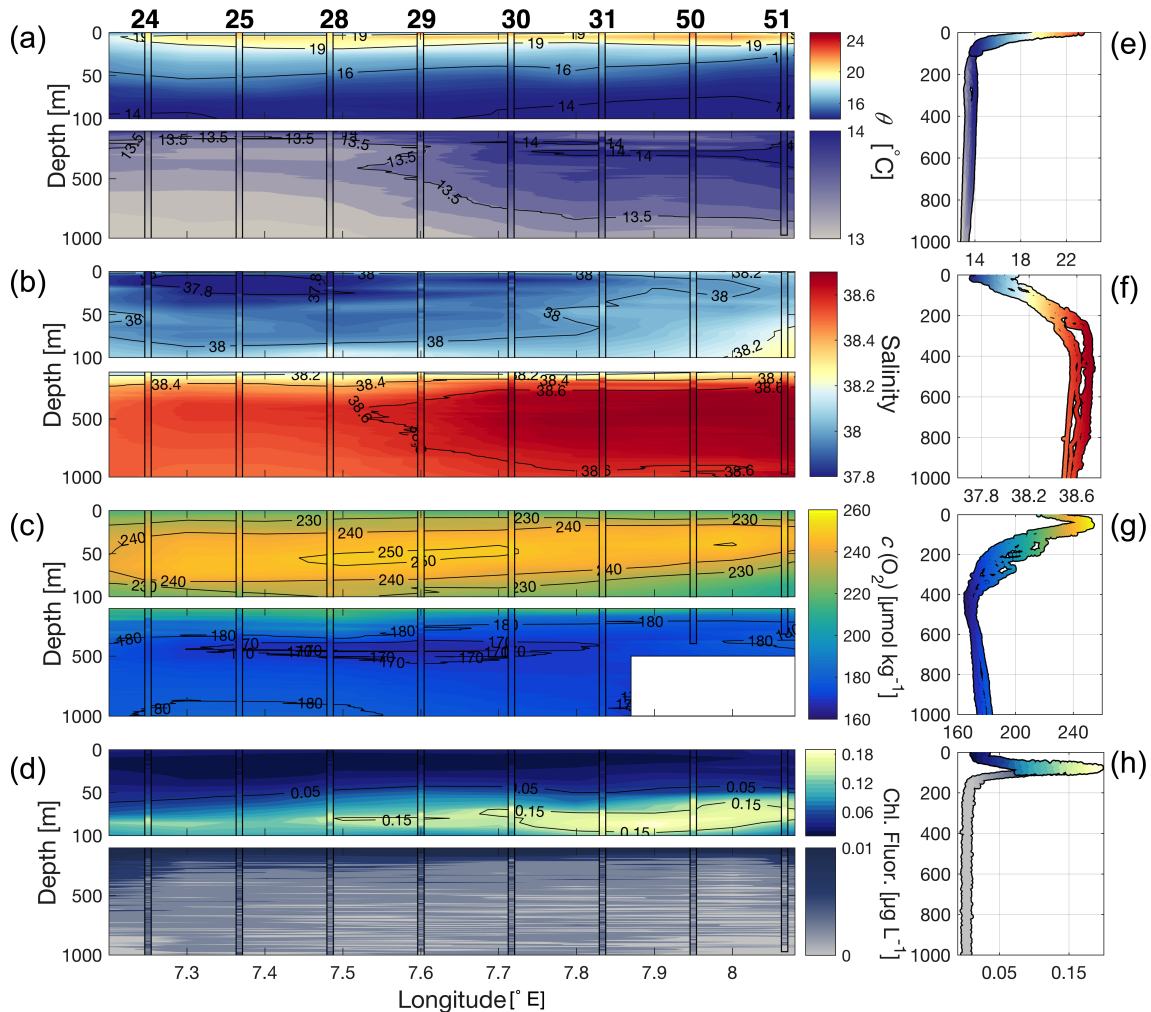


Figure 4. Transects of optimally interpolated (a) potential temperature (θ), (b) salinity, (c) dissolved oxygen concentrations ($c(\text{O}_2)$), and (d) chlorophyll fluorescence, along with their depth profiles (e–h) obtained by ship. These parameters were sorted into 0.1° longitude \times 5 m bins, and the radius of influence used for optimal interpolation was 0.2° longitude \times 20 m. The data retrieved during the eight CTD casts (displayed in Fig. 1) used for optimal interpolation are superimposed on the interpolated fields.

of glider measurements at depths greater than 500 m. These differences in θ , salinity, and $c(\text{O}_2)$ are related to the different spatial distribution of the two datasets, as the glider measured predominantly at 40° N where deep, cooler, and fresher waters were observed in the west (Fig. 4a–b) that are uncommon in other areas of the observational domain (Knoll et al., 2015b).

3.2.2 ISFET pH validation

The mean pH_g and pH_s agreed best between 60 and 250 m (Fig. 7), although pH_g variability was a lot higher than for pH_s . Larger differences between these profiles can be seen above and below this depth range, with a pH_g 0.1 higher at the surface and roughly 0.07 lower between 950 and 1000 m when compared with pH_s . The pH_s maximum at approximately 50 to 70 m of depth was not apparent in the pH_g pro-

file, with the highest pH_g seen at the surface. The standard deviations for pH_g were large, ranging between 0.044 and 0.114 in the top 150 m of the water column and between 0.027 and 0.053 at other points in the water column. Comparing all pH_g dive profiles obtained during the mission suggests a great degree of temporal and spatial variability, with pH ranging from 8.02 to 8.28 at the surface and between 7.97 and 8.13 at 800 m of depth.

