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▶ To cite this version:

C. Karachi, Chantal François. Role of the pedunculopontine nucleus in controlling gait and sleep in normal and parkinsonian monkeys. Journal of Neural Transmission, 2018, 125 (3), pp.471 - 483. 10.1007/s00702-017-1678-y . hal-01722908

HAL Id: hal-01722908 https://hal.sorbonne-universite.fr/hal-01722908

Submitted on 5 Mar 2018

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Role of the pedunculopontine nucleus in controlling gait and sleep in normal and parkinsonian monkeys

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Abstract

Patients with Parkinson's disease (PD) develop cardinal motor symptoms, including akinesia, rigidity, and tremor, that are alleviated by dopaminergic medication and/or subthalamic deep brain stimulation. Over the time course of the disease, gait and balance disorders worsen and become resistant to pharmacological and surgical treatments. These disorders generate debilitating motor symptoms leading to increased dependency, morbidity, and mortality. PD patients also experience sleep disturbance that raise the question of a common physiological basis. An extensive experimental and clinical body of work has highlighted the crucial role of the pedunculopontine nucleus (PPN) in the control of gait and sleep, and its potential major role in PD. Here, we summarise our investigations in the monkey PPN in the normal and parkinsonian states. We first examined the anatomy and connectivity of the PPN and the cuneiform nucleus which both belong to the mesencephalic locomotor region. Second, we conducted experiments to demonstrate the specific effects of PPN cholinergic lesions on locomotion in the normal and

parkinsonian monkey. Third, we aimed to understand how PPN cholinergic lesions impair sleep in parkinsonian monkeys. Our final goal was to develop a novel model of advanced PD with gait and sleep disorders. We believe that this monkey model, even if it does not attempt to reproduce the exact human disease with all its complexities, represents a good biomedical model to characterise locomotion and sleep in the context of PD.

Keywords

Parkinson's disease Pedunculopontine nucleus Gait disorders Sleep disorders Monkeys

Abbreviations

CuN	Cuneiform nucleus
DBS	Deep brain stimulation
FOG	Freezing of gait
MLR	Mesencephalic locomotor region
MPTP	1-Methyl-4-phenyl-1,2,3,6-tetrahydropyridine
PD	Parkinson's disease
PPN	Pedunculopontine nucleus
REM	Rapid eye movement

Introduction

Most patients with advanced Parkinson's disease (PD) develop gait and balance disorders which worsen with time and become resistant to dopaminergic medication. These gait and postural disorders, the so-called axial symptoms, are characterised by freezing of gait (FOG) and balance deficits. Falls, in particular, have been identified as a major feature in the course of PD (Kempster et al. 2010), because they are the source of serious disability leading to dependency, institutionalisation, and poor quality of life. These resistant gait disorders induce high morbidity mainly related to hip fractures and head injuries, and significantly increased mortality. In this context, the poor alleviation of axial symptoms with

dopaminergic medication and deep brain stimulation (DBS) of the subthalamic nucleus or internal pallidum strongly suggests a dysfunction of non-dopaminergic brain networks. An extensive experimental and clinical body of work has highlighted the crucial role of the pedunculopontine nucleus (PPN), whose cholinergic neurons partially degenerate in advanced PD (Hirsch et al. 1987; Zweig et al. 1989; Rinne et al. 2008; Karachi et al. 2010).

The PPN appears to be a complex gait generator of the brainstem linking cortex, basal ganglia, and the spinal cord. Given its anatomical location and its partial neuronal loss in specific neurodegenerative disorders that impair gait, some clinical teams proposed low-frequency PPN-DBS in PD patients with FOG and falls to activate the remaining PPN neurons to restore locomotor function (Mazzone et al. 2005; Plaha and Gill 2005; Stefani et al. 2007; Ferraye et al. 2010; Moro et al. 2010; Thevathasan et al. 2011; Lau et al. 2015; Welter et al. 2015; Mestre et al. 2016). However, despite an initial great optimism (Mazzone et al. 2005), the overall results of controlled clinical studies using PPN-DBS in PD patients have been disappointing and with unpredictable outcome for the majority of patients (Ferraye et al. 2010; Moro et al. 2010; Thevathasan et al. 2011; Welter et al. 2015). Moreover, a recent double-blind clinical study concluded that stimulation of the PPN area has an initial but not sustained benefit for PD gait disorders (Mestre et al. 2016). Many factors could explain the variability of these clinical outcomes, including patient selection, gait assessment, and target location. These basic questions remain unresolved, but the main interrogation remains the exact anatomy of the PPN and the normal physiology of human gait that is poorly understood at the neuronal network level.

The PPN together with the adjacent cuneiform nucleus (CuN) belongs to the mesencephalic locomotor region (MLR), previously named by experimental physiologists, because its stimulation produced locomotion (see the recent review by (Garcia-Rill et al. 2015). Numerous anatomical and physiological studies have been conducted over many years to analyse the PPN inputs and outputs but mainly in rodents. At the present time, the connections of the PPN and above all of the CuN with the basal ganglia, thalamus, and cortex, as well as their descending output pathways are insufficiently described and required new anatomical investigations in primates. Recent physiological studies show that the PPN is not only

involved in motor control, but also in many other functions as various as sleep, sensory integration, attention, and even emotion. Taken together, these results suggest that the primate PPN has a role that extends beyond simple locomotor motor control. In particular, the PPN appears to integrate information that would be necessary for adapting locomotion to environmental demands (Lau et al. 2015). Several other nuclei of the reticular formation (such as raphe nuclei, locus coeruleus, cuneiform nucleus...) likely share this integrative feature, participating in functions as varied as sleep, attention, emotion, and locomotion. A striking example of the non-motor role of the PPN is its involvement in modulating wakefulness and sleep. In particular, pharmacological and neurophysiological studies in animals have emphasised the importance of brainstem cholinergic neurons in the generation of rapid eye movement (REM) sleep (Steriade et al. 1990; Rye 1997; Datta 2002). These cholinergic neurons are in a position to influence thalamo-cortical activity, sleep-wakefulness states, and muscle tone via its ascending projections to the forebrain and descending projections to the reticulospinal pathway (Pare et al. 1988; Jones 1993). Clinical observations have also reported that PPN-DBS in PD patients can significantly modify sleep quality, with increased daytime alertness and a long-term improvement of night-time sleep (Arnulf et al. 2010; Peppe et al. 2012). The PPN is a component of a group of upper brainstem nuclei that constitute the ascending reticular activating system. As such, the PPN and particularly its cholinergic neurons exert a role in regulating sleep and arousal.

Here, we review the anatomy of the PPN and its role in locomotor and sleep control in non-human primate, describing and discussing our recent experimental studies.

Relevance of non-human primates for studies of locomotion and sleep in normal and parkinsonian states

Primate research is essential to better understand the physiological basis of human neurological disorders which have dramatic repercussions on public health. Moreover, this research is of great assistance in performing preclinical therapeutic trials to launch new treatments for patients with the best technical and ethical conditions. Concerning PD, monkeys treated with the neurotoxin 1-methyl-4-phenyl-1,2,3,6-tetrahydropyridine (MPTP)

constitute the most relevant model available to date, since it can reproduce both dopaminergic neuronal loss and the cardinal parkinsonian symptoms (akinesia, rigidity, and episodes of tremor) that closely resemble those observed in PD patients (Emborg 2007). The MPTP primate model presents some limitations that have already been reviewed (Collier et al. 2003; Emborg 2004). In particular, even though postural deficits are frequently reported, whether they are dopa-sensitive and/or associated with episodes of FOG or falls is unknown.

