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## REVIEW

# Rodent models in Down syndrome research: impact and future opportunities

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## ABSTRACT

Down syndrome is caused by trisomy of chromosome 21. To date, a multiplicity of mouse models with Down-syndrome-related features has been developed to understand this complex human chromosomal disorder. These mouse models have been important for determining genotype-phenotype relationships and identification of dosage-sensitive genes involved in the pathophysiology of the condition, and in exploring the impact of the additional chromosome on the whole genome. Mouse models of Down syndrome have also been used to test therapeutic strategies. Here, we provide an overview of research in the last 15 years dedicated to the development and application of rodent models for Down syndrome. We also speculate on possible and probable future directions of research in this fast-moving field. As our understanding of the syndrome improves and genome engineering technologies evolve, it is necessary to coordinate efforts to make all Down syndrome models available to the community, to test therapeutics in models that replicate the whole trisomy and design new animal models to promote further discovery of potential therapeutic targets.

**KEY WORDS:** Down syndrome, Mouse model, Chromosome engineering, Aneuploidy, Dosage-sensitive gene

## Introduction

Trisomy of human chromosome 21 (Hsa21; see Box 1 for a glossary of terms), which affects 1 in 700 live births globally (Canfield et al., 2006), gives rise to Down syndrome (DS), a condition that

significantly impairs health and autonomy of affected individuals (Khoshnood et al., 2011; Parker et al., 2010). Despite the wide availability of prenatal diagnosis since the mid-1960s (Summers et al., 2007) and the introduction of maternal serum screening in 1984 (Inglis et al., 2012), the incidence of DS has not necessarily decreased (Natoli et al., 2012; Loane et al., 2013; de Graaf et al., 2016); in fact, prevalence is going up, largely because of increased lifespan and maternal age (which is the single biggest risk factor) (Sherman et al., 2007; Loane et al., 2013).

A core set of features characterises most cases of DS, including specific cognitive disabilities, hypotonia (Box 1) at birth and characteristic craniofacial changes; however, other traits, such as cardiac defects and susceptibility to leukemias, affect only a subset of individuals with DS (OMIM 190685; ORPHA870). Later in life, the majority of DS individuals will develop Alzheimer's disease (AD; approximately 60% by the age of 65), making trisomy 21 the most common genetic cause of this neurodegenerative disease (Ballard et al., 2016; Dekker et al., 2015; Head et al., 2015; Wiseman et al., 2015).

The phenotypes observed in DS are likely to arise because of dosage sensitivity of Hsa21 genes and associated gene-environment interactions (Antonarakis et al., 2004; Antonarakis, 2016; Beach et al., 2017), and/or a global effect of the extra chromosome on chromatin regulation and methylation (Letourneau et al., 2014; Hervé et al., 2016; Mendioroz et al., 2015). Studies of patients carrying rare segmental duplications of Hsa21 subregions have highlighted the role of specific chromosomal regions in DS pathophysiology (Korbel et al., 2009; Korenberg et al., 1994; Lyle et al., 2009; Delabar et al., 1993). In addition, studies using animal models have confirmed the involvement of homologous regions and shown how some regions with orthologues of individual Hsa21 dosage-sensitive genes are key for DS features (discussed in detail below). A few genes not located on Hsa21 have been shown to contribute to individual phenotypic variation (Roper and Reeves, 2006). Analysis of individuals with a segmental duplication of Hsa21 has been key to building up a phenotypic map and defining a critical DS region (Delabar et al., 1993; Lyle et al., 2009; Korbel et al., 2009; Korenberg et al., 1994; Rahmani et al., 1989). Nevertheless, with only about 60 duplications reported so far in the literature, the resolution of this map is very low. Moreover, duplications that do not induce strong phenotypes, or that lead to embryonic death, are not represented in published studies (Rovelet-Lecrux et al., 2006). A more detailed understanding of the DS genotype-phenotype relationship in humans would require a systematic analysis of very large numbers of individuals and of stillborns. Indeed, 31-54% of DS pregnancies lead to spontaneous foetal loss (Loane et al., 2013; Morris et al., 1999; Morris and Wald, 2007).

This Review focuses on the use of rodent models of DS, which have been essential for the determination of genotype-phenotype relationships for this syndrome. Owing to the genetic tractability of

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**Box 1. Glossary**

**Aneuploid:** having an abnormal or unbalanced number of chromosomes.

**Cardiac septation:** partitioning of the heart.

**Contextual and auditory-cue-conditioned fear task:** a test to study associative memory based on the association of environmental cues (the chamber for the context or a sound for the auditory cue) with an aversive stimulus (a light electric shock). The association of both stimuli will lead to a freezing, with almost no movement of the animal tested. Recording the percentage of immobility of the mouse after being placed back in the environment or with the auditory cue 24 h after the shock gives an assessment of the associative memory.

**Euploid:** having a normal balanced number of chromosomes.

**Hypotonia:** a state of low muscle tone.

**Long-term potentiation (LTP):** an increase in synaptic response after high-frequency stimulation of neurons. A strategy used to test the plasticity and the consolidation of synapses.

**Morris water maze:** a test to study spatial memory, based on the normal behaviour of a mouse to exit a water maze using an external visual cue located outside a pool in which a small platform is hidden below the surface of the water. The path with the distance travelled and the time spent to reach the platform is indicative of function of spatial memory.

**Mosaic:** animal carries cells with different genotypes.

**Novel-object recognition:** a test to study non-spatial episodic memory based on the normal interest of the animal in exploring objects in its environment (an open field). The test evaluates recognition memory for previously explored objects by measuring the time spent sniffing a known versus a novel object.

**Positive thigmotaxis:** a behavioural preference displayed by some animals to be near or in close contact with the solid wall of an enclosure. The time spent in contact or close to the vertical wall of the open field is measured, in the exploration of a new environment.

**Transchromosomal:** a transgenic animal carrying a chromosome from a different species.

**Trisomy:** having a third copy of a given chromosome. Trisomy is associated with a number of human disorders, including Down syndrome.

**Ventriculomegaly:** enlargement or dilation of lateral brain ventricles.

**Y-maze:** a test involving a maze with three arms, which provides the animal with a choice: to visit the arm visited before or go to a new arm. This is a test that measures working memory.

this animal, the most useful DS models to date have been derived from the laboratory mouse. Mice are highly amenable to genome engineering, including through chromosome engineering, to generate precisely defined large genomic segmental duplications to model chromosomal disorders (Brault et al., 2006; Yu and Bradley, 2001; Ramirez-Solis et al., 1995; Hérault et al., 1998; Tybulewicz and Fisher, 2006). Mouse models have also provided platforms for testing interactions between cell and tissue types, responses in the organism, and candidate therapeutics for DS. We highlight the approaches and technologies that have been used to generate mouse models of DS in recent years, and also discuss how the study of these models has brought new knowledge about DS pathophysiology, including key candidate pathways and genes, as well as providing new therapeutic approaches.

**Building up a compendium of DS models**

The rapid development of genetic engineering in recent years has stimulated the generation of multiple DS mouse models. A variety of transgenic models for candidate genes were developed in early attempts at modelling DS in mice (Dierrsens et al., 2009), and we discuss such experiments briefly in a later section. Here, we discuss DS mouse models that contain larger trisomic or duplicated

chromosomal segments, thereby mimicking the trisomy observed in humans.

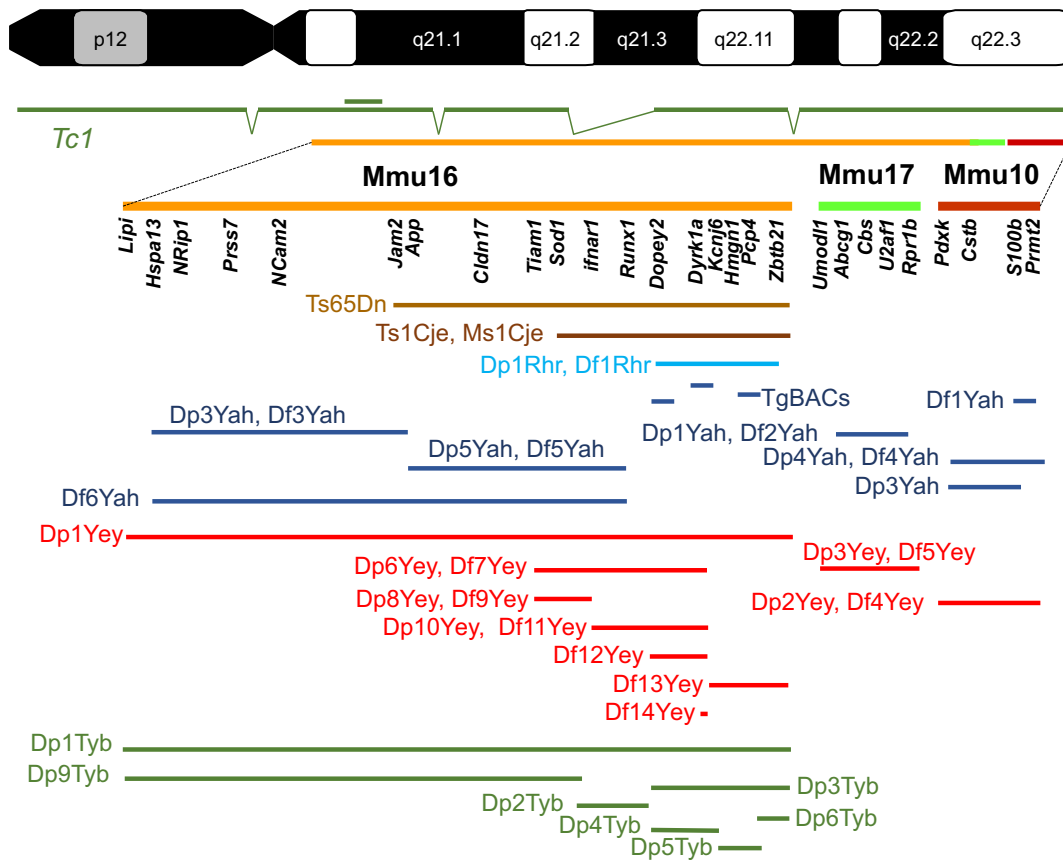
**Early mouse models of trisomy 21**

Over the approximately 75-million years that separate humans and mice in evolutionary time, the chromosomes have rearranged such that Hsa21 has three orthologous regions on mouse chromosomes 10, 16, 17 in which gene order and orientation are conserved (Mmu10, 16, 17; Mmu for *Mus musculus*; Fig. 1). Hsa21 carries 222 protein-coding genes, including 49 that encode keratin-associated proteins and are clustered on Hsa21q, and 325 non-protein-coding genes (Gupta et al., 2016). Of the 158 mouse genes that are homologous to human protein-coding genes, most of them lie on Mmu16 (a total of 102) between *Lipi* and *Zbtb21*, a few on Mmu17 (19) between *Abcg1* and *Rrp1b*, and the rest on Mmu10 (37) between *Pdxk* and *Prmt2*. Of the non-coding genes, 75 elements, such as those encoding miRNAs, are well conserved and are distributed across all three mouse chromosomes (Gupta et al., 2016). Because the equivalent genetic elements are distributed between regions on three different chromosomes, modelling trisomy 21 in the mouse is not straightforward (Antonarakis et al., 2004). In addition, a handful of human genes [such as the prostate-, ovary-, testis- and placenta-expressed ankyrin domain family member D (*POTED*)] are not conserved in the mouse, and there are mouse genes, such as integrin beta-2-like (*Itgb2l*) located between *Igsf5* and *Pcp4* in the Hsa21 homologous regions, that have no human homologues. Furthermore, we do not yet have a clear picture of the role and function (if any) of many pseudogenes (Gupta et al., 2016).

Early modelling was attempted by studying mice with full trisomy of mouse 16 (Gropp et al., 1975; Gropp, 1974). These animals have numerous defects, including, for example, cardiac septation deficits (Box 1) (Webb et al., 1999); however, they do not model DS because the majority of genes that are triplicated in this model are from regions of Mmu16 without homology to Hsa21. Furthermore, these animals die at birth and so cannot give insight into processes beyond this stage.

The field of DS investigation moved forward by the discovery in 1990 and the phenotypic description in 1995 of the Ts65Dn mouse (Fig. 1) (Reeves et al., 1995; Davissou et al., 1990). This mouse has a translocation that results in an extra-small chromosome made up of a fusion of the *App-Zbtb21* region orthologous to Hsa21 found on Mmu16 with the centromeric region of Mmu17; thus, the mouse shows aneuploidy (Box 1). The extra region of Mmu16 includes 90 conserved protein-coding Hsa21 gene orthologues (Choong et al., 2015; Gupta et al., 2016). The Ts65Dn mouse was the main model used to study DS for at least two decades and has provided many new insights (see below). However, the animal carries three copies of an extra segment (arising from Mmu17) with non-DS-related genes, including ~35 protein-coding genes, 15 non-protein-coding genes and 10 pseudogenes (Duchon et al., 2011b; Reinholdt et al., 2011). Moreover, even though some Ts65Dn males are fertile (Moore et al., 2010), transmission is usually achieved through the maternal germline. This might affect the phenotype of the trisomic progeny and their disomic littermates because the mothers are trisomic, generally unlike the situation in humans.

Other models of partial trisomy 16, the Ts1Cje (trisomic for the *Sod1-Zbtb21* region, shown in Fig. 1) and the Ts2Cje (harbours a Robertsonian translocation between the extra chromosome in Ts65Dn and mouse chromosome 12) (Fig. 1, Table 1), have made important contributions to our understanding of DS. Nevertheless,



**Fig. 1. Mouse models of DS.** Human chromosome 21 (p and q arms; G-banding) is depicted at the top of the figure, with the mouse genome orthologous region found on chromosome 16 (Mmu16), Mmu10 and Mmu17 shown respectively in orange, light green and red. A few known genes that are homologous to Hsa21 genes in the DS critical region are listed below each chromosome. The transchromosomal Tc1 mouse model is shown in dark green, with deletions and a duplication (double bar) relative to Hsa21 depicted. Below, the segment of the DS critical region encompassed in different mouse models for DS is illustrated. The original Ts65Dn (Reeves et al., 1995) and Ts1Cje (Sago et al., 1998) models (shown in brown) originated by accidental translocation of Mmu16 segments respectively on Mmu17 and Mmu12, with some additional changes (Duchon et al., 2011b; Reinholdt et al., 2011). Olson et al. (2004) published the first engineered duplication (Dp) and deletion [deficiency (Df)] for the DS critical region (light blue). New models have been developed in the last 10 years by the authors of this Review, as shown in dark blue (Duchon et al., 2011a; Lopes Pereira et al., 2009; Besson et al., 2007; Marechal et al., 2015; Sahun et al., 2014; Raveau et al., 2012; Arbogast et al., 2015; Brault et al., 2015b), red (Jiang et al., 2015; Liu et al., 2011, 2014; Yu et al., 2010a,b,c; Li et al., 2007) and green (Lana-Elola et al., 2016). TgBACs, a few models for BAC or PAC (P1-derived artificial chromosome) transgenic lines.

as with Ts65Dn, they were generated by chance rather than design, and carry additional genetic modifications that could have an impact on phenotypes (Sago et al., 1998; Villar et al., 2005).

**Advances in engineering DS mouse models**

The field of DS modelling in mice changed significantly in the mid-2000s with the advent of two new types of mice: one that is transchromosomal (Box 1) and those that are chromosome engineered.

In 2005, V.L.J.T., E.M.C.F. and colleagues published the first transchromosomal mouse line (O’Doherty et al., 2005), namely Tc1 [formally called Tc(Hsa21)1TybEmcf]. The line was generated using irradiation microcell-mediated chromosome transfer into embryonic stem (ES) cells, leading to a freely segregating copy of Hsa21, transmitted through the germline. In Tc1, human Hsa21 sequences are expressed in the mouse at the mRNA, protein and functional levels (Ahmed et al., 2013; Reynolds et al., 2010; O’Doherty et al., 2005). For example, targeting the overexpressed transcripts encoded by four genes restored VEGF-dependent normal angiogenic responses in Tc1 mice (Reynolds et al., 2010). However, the human chromosome is lost stochastically from cells, and so the resulting mice are mosaics (Box 1): some cells carry the

supernumerary Hsa21, whereas others do not. Also, complete sequencing of the human chromosome (Gribble et al., 2013) has revealed that it was rearranged in the process, probably due to  $\gamma$ -ray-induced *de novo* rearrangements, leading to incomplete trisomy. Nevertheless, this mouse remains a unique complement to the models that help us to understand DS and has given insight into the condition (Hall et al., 2016; Peiris et al., 2016; Powell et al., 2016; Witton et al., 2015).

In the mid-1990s, an approach to generate precise chromosomal rearrangements, including duplications and deletions, was developed in mice (Ramirez-Solis et al., 1995). This technology has radically expanded the available mouse resources for understanding DS by facilitating the design of partial trisomies (Yu and Bradley, 2001; Olson et al., 2004) and producing the most complete model we have: the triple trisomic mouse. This mouse is partially trisomic for the Mmu10, 16 and 17 regions that are homologous to Hsa21 (Yu et al., 2010b). Briefly, the desired chromosomal rearrangement is first engineered in mouse ES cells with two steps of gene targeting to introduce *loxP* sites upstream and downstream of the region of interest with selectable markers, and one additional step leading to the reconstruction of a selectable minigene after Cre expression (Fig. 2). A specific orientation of

**Table 1. Mouse models of DS**

MGI-approved name [lab name]	Type of mutation/ allele	Short description	Genetic background	Chromosomal position (GRCm38 <sup>1</sup> )	First publication	Availability
Tc(HSA21) 1TybEmcf [Tc1]	Transchromosomal	This mouse strain contains a freely segregating copy of Hsa21. The mice are mosaics in that they stochastically lose a copy of Hsa21 from individual cells. Hsa21 has internal deletions and some rearrangements; thus, approximately 75% of Hsa21 is present in the mouse. On average 70% of brain nuclei carry Hsa21 in the Tc1 mouse (O'Doherty et al., 2005).	Mice are bred on F1 of 129S8 and C57BL/6J	NA	O'Doherty et al., 2005	EMMA and Jax
Ts(17<16>)65Dn [Ts65Dn]	Radiation induced	This trisomic mouse contains a segment that is homologous to Hsa21 from <i>Mrp139</i> to the end of the Mmu16, fused to a minichromosome 17 centromeric region.	B6C3B or B6EiC3Sn	Unknown	Reeves et al., 1995	Jax
Ts(16C-tel)1Cje [Ts1Cje]	Spontaneous allele	Mice are trisomic for the genetic interval <i>Sod1</i> to <i>Mx1</i> . <i>Sod1</i> is not functionally trisomic because the <i>Sod1</i> gene in the translocated segment is inactivated by the insertion of the <i>neo</i> sequence. The model is trisomic for 94 genes on Mmu16, covering approximately two-thirds of the trisomic region in Ts65Dn mice. Three genes from Mmu12 are missing (Laffaire et al., 2009).	B6EiC3Sn (or B6 congenic available)	Unknown	Sago et al., 1998	Jax
Ts(17<16>)65Dn 2Cje [Ts2Cje]	Robertsonian translocation	A female mouse with 40 chromosomes and 41 arms was detected by chromosome analysis of progeny from Ts(17<16>)65Dn mice. This mouse had a unique biallelic chromosome that appeared to be the result of a fusion between Mmu12 and the marker chromosome Ts(17<16>)65Dn.	B6EiC3Sn	Unknown	Villar et al., 2005	Jax
Dp(16Cbr1-Fam3b) 1Rhr [Ts1Rhr; Dp1Rhr]	Duplication	Cre-mediated chromosomal rearrangement on Mmu16 between <i>loxP</i> sites in regions proximal to <i>Cbr1</i> and distal to <i>Fam3b</i> results in duplication of the genes, including <i>Cbr1</i> and <i>Fam3b</i> .	B6.129S6	16:93607837-97504936	Olson et al., 2004	Jax
Del(16Cbr1-Fam3b)1Rhr [Ms1Rhr; Df1Rhr]	Deletion	Cre-mediated chromosomal rearrangement on Mmu16 between <i>loxP</i> sites in regions proximal to <i>Cbr1</i> and distal to <i>Fam3b</i> results in deletion of the genes, including <i>Cbr1</i> and <i>Fam3b</i> .	B6.129S6	16:93607837-97504936	Olson et al., 2004	Jax
Dp(10Prmt2-Pdxk) 2Yey [Dp(10) 1Yey; Dp2Yey]	Duplication	Cre-mediated chromosomal rearrangement on Mmu10 between <i>loxP</i> sites in regions proximal to <i>Prmt2</i> and distal to <i>Pdxk</i> results in duplication of the genes, including <i>Prmt2</i> and <i>Pdxk</i> .	B6N4	10:76207227-78464975	Yu et al., 2010b	Jax
Del(10Prmt2-Pdxk) 4Yey [Df(10) 1Yey; Df2Yey]	Deletion	Cre-mediated chromosomal rearrangement on Mmu10 between <i>loxP</i> sites in regions proximal to <i>Prmt2</i> and distal to <i>Pdxk</i> resulted in deletion of the genes, including <i>Prmt2</i> and <i>Pdxk</i> .	Mixed 129S7 and 129S1	10:76207227-78464975	Yu et al., 2010a	Jax
Dp(17Abcg1-Rrp1b)3Yey [Dp (17)1Yey; Dp3Yey]	Duplication	Cre-mediated chromosomal rearrangement on Mmu17 between <i>loxP</i> sites in regions proximal to <i>Abcg1</i> and distal to <i>Rrp1b</i> results in duplication of the genes, including <i>Abcg1</i> and <i>Rrp1b</i> .	B6N4	17:31057698-32062865	Yu et al., 2010b	Jax
Del(17Abcg1-Rrp1b)5Yey [Df (17)1Yey; Df5Yey]	Deletion	Cre-mediated chromosomal rearrangement on Mmu17 between <i>loxP</i> sites in regions proximal to <i>Abcg1</i> and distal to <i>Rrp1b</i> resulted in the deletion of the genes, including <i>Abcg1</i> and <i>Rrp1b</i> .	Mixed 129S7 and 129S1	17:31057698-32062865	Yu et al., 2010a	Jax
Dp(16Lipi-Zbtb21) 1Yey [Dp(16) 1Yey; Dp1Yey]	Duplication	Cre-mediated chromosomal rearrangement on Mmu16 between <i>loxP</i> sites in regions proximal to <i>D930038D03Rik</i> ( <i>Lipi</i> ) and distal to <i>Zfp295</i> ( <i>Zbtb21</i> ) resulted in the duplication of the genes, including <i>Lipi</i> and <i>Zbtb21</i> .	B6N5	NA	Li et al., 2007	Jax

