

Investigation of the Geochemical Preservation of ca. 3.0 Ga Permineralized and Encapsulated Microfossils by Nanoscale Secondary Ion Mass Spectrometry

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 permineralized and encapsulated microfossils by Nanoscale
 secondary ion mass spectrometry.

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5 Running head: Preservation of encapsulated Archean microfossils.

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24 Abstract

25 Observations of Archean organic-walled microfossils suggest that their fossilization took place 26 through both encapsulation by silica and permineralization. In this study, we have investigated 27 microfossils from the ca. 3.0 Gyr-old Farrel Quarzite (Pilbara, Western Australia) using transmitted light microscopy, scanning electron microscopy, Raman microspectrometry and 28 NanoSIMS ion microprobe analyses. In contrast to previous statement, we demonstrated that 29 permineralized microfossils were not characterized by the micrometric spatial relationships 30 between Si and C-N as observed in thin sections. Permineralized microfossils are composed of 31 carbonaceous globules that did not survive the acid treatment whereas, encapsulated microfossils 32 33 were characterized thanks to their resistance to the acid maceration procedure. We have also investigated the microscale relationship between the ${}^{12}C^{14}N^{-}$ and ${}^{12}C_{2}^{-}$ ion emission as a proxy of 34 35 the N/C atomic ratio in both permineralized and encapsulated microfossils. After considering any potential matrix and microtopography effects, we demonstrate that the encapsulated microfossils 36 37 exhibit the highest level of geochemical preservation. This finding shows that the chemical heterogeneity of the microfossils, observed at a spatial resolution of few hundreds of 38 micrometers, can be related to fossilization processes. 39

40

41 **1. Introduction**

Microfossil-like structures have been reported in numerous Archean rocks (e.g., Walsh, 1992; 42 Schopf, 1993; Javaux et al., 2010; Sugitani et al., 2010). As a result of thermal alteration, 43 44 however, their morphological features and their geochemical composition had been often 45 severely modified, making the univocal identification of microorganisms and associated metabolism difficult. Microorganisms are generally thought to be fossilized through 46 permineralization resulting from the õearly infiltration and permeation of tissues by mineral-47 charged waterö (Schopf 1975). Consequently, the organic remnants are progressively replaced by 48 49 silica or carbonates during mineralization. However, the presence of organic-walled microfossils 50 in some Archean rocks (Grey and Sugitani, 2009; Javaux et al. 2010; Sugitani et al., 2015) points to the existence of other mechanisms of fossilization, such as encapsulation of microorganisms 51 through nucleation of adjacent mineral crystals for example (Rainey and Jones, 2010). Through 52 investigation of mineralization of microbial mats from Icelandic hot springs, Konhauser and 53 Ferris (1996) proposed that encapsulation favors morphological and geochemical preservation of 54 microorganisms. This is supported by experimental silicification of modern microorganisms, 55 which shows that the negative effect on geochemical preservation caused by thermal alteration 56 can be counter-balanced by encapsulation (Picard et al., 2015). However, encapsulation has not 57 yet been directly documented in Archean rocks despite its great potential for preservation of 58 microorganisms. 59

Bulk N/C atomic ratio has been classically used as a proxy to characterize the preservation status of organic matter (Watanabe et al., 1997; Beaumont and Robert, 1999). However, this bulk geochemical approach neglects potential heterogeneities in preservation among different organic remnants. Recent technological developments, notably in the field of Secondary Ion Mass Spectrometry (SIMS), have allowed *in situ* elemental and isotopic

