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Pascal Pigeon, Yong Wang, Siden Top, Feten Najlaoui, Maria Concepcion Garcia Alvarez, et al.. A New Series of Succinimido-ferrociphenols and Related Heterocyclic Species Induce Strong Antiproliferative Effects, Especially against Ovarian Cancer Cells Resistant to Cisplatin. *Journal of Medicinal Chemistry*, 2017, 60 (20), pp.8358-8368. 10.1021/acs.jmedchem.7b00743 . hal-01611795

HAL Id: hal-01611795

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A new series of succinimido-ferrociphenols and related heterocyclic species induce strong antiproliferative effects, especially against ovarian cancer cells resistant to cisplatin

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J. Med. Chem., **Just Accepted Manuscript** • DOI: 10.1021/acs.jmedchem.7b00743 • Publication Date (Web): 12 Sep 2017

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Abstract

Ferrociphenols are known to display anticancer properties by original mechanisms dependent on redox properties and generation of active metabolites such as quinone methides. Recent studies have highlighted the positive impact of oxidative stress on chemosensitivity and prognosis of ovarian cancer patients. Ovarian adenocarcinomas are shown to be an excellent model for defining the impact of selected ferrociphenols as new therapeutic drugs for such cancers. This work describes the syntheses and preliminary mechanistic research of unprecedented multitargeting heterocyclic ferrociphenols bearing either a succinimidyl or phthalimidyl group that show exceptional antiproliferative behavior against epithelial ovarian cancer cells resistant to cisplatin. Owing to the failure of the present pharmaceutical options, such as carboplatin a metallodrug based on Pt coordination chemistry, these species may help to overcome the problem of lethal resistance. Currently, ferrociphenolic entities generally operate via apoptotic and senescence pathways. We present here our first results in this new cyclic-imide series.

Keywords. antitumor agents, bioorganometallics, ferrocene, imides, quinones

Introduction

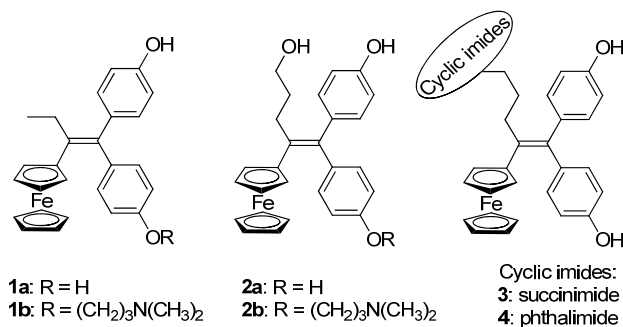
Burgeoning numbers of organometallic complexes bearing ferrocenyl moieties are currently appearing in the literature, with the intended goal of medicinal applicability.¹⁻¹² In this context, our long term project aims to study selected ferrociphenols¹³⁻¹⁵ chosen for their innovative behavior against intractable cancers. We here focus on epithelial ovarian cancer (EOC), a disease that remains a serious therapeutic roadblock with no significant advances because of the failure of the present pharmaceutical options. EOC remains one of the deadliest gynaecological malignancies; standard protocol consists of surgery and chemotherapy. Despite an initial response to therapy, many patients become resistant to treatment and ultimately die. However, although there is increasing evidence that precision therapy targeted to molecular cancer drivers can improve clinical outcomes, high-grade EOC patients are currently treated without consideration of molecular phenotype. Molecular features of high-grade serous EOC could be helpful in fulfilling this need.¹⁶⁻¹⁷ There is compelling evidence that redox regulation plays an essential role in malignancies. Recent studies by Mechta-Grigoriou et al. have highlighted the positive impact of oxidative stress on chemosensitivity and ovarian cancer prognosis.^{16, 18} Interestingly, the redox properties of new ferrociphenols could fit very well with these findings.

The original antiproliferative behavior of ferrociphenols lies in the redox properties of the [ferrocenyl-ene-phenol] motif (Scheme 1) producing ROS (Reactive Oxygen Species)¹⁹ on cancer cells, and subsequent generation of selective electrophiles such as QMs (Quinone Methides).²⁰⁻²³ These active metabolites are able to react with specific receptor proteins or enzymes such as thioredoxin reductases,²³ or AP1, IL-6, IL-8 signaling pathways,²⁴ and induce apoptosis and/or senescence.^{14, 23-25} These multitargeting species can inhibit or slow down resistance problems on cancer cells, and in addition are not at all, or poorly, active on healthy

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6 cells.¹⁴ Given (i) the crucial role of oxidative stress in the biology of some epithelial ovarian
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8 cancers,¹⁶ (ii) the resistance of ovarian cancers to Paclitaxel and Carboplatin regimens, and (iii)
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10 the medicinal need for new therapeutics in this type of cancer, we have selected ovarian
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12 adenocarcinomas as a good model on which to perform *in vitro* mechanistic/biological studies
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14 with selected new ferrociphenols.
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18 Ideally, an improved generation of products is required to address new mechanisms of action.
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20 These compounds should involve unequivocally characterized primary metabolites, and must
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22 also exhibit lower IC₅₀ values than those of the drug candidates known so far. In all the early
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24 modifications of **1** the ethyl entity was left untouched and, when chains were modified, this
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26 occurred mostly on the phenol side.¹³ It is only recently that we have begun to consider
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28 modification of the alkyl chain by attaching an OH group to the end, and we have obtained
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30 particularly low IC₅₀ values (IC₅₀ = 0.11 μM) with **2a** on triple negative breast cancer cell lines
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32 (MDA-MB-231),²⁶ moreover, the bioactivity of their metabolites, all of which have been
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34 identified, was also evaluated.²⁷⁻²⁸ Meanwhile a number of recent research projects have focused
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36 on biologically-oriented ferrocene compounds substituted by heterocyclic skeletons, including for
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38 example nucleobases, sugars or flavonoids.²⁹⁻⁴¹ In this context, the novel series shown below
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40 seems particularly appealing whereby a heterocyclic group such as succinimidyl or phthalimidyl,
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42 as in **3** or **4**, respectively, has been incorporated into the molecule. We note that in
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44 bioorganometallic chemistry, examples of compounds that have significantly low IC₅₀ values on
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46 EOC cells resistant to cisplatin are practically nonexistent, except for rare complexes of
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48 ruthenium or osmium, having structures different from the ferrociphenols, and which appear
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50 promising.⁴²⁻⁴³ Amongst our more recently synthesized and patented compounds,¹⁵ these
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unprecedented molecules bearing a cyclic-imide group are the most promising ones against EOC and deserve a specific study.



Scheme 1. Incorporation into the ferrociphenol skeleton, **1**, of a hydroxyl group, in **2**, or a cyclic-imide, in **3** and **4**.

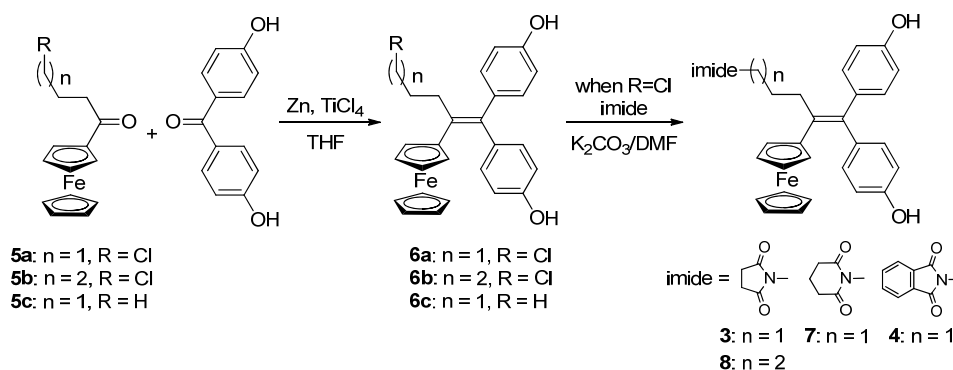
We here report the syntheses, characterization and first biological results of a series of novel complexes obtained by incorporation of imide-type heterocycles at the terminus of the alkyl chain, as in **3** and **4** (Scheme 1). These molecules form a new group of chemical products, but their chief interest lies in the encouraging early biological results obtained for some of these products, in the form of strong antiproliferative effects on ovarian cancer cell lines. They are the founding members of a particularly innovative and effective family of heterocyclic ferrociphenols.

Results and Discussion

Syntheses

Preparation of the chlorinated ferrocene ketones with a three or four methylene chain, **5a** and **5b**, by Friedel-Crafts acylation of ferrocene, followed by McMurry coupling, furnished the heavily substituted alkenes **6a**⁴⁴ and **6b** in yields of 68 and 60%, respectively. Subsequent reaction with an imide (succinimide, glutarimide or phthalimide) in a basic medium (potassium carbonate in DMF) at 80 °C gave the novel mono-heterocyclic ferrociphenols **3**, **7** and **8**, in yields

of 54, 22 and 58%, respectively, and the known phthalimide, **4**, in 83% yield.⁴⁴ The new complexes were characterized spectroscopically and, in the case of the succinimido derivative, **3**, by X-ray crystallography (Figure 1).