A diel cycle in pH_g anomalies (calculated by subtracting the all-time mean from the hourly means within a given depth interval) was found predominantly at depths shallower than 20 m (Fig. 8b). Lower pH was found between 09:00 and 18:00 LT, decreasing by > 0.1 between 12:00 and 14:00 LT. Contrastingly, potential temperature, salinity, and $c(\text{O}_2)$ anomalies (calculated in the same way as pH_g anomalies) did not have strong diel cycles (Fig. 8c–e), suggesting

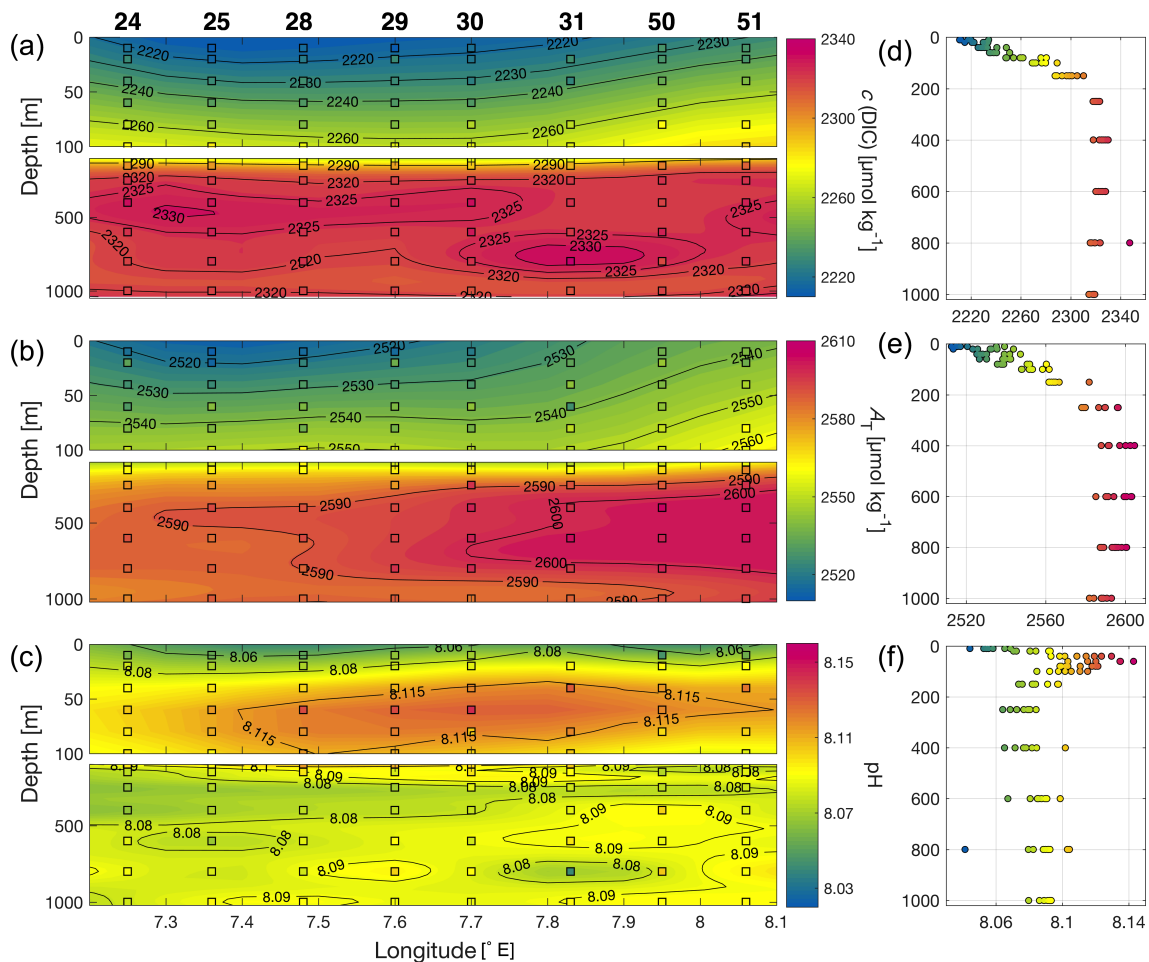


Figure 5. Optimally interpolated fields of (a) dissolved inorganic carbon ($c(\text{DIC})$), (b) total alkalinity (A_T), and (c) pH derived using $c(\text{DIC})$ and A_T are displayed along with their depth profiles (d–f). These parameters were sorted into 0.1° longitude \times 20 m bins, and the radius of influence used for optimal interpolation was 0.3° longitude \times 200 m for $c(\text{DIC})$ and (A_T) and 0.3° longitude \times 80 m for pH. The water sample values retrieved during the eight CTD casts (displayed in Fig. 1) used for optimal interpolation are superimposed on the interpolated fields as squares.

that the decrease in pH was not caused by changing environmental conditions. Particularly, one might expect $c(\text{O}_2)$ to have a similar pattern to pH if it were related to photosynthesis or respiration due to variations in $p(\text{CO}_2)$ (Cornwall et al., 2013; Copin-Montégut and Bégovic, 2002). However, $c(\text{O}_2)$ remained relatively constant throughout the day at all depth ranges, implying that the level of biological activity in the Sardinian Sea did not change on average throughout the day and hence would not have caused this reduction in pH_g .

The decrease in pH_g coincided with increased levels of solar irradiance (Fig. 8a) recorded at meteorological buoy M1 (Fig. 1) during the day at the surface; hence it was likely a light-induced instrumental artefact. The effect of light on the voltage output of FET-based sensors using SiO_2 - and Si_3N_4 -sensitive layers is known (Włodarski et al., 1986), as the presence of photons can excite electrons in the valence band of the semiconductor material, creating holes and allowing

the flow of electrons to the conduction band. This increases the voltage threshold to falsely measure higher hydrogen ion activity, leading to lower apparent pH (Liao et al., 1999).