In our laboratory, we were interested in developing a monkey model which not only displays cardinal parkinsonian symptoms, but also gait and balance disorders which are currently the most debilitating and untreatable symptoms in advanced PD. The physiological mechanisms of gait and posture control have been studied in diverse species, including lamprey, cat, and rodent (Grillner 2003; Clarac 2008; McCrea and Rybak 2008). However, studies on the kinematic and biomechanical properties of gait in macaques have concluded that primate locomotion (including old word primates and humans) is fundamentally different from locomotion in other mammals, and while permanent bipedalism is unique to human (see Vilensky 1983; Courtine et al. 2005; Xiang et al. 2007), it is observed for short durations and over a short distances in non-human primate. It thus seems more appropriate to analyse gait and posture in normal and MPTPtreated macaques than in non-primate species. Since the previous experimental and clinical studies have shown that the PPN is involved in gait control (Pahapill and Lozano 2000), it was of interest to focus on the behavioural effects induced by PPN lesions in normal and parkinsonian macaques to propose a better model of advanced PD.

With regards to the sleep-wake cycle, diurnal habits as well as the nocturnal sleep of monkeys (adult macaques in particular) are much more similar to the sleep architecture in humans than those in rodent or cat. This model could thus allow us to fill a gap between sleep studies in rodent and cat and those in human (Hsieh et al. 2008). MPTP-treated monkeys develop stable sleep disorders with abnormal sleep architecture similar to that seen in PD patients (Barraud et al. 2009). These results demonstrate that monkeys and macaques, in particular, are a good biomedical model to study locomotion and sleep especially in the context of PD.

Anatomical definition and connections of the

PPN in primate

It should be pointed out that only minimal data on the PPN are available for monkeys in comparison to rodent and cat. However, the projections of the PPN are assumed to be similar between these species, the MLR being a highly conserved structure in all vertebrates (Shik et al. 1966; Garcia-Rill et al. 1987; Sirota et al. 2000; Ryczko et al. 2016). Here, the studies available for primates are first discussed together with our recent anatomical data.

The PPN and the adjacent CuN are both located within the reticular formation of the lateral mesencephalon of the upper brainstem. These two structures correspond to open nuclei with unclear boundaries whichever histological stains are used. Both nuclei have a complex cellular organisation with GABAergic and glutamatergic neuronal populations, whereas cholinergic neurons are only present within the PPN itself. This cholinergic cell population has been reported to account for 50% of PPN neurons in human (Manaye et al. 1999). PPN cholinergic neurons are clustered within a dense dorsolateral area (PPN pars compacta) and are scattered more medially (PPN pars dissipata) classically defined as the Ch5 group of cholinergic neurons (Mesulam et al. 1983, 1984, 1989). PPN glutamatergic and GABAergic neurons are observed in the rat (Clements et al. 1991; Wang and Morales 2009; Martinez-Gonzalez et al. 2011) and cat (Jia et al. 2003). In monkeys, GABAergic neurons were not found in the PPN (Lavoie and Parent 1994a), whereas both glutamatergic and GABAergic PPN neurons appear to project to midbrain dopamine neurons (Charara et al. 1996). The possible colocalization of acetylcholine with either glutamate or GABA in PPN neurons is still controversial. About 50% of cholinergic neurons were reported to express GABA (Jia et al. 2003), and about 40% seem to express glutamate in rat and monkey (Clements et al. 1991; Lavoie et al. and Parent 1994a). However, no cholinergic neurons were observed to coexpress GABA or glutamate using a combination of in situ hybridization and immunohistochemistry (Wang et aland Morales- 2009). Recent studies reported that GABAergic neurons are more densely concentrated in the rostral PPN, compared to cholinergic and glutamatergic neurons (Mena-Segovia et al. 2009; Martinez-Gonzalez et al. 2011), suggesting that the PPN should be regarded as an heterogeneous structure. The rostral PPN, which contains significantly more GABAergic neurons, would be interconnected with the basal ganglia, whereas the caudal PPN, which contains a larger number of cholinergic and

glutamatergic neurons, would be more related to thalamo-cortical systems and to regions involved in regulation of gait and posture (Martinez-Gonzalez et al. 2011). Whether such a rostro-caudal organisation of neuronal populations within the PPN and the CuN exists in human remains to be determined.

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Anatomical and physiological studies have identified many connections to the PPN and CuN. However, due to their open boundaries and their neurochemical heterogeneity, it has been difficult to precisely determine which inputs terminate in the PPN or CuN. Overall, the PPN receives dense cortical afferents from the premotor and motor cortices (Matsumura et al. 2000), suggesting that the PPN integrates motor information. The PPN is also highly interconnected with the basal ganglia (see, for review, Pahapill et al. and Lozano 2000). In particular, it has been demonstrated that the PPN integrates information arising from the three different functional territories of the internal pallidum in monkey (Shink et al. 1997). The PPN also sends profuse cholinergic and non-cholinergic ascending efferent fibers to all thalamic nuclei, although the intralaminar and reticular nuclei are particularly densely innervated in monkey (Steriade et al. 1988; Lavoie et al. 1994b Lavoie and Parent 1994b) and human (Heckers et al. 1992). Moreover, the parafascicular nucleus appears more strongly innervated than the centre median in monkey (Olivier et al. 1970). Projections from the deep cerebellar nuclei have also been demonstrated (Hazrati and Parent 1992).

We conducted a series of tract-tracing experiments in monkey to analyse the input and output connections of the PPN and the CuN, whose connections are poorly understood. Our results show that the descending outputs of both the PPN and the CuN target the pontobulbar reticular formation at the origin of the reticulospinal pathway (Rolland et al. 2011). Figure 1 illustrates the anterograde labelling of fibers obtained after tracer injection into the MLR.

Fig. 1

a Photomicrograph and map of biotin dextran amine injection site into PPN (mainly in its dorsal part) with diffusion within the ventral CuN. **b** Maps of anterogradely labelled PPN terminals on transverse sections of the pontomedullary reticular formation. BC brachium conjunctivum, Gi

gigantocellular nucleus, *ml* medial lemniscus, *MLF* medial longitudinal fascicle, *PAG* periaqueductal grey matter, *py* pyramidal tract, *SC* superior colliculus, *V* trigeminal nucleus.

Figure adapted from Rolland et al. (2011)



Whereas the internal pallidum principally innervates the PPN, the substantia nigra pars reticulata innervates both the PPN and CuN leading to the conclusion that the internal pallidum and the substantia nigra control different parts of the MLR and can modulate the descending reticulospinal pathway in the primate.

We have also demonstrated that both the PPN and CuN receive dopamine innervation in the macaque (Rolland et al. 2009). The dopaminergic fibers were mainly found in the anterior part of the PPN and tended to avoid cholinergic neurons (Fig. 2). These data suggest that dopamine may have a role in the modulation of MLR neural activity.

Fig. 2

Distribution of dopaminergic fibers (labelled with dopamine transporter) throughout the PPN and CuN in relation to the cell bodies of cholinergic neurons (*red dots*) (labelled with choline transporter) on transverse sections of a macaque. 4 nucleus of the fourth cranial nerve, *BC* brachium conjunctivum, *DBC* decussation of the brachium conjunctivum, *IC* inferior colliculus, *LL* lateral lemniscus, *ml* medial lemniscus, *MLF* medial longitudinal fascicle, *PAG* periaqueductal grey matter.