investigations of putative microfossils at the micro- to nanoscale (Rasmussen et al., 2008; Oehler 65 et al., 2009; House et al., 2013). Notably, it has been shown that the ${}^{12}C^{14}N^{-12}C_{2}^{-1}$ molecular ionic 66 ratio is strongly correlated with bulk N/C atomic ratio (Thomen et al., 2014; Alleon et al., 2015), 67 68 opening up the possibility to evaluate the geochemical preservation of Archean microfossils at the micrometer scale. In pioneering studies, Oehler et al. (2009; 2010) used NanoSIMS analyses 69 70 to calculate in situ N/C atomic ratios ranging from ca. 0.0125 to 0.05 for Archean spheroid microfossils from 3.0 Gyr-old cherts. However, the possible effects of microtopography on the 71 ${}^{12}C^{14}N^{12}C_2$ molecular ratios determined for the microfossils studied by Oehler et al. (2009; 72 73 2010) have not been thoroughly evaluated, and it is known that microtopography may induce relatively large changes in ${}^{12}C^{14}N^{7/12}C_2^{-7}$ ratios even though precise quantifications are still 74 75 incomplete (e.g., Thomen et al., 2014; Alleon et al., 2015). These potential analytical pitfalls 76 have to be addressed to further evaluate the significance of the relatively high in situ N/C atomic 77 values determined by Oehler et al. (2009; 2010) for Archean microfossils. In this respect, a recent study has highlighted that silicification promoted the exceptional geochemical preservation of 78 organic microfossils in the 1.88 Gyr-old Gunflint cherts that have N/C atomic ratios up to ca. 79 0.25-0.30 (Alleon et al., 2016), which is commensurable with the N/C ratios of modern 80 cyanobacteria and is, by far, higher than the N/C ratios determined by Oehler et al. (2009, 2010). 81

In this study, our purposes are (i) to provide a procedure to determine the preservation status of organic microfossils by studying the relationship between the emissions of the ${}^{12}C_{2}^{-}$ and ${}^{12}C^{14}N^{-}$ molecular ions in pure organic standards, kerogens and microfossils from both thin sections and acid maceration residues and (ii) to we discuss the effect of the process of fossilization, i.e. permineralization vs. encapsulation, on the geochemical preservation of microfossils from the 3.0 Gyr-old Farrel Quartzite. 88

89 2. Material and Methods

90 2.1. Sample locality

91

A black chert sample was collected from the ca. 3.0 Ga Farrel Quartzite at the Mount Grant 92 locality in the Goldsworthy greenstone belt, in the Pilbara Craton in Western Australia. The Farrel 93 Quartzite is composed of a clastic formation up to 80 m thick containing fine- to very coarse-94 95 grained sandstone, including quartzite with minor conglomerate, mafic to ultra-mafic 96 volcanoclastic layers, evaporite beds and black chert layers (Sugitani et al., 2007). This unit underwent greenschist facies metamorphism and was pervasively silicified. The ca. 30 cm thick 97 microfossil-bearing black chert occurs in the uppermost part of the Farrel Quartzite and is closely 98 99 associated with evaporite beds.

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101 **2.2. Analyses**

102 2.2.1. Sample preparations and microscopy

103

Transmission light microscopy (TLM) observations and NanoSIMS analyses were carried out on both rock thick sections (thickness of *ca*. 50 m) and isolated kerogen, whereas Scanning Electron Microscope (SEM) observations were only performed on the kerogen. Kerogen isolation was performed on about 200 g of rock through successive demineralization using HF-HCl (Derenne et al., 2008). Then, a few mg of kerogen were deposited on a microscope glass slide for TLM, SEM and NanoSIMS investigations. Carbonaceous microfossils were first observed using TLM in order to define targets of interest. Then, glass slides were directly gold coated (20 nm thick) for SEM Energy Dispersive X-ray Spectroscopy (EDS) analysis and imaging using a
TESCAN VEGA II at the French National Museum of Natural History (MNHN) with an
accelerating voltage of 15 kV.

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115 2.2.2. Raman microspectrometry

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Raman spectra were obtained using a Renishaw InVIA microspectrometer, equipped with a 532 117 nm argon laser. The laser was focused on the sample by using a DMLM Leica microscope with a 118 50×·objective. The spectrometer was first calibrated with a silicon standard before the analytical 119 session. For each target, we determined the Raman shift intensity in the spectral window from 120 121 1000 to 1900 cm⁻¹ including the first-order disorder carbon (D) and graphite (G) bands. A laser power below 1 mW was used to prevent any thermal alteration during the spectra acquisition. 122 Finally, spectra acquisition was achieved after three successive iterations using a time exposure 123 of 40 seconds. 124

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126 2.2.3. Nanoscale secondary ion mass spectrometry