Continued

Table 1. Continued

MGI-approved name [lab name]	Type of mutation/ allele	Short description	Genetic background	Chromosomal position (GRCm38 <sup>1</sup> )	First publication	Availability
Dp(16Tiam1-Kcnj6)6Yey [Dp(16)2Yey; Dp6Yey]	Duplication	Cre-mediated chromosomal rearrangement on Mmu16 between <i>loxP</i> sites in regions proximal to <i>Tiam1</i> and distal to <i>Kcnj6</i> resulted in the duplication of the genes, including <i>Tiam1</i> and <i>Kcnj6</i> .	Mixed 129S7 and 129S1	16:89787111-94997696	Liu et al., 2011	Jax
Del(16Tiam1-Kcnj6)7Yey [Df(16)2Yey; Df7Yey]	Deletion	Cre-mediated chromosomal rearrangement on Mmu16 between <i>loxP</i> sites in regions proximal to <i>Tiam1</i> and distal to <i>Kcnj6</i> resulted in the deletion of the genes, including <i>Tiam1</i> and <i>Kcnj6</i> .	Mixed 129S7 and 129S1	16:89787111-94997696	Liu et al., 2011	Jax
Dp(16Tiam1-Il10rb)8Yey [Dp(16)3Yey; Dp8Yey]	Duplication	Cre-mediated chromosomal rearrangement on Mmu16 between <i>loxP</i> sites in regions proximal to <i>Tiam1</i> and distal to <i>Il10rb</i> resulted in the duplication of the genes, including <i>Tiam1</i> and <i>Il10rb</i> .	Mixed 129S7 and 129S1	NA	Liu et al., 2014	Jax
Del(16Tiam1-Il10rb)9Yey [Df(16)3Yey; Df9Yey]	Deletion	Cre-mediated chromosomal rearrangement on Mmu16 between <i>loxP</i> sites in regions proximal to <i>Tiam1</i> and distal to <i>Il10rb</i> resulted in the deletion of the genes, including <i>Tiam1</i> and <i>Il10rb</i> .	Mixed 129S7 and 129S1	NA	Liu et al., 2014	Jax
Dp(16Irfar1-Kcnj6)10Yey [Dp(16)4Yey; Dp4Yey]	Duplication	Cre-mediated chromosomal rearrangement on Mmu16 between <i>loxP</i> sites in regions proximal to <i>Irfar1</i> and distal to <i>Kcnj6</i> resulted in the duplication of the genes, including <i>Irfar1</i> and <i>Kcnj6</i> .	Mixed 129S7 and 129S1	NA	Liu et al., 2014	Jax
Del(16Irfar1-Kcnj6)11Yey [Df(16)4Yey; Df11Yey]	Deletion	Cre-mediated chromosomal rearrangement on Mmu16 between <i>loxP</i> sites in regions proximal to <i>Irfar1</i> and distal to <i>Kcnj6</i> resulted in the deletion of the genes, including <i>Irfar1</i> and <i>Kcnj6</i> .	Mixed 129S7 and 129S1	NA	Liu et al., 2014	Jax
Del(16Setd4-Kcnj6)12Yey [Df(16)5Yey; Df12Yey]	Deletion	Cre-mediated chromosomal rearrangement on Mmu16 between <i>loxP</i> sites in regions proximal to <i>Setd4</i> and distal to <i>Kcnj6</i> resulted in the deletion of the genes, including <i>Setd4</i> and <i>Kcnj6</i> .	C57BL/6J (>N6)	NA	Liu et al., 2014	Jax
Del(16Kcnj15-Mx2)13Yey [Df(16)6Yey; Df13Yey]	Deletion	Cre-mediated chromosomal rearrangement on Mmu16 between <i>loxP</i> sites in regions proximal to <i>Kcnj15</i> and distal to <i>Mx2</i> resulted in the deletion of the genes, including <i>Kcnj15</i> and <i>Mx2</i> .	C57BL/6J (>N6)	NA	Jiang et al., 2015	Jax
Del(16Dyrk1a-Kcnj6)14Yey [Df(16)7Yey; Df14Yey]	Deletion	Cre-mediated chromosomal rearrangement on Mmu16 between <i>loxP</i> sites in regions proximal to <i>Dyrk1a</i> and distal to <i>Kcnj6</i> resulted in the deletion of the genes, including <i>Dyrk1a</i> and <i>Kcnj6</i> .	C57BL/6J (>N6)	NA	Jiang et al., 2015	Jax
Dp(17Abcg1-Cbs)1Yah [Ts1Yah; Dp1Yah]	Duplication	Cre-mediated chromosomal rearrangement on Mmu17 between <i>loxP</i> sites inserted in <i>Abcg1</i> and downstream of <i>Cbs</i> (before <i>U2af1</i> ) region resulted in the deletion of <i>Abcg1</i> and the duplication of the segment homologous and syntenic to Hsa21q22.	C57BL/6J (>N6)	17:31057698-31637199	Lopes Pereira et al., 2009	EMMA
Dp(16Hspa13-App)2Yah [Ts2Yah; Dp2Yah]	Duplication	Cre-mediated chromosomal rearrangement on Mmu16 between <i>loxP</i> sites inserted in <i>Hspa13</i> and <i>App</i> resulted in the inactivation of both flanking genes and the duplication of the segment homologous to Hsa21q11-q21.	C57BL/6J (>N6)	16:75755190-85173707	Brault et al., 2015a	EMMA
Dp(10Cstb-Prmt2)3Yah [Ts3Yah; Dp3Yah]	Duplication	Cre-mediated chromosomal rearrangement on Mmu10 between <i>loxP</i> sites inserted in <i>Cstb</i> and <i>Col6a1</i> resulted in the inactivation of both flanking genes and the duplication of the segment homologous to Hsa21.	C57BL/6J (>N6)	NA	Duchon et al., 2008	Y.H. lab
Dp(10Cstb-Prmt2)4Yah [Ts4Yah; Dp4Yah]	Duplication	Cre-mediated chromosomal rearrangement on Mmu10 between <i>loxP</i> sites inserted in <i>Cstb</i> and <i>Prmt2</i> resulted in the inactivation of both flanking genes and the duplication of the segment homologous to Hsa21.	C57BL/6J (>N6)	NA	Duchon et al., 2008	Y.H. lab

Continued

Table 1. Continued

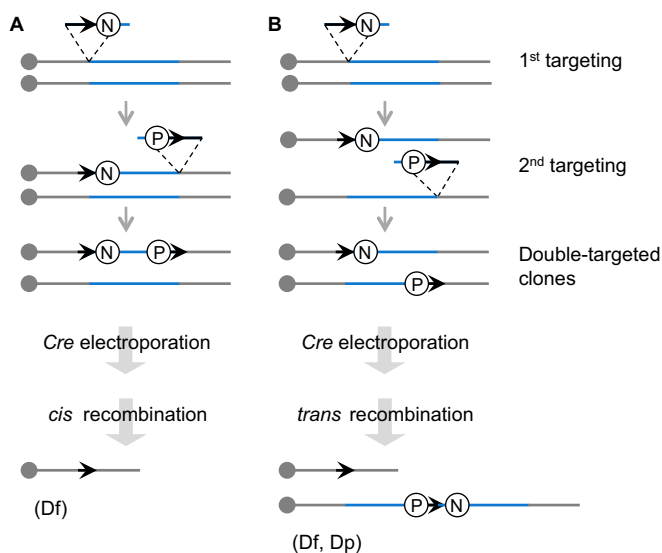
MGI-approved name [lab name]	Type of mutation/ allele	Short description	Genetic background	Chromosomal position (GRCm38 <sup>1</sup> )	First publication	Availability
Dp(16App-Runx1) 5Yah [Ts5Yah; Dp5Yah]	Duplication	Cre-mediated chromosomal rearrangement on Mmu16 between <i>loxP</i> sites inserted in <i>App</i> and <i>Runx1</i> resulted in the inactivation of both flanking genes and the duplication of the segment homologous to Hsa21.	C57BL/6J (>N6)	16:84954440-92826149	Raveau et al., 2012	Y.H. lab
Del(16App-Runx1) 5Yah [Ms5Yah; Df5Yah]	Deletion	Cre-mediated chromosomal rearrangement on Mmu16 between <i>loxP</i> sites inserted in <i>App</i> and <i>Runx1</i> resulted in the inactivation of both flanking genes and the deletion of the segment homologous to Hsa21q22.	C57BL/6J (>N6)	16:84954440-92826149	Raveau et al., 2012	Y.H. lab
Del(16Hspa13-App)3Yah [Ms3Yah; Df3Yah]	Deletion	Cre-mediated chromosomal rearrangement on Mmu16 between <i>loxP</i> sites inserted in <i>Hspa13</i> and <i>App</i> resulted in the inactivation of both flanking genes and the deletion of the segment homologous to Hsa21q22.	C57BL/6J (>N6)	16:75755190-85173707	Brault et al., 2015a	EMMA
Del(17Abcg1-Cbs) 2Yah [Ms2Yah; Df2Yah]	Deletion	Cre-mediated chromosomal rearrangement on Mmu16 between <i>loxP</i> sites inserted in <i>Abcg1</i> and <i>Cbs</i> resulted in the inactivation of both flanking genes and the deletion of the segment homologous to Hsa21q22.	C57BL/6J (>N6)	17:31057698-31637199	Lopes Pereira et al., 2009	EMMA
Del(10Prmt2-Col6a1)1Yah [Ms1Yah; Df1Yah]	Deletion	Cre-mediated chromosomal rearrangement on Mmu16 between <i>loxP</i> sites inserted in <i>Col6a1</i> and <i>Prmt2</i> resulted in the inactivation of both flanking genes and the deletion of the segment homologous to Hsa21q22.	C57BL/6J (>N6)	10:76207227-76726168	Besson et al., 2007	EMMA
Del(10Prmt2-Cstb) 4Yah [Ms4Yah; Df4Yah]	Deletion	Cre-mediated chromosomal rearrangement on Mmu16 between <i>loxP</i> sites inserted in <i>Cstb</i> and <i>Prmt2</i> resulted in the inactivation of both flanking genes and the deletion of the segment homologous to Hsa21q22.	C57BL/6J (>N6)	10:76207227-78427619	Duchon et al., 2008	EMMA
Dp(16Lipi-Zbtb21) 1TybEmcf [Dp1Tyb]	Duplication	Cre-mediated chromosomal rearrangement on Mmu16 to duplicate the region between two <i>loxP</i> sites inserted proximal to <i>Lipi</i> and distal to <i>Zbtb21</i> .	C57BL/6J.129P2	16:74930370-97982380	Lana-Elola et al., 2016	EMMA
Dp(16Mis18a-Runx1)2TybEmcf [Dp2Tyb]	Duplication	Cre-mediated chromosomal rearrangement on Mmu16 to duplicate the region between two <i>loxP</i> sites inserted proximal to <i>Mis18a</i> and distal to <i>Runx1</i> .	C57BL/6J.129P2	16:90563769-93062456	Lana-Elola et al., 2016	EMMA
Dp(16Mir802-Zbtb21) 3TybEmcf [Dp3Tyb]	Duplication	Cre-mediated chromosomal rearrangement on Mmu16 to duplicate the region between two <i>loxP</i> sites inserted proximal to <i>Mir802</i> and distal to <i>Zbtb21</i> .	C57BL/6J.129P2	16:93054020-97982380	Lana-Elola et al., 2016	EMMA
Dp(16Mir802-Dscr3)4TybEmcf [Dp4Tyb]	Duplication	Cre-mediated chromosomal rearrangement on Mmu16 to duplicate the region between two <i>loxP</i> sites inserted proximal to <i>Mir802</i> and distal to <i>Dscr3</i> .	C57BL/6J.129P2	16:93054020-94546849	Lana-Elola et al., 2016	EMMA
Dp(16Dyrk1a-B3galt5) 5TybEmcf [Dp5Tyb]	Duplication	Cre-mediated chromosomal rearrangement on Mmu16 to duplicate the region between two <i>loxP</i> sites inserted proximal to <i>Dyrk1a</i> and distal to <i>B3galt5</i> .	C57BL/6J.129P2	16:94538615-96331804	Lana-Elola et al., 2016	EMMA
Dp(16Igsf5-Zbtb21) 6TybEmcf [Dp6Tyb]	Duplication	Cre-mediated chromosomal rearrangement on Mmu16 to duplicate the region between two <i>loxP</i> sites inserted proximal to <i>Igsf5</i> and distal to <i>Zbtb21</i> .	C57BL/6J.129P2	16:96327324-97982380	Lana-Elola et al., 2016	EMMA
Dp(16Lipi-Hunk) 9TybEmcf [Dp9Tyb]	Duplication	Cre-mediated chromosomal rearrangement on Mmu16 to duplicate the region between two <i>loxP</i> sites inserted proximal to <i>Lipi</i> and distal to <i>Hunk</i> .	C57BL/6J.129P2	16:74930370-90577148	Lana-Elola et al., 2016	EMMA

Further details about the mouse strains listed here can be found at the Jackson Laboratory (Jax; [www.jax.org/](http://www.jax.org/)) or European Mutant Mouse Archive (EMMA; ([www.infrafrontier.eu/search](http://www.infrafrontier.eu/search))) websites.

<sup>1</sup>GRCm28 refers to mouse Genome\_Sequence\_build: GRCm28/UCSC mm10.

Hsa21, human chromosome 21; MGI, mouse genome informatics; Mmu, *Mus musculus* chromosome; NA, not available; neo, neomycin resistance.

*loxP* targeted on a chromosome must be achieved by appropriate design to induce the chromosomal change. Identification of ES cell clones harbouring an engineered chromosomal rearrangement is facilitated by positive selection of expression of the minigene. Chromosomal duplications and deletions can be precisely verified by Southern blot analysis, fluorescence *in situ* hybridisation,



**Fig. 2. Cre-*loxP*-mediated chromosomal engineering in mice.** A *loxP* site (arrow) is targeted into the first endpoint of the engineered segment (blue) in the embryonic stem (ES) cell genome with a positive selectable marker, such as *neo* (the neomycin-resistance gene; N). Next, a second *loxP* site is targeted to the other endpoint with another positive selectable marker such as *puro*, the puromycin resistance gene (P). A Cre expression vector is then transferred by electroporation into double-targeted ES cell clones. If two *loxP* sites are targeted onto the same chromosome homologue and oriented in the same direction (*cis*), recombination between the sites will lead to a deletion (Df; A). If two *loxP* sites are targeted onto two separate homologues and oriented in the same direction (*trans*), the recombination will lead to a duplication (Dp) and the reciprocal deletion (Df) (B).

array-based comparative genome hybridisation or whole-genome sequencing (Yu et al., 2010b; Gribble et al., 2013). Selected ES cells are used to establish the corresponding mouse line. Alternatively, recombination between *loxP* sites can be achieved *in vivo* (Brault et al., 2007; Héroult et al., 1998) in a mouse carrying the two *loxP* sites and one specific Cre driver expressed in the germline. In this case, no additional construct is needed at the frontier of the recombined fragments, minimising potential interference associated with the reconstruction of a minigene. Altogether, these Cre/*loxP*-based technologies, carried out independently in two laboratories, resulted in the generation of several mouse models with segmental duplications encompassing different segments of the mouse chromosomes orthologous to Hsa21 (Héroult et al., 2012).

The crucial step needed for inserting *loxP* sites is now facilitated using recombinant transposon-mediated insertion and further selection (Chen et al., 2013; Ruf et al., 2011). In addition, *loxP* site insertions in the mouse genome generated by transposition have been captured in the TRACER resource with precise location and orientation (<http://tracerdatabase.embl.de>), reducing the time needed to generate new DS models to 3 years (Chen et al., 2013).

However, a major revolution is now underway with the development of an even faster method – CRISPR-mediated rearrangement (CRISMERE) – which is based on CRISPR/Cas9 genome-editing technology (Birling et al., 2017). Two pairs of small guide RNA (sgRNA), each pair selected either upstream or downstream of the region of interest, are injected with the Cas9 nuclease into one-cell mouse embryos that are reimplanted. Newborns are analysed for chromosomal modifications and are bred to select carrier individuals with the new rearrangement in the next generation. The making of *in vivo* duplications, deletions and

inversions of genomic segments of up to 34 Mb using CRISMERE requires less time than it takes to observe germline transmission of recombined ES cells. CRISMERE is not limited to the mouse and has been successfully used in rats (Table 1) (Birling et al., 2017) and can be applied to primates. In the rat (*Rattus norvegicus*) genome, the Hsa21 homologous regions are located on two chromosomes, Rno11 and Rno20. On Rno11, the *Lipi-Zbtb21* segment is almost identical to the homologous region located on the Mmu16, whereas Rno20 harbours a unique segment for the *Umod1-Prmt2* interval (Fig. 1). Using CRISMERE, new models encompassing both regions have been generated (Birling et al., 2017). These models have the potential to facilitate testing of therapies in both mouse and rat models to enable stronger validation prior to assessment in clinical trials.

Nowadays, the development of DS models is no longer limited at the technical level but rather more at the conceptual level, i.e. in terms of the challenges associated with precise delineation of the region of interest for a particular phenotype. A key question is whether it is better to define smaller regions of interest and generate more models or make models for larger segments to recapitulate the human trisomy. In any case, enhancing our understanding of the link between phenotype and genotype is critical, as discussed below.

### Assessing the genotype-phenotype relationship in DS

Many DS features show variable penetrance (<http://omim.org/entry/190685#clinicalFeatures>) (Kruszka et al., 2017; Roizen and Patterson, 2003). Understanding the molecular basis of this huge variability between individuals could inform the development of therapies to modulate specific features of the syndrome. Variability in DS includes the degree of learning difficulties observed – in 39.4% of cases, IQ typically ranges between 50 and 70, but 1% of affected individuals have an IQ around the borderline function range of 70–80 (Antonarakis et al., 2004). The presence of cardiac anomalies, and the incidence of leukaemia, autoimmune diseases, AD pathology and dementia, as well as accelerated ageing, are also quite variable (Antonarakis et al., 2004). In order to better understand the physiopathology of the disease and to correlate genotype with the features observed in patients, studies in animal models and particularly mouse models have been critical (Gupta et al., 2016).

One of the first consequences of DS is the alteration of embryonic development, leading at the extreme to gestational loss in humans (Loane et al., 2013; Morris et al., 1999; Morris and Wald, 2007). This phenotype is also observed in Tc1 mice, in Dp1Yey and Dp1Tyb mice (generated in two independent groups), which duplicate the region from *Lipi* to *Zbtb21* (Fig. 1), in the Ts65Dn mouse, and to some extent in the Ts1Cje mouse (Arbogast et al., 2015; Raveau et al., 2012; Yu et al., 2010b; Li et al., 2007; Guedj et al., 2016; Lana-Elola et al., 2016; O'Doherty et al., 2005). Reducing the dosage of the 7.7 Mb *App-Runx1* region, containing 54 protein-coding genes and 25 keratin genes, in Ts65Dn mice rescued impaired postnatal viability, and deletion of this region resulted in severe phenotypes and lowered viability, suggesting the presence of critical genes in the interval (Arbogast et al., 2015; Raveau et al., 2012).