Scheme 2. Synthesis of the imides **3**, **4**, **7**, **8**, and of the non-substituted propyl derivative **6c**.

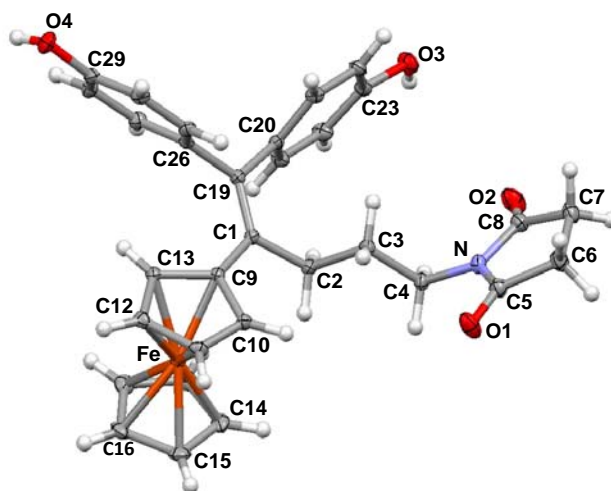


Figure 1. X-ray structure of *N*-[4-ferrocenyl-5,5-bis-(4-hydroxyphenyl)-pent-4-enyl]succinimide, **3** (thermal ellipsoids are shown at 50%).

To better assess the effect of attaching an imide at the end of the chain, we compared the cytotoxicity of each of these species with that of the reference compound **1a**, which has no such attachment. Moreover, for even better comparison, we also specifically synthesized compound **6c**,

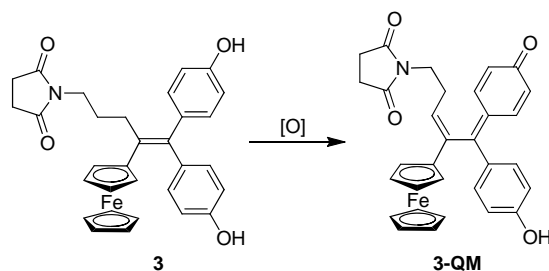
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6 which has a propyl group, with the same number of carbon atoms in the chain as compounds **3**, **4**
7 and **7**. The synthetic route is the same as for the chlorinated compounds, starting from the
8 commercially-available ferrocenyl propyl ketone **5c**.
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14 In order to investigate possible mechanisms for the antiproliferative action of our cyclic-imidyl
15 ferrociphenols, we based our studies on two types of potential metabolites. We focused initially
16 on the “ferrociphenol” motif (Fc-C=C-C₆H₄-OH), known to lead to oxidation with formation of a
17 QM in a biological medium.²⁷⁻²⁸ Subsequently, we looked at the imide component, which is
18 capable of producing specific ring-opened metabolites.
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27 One of the likely mechanisms that we have previously identified to explain the cytotoxic effect
28 of ferrocenyl phenols on various cancer cells is based on their *in situ* transformation to
29 electrophilic QMs, which are potentially cytotoxic species. This process is mediated by the
30 ferrocenyl group and allows QM formation to occur under comparatively mild (i.e. biologically
31 relevant) oxidation conditions. We propose that the strong antiproliferative activity of **3** as a
32 precursor compound is also related to QM formation inside cells. Thus, we first explored the
33 chemical oxidation of **3** (by Ag₂O) to produce the vinyl **3-QM** (Scheme 3) that was fully
34 characterized by ¹H and ¹³C NMR spectroscopy and mass spectrometry. The ¹H NMR spectrum
35 revealed an approximate 3:1 ratio of *E/Z* isomers, which can be explained by the steric issue we
36 proposed recently.²⁸
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50 Molecule **3-QM** showed a far better stability profile (half-life in acetone of about 10 days!)
51 than the corresponding QM of **1a** which was unstable, and even started to decompose before
52 oxidation was complete.²⁰ It was also superior to the tetrahydrofuran-substituted **2a-QM** (t_{1/2} in
53 acetone ~30 h), resulting from the oxidation of **2a**.²⁶ These preliminary results have been
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obtained by oxidation using rat liver microsomes (containing cytochrome P450, to be published). Since the two-electron chemical oxidation data of **3** with Ag₂O confirmed that the vinyl derivative **3-QM** is main species formed, the probable formation of **3-QM** inside cells may be anticipated, as with other ferrociphenols.¹⁴



Scheme 3. Formation of **3-QM** by chemical or metabolic oxidation.

The redox behavior of ferrociphenols mediated by the ferrocenyl can also be followed by electrochemistry. Cyclic voltammetric experiments (Figure 2) show that, in the absence of imidazole, compounds **3** and **4** exhibit a reversible monoelectronic oxidation process at 0.51 V and 0.49 V, respectively, ascribed to ferrocenyl oxidation, as already established for other ferrociphenols.²¹ Upon addition of 20 equivalents of base, they showed very similar electrochemical behavior with that we reported before,²¹ involving a base-promoted intramolecular electron transfer between the phenol and the ferrocenium cation to give a quinone methide.

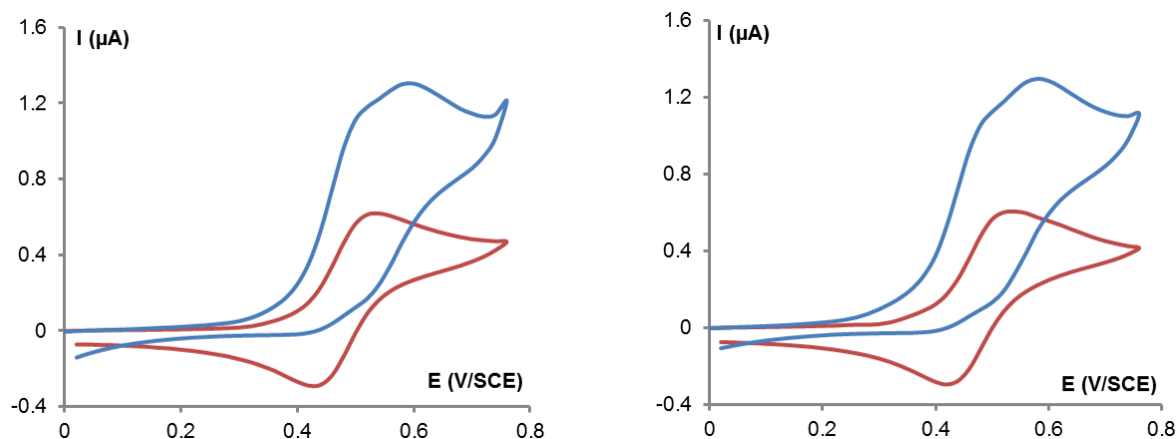
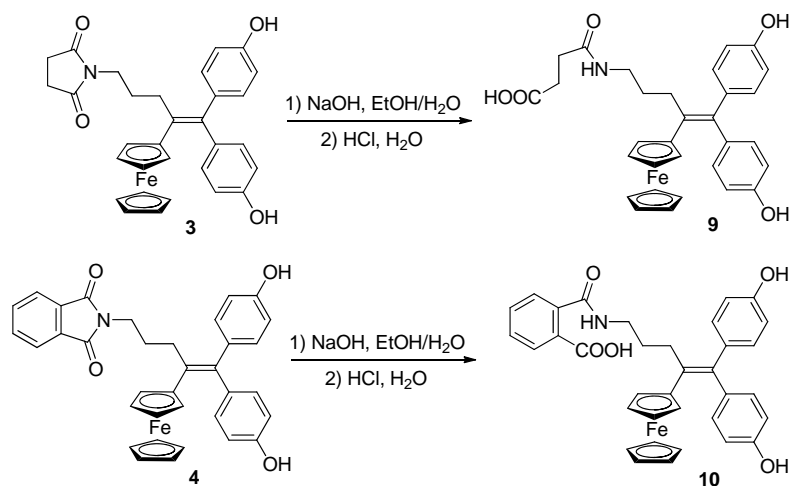
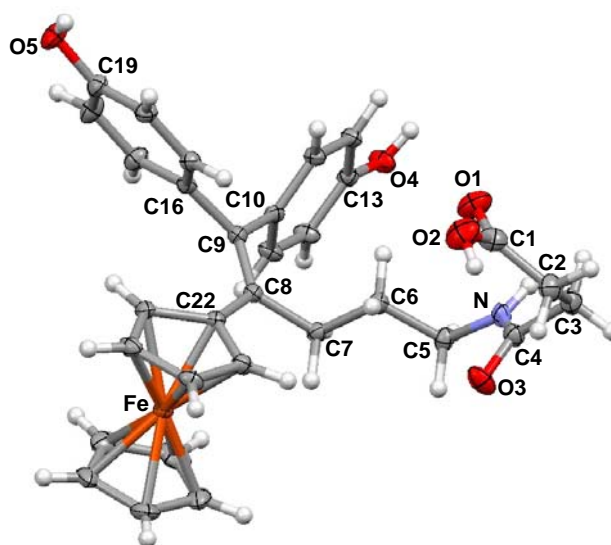


Figure 2. Cyclic voltammograms of **3** (approximately 1 mM, scan rate 200 mV/s, left) and **4** (approximately 1 mM, scan rate 200 mV/s, right) in the absence of base (red curve) and in the presence of 20 mM imidazole (blue curve).