Figure adapted from Rolland et al. (2009)



We recently conducted experiments to determine the organisation of both PPN and CuN inputs and outputs in macaque and human with a special focus on the three anatomo-functional sensorimotor, associative and limbic territories of the basal ganglia, and on the thalamus, amygdala, and cortex (Sebille et al. 2016). We found that PPN and CuN connections are generally similar in both species, using tract-tracing methods in monkey, and diffusion-weighted imaging-based methods in healthy human volunteers. The PPN projects to the three anatomo-functional territories of the basal ganglia nuclei, to the centre median-parafascicular complex of the thalamus, and to the central nucleus of the amygdala. The PPN receives from the motor cortical areas, especially in its anterior part, and less strongly from the associative and limbic cortices. The CuN projects to similar cerebral structures as the PPN. However, CuN projections were much stronger to the limbic territories of the basal ganglia and thalamus, and to the basal forebrain (extended amygdala) and the central nucleus of the amygdala. We conclude that the PPN integrates sensorimotor, cognitive, and emotional information, whereas the CuN participates in a more restricted network predominantly integrating emotional information (Karachi et al. 2014; Sebille et al. 2016) (Fig. 3).

Fig. 3

a Sagittal view of fibers (*yellow*) connecting the CuN (which represents the seed) and the limbic part of the subthalamic nucleus (STN) (which represents the target) in a human subject (data provided from the Human

Connectome Project) using diffusion-weighted imaging-based tractography. The sensorimotor, associative, and limbic anatomo-functional territories of the STN are represented in *green*, *pink*, and *yellow*, respectively. In this sagittal view, we have also chosen to illustrate the fiber tract originating from the CuN and descending to the spinal cord to validate the specificity of the reconstructed tracts. Histograms showing percentages of connected voxels from the anatomo-functional territories of the STN to the PPN and CuN, averaged in 30 human subjects. The *error bars* represent standard deviation. Wilcoxon signed-rank test: ***p < 0.001.

Figure adapted from Sebille et al. (2016)



Involvement of the PPN in normal locomotor control

The previous experimental studies have shown that a train of electrical stimulation, at 20–60 Hz, within the MLR can produce locomotion in rat,

cat, and monkey decerebrate preparations (Shik et al. 1966; Eidelberg et al. 1981; Garcia-Rill et al. 1987; Garcia-Rill and Skinner 1987). However, a delay is observed between stimulation onset and the induction of the first step, suggesting that the MLR is not a step generator per se and that locomotion is gradually recruited (Garcia-Rill 1991). Notably, increasing current intensity resulted in changes in the frequency of the step cycle, going from a walk to a gallop (Grillner 1981). Moreover, the activity of some PPN neurons was often related to the duration of the stepping episode, while other neurons were shown to be active in relation to left-right alternation (Garcia-Rill and Skinner 1988; Garcia-Rill 1991). The locomotor behaviour obtained following MLR stimulation varied with the location of the stimulating electrode, since stimulation of the CuN in decerebrate cat seemed to elicit locomotor patterns, whereas stimulation of the PPN could change muscle tone (Takakusaki et al. 2003). In the intact cat, stimulation of the PPN area induces complex motor behaviour, the animal exhibiting fast walking and running movements, avoiding collision with obstacles (Mori et al. 1989). In rat, PPN lesions produce motor deficits (MacLaren et al. 2014) if the lesion is restricted to the anterior part of the PPN (Alderson et al. 2008). Moreover, a recent study using an optogenetic approach in mice enabled the identification of distinct locomotor functions of the three neurochemically identified cell types within the MLR showing that PPN glutamatergic neurons, and not cholinergic ones, could provoke locomotion (Roseberry et al. 2016). In summary, while many experimental studies have shown that electrical or pharmacological manipulations of the PPN area can modulate gait on a treadmill in cat and rodent, little is known about the role of these structures in primates.

In monkey, widespread lesion or inactivation of the whole PPN region (by excitotoxic or radiofrequency lesions, local injection of GABAergic agonists, or high-frequency stimulation), induces motor symptoms which have been identified as parkinsonism, since they include akinesia and rigidity (Kojima et al. 1997; MunroDavies et al. 1999; Matsumura and Kojima 2001; Nandi et al. 2002; Pereira et al. 2011). However, dopamine therapy was not tested in these animals, gait and balance were not carefully examined, and it was not known whether these effects are due to degeneration of cholinergic neurons, non-cholinergic neurons, or both.

We thus conducted an initial series of experiments to demonstrate whether

a PPN cholinergic lesion in monkey can induce gait disorders (Karachi et al. 2010). For this study, young macaques (3-5 years old) were trained to walk along a corridor, and various parameters of gait and posture were quantified. Akinesia, rigidity, and tremor were assessed together with the general activity. Stereotaxic injections of diphtheria toxin conjugated with urotensin II specific to cholinergic neurons (Clark et al. 2007) and restricted to the PPN were performed uni and bilaterally. Such injections resulted in a mean reduction of 39% of cholinergic PPN neurons, which is close to the partial PPN neuronal degeneration reported in the PPN of PD patients (Hirsch et al. 1987). Lesioned monkeys developed postural and gait abnormalities. These symptoms were characterised by strong axial rigidity (increase of the back curvature and modification of the tail position), a mild peripheral rigidity (decrease in the knee angle), but without akinesia (normal overall activity) or tremor. If this rigidity looked partially like that obtained following MPTP intoxication, it predominantly involved axial body parts and no cog-wheel rigidity of the limbs was noticed. These postural and gait deficits were not improved by dopaminergic medication, demonstrating that they were unrelated to dopaminergic depletion.

We concluded from these experiments that cholinergic PPN neurons are involved in gait and postural control. However, a small proportion of noncholinergic PPN neurons was also affected by the toxin and could, therefore, have been involved in the expression of these symptoms.

Involvement of the PPN in parkinsonian locomotor dysfunction

As previously discussed, MPTP intoxication in young macaques induces cardinal parkinsonian symptoms responsive to dopaminergic treatment. However, gait and balance were rarely examined and never described in these animals. Histological analysis and cell counts failed to show any loss of PPN cholinergic neurons (Herrero et al. 1993; Heise et al. 2005). We can conclude, therefore, that this PD model does not produce all the clinical and histological features of advanced PD patients.

Our aim was thus to develop a new monkey model of advanced PD which displays the cardinal parkinsonian symptoms together with gait and balance disorders with dopaminergic and cholinergic cell loss. Initially, we looked to see whether cholinergic lesions of the PPN in young MPTP-

treated macaques (Macaca fascicularis, 3-5 years old) would add gait and balance disorders resistant to dopaminergic drug treatment to the cardinal parkinsonian symptoms (Grabli et al. 2013). Experiments were conducted using similar procedures as those explained above (Karachi et al. 2010). MPTP intoxication was first performed to obtain a mild, stable parkinsonian state. We then performed stereotaxic lesions of the PPN specific to its cholinergic neurons (Karachi et al. 2010). We first observed that adding bilateral partial cholinergic PPN lesions in young MPTPtreated macaque (41% loss of cholinergic PPN neurons) induced a worsening of axial posture (flexed trunk displaced towards the side contralateral to the lesion and erect tail), an increase of knee angle and height of the pelvis, and a weak but persistent balance deficit that induced more falls compared to the parkinsonian state. These parameters did not improve with dopaminergic medication. However, we observed that after PPN lesions, tremor and akinesia improved significantly, with an increase of gait speed but no significant change of step length. Finally, additional MPTP injections gave the clinical association of both a severe DOPAresponsive parkinsonism and a DOPA-unresponsive gait and balance disorders. These complex changes in axial muscle tone induced by PPN lesions may be mediated via its descending input to the reticulospinal tract (Rye et al. 1988; Lai and Siegel 1990; Takakusaki et al. 2003). We also observed that PPN lesions improved tremor and akinesia, with an increase of gait speed but no statistical change of step length compared with parkinsonian state. Such improvement of parkinsonian symptoms could be explained by the reduction of excitatory outputs from the PPN cholinergic neurons to the hyperactive subthalamic nucleus that strongly participates in PD symptom severity.