The effect of light on our sensor was investigated further by exposing two ISFET pH sensors to artificial light whilst placed in reference buffer solutions (TRIS and AMP) under laboratory conditions. The results (not shown here) confirmed that our ISFET sensor is affected by light. The light-induced offset depended on the strength and type of the light source and which sensor was being used. The offset remained relatively constant whilst the light was turned on. Maximum pH offsets of -0.7 (-6×10^6 counts) and -0.15 (-3×10^6 counts) were found when the LED and halogen lights were used, respectively.

There were not enough dives for a robust light correction, and an irradiance-measuring sensor was not attached to the glider; hence data collected within the top 50 m be-

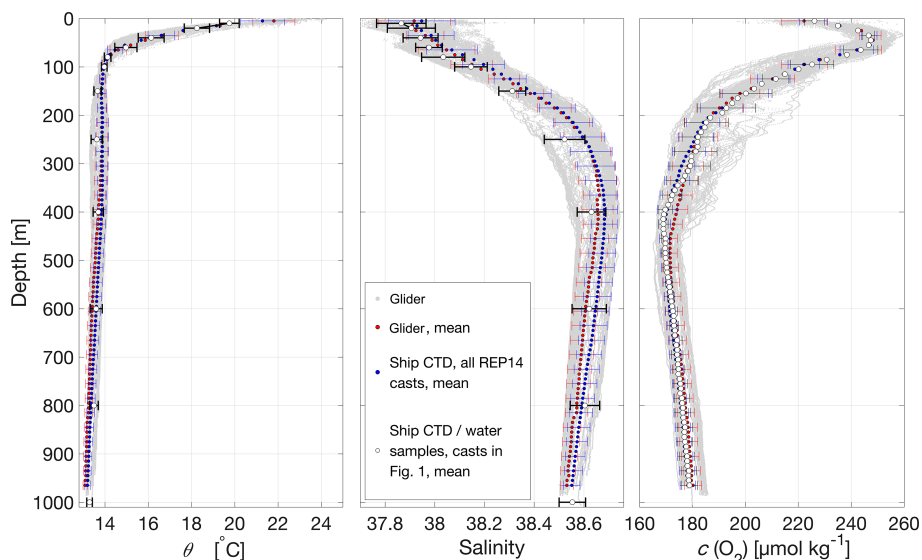


Figure 6. A comparison between the measurements retrieved by the glider and those obtained by the ship CTD package. The binned means (red) calculated using glider measurements (grey) are compared with the binned means of the CTD casts obtained from the entire REP14-MED observational domain (blue) and the binned mean values obtained from water samples (SBE oxygen optode sensor for dissolved oxygen concentrations ($c(\text{O}_2)$) during the eight CTD casts in Fig. 1 (white) for (a) potential temperature (θ), (b) salinity, and (c) $c(\text{O}_2)$. Standard deviations (calculated for every 10 m bin) are displayed as error bars in this figure every 30 m for glider and CTD measurements and at every sampled depth for water samples.

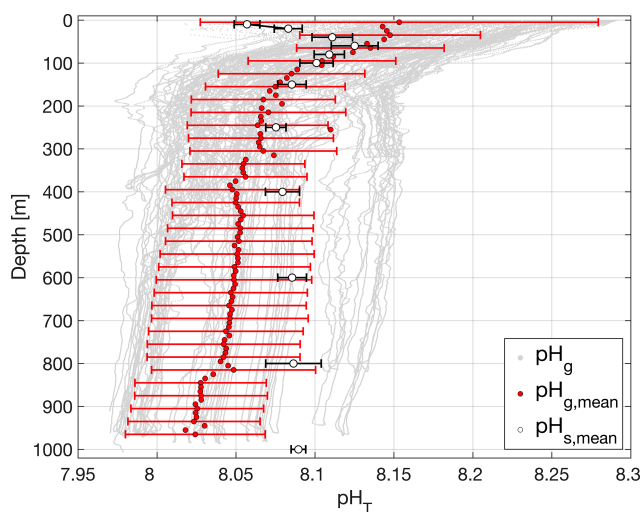


Figure 7. The pH obtained by the glider ISFET sensor (pH_g ; grey) is compared with the depth-binned mean of these profiles (red) along with the corresponding standard deviation error bars (using 10 m bins) displayed in this figure every 30 m. The mean pH values obtained by the ship (pH_s) during the eight CTD casts displayed in Fig. 1 are shown (white), with their relevant standard deviations displayed as error bars. The mean pH_g values and their corresponding standard deviations were calculated using 10 m depth bins, whereas the mean pH_s values and standard deviations were calculated at each sampled depth.

tween 05:00 and 21:00 LT, representing roughly 5 % of all pH_g measurements, were not used in later analysis. In order to reduce this light effect on pH measurements in future, ISFET sensors will have to be placed on the underside of the glider or equipped with a light shield.

Comparing pH_g to pH_s indicated that the range observed by the ISFET sensor was much larger. It could be argued that this difference in range is due to the differing temporal and spatial resolution between the glider and the ship measurements. However, comparing pH_g further with pH measurements in the literature on a similar timescale and spatial scale (Álvarez et al., 2014) suggests that this is not an issue with resolution. The $\text{pH}_{T,25}$ collected in the western Mediterranean Sea over a period of around 2 weeks (Álvarez et al., 2014), which is comparable in length to this trial, varied by roughly 0.02 at the surface and by around 0.08 at depths greater than 100 m. The range observed by the glider ISFET sensor was therefore 13 times larger at the surface and roughly 3 times larger at depths below 100 m. This difference in range cannot be explained by the high sampling frequency of the glider. Furthermore, the larger variations in pH_g were not a result of changing environmental conditions, as evidenced by the relatively stable $c(\text{O}_2)$, θ , and salinity measured by the glider, as discussed in Sects. 3.2.1 and 3.2.3.