Another possible mechanism involves opening the imide rings. It is known that thalidomide, a compound of the imide family, is at least partly metabolized by ring-opening its two imide functions (phthalimide and glutarimide).⁴⁵ We decided to examine the possibility that this opening of the imide ring could be occurring and, if so, would it explain the unusual biological activity (see below) of our compounds, particularly **3** and **4**. In order to test their activity (IC_{50}), and to compare it with that of the parent products, we synthesized these presumed metabolites by basic hydrolysis, thus opening the imido rings. The action of sodium hydroxide on these compounds dissolved in a water/ethanol mixture at room temperature gave the ring-opened products **9** and **10** in yields of 98 and 78%, respectively. These products were characterized spectroscopically and, in the case of **9**, by X-ray crystallography; its structure is shown as Figure 3.



24 **Scheme 4.** Formation of the presumed ring-opened metabolites, **9** and **10**, from the succinimidyl-
25 and phthalimidyl-ferrociphenols **3** and **4**, respectively.



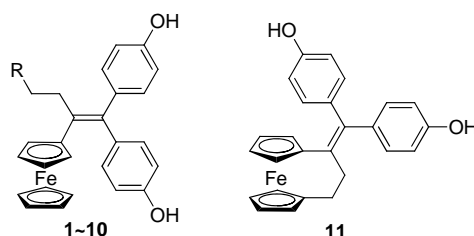
49 **Figure 3.** Molecular structure of 5-(2'-carboxypropamido)-2-ferrocenyl-1,1-bis-(4-hydroxy-
50 phenyl)-pent-1-ene, **9** (thermal ellipsoids are shown at 50%).

Biological studies

In this laboratory, preliminary cytotoxicity tests on MDA-MB-231 cancer cells have been routinely used to evaluate the anticancer activity of our molecules. Historically, our molecules were seen as derivatives of tamoxifen and thus were initially compared to its behavior against hormone-independent (MDA-MB-231) and hormone-dependent (MCF-7) breast cancer cell lines. However, over time the structure of these molecules has departed more and more from that of tamoxifen, with the result that limiting them to only breast cancers is no longer justified. Thus, we have chosen to test the activity of our new heterocyclic ferrociphenols on three ovarian epithelial cancer cell lines, one of which is resistant to cisplatin (A2780, A2780-Cis and SK-OV-3). As shown in Table 1, we also compared the new species to molecules from our library of ferrociphenols known for their strong activity: the first ferrociphenol that we synthesized, **1a**, its cyclic version [3]ferrocenophane, **11**, and the hydroxypropyl product **2a**. Finally, **6c** was added to the list to keep the same number of carbon atoms in the chain, and so better assess the effect of the attachment of a hydroxyl or cyclic-imidyl group to the end of the propyl chain. We can already see that lengthening the carbon chain from three to four methylene units diminishes the anticancer activity, and this holds true for all the lines tested.

Since increasing the length of the non-polar carbon chain on the north-west side of the molecule, which affects both steric hindrance and lipophilicity, seems to have a negative influence, we chose to study the effect of attaching a polar group onto the propyl chain. Thus, the hydroxyl group, which is small, makes it possible to improve the activity of **2a** on all the lines tested compared to **6c**, and even compared to **1a**. This shows that the positive effect of the hydroxyl overcomes the negative effect linked to the steric hindrance such that this latter parameter seems less important than the presence of this polar and protic group.

Table 1. Activity of products (IC₅₀ μM) on ovarian cancer (A2780, A2780-Cis and SK-OV3) and log Po/w results for selected ferrocenyl compounds.



Product	R	LogPo/w	SK-OV-3	A2780	A2780-Cis	SF ^a
11	--	4.6	1.91 ± 0.24	0.359 ± 0.023	0.165 ± 0.044	0.46
1a	H	5.0	2.05 ± 0.67	3.50 ± 0.94	2.3 ± 0.7	0.66
2a		4.2	0.47 ± 0.06	0.53 ± 0.01	0.30 ± 0.03	0.57
3		4.0	0.30 ± 0.02	0.035 ± 0.005	0.049 ± 0.003	1.40
4		4.8	0.31 ± 0.04	0.16 ± 0.02	0.074 ± 0.008	0.46
6c		--	12.9 ± 1.2	3.91 ± 0.37	4.99 ± 0.15	1.28
7		4.2	0.58 ± 0.16	0.15 ± 0.02	0.30 ± 0.67	2.00
8		4.4	0.26 ± 0.05	0.21 ± 0.03	1.51 ± 0.52	7.19
9		--	1.07 ± 0.63	0.150 ± 0.004	1.48 ± 0.38	9.87
10		--	8.75 ± 0.28	2.78 ± 0.47	2.91 ± 0.39	1.05
Cisplatin^b	--	--	--	1.2 ± 0.2	11.5 ± 0.3	9.6

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6 (a) Selectivity Factor (Ratio IC_{50} A2780-Cis / IC_{50} A2780); (b) Values taken from ref. 53.
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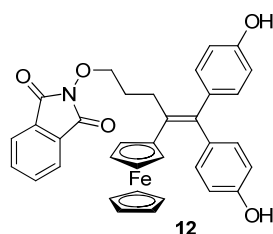
9 In light of these data, it was decided to test the attachment of a larger polar group, this time
10 aprotic, such as the imide rings. Compounds **3** and **4**, bearing succinimido or phthalimido
11 substituents, respectively, show a large improvement in antitumoral activity on the three ovarian
12 cancer cell lines with IC_{50} values below 0.08 μ M on A2780-Cis, resistant to cisplatin, as well as
13 on the A2780 line for **3**. These results on ovarian cancer are systematically better than those of **2a**,
14 even though the size of these two imides is substantially greater than that of a single hydroxyl
15 group. The results are also better than those of [3]ferrocenophane, **11**. This is an important
16 finding, considering that the cyclic compounds ([3]ferrocenophanes) were previously always
17 thought to be more antiproliferative than non-cyclic compounds,^{22, 46} and that none of our non-
18 cyclic compounds had ever had an IC_{50} value below 0.1 μ M on the lines tested. Evidently, the
19 cyclic imido species are the start of a new paradigm.
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34 To examine what makes these imides so unusual, in particular, the most active of them, **3**, we
35 made slight structural modifications by increasing the number of carbons through addition of a
36 methylene group. However, enlargement of the succinimide ring to a glutarimide ring, as in **7**,
37 resulted in loss of activity by a factor of four relative to A2780 and A2780-Cis. Even more
38 strikingly, incorporation of the additional methylene unit to **3** by increasing the chain length from
39 3 to 4 carbon atoms, as in **8**, triggers a global loss of activity that is even greater than that for the
40 glutarimide, **7**. The best combination therefore appears to be a chain of three methylenes and a
41 planar five-membered imide ring. It should be noted that the phthalimide **4**, which also possesses
42 these two characteristics, is the closest compound to **3** in terms of activity, despite having an
43 additional non-polar benzene ring that increases steric hindrance and lipophilicity. However, **4** is
44 unusual in being twice as active (selectivity factor SF = 0.46) on the cisplatin-resistant line
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6 A2780-Cis compared to A2780 (comparable to the non-heterocyclic ferrociphenols **1a**, **2a** and **11**,
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8 but with much higher activity). Although the succinimido complex, **3**, is a little more selective in
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10 terms of A2780, its IC₅₀ value on A2780-Cis is 1.5 times better than that of its phthalimido
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12 counterpart, **4**.
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16 We next attempted to investigate the possible metabolites of **3** and **4**. It has already been
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18 shown that ferrociphenols undergo oxidation to QMs which can influence activity.²⁷⁻²⁸ We now
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20 see that molecule **3** follows the same path, and its QM has been isolated (see above). However,
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22 these new molecules contain cyclic imides that are known to form metabolites by ring opening,
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24 as in thalidomide.⁴⁵ Having independently synthesized compounds **9** and **10**, and finding that
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26 their IC₅₀ values were not as potent as those of the parent imides, one might suggest that these
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28 species are broadly not involved as metabolites, or are metabolites that are not active on these
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30 targets. This statement must be tempered, however, by the fact that we have previously shown
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32 that QMs and their indenyl ring products are less active than their ferrociphenol precursors,
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34 despite admittedly being active metabolites.²⁷ We justified this finding by the fact that the
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36 metabolite must form in the cell very close to its biological target to avoid any interaction with
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38 other molecules in its path as it approaches the target.²⁷ The ring-opened molecules **9** and **10**
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40 could thus also potentially form inside the cell, but this remains to be determined. Their higher
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42 polarity compared to the precursor imide (attributable to their two polar protic functions, the
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44 secondary amide, and particularly the carboxylic acid), could also be an issue when they are
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46 introduced directly into the culture medium. Indeed, lipophilicity is an important parameter for
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48 the entry of molecules into the cell,⁴⁷ and the carboxylic function, which must exist as a
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50 carboxylate at the pH level of the culture medium should play an important role.
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We note that the phthalimide **4** is structurally very close to one of our recently published compounds, **12**,⁴⁸ the difference being insertion of an oxygen atom between the nitrogen atom of the phthalimide group and the chain. Activity on MDA-MB-231 was 13.31 μM , compared to IC_{50} values of 0.15 μM for **4** and 0.30 μM for **8** (Table 2), less potent by a factor of 89 and 44 respectively, despite the fact that this compound bears a group that is fairly equivalent to that of **4** in terms of polarity and size. This demonstrates the highly unusual properties of our imides and in particular of their linkages [imide—chain carbons—ferrociphenol].