To reinforce our new monkey model of advanced PD, we performed additional MPTP injections. These dramatically worsened akinesia as well as balance deficits and falls during walking. The symptoms were worsened by dopaminergic treatment, increasing the ipsilateral hypotonia (Fig. 4b), which is consistent with clinical observations of PD patients (Giladi et al. 2001). With these experiments, we were able to obtain robust dopamineresistant gait and balance disorders associated with the cardinal parkinsonian symptoms in the macaque. This new monkey model of advanced PD will be useful especially for testing new medical or surgical approaches.

Fig. 4

a Photomicrographs of the same animal in control state, after MPTP administration, and then following a PPN lesion (DA and PPN lesion). **b** Development of balance deficit observed after additional doses of MPTP and apomorphine injection. **c** Total number of cholinergic neurons quantified in young and aged MPTP-treated macaques compared with their respective controls (n = 4 per group). MPTP did not affect cholinergic neurons in young animals, but induced a 30% loss of cholinergic neurons in aged monkeys. **p < 0.01, Mann–Whitney U test.

Figure adapted from Grabli et al. (2013)

In parallel, we also used aged macaques to determine whether balance disorders could be induced using MPTP intoxication only (Karachi et al.

2010; Grabli et al. 2013). We suspected that this could be the case, because aged MPTP-treated macaques have been reported to develop abnormal postures and poor balance (Ovadia et al. 1995). We thus administered MPTP to aged macaques (20-30 years old) of two different species (Macaca arctoides and M. fascicularis). The macaques developed severe cardinal parkinsonian symptoms that were improved by dopaminergic treatment, but also balance problems during walking with occasional falls that were worsened following dopaminergic medication. Histological examination showed severe dopaminergic neuronal loss in the substantia nigra and a significant partial loss of PPN cholinergic neurons (30% in *M. arctoides* and 22% in *M. fascicularis*) (Fig. 4c). These data strongly suggested a relationship between cholinergic neuronal loss and axial symptoms. MPTP is known to be a selective toxin for the DA neurons. However, the possibility that MPTP finally also kills some PPN cholinergic neurons cannot be ruled out. Indeed, chronically MPTP injections appear to affect non-dopaminergic neurons in the thalamus (Villalba et al. 2014) as well as noradrenergic neurons in the locus coeruleus (Masilamoni et al. 2011). Thus, the cholinergic neuronal loss that we observed in MPTP-treated monkeys is due to both a direct action of MPTP on these neurons as well as the consequences of functional modifications induced by MPTP in basal ganglia activity. However, because PPN cholinergic loss was too low and gait disorders too mild in these aged MPTP-treated monkeys, we considered that these animals could not be used as a model of advanced PD.

Involvement of the PPN in normal and parkinsonian sleep

Many pharmacological and neurophysiological studies have indicated the importance of cholinergic neurons of the PPN and the adjacent dorsolateral tegmental nuclei in the regulation of arousal and REM sleep. Large PPN lesions in cats severely reduce REM sleep with muscle atonia (Webster and Jones 1988; Shouse and Siegel 1992). However, more restricted lesions of the rat PPN have failed to show any alteration in sleep architecture (Deurveilher and Hennevin 2001). It has been shown that pharmacological or electrical stimulation of the PPN in rat or cat is able to elicit REM sleep together with cortical activation and muscle atonia (Jones 1993; Takakusaki et al. 2004). Recently, cholinergic PPN neurons have been shown to discharge maximally during waking and during REM sleep in rat

(Boucetta et al. 2014). Taken together, these experimental data highlight the complex role of the cholinergic PPN neurons in the control of normal sleep, and raise the question of whether this specific cell loss could participate in the emergence of sleep disorders in the context of PD.

Sleep disorders are an important non-motor symptom of PD. They are characterised by excessive daytime sleepiness and nocturnal sleep disorders among which are difficulty in falling and staying asleep with sleep fragmentation (Wailke et al. 2011; Schrempf et al. 2014). Whether these sleep disorders are primarily caused by the PD itself or by the dopaminergic treatment or both is difficult to determine. Moreover, the effects of dopaminergic drugs on sleep are variable, depending on the nature of the pharmacological agent and the doses used. Dopamine agonists have been reported to provoke sleepiness, particularly at high doses, whereas L-dopa seems to have the opposite effect (Bliwise et al. 2012). Cholinergic PPN cell loss may also have a relevant role in the development of sleep disorders in PD, since PPN low-frequency stimulation has been shown to significantly improve sleep efficiency with an increase of REM sleep episodes (Romigi et al. 2008; Alessandro et al. 2010; Arnulf et al. 2010; Peppe et al. 2012). These data suggest that further investigations are necessary to disentangle the roles of dopaminergic cell death, dopaminergic treatment, and PPN degeneration in PD sleep disorders.

The suitability of the macaque model to replicate sleep–wake disturbances in PD patients has been well demonstrated (Barraud et al. 2009). Using a similar macaque model, we performed long-term continuous electroencephalographic monitoring of vigilance states using an implanted miniaturised telemetry device to examine the sleep–wake parameters. We designed a longitudinal study to record sleep in macaques during different stages: baseline, parkinsonian without and with dopaminergic treatment, and following PPN cholinergic lesions (Belaid et al. 2014). We confirmed that normal macaques slept for 42% of the 24 h, mostly during the night. Sleep includes rapid eye movements (REM), non-REM cycles with light and slow-wave, deep sleep, and waking episodes (Hsieh et al. 2008; Barraud et al. 2009). MPTP-treated macaques displayed a reduction in time spent in REM and slow-wave sleep, and an increased number of nocturnal awakenings. These sleep disorders were similar to those previously reported (Almirall et al. 1999; Barraud et al. 2009; Hyacinthe

et al. 2014) and to those recorded in advanced PD patients (Wailke et al. 2011; Yong et al. 2011). Our MPTP-treated macaques also increased their daytime sleepiness as previously reported (Rye 2010). We concluded that DA lesion alone (severe in our macaques) accounts for the development of sleep disorders, supporting the involvement of the DA system in the regulation of sleep–wake cycle.