In light of the characteristic intellectual deficiency in human DS, many types of learning and memory have been monitored in DS mouse models to explore which part of the brain is affected by the trisomy (Das and Reeves, 2011; Gupta et al., 2016; Xing et al., 2016; Belichenko et al., 2015; Jiang et al., 2015; Zhang et al., 2014; Arbogast et al., 2015; Brault et al., 2015b; Marechal et al., 2015; Sahun et al., 2014; Héroult et al., 2012). The open-field (OF) test (Stanford, 2007) has been used in these studies to assay locomotor



activity, exploration and anxiety. Episodic memory (involving the perirhinal cortex and the hippocampus) was a particular focus, with tests for non-spatial learning such as the novel-object recognition (NOR; Box 1) applied with two retention times: 1 h or 24 h (Cohen and Stackman, 2015). Short-term working memory was assessed with continuous spontaneous alternation behaviour mostly using the Y-maze test (Box 1) (Hughes, 2004). Spatial memory, which involves several regions of the brain (hippocampus, striatum, basal forebrain, cerebellum and cerebral cortex), has been explored in DS mice using the Morris water maze (MWM; Box 1) test (D'Hooge and De Deyn, 2001; Morris, 1984). For associative memory, the contextual and auditory-cue-conditioned fear task [FC (fear conditioning); Box 1] (Paylor et al., 1994; Mátyás et al., 2014; Lee et al., 2011) was used to test the connection between the hippocampus, frontal cortex, cingulate cortex and amygdala and the mediodorsal thalamic nucleus. These studies point to an important role for *DYRK1A* (discussed in the section below). *DYRK1A*, the mammalian orthologue of *Drosophila minibrain kinase (mnb)* (Tejedor et al., 1995), encodes a proline/arginine-directed dual-specificity kinase, and is overexpressed both in the brain of trisomic mice and of individuals with DS (Dowjat et al., 2007). Three copies of *Dyrk1a* are necessary and sufficient to induce several deficits in NOR (at 1 and 24 h) and FC, but only result in delayed learning in the MWM (García-Cerro et al., 2014; Pons-Espinal et al., 2013; Dierssen and de Lagrán, 2006; Altafaj et al., 2001; Duchon and Herault, 2016). Interestingly, the Ts1Rhr trisomy mouse, which is trisomic for the DS critical region with 33 genes including *Dyrk1a*, displayed deficits in the OF test and in NOR, with 24 h of retention (Belichenko et al., 2009); additionally, trisomy of this region was necessary to alter spatial memory in Ts65Dn mice (Olson et al., 2007). In Ts65Dn, Ts1Cje and Ts1Rhr mice, long-term potentiation (LTP; Box 1), a measure of synaptic plasticity, could be induced only after blocking GABA(A)-dependent inhibitory neurotransmission in the fascia dentata, a structure that receives inputs from the perirhinal cortex (Belichenko et al., 2009; Kleschevnikov et al., 2012b); this result is indicative of excessive neuronal inhibition and is consistent with previous observations of Ts65Dn mice (Kleschevnikov et al., 2012b; Fernandez et al., 2007). In addition, widespread enlargement of dendritic spines and decreased density of spines in the fascia dentata were observed, which could explain the overall reduced activation of neuronal activity (Belichenko et al., 2009; Haas et al., 2013). Thus, cognitive impairment in DS seems to derive from molecular and structural changes related to an altered copy number within this 33-gene region. This conclusion was confirmed when combining Dp1Yey mice either with deletion of the *Std4-Kcnj6* interval or *Kcnj15-Mx2*, which showed that both regions contain dosage-sensitive genes contributing to cognitive phenotypes (Jiang et al., 2015).

The Ts65Dn mouse model also displays lower performance in finding a hidden platform compared to controls in the MWM task at 4 months of age (Reeves et al., 1995; Netzer et al., 2010; Olmos-Serrano et al., 2016b), but this phenotype is not consistently observed in 2- to 4-month-old Dp1Yey mice (Yu et al., 2010c; Goodliffe et al., 2016). Nevertheless, learning is impaired for both models in a variant of the MWM test at the age of 2-3 months (Goodliffe et al., 2016; Olmos-Serrano et al., 2016a). Overall, the results obtained from these studies are difficult to compare owing to differences in age of tested individuals and more importantly in the protocols or the genetic background used. Thus, there is a strong need to better standardise experimental protocols to allow for more equivalent cross-laboratory comparisons.

Another important point is that combining models with different segmental trisomies can alter the phenotypic outcomes (Jiang et al., 2015; Duchon et al., 2011a; García-Cerro et al., 2014; Salehi et al., 2006). Studies in such mice strengthen the evidence for the multigenic nature of DS, already pointed to in human genetic studies (Korbel et al., 2009; Korenberg et al., 1994; Lyle et al., 2009), with multiple genes interacting to induce the frequently observed intellectual disability that characterises DS. One of the main conclusions is that the hippocampus is a key hub whose dysfunction is observed in many DS mouse models, altering many types of memory, including, for example, the function of the place cells, a type of hippocampal pyramidal neuron that acts to define a cognitive map needed for spatial memory (Witton et al., 2015).

DS universally causes the typical plaques and tangles of AD to appear in the brain by the age of 40, and current figures show that two-thirds of people with DS develop dementia by the age of 60 (Wiseman et al., 2015). Individuals with DS develop Alzheimer's-like pathologies comparatively early in life, including progressive degeneration of basal forebrain cholinergic neurons (BFCNs). The Ts65Dn mouse model exhibits elevated levels of  $\beta$ -amyloid (A $\beta$ ) peptide, as well as atrophy of BFCNs. Although the mechanisms are not yet fully understood, the appearance of the pathology almost certainly arises from overexpression of the Hsa21 gene *APP*, which is known to cause early-onset AD when present in three copies, as shown in very rare families with small internal chromosomal duplications that include this gene (Rovelet-Lecrux et al., 2007, 2006). In line with this hypothesis, *App* triplication is necessary for the age-dependent BFCN loss observed in the Ts65Dn mouse (Salehi et al., 2006; Granholm et al., 2000) and for neuronal abnormalities in the endosomal compartment, also reported in these mice (Cataldo et al., 2008, 2000).

Intriguingly, not everyone with DS develops dementia, although all individuals with DS over the age of 40 show evidence of amyloid plaques. Individuals with duplicated *APP* have dementia onset that is fully penetrant, between 39 to 64 years of age (Wiseman et al., 2015). Conversely, individuals with DS (and thus *APP* in triplicate) show a wide range in age-of-onset of dementia, and can live into their 70s with no sign of dementia (Karmiloff-Smith et al., 2016; Krinsky-McHale et al., 2008; Ghezzi et al., 2014). Thus, it seems likely that there are also protective factors for AD on Hsa21, and these might be important for understanding and treating dementia in the euploid (Box 1) population. Although the genes involved in modulating AD phenotypes remain to be determined, crosses of different types of AD and DS mouse models to humanised *APP* will give insight into molecular processes (Choong et al., 2015; Hamlett et al., 2016).

Insights into congenital heart defects (CHDs) in DS have also been gained using mouse models. In humans, 40% of DS newborns present with a CHD (Antonarakis et al., 2004). In mice, CHDs were observed during development in Tc1 (O'Doherty et al., 2005), Ts65Dn (Moore, 2006), Dp1Yey (Li et al., 2007), Dp1Tyb (Lana-Elola et al., 2016) and Ts1Cje (Guedj et al., 2016) embryos. Detailed investigation with several trisomic models refined a critical interval between *Ifnar1* and *Kcnj6* (Fig. 1) (Raveau et al., 2012; Liu et al., 2011) and a role for the most distal part of Mmu16 from *mir802* to *Zbtb21* (Lana-Elola et al., 2016). In addition, overexpression of *Jam2*, a gene located upstream of *App* that encodes junctional adhesion molecule 2 found in the tight junctions of endothelial cells, modifies the activity of the matricellular protein cysteine-rich with EGF-like domain protein 1 (CRELD1), leading to enhanced septal defects in Ts65Dn mice (Li et al., 2016). Reducing the dosage of another key gene for heart development located outside of Hsa21 homologous regions, T-box transcription

factor *Tbx5* worsens CHDs in the Ts65Dn mouse model, evidenced by increasing aortic and atrial-ventricular septal defects (Polk et al., 2015). These studies indicate that several Hsa21 homologous regions contribute to CHDs in DS and that additional genes involved in normal heart development can modify the severity of the heart phenotypes observed in DS models.

Additional DS-related features, such as craniofacial changes, have been described in mouse models, and certain trisomic genes or regions have been implicated in these phenotypes (Starbuck et al., 2014; Richtsmeier et al., 2002, 2000). A mouse-based study has also shed light on hypotonia, a major phenotype observed in DS newborns (Vicari, 2006). Analysis of a mouse model carrying three copies of the *Hspa13-App* region (Fig. 1) shows changes in locomotor activity and in muscle strength and physiology, suggesting the contribution of muscular deficits in the periphery (Braut et al., 2015a).

This summary is not exhaustive and many additional DS features have been explored using mouse models, such as the mineralisation of long bones (Blazek et al., 2015), the risk of chronic otitis media (Bhutta et al., 2013), and the higher risk of developing myeloproliferative disorders and cancer (Ng et al., 2015; Sussan et al., 2008; Malinge et al., 2012; Alford et al., 2010; Yang et al., 2016). Each mouse strain has the potential to provide unique information, and a range of animals, aneuploid and partially trisomic, are important to pinpoint specific candidate genes and dissect the underlying molecular mechanisms.

#### Identification of molecular mechanisms and candidate genes for DS cognitive features

The analysis of DS mouse models has facilitated the identification of specific Hsa21 genes involved in DS features. Initially, DS candidate genes were pinpointed by human genetic analyses or by parallel knowledge of gene function (Antonarakis, 2016; Antonarakis et al., 2004). Two basic experimental approaches in mice have been applied either to increase or decrease the expression of a candidate gene. As described above, a 33-gene region has been identified as being crucial for cognitive impairment in DS, based on a number of mouse behavioural studies (Belichenko et al., 2009; Olson et al., 2007). Among the genes from this region, *Dyrk1a* was an attractive candidate for inducing cognitive-impairment phenotypes.

Large genomic fragments such as yeast artificial chromosomes (YACs) containing mouse DNA of the locus (Smith and Rubin, 1997; Smith et al., 1995) or bacterial artificial chromosome (BAC) constructs covering the human (Ahn et al., 2006) or the mouse (Guedj et al., 2012) gene, were developed in order to express *Dyrk1a* with a pattern of expression similar to the endogenous gene. The YAC transgenic mouse was used to demonstrate the role of *Dyrk1a* in DS cognitive impairment (Sebrié et al., 2008; Rachidi et al., 2007; Roubertoux et al., 2006; Branchi et al., 2004). The evidence for this was reinforced by applying transgenic approaches to overexpress *Dyrk1a* alone in mice either by using expression vectors driven by an exogenous promoter (Altafaj et al., 2001, 2013; Grau et al., 2014; Martinez de Lagrán et al., 2004) or by using BAC encoding human or mouse *DYRK1A* (Ahn et al., 2006; Guedj et al., 2012). All lines played an important role in understanding the molecular consequences induced by *DYRK1A* overdosage and provided important support for demonstrating molecular alterations in synaptic plasticity pathways, particularly expression changes in GABAergic- and glutamatergic-related proteins (Ahn et al., 2006; Park et al., 2012, 2010; Song et al., 2015, 2013; Souchet et al., 2015; Rachdi et al., 2014; Laguna et al., 2013; Guedj et al., 2012; Souchet

et al., 2014). Similar alterations were observed in models with partial trisomy of Mmu16, Ts65Dn and Dp(16)1Yey, and were reversed in the *Dyrk1a*<sup>+/-</sup> model (Souchet et al., 2014). Overexpression of *Dyrk1a* also decreased firing rate and  $\gamma$ -frequency power in the prefrontal network of anaesthetised and awake mice, indicating that excess levels of this gene reinforce neuronal inhibition (Ruiz-Mejias et al., 2016).

Reducing the *Dyrk1a* dosage in Ts65Dn mice by crossing Ts65Dn females with heterozygous *Dyrk1a*<sup>+/-</sup> male mice revealed that normalization of the *Dyrk1a* copy number improves spatial working, reference memory and contextual conditioning, as well as rescuing hippocampal LTP (García-Cerro et al., 2014). Similar results were obtained by crossing Dp1Yey mice with *Dyrk1a*-knockout mice: rescued trisomic mice with only two functional copies of *Dyrk1a* showed a better performance in the T-maze and FC assays (Jiang et al., 2015). Concomitant with these functional improvements, normalisation of the *Dyrk1a* expression level in trisomic mice restored the proliferation and differentiation of hippocampal cells in the adult dentate gyrus (DG), and the density of GABAergic- and glutamatergic-synapse markers in the molecular layer of the hippocampus (García-Cerro et al., 2014).

Additional genes have been implicated in DS phenotypes in mice. *App* triplication was shown to impact BFCN degeneration, consistent with a role for this AD-associated gene in DS impaired cognition (Salehi et al., 2006). Regulator of calcineurin 1 (*Rcan1*) inhibits the calcineurin-dependent signalling pathway and its aggregation is controlled by *Dyrk1a* phosphorylation (Song et al., 2013). Increasing *RCAN1* gene dosage impairs hippocampal LTP (Xing et al., 2013), whereas genetic rescue experiments restore sympathetic nervous system development in Dp1Yey mice (Patel et al., 2015). The gene encoding calmodulin regulator protein Purkinje cell protein 4 (*Pcp4*) has been overexpressed using a P1-phage vector (PAC) to generate transgenic mice that display cerebellar defects (Mouton-Liger et al., 2014, 2011). Recently, overexpression of this gene was implicated in the brain ventriculomegaly (Box 1) observed in Ts1Cje mice and in DS-affected humans, and it is thought that the mechanism involves impaired cilia function in ependymal cells, which form the lining of the brain ventricular system (Raveau et al., 2017). Another gene that has been linked to DS is *Kcnj6*, which encodes potassium-voltage-gated channel subfamily J member 6. This gene has been shown to contribute to CHDs (Lignon et al., 2008) and to cognitive defects, together with another gene not yet identified (Jiang et al., 2015; Joshi et al., 2016; Cooper et al., 2012). The gene encoding cystathionine  $\beta$ -synthase (*Cbs*), which is involved in the methionine/cysteine cycle, is overexpressed in the DS brain (Ichinohe et al., 2005). Comparable overexpression of *Cbs* using a PAC transgenic line leads to changes in behaviour and LTP in mouse (Régnier et al., 2012), with similar phenotypes being observed in trisomic mice involving larger segments that include *Cbs* (Lopes Pereira et al., 2009; Yu et al., 2010c).

Several pathways that are perturbed in DS mouse models have been brought to light using transcriptomic and proteomic approaches. A meta-analysis of DS data, selected from human and mouse studies, unveiled perturbed neurological processes involved in neurodegeneration, axon guidance and nerve growth factor (NGF) signalling (Vilardell et al., 2011). A few Hsa21 genes (*SOD1*, *APP*, *DONSON*, *TIAM1*, *COL6A2*, *ITSN1* and *BACE2*) and the brain-derived neurotrophic factor (BDNF)-dependent pathway, involved in growth, differentiation and survival of neurons, were found to be altered. An elevated level of BDNF and of Akt-mTOR/Ras-ERK signalling was observed in the hippocampus of Ts1Cje

mice, and normal mTOR activity could be restored by treatment with the mTOR inhibitor rapamycin (Troca-Marín et al., 2014, 2011). In agreement, Ahmed et al. (2013) also showed that mTOR signalling is deregulated in Tc1 mouse brains.

There are likely to be multiple signalling pathways affected in the DS brain, however. In a recent systems biology study, the transcriptomes of cells taken from DS-affected human fetuses (and unaffected controls) were compared to transcriptome data from the embryonic forebrains of three mouse models (Dp1Yey, Ts65Dn and Ts1Cje) (Guedj et al., 2016). Their analyses revealed that a large panel of cellular processes (cellular stress response, DNA-repair signalling, regulation of cell cycle checkpoints, kinetochore organisation, proteolytic activity and anti-apoptotic genes) and molecular pathways (neurogenesis and neuronal differentiation, mitochondrial function, oxidative stress response, and inflammation) is dysregulated in DS mouse models and humans, indicating that the disease mechanisms are likely to be similar in both species. Another study, involving multi-regional transcriptome analysis of human DS and euploid foetal brains, pointed to misregulation of genes involved in the differentiation of oligodendrocytes and in myelination (Olmos-Serrano et al., 2016a). These findings were confirmed by analysis of Ts65Dn trisomy mice in the same study, highlighting that defects in white matter function could play a part in DS physiopathology. Lastly, epigenetic profiling has revealed multiple loci with altered CpG methylation in human DS and in mouse models, and this phenomenon may reflect increased dosage of Hsa21-linked methylation-pathway genes, such as *DNMT3L*, *SLC19A1* and others, as well as overexpression of key Hsa21-linked transcription factors, such as *RUNX1*, which could affect epigenetic patterns where they bind DNA (reviewed in Do et al., 2017). Collectively, these studies give an idea of the complexity of DS and emphasise the need to use an integrative approach that includes human samples and animal models, analysed at different periods in development (for example, foetal, early postnatal, young and late adult stages), to better understand the sequence of altered cellular processes and affected pathways in this disease.

### Therapeutic proof-of-concept for DS

In the last 10 years, a number of studies have sought to assess the efficacy of candidate therapeutic interventions for DS using mouse models. A summary of studies that have explored strategies for rectifying molecular, cellular or systemic defects in DS using mice is given in Table 2.

Some studies have attempted to target defects in neurogenesis and brain development in DS mouse models. The origin of the granule cell deficit in Ts65Dn has been traced to precursors in early postnatal development, which show a substantially reduced mitogenic response to hedgehog protein signaling (Roper et al., 2006; Baxter et al., 2000), a crucial pathway in development. Activation of the sonic hedgehog (SHH) pathway can be achieved by intraperitoneal injection of smoothened agonist (SAG) at postnatal day 2 (P2). Treated Ts65Dn pups have partially corrected cerebellar neurogenesis and hippocampal LTP defects, resulting in improved spatial memory (Das et al., 2013; Gutierrez-Castellanos et al., 2013; Roper et al., 2006). However, SAG treatment failed to restore normal cerebellar long-term depression and working memory. Moreover, a key limitation of systemic targeting of the SHH pathways is the potential for such treatment to increase the risk of several types of human cancer (Taipale and Beachy, 2001; Jiang and Hui, 2008). An alternative strategy has been proposed based on use of a  $\gamma$ -secretase inhibitor to reduce

overexpression of the SHH receptor PATCHED1 (PTCH1), which represses the SHH pathway. Overexpression of PTCH1 has been reported in Ts65Dn neural precursor cells (NPCs) (Trazzi et al., 2013, 2011) and is thought to result from the over-accumulation of the amyloid precursor protein intracellular domain (AICD), which is produced by cleavage of the APP precursor. The increase in AICD leads to overexpression of *PTCH1* in trisomic NPCs, impairing neurogenesis and neurite development (Trazzi et al., 2013, 2011). Using an inhibitor of APP  $\gamma$ -secretase, Giacomini et al. (2015) restored neuronal differentiation of Ts65Dn-derived NPCs and, with postnatal treatment, restored neurogenesis in the DG and the subventricular zone of Ts65Dn mice, while also normalising processing of APP. Thus, indirect targeting of the SHH pathway could form the basis of new therapeutic strategies to restore neurogenesis in trisomic brains.

An alternative is stem-cell-based therapy, a promising strategy for many diseases, including DS. Several studies have attempted to implant euploid neural stem cells into the brains of Ts65Dn mice and there is growing evidence that injected cells migrate to sites of damage where they provide neuroprotection. When NSCs were implanted in 12-month-old Ts65Dn mice, extrasomatic granules positive for expression of TAU and REELIN, associated with neuronal ageing, were reduced (Kern et al., 2011). When injected earlier, at P2, these cells induced a significant increase in the density of dentate granule cells and had a long-lasting positive effect on cognitive performance (learning) (Rachubinski et al., 2012a,b). Nevertheless, the stem-cell-based strategy has three limiting factors: (1) cells need to be injected directly into the brain for maximum efficacy; (2) the short-term effect is limited to the injection site and close vicinity; and (3) benefits are transitory and, when they persist, effects are probably linked to NSC-dependent neurotrophin production (Rachubinski et al., 2012b).

Among the other strategies that have been tested in DS mouse models is long-term peripheral administration of peptide 6 – an 11-mer corresponding to an active region of ciliary neurotrophic factor – which can enhance the pool of neural progenitor cells, and ameliorate learning and memory impairments in Ts65Dn mice (Blanchard et al., 2011). Three months of treatment with P7C3, an aminopropyl carbazole that enhances hippocampal neurogenesis, is sufficient to restore the neurogenic deficits observed in the Ts65Dn model (Latchney et al., 2015). Prenatal treatment with epigallocatechin gallate (EGCG), an inhibitor of DYRK1A, normalised some craniofacial phenotypes, including the increased cranial vault, in Ts65Dn mice (McElyea et al., 2016). Different protocols have demonstrated a beneficial impact of environmental enrichment – a widely used paradigm that increases sensory-motor stimulation – on learning, memory and motor activity in Ts65Dn mice (Llorens-Martín et al., 2010; Kida et al., 2013; Begenisic et al., 2015). The underlying molecular mechanisms responsible for this rescue are poorly understood, although it is thought that increased neurogenesis and synaptogenesis might be involved.