It is likely that the ISFET pH measurements were not only related to the amount of hydrogen ion activity in the water, but also to the temperature and pressure that the sensor experienced, which was unexpected considering the sensor has previously shown good temperature and pressure character-

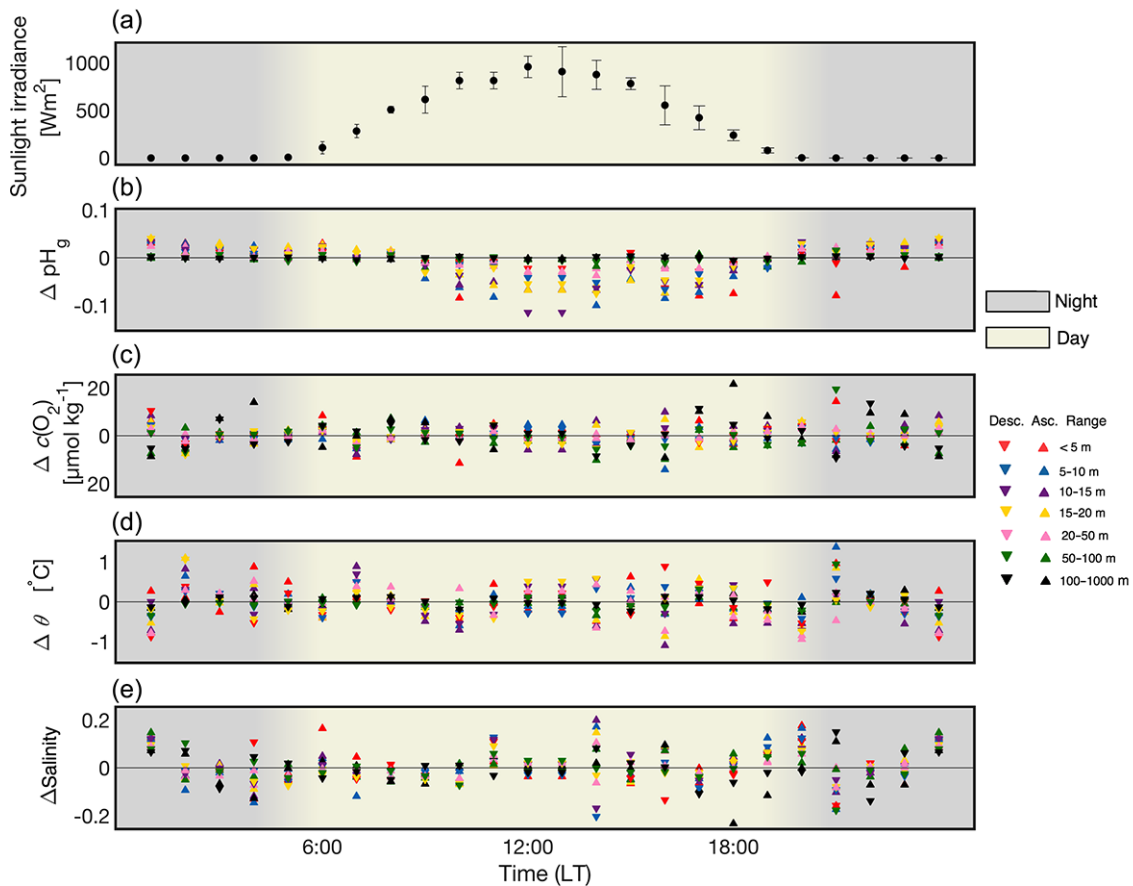


Figure 8. (a) Solar irradiance measured using a pyranometer on meteorological buoy M1 (Fig. 1). (b) Glider-retrieved pH (pH_g), (c) dissolved oxygen concentrations ($c(\text{O}_2)$), (d) potential temperature (θ), and (e) salinity average anomalies (calculated by subtracting the all-time mean from the hourly means within a given depth interval) for each hour of the day in local time (LT) for five near-surface depth ranges: < 5 m, 5–10, 10–15, 15–20, and 20–50 m, as well as two deeper depth ranges of 50–100 and 100–1000 m for both ascending (upward triangle) and descending (downward triangle) dive profiles. The grey shaded area represents the nighttime, whilst the lightly shaded area represents the daytime.

istics (Shitashima et al., 2002, 2013; Shitashima, 2010). Furthermore, comparing ISFET measurements with the pH_s and pH presented by Álvarez et al. (2014) suggests that the accuracy of the sensor was not as good as previously claimed (Shitashima et al., 2002). Therefore, it was necessary to correct the pH_g measurements for instrumental drift, temperature, and pressure.

The response of the ISFET sensor can be described by the Nernst equation (Eq. 1), which relates sensor voltage to hydrogen ion activity:

$$E = E^* - m_N \lg(a(\text{H}^+) a(\text{Cl}^-)), \quad (1)$$

which incorporates the Nernst slope (Eq. 2),

$$m_N = RT \ln(10)/F, \quad (2)$$

where T is temperature (K), R is the gas constant ($8.3145 \text{ J K}^{-1} \text{ mol}^{-1}$), F is the Faraday constant (96485 C mol^{-1}), $a(\text{H}^+)$ and $a(\text{Cl}^-)$ are the proton

and chloride ion activities, E is the measured voltage by the sensor (i.e. electromotive force), and E^* is representative of the two half-cells in the ISFET sensor forming a circuit (i.e. interface potential) (Martz et al., 2010). It is known that temperature and pressure have an effect on E^* (strong linear relationship) and that the Nernst slope is a function of temperature. Studies have also shown that it is possible for ISFET sensors to experience some form of hysteresis as a result of changing T and pressure (Martz et al., 2010; Bresnahan et al., 2014; Johnson et al., 2016).

