

### A new Burmese amber hawker dragonfly helps to redefine the position of the aeshnopteran family Burmaeshnidae (Odonata: Anisoptera: Aeshnoidea)

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1	A new Burmese amber hawker dragonfly helps to redefine the position of the aeshnopteran
2	family Burmaeshnidae (Odonata: Anisoptera: Aeshnoidea)
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16	
17	ABSTRACT
18	The new genus and species Angustaeshna magnifica of Burmaeshnidae is described on the basis
19	of a new fossil from Burmese amber. The genus Cretaeshna from the same amber is transferred
20	from the Telephlebiidae into the Burmaeshnidae. We redefine this last family, no longer
21	considered as the sister group of the Late Cretaceous Enigmaeshnidae, but as putative sister
22	group of the Telephlebiidae in the Aeshnoidea. No known fossil belongs to the Telephlebiidae.
23	
24	Keywords: Insecta; Aeshnoptera; gen. et sp. nov.; Myanmar; phylogeny
25	

#### 26 **1. Introduction**

27 Mesozoic Aeshnoptera are very diverse but they mainly belong to the stem group of the extant clade Aeshnodea Bechly, 1996. Only two Allopetaliidae Cockerell, 1913, one Telephlebiidae 28 29 Cockerell, 1913, and no Aeshnidae Leach, 1815 are recorded in the Cretaceous (Bechly et al., 2001; Zheng et al., 2017). Huang et al. (2017) recently described the new family Burmaeshnidae 30 from the Burmese amber, on the basis of the basal parts of a fore- and a hindwing, and 31 32 tentatively considered it as the sister group of the Late Cretaceous family Enigmaeshnidae Nel et al., 2008. It was not possible to compare Burmaeshna azari Huang et al., 2017 to the alleged 33 telephlebiid Cretaeshna Zheng et al., 2017 because the latter is based on distal parts of wings. 34 35 Here we describe a third hawker dragonfly from the Burmese amber, showing nearly complete fore- and hindwings. This fossil allows to compare Burmaeschna azari with Cretaeshna and to 36 put all these in the same family Burmaeshnidae, as sister group of the extant Telephlebiidae. 37

38

#### 39 2. Materials and methods

The fossil was examined and measured using an incident light stereomicroscope (Olympus SZX9) and a stereomicroscope (Nikon SMZ 1500), as well as a Leitz Wetzlar binocular microscope. Photographs were taken using a Zeiss Discovery V20 microscope system. Optical instruments were equipped by camera lucida and digital cameras. The raw digital images were processed with focus stacking software, and figure plates prepared with Adobe Photoshop<sup>TM</sup>.

The nomenclature of the odonatan wing venation used in this paper is based on the interpretations of Riek & Kukalová-Peck (1984), as modified by Nel et al. (1993) and Bechly (1996). The higher classification of fossil and extant Aeshnoptera follows Bechly et al. (2001). Wing abbreviations are as follows: CuA, cubitus anterior; IR1, intercalary radial veins; MA, median anterior; MP, median posterior; N, nodus; Pt, pterostigma; RA, radius anterior; RP, radius posterior; Sn, subnodal crossvein. All measurements are given in mm. The specimen is preserved in a piece of relatively clear, yellow Burmese amber. The amber piece was polished before being examined and photographed. All amber material was legally acquired in Myanmar from local traders with government registration, and legally exported according to the official regulations in Myanmar.