As highlighted above, the mTOR pathway has been implicated in DS. Treatment with the specific mTOR inhibitor rapamycin improved the spatial-memory performance of Ts1Cje mice and restored BDNF-dependent LTP in hippocampal slices (Andrade-Talavera et al., 2015). The same authors showed that deficits in synaptic plasticity (i.e. BDNF-LTP) and in the persistence of spatial memory were fully reversed using rapamycin in the Ts65Dn model (Andrade-Talavera et al., 2015), indicating that targeting mTOR hyperactivation may be a novel pharmacotherapeutic approach for DS. Consistent with this, administration of  $\alpha$ -Tocopherol (vitamin E), also known to act upon the mTOR pathway, led to

**Table 2. Candidate therapeutic approaches for DS**

Target; drug and dose	Mouse model	Treatment start/ duration/age at test	Main impact	Long term effect observed? (age)	Publication
<b>Neurogenesis, brain development</b>					
SHH and neurogenesis; SHH agonist (SAG-1.1; 20 mg/kg)	Ts65Dn	P0/1 injection/P6	Restored cerebellar neurogenesis		Roper et al., 2006
SHH and neurogenesis; SHH agonist (SAG-1.1; 20 mg/kg)	Ts65Dn	P0/1 injection/4 m	Improved spatial memory in MWM, hippocampal LTP, cerebellar LTD, neurogenesis and normalisation of cerebellar morphology but no effect in working memory	Yes (4 m)	Das et al., 2013
SHH and neurogenesis; SHH agonist (SAG-1.1; 20 mg/kg)	Ts65Dn	P0/1 injection/4 m	No impact on the deficient vestibulo-ocular reflex, a cerebellar function found defective in the Ts65Dn mouse model		Gutierrez-Castellanos et al., 2013
SHH and neurogenesis; $\gamma$ -secretase inhibitor (ELND006)	Ts65Dn	P3/12 d/P15	Restored neurogenesis and synapse development, reduction of expression of AICD and APP		Giacomini et al., 2015
Brain development; injections with neural progenitor cells	Ts65Dn	P2/acute/P2-P15 and 4 m	Migration of implanted neuronal progenitors and increased density of dentate granule cells but no change in the cognitive deficits observed in Ts65Dn mice during the neonatal period or adulthood (plus maze, MWM, conditioned task avoidance)		Rachubinski et al., 2012b
Neurotrophic factors; peptide 6 (CNTF)	Ts65Dn	11-15 m/1 m/12-16 m	Improved spatial memory (MWM), neurogenesis and connectivity		Blanchard et al., 2011
Neurogenesis; P7C3 (18 mg/kg)	Ts65Dn	1-2.5 m/3 m/4-6 m	Improved neurogenesis with increase in total Ki67+, DCX+ and surviving BrdU+ cells		Latchney et al., 2015
DYRK1A; EGCG (200 mg/kg)	Ts65Dn	E7/2 d/6 w	Normalisation of some craniofacial phenotypes, and of <i>Ptch1</i> , <i>Ets2</i> , <i>Rcan1</i> and <i>Shh</i> expression		McElyea et al., 2016
Voluntary exercise	Ts65Dn	10-12 m/7 w/12-14 m	Improved performance in MWM, increased in DCX+ and phospho-histone-3+ hippocampal neurons		Llorens-Martin et al., 2010
Voluntary exercise/running	Ts65Dn	21 d/9 m/10 m	Improved performance in the rotarod and NOR tests, and reduced overexpression of <i>DYRK1A</i> , <i>APP</i> , <i>MAP2ab</i> and <i>SOD1</i>		Kida et al., 2013
Voluntary exercise/running	Ts65Dn	6 m/7 m/13 m	Only improved performance in the rotarod not in NOR		Kida et al., 2013
Neurogenesis, brain function; EET	Ts65Dn	Gestation/2 m/2 m	Improvement in maternal care and normalisation of cognition in NOR and of hippocampal LTP in adults treated during gestation		Begenisic et al., 2015
<b>Oxidative stress, development</b>					
Astrocytes, brain development, oxidative stress; NAP+SAL (20 $\mu$ g; IP)	Ts65Dn	E8/4 d/P5-P20	Better motor and sensory milestones	Yes (8-10 m)	Begenisic et al., 2015
Astrocytes, brain development, oxidative stress; NAP+SAL (20 $\mu$ g; IP)	Ts65Dn	10 m/9 d/10 m	Improved MWM learning performance	Yes (8-10 m)	Incerti et al., 2011
Astrocytes, brain development, oxidative stress; NAP+SAL (20 $\mu$ g; IP)	Ts65Dn	E8/4 d/8 m	Decreased latency to find the platform in the MWM but no probe test shown		Incerti et al., 2012
ROS; vitamin E (tocopherol; 400 mg/kg in food)	Ts65Dn	4 m/6 m/10 m	Improved working memory in the RAWM with reduced production of cortical ROS, reduced degeneration of cholinergic neurons (TrkA+) in the medial septal nucleus with no change in brain APP level, and reduced loss of calbindin hippocampal CA1 neurons in 10-m-old mice		Lockrow et al., 2009
ROS; vitamin E ( $\alpha$ -tocopherol; 160 mg/kg in food)	Ts65Dn	Gestation/3 m/3 m	Reduced anxiety in the elevated plus maze, improved latency to find the platform with rescue in the probe test of the MWM, no impact on hyperactivity during circadian activity, variable normalisation of several lipid peroxidation products in plasma, cortex and hippocampus		Shichiri et al., 2011

Continued

Table 2. Continued

Target; drug and dose	Mouse model	Treatment start/ duration/age at test	Main impact	Long term effect observed? (age)	Publication
<b>Neurotransmission</b>					
Endocannabinoid signalling; inhibitor of monoacylglycerol lipase	Ts65Dn	11 m/4 w/12 m	Reduced ambulatory activity (but no change in the thigmotaxic behaviour) in the open field and the Y-maze, no improvement in spontaneous alternation (Y-maze) or NOR with 10-min retention time but improvement in the NOR with 24 h retention, normalisation of LTP in the CA1, changes in the level of brain endocannabinoids and their metabolites, reduced levels of A $\beta$ 40 and A $\beta$ 42 (APP-derived peptides)		Lysenko et al., 2014
GABA-A receptor; PTZ (3 mg/kg in food)	Ts65Dn	3-4 m/17 d/4-5 m	Rescued performance in the NOR (24-h retention time) and spontaneous alternation in the T-maze, improved hippocampal LTP (DG)		Fernandez et al., 2007
GABA-A receptor; PTZ (3 mg/kg in food)	Ts65Dn	4 m/7 w/5-6 m	No effect on hyperactivity during circadian monitoring and on sensorimotor tests, including rotarod; normalisation of learning performance in the MWM		Rueda et al., 2008a
GABA-A $\alpha$ 5; $\alpha$ 5IA (5 mg/kg; IP)	Ts65Dn	3 m/1 d-2 w/3-4 m	Improvement in NOR (10-min retention time) and in the learning phase of the MWM, no change in the locomotor activity (open field) or the anxiety level (open field and elevated plus maze), enhancement of the early gene <i>c-fos</i> products in brain region		Braudeau et al., 2011b
GABA synthesis; gabapentin (10 mg/kg; IP)	Ts65Dn	3 m/10 w/4-6 m	No effect on sensorimotor abilities, motor coordination or spontaneous activity, on anxiety (open field and elevated plus maze), in the MWM and in the FC test		Vidal et al., 2012
GABA-A receptor; PTZ (0.03 mg/kg; IP)	Ts65Dn	2-3 m, 12-15 m/2-3 w/ 5-12 m, 5-15.5 m	Improvement in NOR (24-h retention time) 8 days after 2-w treatment		Colas et al., 2013
GABA-A receptor; PTX	Ts65Dn	3-4 m/acute/3-4 m	Restoration of the LTP in the DG		Kleschevnikov et al., 2004
GABA-A receptor; PTX	Ts65Dn	4-6 m/acute/4-6 m	Restoration of the LTP-induced theta-burst stimulation in the CA1		Costa and Grybko, 2005
GABA-B; CGP55845 (0.5 mg/kg; IP)	Ts65Dn	2-3 m/3 w/3-4 m	Memory restored in the novel-place recognition, NOR and FC tasks, but no effect on locomotion and performance in T-maze; increased production of BDNF and restoration of the LTP in the DG		Kleschevnikov et al., 2012a
GABA-A $\alpha$ 5; RO4938581 (20 mg/kg in food)	Ts65Dn	3-4 m/6 w/4-6 m	Improved spatial learning and memory (MWM), hippocampal synaptic plasticity (LTP) and adult neurogenesis		Martinez-Cue et al., 2013
NMDA receptor; memantine (5 mg/kg; IP)	Ts65Dn	4-6 m/acute/4-6 m	Analysis of proteomic changes after rescue in the FC test show increased NR2A and NR1 in the hippocampus and cortex		Costa et al., 2008; Ahmed et al., 2015
NMDA receptor; memantine (30 mg/kg; IP)	Ts65Dn	9 m/8-9 w/11 m	Improved spatial learning in MWM with reduced brain A $\beta$ level and increased brain VGLUT1 level		Rueda et al., 2010
NMDA receptor; memantine (20 mg/kg in food)	Ts65Dn	4/10 m	Memory improvement in the RAWM and the NOR (24-h retention time) but no effect on hyper-exploration activity with increased <i>BDNF</i> gene expression in the hippocampus and BDNF protein in the frontal cortex		Lockrow et al., 2011
NMDA receptor; R025-6981	Ts65Dn	3-6 m/acute or 2 w/3- 6 m or 3.5-6.5 m	Reduced synaptic NMDA-receptor responses in CA1 and $\gamma$ -oscillation deficits, with restoration of LTP in CA1 but no effect on spontaneous alternation (Y-maze) or in the Barnes maze		Hanson et al., 2013
mTOR; rapamycin (10 mg/kg)	Ts1Cje	2 m/5 d/2 m	Restoration of BDNF-induced LTP at CA3-CA1 hippocampal synapses, and increase spatial learning and memory in the Barnes maze		Andrade-Talavera et al., 2015

Continued

Table 2. Continued

Target; drug and dose	Mouse model	Treatment start/ duration/age at test	Main impact	Long term effect observed? (age)	Publication
Neurogenesis, 5-HT reuptake; fluoxetine (5-10 mg/kg; IP)	Ts65Dn	P3/13 d/P45	Increased thickness of the mossy fibres and increased synaptophysin and VGLUT1 reactivity in the CA3 with improvement of the spine density and better basal synaptic plasticity in CA3 neurons with higher BDNF level	Yes (1 m)	Stagni et al., 2013
Neurogenesis, 5-HT reuptake; fluoxetine	Ts65Dn	E10/10 d/1.5 m	Restoration of neuronal precursor proliferation and cellularity in the forebrain with normal dendritic development, cortical and hippocampal synapse development and brain volume associated with normalisation of activity in the open field, and in the FC test but no effect on NOR (4-h or 24-h retention time)	Yes (1.5 m)	Guidi et al., 2014
Neurogenesis, 5-HT reuptake; fluoxetine (5-10 mg/kg; IP)	Ts65Dn	P3/13 d/3 m	Maintenance of the cellular restoration observed at P3 and P10 and improved hippocampal function in the MWM, NOR (1-h retention time) and passive avoidance tests	Yes (3 m)	Stagni et al., 2015
Neurogenesis, 5-HT reuptake; fluoxetine (5-10 mg/kg; IP)	Ts65Dn	2-5 m/24 d/2.5-5.5 m	Increased neurogenesis and neuronal survival in the hippocampus		Clark et al., 2006
Neurogenesis, 5-HT reuptake; fluoxetine (5-10 mg/kg; IP)	Ts65dn	P3-P15/2 h or 1 m	Restored proliferation in the hippocampus with normal expression of R-HT1A receptors and BDNF associated with a complete recovery in the FC test		Bianchi et al., 2010
Neurogenesis, 5-HT reuptake; fluoxetine (0.2 mg/ml water)	Ts65Dn	4-6 m/4 w/5-7 m	Reduced exploratory activity in the open field but no effect in the MWM and increased adverse effects (seizure and mortality)		Heinen et al., 2012
Neurogenesis, 5-HT reuptake; fluoxetine (10 mg/kg in water)	Ts65Dn	2 m/8 w/4 m	Recovery of spatial memory in the novel-place recognition and spontaneous alternation (Y-maze) tests, and of hippocampal LTP in CA1 with normalisation of GABA release in the hippocampus		Begenisic et al., 2014
Neurogenesis; lithium	Ts65Dn	5-6 m/4 w/6-7 m	Restoration of the number of newborn neurons, the adult neurogenesis and the LTP in DG, associated with improved FC and novel-place recognition (24 h) but no change in the spontaneous alternation (T-maze)		Contestabile et al., 2013
Glutamatergic and cholinergic pathways; piracetam (75, 150 or 300 mg/kg; IP)	Ts65Dn	1.5-2 m/4 w/2.1-3 m	Improved performance in the MWM (75-150 mg/kg) and only the 300 mg/kg reduced the nocturnal spontaneous activity		Moran et al., 2002
Glutamatergic and cholinergic pathways; piracetam, SGS- 111	Ts65Dn	4-6 m/6 w/5.5-7.5 m	Reduced hyperactivity (home-cage, open-field and hole board tests) but no improvement in spatial learning in the MWM or in the passive avoidance test		Rueda et al., 2008b
Glutamatergic and cholinergic pathways; piracetam, SGS- 111	Ts65Dn	gestation/5 m/5 m	No effect on hyperactivity or spatial memory in the MWM or in the passive avoidance test		Rueda et al., 2008b
NE, locus coeruleus neurons; DREADDs	Ts65Dn	11-14 m/acute/ 11-14 m	Improved NOR, normalised spontaneous activity in the four-arm maze		Fortress et al., 2015
NE; formoterol (2 mg/kg) +nadolol (4 mg/kg) (IP)	Ts65Dn	5-6 m/15 d/5.5-6.5 m	Reversal of FC failure with restored synaptic density and neuronal activity ( <i>c-fos</i> after stimulation), and increased cell proliferation and dendritic arborisation		Dang et al., 2014
NE; L-DOPS	Ts65Dn	11 m/2 w/12 m	Improved NOR, normalised spontaneous activity in the four-arm maze with reduced $\beta$ 1-adrenergic-receptor staining in the hippocampus		Fortress et al., 2015

Continued

Table 2. Continued

Target; drug and dose	Mouse model	Treatment start/ duration/age at test	Main impact	Long term effect observed? (age)	Publication
NE; L-DOPS, xamoterol	Ts65Dn	3-5 m/acute/3-5 m	Recovery of contextual fear learning and nesting behaviour, with increased NE hippocampal concentration. Xamoterol, an $\beta$ 1-adrenergic-receptor agonist, failed to rescue contextual learning, suggesting a $\beta$ 1-adrenergic-receptor-dependent mechanism in the L-DOPS rescue		Salehi et al., 2009
Norepinephrine; xamoterol, (1-10 mg/kg; IP)	Ts65Dn	9-12 m/acute/9-12 m	Restoration of the memory deficit in FC, spontaneous alternation (Y-maze and T-maze) and NOR (24-h retention time) but no effect on ambulatory activity (open field, PhenoTyper)		Faizi et al., 2011
Cholinergic pathway; donepezil (3.3 mg/kg in food)	Ts65Dn	4 m/7 w/5-6 m	No modification of learning and memory in the MWM		Rueda et al., 2008a
Cholinergic pathway; physostigmine (50 $\mu$ g/kg)	Ts65Dn	4, 10, 16 m/acute/ 4, 10, 16 m	Reversal of spontaneous alternation in the four-arm maze only at 4 months of age		Chang and Gold, 2008
Cholinergic pathway; galantamine	Ts65Dn	3-6 m/acute/3-6 m	Better performance in olfactory learning		de Souza et al., 2011
Cholinergic pathway; choline (1.1 and 5.0 g/kg in the diet)	Ts65Dn	Gestation/13 m	Improved spatial recognition in RAWM and neurogenesis	Yes (13 m)	Velazquez et al., 2013
Glutamatergic pathway; melatonin (20 mg/kg)	Ts65Dn	5 m/5 m/10 m	Recovered spatial learning (latency) and memory in the MWM and increased the number of ChAT+ cells in the medial septum but no effect on APP level, on sensorimotor tests, in rotarod, on anxiety (elevated plus maze and open field) or on spontaneous locomotor activity. Increased exploratory activity (hole board and open field). Rescued adult neurogenesis, density of hippocampal granule cells and hippocampal cortical 4-hydroxynonenal protein adducts, and LTP (CA1). Increased VGLUT1 signal in the hippocampal DG		Corrales et al., 2013; Corrales et al., 2014
<b>Individual genes</b>					
KCNJ6; ETH (150 mg/kg; IP)	Ts65Dn	3 m/10 w/4-6 m	No effect on sensorimotor abilities, motor coordination or spontaneous activity, on anxiety (open field and elevated plus maze), in the MWM and/or in the FC test		Vidal et al., 2012
DYRK1a; shRNA, AAV (IC – striatum)	Tg(MT1A-Dyrk1a)	2-3 m/4-5 m	Attenuation of hyperactivity (open field) and motor alterations (treadmill)		Ortiz-Abalia et al., 2008
DYRK1a; shRNA, AAV (IC – hippocampus)	Ts65Dn	2 m/1 injection/3 m	Normalised ratio of pERK1:ERK1 and pCREB:CREB, improvement of hippocampal CA1 LTP, no rescue of MWM but reduced thigmotaxis		Altafaj et al., 2013
DYRK1a; EGCG (10 $\mu$ M)	Ts65Dn	2-5 m/acute/2-5 m	Normalisation of hippocampal CA3-CA1 LTP		Xie et al., 2008
DYRK1a; EGCG-GT (30-60 mg/kg in food)	Tg(CEPHY152F7) 12Hgc	Gestation-4.5 m/ 4.5 m	Rescued NOR with effect on brain volume and normalised level of BDNF and TrkB		Guedj et al., 2009
DYRK1a; POL60-EGCG (60 mg/kg in food)	Tg(MT1A-Dyrk1a), Ts65Dn	4.5 m/1 m/5.5 m	Restoration of spatial memory in the MWM and NOR, with normalisation of homocysteine plasma level		De la Torre et al., 2014
DYRK1a; MGTE-EGCG (60 mg/kg in food)	Tg(Dyrk1a) 189N3YahJmd	4 m/1 m/5 m	Rescued prefrontal LTP (but not long-term depression) and normalisation of pCAMK2:CAMK2 ratio in the prefrontal cortex		Thomazeau et al., 2014
DYRK1a; EGCG	Ts65Dn	P3/13 d/P15-P45	Restored neurogenesis in the subventricular zone and the DG, normalisation of total hippocampal granule cells and level of pre- and post-synaptic proteins (SYN and PSD95), but no long-term effect at P45		Stagni et al., 2016
DYRK1a; POL60-EGCG (60 mg/kg in food)	Tg(Dyrk1a) 189N3YahJmd, Ts65Dn	4.5 m/1 m/5.5 m	Normalisation of the component level of the GABAergic and glutamatergic pathway in cortex and hippocampus but not cerebellum, and rescue of the spontaneous alternation in the Y-maze		Souchet et al., 2015

Continued

Table 2. Continued

Target; drug and dose	Mouse model	Treatment start/ duration/age at test	Main impact	Long term effect observed? (age)	Publication
<b>Neurodegeneration</b>					
A $\beta$ ; $\gamma$ -secretase inhibitor (DAPT; 100 mg/kg; SC)	Ts65Dn (female)	4 m/acute/4 m	Improved spatial learning and memory in the MWM with increased brain APP-CTF and lower A $\beta$ levels		Netzer et al., 2010
A $\beta$ ; anti-A $\beta$ vaccine	Ts65Dn	5 m/1 injection/8 m	Reduction of brain A $\beta$ level and restoration of NOR and FC, with reduction of cholinergic neuron atrophy		Belichenko et al., 2016
A $\beta$ ; EET	Ts65Dn	2.5 m/9.5 m/12 m	Normalisation of hippocampal A $\beta$ oligomer and improved spatial learning and memory (MWM) and spontaneous alternation (Y-maze)		Sansevero et al., 2016
Cholinergic pathway; choline-supplemented diet	Ts65Dn	Pregnancy-weaning/ 40 d/14-18 m	Improved in the RAWM and increased hippocampal ChAT innervation		Kelley et al., 2016
ChAT+ neurons; genetic, <i>Bace1</i> <sup>+/-</sup>	Ts2	4 m/9 m/16 m	Prevention of endosomal pathology in the medial septal nucleus and the cerebral cortex, normalised the number of ChAT+ neurons		Jiang et al., 2016
BFCNs, neurodegeneration; estrogen (class B)	Ts65Dn (female)	11-15 m/2 m/13-17 m	Improved learning in the T-maze escape task (plus reversal) with increased number of ChAT+ neurons in the medial septal nucleus and normalisation of NGF level in the basal forebrain		Granhholm et al., 2002
BFCNs; estrogen (class B)	Ts65Dn (female)	9-15 m/2 m/11-17 m	Normalisation of MAP2 and AChE staining in the hippocampus		Granhholm et al., 2003
BFCNs; estrogen (class B)	Ts65Dn (male)	5 m/12 d/6 m	No improvement of cognitive ability in the RAWM		Hunter et al., 2004b
Inflammation; minocycline	Ts65Dn	7 m/3 m/10 m	Prevention of microglial activation, progressive BFCN decline and improved performance in the RAWM		Hunter et al., 2004a
BFCNs; choline (25 mM in the diet)	Ts65Dn	Pregnancy-lactation/ 40 d/6 m	Improved attention tasks	Yes (6 m)	Moon et al., 2010
Cholinergic pathway; choline (1.1 or 5 g/kg in the diet)		Gestation/13-17 m	Improved spatial mapping in the RAWM, and increased number, density and size of medial septum BFCNs	Yes (13 m)	Ash et al., 2014

5-HT, serotonin; A $\beta$ ,  $\beta$ -amyloid; AAV, adeno-associated virus; AICD, amyloid precursor protein intracellular domain; BDNF, brain-derived neurotrophic factor; BFCNs, basal forebrain cholinergic neurons; ChAT, choline acetyltransferase; CNTF, ciliary neurotrophic factor; d, day; DCX, doublecortin; DG, dentate gyrus; DREADDs, designer receptors exclusively activated by designer drugs; E, embryonic day; EET, enriched environment treatment; EGCG, epigallocatechin gallate; ETH, ethosuximide; FC, contextual and auditory-cue-conditioned fear task (fear conditioning); GT, green tea extract; IC, internal capsule; IP, intraperitoneal injection; L-DOPS, droxidopa; LTP, long-term potentiation; m, month; MGTE, modified green tea extract; MWM, Morris water maze; NAP, peptide D-NAPVSIPQ; NE, norepinephrine; NGF, nerve growth factor; NOR, novel-object recognition; P, postnatal day; POL60, polyphenon 60; PTZ, pentylenetetrazol; RAWM, radial-arm water maze; ROS, reactive oxygen species; SAG, smoothed agonist; SAL, peptide D-SALLRSIPA; SC, subcutaneous; w, week.

attenuation of cognitive impairments in the Ts65Dn model (Lockrow et al., 2009; Shichiri et al., 2011).