The first step in correcting pH_g aimed to reduce the measured extent of variability to within the measured limits of pH_s . This in part removed the non-monotonous instrumental drift experienced by the sensor, which we think was likely due to the E^* between the two n-type silicon parts of the semiconductor being affected. A depth-constant time-varying offset correction (i.e. one offset value determined for each dive and applied to the entire profile) was applied

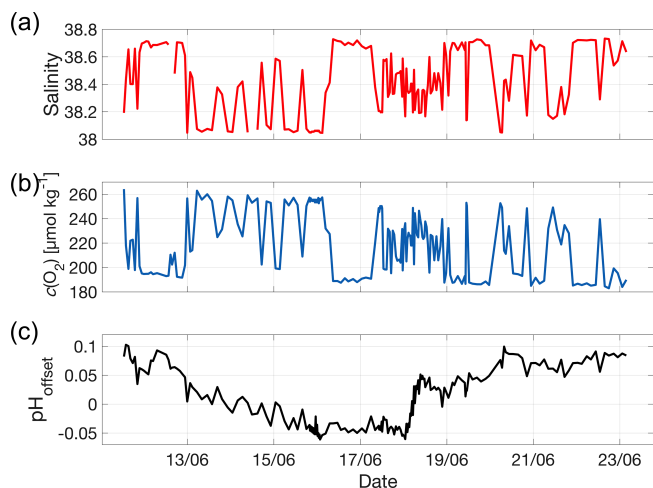


Figure 9. Salinity (a), dissolved oxygen concentrations ($c(\text{O}_2)$) (b), and the calculated pH offset values (c) as a function of time at the depth where the in situ temperature was $14 \pm 0.1^\circ\text{C}$.

(Eq. 3) using the difference between the mean pH_s and each pH_g dive measurement where the in situ temperature was 14.0°C , as water with this temperature was situated at a depth below the thermocline for most dives where the density gradient was weak:

$$\text{pH}_{\text{Offset}} = \text{pH}_s(T)_{\text{mean}} - \text{pH}_g(T) \text{ for } T = 14 \pm 0.1^\circ\text{C}. \quad (3)$$

The calculated offset values as a function of time were compared with salinity and $c(\text{O}_2)$ where the in situ temperature was constant at 14°C (Fig. 9). Variability in salinity and $c(\text{O}_2)$ with time were strongly related ($r^2 = 0.97$), whereas the relationship between pH offset values, salinity, and $c(\text{O}_2)$, were not (r^2 of around 0.2). Furthermore, the majority of the offset values were calculated below 100 m (Fig. 12d–f) where the density and pH gradients were weak. This indicated that our depth-constant time-varying offset correction decreased the apparent range of pH variability by an amount that was mostly associated with instrumental drift rather than physical and biogeochemical variability. Applying these offsets to the data decreased the range of pH measured by the ISFET sensor by approximately two-thirds (Fig. 11), with new pH_g standard deviations ranging between 0.009 and 0.048 within the top 150 m and between 0.008 and 0.017 at greater depths.

After applying this offset correction, pH_g was further corrected for in situ temperature and pressure using linear regression models. The method is outlined below.

1. Calculate ΔpH (Eq. 4) as the difference between the mean pH_s and pH_g :

$$\Delta\text{pH} = \text{pH}_{s,\text{mean}} - \text{pH}_g. \quad (4)$$

2. Determine the line of best fit between ΔpH and in situ temperature in the top 100 m of the water column where

the temperature gradient was the strongest using linear regression.

3. Correct pH_g for in situ temperature for the entire water column using the slope (m) and intercept (c) coefficients of the best fit line in step 2 to obtain $\text{pH}_{g,\text{tc}}$, where “tc” stands for “temperature-corrected” values.
4. Calculate the difference between $\text{pH}_{g,\text{tc}}$ profiles and mean pH_s , producing $\Delta\text{pH}_{\text{tc}}$, using an equation similar to Eq. (4).
5. Determine the line of best fit between $\Delta\text{pH}_{\text{tc}}$ and pressure for the lower 900 m of the water column using linear regression.
6. Correct $\text{pH}_{g,\text{tc}}$ for pressure for the entire water column using the coefficients m and c in a similar way to step 3 to obtain $\text{pH}_{g,\text{tpc}}$, where “tpc” stands for “temperature- and pressure-corrected” values.

The derived equation used for correcting pH_g is as follows:

$$\text{pH}_{g,\text{tpc}} = \text{pH}_g - 0.021t/^\circ\text{C} + 4.5 \times 10^{-5} P/\text{dbar} + 0.261, \quad (5)$$

where t is in situ temperature and P is pressure. A good fit was found between pH_g and in situ temperature, and a reasonable fit was found with pressure (Fig. 10). The standard deviations of $\text{pH}_{g,\text{tpc}}$ ranged between 0.008 and 0.039 in the top 150 m of the water column and between 0.007 and 0.013 at greater depths, corresponding to a further decrease in apparent variability of 13 to 23 and 14 to 31 % respectively (Fig. 11).