Fossil-bearing has mostly been collected from the Hukawng Valley in northern 55 Myanmar (formerly known as Burma). For an overview of the amber deposit and its geological 56 57 setting see, e.g., Zherikhin and Ross (2000), Grimaldi et al. (2002), Cruickshank and Ko (2003), and Ross et al. (2010). Radiometric U-Pb zircon dating (Shi et al., 2012) recently constrained 58 this amber to a minimum age of 98.79±0.62 Ma, which is equivalent to the mid-Cretaceous 59 60 (earliest Cenomanian). The original habitat of the amber forest is still controversial, in fact it has originally been assumed to be a tropical araucarian forest (Grimaldi et al., 2002; Poinar et 61 al., 2007), possibly with Dipterocarpaceae as another source for the fossil resin. However, the 62 63 first detailed report on the macromolecular nature and palaeobotanical affinity of Burmite (Dutta et al., 2011), based on gas chromatography - mass spectrometry, rejected Araucariaceae 64 65 and Dipterocarpaceae in favour of Pinaceae as the Burmese amber tree. Grimaldi (2016), after Grimaldi and Ross (in press), considered 'based on the abundant inclusions of leafy shoots' that 66 it was formed by a conifer, and 'amber produced possibly by Metasequoia (Taxodiaceae) or a 67 68 close relative'.

69 The family, genus, and species are registered in Zoobank under the urn: xxxx

70

#### 71 **3. Systematic palaeontology**

72 Odonata Fabricius, 1793

73 Anisoptera Selys in Selys and Hagen, 1854

74 Aeshnoptera Bechly, 1996

75 Burmaeshnidae Huang et al., 2017

- 76 Type species. *Burmaeshna azari* Huang et al., 2017
- 77 Other taxa. Cretaeshna lini Zheng et al., 2017 nov. sit., transferred from Telephlebiidae into
- 78 Burmaeshnidae; *Angustaeshna magnifica* gen. et sp. nov.
- 79 *Emended diagnosis*. Two characters have to be added to the diagnosis of the Burmaeshnidae:
- 80 IR2 with a distal fork; accessory anal loop between the two main branches of CuA in hindwing
- 81 present.
- 82
- 83 Angustaeshna gen. nov.
- 84 Type species: *Angustaeshna magnifica* sp. nov.
- 85 *Etymology*. Named after *angustus*, narrow in Latin for the very narrow wings, and *Aeshna*.

86 *Diagnosis*. Wing venation characters only. One oblique veins 'O'; accessory anal loop between

the two main branches of CuA in hindwing very well-defined; only one antesubnodal crossvein

distal of base of IR2; discoidal triangles divided into two smaller cells; base of IR1 1-2 cells

89 basal of pterostigma.

- 90
- 91 *Angustaeshna magnifica* sp. nov.
- 92 Figs. 1-5

*Holotype*. NIGP166238 (two incomplete forewings and a complete hindwing), stored at the
Nanjing institute of Geology and Palaeontology.

95 Locality and Horizon. Hukawng Valley, Kachin Province, Myanmar; late Upper Albian to

- 96 lowermost Cenomanian, Upper Cretaceous.
- 97 Etymology. Named after the wonderful state of preservation of the wings.
- 98 *Diagnosis*. As for genus. Hindwing discoidal triangle divided into two cells.
- 99 Description. Mid part of a forewing and distal half of the other one, hyaline, wing ca 31.7 mm
- long, 6.1 mm wide; distance from base to arculus ca. 3.5 mm; from arculus to base of RP3/4