Different neurotransmission pathways have been shown to be altered in mouse models of DS, and these pathways have been targeted in attempts to improve cognition. In one example, Ts65Dn mice were treated with JZL 184, a selective inhibitor of monoacylglycerol lipase. The lipase degrades the most abundant endocannabinoid and its inactivation improves synaptic plasticity and memory in mouse (Pan et al., 2011). Thus, hippocampal LTP, long-term memory and locomotor activity were restored to a normal level in JZL-184-treated Ts65Dn mice; however, positive thigmotaxis (Box 1) and short-term memory defects remained unchanged (Lysenko et al., 2014).

Ts65Dn mice show perturbations of the excitatory/inhibitory balance towards an excess of GABA transmission; this imbalance may also explain cognitive dysfunction in human DS. The ability of antagonists of GABA receptors [including pentylenetetrazol (PTZ), RO4938581 and CGP55845] to normalise this balance has been tested in mouse models. These compounds rescued long-term and

spatial memory, hippocampal LTP and the expression of several key molecular markers, including BDNF, GAD65/67 and VGAT (Fernandez et al., 2007; Rueda et al., 2008a; Braudeau et al., 2011a,b; Colas et al., 2013; Vidal et al., 2012; Kleschevnikov et al., 2004, 2012b; Martinez-Cue et al., 2013; Costa and Grybko, 2005). Similarly, the N-methyl-D-aspartate receptor antagonists memantine and MK-801 successfully rescued learning and improved memory, partly by normalising the levels of APP, vGlut1 and BDNF (Costa et al., 2008; Ahmed et al., 2015; Rueda et al., 2010; Lockrow et al., 2011; Hanson et al., 2013). A memantine clinical study is currently underway to assess the effects of this treatment on adolescents with DS (<https://www.clinicaltrials.gov/>).

Prenatal or early postnatal treatment with fluoxetine, which targets the serotonergic system, rescued neurogenesis, long-term memory and synaptic plasticity in Ts65Dn mice (Guidi et al., 2014; Stagni et al., 2015). Adult treatment enhanced neurogenesis without improving learning/memory deficits at a high dose of fluoxetine (Heinen et al., 2012); however, an improvement in short-term memory was observed using a low dose (Begenisic



et al., 2014). As seen with fluoxetine treatment, early administration of lithium, another antidepressant drug acting also upon the serotonin pathway, fully rescued short- and long-term memory, and LTP as well as neurogenesis in Ts65Dn mice (Contestabile et al., 2013).

Therapeutic targeting of key candidate genes implicated in DS has also been attempted. Partial display of DS phenotypes in mice harbouring a single trisomy of *Kcnj6* provides compelling evidence for a functional role of increased channel expression in some of the abnormal neurological phenotypes found in DS. However, treatment of Ts65Dn mice with ethosuximide (ETH), an inhibitor of KCNJ6, failed to rescue impairments of motor coordination or cognitive performance (MWM or FC) (Vidal et al., 2012). Given its putative role in DS, researchers have also explored the potential of *DYRK1A* as a therapeutic target. Studies involving inhibition of DYRK1A enzymatic activity using a safe and naturally available inhibitor (EGCG) demonstrated that learning/memory phenotypes could be rescued in different models, including in Ts65Dn mice (Xie et al., 2008; Guedj et al., 2009; De la Torre et al., 2014; Thomazeau et al., 2014), despite the fact that other genes are trisomic in this model.

As mentioned above, individuals with DS are at increased risk of developing AD during ageing, and the presence of three copies of APP is known to contribute to this phenomena. Thus, several therapies targeting the APP pathways have been tested. For example, the  $\gamma$ -secretase inhibitor DAPT was shown to improve memory and prevented neurodegeneration in Ts65Dn mice (Netzer et al., 2010). Anti-A $\beta$  vaccine failed to rescue a normal level of A $\beta$ 40 and 42 but improved spatial learning of trisomic mice (Belichenko et al., 2016), suggesting that there is no direct relationship between the levels of A $\beta$  and defects in spatial learning. In wild-type rodents, supplementing the maternal diet with additional choline (~4.5 $\times$  the amount in normal chow) enhances spatial memory and attention in the offspring, and exerts structural and functional changes in the septo-hippocampal cholinergic system (McCann et al., 2006); these results suggested that maternal choline supplementation (MCS) could enhance cognitive function and protect against BFCN degeneration. MCS given to Ts65Dn mice significantly improved spatial mapping and increased the number, density and size of medial septum BFCNs (Moon et al., 2010; Ash et al., 2014). MCS also seems to protect against hippocampal cholinergic projection system degeneration (Kelley et al., 2016).

Long-term exposure to environmental enrichment reduces A $\beta$  oligomers and rescues spatial-memory abilities in 12-month-old trisomic mice (Sansevero et al., 2016). Estrogen treatment partially rescued working memory (T-maze test) and prevented neurodegeneration in aged Ts65Dn animals (11- to 17-months old) (Granholm et al., 2002, 2003). Consistent with a role for inflammatory processes in BFCN degeneration, minocycline treatment inhibits microglial activation, prevents progressive BFCN decline and markedly improves performance of Ts65Dn mice on a working and reference memory task (Hunter et al., 2004a). Partially reducing  $\beta$ -secretase 1 (BACE1) by deleting one *BACE1* allele blocked development of age-related endosome enlargement in the medial septal nucleus, cerebral cortex and hippocampus, and prevented loss of choline acetyltransferase (ChAT)-positive medial septal nucleus neurons (Jiang et al., 2016).

These potential therapeutic approaches have been tested ‘preclinically’ in DS mouse models (mostly in Ts65Dn mice) across different laboratories. Many different compounds and manipulations have produced beneficial effects. However, apart

from EGCG and GABA-targeting compounds, the efficacy of specific strategies have not been validated in an independent study. Testing for interlaboratory reproducibility of the results, and reproducibility in more than one mouse model, will be crucial next steps in this exciting research area.

#### Future directions for DS research

As highlighted here, DS mouse models are providing important insight into this complex disorder. Chromosome engineering technologies now enable us to address more refined questions, including, for example, on synergy and epistatic interaction between Hsa21 genes. The panel of new technologies provides the opportunity to broaden the range of animal models to include the rat or primates, although it must be emphasised that such models should only be developed where justified by the scientific question and there is clear added value compared to available models. A key limitation of DS mouse models is that most of the phenotypes have been described in a few genetic backgrounds, mostly pure inbred C57BL/6J mice, and for a few lines only in the F1, potentially reducing complex genetic interactions and their influence on the penetrance and expressivity of the phenotypes. It is widely known that the genetic background can impact on phenotypic outcome in mice; for example, in some backgrounds, homozygous knockout of epidermal growth factor receptor (EGFR) is lethal, whereas, in other backgrounds, the mutant is viable (Threadgill et al., 1995; Sanford et al., 2001). An impact on phenotype is also observed when modelling intellectual disabilities, autism spectrum disorders or psychiatric diseases (Sittig et al., 2016; Arbogast et al., 2016). Thus, addressing the interindividual phenotypic variability in DS will require strategies either to develop models in various genetic mixed backgrounds, e.g. with the F1 modelling approach proposed by Sittig et al. (2016), or to bring new genetic diversity using, for example, the Collaborative Cross lines (<http://csbio.unc.edu/CCstatus/index.py>) or using large cohorts derived in an outbred genetic background. In parallel, the use of large cohorts of ethnically and genetically diverse DS individuals will help to capture the variability in the human population.

The complex nature of DS creates the need for collaborative, multidisciplinary research to understand its biology. One key step was the formation of the T21 Research Society (T21RS), a global initiative that provides freely available, curated and therefore up-to-date web resources, including validated models and protocols for behavioural and pharmacological treatment. The T21RS society is unique and aims to develop and coordinate action to generate new models to make them available and to propose standard operating protocols for behavioural investigation of DS. Moreover, mouse-based research into DS has resulted in the advancement of a few different treatments to human clinical trials. Promising results have been obtained in terms of improvement of cognitive function and the autonomy of individuals with DS (de la Torre et al., 2016, 2014), but more work is needed to better assess drug efficacy, the consequence of long-term treatment and the impact of treatment timing. In addition, most of the treatments were evaluated in a Ts65Dn mouse model, which only recapitulates the trisomy partially, lacking about half of the mouse genes that are homologous to those on Hsa21. The use of more complex mouse models, such as Dp1Yey and Dp1Tyb or those that combine trisomies for Mmu16, Mmu17 and Mmu10 homologous regions (Yu et al., 2010b), should be developed for reliable therapeutic assessment.

Models developed in other species, such as rat, are likely to be helpful to validate genotype-phenotype relationships and the outcome of preclinical treatments. Furthermore, we may have

access to new avenues for therapeutic intervention. Firstly, we could adapt a treatment to the age of DS individuals, so aiming to counteract neurodevelopmental defects as early as possible, and decreasing the risk of comorbidities in the adult and during ageing. Before devising strategies for prenatal treatment, it is important to understand in depth the development of disomic versus trisomic human foetuses. New tools will also certainly contribute to new strategies. Notably, the use of CRISPR/Cas9 or CRISMERE tools could lead to therapeutic strategies for reducing the overdosage of a given gene or a genomic region in trisomic individuals by inactivating one of the three alleles, for example, in transplantable stem cells or induced pluripotent stem cells. Finally, DS models will continue to be improved in the next few years. As noted above, most drug candidates to date have been tested in the Ts65Dn model, which carries only ~90 of the Hsa21 homologous genes and includes ~60 additional genes that are not orthologous to Hsa21. Thus, we must consider more complete models to evaluate therapeutic options even though these may be difficult to breed at present (Belichenko et al., 2015). An intriguing alternative is provided by the transchromosomal strategy that brings a complete set of Hsa21 genes into the mouse genome. However, this approach has some pitfalls: (1) human proteins or non-coding RNAs may not be able to interact with the same efficiency and efficacy with their mouse partners; (2) the regulation of human sequences may differ from the mouse genes, leading to unexpected domains of expression; and (3) the human allelic repertoire will be fixed in each strain. Despite the limitations, the progress to date has been promising and there is little doubt that mouse-based DS research will be important for translation into therapies for DS in the future.

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#### References

Ahmed, M. M., Dhanasekaran, A. R., Tong, S., Wiseman, F. K., Fisher, E. M. C., Tybulewicz, V. L. J. and Gardiner, K. J. (2013). Protein profiles in Tc1 mice implicate novel pathway perturbations in the Down syndrome brain. *Hum. Mol. Genet.* **22**, 1709-1724.

Ahmed, M. M., Dhanasekaran, A. R., Block, A., Tong, S., Costa, A. C. S., Stasko, M. and Gardiner, K. J. (2015). Protein dynamics associated with failed and

rescued learning in the Ts65Dn mouse model of Down syndrome. *PLoS ONE* **10**, e0119491.

Ahn, K.-J., Jeong, H. K., Choi, H.-S., Ryoo, S.-R., Kim, Y. J., Goo, J.-S., Choi, S.-Y., Han, J.-S., Ha, I. and Song, W.-J. (2006). DYRK1A BAC transgenic mice show altered synaptic plasticity with learning and memory defects. *Neurobiol. Dis.* **22**, 463-472.

Alford, K. A., Slender, A., Vanes, L., Li, Z., Fisher, E. M. C., Nizetic, D., Orkin, S. H., Roberts, I. and Tybulewicz, V. L. J. (2010). Perturbed hematopoiesis in the Tc1 mouse model of Down syndrome. *Blood* **115**, 2928-2937.

Altajaf, X., Dierssen, M., Baamonde, C., Marti, E., Visa, J., Guimera, J., Oset, M., Gonzalez, J. R., Florez, J., Fillat, C. et al. (2001). Neurodevelopmental delay, motor abnormalities and cognitive deficits in transgenic mice overexpressing Dyk1A (minibrain), a murine model of Down's syndrome. *Hum. Mol. Genet.* **10**, 1915-1923.

Altajaf, X., Martin, E. D., Ortiz-Abalia, J., Valderrama, A., Lao-Peregrin, C., Dierssen, M. and Fillat, C. (2013). Normalization of Dyk1A expression by AAV2/1-shDyk1A attenuates hippocampal-dependent defects in the Ts65Dn mouse model of Down syndrome. *Neurobiol. Dis.* **52**, 117-127.

Andrade-Talavera, Y., Benito, I., Casañas, J. J., Rodríguez-Moreno, A. and Montesinos, M. L. (2015). Rapamycin restores BDNF-LTP and the persistence of long-term memory in a model of Down's syndrome. *Neurobiol. Dis.* **82**, 516-525.

Antonarakis, S. E. (2016). Down syndrome and the complexity of genome dosage imbalance. *Nat. Rev. Genet.* **18**, 147-163.

Antonarakis, S. E., Lyle, R., Dermitzakis, E. T., Reymond, A. and Deutsch, S. (2004). Chromosome 21 and down syndrome: from genomics to pathophysiology. *Nat. Rev. Genet.* **5**, 725-738.

Arbogast, T., Raveau, M., Chevalier, C., Nalesso, V., Dembele, D., Jacobs, H., Wendling, O., Roux, M., Duchon, A. and Héault, Y. (2015). Deletion of the App-Runx1 region in mice models human partial monosomy 21. *Dis. Model. Mech.* **8**, 623-634.

Arbogast, T., Ouagazzal, A.-M., Chevalier, C., Kopanitsa, M., Afinowi, N., Migliavacca, E., Cowling, B. S., Birling, M.-C., Champy, M.-F., Reymond, A. et al. (2016). Reciprocal effects on neurocognitive and metabolic phenotypes in mouse models of 16p11.2 deletion and duplication syndromes. *PLoS Genet.* **12**, e1005709.

Ash, J. A., Velazquez, R., Kelley, C. M., Powers, B. E., Ginsberg, S. D., Mufson, E. J. and Strupp, B. J. (2014). Maternal choline supplementation improves spatial mapping and increases basal forebrain cholinergic neuron number and size in aged Ts65Dn mice. *Neurobiol. Dis.* **70**, 32-42.

Ballard, C., Mobley, W., Hardy, J., Williams, G. and Corbett, A. (2016). Dementia in Down's syndrome. *Lancet Neurol.* **15**, 622-636.

Baxter, L. L., Moran, T. H., Richtsmeier, J. T., Troncoso, J. and Reeves, R. H. (2000). Discovery and genetic localization of Down syndrome cerebellar phenotypes using the Ts65Dn mouse. *Hum. Mol. Genet.* **9**, 195-202.

Beach, R. R., Ricci-Tam, C., Brennan, C. M., Moomau, C. A., Hsu, P.-H., Hua, B., Silberman, R. E., Springer, M. and Amon, A. (2017). Aneuploidy causes non-genetic individuality. *Cell* **169**, 229-242.e21.

Begenisic, T., Baroncelli, L., Sansevero, G., Milanese, M., Bonifacino, T., Bonanno, G., Cioni, G., Maffei, L. and Sale, A. (2014). Fluoxetine in adulthood normalizes GABA release and rescues hippocampal synaptic plasticity and spatial memory in a mouse model of Down syndrome. *Neurobiol. Dis.* **63**, 12-19.

Begenisic, T., Sansevero, G., Baroncelli, L., Cioni, G. and Sale, A. (2015). Early environmental therapy rescues brain development in a mouse model of Down syndrome. *Neurobiol. Dis.* **82**, 409-419.

Belichenko, N. P., Belichenko, P. V., Kleschevnikov, A. M., Salehi, A., Reeves, R. H. and Mobley, W. C. (2009). The "Down syndrome critical region" is sufficient in the mouse model to confer behavioral, neurophysiological, and synaptic phenotypes characteristic of Down syndrome. *J. Neurosci.* **29**, 5938-5948.

Belichenko, P. V., Kleschevnikov, A. M., Becker, A., Wagner, G. E., Lysenko, L. V., Yu, Y. E. and Mobley, W. C. (2015). Down syndrome cognitive phenotypes modeled in mice trisomic for All HSA 21 homologues. *PLoS ONE* **10**, e0134861.

Belichenko, P. V., Madani, R., Rey-Bellet, L., Pihlgren, M., Becker, A., Plassard, A., Vuillermot, S., Giriens, V., Nosheny, R. L., Kleschevnikov, A. M. et al. (2016). An anti-beta-amyloid vaccine for treating cognitive deficits in a mouse model of Down syndrome. *PLoS ONE* **11**, e0152471.

Besson, V., Brault, V., Duchon, A., Togbe, D., Bizot, J.-C., Quesniaux, V. F. J., Ryffel, B. and Héault, Y. (2007). Modeling the monosomy for the telomeric part of human chromosome 21 reveals haploinsufficient genes modulating the inflammatory and airway responses. *Hum. Mol. Genet.* **16**, 2040-2052.

Bhutta, M. F., Cheeseman, M. T., Héault, Y., Yu, Y. E. and Brown, S. D. M. (2013). Surveying the Down syndrome mouse model resource identifies critical regions responsible for chronic otitis media. *Mamm. Genome* **24**, 439-445.

Bianchi, P., Ciani, E., Guidi, S., Trazzi, S., Felice, D., Grossi, G., Fernandez, M., Giuliani, A., Calza, L. and Bartesaghi, R. (2010). Early pharmacotherapy restores neurogenesis and cognitive performance in the Ts65Dn mouse model for Down syndrome. *J. Neurosci.* **30**, 8769-8779.

Birling, M.-C., Schaeffer, L., André, P., Lindner, L., Maréchal, D., Ayadi, A., Sorg, T., Pavlovic, G. and Héault, Y. (2017). Efficient and rapid generation of large genomic variants in rats and mice using CRISMERE. *Sci. Rep.* **7**, 43331.