Johnson et al. (2016) ran a series of temperature and pressure cycling experiments when testing an ISFET pH sensor based on the Honeywell Durafet ISFET. They found a temperature sensitivity of around $\partial\text{pH}/\partial T = -0.018 \text{ K}^{-1}$, which is similar to our calculated slope, and a pressure hysteresis of 0.5 mV (pH of around 0.01) at maximum compression (2000 dbar). This is equivalent to a pressure sensitivity of roughly $\partial\text{pH}/\partial P = 5 \times 10^{-6} \text{ dbar}^{-1}$, which is an order of magnitude smaller than in this study. This difference in pressure sensitivity could be related to the different housing materials used, as Johnson et al. (2016) used polyether ether ketone (PEEK), whereas acrylic resin was used for our sensor.

Salinity covaries with temperature and pressure, and some of the salinity dependence of the offset between pH_s and pH_g might have been misattributed to the regression coefficients associated with temperature and pressure. The sensor characteristics should therefore be studied in detail under controlled laboratory conditions. However, for the purposes of calibrating the high-resolution poor-accuracy measurements (relative to pH_s) obtained from the ISFET pH sensor, the present empirical correction based on temperature and pressure appears to be sufficient to achieve a match within the pH repeatability of the discrete samples between 0.004 and 0.017.

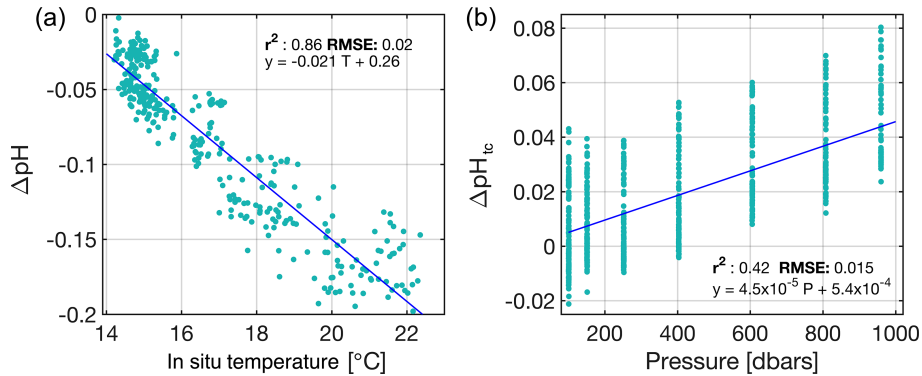


Figure 10. Linear regression fits are displayed for (a) ΔpH (the difference between the mean pH_S and pH_g corrected for drift) vs. in situ temperature in the top 100 m of the water column and (b) ΔpH corrected for in situ temperature ($\Delta\text{pH}_{\text{TC}}$) vs. pressure between 100 and 1000 m using all available dives. The r^2 , root mean square error (RMSE), and the equation of the line are displayed for each linear fit.

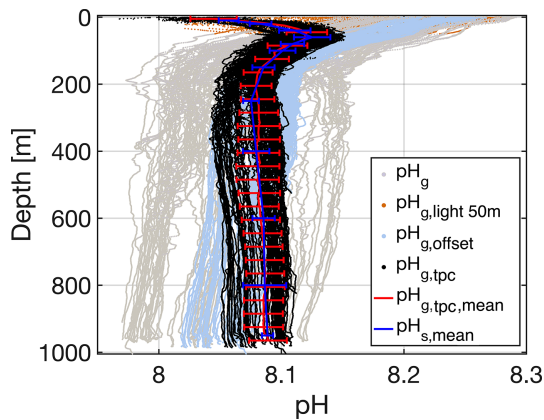


Figure 11. Profiles of glider-retrieved pH (pH_g) pre-correction (grey) with a depth-constant time-varying offset correction applied (light blue) and further corrected for in situ temperature and pressure (black). The pH_g measurements affected by light in the top 50 m of the water column (orange) and not used for the drift; temperature and pressure corrections are also shown. The depth-binned mean profile of drift, temperature, and pressure-corrected (tpc) pH_g (using 10 m bins) is shown in the foreground (red) along with the standard deviation ranges displayed every 40 m in this figure. The depth-binned mean profile of the pH measurements retrieved by ship (pH_S) is plotted for comparison (dark blue) with standard deviation ranges displayed at each sampled depth.

3.2.3 Coast to open ocean high-resolution hydrographic and biogeochemical variability

Spatial and temporal variability can be seen in $\text{pH}_{\text{g,tpc}}$ for three individual east–west transects using measurements obtained within different time periods (Fig. 12a–c). This pH variability is likely related to the air–sea exchange of carbon dioxide (weak), changes in temperature (indirectly), and biological activity (Yao et al., 2016). In the top 100 m, pH higher than 8.12 was found at depths ranging from 20 to 95 m, whereas lower pH values ranging from 8.06 to 8.09

were present closer to the surface at some locations (e.g. between 7.5 and 7.7° E and east of 8° E). The pH maxima were found at depths between 40 and 70 m where θ was around 15 °C (Fig. 12d–f) within the pycnocline (Fig. 12j–l). This band of high pH situated at 20 to 95 m of depth corresponded with a thick layer of $c(\text{O}_2)$ -rich water at similar depths (Fig. 12m–o). The pH and $c(\text{O}_2)$ in the top 200 m of the water column more or less followed isopycnal surfaces at a range of points in time and space. For example, the slanted isopycnals closer to the coast (east of 7.95° E) associated with geostrophic shear corresponded with horizontal gradients in pH and $c(\text{O}_2)$. Below 100 m, $c(\text{O}_2)$ decreased to a minimum of $< 170 \mu\text{mol kg}^{-1}$; although not spatially homogeneous, this corresponded with generally colder, saltier, and lower-pH waters.