To determine whether dopaminergic treatment can alleviate sleep disorders, we administered L-dopa twice a day for 8-12 days to MPTPtreated macaques. This dopaminergic treatment resulted in a partial but a significant improvement of almost all sleep parameters (reduction of daytime sleepiness and of nocturnal awakenings, sleep pattern benefits). Using this macaque model, it has further been reported that selective D1 receptor agonist administration can improve REM sleep and excessive daytime sleepiness, whereas D2 receptor agonists have no effect (Hyacinthe et al. 2014). These results differ from those reported in PD patients, since it has been shown that dopaminergic treatment has no impact on the altered sleep structure and could even decrease the total sleep period (Wailke et al. 2011). These discrepancies may be explained by the fact that our animals were young and were treated with L-dopa after a 2 week habituation period, whereas L-dopa is administered in middleaged patients and for years. It is also possible that, compared to MPTPtreated macaques, PD patients have non-dopaminergic lesions which could limit the beneficial effects of the dopaminergic treatment. In conclusion, these experiments demonstrated the efficacy of the dopaminergic treatment in improving sleep disorders in MPTP-treated macaque.

To determine the role of the PPN on sleep disorders, and in particular whether PPN lesions could reduce REM sleep in MPTP-treated macaques, we added a specific cholinergic PPN lesion to the dopaminergic cell loss in the same animal (Belaid et al. 2014). This partial cholinergic PPN lesion induced transient acute sleep disturbances characterised by an increase in sleep fragmentation and a decrease in sleep efficiency and REM sleep. These results confirm the role of cholinergic PPN neurons in REM sleep as proposed by Lu et al. (2006). We also observed the occurrence of highamplitude slow-wave sleep during daytime, which resembles that described following lesions of the reticular formation in cat (Denoyer et al. 1991) or its pharmacological manipulation in rabbit (Longo 1956). These changes may be explained by the fact that the cholinergic PPN neurons are no

longer able to activate the thalamo-cortical pathway, inducing a suppression of the transition from slow-wave sleep to either arousal or REM sleep (Steriade et al. 1991). However, all the changes we observed were only transient. Surprisingly, 3 weeks after the PPN lesion, monkeys slept slightly better than after MPTP intoxication, with an increase in sleep efficiency and a decrease of nocturnal awakenings (Fig. 5). Several hypotheses could be advanced to explain this benefit. First, compensatory processes could occur and could have masked the acute effects of the PPN lesions. These compensatory processes may involve the remaining PPN cholinergic neurons or other modulatory neurons from systems that are known to be involved in sleep-wake control, such as the adjacent dorsolateral tegmental nucleus, the locus coeruleus, or the raphe nucleus. Second, the cholinergic PPN lesions could be responsible for a reduction of PPN excitatory inputs to the subthalamic nucleus, which is overactive in PD, leading to an improvement of PD motor symptoms and then sleep. This fits well with the reversal of subthalamic hyperactivity reported in a 6-hydroxydopamine rat model after a PPN lesion (Breit et al. 2006). This might suggest that the improvement of sleep quality after a PPN lesion is, at least in part, a consequence of a reduction in night-time akinesia and painfulness.

Fig. 5

Examples of hypnograms of 24 h recordings in the same macaque at baseline (CTL), after MPTP intoxication, after L-dopa administration, and 3 weeks after subsequent PPN lesion.

Figure published in Belaid et al. (2014)

In summary, the combination of dopaminergic and cholinergic PPN lesions allowed us to characterise sleep disorders in a monkey model of advanced PD. We concluded that a partial cholinergic PPN lesion in MPTP-treated macaque is not sufficient to induce consistent additional sleep disorders compared to the parkinsonian state.

Conclusion

From our primate research, we proposed a new monkey model of advanced PD that displays the cardinal parkinsonian symptoms together with gait and balance disorders, and dopaminergic and cholinergic cell loss. However, we are fully aware of several limitations to using a monkey model of PD. In particular, the regular quadrupedal locomotion of macaques cannot be directly compared to the bipedal gait in humans. In that sense, a macaque model of bipedal locomotion has been developed (Goetz et al. 2012) and has been further used as a pre-clinical tool to study gait parameters and their neuronal control after MPTP intoxication (Goetz et al. 2016). However, it should be noted that the bipedal locomotion of macaque, observed over a short distance in its natural environment (Hemmi and Menzel 1995), is obtained under restrained experimental conditions by inducing postural changes that cannot be directly compared

with natural monkey or human gait. Our monkey model that used natural quadrupedal locomotion also cannot be directly compared to bipedal gait in humans. These two approaches are complementary and both are potentially useful in developing monkey models of gait disorders, especially with a pretherapeutic aim.

The pre-clinical studies using monkeys reported here have been made possible by a collaborative interdisciplinary clinical and experimental approach. We believe that this work represents an important step in the validation of the safety and efficacy of new therapeutic procedures prior to their transfer to larger scale clinical trials.

Acknowledgements

The authors would like to thank Max Westby for language editing. The research leading to these results received funding from the "Investissements d'avenir" (investing in the future) programme ANR-10-IAIHU-06.

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Compliance with ethical standards

Conflict of interest The authors declare no other conflicts of interest.

References

Alderson HL, Latimer MP, Winn P (2008) A functional dissociation of the anterior and posterior pedunculopontine tegmental nucleus: excitotoxic lesions have differential effects on locomotion and the response to nicotine. Brain Struct Funct 213:247–253

Alessandro S, Ceravolo R, Brusa L, Pierantozzi M, Costa A, Galati S, Placidi F, Romigi A, Iani C, Marzetti F, Peppe A (2010) Non-motor functions in parkinsonian patients implanted in the pedunculopontine nucleus: focus on sleep and cognitive domains. J Neurol Sci 289:44–48

Almirall H, Pigarev I, de la Calzada MD, Pigareva M, Herrero MT, Sagales T (1999) Nocturnal sleep structure and temperature slope in MPTP treated monkeys. J Neural Transm (Vienna) 106:1125–1134

Arnulf I, Ferraye M, Fraix V, Benabid AL, Chabardes S, Goetz L, Pollak P, Debu B (2010) Sleep induced by stimulation in the human pedunculopontine nucleus area. Ann Neurol 67:546–549

Barraud Q, Lambrecq V, Forni C, McGuire S, Hill M, Bioulac B, Balzamo E, Bezard E, Tison F, Ghorayeb I (2009) Sleep disorders in Parkinson's disease: the contribution of the MPTP non-human primate model. Exp Neurol 219:574–582

Belaid H, Adrien J, Laffrat E, Tande D, Karachi C, Grabli D, Arnulf I, Clark SD, Drouot X, Hirsch EC, Francois C (2014) Sleep disorders in Parkinsonian macaques: effects of L-dopa treatment and pedunculopontine nucleus lesion. J Neurosci 34:9124–9133

Bliwise DL, Trotti LM, Wilson AG, Greer SA, Wood-Siverio C, Juncos JJ, Factor SA, Freeman A, Rye DB (2012) Daytime alertness in Parkinson's disease: potentially dose-dependent, divergent effects by drug class. Mov Disord 27:1118–1124

Boucetta S, Cisse Y, Mainville L, Morales M, Jones BE (2014) Discharge profiles across the sleep–waking cycle of identified cholinergic, GABAergic, and glutamatergic neurons in the pontomesencephalic tegmentum of the rat. J Neurosci 34:4708–4727

Breit S, Lessmann L, Unterbrink D, Popa RC, Gasser T, Schulz JB (2006) Lesion of the pedunculopontine nucleus reverses hyperactivity of the subthalamic nucleus and substantia nigra pars reticulata in a 6-hydroxydopamine rat model. Eur J Neurosci 24:2275–2282

Charara A, Smith Y, Parent A (1996) Glutamatergic inputs from the pedunculopontine nucleus to midbrain dopaminergic neurons in primates: phaseolus vulgaris-leucoagglutinin anterograde labeling combined with postembedding glutamate and GABA immunohistochemistry. J Comp Neurol 364:254–266