- Blanchard, J., Bolognin, S., Chohan, M. O., Rabe, A., Iqbal, K. and Grundke-Iqbal, I. (2011). Rescue of synaptic failure and alleviation of learning and memory impairments in a trisomic mouse model of down syndrome. *J. Neuropathol. Exp. Neurol.* **70**, 1070-1079.
- Blazek, J. D., Malik, A. M., Tischbein, M., Arbones, M. L., Moore, C. S. and Roper, R. J. (2015). Abnormal mineralization of the Ts65Dn Down syndrome mouse appendicular skeleton begins during embryonic development in a Dyrk1a-independent manner. *Mech. Dev.* **136**, 133-142.
- Branchi, I., Bichler, Z., Minghetti, L., Delabar, J. M., Malchiodi-Albedi, F., Gonzalez, M.-C., Chettouh, Z., Nicolini, A., Chabert, C., Smith, D. J. et al. (2004). Transgenic mouse in vivo library of human Down syndrome critical region 1: association between DYRK1A overexpression, brain development abnormalities, and cell cycle protein alteration. *J. Neuropathol. Exp. Neurol.* **63**, 429-440.
- Braudeau, J., Dauphinot, L., Duchon, A., Loistron, A., Dodd, R. H., Héroult, Y., Delatour, B. and Potier, M. C. (2011a). Chronic treatment with a promnesiant GABA-A alpha5-selective inverse agonist increases immediate early genes expression during memory processing in mice and rectifies their expression levels in a Down syndrome mouse model. *Adv. Pharmacol. Sci.* **2011**, 153218.
- Braudeau, J., Delatour, B., Duchon, A., Pereira, P. L., Dauphinot, L., De Chaumont, F., Olivo-Marin, J.-C., Dodd, R. H., Héroult, Y. and Potier, M.-C. (2011b). Specific targeting of the GABA-A receptor alpha5 subtype by a selective inverse agonist restores cognitive deficits in Down syndrome mice. *J. Psychopharmacol.* **25**, 1030-1042.
- Braut, V., Pereira, P., Duchon, A. and Héroult, Y. (2006). Modeling chromosomes in mouse to explore the function of genes, genomic disorders, and chromosomal organization. *PLoS Genet.* **2**, e86.
- Braut, V., Besson, V., Magnol, L., Duchon, A. and Héroult, Y. (2007). Cre/loxP-mediated chromosome engineering of the mouse genome. *Handb. Exp. Pharmacol.* **178**, 29-48.
- Braut, V., Duchon, A., Romestaing, C., Sahun, I., Pothion, S., Karout, M., Borel, C., Dembele, D., Bizot, J.-C., Messaddeq, N. et al. (2015a). Opposite phenotypes of muscle strength and locomotor function in mouse models of partial trisomy and monosomy 21 for the proximal Hspa13-App region. *PLoS Genet.* **11**, e1005062.
- Braut, V., Romestaing, C., Sahun, I., Karout, M., Borel, C., Dembele, D., Bizot, J.-C., Messaddeq, N., Sharp, A. J., Roussel, D. et al. (2015b). Locomotor dysfunction and hypotonia in Down syndrome mouse models for the HSPA13-APP region as a consequence of dosage sensitive genes controlling muscular metabolism and mitochondrial function. *Acta Physiol.* **214**, 11-12.
- Canfield, M. A., Honein, M. A., Yuskiv, N., Xing, J., Mai, C. T., Collins, J. S., Devine, O., Petrini, J., Ramadhani, T. A., Hobbs, C. A. et al. (2006). National estimates and race/ethnic-specific variation of selected birth defects in the United States, 1999-2001. *Birth Defects Res. A Clin. Mol. Teratol.* **76**, 747-756.
- Cataldo, A. M., Peterhoff, C. M., Troncoso, J. C., Gomez-Isla, T., Hyman, B. T. and Nixon, R. A. (2000). Endocytic pathway abnormalities precede amyloid beta deposition in sporadic Alzheimer's disease and Down syndrome: differential effects of APOE genotype and presenilin mutations. *Am. J. Pathol.* **157**, 277-286.
- Cataldo, A. M., Mathews, P. M., Boiteau, A. B., Hassinger, L. C., Peterhoff, C. M., Jiang, Y., Mullaney, K., Neve, R. L., Gruenberg, J. and Nixon, R. A. (2008). Down syndrome fibroblast model of Alzheimer-related endosome pathology: accelerated endocytosis promotes late endocytic defects. *Am. J. Pathol.* **173**, 370-384.
- Chang, Q. and Gold, P. E. (2008). Age-related changes in memory and in acetylcholine functions in the hippocampus in the Ts65Dn mouse, a model of Down syndrome. *Neurobiol. Learn. Mem.* **89**, 167-177.
- Chen, C.-K., Symmons, O., Uslu, V. V., Tsujimura, T., Ruf, S., Smedley, D. and Spitz, F. (2013). TRACER: a resource to study the regulatory architecture of the mouse genome. *BMC Genomics* **14**, 215.
- Choong, X. Y., Tosh, J. L., Pulford, L. J. and Fisher, E. M. C. (2015). Dissecting Alzheimer disease in Down syndrome using mouse models. *Front. Behav. Neurosci.* **9**, 268.
- Clark, S., Schwalbe, J., Stasko, M. R., Yarowsky, P. J. and Costa, A. C. S. (2006). Fluoxetine rescues deficient neurogenesis in hippocampus of the Ts65Dn mouse model for Down syndrome. *Exp. Neurol.* **200**, 256-261.
- Cohen, S. J. and Stackman, R. W. Jr. (2015). Assessing rodent hippocampal involvement in the novel object recognition task. A review. *Behav. Brain Res.* **285**, 105-117.
- Colas, D., Chuluun, B., Warrior, D., Blank, M., Wetmore, D. Z., Buckmaster, P., Garner, C. C. and Heller, H. C. (2013). Short-term treatment with the GABA receptor antagonist pentylentetrazole produces a sustained pro-cognitive benefit in a mouse model of Down's syndrome. *Br. J. Pharmacol.* **169**, 963-973.
- Contestabile, A., Greco, B., Ghezzi, D., Tucci, V., Benfenati, F. and Gasparini, L. (2013). Lithium rescues synaptic plasticity and memory in Down syndrome mice. *J. Clin. Invest.* **123**, 348-361.
- Cooper, A., Grigoryan, G., Guy-David, L., Tsoory, M. M., Chen, A. and Reuveny, E. (2012). Trisomy of the G protein-coupled K<sup>+</sup> channel gene, *Kcnj6*, affects reward mechanisms, cognitive functions, and synaptic plasticity in mice. *Proc. Natl. Acad. Sci. USA* **109**, 2642-2647.
- Corrales, A., Martínez, P., García, S., Vidal, V., García, E., Flórez, J., Sanchez-Barceló, E. J., Martínez-Cué, C. and Rueda, N. (2013). Long-term oral administration of melatonin improves spatial learning and memory and protects against cholinergic degeneration in middle-aged Ts65Dn mice, a model of Down syndrome. *J. Pineal Res.* **54**, 346-358.
- Corrales, A., Vidal, R., García, S., Vidal, V., Martínez, P., García, E., Flórez, J., Sanchez-Barceló, E. J., Martínez-Cué, C. and Rueda, N. (2014). Chronic melatonin treatment rescues electrophysiological and neuromorphological deficits in a mouse model of Down syndrome. *J. Pineal Res.* **56**, 51-61.
- Costa, A. C. S. and Grybko, M. J. (2005). Deficits in hippocampal CA1 LTP induced by TBS but not HFS in the Ts65Dn mouse: a model of Down syndrome. *Neurosci. Lett.* **382**, 317-322.
- Costa, A. C. S., Scott-Mckean, J. J. and Stasko, M. R. (2008). Acute injections of the NMDA receptor antagonist memantine rescue performance deficits of the Ts65Dn mouse model of Down syndrome on a fear conditioning test. *Neuropsychopharmacology* **33**, 1624-1632.
- Dang, V., Medina, B., Das, D., Moghadam, S., Martin, K. J., Lin, B., Naik, P., Patel, D., Nosheny, R., Wesson Ashford, J. et al. (2014). Formoterol, a long-acting beta2 adrenergic agonist, improves cognitive function and promotes dendritic complexity in a mouse model of Down syndrome. *Biol. Psychiatry* **75**, 179-188.
- Das, I. and Reeves, R. H. (2011). The use of mouse models to understand and improve cognitive deficits in Down syndrome. *Dis. Model. Mech.* **4**, 596-606.
- Das, I., Park, J.-M., Shin, J. H., Jeon, S. K., Lorenzi, H., Linden, D. J., Worley, P. F. and Reeves, R. H. (2013). Hedgehog agonist therapy corrects structural and cognitive deficits in a Down syndrome mouse model. *Sci. Transl. Med.* **5**, 201ra120.
- Davisson, M. T., Schmidt, C. and Akeson, E. C. (1990). Segmental trisomy of murine chromosome 16: a new model system for studying Down syndrome. *Prog. Clin. Biol. Res.* **360**, 263-280.
- De Graaf, G., Buckley, F. and Skotko, B. G. (2016). Live births, natural losses, and elective terminations with Down syndrome in Massachusetts. *Genet. Med.* **18**, 459-466.
- De la Torre, R., De Sola, S., Pons, M., Duchon, A., De Lagran, M. M., Farré, M., Fitó, M., Benejam, B., Langohr, K., Rodriguez, J. et al. (2014). Epigallocatechin-3-gallate, a DYRK1A inhibitor, rescues cognitive deficits in Down syndrome mouse models and in humans. *Mol. Nutr. Food Res.* **58**, 278-288.
- De la Torre, R., De Sola, S., Hernandez, G., Farré, M., Pujol, J., Rodriguez, J., Espadaler, J. M., Langohr, K., Cuenca-Royo, A., Principe, A. et al. (2016). Safety and efficacy of cognitive training plus epigallocatechin-3-gallate in young adults with Down's syndrome (TESDAD): a double-blind, randomised, placebo-controlled, phase 2 trial. *Lancet Neurol.* **15**, 801-810.
- De Souza, F. M. S., Busquet, N., Blatner, M., Maclean, K. N. and Restrepo, D. (2011). Galantamine improves olfactory learning in the Ts65Dn mouse model of Down syndrome. *Sci. Rep.* **1**, 137.
- Dekker, A. D., Strydom, A., Coppus, A. M. W., Nizetic, D., Vermeiren, Y., Naudé, P. J. W., Van Dam, D., Potier, M.-C., Fortea, J. and De Deyn, P. P. (2015). Behavioural and psychological symptoms of dementia in Down syndrome: Early indicators of clinical Alzheimer's disease? *Cortex* **73**, 36-61.
- Delabar, J. M., Theophile, D., Rahmani, Z., Chettouh, Z., Blouin, J. L., Prieur, M., Noel, B. and Sinet, P. M. (1993). Molecular mapping of twenty-four features of Down syndrome on chromosome 21. *Eur. J. Hum. Genet.* **1**, 114-124.
- D'hooge, R. and De Deyn, P. P. (2001). Applications of the Morris water maze in the study of learning and memory. *Brain Res. Rev.* **36**, 60-90.
- Dierssen, M. and De Lagrán, M. M. (2006). DYRK1A (dual-specificity tyrosine-phosphorylated and -regulated kinase 1A): a gene with dosage effect during development and neurogenesis. *ScientificWorldJournal* **6**, 1911-1922.
- Dierssen, M., Herault, Y. and Estivill, X. (2009). Aneuploidy: from a physiological mechanism of variance to Down syndrome. *Physiol. Rev.* **89**, 887-920.
- Do, C., Xing, Z., Yu, Y. E. and Tycko, B. (2017). Trans-acting epigenetic effects of chromosomal aneuploidies: lessons from Down syndrome and mouse models. *Epigenomics* **9**, 189-207.
- Dowjat, W. K., Adayev, T., Kuchna, I., Nowicki, K., Palmieriello, S., Hwang, Y. W. and Wegiel, J. (2007). Trisomy-driven overexpression of DYRK1A kinase in the brain of subjects with Down syndrome. *Neurosci. Lett.* **413**, 77-81.
- Duchon, A. and Herault, Y. (2016). DYRK1a, a dosage-sensitive gene involved in neurodevelopmental disorders, is a target for drug development in Down syndrome. *Front. Behav. Neurosci.* **10**, 104.
- Duchon, A., Besson, V., Pereira, P. L., Magnol, L. and Herault, Y. (2008). Inducing segmental aneuploid mosaicism in the mouse through targeted asymmetric sister chromatid event of recombination. *Genetics* **180**, 51-59.
- Duchon, A., Pothion, S., Braut, V., Sharp, A. J., Tybulewicz, V. L. J., Fisher, E. M. C. and Herault, Y. (2011a). The telomeric part of the human chromosome 21 from *Cstb* to *Prrtm2* is not necessary for the locomotor and short-term memory deficits observed in the Tc1 mouse model of Down syndrome. *Behav. Brain Res.* **217**, 271-281.
- Duchon, A., Raveau, M., Chevalier, C., Nalesso, V., Sharp, A. J. and Herault, Y. (2011b). Identification of the translocation breakpoints in the Ts65Dn and Ts1Cje mouse lines: relevance for modeling Down syndrome. *Mamm. Genome* **22**, 674-684.
- Faizi, M., Bader, P. L., Tun, C., Encarnacion, A., Kleschevnikov, A., Belichenko, P., Saw, N., Priestley, M., Tsien, R. W., Mobley, W. C. et al. (2011). Comprehensive behavioral phenotyping of Ts65Dn mouse model of Down

- syndrome: activation of beta1-adrenergic receptor by xamoterol as a potential cognitive enhancer. *Neurobiol. Dis.* **43**, 397-413.
- Fernandez, F., Morishita, W., Zuniga, E., Nguyen, J., Blank, M., Malenka, R. C. and Garner, C. C.** (2007). Pharmacotherapy for cognitive impairment in a mouse model of Down syndrome. *Nat. Neurosci.* **10**, 411-413.
- Fortress, A. M., Hamlett, E. D., Vazey, E. M., Aston-Jones, G., Cass, W. A., Boger, H. A. and Granholm, A.-C. E.** (2015). Designer receptors enhance memory in a mouse model of Down syndrome. *J. Neurosci.* **35**, 1343-1353.
- García-Cerro, S., Martínez, P., Vidal, V., Corrales, A., Flórez, J., Vidal, R., Rueda, N., Arbonés, M. L. and Martínez-Cuá, C.** (2014). Overexpression of Dyrk1A is implicated in several cognitive, electrophysiological and neuromorphological alterations found in a mouse model of Down syndrome. *PLoS ONE* **9**, e106572.
- Ghezzi, A., Salvioli, S., Solimando, M. C., Palmieri, A., Chiostergi, C., Scurti, M., Lomartire, L., Bedetti, F., Cocchi, G., Follo, D. et al.** (2014). Age-related changes of adaptive and neuropsychological features in persons with Down Syndrome. *PLoS ONE* **9**, e113111.
- Giacomini, A., Stagni, F., Trazzi, S., Guidi, S., Emili, M., Brigham, E., Ciani, E. and Bartesaghi, R.** (2015). Inhibition of APP gamma-secretase restores Sonic Hedgehog signaling and neurogenesis in the Ts65Dn mouse model of Down syndrome. *Neurobiol. Dis.* **82**, 385-396.
- Goodliffe, J. W., Olmos-Serrano, J. L., Aziz, N. M., Pennings, J. L. A., Guedj, F., Bianchi, D. W. and Haydar, T. F.** (2016). Absence of prenatal forebrain defects in the Dp(16)1Yey/+ mouse model of Down syndrome. *J. Neurosci.* **36**, 2926-2944.
- Granholm, A.-C. E., Sanders, L. A. and Crnic, L. S.** (2000). Loss of cholinergic phenotype in basal forebrain coincides with cognitive decline in a mouse model of Down's syndrome. *Exp. Neurol.* **161**, 647-663.
- Granholm, A.-C. E., Ford, K. A., Hyde, L. A., Bimonte, H. A., Hunter, C. L., Nelson, M., Albeck, D., Sanders, L. A., Mufson, E. J. and Crnic, L. S.** (2002). Estrogen restores cognition and cholinergic phenotype in an animal model of Down syndrome. *Physiol. Behav.* **77**, 371-385.
- Granholm, A.-C., Sanders, L., Seo, H., Lin, L., Ford, K. and Isacson, O.** (2003). Estrogen alters amyloid precursor protein as well as dendritic and cholinergic markers in a mouse model of Down syndrome. *Hippocampus* **13**, 905-914.
- Grau, C., Arató, K., Fernández-Fernández, J. M., Valderrama, A., Sindreu, C., Fillat, C., Ferrer, I., DE, L. A. Luna, S. and Altfaj, X.** (2014). DYRK1A-mediated phosphorylation of GluN2A at Ser(1048) regulates the surface expression and channel activity of GluN1/GluN2A receptors. *Front. Cell Neurosci.* **8**, 331.
- Gribble, S. M., Wiseman, F. K., Clayton, S., Prigmore, E., Langley, E., Yang, F., Maguire, S., Fu, B., Rajan, D., Sheppard, O. et al.** (2013). Massively parallel sequencing reveals the complex structure of an irradiated human chromosome on a mouse background in the Tc1 model of Down syndrome. *PLoS ONE* **8**, e60482.
- Gropp, A.** (1974). Animal model of human disease. Autosomal trisomy, developmental impairment and fetal death. *Am. J. Pathol.* **77**, 539-542.
- Gropp, A., Kolbus, U. and Giers, D.** (1975). Systematic approach to the study of trisomy in the mouse. II. *Cytogenet. Cell Genet.* **14**, 42-62.
- Guedj, F., Sébrié, C., Rivals, I., Ledru, A., Paly, E., Bizot, J. C., Smith, D., Rubin, E., Gillet, B., Arbones, M. et al.** (2009). Green tea polyphenols rescue of brain defects induced by overexpression of DYRK1A. *PLoS ONE* **4**, e4606.
- Guedj, F., Pereira, P. L., Najas, S., Barallobre, M.-J., Chabert, C., Souchet, B., Sebric, C., Verney, C., Hérault, Y., Arbones, M. et al.** (2012). DYRK1A: a master regulatory protein controlling brain growth. *Neurobiol. Dis.* **46**, 190-203.
- Guedj, F., Pennings, J. L. A., Massingham, L. J., Wick, H. C., Siegel, A. E., Tantravahi, U. and Bianchi, D. W.** (2016). An integrated human/murine transcriptome and pathway approach to identify prenatal treatments for Down syndrome. *Sci. Rep.* **6**, 32353.
- Guidi, S., Stagni, F., Bianchi, P., Ciani, E., Giacomini, A., De Franceschi, M., Moldrich, R., Kurniawan, N., Mardon, K., Giuliani, A. et al.** (2014). Prenatal pharmacotherapy rescues brain development in a Down's syndrome mouse model. *Brain* **137**, 380-401.
- Gupta, M., Dhanasekaran, A. R. and Gardiner, K. J.** (2016). Mouse models of Down syndrome: gene content and consequences. *Mamm. Genome* **27**, 538-555.
- Gutierrez-Castellanos, N., Winkelman, B. H. J., Tolosa-Rodriguez, L., Devenney, B., Reeves, R. H. and De Zeeuw, C. I.** (2013). Size does not always matter: Ts65Dn Down syndrome mice show cerebellum-dependent motor learning deficits that cannot be rescued by postnatal SAG treatment. *J. Neurosci.* **33**, 15408-15413.
- Haas, M. A., Bell, D., Slender, A., Lana-Elola, E., Watson-Scales, S., Fisher, E. M. C., Tybulewicz, V. L. J. and Guillemot, F.** (2013). Alterations to dendritic spine morphology, but not dendrite patterning, of cortical projection neurons in Tc1 and Ts1Rhr mouse models of Down syndrome. *PLoS ONE* **8**, e78561.
- Hall, J. H., Wiseman, F. K., Fisher, E. M. C., Tybulewicz, V. L. J., Harwood, J. L. and Good, M. A.** (2016). Tc1 mouse model of trisomy-21 dissociates properties of short- and long-term recognition memory. *Neurobiol. Learn. Mem.* **130**, 118-128.
- Hamlett, E. D., Boger, H. A., Ledreux, A., Kelley, C. M., Mufson, E. J., Falangola, M. F., Guilfoyle, D. N., Nixon, R. A., Patterson, D., Duval, N. et al.** (2016). Cognitive impairment, neuroimaging, and Alzheimer neuropathology in mouse models of Down syndrome. *Curr. Alzheimer Res.* **13**, 35-52.
- Hanson, J. E., Weber, M., Meilandt, W. J., Wu, T., Luu, T., Deng, L., Shamloo, M., Sheng, M., Scearce-Levie, K. and Zhou, Q.** (2013). GluN2B antagonism affects interneurons and leads to immediate and persistent changes in synaptic plasticity, oscillations, and behavior. *Neuropsychopharmacology* **38**, 1221-1233.
- Head, E., Lott, I. T., Wilcock, D. M. and Lemere, C. A.** (2015). Aging in Down Syndrome and the Development of Alzheimer's Disease Neuropathology. *Curr. Alzheimer Res.* **13**, 18-29.
- Heinen, M., Hettich, M. M., Ryan, D. P., Schnell, S., Paesler, K. and Ehninger, D.** (2012). Adult-onset fluoxetine treatment does not improve behavioral impairments and may have adverse effects on the Ts65Dn mouse model of Down syndrome. *Neural Plast.* **2012**, 467251.
- Hérault, Y., Rassoulzadegan, M., Cuzin, F. and Duboule, D.** (1998). Engineering chromosomes in mice through targeted meiotic recombination (TAMERE). *Nat. Genet.* **20**, 381-384.
- Hérault, Y., Duchon, A., Velot, E., Maréchal, D. and Brault, V.** (2012). The in vivo Down syndrome genomic library in mouse. *Prog. Brain Res.* **197**, 169-197.
- Hervé, B., Coussement, A., Gilbert, T., Dumont, F., Jacques, S., Cuisset, L., Chicard, M., Hizem, S., Bourdoncle, P., Letourneur, F. et al.** (2016). Aneuploidy: the impact of chromosome imbalance on nuclear organization and overall genome expression. *Clin. Genet.* **90**, 35-48.
- Hughes, R. N.** (2004). The value of spontaneous alternation behavior (SAB) as a test of retention in pharmacological investigations of memory. *Neurosci. Biobehav. Rev.* **28**, 497-505.
- Hunter, C. L., Bachman, D. and Granholm, A.-C.** (2004a). Minocycline prevents cholinergic loss in a mouse model of Down's syndrome. *Ann. Neurol.* **56**, 675-688.
- Hunter, C. L., Bimonte-Nelson, H. A., Nelson, M., Eckman, C. B. and Granholm, A.-C.** (2004b). Behavioral and neurobiological markers of Alzheimer's disease in Ts65Dn mice: effects of estrogen. *Neurobiol. Aging* **25**, 873-884.
- Ichinohe, A., Kanaumi, T., Takashima, S., Enokido, Y., Nagai, Y. and Kimura, H.** (2005). Cystathionine beta-synthase is enriched in the brains of Down's patients. *Biochem. Biophys. Res. Commun.* **338**, 1547-1550.
- Incerti, M., Toso, L., Vink, J., Roberson, R., Nold, C., Abebe, D. and Spong, C. Y.** (2011). Prevention of learning deficit in a Down syndrome model. *Obstet. Gynecol.* **117**, 354-361.
- Incerti, M., Horowitz, K., Roberson, R., Abebe, D., Toso, L., Caballero, M. and Spong, C. Y.** (2012). Prenatal treatment prevents learning deficit in Down syndrome model. *PLoS ONE* **7**, e50724.
- Inglis, A., Hippman, C. and Austin, J. C.** (2012). Prenatal testing for Down syndrome: the perspectives of parents of individuals with Down syndrome. *Am. J. Med. Genet. A* **158A**, 743-750.
- Jiang, J. and Hui, C.** (2008). Hedgehog signaling in development and cancer. *Dev. Cell* **15**, 801-812.
- Jiang, X., Liu, C., Yu, T., Zhang, L., Meng, K., Xing, Z., Belichenko, P. V., Kleschevnikov, A. M., Pao, A., Peresie, J. et al.** (2015). Genetic dissection of the Down syndrome critical region. *Hum. Mol. Genet.* **24**, 6540-6551.
- Jiang, Y., Rigoglioso, A., Peterhoff, C. M., Pawlik, M., Sato, Y., Bleiwas, C., Stavrides, P., Smiley, J. F., Ginsberg, S. D., Mathews, P. M. et al.** (2016). Partial BACE1 reduction in a Down syndrome mouse model blocks Alzheimer-related endosomal anomalies and cholinergic neurodegeneration: role of APP-CTF. *Neurobiol. Aging* **39**, 90-98.
- Joshi, K., Shen, L., Michaeli, A., Salter, M., Thibault-Messier, G., Hashmi, S., Eubanks, J. H., Cortez, M. A. and Snead, O. C.** (2016). Infantile spasms in Down syndrome: rescue by knockdown of the GIRK2 channel. *Ann. Neurol.* **80**, 511-521.
- Karmiloff-Smith, A., Al-Janabi, T., D'souza, H., Groet, J., Massand, E., Mok, K., Startin, C., Fisher, E., Hardy, J., Nizetic, D. et al.** (2016). The importance of understanding individual differences in Down syndrome. *F1000Res* **5**, 389.
- Kelley, C. M., Ash, J. A., Powers, B. E., Velazquez, R., Alldred, M. J., Ikonomic, M. D., Ginsberg, S. D., Strupp, B. J. and Mufson, E. J.** (2016). Effects of maternal choline supplementation on the septohippocampal cholinergic system in the Ts65Dn mouse model of Down syndrome. *Curr. Alzheimer Res.* **13**, 84-96.
- Kern, D. S., Maclean, K. N., Jiang, H., Synder, E. Y., Sladek, J. R., Jr. and Bjugstad, K. B.** (2011). Neural stem cells reduce hippocampal tau and reelin accumulation in aged Ts65Dn Down syndrome mice. *Cell Transplant.* **20**, 371-379.
- Khoshnood, B., Greenlees, R., Loane, M., Dolk, H., Committee, E. P. M. and Group, E. W.** (2011). Paper 2: EUROCAT public health indicators for congenital anomalies in Europe. *Birth Defects Res. A Clin. Mol. Teratol.* **91** Suppl. 1, S16-S22.
- Kida, E., Rabe, A., Walus, M., Albertini, G. and Golabek, A. A.** (2013). Long-term running alleviates some behavioral and molecular abnormalities in Down syndrome mouse model Ts65Dn. *Exp. Neurol.* **240**, 178-189.
- Kleschevnikov, A. M., Belichenko, P. V., Villar, A. J., Epstein, C. J., Malenka, R. C. and Mobley, W. C.** (2004). Hippocampal long-term potentiation suppressed by increased inhibition in the Ts65Dn mouse, a genetic model of Down syndrome. *J. Neurosci.* **24**, 8153-8160.
- Kleschevnikov, A. M., Belichenko, P. V., Faizi, M., Jacobs, L. F., Htun, K., Shamloo, M. and Mobley, W. C.** (2012a). Deficits in cognition and synaptic plasticity in a mouse model of Down syndrome ameliorated by GABAB receptor antagonists. *J. Neurosci.* **32**, 9217-9227.
- Kleschevnikov, A. M., Belichenko, P. V., Gall, J., George, L., Nosheny, R., Maloney, M. T., Salehi, A. and Mobley, W. C.** (2012b). Increased efficiency of the