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Standards and microfossils were analyzed using the CAMECA NanoSIMS 50 at the MNHN. Before measurements, pre-sputtering is required (i) to avoid surficial contamination and (ii) to achieve the saturation fluence of implemented Cesium (Cs⁺) in order to obtain constant secondary ion count rates and then, a constant ${}^{12}C{}^{14}N{}^{-/12}C{}_{2}{}^{-}$ ionic ratio (Fig. 1). Hence, Cs⁺ was implanted using a 200 pA primary current (300 µm aperture diaphragm) on 50×50 to 75×75 µm² areas, depending of the size of each target. Analyses were then carried out using a 5 pA primary current (150 µm aperture diaphragm) on smaller areas to avoid pre-sputtering edge artifacts. Secondary

molecular ions and species of ${}^{12}C_2^{-}$, ${}^{12}C^{14}N^{-}$ and ${}^{28}Si^{-}$ were collected simultaneously in electron 135 multipliers. NanoSIMS raw data were corrected for a 44 ns dead time on each electron multiplier 136 and were processed using the Limage software (developed by L. Nittler, Carnegie Institution, 137 138 Washington DC, USA). The external reproducibility was determined through multiple measurements of the emissions of the ${}^{12}C^{14}N^{-}$ and ${}^{12}C_{2}^{-}$ molecules on a coal standard used in 139 140 Thomen et al. (2014). A second NanoSIMS session was dedicated to the analyses of a blank (polycarbonate filter), of pure organic standards (resin and tryptophan) and of a type III kerogen 141 (land plant-derived carbonaceous matter). These pure standards and the type III kerogen 142 correspond to the standards previously used in Alleon et al. (2015). Hence, Cs+ was implanted 143 using a 400 pA primary current (150 μ m aperture diaphragm) on 45×45 μ m² areas. Analyses 144 were then carried out using a 1 pA primary current (150 µm aperture diaphragm) on smaller areas 145 to avoid pre-sputtering edge artifacts. 146

147

148 **2.2.4. Statistics and errors**

149

150 Correlations between the ${}^{12}C_{2}^{-}$ and ${}^{12}C^{14}N^{-}$ and ${}^{28}Si^{-}$ ion emissions were tested using Spearman's 151 rank correlation. A *p*-value inferior to 0.05 is indicative of a significant correlation. In the 152 presence of a significant spatial relationship between the emissions of ions, linear regressions 153 were performed to calculate the value of the slope and its associated standard error (1 _{reg}) 154 following:

155

156 1 =
$$\sum yi - \hat{y}i / (-2) / \sum xi - x$$
 (1)

where yi is the emission of the ${}^{12}C^{14}N^{-}$ ion measured by NanoSIMS, i is the emission of the 159 ${}^{12}C^{14}N^{-}$ ion determined by linear regression, xi is the emission of the ${}^{12}C_{2}^{-}$ ion measured by 160 NanoSIMS, \overline{x} the average value of the emissions of the ${}^{12}C_{2}^{-}$ ion and where n is the number of 161 ROIs.

162

163 The external reproducibility was determined by determining the slope of the regression line 164 between the ${}^{12}C_{2}^{-}$ and ${}^{12}C^{14}N^{-}$ ion emissions of a coal standard (n = 7). Then, the standard error 165 of the mean slope $\tilde{0}$ $\tilde{0}$ (1 _{rep}) was calculated.

166

167 Finally, the total error (1_{tot}) was determined as follow:

168 1 = $\overline{1 + 1}$ (2)

169

170 **3. Results**

171 **3.1.** Carbonaceous microfossils in thin section and kerogen

172

A morphological diversity of microfossils was observed in thin section, with assemblages of lenticular (formerly described as spindle-like=*ca*. 20-40 μ m=Sugitani et al., 2007=Grey and Sugitani, 2009=Sugitani et al., 2009), film-like (>100 μ m), and spheroidal (mainly <15 μ m) microfossils occurring either as isolated specimens or as clusters (Fig. 2). In both spheroids and lenticular structures analyzed in thin sections, the ¹²C₂⁻ and ¹²C¹⁴N⁻ ion emissions (Figs. 2a, b) are found within the siliceous matrix. In the film-like microstructure (Fig. 2c), the ¹²C₂⁻ and 179 ${}^{12}C^{14}N^{-}$ are emitted with almost no emission of ${}^{28}Si^{-}$. This observation illustrates the fact that this 180 microstructure was encapsulated by the siliceous matrix.