6.0 mm; from arculus to nodus 6.4 mm; nodal crossvein and subnodus strongly oblique, ScP 101 102 making a right angle in nodus; six preserved antenodal crossveins distal of Ax2, the first one being completed while the others are not aligned with those of second row between ScP and 103 104 RA, Ax2 between arculus and discoidal triangle, a secondary antenodal crossvein between Ax2 and Ax1, Ax1 not preserved; anterior part of arculus slightly curved, RP and MA separated in 105 angle of arculus where posterior part of arculus touches anterior part; hypertriangle 3.5 mm 106 107 long, free; discoidal triangle 1.0 mm distal of arculus, divided into two smaller cells, not very elongate and narrow, with anterior side 2.5 mm long, and MAb 2.2 mm long, with a strong 108 angle from which emerges distinct convex trigonal planate, short and zigzagged; postdiscoidal 109 110 area with two rows of cells just after discoidal triangle and three more distally; Mspl welldefined with one row of cells between it and MAa; base of Mspl four cells distal of discoidal 111 triangle; basal part of area between RA and RP with four crossveins basal of RP3/4, and two 112 distal of base of RP3/4 and basal of subnodus; area between RP and MA with three crossveins 113 basal of RP3/4; Bq space long, but two Bq crossveins; median area free; submedian area crossed 114 by curved CuP; subdiscoidal space free; one row of cells between MP and CuA; two rows of 115 cells in a narrow anal area; two rows of cells in area below CuA; CuA without well-defined 116 posterior branches; base of RP2 aligned with subnodus; RP2 straight; one row of cell between 117 118 RP2 and RP1; IR2 with a clear distal fork, and two rows of cells between its branches; Rspl well-defined straight, with one row of cell between it and IR2; one oblique vein 'O' one cell 119 distal of subnodus; six preserved posnodal crossveins, not aligned with the eight postsubnodal 120 121 crossveins; pterostigma and wing apex not preserved in one forewing, present on the second one, covering one cell and a half; a short pseudo-IR1 with its base below middle of pterostigma 122 123 but aligned with a zigzagged vein that begins two cells basal of pterostigma.

Hind wing complete, hyaline, 30.0 mm long, 7.3 mm wide; with a rather long petiole,
with stem of anal vein 2.2 mm long; distance from base to arculus 5.5 mm; from arculus to

nodus 7.7 mm; from arculus to base of RP3/4 5.3 mm; four antenodal crossveins of primary 126 127 type, viz. with a triangular membrane between C, ScP and radius, Ax0 at extreme base of wing, Ax1 3.2 mm distally, 'Ax2' 3.5 mm distally and a supplementary one 'Ax3' 1.7 mm distally, 128 weaker than others and with a weaker membrane between C, ScP and RA; a secondary 129 antenodal crossvein between Ax1 and Ax2 and three secondary antenodal crossveins of first 130 row not well aligned with the three crossveins of second row, distal of most distal primary 131 132 antenodal; 11 postnodal crossveins not well aligned with postsubnodal crossveins; one row of cells between RP1 and RP2; nodal veins with the same pattern as in forewing; arculus 1.1 mm 133 basal of Ax2; Ax2 aligned with basal side of discoidal triangle; RP and MA separated in angle 134 135 of arculus where posterior part of arculus touches curved anterior part; hypertriangle 3.9 mm long, free; discoidal triangle 1.1 mm distal of arculus, divided into two smaller cells, more 136 elongate and narrower than in forewing, with basal side 1.2 mm long, anterior side 2.8 mm 137 long, and MAb 2.2 mm long, sigmoidal; postdiscoidal area with two rows of cells just after 138 discoidal triangle, distally broadened with 10 rows of cells along posterior wing margin; a 139 140 distinct convex trigonal planate, short and zigzagged; Mspl well-defined, straight, with one row of cells between it and MAa; base of Mspl three cells distal of discoidal triangle; basal part of 141 area between RA and RP with two crossveins basal of RP3/4, and two distal of base of RP3/4 142 143 and basal of subnodus; area between RP and MA with two crossveins basal of RP3/4; one oblique vein "O", one cell distal of base of RP2; one row of cells between IR2 and RP3/4 at 144 least till two cells distal of subnodus; base of RP2 aligned with subnodus; Rspl straight with 145 one row of cells between it and IR2; IR2 forked well basal of pterostigma, two rows of cells 146 between its branches; pterostigma short, 1.5 mm long, 0.8 mm wide, covering one cell and a 147 half; pterostigmal brace aligned with basal side of pterostigma and oblique; a short pseudo-IR1 148 with its base below middle of pterostigma but aligned with a zigzagged vein that begins one 149 cell basal of pterostigma; one row of cells between MP and CuAa; median area free; submedian 150