Scheme 5. Compound **12**, analogue of **4** and **8**.

Encouraged by the exceptional results obtained with our two best products, **3** and **4**, on these three ovarian cancer cell lines, we wished to establish whether they showed an equally potent activity on other cancers (Table 2). We thus measured their IC_{50} values on triple negative breast cancer cell lines (MDA-MB-231), colorectal cancer (HCT-116) and pancreatic cancer (Mia-PaCa-2) cell lines, as well as those of two leukemias, one of them resistant to Doxorubicin (K562 and K562R). For comparison, tests were also carried out on our structurally close flagship products, **1a** and **2a**, as well as **8**, **9** and **10**. Compound **3** is the most active overall, followed by **2a** and **4**. A decrease in the cytotoxic activity of these compounds on the K562R cell line compared to the K562 line seems to indicate that they may be partial substrates of the Pgp protein overexpressed in this cell line and responsible for its resistance to Doxorubicine. However, with

selectivity factors (IC_{50} K562R / IC_{50} K562) of under 5, while the equivalent value is 745 for Doxorubicine,⁴⁹ **1a**, **2a**, **4** and **10** seem resistant to this phenomenon, unlike **3** (85) and **9** (>357).

Table 2. IC_{50} (μ M) of certain molecules on other cell lines: triple negative breast cancer (MDA-MB-231), colorectal cancer (HCT 116), pancreatic cancer (Mia-PaCa-2), leukemia (K562 and K562R).

Molecule	MDA-MB-231	HCT116	Mia-Paca-2	K562	K562R
1a	0.66 ± 0.02	4.24 ± 0.02	11.3 ± 1.8	3.32 ± 1.10	2.74 ± 0.27
2a	0.11 ± 0.02	1.21 ± 0.25	1.23 ± 0.01	1.99 ± 0.52	3.11 ± 1.41
3	0.035 ± 0.005	0.140 ± 0.001	0.13 ± 0.04	0.048 ± 0.003	4.1 ± 0.9
4	0.145 ± 0.005	0.33 ± 0.03	10.4 ± 1.5	0.50 ± 0.14	2.48 ± 0.59
8	0.30 ± 0.08	0.160 ± 0.001	15.94 ± 2.17	2.71 ± 0.41	2.74 ± 0.20
9	7.75 ± 0.31	0.11 ± 0.02	0.48 ± 0.08	0.14 ± 0.02	>50
10	1.01 ± 0.35	4.78 ± 0.34	2.75 ± 0.36	2.81 ± 0.15	14.0 ± 3.90

Five compounds were next tested on human pulmonary fibroblasts (MRC5) in order to evaluate their cytotoxic activity on non-cancerous cells (Table 3). For all the compounds tested, lower cytotoxicity was observed than that seen on A2780 and A2780-Cis cell lines, translating to a ratio greater than 1. Thus, in order of decreasing IC_{50} MRC5 relative to IC_{50} A2780-Cis we see: **2a** = 28.5 > **4** = 27.8 >> **11** = 3.8 > **1a** = 3.2 > **3** = 2.0. Compound **3**, which is the most active on ovarian cancer cells, does not have the highest ratio. However, this compound is 17 times less toxic on MRC5 than Calicheamicin, for which we recorded an IC_{50} value of 5.87 nM, and 11

times less than Paclitaxel (9.2 nM), both of which are used as anticancer drugs. On the other hand, compounds **2a** and **4** are 30 times less active on healthy cells than on cancer cells, for which they possess a high level of cytotoxicity, and this indicative criterion suggests them as good candidates for drug development.

Table 3. IC₅₀ in μM of MRC5 and the ratios IC₅₀ of MRC5 / IC₅₀ of A2780 or A2780-Cis

Product	IC ₅₀	Ratio IC ₅₀	Ratio IC ₅₀
	MRC5	MRC5/A2780	MRC5/ A2780-Cis
11	0.63 ± 0.15	1.8	3.8
1a	7.28 ± 0.39	2.1	3.2
2a	8.54 ± 0.32	16.1	28.5
3	0.10 ± 0.03	2.9	2.0
4	2.06 ± 0.97	12.9	27.8

Cell cycle analysis and apoptosis

The effect of compound **3** on the progression of A2780-Cis cells was evaluated in the different phases of the cell cycle. The cancer cells were cultivated in the absence or presence of **3** and their distribution in the different phases of the cycle were analysed by flow cytometry after double labeling with propidium iodide and incorporation of BrdU (Figure 4). When A2780-Cis cells were treated for 24 h with **3**, it can be clearly observed that this compound leads in a dose-dependent fashion to an increase in the number of cells in the synthesis phase (phase S) of the

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cell cycle. This has already been observed for **11** and **1a**, where a senescence route has been identified.²⁴ From 50 nM, it can be seen that there are no more cells that have incorporated BrdU (synthetic phase marker), suggesting that a portion of the cells that were sensitive to the action of **3** have been destroyed and that they are no longer quantifiable by flow cytometry. The rest of the cells affected by **3** have a DNA content between 2n and 4n after incorporation of propidium iodide and are found to be localized between the G0-G1 and G2-M phases.

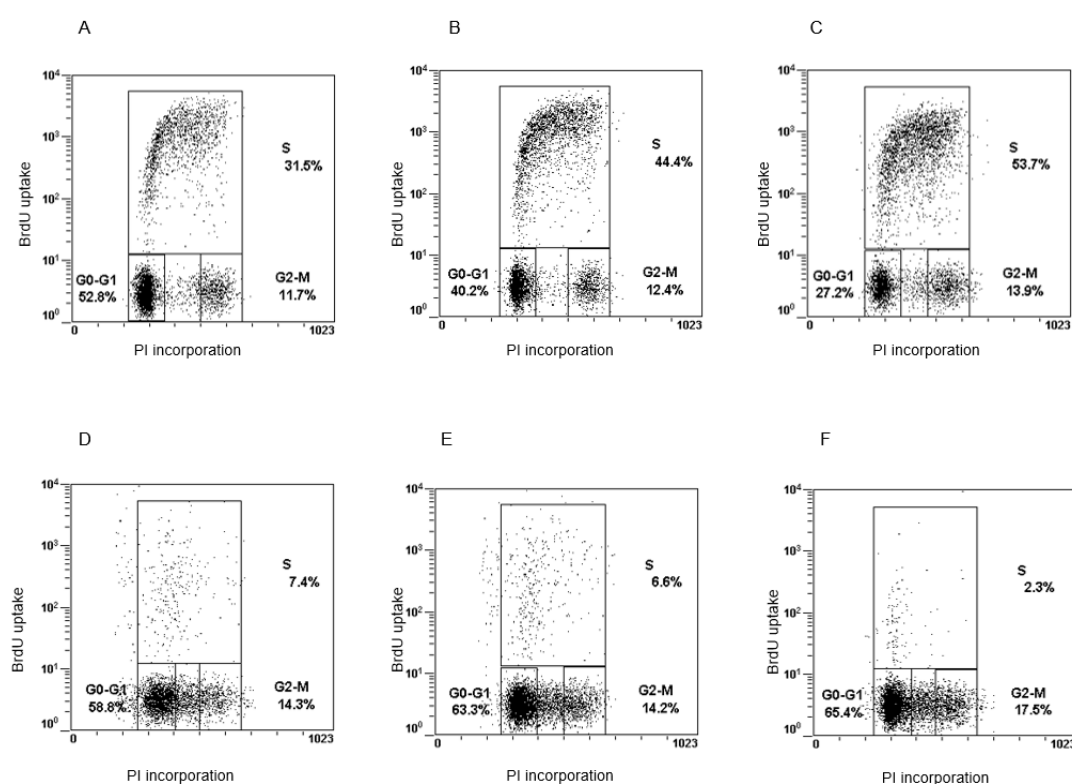
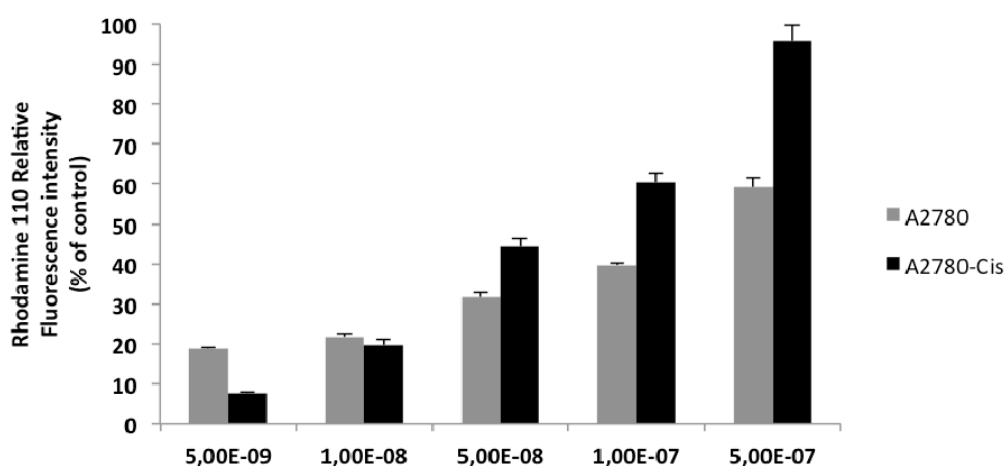