All three east–west transects can be separated into two parts roughly either side of 7.7° E for depths greater than 100 m. Lower pH values between 8.05 and 8.1 were found in the western part, whereas a higher pH ranging from 8.07 to 8.12 was found in the eastern part, which was partially seen in the pH_S measurements (Fig. 5). The spatial variability in these eastern and western parts differed for each of the three time periods (times labelled in Fig. 12) with both the eastern high and western low pH patches changing in size vertically and horizontally, corresponding to spatial changes in θ and salinity. Furthermore, salinity, θ , and $c(\text{O}_2)$ were lower in the western part compared with the values found at similar depths in the eastern section (Fig. 12d–i, m–o).

In the top 100 m of the water column, the variability in pH and $c(\text{O}_2)$ is likely related to biological activity and air–sea gas exchange. As discussed in Sect. 3.1, a DCM within this depth range is common in the Mediterranean Sea when waters are thermally stratified and sufficient nutrients and light are available below the mixed layer (Estrada, 1996). High chlorophyll fluorescence was observed by the ship’s sensor here (Fig. 4d). Enhanced $c(\text{O}_2)$ values at these depths are likely the by-product of photosynthesis, and the higher pH

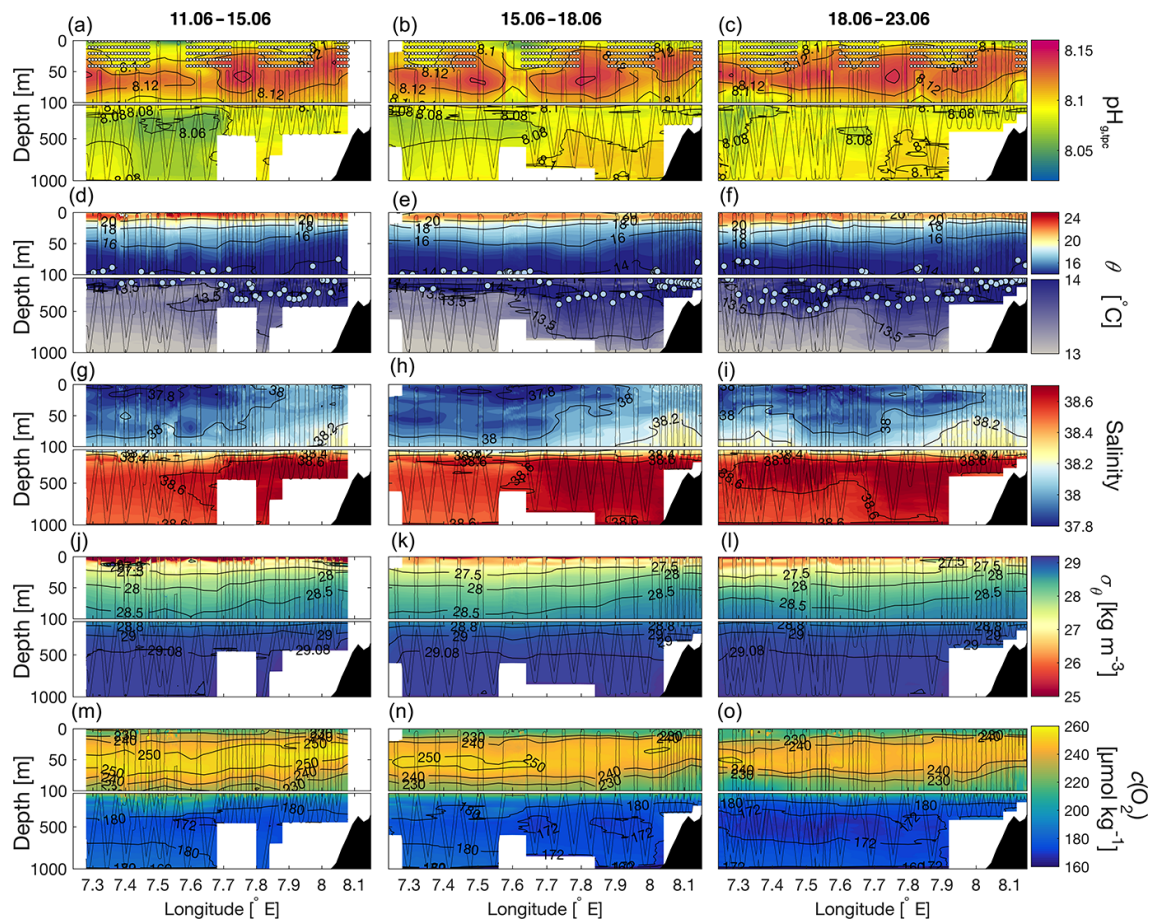


Figure 12. Objectively mapped transects of glider-retrieved (a–c) pH corrected for drift, temperature, and pressure ($\text{pH}_{g,tpc}$). (d–f) Potential temperature (θ), (g–i) salinity, (j–l) potential density anomalies (σ_θ), and (m–o) dissolved oxygen concentrations ($c(O_2)$) for three different time periods between 11 and 15, 15 and 18, and 18 and 23 June 2014. The spatial ranges of pH measurements affected by light and removed prior to corrections are represented by small white points (a–c). The depth–longitude points at which pH offsets were calculated at a temperature of 14 °C are indicated by pale blue points (d–f). Glider measurements were sorted into 0.04° longitude \times 2 m bins, and the radius of influence used for optimal interpolation was 0.1° longitude \times 10 m. Glider measurements used for optimal interpolation are superimposed on the interpolated fields for reference.

values are likely the result of changes in the carbon equilibrium due to the consumption of CO_2 (Cornwall et al., 2013; Rivarolo et al., 2010; Copin-Montégut and Bégovic, 2002). A similar relationship between pH and primary production was described by Álvarez et al. (2014) in the western Mediterranean Sea. As discussed in Sect. 3.1, the fresher waters found in the top 100 m are likely MAW.