area crossed by curved CuP, situated between Ax1 and Ax2; subdiscoidal space free; no anal 151 152 triangle; no clear anal angle (female specimen?); postero-basal wing margin (AP) nearly straight; anal area very long but not very broad, 5.5 mm long, 2.8 mm wide, with three rows of 153 large cells between AA and AP basal of anal loop and no clear posterior branches of AA; anal 154 loop two cells broad, much more elongate than broad, 3.9 mm long, 1.5 mm wide, hexagonal, 155 posteriorly closed, divided into five cells; CuAb well-defined; cubito-anal area narrow, with 156 157 three rows of cells between CuAa and posterior wing margin; CuAa with only one strong posterior branch that delimitates a clear subanal loop. 158

*Discussion. Angustaeshna* gen. nov. can be attributed to the Neoaeshnida Bechly, 1996 (= Gomphaeschnidae Tillyard & Fraser, 1940 + Aeshnodea Bechly, 1996) because of the very elongate discoidal triangles; presence of only one row of cells between RP1 and RP2; welldefined Mspl and Rspl, parallel to MA and IR2; both pairs of wings with a strong convex secondary longitudinal vein (trigonal planate) in postdiscoidal area; in both pairs of wings MP and CuA closely parallel with only one row of cells between them up to wing margin; only one oblique vein 'O' (Bechly, 1996; 2016).

Angustaeshna has several characters currently considered as synapomorphies of the 166 Gomphaeschnidae, viz. the most distal part of antesubnodal area between RA and RP free of 167 168 antesubnodal crossveins (such a 'cordulegastrid gap' is not present in the fossil gomphaeschnid Alloaeschna quadrata Wighton and Wilson, 1986); no accessory cubito-anal crossveins in 169 submedian space between CuP and PsA; discoidal triangles only divided into two cells by a 170 single crossvein; hypertriangles secondarily unicellular. Nevertheless, Angustaeshna has also a 171 crucial apomorphy of the Eueshnodea Bechly et al., 2001, viz. IR2 with a distal dichotomic 172 furcation, absent in all Gomphaeschnidae, recent and fossil. It cannot be attributed to the 173 Brachytronidae Cockerell, 1913 because its pterostigmal brace is not reduced and MP and CuAa 174 are distally not divergent. Angustaeshna has also a crucial synapomorphy of the Aeshnoidea 175

Leach, 1815, viz. a very well-defined accessory anal loop between the two main branches of
CuA in hindwing, also absent in all Gomphaeschnidae. *Angustaeshna* also has a synapomorphy
(proposed by Bechly, 1996, 2016) of the extant Telephlebiidae, viz. Ax2 recessed to basal angle
of discoidal triangle (close to, at, or even basal).

Nevertheless, Angustaeshna strongly differs from all the recent representatives of the 180 Telephlebiidae in the shape of the anal loop, clearly longer than wide, while it is wider than 181 182 long in the extant Telephlebiidae, as in the Aeshnidae. Also, its anal stem is distinctly longer than in the modern Aeshnoidea. These structures are synapomorphies of the Burmese amber 183 family Burmaeshnidae Huang et al., 2017, together with the presence of three primary antenodal 184 185 crossveins Ax1, 'Ax2' and 'Ax3' in hindwing. Peters & Theischinger (2007) proposed a series of synapomorphies for the Telephlebiidae, all based on body structures, unknown in these 186 187 fossils.

The genus Burmaeshna Huang et al., 2017 differs from Angustaeshna in several 188 characters, the most important being the presence of two oblique veins 'O' instead of one (a 189 190 character of the Neoaeshnida). This character is subject to homoplasies in the whole clade Aeshnoptera, so it is possibly also the case here. The accessory anal loop between the two main 191 branches of CuA in hindwing is less well-defined in Burmaeshna than in Angustaeshna. 192 Burmaeshna has also more antesubnodal crossveins distal of the base of IR2 than in 193 Angustaeshna; and discoidal triangles divided into three smaller cells instead of two in 194 195 Angustaeshna.