Clarac F (2008) Some historical reflections on the neural control of locomotion. Brain Res Rev 57:13–21

Clark SD, Alderson HL, Winn P, Latimer MP, Nothacker HP, Civelli O

(2007) Fusion of diphtheria toxin and urotensin II produces a neurotoxin selective for cholinergic neurons in the rat mesopontine tegmentum. J Neurochem 102:112–120

Clements JR, Toth DD, Highfield DA, Grant SJ (1991) Glutamate-like immunoreactivity is present within cholinergic neurons of the laterodorsal tegmental and pedunculopontine nuclei. Adv Exp Med Biol 295:127–142

Collier TJ, Steece-Collier K, Kordower JH (2003) Primate models of Parkinson's disease. Exp Neurol 183:258–262

Courtine G, Roy RR, Hodgson J, McKay H, Raven J, Zhong H, Yang H, Tuszynski MH, Edgerton VR (2005) Kinematic and EMG determinants in quadrupedal locomotion of a non-human primate (Rhesus). J Neurophysiol 93:3127–3145

Datta S (2002) Evidence that REM sleep is controlled by the activation of brain stem pedunculopontine tegmental kainate receptor. J Neurophysiol 87:1790–1798

Denoyer M, Sallanon M, Buda C, Kitahama K, Jouvet M (1991) Neurotoxic lesion of the mesencephalic reticular formation and/or the posterior hypothalamus does not alter waking in the cat. Brain Res 539:287–303

Deurveilher S, Hennevin E (2001) Lesions of the pedunculopontine tegmental nucleus reduce paradoxical sleep (PS) propensity: evidence from a short-term PS deprivation study in rats. Eur J Neurosci 13:1963– 1976

Eidelberg E, Walden JG, Nguyen LH (1981) Locomotor control in macaque monkeys. Brain 104:647–663

Emborg ME (2004) Evaluation of animal models of Parkinson's disease for neuroprotective strategies. J Neurosci Methods 139:121–143

Emborg ME (2007) Nonhuman primate models of Parkinson's disease. ILAR J 48:339–355

Ferraye MU, Debu B, Fraix V, Goetz L, Ardouin C, Yelnik J, Henry-Lagrange C, Seigneuret E, Piallat B, Krack P, Le Bas JF, Benabid AL, Chabardes S, Pollak P (2010) Effects of pedunculopontine nucleus area stimulation on gait disorders in Parkinson's disease. Brain 133:205–214

Garcia-Rill E (1991) The pedunculopontine nucleus. Prog Neurobiol 36:363–389

Garcia-Rill E, Skinner RD (1987) The mesencephalic locomotor region. I. Activation of a medullary projection site. Brain Res 411:1–12

Garcia-Rill E, Skinner RD (1988) Modulation of rhythmic function in the posterior midbrain. Neuroscience 27:639–654

Garcia-Rill E, Houser CR, Skinner RD, Smith W, Woodward DJ (1987) Locomotion-inducing sites in the vicinity of the pedunculopontine nucleus. Brain Res Bull 18:731–738

Garcia-Rill E, Hyde J, Kezunovic N, Urbano FJ, Petersen E (2015) The physiology of the pedunculopontine nucleus: implications for deep brain stimulation. J Neural Transm 122:225–235

George O, Parducz A, Dupret D, Kharouby M, Le Moal M, Piazza PV, Mayo W (2006) Smad-dependent alterations of PPT cholinergic neurons as a pathophysiological mechanism of age-related sleep-dependent memory impairments. Neurobiol Aging 27:1848–1858 AQ3

Giladi N, McDermott MP, Fahn S, Przedborski S, Jankovic J, Stern M, Tanner C, Parkinson Study Group (2001) Freezing of gait in PD: prospective assessment in the DATATOP cohort. Neurology 56:1712– 1721

Goetz L, Piallat B, Thibaudier Y, Montigon O, David O, Chabardes S (2012) A non-human primate model of bipedal locomotion under restrained condition allowing gait studies and single unit brain recordings. J Neurosci Methods 204:306–317

Goetz L, Piallat B, Bhattacharjee M, Mathieu H, David O, Chabardes S

(2016) On the role of the pedunculopontine nucleus and mesencephalic reticular formation in locomotion in nonhuman primates. J Neurosci 36:4917–4929

Grabli D, Karachi C, Folgoas E, Monfort M, Tande D, Clark S, Civelli O, Hirsch EC, Francois C (2013) Gait disorders in parkinsonian monkeys with pedunculopontine nucleus lesions: a tale of two systems. J Neurosci 33:11986–11993

Grillner S (1981) Control of locomotion in bipeds, tetrapods and fish. In: Brooks VB (ed) Handbook of physiology. The nervous system II, vol V. American Physiological Society, Baltimore, pp 1199–1236

Grillner S (2003) The motor infrastructure: from ion channels to neuronal networks. Nat Rev Neurosci 4:573–586

Hazrati LN, Parent A (1992) Projection from the deep cerebellar nuclei to the pedunculopontine nucleus in the squirrel monkey. Brain Res 585:267–271

Heckers S, Geula C, Mesulam MM (1992) Cholinergic innervation of the human thalamus: dual origin and differential nuclear distribution. J Comp Neurol 325:68–82

Heise CE, Teo ZC, Wallace BA, Ashkan K, Benabid AL, Mitrofanis J (2005) Cell survival patterns in the pedunculopontine tegmental nucleus of methyl-4-phenyl-1,2,3,6-tetrahydropyridine-treated monkeys and 6OHDA-lesioned rats: evidence for differences to idiopathic Parkinson disease patients? Anat Embryol (Berl) 210:287–302

Hemmi JM, Menzel CR (1995) Foraging strategies of long-tailed macaques, *Macaca fascicularis*—directional extrapolation. Anim Behav 49:457–464

Herrero MT, Hirsch EC, Javoy-Agid F, Obeso JA, Agid Y (1993) Differential vulnerability to 1-methyl-4-phenyl-1,2,3,6tetrahydropyridine of dopaminergic and cholinergic neurons in the monkey mesopontine tegmentum. Brain Res 624:281–285

Hirsch EC, Graybiel AM, Duyckaerts C, Javoy-Agid F (1987) Neuronal loss in the pedunculopontine tegmental nucleus in parkinson disease and in progressive supranuclear palsy. Proc Natl Acad Sci USA (Washington) 84:5976–5980

Hsieh KC, Robinson EL, Fuller CA (2008) Sleep architecture in unrestrained rhesus monkeys (*Macaca mulatta*) synchronized to 24hour light-dark cycles. Sleep 31:1239–1250

Hyacinthe C, Barraud Q, Tison F, Bezard E, Ghorayeb I (2014) D1 receptor agonist improves sleep–wake parameters in experimental parkinsonism. Neurobiol Dis 63:20–24

Jia HG, Yamuy J, Sampogna S, Morales FR, Chase MH (2003) Colocalization of gamma-aminobutyric acid and acetylcholine in neurons in the laterodorsal and pedunculopontine tegmental nuclei in the cat: a light and electron microscopic study. Brain Res 992:205–219

Jones BE (1993) The organization of central cholinergic systems and their functional importance in sleep-waking states. Prog Brain Res 98:61–71