181 Characterizing microfossils from thin sections using NanoSIMS implies that the analyzed targets 182 occur at the very surface of the sample because the intensity of the primary beam cannot sputter more than a few atomic layers in depth. Therefore, the amount of microfossil targets in thin 183 sections is limited. On the contrary, the kerogen fraction obtained by HF-HCl maceration of the 184 fossil-bearing black cherts contains some microfossils morphologically equivalents to those in 185 thin section (Grey and Sugitani, 2009). Although spheroids identified in the thin section were not 186 187 found in the kerogen residue, lenticular and film-like microfossils were also observed in the kerogen fraction (Fig. 3). These lenticular and film-like microfossils are characterized by Raman 188 line shape (Fig. 4), which are consistent with previous Raman spectra determined on microfossils 189 190 from thin section (Sugitani et al., 2007)

191

192 **3.2 NanoSIMS quantitative investigation**

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194 The N/C atomic ratio has been classically used to assess the preservation status of ancient organic matter (Watanabe et al., 1997=Beaumont and Robert, 1999). Using NanoSIMS analysis, the 195 ${}^{12}C^{14}N^{12}C_2^{-1}$ ionic ratio has been regarded as a proxy of the N/C atomic ratio for silica-free 196 197 kerogens or pure organic standards (Thomen et al., 2014=Alleon et al., 2015). However, in case of microfossils, two additional analytical artefacts should be considered (1) matrix effects 198 199 (chemical heterogeneity) linked to the occurrence of silicate minerals and (2) microtopographic effects yielding a difference in the emissivity of the ${}^{12}C_2^{-1}$ and ${}^{12}C_2^{-14}N^{-1}$ ions. Owing to the imaging 200 capability of the NanoSIMS, the ${}^{12}C_2^{-1}$ and ${}^{12}C^{14}N^{-1}$ variations have been recorded at a high spatial 201 resolution. In the following, we demonstrate that the spatial variation between ${}^{12}C_2^{-1}$ and ${}^{12}C^{14}N^{-1}$ is 202

linear and that its corresponding slope $\tilde{o}\alpha\tilde{o}$ is correlated with the N/C atomic ratio. However, this linear variation between ${}^{12}C_{2}^{-1}$ and ${}^{12}C^{14}N^{-1}$ shows a non-zero intercept \ddot{o} \ddot{o} , possibly related to the sample surface microtopography. No relation was found between and α , justifying in turn, the use of α to record the relative variations of the N/C ratio. In addition, no measurable variation in the emissivity of the ${}^{12}C_{2}^{-1}$ and ${}^{12}C^{14}N^{-1}$ ions has been detected in the presence of silicate minerals, avoiding measurable matrix effects on α .

209

210 **3. 2..1** The slope õ ö, a record of the N/C atomic ratio

211

Emissions of ${}^{12}C_2^{-1}$ and ${}^{12}C^{14}N^{-1}$ in resin, tryptophan and a type III kerogen are systematically 212 correlated (Figs. 5a and 5b=Table 1). Although both emissions converge towards 0 for lower ion 213 214 counting rates, the linear regression calculated on the whole range of emissions yields a non-zero intercept õ ö. This relationship is characterized by a slope õ ö. In Fig. 5c, the slopes of pure 215 kerogen and standards are reported versus bulk N/C atomic ratio. Note that for a N-free sample 216 (polycarbonate filter) no relationship between the emissions of the ${}^{12}C_2^{-1}$ and ${}^{12}C^{14}N^{-1}$ ions is found 217 (Table 1). Hence, a significantly linear relationship between the ${}^{12}C_2^{-1}$ and ${}^{12}C_2^{-14}N^{-1}$ ion emissions is 218 the preliminary condition in order to define a slope \tilde{o} of that can be used to record the N/C atomic 219 ratio. 220