Figure 4. Cell cycle distribution, measured by bromodeoxyuridine (BrdU) uptake and propidium iodide (PI) staining, of A2780-Cis cells (control, A) and A2780-Cis cells treated with compound **3** at 5 nM (B), 10 nM (C), 50 nM (D), 100 nM (E) and 500 nM (F) for 24 h.

The blocking of the cells in one of the phases of the cell cycle is often followed by fragmentation of the DNA and other morphological characteristics of apoptosis. The ability of **3**

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6 to induce apoptosis was evaluated using a specific test that measures the cleaving of pro-caspases
7 into active caspases. A2780 and A2780-Cis cells were treated for 24 h with different
8 concentrations of **3**, and induction of apoptosis was evaluated by measuring the activity of
9 caspases 3 and 7. In Figure 5, a dose-dependent effect of **3** can clearly be seen on the two cell
10 lines with a greater sensitivity in the cisplatin-resistant cells compared to the A2780 cell line.
11 Unfortunately, an increase of active caspase-3 at the protein level was not observed, and a greater
12 understanding of the mechanisms by which **3** induces cell death in an apoptotic and non-
13 apoptotic manner is the focus of an ongoing study.
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43 **Figure 5.** Apoptotic effects of **3** in A2780 and A2780-Cis cells. The results are expressed in the
44 percentage of apoptotic cells detected following 24 h of treatment with **3** at
45 different concentrations.
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50 51 **Conclusions**

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53 The ferrociphenol family is now a rich group of more than four hundred members, some of
54 them endowed with unique antitumoral properties. All are characterized by the presence on their
55 molecular skeleton of a redox motif of the {ferrocenyl-ene-phenol} type which can be activated
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6 to form a selective electrophilic QM in cancer cells.¹³⁻¹⁴ The importance of a functionalized chain
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8 on the olefin carbon of the ferrocenyl group, such as $-(\text{CH}_2)_3\text{-OH}$ in **2a**, has recently been shown
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10 to increase antiproliferative effects. We have shown here that the antitumoral effect can be
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12 improved still further with the help of specific cyclic-imides positioned at the end of the
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14 aforementioned $-(\text{CH}_2)_3\text{-(heterocycle)}$ chain, the most potent examples having been obtained with
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16 **3** (succinimide) and **4** (phthalimide) on ovarian epithelial cancer cells. In fact, with A2780-cis, a
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18 cell resistant to cisplatin, the IC_{50} values obtained are 49 ± 3 nM for **3** and 74 ± 8 nM for **4**, a
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20 remarkable result for a type of cancer whose resistance induced by carboplatin is a major
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22 therapeutic obstacle. The exceptional activity of these two cyclic-imidyl ferrociphenol species
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24 calls for an in-depth study of their chemical and biological mechanisms in the near future.
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30 For the time being, we have started a few preliminary behavioral studies that may help
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32 establish directions for future exploration. The IC_{50} values obtained on healthy cells such as
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34 MRC5, normal human fibroblast cells, show that our compounds are more active, although in
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36 various degrees, on cancer cells. The selectivity factor SF gives an IC_{50} ratio for MRC5/A2780-
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38 cis of 2 for **3**, 27.8 for **4** and 28.5 for **2a**. It is therefore possible to take into account, where
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40 necessary, the balance between the highest antiproliferative activity with **3** and the more qualified
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42 SF values on healthy cells.
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46 For **3**, it has been shown that the major metabolite resulting from chemical oxidation is a
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48 cyclic-imidyl QM using the ferrocene entity as an oxidizing intermediary. This mechanism is
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50 similar to previous observations,²⁰⁻²³ the difference being the noteworthy presence of the
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52 succinimido functionality whose exact role remains to be clarified. The cell cycle studied with
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54 this product shows the unusual effect of the stopping of the cycle in the S phase, often associated
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56 in these series with senescence,²⁴⁻²⁵ and the occurrence of apoptosis has been shown.
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6 The antiproliferative activity observed on these ovarian cells is quite similar, whether or not
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8 they are resistant to cisplatin. Compound **3** has the lowest IC₅₀ values for the two categories,
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10 whereas **4** is twice as active on the resistant line than on A2780. The non-heterocyclic complexes
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12 (**11**, **2a**, **1a**) are also more active on the resistant line, but with IC₅₀ values mostly around one
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14 order of magnitude higher, which still remains acceptable.
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18 Comparison of the activity results of seven ferrociphenol compounds on K562 and
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20 doxorubicine-resistant K562R shows that activity is less on the resistant line for five of the
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22 compounds. This may suggest that the compounds are partial substrates of P glycoprotein (PgP)
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24 which is responsible for efflux-induced resistance to anticancer drugs. The two compounds that
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26 do not display these characteristics are **1a** and **8**, the only two compounds without a trimethylene
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28 carbon chain, but their IC₅₀ values on K562R are modest (2.74 μM). Following on from these
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30 results, the unusual behavior of the two cyclic-imidyl molecules **3** and **4** is noteworthy. It may be
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32 remarked that if, in terms of IC₅₀ values, **3** appears in general to be the more active, when other
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34 parameters such as SF are considered, the interest in **4** is increased. Biological studies are
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36 currently being carried out on the two species in parallel.
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43 **Experimental section**

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46 All reagents and solvents were obtained from commercial suppliers. THF was obtained by
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48 distillation from sodium/benzophenone. Thin layer chromatography (TLC) was performed on
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50 silica gel 60 GF₂₅₄. Column chromatography was performed on silica gel Merck 60 (40-63 μm).
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52 All NMR experiments (¹H, ¹³C) were carried out at room temperature on Bruker 300 and 400
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54 NMR spectrometers. Assignments were made based on standard 1D and 2D methods, in
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56 particular HMBC, to assign ortho and meta phenolic resonances on compound **3**. Mass
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6 spectrometry was performed with a Nermag R 10–10C spectrometer. HRMS measurements were
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8 performed on a Thermo Fischer LTQ-Orbitrap XL apparatus equipped with an electrospray
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10 source by IPCM (UMR 8232). Elemental analyses were performed by the microanalysis service
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12 of ICSN (Gif sur Yvette, France), purity of $\geq 95\%$ was confirmed via elemental analysis for
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14 compound **3**, **6c**, **7**, **8** and analytical HPLC for **9**, **10**. Cytotoxicity measurements were performed
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16 by Centre de Recherche de Gif, Institut de Chimie des Substances Naturelles, UPR 2301 du
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18 CNRS, Université Paris-Saclay, Gif-sur-Yvette Cedex, France.

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22 **General procedure for the synthesis of imides.** A mixture of potassium carbonate and imide
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24 in dimethylformamide (DMF) was heated at 80 °C for 15 min. Compound **6a** or **6b** was added
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26 and the stirring was continued at 80 °C overnight. The mixture was allowed to cool to room
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28 temperature, poured into a dilute hydrochloric acid solution, and extracted twice with diethyl
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30 ether, then the organic layer was dried on magnesium sulfate and concentrated under reduced
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32 pressure. The residue was purified by flash-chromatography with an ethyl acetate/cyclohexane
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34 mixture to afford the imide.