Karachi C, Grabli D, Bernard FA, Tande D, Wattiez N, Belaid H, Bardinet E, Prigent A, Nothacker HP, Hunot S, Hartmann A, Lehericy S, Hirsch EC, Francois C (2010) Cholinergic mesencephalic neurons are involved in gait and postural disorders in Parkinson disease. J Clin Invest 120:2745–2754

Karachi KL, André A, Tandé D, Hirsch EC, Francois C (2014) The mesencephalic locomotor region integrates motor, cognitive, and emotional information: an anatomical substrate for differential roles of the pedunculopontine and the cuneiform nuclei. In: 44th annual meeting of the Society for Neuroscience, Chicago

Kempster PA, O'Sullivan SS, Holton JL, Revesz T, Lees AJ (2010) Relationships between age and late progression of Parkinson's disease: a clinico-pathological study. Brain 133:1755–1762

Kojima J, Yamaji Y, Matsumura M, Nambu A, Inase M, Tokuno H,

Takada M, Imai H (1997) Excitotoxic lesions of the pedunculopontine tegmental nucleus produce contralateral hemiparkinsonism in the monkey. Neurosci Lett 226:111–114

Lai YY, Siegel JM (1990) Muscle tone suppression and stepping produced by stimulation of midbrain and rostral pontine reticular formation. J Neurosci 10:2727–2734

Lau B, Welter ML, Belaid H, Fernandez Vidal S, Bardinet E, Grabli D, Karachi C (2015) The integrative role of the pedunculopontine nucleus in human gait. Brain 138:1284–1296

Lavoie B, Parent A (1994a) Pedunculopontine nucleus in the squirrel monkey: distribution of cholinergic and monoaminergic neurons in the mesopontine tegmentum with evidence for the presence of glutamate in cholinergic neurons. J Comp Neurol 344:190–209

Lavoie B, Parent A (1994b) Pedunculopontine nucleus in the squirrel monkey: projections to the basal ganglia as revealed by anterograde tract-tracing methods. J Comp Neurol 344:210–231

Longo VG (1956) Effects of scopolamine and atropine electroencephalographic and behavioral reactions due to hypothalamic stimulation. J Pharmacol Exp Ther 116:198–208

Lu J, Sherman D, Devor M, Saper CB (2006) A putative flip-flop switch for control of REM sleep. Nature 441:589–594

MacLaren DA, Santini JA, Russell AL, Markovic T, Clark SD (2014) Deficits in motor performance after pedunculopontine lesions in rats– impairment depends on demands of task. Eur J Neurosci 40:3224–3236

Manaye KF, Zweig R, Wu D, Hersh LB, De Lacalle S, Saper CB, German DC (1999) Quantification of cholinergic and select noncholinergic mesopontine neuronal populations in the human brain. Neuroscience 89:759–770

Martinez-Gonzalez C, Bolam JP, Mena-Segovia J (2011) Topographical organization of the pedunculopontine nucleus. Front Neuroanat 5:22

Martinez-Gonzalez C, Wang HL, Micklem BR, Bolam JP, Mena-Segovia J (2012) Subpopulations of cholinergic, GABAergic and glutamatergic neurons in the pedunculopontine nucleus contain calcium-binding proteins and are heterogeneously distributed. Eur J Neurosci 35:723–734

Masilamoni GJ, Bogenpohl JW, Alagille D, Delevich K, Tamagnan G, Votaw JR, Wichmann T, Smith Y (2011) Metabotropic glutamate receptor 5 antagonist protects dopaminergic and noradrenergic neurons from degeneration in MPTP-treated monkeys. Brain 134:2057–2073

Matsumura M, Kojima J (2001) The role of the pedunculopontine tegmental nucleus in experimental parkinsonism in primates. Stereotact Funct Neurosurg 77:108–115

Matsumura M, Nambu A, Yamaji Y, Watanabe K, Imai H, Inase M, Tokuno H, Takada M (2000) Organization of somatic motor inputs from the frontal lobe to the pedunculopontine tegmental nucleus in the macaque monkey. Neuroscience 98:97–110

Mazzone P, Lozano A, Stanzione P, Galati S, Scarnati E, Peppe A, Stefani A (2005) Implantation of human pedunculopontine nucleus: a safe and clinically relevant target in Parkinson's disease. NeuroReport 16:1877–1881

McCrea DA, Rybak IA (2008) Organization of mammalian locomotor rhythm and pattern generation. Brain Res Rev 57:134–146

Mena-Segovia J, Micklem BR, Nair-Roberts RG, Ungless MA, Bolam JP (2009) GABAergic neuron distribution in the pedunculopontine nucleus defines functional subterritories. J Comp Neurol 515:397–408

Mestre TA, Sidiropoulos C, Hamani C, Poon YY, Lozano AM, Lang AE, Moro E (2016) Long-term double-blinded unilateral pedunculopontine area stimulation in Parkinson's disease. Mov Disord 31:1570–1574

Mesulam MM, Mufson EJ, Wainer BH, Levey AI (1983) Central cholinergic pathways in the rat: an overview based on an alternative

nomenclature (Ch1-Ch6). Neuroscience 10:1185-1201

Mesulam MM, Mufson EJ, Levey AI, Wainer BH (1984) Atlas of cholinergic neurons in the forebrain and upper brainstem of the macaque based on monoclonal choline acetyltransferase immunohistochemistry and acetylcholinesterase histochemistry. Neuroscience 12:669–686

Mesulam MM, Geula C, Bothwell MA, Hersh LB (1989) Human reticular formation: cholinergic neurons of the pedunculopontine and laterodorsal tegmental nuclei and some cytochemical comparisons to forebrain cholinergic neurons. J Comp Neurol 283:611–633

Mori S, Sakamoto T, Ohta Y, Takakusaki K, Matsuyama K (1989) Sitespecific postural and locomotor changes evoked in awake, freely moving intact cats by stimulating the brainstem. Brain Res 505:66–74

Moro E, Hamani C, Poon YY, Al-Khairallah T, Dostrovsky JO, Hutchison WD, Lozano AM (2010) Unilateral pedunculopontine stimulation improves falls in Parkinson's disease. Brain 133:215–224

MunroDavies LE, Winter J, Aziz TZ, Stein JF (1999) The role of the pedunculopontine region in basal-ganglia mechanisms of akinesia. Exp Brain Res 129:511–517

Nandi D, Aziz TZ, Giladi N, Winter J, Stein JF (2002) Reversal of akinesia in experimental parkinsonism by GABA antagonist microinjections in the pedunculopontine nucleus. Brain 125:2418–2430

Olivier A, Parent A, Poirier LJ (1970) Identification of the thalamic nuclei on the basis of their cholinesterase content in the monkey. J Anat 106:37–50

Ovadia A, Zhang Z, Gash DM (1995) Increased susceptibility to MPTP toxicity in middle-aged rhesus monkeys. Neurobiol Aging 16:931–937

Pahapill PA, Lozano AM (2000) The pedunculopontine nucleus and Parkinson's disease. Brain 123:1767–1783

Pare D, Smith Y, Parent A, Steriade M (1988) Projections of brainstem core cholinergic and non-cholinergic neurons of cat to intralaminar and reticular thalamic nuclei. Neuroscience 25:69–86

Peppe A, Pierantozzi M, Baiamonte V, Moschella V, Caltagirone C, Stanzione P, Stefani A (2012) Deep brain stimulation of pedunculopontine tegmental nucleus: role in sleep modulation in advanced Parkinson disease patients: one-year follow-up. Sleep 35:1637–1642