221

222 **3.2.2.Matrix effect**

223

Oehler et al. (2009) defined a matrix effect in their NanoSIMS measurements as the enhancement of the ${}^{28}Si^{-}$ and ${}^{16}O^{-}$ ion emissions when these ions are closely and spatially associated with carbon. Such a matrix effect may be linked to a higher conductibility of Si associated with carbonaceous globules compared to that of Si in the surrounding siliceous minerals. In microfossils from chert thick section (Fig. 6a), no relationship between ²⁸Si⁻ on the one hand and ¹²C₂⁻ (Fig. 6b) and ¹²C¹⁴N⁻ (Fig. 6c) ion emissions on the other hand has been found. Hence, is not affected by the occurrence of Si.

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232 **3.2.3. Microtopography**

233

To investigate the effect of microtopography, we have compared the ${}^{12}C_{2}^{-}$ and ${}^{12}C^{14}N^{-}$ ion emissions on two selected flat and non-flat (microtopograhic features between 1 to 10µm) areas from a chemically homogeneous resin standard (Fig. 7a). Fig. 7, it can be seen that microtopography does not cause measurable shift in (Fig. 7b=flat area: = 0.52 ± 0.05 =non flat area = 0.56 ± 0.02).

Fig. 7c shows that the value of the ${}^{12}C{}^{14}N{}^{-/12}C{}_2{}^{-}$ ratio is not affected by small micrometric scale 239 microtopography up to 2 μ m. Although a 10 μ m of topography can enhance the ionic ${}^{12}C^{14}N^{-}/{}^{12}C_{2}^{-}$ 240 ratio by a factor of up to 4, the slope of the correlated variations between ${}^{12}C^{14}N^{-}$ and ${}^{12}C_{2}^{-}$ is only 241 related to the N/C atomic ratio of the sample. Such a bias in the determination of the ${}^{12}C^{14}N^{-}/{}^{12}C_{2}^{-}$ 242 ratio is also related to the value of the non-zero interceptõ ö, which seems to rise through 243 enhanced microtopography (Figs. 7b and c). In contrast to the ${}^{12}C^{14}N^{-}/{}^{12}C_{2}^{-}$ ratio, the slope is 244 constant in topographic domain covering 1 to 10 micrometers (Figure 7c). ${}^{12}C^{14}N^{-/12}C_2^{-1}$. Note 245 246 that due to the size of the presently studied microfossils and the fact that microfossil edges were not considered, the microtopographic features cannot exceed a few µm. 247

248 Consequently, the matrix and microtopograhic effects do not bias the use of the slope $\tilde{\alpha}\alpha$ ö, as a 249 record of the N/C atomic ratio.

250

251 **4. Discussion**

252 Evidence for the permineralization of a part of the microfossils from the Farrel Quartzite was previously suggested by Oehler et al. (2009) owing to the co-emissions of the Si⁻, C₂⁻ and 253 CN^{-} ions in microfossils from thin section. Here, ²⁸Si⁻ and ¹²C₂ ions on the one hand and ²⁸Si⁻ and 254 ${}^{12}C^{14}N^{-1}$ ions on the other hand, were not spatially associated at the pixel scale (Fig. 6). Such 255 findings may echo results observed in the 3.4 Gyr-old Strelley Pool Formation, in which Lepot et 256 257 al (2013) observed lenticular microfossils composed of carbonaceous globules that were interpreted as degradation by-products of Archean microorganisms. However, in the present 258 study, no 3D carbonaceous globules were observed in the isolated kerogen. Since they did not 259 survive the acid treatment, they must be not considered as encapsulated but rather as 260 permineralized. 261

262

In contrast to carbonaceous globules, carbonaceous microfossils were recovered in the acid maceration residue. Among these microfossils, one example of an exceptional morphological preservation of a lenticular microfossil is shown. Classically, in the literature, lenticular microfossils exhibit two kinds of flange-like appendages situated either in the equatorial plane or at the apical part of the vesicle body (Sugitani et al., 2009=House et al., 2013). Here, the flange-like appendage was situated at the apical part of the vesicle body. Lenticular but also film-like microfossils consist almost entirely of organic matter, suggesting that they are organic-walled microfossils removed from the silica matrix by the HF treatment. In turn, this result implies the preservation of some organic-walled microfossils by encapsulation rather than by permineralization. These organic-walled microfossils are characterized by equivalent Raman line shape (Fig. 4), corresponding to advanced carbonization/greenschist facies metamorphism in silicified cherts (Delarue et al., 2016). Raman characteristics of the microfossils are then consistent with the thermal history of the Farrel Quartzite cherts (Sugitani et al., 2007) revealing, in turn, their syngenecity.