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38 ***N*-[4-Ferrocenyl-5,5-bis-(4-hydroxyphenyl)-pent-4-enyl]succinimide, 3.** Following the
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40 general procedure for the synthesis of imides, potassium carbonate (0.478 g, 3.5 mmoles),
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42 succinimide (0.457 g, 4.6 mmoles), compound **6a** (1.09 g, 2.31 mmoles) furnished compound **3**
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44 as a yellow solid in a yield of 54% (0.662 g). Mp: 225 °C decomp. ¹H NMR (300 MHz, acetone-
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46 d₆): δ 1.70-1.81 (m, 2H, CH₂), 2.56-2.64 (m, 6H, 2CH₂ of succinimide + CH₂-C=C), 3.38 (t, J =
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48 6.6 Hz, 2H, CH₂N), 3.95 (t, J = 1.9 Hz, 2H, C₅H₄), 4.10 (t, J = 1.9 Hz, 2H, C₅H₄), 4.16 (s, 5H,
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50 Cp), 6.74 (d, J = 8.6 Hz, 2H, C₆H₄ ortho), 6.85 (d, J = 8.6 Hz, 2H, C₆H₄ ortho), 6.91 (d, J = 8.6 Hz,
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52 2H, C₆H₄ meta), 7.04 (d, J = 8.6 Hz, 2H, C₆H₄ meta), 8.25 (s, 1H, OH), 8.36 (s, 1H, OH). ¹³C NMR
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54 (75 MHz, acetone-d₆) : δ 29.4 (2CH₂, succinimide), 30.8 (CH₂), 33.8 (CH₂), 39.7 (CH₂), 69.5
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6 (2CH, C₅H₄), 70.7 (7CH, Cp+2CH, C₅H₄), 88.8 (C, C₅H₄), 116.5 (2CH, C₆H₄ ortho), 116.6 (2CH,
7 C₆H₄ ortho), 132.0 (2CH, C₆H₄ meta), 132.4 (2CH, C₆H₄ meta), 135.3 (C), 137.7 (C), 137.9 (C), 140.2
8 (C), 157.3 (C), 157.5 (C), 178.6 (2CO). IR (KBr, ν cm⁻¹): 3421 (OH), 3096, 2967, 2936 (CH,
9 CH₂), 1697 (CO). MS (ESI) m/z: 535 [M]⁺, 342, 279, 224, 143, 83. HRMS (ESI, C₃₁H₂₉FeNO₄:
10 [M]⁺) calcd: 535.1446, found: 535.1460. Anal. Calcd for C₃₁H₂₉FeNO₄(H₂O)_{0.3}: C, 68.85; H,
11 5.51; N, 2.58. Found: C, 68.76; H, 5.14; N, 2.37.
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20 **6-Chloro-2-ferrocenyl-1,1-bis-(4-hydroxyphenyl)-hex-1-ene, 6b.** Titanium chloride (14.228
21 g, 8.24 mL, 75 mmol) was added dropwise to a suspension of zinc powder (5.883 g, 90 mmol)
22 in dry THF (500 mL) at 10-20 °C. The mixture was heated at reflux for 2 hours. A second
23 solution was prepared by dissolving compound **5b** (4.569 g, 15 mmol) and 4,4'-
24 dihydroxybenzophenone (3.222 g, 15 mmol) in dry THF (50 mL). This latter solution was
25 added dropwise to the first solution and then the reflux was continued overnight. After cooling to
26 room temperature, the mixture was stirred with water and dichloromethane. The mixture was
27 acidified with dilute hydrochloric acid until the dark color disappeared, and was then decanted.
28 The aqueous layer was extracted with dichloromethane and the combination of organic layers
29 was dried on magnesium sulfate. After concentration under reduced pressure, the crude product
30 was chromatographed on silica gel column with dichloromethane/acetone 90/10 as the eluent and
31 crystallized from dichloromethane, to afford **6b** in a yield of 60%, as an orange solid. ¹H NMR
32 (300 MHz, acetone-d₆): δ 1.61-1.74 (m, 4H, CH₂-CH₂), 2.69 (t, J = 7.4 Hz, 2H, CH₂-C=C), 3.50
33 (t, J = 6.2 Hz, 2H, CH₂Cl), 3.96 (t, J = 1.9 Hz, 2H, C₅H₄), 4.09 (t, J = 1.9 Hz, 2H, C₅H₄), 4.15 (s,
34 5H, Cp), 6.74 (d, J = 8.6 Hz, 2H, C₆H₄ ortho), 6.85 (d, J = 8.6 Hz, 2H, C₆H₄ ortho), 6.91 (d, J = 8.6
35 Hz, 2H, C₆H₄ meta), 7.09 (d, J = 8.6 Hz, 2H, C₆H₄ meta), 8.28 (s, 1H, OH), 8.32 (s, 1H, OH). ¹³C
36 NMR (75 MHz, acetone-d₆): δ 28.4 (CH₂), 33.0 (CH₂), 34.2 (CH₂), 45.3(CH₂Cl), 68.4 (2CH,
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6 C₅H₄), 69.6 (5CH, Cp), 69.7 (2CH, C₅H₄), 87.9 (C, C₅H₄), 115.5 (2CH, C₆H₄ ortho), 115.6 (2CH,
7 C₆H₄ ortho), 131.0 (2CH, C₆H₄ meta), 131.5 (2CH, C₆H₄ meta), 134.5 (C), 136.7 (C), 137.0 (C), 139.2
8 (C), 156.4 (C), 156.5 (C). IR (KBr, ν cm⁻¹): 3419 (OH), 2987, 2952, 2863 (CH, CH₂). MS (EI, 70
9 eV) m/z: 486 [M]⁺, 421, 343, 286, 186. HRMS (ESI, C₂₈H₂₇ClFeO₂: [M]⁺) calcd: 486.1049,
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12 found: 486.1058.
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18 **2-Ferrocenyl-1,1-bis-(4-hydroxyphenyl)-pent-1-ene, 6c.** The same procedure as for **6b** was
19 used with commercially-available ferrocenyl propyl ketone **5c** (1.28 g, 5 mmoles), titanium
20 chloride (3.11 g, 1.8 mL, 16.4 mmoles), zinc powder (2.0 g, 30.6 mmoles), 4,4'-
21 dihydroxybenzophenone (3.222 g, 15 mmoles), dry THF (120 + 15 mL), reflux 2 hours + 2 hours
22 to furnish compound **6c** as an orange solid with a yield of 37 %. Mp: 186 °C. ¹H NMR (300 MHz,
23 acetone-d₆): δ 0.85 (t, J = 7.5 Hz, 3H, CH₃), 1.45-1.60 (m, 2H, CH₂), 2.64 (t, J = 8.0 Hz, 2H,
24 CH₂), 3.94 (t, J = 1.9 Hz, 2H, C₅H₄), 4.09 (t, J = 1.9 Hz, 2H, C₅H₄), 4.15 (s, 5H, Cp), 6.74 (d, J =
25 8.7 Hz, 2H, C₆H₄ ortho), 6.85 (d, J = 8.7 Hz, 2H, C₆H₄ ortho), 6.90 (d, J = 8.7 Hz, 2H, C₆H₄ meta),
26 7.09 (d, J = 8.7 Hz, 2H, C₆H₄ meta), 8.23 (s, 1H, OH), 8.26 (s, 1H, OH). ¹³C NMR (75 MHz,
27 acetone-d₆) : δ 15.4 (CH₃), 25.4 (CH₂), 38.4 (CH₂), 69.4 (2CH, C₅H₄), 70.6 (5CH, Cp), 70.8
28 (2CH, C₅H₄), 89.3 (C, C₅H₄), 116.5 (2x2CH, C₆H₄ ortho), 132.1 (2CH, C₆H₄ meta), 132.5 (2CH,
29 C₆H₄ meta), 136.0 (C), 138.1 (C), 138.3 (C), 140.0 (C), 157.4 (C), 157.5 (C). IR (KBr, ν cm⁻¹):
30 3392 (OH), 2957, 2869 (CH₂, CH₃). MS (EI, 70 eV) m/z: 438 [M]⁺, 395 [M-Pr]⁺, 373 [M-Cp]⁺,
31 121 [CpFe]⁺. HRMS (FAB, C₂₇H₂₆FeO₂: [M]⁺) calcd: 438.1283, found: 438.1287. Anal. Calcd
32 for C₂₇H₂₆FeO₂: C, 73.98; H, 5.97. Found: C, 73.85; H, 5.91.
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54 **N-[4-Ferrocenyl-5,5-bis-(4-hydroxyphenyl)-pent-4-enyl]glutarimide, 7.** Following the
55 procedure used for **3**, potassium carbonate (0.175 g, 1.3 mmoles), glutarimide (0.191 g, 1.7
56 mmoles), compound **6a** (0.4 g, 0.85 mmoles) gave compound **7** as an orange solid in a yield of
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22% (0.10 g). Mp: 216 °C decomp. ¹H NMR (300 MHz, acetone-d₆): δ 1.60-1.76 (m, 2H, CH₂), 1.79-1.93 (m, 2H, CH₂), 2.50-2.63 (m, 6H, glutarimide+CH₂), 3.65 (t, *J* = 7.0 Hz, 2H, CH₂), 3.92 (t, *J* = 1.9 Hz, 2H, C₅H₄), 4.06 (t, *J* = 1.9 Hz, 2H, C₅H₄), 4.11 (s, 5H, Cp), 6.70 (d, *J* = 8.7 Hz, 2H, C₆H₄ ortho), 6.81 (d, *J* = 8.5 Hz, 2H, C₆H₄ ortho), 6.87 (d, *J* = 8.7 Hz, 2H, C₆H₄ meta), 7.02 (d, *J* = 8.5 Hz, 2H, C₆H₄ meta), 8.24 (s, 1H, OH), 8.31 (s, 1H, OH). ¹³C NMR (75 MHz, acetone-d₆): δ 17.8 (CH₂, glutarimide), 29.6 (CH₂), 33.0 (CH₂), 33.2 (2CH₂, glutarimide), 39.6 (CH₂), 68.6 (2CH, C₅H₄), 69.9 (5CH, Cp), 70.0 (2CH, C₅H₄), 88.1 (C, C₅H₄), 115.7 (2CH, C₆H₄ ortho), 115.9 (2CH, C₆H₄ ortho), 131.2 (2CH, C₆H₄ meta), 131.7 (2CH, C₆H₄ meta), 134.6 (C), 137.0 (C), 137.3 (C), 139.3 (C), 156.6 (C), 156.7 (C), 173.0 (2CO). IR (KBr, ν cm⁻¹): 3358 (OH), 1723 (CO). HRMS (ESI, C₃₂H₃₁FeNNaO₄: [M+Na]⁺) calcd: 572.1500, found: 572.1495. Anal. Calcd for C₃₂H₃₁FeNO₄(H₂O)_{0.5}: C, 68.82; H, 5.77; N, 2.50. Found: C, 68.86; H, 5.46; N, 2.49.