Pereira EA, Nandi D, Jenkinson N, Stein JF, Green AL, Aziz TZ (2011) Pedunculopontine stimulation from primate to patient. J Neural Transm (Vienna) 118:1453–1460

Plaha P, Gill SS (2005) Bilateral deep brain stimulation of the pedunculopontine nucleus for Parkinson's disease. NeuroReport 16:1883–1887

Rinne JO, Ma SY, Lee MS, Collan Y, Roytta M (2008) Loss of cholinergic neurons in the pedunculopontine nucleus in Parkinson's disease is related to disability of the patients. Parkinsonism Relat Disord 14:553–557

Rolland AS, Tande D, Herrero MT, Luquin MR, Vazquez-Claverie M, Karachi C, Hirsch EC, Francois C (2009) Evidence for a dopaminergic innervation of the pedunculopontine nucleus in monkeys, and its drastic reduction after MPTP intoxication. J Neurochem 110:1321–1329

Rolland AS, Karachi C, Muriel MP, Hirsch EC, Francois C (2011) Internal pallidum and substantia nigra control different parts of the mesopontine reticular formation in primate. Mov Disord 26:1648–1656

Romigi A, Placidi F, Peppe A, Pierantozzi M, Izzi F, Brusa L, Galati S, Moschella V, Marciani MG, Mazzone P, Stanzione P, Stefani A (2008) Pedunculopontine nucleus stimulation influences REM sleep in Parkinson's disease. Eur J Neurol 15:e64–e65

Roseberry TK, Lee AM, Lalive AL, Wilbrecht L, Bonci A, Kreitzer AC (2016) Cell-type-specific control of brainstem locomotor circuits by

basal ganglia. Cell 164:526-537

Ryczko D, Auclair F, Cabelguen JM, Dubuc R (2016) The mesencephalic locomotor region sends a bilateral glutamatergic drive to hindbrain reticulospinal neurons in a tetrapod. J Comp Neurol 524:1361–1383

Rye DB (1997) Contributions of the pedunculopontine region to normal and altered REM sleep. Sleep 20:757–788

Rye D (2010) Seeing beyond one's nose: sleep disruption and excessive sleepiness accompany motor disability in the MPTP treated primate. Exp Neurol 222:179–180

Rye DB, Lee HJ, Saper CB, Wainer BH (1988) Medullary and spinal efferents of the pedunculopontine tegmental nucleus and adjacent mesopontine tegmentum in the rat. J Comp Neurol 269:315–341

Schrempf W, Brandt MD, Storch A, Reichmann H (2014) Sleep disorders in Parkinson's disease. J Parkinsons Dis 4:211–221

Sebille SB, Belaid H, Philippe AC, Andre A, Lau B, Francois C, Karachi C, Bardinet E (2016) Anatomical evidence of the functional diversity of the mesencephalic locomotor region in primate. Neuroimage 147:66–78

Shik ML, Severin FV, Orlovskii GN (1966) Control of walking and running by means of electric stimulation of the midbrain. Biofizika 11:659–666

Shink E, Sidibe M, Smith Y (1997) Efferent connections of the internal globus pallidus in the squirrel monkey: II. Topography and synaptic organization of pallidal efferents to the pedunculopontine nucleus. J Comp Neurol 382:348–363

Sirota MG, Di Prisco GV, Dubuc R (2000) Stimulation of the mesencephalic locomotor region elicits controlled swimming in semiintact lampreys. Eur J Neurosci 12:4081–4092

Stefani A, Lozano AM, Peppe A, Stanzione P, Galati S, Tropepi D, Pierantozzi M, Brusa L, Scarnati E, Mazzone P (2007) Bilateral deep brain stimulation of the pedunculopontine and subthalamic nuclei in severe Parkinson's disease. Brain 130:1596–1607

Steriade M, Pare D, Parent A, Smith Y (1988) Projections of cholinergic and non-cholinergic neurons of the brainstem core to relay and associational thalamic nuclei in the cat and macaque monkey. Neuroscience 25:47–67

Steriade M, Datta S, Pare D, Oakson G, Curro Dossi RC (1990) Neuronal activities in brain-stem cholinergic nuclei related to tonic activation processes in thalamocortical systems. J Neurosci 10:2541– 2559

Steriade M, Dossi RC, Pare D, Oakson G (1991) Fast oscillations (20-40 Hz) in thalamocortical systems and their potentiation by mesopontine cholinergic nuclei in the cat. Proc Natl Acad Sci USA 88:4396–4400

Takakusaki K, Habaguchi T, Ohtinata-Sugimoto J, Saitoh K, Sakamoto T (2003) Basal ganglia efferents to the brainstem centers controlling postural muscle tone and locomotion: a new concept for understanding motor disorders in basal ganglia dysfunction. Neuroscience 119:293–308

Takakusaki K, Saitoh K, Harada H, Okumura T, Sakamoto T (2004) Evidence for a role of basal ganglia in the regulation of rapid eye movement sleep by electrical and chemical stimulation for the pedunculopontine tegmental nucleus and the substantia nigra pars reticulata in decerebrate cats. Neuroscience 124:207–220

Thevathasan W, Coyne TJ, Hyam JA, Kerr G, Jenkinson N, Aziz TZ, Silburn PA (2011) Pedunculopontine nucleus stimulation improves gait freezing in Parkinson disease. Neurosurgery 69:1248–1253

Vilensky JA (1983) Gait characteristics of two macaques, with emphasis on relationships with speed. Am J Phys Anthropol 61:255– 265

Villalba RM, Wichmann T, Smith Y (2014) Neuronal loss in the caudal intralaminar thalamic nuclei in a primate model of Parkinson's disease. Brain Struct Funct 219:381–394

Wailke S, Herzog J, Witt K, Deuschl G, Volkmann J (2011) Effect of controlled-release levodopa on the microstructure of sleep in Parkinson's disease. Eur J Neurol 18:590–596

Wang HL, Morales M (2009) Pedunculopontine and laterodorsal tegmental nuclei contain distinct populations of cholinergic, glutamatergic and GABAergic neurons in the rat. Eur J Neurosci 29:340–358

Webster HH, Jones BE (1988) Neurotoxic lesions of the dorsolateral pontomesencephalic tegmentum-cholinergic cell area in the cat. II. Effects upon sleep-waking states. Brain Res 458:285–302

Welter ML, Demain A, Ewenczyk C, Czernecki V, Lau B, El Helou A, Belaid H, Yelnik J, Francois C, Bardinet E, Karachi C, Grabli D (2015) PPNa-DBS for gait and balance disorders in Parkinson's disease: a double-blind, randomised study. J Neurol 262:1515–1525

Xiang Y, John P, Yakushin SB, Kunin M, Raphan T, Cohen B (2007) Dynamics of quadrupedal locomotion of monkeys: implications for central control. Exp Brain Res 177:551–572

Yong MH, Fook-Chong S, Pavanni R, Lim LL, Tan EK (2011) Case control polysomnographic studies of sleep disorders in Parkinson's disease. PLoS One 6:e22511

Zhang JH, Sampogna S, Morales FR, Chase MH (2005) Age-related changes in cholinergic neurons in the laterodorsal and the pedunculopontine tegmental nuclei of cats: a combined light and electron microscopic study. Brain Res 1052:47–55

Zweig RM, Jankel WR, Hedreen JC, Mayeux R, Price DL (1989) The pedunculopontine nucleus in Parkinson's disease. Ann Neurol 26:41–46

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