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The slope $\tilde{0}$ ö parameter for both permineralized and encapsulated microfossils was calculated as a proxy of the *in situ* N/C atomic ratio (Fig. 8=Table 2). The correlation between ${}^{12}C_{2}^{-1}$ and ${}^{12}C^{14}N^{-1}$ is statistically significant for eleven out of the fourteen analyzed microfossils (Table 2), which will be considered in the following discussion.

In the permineralized lenticular and spheroid microfossils, ranges from 0.03 to 0.19 282 whereas it ranges from 0.05 to 1.84 in the encapsulated lenticular and film-like ones (Table 2). 283 First of all, these data suggest that there is an unexpected geochemical heterogeneity among the 284 microfossils preserved in the 3.0 Gyr-old Farrel Quartzite. Most of the encapsulated microfossils 285 are characterized by a greater compared to permineralized ones (Fig. 8=Table 2). This indicates 286 that encapsulated microfossils present higher geochemical preservation level than the 287 288 permineralized ones, and that the mode of fossilization may a key controlling factor in the geochemical heterogeneity in the Farrel Quartzite carbonaceous matter. Finally, this difference in 289 290 the extent of geochemical preservation between permineralized and encapsulated microfossils is consistent with observations made on modern microbial mat that show that microorganisms are 291 better preserved through encapsulation (Konhauser and Ferris, 1996). Focusing future studies on 292

Archean encapsulated microfossils may then provide the best geochemical evidence in the searchfor traces of terrestrial early-life.

295

296 Conclusion

In this study, we provide new lines of evidence supporting the partial fossilization of 297 carbonaceous microfossils through encapsulation in the ca. 3.0 billion-years-old cherts from the 298 Farrel Quarzite in the Pilbara craton, Western Australia. Encapsulated microfossils were observed 299 300 both in the thin section and in the kerogen fraction. Using the slope õ ö parameter relating the ${}^{12}C_2$ and ${}^{12}C_1{}^{14}N$ - NanoSIMS emissions as an index of geochemical preservation of the studied 301 microfossils, we demonstrate that encapsulated microfossils present higher level of geochemical 302 303 preservation than permineralized ones. Thus, the mechanism of fossilization of microorganisms may be considered as a key controlling factor in preserving geochemical heterogeneity. 304

305 Overall our results suggest that focusing *in situ* investigations on well preserved encapsulated 306 carbonaceous matter may provide the best chance to recover information on the earliest forms of 307 terrestrial life that are likely to be lost in bulk investigations.

308

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319

320 **Disclosure statement**

321 No competing financial interests exist.

322

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1 Figure captions

2

FIG.1. ¹²C¹⁴N⁻/¹²C₂⁻ ratio recorded as a function of the presputerring duration.
The grey area indicates the time window in which the saturation fluence was
achieved on the resin standard. Saturation fluence was systematically controlled
for each studied microfossils.

7

FIG.2. Transmitted light photomicrographs and NanoSIMS ion images (${}^{12}C_{2}$, 9 ${}^{12}C^{14}N^{-}$ and ${}^{28}Si^{-}$) of (**a**) a lenticular-like microfossil, (**b**) a spheroid cluster, and 10 (**c**) a film-like microfossil. The microfossils were all observed on thin section. On 11 each photomicrograph, the red dashed square outline indicates the area 12 investigated by NanoSIMS.

13

FIG.3. (a) Secondary electron and X-ray images of selected elements for a filmlike and a well-preserved lenticular-like isolated microfossils. (b) NanoSIMS ion images (¹²C₂⁻, ¹²C¹⁴N⁻ and ²⁸Si⁻) determined on the filament-like microfossil and on the two distinct ultrastructures of the well-preserved lenticular-like microfossil, namely the vesicle body and the flange-like appendage.