***N*-[5-Ferrocenyl-6,6-bis-(4-hydroxyphenyl)-hex-5-enyl]succinimide, 8.** Likewise, potassium carbonate (0.415 g, 3 mmoles), succinimide (0.396 g, 4 mmoles), compound **6b** (0.974 g, 2 mmoles) gave compound **8** as an orange solid in a yield of 58% (0.632 g). mp: 207 °C decomp. ¹H NMR (300 MHz, acetone-d₆): δ 1.42-1.54 (m, 4H, CH₂-CH₂), 2.66-2.71 (m, 6H, succinimide+CH₂-C=C), 3.35 (t, *J* = 6.6 Hz, 2H, CH₂N), 3.95 (t, *J* = 1.9 Hz, 2H, C₅H₄), 4.10 (t, *J* = 1.9 Hz, 2H, C₅H₄), 4.17 (s, 5H, Cp), 6.73 (d, *J* = 8.6 Hz, 2H, C₆H₄ ortho), 6.84 (d, *J* = 8.6 Hz, 2H, C₆H₄ ortho), 6.89 (d, *J* = 8.6 Hz, 2H, C₆H₄ meta), 7.08 (d, *J* = 8.6 Hz, 2H, C₆H₄ meta), 8.25 (s, 1H, OH), 8.30 (s, 1H, OH). ¹³C NMR (75 MHz, acetone-d₆): δ 28.1 (2CH₂), 28.3 (2CH₂), 34.6 (CH₂), 38.4 (CH₂), 68.3 (2CH, C₅H₄), 69.5 (5CH, Cp), 69.7 (2CH, C₅H₄), 88.2 (C, C₅H₄), 115.4 (2CH, C₆H₄ ortho), 115.5 (2CH, C₆H₄ ortho), 130.9 (2CH, C₆H₄ meta), 131.4 (2CH, C₆H₄ meta), 134.6 (C), 136.7 (C), 137.0 (C), 139.0 (C), 156.3 (C), 156.4 (C), 177.7 (2CO). IR (KBr, ν cm⁻¹): 3393 (OH), 3096, 3023, 2940, 2879 (CH, CH₂), 1681 (CO). MS (EI, 70 eV) m/z: 549 [M]⁺, 484 [M-Cp]⁺, 442, 382,

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6 343, 286, 186. HRMS (ESI, $C_{32}H_{31}FeNO_4$: $[M]^{+}$) calcd: 549.1602, found: 549.1622. Anal. Calcd
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8 for $C_{32}H_{31}FeNO_4(H_2O)$: C, 67.73; H, 5.85; N, 2.46. Found: C, 67.33; H, 5.53; N, 2.28.