Huang et al. (2017) indicated that *Burmaeshna* shares with the Late Cretaceous family
Enigmaeshnidae the very elongate anal area with an anal loop distinctly longer than wide, as
putative synapomorphies. But *Enigmaeshna* Nel et al., 2008 strongly differs from *Burmaeshna*and *Angustaeshna* in the complete absence of the accessory anal loop between the two main
branches of CuA; absence of a stem of anal vein; presence of numerous crossveins in the

hindwing subdiscoidal space; presence of numerous secondary antenodal crossveins between
Ax1 and Ax2; Ax2 well distal of base of discoidal triangle; and absence supplementary primary
antenodal crossvein in hindwing. Thus the similar shapes of the anal loops of *Enigmaeshna*with those of *Angustaeshna* and *Burmaeshna* is a parallelism.

In conclusion, we consider that *Angustaeshna* belongs to the family Burmaeshnidae, but to a genus different from *Burmaeshna*. Also the family Burmaeshnidae belongs to the clade Aeshnoidea (Telephlebiidae + Aeshnidae), and is probably the sister group of the modern family Telephlebiidae for the Ax2 recessed close to the basal angle of discoidal triangle.

Zheng et al. (2017) described the Burmese amber genus *Cretaeshna* on the basis of weak 209 210 arguments, viz. a distal half of a wing. They attributed it to the family Telephlebiidae on the basis of the shape of the nearly straight vein Rspl; absence of the bulge in the distal part of MAa 211 (plesiomorphies); and presence of a forked IR2 (a synapomorphy of the Eueshnodea). 212 213 Nevertheless the preserved structures of *Cretaeshna* are very similar to those of *Angustaeshna*, the unique clear difference being the shorter vein IR1 that begins below the pterostigma in 214 215 Cretaeshna while it begins 1-2 cells basal of pterostigma in Angustaeshna. Cretaeshna differs from Burmaeshna in the presence of only one oblique vein 'O'. The lack of information on all 216 the structures of the basal halves of the wings in Cretaeshna forbids us to better compare it to 217 218 Burmaeshna and Angustaeshna. Nevertheless the quasi identity in venation strongly indicates that Cretaeshna also belongs to the Burmaeshnidae. Therefore the family Telephlebiidae sensu 219 stricto is still unknown in the fossil record. 220

221

#### **4.** Conclusions

The Burmese amber family Burmaeshnidae is not related to the Enigmaeshnidae, but belongsto the Aeshnoidea, as putative sister group of the extant family Telephlebiidae, suggesting that

the modern aeshnoids began to diversify during the mid-Cretaceous, if the most diverse extantAeshnidae are still only known from the Paleogene.

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- 310 Figures
- Fig. 1. *Angustaeshna magnifica* gen. et sp. nov., holotype NIGP166238, photograph of two
  fore- and one hindwing. Scale bar = 2 mm.
- Fig. 2. Angustaeshna magnifica gen. et sp. nov., holotype NIGP166238, photograph of
  hindwing base. Scale bar = 0.5 mm.
- **Fig. 3**. *Angustaeshna magnifica* gen. et sp. nov., holotype NIGP166238, photograph of mid part
- 316 of hindwing. Scale bar = 1 mm.
- Fig. 4. Angustaeshna magnifica gen. et sp. nov., holotype NIGP166238, photograph of
  hindwing pterostigma. Scale bar = 0.2 mm.
- Fig. 5. Angustaeshna magnifica gen. et sp. nov., holotype NIGP166238, photograph of
  hindwing apex. Scale bar = 2 mm.
- 321

## forewing

# forewing

hindwing







# pseudo-IR1



### Pt brace

### IR1

# IR2 fork IR1 Pt brace

-

RP2

2 mm

Pt

Rspl