FIG.4. First-order Raman spectrum of the filaments and the lenticular
microfossils presented in Fig. 3.

22

FIG.5. (a) NanoSIMS ion images $({}^{12}C_2^{-1}$ and ${}^{12}C^{14}N^{-1}$) of type III kerogen, resin 23 and tryptophan. (b) Relationship between the emissions of ${}^{12}C_2^-$ and ${}^{12}C_2^{14}N^-$ ions 24 in each standard. Each point is related to a ROI (n=30) manually drawn on the 25 flattest part of the standards following the procedure described in Alleon et al. 26 (2015; see Table 1 for further information about the number and size of ROIs). (c) 27 Determination of the N/C atomic ratio by the use of the ${}^{12}C^{14}N^{-}/{}^{12}C_{2}^{-}$ ionic ratio 28 (Thomen et al., 2014; Alleon et al., 2015) and the slope "". Note that coal and 29 30 microfossils (session 1) were not studied in the same analytical session than the 3 31 standards (session 2) used to calibrate the use of the slope α as a proxy of the N/C 32 atomic ratio. Accordingly, this prevents any calculation of in situ N/C atomic 33 ratios of microfossils.. Also, note the linear relationship in each case, which evidences the absence of matrix effect on the determination of both ${}^{12}C^{14}N^{-/12}C_2^{-14}$ 34 ionic ratio and the slope " α ". 35

36

FIG.6. (a) NanoSIMS images $({}^{12}C_2^{-})$ of microfossils from thin section (presented in Fig. 2) displaying the ROIs used to study the potential relationship between (b) the ${}^{28}Si^{-}$ and the ${}^{12}C_2^{-}$ ions and (c) the ${}^{28}Si^{-}$ and the ${}^{12}C^4N^{-}$ ions. In order to constrain the spatial variability of the emissions of the ${}^{12}C_2^{-}$ and ${}^{12}C^{14}N^{-}$ ions,

41 ROIs were manually drawn around carbon/silica in permineralized microfossils.

42 For encapsulated microfossils, a grid was used (see Table 1 for further43 information on the number and the size of the ROIs).

44

FIG.7. Investigation of the effect of microtopography on the determination of the 45 slope "" and its error on a (a) ${}^{12}C_2$ and ${}^{12}C^{14}N$ ion flat (left images) and non-flat 46 (right images) areas from the resin standards. In the non-flat area, photonic 47 microtopography was created through extensive sputtering (1, 2 and 12 hours) 48 49 leading to the creation of micrometric scale microtopography estimated through microscopy (ca. 1, 2 and 10 μ m depth, respectively). The relationship between the 50 emissions of ${}^{12}C_2^{-1}$ and ${}^{12}C^{14}N^{-1}$ ions was estimated (i) on the above (i) flat area 51 52 using 30 ROIs (yellow circles of 0.8 µm diameter) as suggested by Alleon et al. (2015) and (ii) non-flat area using 121 ROIs (red grid, each square has a width of 53 2.2 μ m). (b) Relationships between the emissions of ${}^{12}C_2^{-1}$ and ${}^{12}C^{14}N^{-1}$ ions in the 54 flat and the non-flat areas. (c) Comparison between the ${}^{12}C^{14}N^{-/12}C_2^{-10}$ ionic ratio 55 calculated as in Alleon et al. (2015) and the slope "" " (this study) values across 56 microscale microtopography. The red area indicates the values of the slope " " 57 determined on the flat area. The intercept of the linear regression between the 58 emissions of ${}^{12}C_2^-$ and ${}^{12}C^{14}N^-$ ions is noted " β ". 59

FIG.8. (a) Relationships between the emissions of ${}^{12}C_2^-$ and ${}^{12}C^{14}N^-$ ions in microfossils from thin section (b) Relationship between the emissions of ${}^{12}C_2^-$ and ${}^{12}C^{14}N^-$ ions in encapsulated microfossils from the kerogen residue.

