- GABAA and GABAB receptor-mediated neurotransmission in the Ts65Dn mouse model of Down syndrome. *Neurobiol. Dis.* **45**, 683-691.
- Korbel, J. O., Tirosh-Wagner, T., Urban, A. E., Chen, X.-N., Kasowski, M., Dai, L., Grubert, F., Erdman, C., Gao, M. C., Lange, K. et al. (2009). The genetic architecture of Down syndrome phenotypes revealed by high-resolution analysis of human segmental trisomies. *Proc. Natl. Acad. Sci. USA* **106**, 12031-12036.
- Korenberg, J. R., Chen, X. N., Schipper, R., Sun, Z., Gonsky, R., Gerwehr, S., Carpenter, N., Daumer, C., Dignan, P. and Distèche, C. (1994). Down syndrome phenotypes: the consequences of chromosomal imbalance. *Proc. Natl. Acad. Sci. USA* **91**, 4997-5001.
- Krinsky-Mchale, S. J., Devenny, D. A., Gu, H., Jenkins, E. C., Kittler, P., Murty, V. V., Schupf, N., Scott, L., Tycko, B., Urv, T. K. et al. (2008). Successful aging in a 70-year-old man with down syndrome: a case study. *Intellect. Dev. Disabil.* **46**, 215-228.
- Kruszka, P., Porras, A. R., Sobering, A. K., Ikolo, F. A., La Qua, S., Shotelersuk, V., Chung, B. H. Y., Mok, G. T. K., Uwineza, A., Mutesa, L. et al. (2017). Down syndrome in diverse populations. *Am. J. Med. Genet. A* **173**, 42-53.
- Laffaire, J., Rivals, I., Dauphinot, L., Pasteau, F., Wehrle, R., Larrat, B., Vitalis, T., Moldrich, R. X., Rossier, J., Sinkus, R. et al. (2009). Gene expression signature of cerebellar hypoplasia in a mouse model of Down syndrome during postnatal development. *BMC Genomics* **10**, 138.
- Laguna, A., Barallobre, M.-J., Marchena, M., Mateus, C., Ramírez, E., Martínez-Cue, C., Delabar, J. M., Castelo-Branco, M., de la Villa, P. and Arbonés, M. L. (2013). Triplication of DYRK1A causes retinal structural and functional alterations in Down syndrome. *Hum. Mol. Genet.* **22**, 2775-2784.
- Lana-Elola, E., Watson-Scales, S., Slender, A., Gibbins, D., Martineau, A., Douglas, C., Mohun, T., Fisher, E. M. C. and Tybulewicz, V. L. J. (2016). Genetic dissection of Down syndrome-associated congenital heart defects using a new mouse mapping panel. *Life* **5**, e11614.
- Latchney, S. E., Jaramillo, T. C., Rivera, P. D., Eisch, A. J. and Powell, C. M. (2015). Chronic P7C3 treatment restores hippocampal neurogenesis in the Ts65Dn mouse model of Down Syndrome [Corrected]. *Neurosci. Lett.* **591**, 86-92.
- Lee, S., Ahmed, T., Kim, H., Choi, S., Kim, D.-S., Kim, S. J., Cho, J. and Shin, H.-S. (2011). Bidirectional modulation of fear extinction by mediadorsal thalamic firing in mice. *Nat. Neurosci.* **15**, 308-314.
- Letourneau, A., Santoni, F. A., Bonilla, X., Sailani, M. R., Gonzalez, D., Kind, J., Chevalier, C., Thurman, R., Sandstrom, R. S., Hibaoui, Y. et al. (2014). Domains of genome-wide gene expression dysregulation in Down's syndrome. *Nature* **508**, 345-350.
- Li, Z., Yu, T., Morishima, M., Pao, A., Laduca, J., Conroy, J., Nowak, N., Matsui, S.-I., Shiraiishi, I. and Yu, Y. E. (2007). Duplication of the entire 22.9 Mb human chromosome 21 syntenic region on mouse chromosome 16 causes cardiovascular and gastrointestinal abnormalities. *Hum. Mol. Genet.* **16**, 1359-1366.
- Li, H., Edie, S., Klinedinst, D., Jeong, J. S., Blackshaw, S., Maslen, C. L. and Reeves, R. H. (2016). Penetrance of congenital heart disease in a mouse model of Down syndrome depends on a trisomic potentiator of a disomic modifier. *Genetics* **203**, 763-770.
- Lignon, J. M., Bichler, Z., Hivert, B., Gannier, F. E., Cosnay, P., Del Rio, J. A., Migliore-Samour, D. and Malécot, C. O. (2008). Altered heart rate control in transgenic mice carrying the KCNJ6 gene of the human chromosome 21. *Physiol. Genomics* **33**, 230-239.
- Liu, C., Morishima, M., Yu, T., Matsui, S.-I., Zhang, L., Fu, D., Pao, A., Costa, A. C., Gardiner, K. J., Cowell, J. K. et al. (2011). Genetic analysis of Down syndrome-associated heart defects in mice. *Hum. Genet.* **130**, 623-632.
- Liu, C., Morishima, M., Jiang, X., Yu, T., Meng, K., Ray, D., Pao, A., Ye, P., Parmacek, M. S. and Yu, Y. E. (2014). Engineered chromosome-based genetic mapping establishes a 3.7 Mb critical genomic region for Down syndrome-associated heart defects in mice. *Hum. Genet.* **133**, 743-753.
- Llorens-Martín, M. V., Rueda, N., Tejada, G. S., Flórez, J., Trejo, J. L. and Martínez-Cué, C. (2010). Effects of voluntary physical exercise on adult hippocampal neurogenesis and behavior of Ts65Dn mice, a model of Down syndrome. *Neuroscience* **171**, 1228-1240.
- Loane, M., Morris, J. K., Addor, M.-C., Arriola, L., Budd, J., Doray, B., Garne, E., Gatt, M., Haeusler, M., Khoshnood, B. et al. (2013). Twenty-year trends in the prevalence of Down syndrome and other trisomies in Europe: impact of maternal age and prenatal screening. *Eur. J. Hum. Genet.* **21**, 27-33.
- Lockrow, J., Prakasam, A., Huang, P., Bimonte-Nelson, H., Sambamurti, K. and Granholm, A.-C. (2009). Cholinergic degeneration and memory loss delayed by vitamin E in a Down syndrome mouse model. *Exp. Neurol.* **216**, 278-289.
- Lockrow, J., Boger, H., Bimonte-Nelson, H. and Granholm, A.-C. (2011). Effects of long-term memantine on memory and neuropathology in Ts65Dn mice, a model for Down syndrome. *Behav. Brain Res.* **221**, 610-622.
- Lopes Pereira, P., Magnol, L., Sahún, I., Brault, V., Duchon, A., Prandini, P., Gruart, A., Bizot, J.-C., Chadeaux-Vekemans, B., Deutsch, S. et al. (2009). A new mouse model for the trisomy of the Abcg1-U2af1 region reveals the complexity of the combinatorial genetic code of down syndrome. *Hum. Mol. Genet.* **18**, 4756-4769.
- Lyle, R., Béna, F., Gagos, S., Gehrig, C., Lopez, G., Schinzel, A., Lespinasse, J., Bottani, A., Dahoun, S., Taine, L. et al. (2009). Genotype-phenotype correlations in Down syndrome identified by array CGH in 30 cases of partial trisomy and partial monosomy chromosome 21. *Eur. J. Hum. Genet.* **17**, 454-466.
- Lysenko, L. V., Kim, J., Henry, C., Tyrtshnaia, A., Kohnz, R. A., Madamba, F., Simon, G. M., Kleschevnikova, N. E., Nomura, D. K., Ezekowitz, R. A. B. et al. (2014). Monoacylglycerol lipase inhibitor JZL184 improves behavior and neural properties in Ts65Dn mice, a model of down syndrome. *PLoS ONE* **9**, e114521.
- Malinge, S., Bliss-Moreau, M., Kirsammer, G., Diebold, L., Chlon, T., Gurbuxani, S. and Crispino, J. D. (2012). Increased dosage of the chromosome 21 ortholog Dyrk1a promotes megakaryoblastic leukemia in a murine model of Down syndrome. *J. Clin. Invest.* **122**, 948-962.
- Marechal, D., Lopes Pereira, P., Duchon, A. and Herault, Y. (2015). Dosage of the Abcg1-U2af1 region modifies locomotor and cognitive deficits observed in the Tc1 mouse model of Down syndrome. *PLoS ONE* **10**, e0115302.
- Martinez-Cue, C., Martinez, P., Rueda, N., Vidal, R., Garcia, S., Vidal, V., Corrales, A., Montero, J. A., Pazos, A., Florez, J. et al. (2013). Reducing GABAA alpha5 receptor-mediated inhibition rescues functional and neuromorphological deficits in a mouse model of Down syndrome. *J. Neurosci.* **33**, 3953-3966.
- Martinez de Lagrán, M., Altafaj, X., Gallego, X., Martí, E., Estivill, X., Sahún, I., Fillat, C. and Dierssen, M. (2004). Motor phenotypic alterations in TgDyrk1a transgenic mice implicate DYRK1A in Down syndrome motor dysfunction. *Neurobiol. Dis.* **15**, 132-142.
- Mátyás, F., Lee, J. H., Shin, H.-S. and Acsády, L. (2014). The fear circuit of the mouse forebrain: connections between the mediadorsal thalamus, frontal cortices and basolateral amygdala. *Eur. J. Neurosci.* **39**, 1810-1823.
- McCann, J. C., Hudes, M. and Ames, B. N. (2006). An overview of evidence for a causal relationship between dietary availability of choline during development and cognitive function in offspring. *Neurosci. Biobehav. Rev.* **30**, 696-712.
- McElyea, S. D., Starbuck, J. M., Tumbleson-Brink, D. M., Harrington, E., Blazek, J. D., Ghoneima, A., Kula, K. and Roper, R. J. (2016). Influence of prenatal EGCG treatment and Dyrk1a dosage reduction on craniofacial features associated with Down syndrome. *Hum. Mol. Genet.* **25**, 4856-4869.
- Mendioroz, M., Do, C., Jiang, X., Liu, C., Darbary, H. K., Lang, C. F., Lin, J., Thomas, A., Abu-Amero, S., Stanier, P. et al. (2015). Trans effects of chromosome aneuploidies on DNA methylation patterns in human Down syndrome and mouse models. *Genome Biol.* **16**, 263.
- Moon, J., Chen, M., Gandhi, S. U., Strawderman, M., Levitsky, D. A., Maclean, K. N. and Strupp, B. J. (2010). Perinatal choline supplementation improves cognitive functioning and emotion regulation in the Ts65Dn mouse model of Down syndrome. *Behav. Neurosci.* **124**, 346-361.
- Moore, C. S. (2006). Postnatal lethality and cardiac anomalies in the Ts65Dn Down syndrome mouse model. *Mamm. Genome* **17**, 1005-1012.
- Moore, C. S., Hawkins, C., Franca, A., Lawler, A., Devenney, B., Das, I. and Reeves, R. H. (2010). Increased male reproductive success in Ts65Dn "Down syndrome" mice. *Mamm. Genome* **21**, 543-549.
- Moran, T. H., Capone, G. T., Knipp, S., Davissson, M. T., Reeves, R. H. and Gearhart, J. D. (2002). The effects of piracetam on cognitive performance in a mouse model of Down's syndrome. *Physiol. Behav.* **77**, 403-409.
- Morris, R. (1984). Developments of a water-maze procedure for studying spatial learning in the rat. *J. Neurosci. Methods* **11**, 47-60.
- Morris, J. K. and Wald, N. J. (2007). Estimating the risk of Down's syndrome in antenatal screening and the gestation at which this risk applies. *J. Med. Screen* **14**, 5-7.
- Morris, J. K., Wald, N. J. and Watt, H. C. (1999). Fetal loss in Down syndrome pregnancies. *Prenat. Diagn.* **19**, 142-145.
- Mouton-Liger, F., Thomas, S., Rattenbach, R., Magnol, L., Larigaldie, V., Ledru, A., Héault, Y., Verney, C. and Créau, N. (2011). PCP4 (PEP19) overexpression induces premature neuronal differentiation associated with Ca(2+)-calmodulin-dependent kinase II delta activation in mouse models of down syndrome. *J. Comp. Neurol.* **63**, 92-106.
- Mouton-Liger, F., Sahún, I., Collin, T., Lopes Pereira, P., Masini, D., Thomas, S., Paly, E., Lullier, S., Mème, S., Jouhault, Q. et al. (2014). Developmental molecular and functional cerebellar alterations induced by PCP4/PEP19 overexpression: implications for Down syndrome. *Neurobiol. Dis.* **63**, 92-106.
- Natoli, J. L., Ackerman, D. L., Mcdermott, S. and Edwards, J. G. (2012). Prenatal diagnosis of Down syndrome: a systematic review of termination rates (1995-2011). *Prenat. Diagn.* **32**, 142-153.
- Netzer, W. J., Powell, C., Nong, Y., Blundell, J., Wong, L., Duff, K., Flajolet, M. and Greengard, P. (2010). Lowering beta-amyloid levels rescues learning and memory in a Down syndrome mouse model. *PLoS ONE* **5**, e10943.
- Ng, A. P., Hu, Y., Metcalf, D., Hyland, C. D., Ierino, H., Phipson, B., Wu, D., Baldwin, T. M., Kauppi, M., Kiu, H. et al. (2015). Early lineage priming by trisomy of Erg leads to myeloproliferation in a Down syndrome model. *PLoS Genet.* **11**, e1005211.
- O'Doherty, A., Ruf, S., Mulligan, C., Hildreth, V., Errington, M. L., Cooke, S., Sesay, A., Modino, S., Vanes, L., Hernandez, D. et al. (2005). An aneuploid mouse strain carrying human chromosome 21 with Down syndrome phenotypes. *Science* **309**, 2033-2037.
- Olmos-Serrano, J. L., Kang, H. J., Tyler, W. A., Silbereis, J. C., Cheng, F., Zhu, Y., Pletikos, M., Jankovic-Rapan, L., Cramer, N. P., Galdzicki, Z. et al. (2016a). Down syndrome developmental brain transcriptome reveals defective oligodendrocyte differentiation and myelination. *Neuron* **89**, 1208-1222.