The difference in pH between the eastern and western parts at depths greater than 100 m depth highlighted the variability of water masses in this region. In particular, the higher pH found in the eastern part of the transect (east of 7.7° E), coinciding with high A_T and $c(\text{DIC})$ (Fig. 5), was likely related to the flow of LIW, as described in Sect. 3.1. The LIW flows from the eastern Mediterranean basin (east of the Strait of Sicily) where pH is higher than in the western Mediterranean basin (Álvarez et al., 2014) towards the west along the continental shelf edge (Millot, 1999). This high pH found

in the eastern section of the glider transect may therefore be a remnant of these eastern Mediterranean waters. The low-pH low- $c(O_2)$ waters found deeper than 100 m result from increased respiration and remineralisation of organic matter (Lefèvre and Merlivat, 2012), coinciding with higher levels of $c(\text{DIC})$ deeper than 200 m (Merlivat et al., 2015), which may have been more prominent in the western part of the transect (west of 7.7° E) leading to decreased levels of pH.

The pycnocline shallowed east of 7.7° E in the top 100 m of the water column during all three time periods (times labelled in Fig. 12), which corresponded with shoaling, high-salinity, low-pH, low- $c(O_2)$ waters and high $c(\text{DIC})$, A_T , and chlorophyll fluorescence obtained by ship (Figs. 4 and 5). These features may be related to upwelling. Meteorological buoy M1 located south of the glider transect recorded an average surface wind direction of 198° towards the south-southwest, which would be favourable for coastal upwelling.

However, the mean wind speed was only 2 m s^{-1} , which is weak. On the other hand, salinity maxima seen at depths of 200 to 700 m seem to suggest an intrusion of LIW westward. An intrusion of water away from the coast towards the open ocean has been shown to increase divergence in regions close to shore with strong alongshore currents (Roughan et al., 2005). Upwelling signatures at this longitudinal range along the Sardinian coast have been simulated, particularly in the summer, by Olita et al. (2013) using a hydrodynamic 3-D mesoscale-resolving numerical model. They suggest that a mixture of both current flow and wind preconditioned and enhanced upwelling in this region, which may have also been the case during our deployment. Furthermore, chlorophyll fluorescence (Fig. 4) obtained by ship was higher closer to shore, which is indicative of a greater abundance of biomass in the top 100 m, perhaps fuelled by upwelled nutrients (Porter et al., 2016; El Sayed et al., 1994).

4 Conclusions

Our trials of an experimental pH sensor in the Mediterranean Sea uncovered instrumental problems that were unexpected and will need to be addressed in future usage. These are summarised here.

1. The data retrieved from the dual pH– $p(\text{CO}_2)$ integrated sensor and from the $p(\text{CO}_2)$ unit of the stand-alone dual sensor could not be used due to quality issues. It is unclear why there was a problem with the measurements obtained by the stand-alone $p(\text{CO}_2)$ unit; however, we think the regular on/off cycling of power to the integrated dual pH– $p(\text{CO}_2)$ sensor between sampling did not allow it to function properly. In future, we would suggest the addition of backup batteries to supply electricity to the sensor between sampling.
2. The stand-alone pH sensor was subject to drift. This could be reduced by subtracting a depth-constant time-varying offset from each dive using the difference between pH_g and pH_s at a more dynamically stable depth, but such an approach is not generally recommended or valid. We think that a change in E^* between the two n-type silicon parts of the semiconductor might be the cause of the drift. To elucidate this drift further, in future two ISFET sensors should be tested in laboratory conditions within a bridge circuit to attempt to isolate possible factors contributing to drift. Focussing on the root cause of the sensor drift rather than correcting the pH data for drift after the deployment would be more beneficial to the long-term study of ISFET pH– $p(\text{CO}_2)$ sensors.
3. The sensor was apparently affected by temperature and pressure, but it is unclear to what extent the empirical relationship between in situ temperature and ΔpH in

the thermocline (top 100 m) and between pressure and $\Delta\text{pH}_{\text{TC}}$ in the deeper water (100–900 m) can be generalised.

4. The effect of light caused the sensor to measure lower levels of pH_g in surface waters. This effect is expected to be ubiquitous wherever the sensor nears the surface during daytime. In future, the sensor will have to be positioned on the underside of the glider or equipped with a light shield to limit the effect of the sun when close to the surface.

Despite the overall disappointing performance, we were able to demonstrate the potential use of the corrected glider pH measurements to uncover the biogeochemical variability associated with biological and physical mesoscale features. The pH_g corrected for drift, temperature, and pressure was compared temporally and spatially with the other physical and biogeochemical parameters obtained by the glider. This comparison indicated that the pH in the top 100 m of the water column was mostly related to biological activity where $c(\text{O}_2)$ was high. Below 100 m, low pH west of 7.7° E was likely linked to the remineralisation of organic matter, whilst east of this point, higher pH may have been transported from the eastern Mediterranean basin via LIW. Shoaling isopycnals east of 7.7° E closer to shore may have been indicative of upwelling, and possible upwelling signatures at the same location could be seen in salinity, θ , pH, $c(\text{O}_2)$, $c(\text{DIC})$, A_T , and chlorophyll fluorescence.

Data availability. All REP14-MED experiment data are available on the CMRE data server at <http://geos3.cmre.nato.int/portal/> (NATO Centre for Maritime Research and Experimentation (CMRE), 2016). The data are NATO UNCLASSIFIED and available only for the partners of the experiment. However, interested institutions can sign up for partnership at any time. Requests may be directed to the author or to geos-webmaster@nrc.nato.int.

Competing interests. The authors declare that they have no conflict of interest.

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