Table 1. Bulk N/C atomic ratio (Thomen et al., 2014; Alleon et al., 2015) of studied standards, number and size of regions of interest (ROIs) used (i) to test the correlation between the emissions of the ${}^{12}C_2^{-1}$ and ${}^{12}C^{14}N^{-1}$ ions by using Spearman's rank correlation and (ii) to determine the slope õ ö on standards. To minimize the effect of microtopography, ROIs were selected in the flattest part of the standard as recommended by Alleon et al. (2015). In the specific case of the coal standard, our purpose was to measure the external reproducibility. Thus, a maximum of ROIs were selected to characterize the strict effect of analytical drift avoiding then to take into account twice the effect of microtopography during the analyses of microfossils Note that the polycarbonate filter was used as a nitrogen blank in which the ${}^{12}C_2^{-1}$ and ${}^{12}C^{14}N^{-1}$ ion emissions are not significantly correlated

Purpose	Sample	Bulk N/C atomic ratio	number of ROIs	ROIs diameter (µm)	Spearman <i>p</i> -value	±1 reg
Blank	Polycarbonate Filter	-	30	1.5	0.50	-
Calibration line	Type III kerogen	0.016	30	1.5	< 0.0001	0.13 ± 0.01
	Resin	0.053	30	0.8	< 0.0001	0.52 ± 0.05
	Tryptophan	0.182	30	1.5	< 0.0001	1.20 ± 0.13
External	Coal 1	0.0022	545	1.1	< 0.0001	0.11 ± 0.004
reproduciblity	Coal 2	0.0022	460	1.1	< 0.0001	0.10 ± 0.002
	Coal 3	0.0022	457	1.3	< 0.0001	0.09 ± 0.002
	Coal 4	0.0022	478	1.3	< 0.0001	0.09 ± 0.004
	Coal 5	0.0022	321	1.3	< 0.0001	0.13 ± 0.003
	Coal 6	0.0022	365	1.3	< 0.0001	0.09 ± 0.002
	Coal 7	0.0022	367	1.4	< 0.0001	0.08 ± 0.004

Table 2. Number and diameter of ROIs used on studied microfossils (microfossils presented in Figs; 2, 3 and 8 are indicated by the superscript a,b and c, respectively). Spearman's rank correlation, slope $\tilde{0}$ öand associated error (1 tot) determined through the linear relationship between the emissions of the ${}^{12}C_{2}^{-1}$ and ${}^{12}C^{14}N^{-1}$ ions in both permineralized and encapsulated microfossils from thin section and kerogen.

		Encapsulated (E)				
	Thin section (TS)	VS	number			
	VS	Permineralized	of	ROI diameter	Spearman	
Microfossil type	Kerogen (K)	(P)	ROIs	$(\mu m \pm SD)$	<i>p</i> -value	± 1 tot
Lenticular	TS	Р	25	0.63 ± 0.15	0.0013	$0.19\pm\ 0.05$
Lenticular ^{a,c}	TS	Р	49	0.66 ± 0.12	< 0.0001	0.06 ± 0.01
Lenticular	TS	Р	42	0.78 ± 0.20	0.0002	0.07 ± 0.02
Spheroid	TS	Р	52	0.66 ± 0.11	< 0.0001	$0.03\pm\ 0.01$
Spheroid	TS	Р	25	0.65 ± 0.40	0.14	-
Spheroid 1 ^{a,c}	TS	Р	23	0.94 ± 0.27	< 0.0001	0.07 ± 0.01
Spheroid 2	TS	Р	20	1.08 ± 0.52	0.01	0.11 ± 0.05
Spheroid 3	TS	Р	13	0.70 ± 0.38	0.11	-
Film	TS	Е	252	2.9	< 0.0001	1.48 ± 0.13
Filament ^{b,c}	K	E	146	2.4	< 0.0001	0.05 ± 0.00
Film	K	E	45	1.3	0.11	-
Film ^{a,c}	TS	E	195	2.2	< 0.0001	1.84 ± 0.14
Lenticular						
(vesicle) ^{0,c}	K	E	121	1.2	< 0.0001	0.83 ± 0.09
(flange) ^{b,c}	К	E	28	0.7	< 0.0001	1.12 ± 0.18