- Olmos-Serrano, J. L., Tyler, W. A., Cabral, H. J. and Haydar, T. F. (2016b). Longitudinal measures of cognition in the Ts65Dn mouse: Refining windows and defining modalities for therapeutic intervention in Down syndrome. *Exp. Neurol.* **279**, 40-56.
- Olson, L. E., Richtsmeier, J. T., Leszl, J. and Reeves, R. H. (2004). A chromosome 21 critical region does not cause specific Down syndrome phenotypes. *Science* **306**, 687-690.
- Olson, L. E., Roper, R. J., Sengstaken, C. L., Peterson, E. A., Aquino, V., Galdzicki, Z., Siarey, R., Pletnikov, M., Moran, T. H. and Reeves, R. H. (2007). Trisomy for the Down syndrome 'critical region' is necessary but not sufficient for brain phenotypes of trisomic mice. *Hum. Mol. Genet.* **16**, 774-782.
- Ortiz-Abalia, J., Sahún, I., Altafaj, X., Andreu, N., Estivill, X., Dierssen, M. and Fillat, C. (2008). Targeting Dyrk1A with AAVshRNA attenuates motor alterations in TgDyrk1a, a mouse model of Down syndrome. *Am. J. Hum. Genet.* **83**, 479-488.
- Pan, B., Wang, W., Zhong, P., Blankman, J. L., Cravatt, B. F. and Liu, Q.-S. (2011). Alterations of endocannabinoid signaling, synaptic plasticity, learning, and memory in monoacylglycerol lipase knock-out mice. *J. Neurosci.* **31**, 13420-13430.
- Park, J., Oh, Y., Yoo, L., Jung, M.-S., Song, W.-J., Lee, S.-H., Seo, H. and Chung, K. C. (2010). Dyrk1A phosphorylates p53 and inhibits proliferation of embryonic neuronal cells. *J. Biol. Chem.* **285**, 31895-31906.
- Park, J., Sung, J. Y., Song, W.-J., Chang, S. and Chung, K. C. (2012). Dyrk1A negatively regulates the actin cytoskeleton through threonine phosphorylation of N-WASP. *J. Cell Sci.* **125**, 67-80.
- Parker, S. E., Mai, C. T., Canfield, M. A., Rickard, R., Wang, Y., Meyer, R. E., Anderson, P., Mason, C. A., Collins, J. S., Kirby, R. S. et al. (2010). Updated National Birth Prevalence estimates for selected birth defects in the United States, 2004-2006. *Birth Defects Res. A Clin. Mol. Teratol.* **88**, 1008-1016.
- Patel, A., Yamashita, N., Ascaño, M., Bodmer, D., Boehm, E., Bodkin-Clarke, C., Ryu, Y. K. and Kuruvilla, R. (2015). RCAN1 links impaired neurotrophin trafficking to aberrant development of the sympathetic nervous system in Down syndrome. *Nat. Commun.* **6**, 10119.
- Paylor, R., Tracy, R., Wehner, J. and Rudy, J. W. (1994). DBA/2 and C57BL/6 mice differ in contextual fear but not auditory fear conditioning. *Behav. Neurosci.* **108**, 810-817.
- Peiris, H., Duffield, M. D., Fadista, J., Jessup, C. F., Kashmir, V., Genders, A. J., Mcgee, S. L., Martin, A. M., Saiedi, M., Morton, N. et al. (2016). A syntenic cross species aneuploidy genetic screen links RCAN1 expression to  $\beta$ -cell mitochondrial dysfunction in type 2 diabetes. *PLoS Genet.* **12**, e1006033.
- Polk, R. C., Gergics, P., Steimle, J. D., Li, H., Moskowitz, I. P., Camper, S. A. and Reeves, R. H. (2015). The pattern of congenital heart defects arising from reduced Tbx5 expression is altered in a Down syndrome mouse model. *BMC Dev. Biol.* **15**, 30.
- Pons-Espinal, M., Martínez de Lagran, M. and Dierssen, M. (2013). Environmental enrichment rescues DYRK1A activity and hippocampal adult neurogenesis in TgDyrk1A. *Neurobiol. Dis.* **60**, 18-31.
- Powell, N. M., Modat, M., Cardoso, M. J., Ma, D., Holmes, H. E., Yu, Y., O'callaghan, J., Cleary, J. O., Sinclair, B., Wiseman, F. K. et al. (2016). Fully-automated  $\mu$ MRI morphometric phenotyping of the Tc1 mouse model of Down syndrome. *PLoS One* **11**, e0162974.
- Rachdi, L., Kariyawasam, D., Aiello, V., Herault, Y., Janel, N., Delabar, J.-M., Polak, M. and Scharfmann, R. (2014). Dyrk1A induces pancreatic  $\beta$  cell mass expansion and improves glucose tolerance. *Cell Cycle* **13**, 2221-2229.
- Rachidi, M., Lopes, C., Vayssettes, C., Smith, D. J., Rubin, E. M. and Delabar, J. M. (2007). New cerebellar phenotypes in YAC transgenic mouse in vivo library of human Down syndrome critical region-1. *Biochem. Biophys. Res. Commun.* **364**, 488-494.
- Rachubinski, A. L., Crowley, S. K., Sladek, J. R., Jr, Maclean, K. N. and Bjugstad, K. B. (2012a). Effects of neonatal neural progenitor cell implantation on adult neuroanatomy and cognition in the Ts65Dn model of Down syndrome. *PLoS ONE* **7**, e36082.
- Rachubinski, A. L., Maclean, K. N., Evans, J. R. and Bjugstad, K. B. (2012b). Modulating cognitive deficits and tau accumulation in a mouse model of aging Down syndrome through neonatal implantation of neural progenitor cells. *Exp. Gerontol.* **47**, 723-733.
- Rahmani, Z., Blouin, J. L., Creau-Goldberg, N., Watkins, P. C., Mattei, J. F., Poissonnier, M., Prieur, M., Chettouh, Z., Nicole, A., Aurias, A. et al. (1989). Critical role of the D21S55 region on chromosome 21 in the pathogenesis of Down syndrome. *Proc. Natl. Acad. Sci. USA* **86**, 5958-5962.
- Ramirez-Solis, R., Liu, P. and Bradley, A. (1995). Chromosome engineering in mice. *Nature* **378**, 720-724.
- Raveau, M., Lignon, J. M., Nalesso, V., Duchon, A., Groner, Y., Sharp, A. J., Dembele, D., Brault, V. and Héralut, Y. (2012). The app-runtx1 region is critical for birth defects and electrocardiographic dysfunctions observed in a down syndrome mouse model. *PLoS Genet.* **8**, e1002724.
- Raveau, M., Nakahari, T., Asada, S., Ishihara, K., Amano, K., Shimohata, A., Sago, H. and Yamakawa, K. (2017). Brain ventriculomegaly in Down syndrome mice is caused by Pcp4 dose-dependent cilia dysfunction. *Hum. Mol. Genet.* **26**, 923-931.
- Reeves, R. H., Irving, N. G., Moran, T. H., Wohn, A., Kitt, C., Sisodia, S. S., Schmidt, C., Bronson, R. T. and Davison, M. T. (1995). A mouse model for Down syndrome exhibits learning and behaviour deficits. *Nat. Genet.* **11**, 177-184.
- Régnier, V., Billard, J.-M., Gupta, S., Potier, B., Woerner, S., Paly, E., Ledru, A., David, S., Luilier, S., Bizot, J.-C. et al. (2012). Brain phenotype of transgenic mice overexpressing cystathionine  $\beta$ -synthase. *PLoS One* **7**, e29056.
- Reinholdt, L. G., Ding, Y., Gilbert, G. J., Gilbert, G. T., Czechanski, A., Solzak, J. P., Roper, R. J., Johnson, M. T., Donahue, L. R., Lutz, C. et al. (2011). Molecular characterization of the translocation breakpoints in the Down syndrome mouse model Ts65Dn. *Mamm. Genome* **22**, 685-691.
- Reynolds, L. E., Watson, A. R., Baker, M., Jones, T. A., D'amico, G., Robinson, S. D., Joffre, C., Garrido-Urbani, S., Rodriguez-Manzanares, J. C., Martino-Echarri, E. et al. (2010). Tumour angiogenesis is reduced in the Tc1 mouse model of Down's syndrome. *Nature* **465**, 813-817.
- Richtsmeier, J. T., Baxter, L. L. and Reeves, R. H. (2000). Parallels of craniofacial maldevelopment in Down syndrome and Ts65Dn mice. *Dev. Dyn.* **217**, 137-145.
- Richtsmeier, J. T., Zumwalt, A., Carlson, E. J., Epstein, C. J. and Reeves, R. H. (2002). Craniofacial phenotypes in segmentally trisomic mouse models for Down syndrome. *Am. J. Med. Genet.* **107**, 317-324.
- Roizen, N. J. and Patterson, D. (2003). Down's syndrome. *Lancet* **361**, 1281-1289.
- Roper, R. J. and Reeves, R. H. (2006). Understanding the basis for Down syndrome phenotypes. *PLoS Genet.* **2**, e50.
- Roper, R. J., Baxter, L. L., Saran, N. G., Klinedinst, D. K., Beachy, P. A. and Reeves, R. H. (2006). Defective cerebellar response to mitogenic Hedgehog signaling in Down [corrected] syndrome mice. *Proc. Natl. Acad. Sci. USA* **103**, 1452-1456.
- Roubertoux, P. L., Bichler, Z., Pinoteau, W., Jamon, M., Sérégaza, Z., Smith, D. J., Rubin, E. and Migliore-Samour, D. (2006). Pre-weaning sensorial and motor development in mice transpolygenic for the critical region of trisomy 21. *Behav. Genet.* **36**, 377-386.
- Rovelet-Lecrux, A., Hannequin, D., Raux, G., Le Meur, N., Laquerrière, A., Vital, A., Dumanchin, C., Feuillet, S., Brice, A., Vercelletto, M. et al. (2006). APP locus duplication causes autosomal dominant early-onset Alzheimer disease with cerebral amyloid angiopathy. *Nat. Genet.* **38**, 24-26.
- Rovelet-Lecrux, A., Frebourg, T., Tuominen, H., Majamaa, K., Campion, D. and Remes, A. M. (2007). APP locus duplication in a Finnish family with dementia and intracerebral haemorrhage. *J. Neurol. Neurosurg. Psychiatry* **78**, 1158-1159.
- Rueda, N., Flórez, J. and Martínez-Cué, C. (2008a). Chronic pentylentetrazole but not donepezil treatment rescues spatial cognition in Ts65Dn mice, a model for Down syndrome. *Neurosci. Lett.* **433**, 22-27.
- Rueda, N., Florez, J. and Martínez-Cue, C. (2008b). Effects of chronic administration of SGS-111 during adulthood and during the pre- and post-natal periods on the cognitive deficits of Ts65Dn mice, a model of Down syndrome. *Behav. Brain Res.* **188**, 355-367.
- Rueda, N., Llorens-Martín, M., Flórez, J., Valdizán, E., Banerjee, P., Trejo, J. L. and Martínez-Cué, C. (2010). Memantine normalizes several phenotypic features in the Ts65Dn mouse model of Down syndrome. *J. Alzheimers Dis.* **21**, 277-290.
- Ruf, S., Symmons, O., Uslu, V. V., Dolle, D., Hot, C., Ettwiller, L. and Spitz, F. (2011). Large-scale analysis of the regulatory architecture of the mouse genome with a transposon-associated sensor. *Nat. Genet.* **43**, 379-386.
- Ruiz-Mejias, M., Martínez de Lagran, M., Mattia, M., Castano-Prat, P., Perez-Mendez, L., Ciria-Suarez, L., Gener, T., Sancristobal, B., García-Ojalvo, J., Gruart, A. et al. (2016). Overexpression of Dyrk1a, a Down Syndrome Candidate, Decreases Excitability and Impairs Gamma Oscillations in the Prefrontal Cortex. *J. Neurosci.* **36**, 3648-3659.
- Sago, H., Carlson, E. J., Smith, D. J., Kilbridge, J., Rubin, E. M., Mobley, W. C., Epstein, C. J. and Huang, T.-T. (1998). Ts1Cje, a partial trisomy 16 mouse model for Down syndrome, exhibits learning and behavioral abnormalities. *Proc. Natl. Acad. Sci. USA* **95**, 6256-6261.
- Sahun, I., Marechal, D., Pereira, P. L., Nalesso, V., Gruart, A., Delgado Garcia, J. M., Antonarakis, S. E., Dierssen, M. and Herault, Y. (2014). Cognition and Hippocampal Plasticity in the Mouse Is Altered by Monosomy of a Genomic Region Implicated in Down Syndrome. *Genetics* **197**, 899-912.
- Salehi, A., Delcroix, J.-D., Belichenko, P. V., Zhan, K., Wu, C., Valletta, J. S., Takimoto-Kimura, R., Kleschevnikov, A. M., Sambamurti, K., Chung, P. P. et al. (2006). Increased App expression in a mouse model of Down's syndrome disrupts NGF transport and causes cholinergic neuron degeneration. *Neuron* **51**, 29-42.
- Salehi, A., Faizi, M., Colas, D., Valletta, J., Laguna, J., Takimoto-Kimura, R., Kleschevnikov, A., Wagner, S. L., Aisen, P., Shamloo, M. et al. (2009). Restoration of norepinephrine-modulated contextual memory in a mouse model of Down syndrome. *Sci. Transl. Med.* **1**, Tra17.
- Sanford, L. P., Kallapur, S., Ormsby, I. and Doetschman, T. (2001). Influence of genetic background on knockout mouse phenotypes. *Methods Mol. Biol.* **158**, 217-225.
- Sansevero, G., Begenisic, T., Mainardi, M. and Sale, A. (2016). Experience-dependent reduction of soluble beta-amyloid oligomers and rescue of cognitive abilities in middle-age Ts65Dn mice, a model of Down syndrome. *Exp. Neurol.* **283**, 49-56.

- Sebrié, C., Chabert, C., Ledru, A., Guedj, F., Po, C., Smith, D. J., Rubin, E., Rivals, I., Beloeil, J.-C., Gillet, B. et al. (2008). Increased Dosage of DYRK1A and Brain Volumetric Alterations in a YAC Model of Partial Trisomy 21. *Anat. Rec. (Hoboken)* **291**, 254-262.
- Sherman, S. L., Allen, E. G., Bean, L. H. and Freeman, S. B. (2007). Epidemiology of Down syndrome. *Genet. Retard. Dev. Disabil. Res. Rev.* **13**, 221-227.
- Shichiri, M., Yoshida, Y., Ishida, N., Hagihara, Y., Iwahashi, H., Tamai, H. and Niki, E. (2011). alpha-Tocopherol suppresses lipid peroxidation and behavioral and cognitive impairments in the Ts65Dn mouse model of Down syndrome. *Free Radic. Biol. Med.* **50**, 1801-1811.
- Sittig, L. J., Carbonetto, P., Engel, K. A., Krauss, K. S., Barrios-Camacho, C. M. and Palmer, A. A. (2016). Genetic Background Limits Generalizability of Genotype-Phenotype Relationships. *Neuron* **91**, 1253-1259.
- Smith, D. J. and Rubin, E. M. (1997). Functional screening and complex traits: human 21q22.2 sequences affecting learning in mice. *Hum. Mol. Genet.* **6**, 1729-1733.
- Smith, D. J., Zhu, Y. W., Zhang, J. L., Cheng, J. F. and Rubin, E. M. (1995). Construction of a panel of transgenic mice containing a contiguous 2-MB set of YAC/P1 clones from human chromosome 21Q22.2. *Genomics* **27**, 425-434.
- Song, W.-J., Song, E.-A. C., Choi, S.-H., Baik, H.-H., Jin, B. K., Kim, J. H. and Chung, S.-H. (2013). Dyrk1A-mediated phosphorylation of RCAN1 promotes the formation of insoluble RCAN1 aggregates. *Neurosci. Lett.* **554**, 135-140.
- Song, W.-J., Song, E.-A. C., Jung, M.-S., Choi, S.-H., Baik, H.-H., Jin, B. K., Kim, J. H. and Chung, S.-H. (2015). Phosphorylation and inactivation of glycogen synthase kinase 3 $\beta$  (GSK3 $\beta$ ) by dual-specificity tyrosine phosphorylation-regulated kinase 1A (Dyrk1A). *J. Biol. Chem.* **290**, 2321-2333.
- Souchet, B., Guedj, F., Sahún, I., Duchon, A., Daubigney, F., Badel, A., Yanagawa, Y., Barallobre, M. J., Dierssen, M., Yu, E. et al. (2014). Excitation/inhibition balance and learning are modified by Dyrk1a gene dosage. *Neurobiol. Dis.* **69**, 65-75.
- Souchet, B., Guedj, F., Penke-Verdier, Z., Daubigney, F., Duchon, A., Herault, Y., Bizot, J.-C., Janel, N., Créau, N., Delatour, B. et al. (2015). Pharmacological correction of excitation/inhibition imbalance in Down syndrome mouse models. *Front. Behav. Neurosci.* **9**, 267.
- Stagni, F., Magistretti, J., Guidi, S., Ciani, E., Mangano, C., Calzà, L. and Bartesaghi, R. (2013). Pharmacotherapy with fluoxetine restores functional connectivity from the dentate gyrus to field CA3 in the Ts65Dn mouse model of down syndrome. *PLoS ONE* **8**, e61689.
- Stagni, F., Giacomini, A., Guidi, S., Ciani, E., Ragazzi, E., Filonzi, M., De lasio, R., Rimondini, R. and Bartesaghi, R. (2015). Long-term effects of neonatal treatment with fluoxetine on cognitive performance in Ts65Dn mice. *Neurobiol. Dis.* **74**, 204-218.
- Stagni, F., Giacomini, A., Emili, M., Trazzi, S., Guidi, S., Sassi, M., Ciani, E., Rimondini, R. and Bartesaghi, R. (2016). Short- and long-term effects of neonatal pharmacotherapy with epigallocatechin-3-gallate on hippocampal development in the Ts65Dn mouse model of Down syndrome. *Neuroscience* **333**, 277-301.
- Stanford, S. C. (2007). The Open Field Test: reinventing the wheel. *J. Psychopharmacol.* **21**, 134-135.
- Starbuck, J. M., Dutka, T., Ratliff, T. S., Reeves, R. H. and Richtsmeier, J. T. (2014). Overlapping trisomies for human chromosome 21 orthologs produce similar effects on skull and brain morphology of Dp(16)1Yey and Ts65Dn mice. *Am. J. Med. Genet. A* **164**, 1981-1990.
- Summers, A. M., Langlois, S., Wyatt, P., Wilson, R. D. and Canada, S. O. O. A. G. O. (2007). Prenatal screening for fetal aneuploidy. *J. Obstet. Gynaecol. Can.* **29**, 146-179.
- Sussan, T. E., Yang, A., Li, F., Ostrowski, M. C. and Reeves, R. H. (2008). Trisomy represses Apc(Min)-mediated tumours in mouse models of Down's syndrome. *Nature* **451**, 73-75.
- Taipale, J. and Beachy, P. A. (2001). The Hedgehog and Wnt signaling pathways in cancer. *Nature* **411**, 349-354.
- Tejedor, F., Zhu, X. R., Kaltenebach, E., Ackermann, A., Baumann, A., Canal, I., Heisenberg, M., Fischbach, K. F. and Pongs, O. (1995). minibrain: a new protein kinase family involved in postembryonic neurogenesis in Drosophila. *Neuron* **14**, 287-301.
- Thomazeau, A., Lassalle, O., Iafraji, J., Souchet, B., Guedj, F., Janel, N., Chavis, P., Delabar, J. and Manzoni, O. J. (2014). Prefrontal deficits in a murine model overexpressing the down syndrome candidate gene dyrk1a. *J. Neurosci.* **34**, 1138-1147.
- Threadgill, D. W., Dlugosz, A. A., Hansen, L. A., Tennenbaum, T., Lichti, U., Yee, D., Lamantia, C., Mouton, T., Herrup, K. and Harris, R. C. (1995). Targeted disruption of mouse EGF receptor: effect of genetic background on mutant phenotype. *Science* **269**, 230-234.
- Trazzi, S., Mitrugno, V. M., Valli, E., Fuchs, C., Rizzi, S., Guidi, S., Perini, G., Bartesaghi, R. and Ciani, E. (2011). APP-dependent up-regulation of Ptch1 underlies proliferation impairment of neural precursors in Down syndrome. *Hum. Mol. Genet.* **20**, 1560-1573.
- Trazzi, S., Fuchs, C., Valli, E., Perini, G., Bartesaghi, R. and Ciani, E. (2013). The amyloid precursor protein (APP) triplicated gene impairs neuronal precursor differentiation and neurite development through two different domains in the Ts65Dn mouse model for Down syndrome. *J. Biol. Chem.* **288**, 20817-20829.
- Troca-Marin, J. A., Alves-Sampaio, A. and Montesinos, M. L. (2011). An increase in basal BDNF provokes hyperactivation of the Akt-mammalian target of rapamycin pathway and deregulation of local dendritic translation in a mouse model of Down's syndrome. *J. Neurosci.* **31**, 9445-9455.
- Troca-Marin, J. A., Casañas, J. J., Benito, I. and Montesinos, M. L. (2014). The Akt-mTOR pathway in Down's syndrome: the potential use of rapamycin/rapalogs for treating cognitive deficits. *CNS Neurol. Disord. Drug Targets* **13**, 34-40.
- Tybulewicz, V. L. J. and Fisher, E. M. (2006). New techniques to understand chromosome dosage: mouse models of aneuploidy. *Hum. Mol. Genet.* **15**, R103-R109.
- Velazquez, R., Ash, J. A., Powers, B. E., Kelley, C. M., Strawderman, M., Luscher, Z. I., Ginsberg, S. D., Mufson, E. J. and Strupp, B. J. (2013). Maternal choline supplementation improves spatial learning and adult hippocampal neurogenesis in the Ts65Dn mouse model of Down syndrome. *Neurobiol. Dis.* **58**, 92-101.
- Vicari, S. (2006). Motor development and neuropsychological patterns in persons with Down syndrome. *Behav. Genet.* **36**, 355-364.
- Vidal, V., García, S., Martínez, P., Corrales, A., Flórez, J., Rueda, N., Sharma, A. and Martínez-Cué, C. (2012). Lack of behavioral and cognitive effects of chronic ethosuximide and gabapentin treatment in the Ts65Dn mouse model of Down syndrome. *Neuroscience* **220**, 158-168.
- Villardell, M., Rasche, A., Thormann, A., Maschke-Dutz, E., Pérez-Jurado, L. A., Lehrach, H. and Herwig, R. (2011). Meta-analysis of heterogeneous Down Syndrome data reveals consistent genome-wide dosage effects related to neurological processes. *BMC Genomics* **12**, 229.
- Villar, A. J., Belichenko, P. V., Gillespie, A. M., Kozy, H. M., Mobley, W. C. and Epstein, C. J. (2005). Identification and characterization of a new Down syndrome model, Ts Rb(12.17(16))2Cje, resulting from a spontaneous Robertsonian fusion between T(17(16))65Dn and mouse Chromosome 12. *Mamm. Genome* **16**, 79-90.
- Webb, S., Anderson, R. H., Lamers, W. H. and Brown, N. A. (1999). Mechanisms of deficient cardiac septation in the mouse with trisomy 16. *Circ. Res.* **84**, 897-905.
- Wiseman, F. K., Al-Janabi, T., Hardy, J., Karmiloff-Smith, A., Nizetic, D., Tybulewicz, V. L. J., Fisher, E. M. C. and Strydom, A. (2015). A genetic cause of Alzheimer disease: mechanistic insights from Down syndrome. *Nat. Rev. Neurosci.* **16**, 564-574.
- Witton, J., Padmashri, R., Zinyuk, L. E., Popov, V. I., Kraev, I., Line, S. J., Jensen, T. P., Tedoldi, A., Cummings, D. M., Tybulewicz, V. L. J. et al. (2015). Hippocampal circuit dysfunction in the Tc1 mouse model of Down syndrome. *Nat. Neurosci.* **18**, 1291-1298.
- Xie, W., Ramakrishna, N., Wieraszko, A. and Hwang, Y.-W. (2008). Promotion of neuronal plasticity by (-)-epigallocatechin-3-gallate. *Neurochem. Res.* **33**, 776-783.
- Xing, L., Salas, M., Zhang, H., Gittler, J., Ludwig, T., Lin, C.-S., Murty, V. V., Silverman, W., Arancio, O. and Tycko, B. (2013). Creation and characterization of BAC-transgenic mice with physiological overexpression of epitope-tagged RCAN1 (DSCR1). *Mamm. Genome* **24**, 30-43.
- Xing, Z., Li, Y., Pao, A., Bennett, A. S., Tycko, B., Mobley, W. C. and Yu, Y. E. (2016). Mouse-based genetic modeling and analysis of Down syndrome. *Br. Med. Bull.* **120**, 111-122.
- Yang, A., Currier, D., Poitras, J. L. and Reeves, R. H. (2016). Increased skin tumor incidence and keratinocyte hyper-proliferation in a mouse model of Down syndrome. *PLoS ONE* **11**, e0146570.
- Yu, Y. and Bradley, A. (2001). Engineering chromosomal rearrangements in mice. *Nat. Rev. Genet.* **2**, 780-790.
- Yu, T., Clapcote, S. J., Li, Z., Liu, C., Pao, A., Bechard, A. R., Carattini-Rivera, S., Matsui, S.-I., Roder, J. C., Baldini, A. et al. (2010a). Deficiencies in the region syntenic to human 21q22.3 cause cognitive deficits in mice. *Mamm. Genome* **21**, 258-267.
- Yu, T., Li, Z., Jia, Z., Clapcote, S. J., Liu, C., Li, S., Asrar, S., Pao, A., Chen, R., Fan, N. et al. (2010b). A mouse model of Down syndrome trisomic for all human chromosome 21 syntenic regions. *Hum. Mol. Genet.* **19**, 2780-2791.
- Yu, T., Liu, C. H., Belichenko, P., Clapcote, S. J., Li, S. M., Pao, A. N., Kleschevnikov, A., Bechard, A. R., Asrar, S., Chen, R. Q. et al. (2010c). Effects of individual segmental trisomies of human chromosome 21 syntenic regions on hippocampal long-term potentiation and cognitive behaviors in mice. *Brain Res.* **1366**, 162-171.
- Zhang, L., Meng, K., Jiang, X., Liu, C., Pao, A., Belichenko, P. V., Kleschevnikov, A. M., Josselyn, S., Liang, P., Ye, P. et al. (2014). Human chromosome 21 orthologous region on mouse chromosome 17 is a major determinant of Down syndrome-related developmental cognitive deficits. *Hum. Mol. Genet.* **23**, 578-589.