10 **5-(2'-Carboxypropamido)-2-ferrocenyl-1,1-bis-(4-hydroxyphenyl)-pent-1-ene, 9**. Imide **3**
11 (3.18 g, 5.94 mmoles) was dissolved into an ethanol/water solution (300 mL/150 mL) and
12 sodium hydroxide (6.2 g, 155 mmoles) was added. The solution was stirred overnight at room
13 temperature, then was poured into water and acidified (HCl) in order to protonate phenolates.
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15 Then the pH was adjusted to 5-6 with sodium hydrogen carbonate in order to deprotonate the
16 carboxylic function only. Ethyl acetate was added, the organic layer was discarded and the
17 aqueous layer was washed with ethyl acetate, then was acidified (HCl). Acid **9** was extracted
18 twice with ethyl acetate, then the solution was concentrated under reduced pressure. Acid **9** was
19 obtained as an orange solid in a yield of 98 % and was recrystallized from an
20 acetone/dichloromethane solution. Mp: 198 °C. 1H NMR (300 MHz, acetone- d_6): δ 1.61-1.74 (m,
21 2H, CH_2), 2.43 (t, $J = 6.6$ Hz, 2H, CH_2), 2.57 (t, $J = 6.6$ Hz, 2H, CH_2), 2.62-2.71(m, 2H, CH_2 -
22 C=C), 3.10 (t, $J = 6.6$ Hz, 2H, CH_2N), 3.93 (t, $J = 1.9$ Hz, 2H, C_5H_4), 4.05 (t, $J = 1.9$ Hz, 2H,
23 C_5H_4), 4.12 (s, 5H, Cp), 6.70 (d, $J = 8.5$ Hz, 2H, C_6H_4 ortho), 6.82 (d, $J = 8.5$ Hz, 2H, C_6H_4 ortho),
24 6.86 (d, $J = 8.5$ Hz, 2H, C_6H_4 meta), 6.98 (s broad, 1H, NH), 7.05 (d, $J = 8.5$ Hz, 2H, C_6H_4 meta).
25 ^{13}C NMR (75 MHz, acetone- d_6) : δ 29.2 (CH_2), 31.2 (CH_2), 31.5 (CH_2), 32.9 (CH_2), 40.1 (CH_2),
26 68.7 (2CH, C_5H_4), 69.9 (5CH, Cp), 70.0 (2CH, C_5H_4), 88.3 (C, C_5H_4), 115.8 (2CH, C_6H_4 ortho),
27 115.9 (2CH, C_6H_4 ortho), 131.3 (2CH, C_6H_4 meta), 131.7 (2CH, C_6H_4 meta), 134.7 (C), 137.0 (C),
28 137.3 (C), 139.4 (C), 156.7 (C), 156.8 (C), 172.6 (CO), 174.1 (CO). IR (ATR, ν cm^{-1}): 2970
29 (OH), 1727 (CO), 1711 (CO). HRMS (ESI, $C_{31}H_{31}FeNNaO_5$: $[M+Na]^{+}$) calcd: 576.1449, found:
30 576.1443.
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6 **5-(o-Carboxybenzamido)-2-ferrocenyl-1,1-bis-(4-hydroxyphenyl)-pent-1-ene, 10.** The
7
8 same procedure as for **9** was used with imide **4** (0.226 g, 0.39 mmoles), sodium hydroxide (0.24 g,
9
10 6 mmoles), ethanol (20 mL), water (10 mL) to furnish acid **10** as an orange solid with a yield of
11
12 78 %. Mp: 175 °C. ¹H NMR (300 MHz, DMSO-d₆): δ 1.61-1.77 (m, 2H, CH₂), 2.51-2.60 (m, 2H,
13
14 CH₂), 3.08 (q, *J* = 6.5 Hz, 2H, CH₂N), 3.85 (t, *J* = 1.9 Hz, 2H, C₅H₄), 4.07 (t, *J* = 1.9 Hz, 2H,
15
16 C₅H₄), 4.11 (s, 5H, Cp), 6.64 (d, *J* = 8.3 Hz, 2H, C₆H₄ ortho), 6.73 (d, *J* = 8.3 Hz, 2H, C₆H₄ ortho),
17
18 6.80 (d, *J* = 8.3 Hz, 2H, C₆H₄ meta), 6.98 (d, *J* = 8.3 Hz, 2H, C₆H₄ meta), 7.32 (d, *J* = 7.4 Hz, 1H,
19
20 H_{arom}), 7.49 (t, *J* = 7.4 Hz, 1H, H_{arom}), 7.58 (t, *J* = 7.5 Hz, 1H, H_{arom}), 7.74 (d, *J* = 7.5 Hz, 1H,
21
22 H_{arom}), 8.25 (s broad, 1H, NH), 9.26 (s, 1H, OH), 9.31 (s, 1H, OH). ¹³C NMR (75 MHz, DMSO-
23
24 d₆): δ 30.3 (CH₂), 31.9 (CH₂), 39.8 (CH₂), 67.7 (2CH, C₅H₄), 68.8 (2CH, C₅H₄), 69.0 (5CH, Cp),
25
26 86.6 (C, C₅H₄), 115.0 (2x2CH, C₆H₄ ortho), 127.6 (CH_{arom}), 129.0 (CH_{arom}), 129.1 (CH_{arom}), 129.9
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28 (2CH, C₆H₄ meta), 130.3 (2CH, C₆H₄ meta), 130.8 (C), 131.0 (CH_{arom}), 133.3 (C), 135.3 (C), 135.7
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30 (C), 138.0 (C), 138.6 (C), 155.57 (C), 155.64 (C), 168.0 (CO), 168.2 (CO). IR (ATR, ν cm⁻¹):
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32 2970 (OH), 1738 (CO). HRMS (ESI, C₃₅H₃₁FeNNaO₅: [M+Na]⁺) calcd: 624.1449, found:
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34 624.1445.
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42 **X-Ray crystal structure determination.** Single crystals were selected, mounted and transferred
43
44 into a cold nitrogen gas stream. Intensity data was collected with Bruker Kappa-APEX2 systems
45
46 using micro-source Cu-Kα radiation. Unit-cell parameters, data collection strategy, integration
47
48 and absorption correction were carried out with the Bruker APEX2 suite of programs. Structures
49
50 were solved with SHELXT-2014⁵⁰ and refined anisotropically by full-matrix least-squares
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52 methods with SHELXL-2014⁵⁰ using the WinGX suite.⁵¹ Relevant data have been deposited at
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54 the Cambridge Crystallographic Data Centre with numbers CCDC 1542075 and 1542077 and can
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56 be obtained free of charge via www.ccdc.cam.ac.uk.
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6 **Electrochemical oxidation of 3 and 4.** Cyclic voltammograms (CVs) were obtained using a
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8 three electrode cell with a 0.5 mm Pt working electrode, stainless steel rod counter electrode, and
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10 Ag/AgCl ethanol reference electrode, with an μ -Autolab 3 potentiostat driven by GPES software
11
12 (General Purpose Electrochemical System, Version 4.8, EcoChemie B.V., Utrecht, the
13
14 Netherlands). Solutions consisted of 5 mL CH₃CN, approximately 1 mM analyte, and 0.1 M
15
16 Bu₄NPF₆ supporting electrolyte. Scan rate of CVs was 200 mV/s.
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20 **Lipophilicity.** Measurements of the octanol/water partition coefficient (log Po/w) were made by
21
22 the HPLC technique. Measurement of the chromatographic capacity factors (k) for each molecule
23
24 was done at various concentrations in the range of 95–75% methanol containing 0.25% (v/v) 1-
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26 octanol and an aqueous phase consisting of 0.15% (v/v) n-decylamine in the buffering agent
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28 MOPS (3-morpholinopropane-1-sulfonic acid, prepared in 1-octanol saturated water) adjusted to
29
30 pH 7.4. These capacity factors (k') are extrapolated to 100% of the aqueous component given the
31
32 value of k'w. The log Po/w is obtained by the formula $\log Po/w = 0.13418 + 0.98452 \log k'$.
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37 **Cell Culture and Proliferation Assay.** Cancer cell lines were obtained from the American Type
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39 Culture Collection (Rockville, MD, USA) or from the European collection of cell culture
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41 (ECACC, England) and were cultured according to the supplier's instructions. Human HCT-116
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43 colorectal carcinoma and SK-OV-3 ovary carcinoma were grown in Gibco McCoy's 5A
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45 supplemented with 10% fetal calf serum (FCS) and 1% glutamine. K562 and K562R
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47 myelogenous leukemia, A2780 and A2780-Cis ovarian carcinoma cells were grown in RPMI
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49 1640 supplemented with 10% fetal calf serum (FCS) and 1% glutamine. Mia-Paca2 carcinoma
50
51 cells were grown in Gibco medium DMEM supplemented with 10% fetal calf serum (FCS) and
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53 1% glutamine. Cells were maintained at 37 °C in a humidified atmosphere containing 5% CO₂.
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55 Cell growth inhibition was determined by an MTS assay according to the manufacturer's
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6 instructions (Promega, Madison, WI, USA). Briefly, the cells were seeded in 96-well plates (2.5
7 $\times 10^3$ cells/well) containing 100 μL of growth medium. After 24 h of culture, the cells were
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9 treated with the tested compounds at 10 different final concentrations. After 72 h of incubation,
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11 20 μL of CellTiter 96® AQueous One Solution Reagent was added for 2h before recording
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13 absorbance at 490 nm with a spectrophotometric plate reader PolarStar Omega (BMG Labtech,
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15 USA). The dose-response curves were plotted with Graph Prism software and the IC_{50} values
16
17 were calculated using the Graph Prism software from polynomial curves (four or five-parameter
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19 logistic equations).
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24 MDA-MB-231: Stock solutions (10 mM) of the compounds to be tested were prepared in DMSO
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26 and were kept at $-20\text{ }^\circ\text{C}$ in the dark. Serial dilutions in Dulbecco's modified eagle medium
27
28 (DMEM) without phenol red/Glutamax I were prepared just prior to use. DMEM without phenol
29
30 red, Glutamax I and fetal bovine serum were purchased from Gibco; MDA-MB-231 cells were
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32 obtained from ATCC (Manassas, VA, USA). Cells were maintained in a monolayer culture in
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34 DMEM with phenol red/Glutamax I supplemented with 9% fetal bovine serum at $37\text{ }^\circ\text{C}$ in a 5%
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36 CO_2 /air-humidified incubator. For proliferation assays, MDA-MB-231 cells were plated in 1 mL
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38 of DMEM without phenol red, supplemented with 9% decompemented and hormone-depleted
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40 fetal bovine serum, 1% kanamycin, 1% Glutamax I and incubated. The following day(D0), 1 mL
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42 of the same medium containing the compounds to be tested was added to the plates. After 3 days
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44 (D3) the incubation medium was removed and 2 mL of the fresh medium containing the
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46 compounds was added. At different days (D4, D5), the protein content of each well was
47
48 quantified by methylene blue staining as follows: cell monolayers were fixed for 1 h at room
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50 temperature with methylene blue (1mg mL^{-1} in 50:50 water/MeOH mixture), then washed with
51
52 water. After addition of HCl (0.1 M, 2 mL), the plate was incubated for 1 h at $37\text{ }^\circ\text{C}$ and then the
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54 absorbance of each well (4 wells for each concentration) was measured at 655 nm with a Biorad
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6 spectrophotometer. The results are expressed as the percentage of proteins versus the control.
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8 Two independent experiments, run in quadruplicate, were performed.
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11 **Cell Cycle Analysis.** Exponentially growing A2780-Cis cancer cells were incubated with the
12 tested compound or DMSO for 24 h. Cell-cycle profiles were determined by flow cytometry on a
13 FC500 flow cytometer (Beckman-Coulter, France) as described previously.⁵²
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18 **Apoptosis Assay.** Apoptosis was measured by the Apo-one homogeneous caspase-3/7 assay
19 (Promega Co, WI) according to the manufacturer's recommendations. Briefly, cells were
20 subcultured on a 96-well plate with 5×10^4 cells/well in 50 μ L medium. After 24 h of culture, the
21 cells were supplemented with 50 μ L of medium containing different concentrations of compound
22 **6** (5, 10, 50, 100 and 500 nM) or 0.1% DMSO (as negative control). The treated cells were
23 incubated for 24 h, each well then received 100 μ L of a mixture of caspase substrate and Apo-one
24 caspase 3/7 buffer. After 6 h of incubation, the fluorescence of sample was measured using a
25 PolarStar Omega microtiter plate fluorimeter (BMG Labtech, USA) at 527 nm.
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38 **Supporting Information Available**

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41 Crystallographic Data for **3** and **9**, molecular formula strings of related compounds, ¹H NMR
42 spectrum of **3-QM** in acetone-*d*₆. Mass spectrometry of **3**, **6b** and **8**. This material is available
43 free of charge via the Internet at <http://pubs.acs.org>.
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6 Dr P. Pascal and Dr Y. Wang equally contributed to this work.
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9 **ACKNOWLEDGEMENTS**

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11
12 The authors acknowledge Geoffrey Gontard of the Institut Parisien de Chimie Moléculaire for all
13
14 X-ray diffraction studies, PSL, Feroscan and PGG for financial support of Y. W., Barbara
15
16 McGlinchey for editorial assistance, and Dr F. Mechta-Grigoriou, Curie Hospital, for helpful
17
18 discussions.
19
20

21 **ABBREVIATIONS USED**

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25 EOC, epithelial ovarian cancer; ROS, reactive oxygen species; QM, quinone methide; IC₅₀, half
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27 inhibitory concentration; AP1, activator protein 1; IL-6, interleukin-6; IL-8, interleukin-8;
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29 A2780-Cis, A2780 resistant to cisplatin; SF, selectivity factor; BrDU, bromodeoxyuridine; PI,
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31 propidium iodide; PgP, P glycoprotein.
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