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1 **Silver ion chromatography for peak resolution enhancement: Application to the**  
2 **preparative separation of two sesquiterpenes using online heart-cutting LC-LC**  
3 **technique**

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29 **Abstract**

30 Silver ion chromatography, utilizing columns packed with silver ions bonded to silica gel, has  
31 proved to be an invaluable technique for the analysis of some positional isomers. In this work,  
32 silver ion chromatography by combination with online heart-cutting LC-LC technique for the  
33 preparative separation of two sesquiterpenes positional isomers from a natural product was  
34 investigated. On the basis of the evaluation that silver ion content impacts on the separation, the  
35 laboratory-made silver ion columns, utilizing silica gel impregnated with 15% silver nitrate as  
36 column packing materials, were used for peak resolution improvement of these two isomers and  
37 the preparative separation of them in heart-cutting LC-LC. The relationship among the maximal  
38 sample load, flow rate and peak resolution in the silver ion column were optimized, and the  
39 performance of the silver ion column was compared with conventional C<sub>18</sub> column and silica gel  
40 column. Based on the developed chromatographic conditions, online heart-cutting LC-LC  
41 chromatographic separation system in combination with a silica gel column and a silver ion  
42 column that was applied to preparative separation of these two isomers from a traditional Chinese  
43 medicine, *Inula racemosa* Hook.f., was established. The results showed that the online  
44 heart-cutting LC-LC technique by combination of a silica gel column and a silver ion column for  
45 the preparative separation of these two positional isomers from this natural plant was superior to  
46 the preparative separation performed on a single-column system with C<sub>18</sub> column or silica gel  
47 column.

48 **Keywords:** Positional isomers; Argentation; Peak resolution enhancement; Heart-cutting LC-LC;  
49 Preparative separation; Natural product

50 **1. Introduction**

51 Silver ion chromatography, utilizing columns packed with silver ions bonded to a silica gel  
52 or similar substrate, has proved to be an invaluable technique for the analysis of complex  
53 triacylglycerol mixtures, fatty acids, lipids and some positional isomers [1-4]. This application  
54 based on the mechanism that the different number, or/and geometrical configuration, or/and  
55 position of unsaturated C-C bonds provide the different amount of  $\pi$  donors. Since silver ions act  
56 as  $\pi$  acceptors while unsaturated C-C bonds act as  $\pi$  donors, causing molecules that contain fewer  $\pi$   
57 donors or do not contain  $\pi$  donors to be eluted firstly on silver ion chromatography [5].

58           Although silver ion chromatography by a single-column system is inexpensive and easy to  
59   operate for the trace separation and determination of active ingredients, or preparative separation  
60   compounds from simple sample on small-scale [6], it still restricts to the large-scale preparative  
61   separation of compounds from complex natural products. Due to adsorption of excessive  
62   impurities, the action of silver ions becomes worse, causing the decrease of column separation  
63   efficiency, even resulting in bleeding of silver ions and affecting the UV detector. In addition, a  
64   single-column system sometimes has limited sample capacity and poor relative peak separation [7].  
65   Notably, heart-cutting two-dimensional liquid chromatography (LC-LC) offers the probability to  
66   improve these drawbacks based on a tandem combination of two independent liquid phase  
67   separation systems [8-11].

68           *Inula racemosa* Hook.f., a medicinal plant, is widely distributed in Europe, Asia and Africa  
69   [12]. Its root as a traditional medicine has been most frequently applied for issues related to peptic  
70   disorders, phlegm, detumescence, inflammatory and vermifuge [13]. Alantolactone (AL) and  
71   isoalantolactone (IS) (Figure.1) are two sesquiterpenes positional isomers and have abundant  
72   content in this medicinal plant. Previous studies suggested that they show many biological  
73   activities [14-25], especially in killing of cancer cells. In addition, researchers also have examined  
74   the relationship between their structural modifications and biological activities [26-28]. These  
75   studies suggested that AL and IS may be good potential lead compounds for future anticancer  
76   agent development or act as chemical templates for the design, synthesis, and semisynthesis of  
77   new substances. Nevertheless, further studies are required in preclinical and clinical applications  
78   and to explain their potential role. For these studies, it certainly will need a large number of  
79   materials. In the current ways, acquisition of AL and IS from natural plants is a short-cut.  
80   Therefore, a robust method for the preparative separation of AL and IS from this medicinal plant  
81   may be necessary. Unfortunately, the structural similarity of AL and IS increases the preparative  
82   separation difficulty of them from this natural plant. Although the traditional separation methods  
83   used in the trace separation and determination of these two compounds have been performed with  
84   capillary electrophoresis (CE) [29], gas chromatography (GC) [30] and RP-HPLC [31, 32], when  
85   these methods are used to isolate AL and IS from *Inula racemosa* Hook.f. on a larger scale, it  
86   becomes difficult due to the limitation of the resolution between their peaks. Hence, the

87 establishment of an effective method for increase of the peak resolution and its application to their  
88 preparative separation from this plant becomes a key problem. Since AL and IS constitute a pair of  
89 positional isomers related to C=C bond, silver ion chromatography provides the probability to  
90 improve the separation of them (Figure.1). Therefore, the aim of this study was to improve the  
91 separation of these two isomers by silver ion chromatography in preparative scale and obtain them  
92 from a crude extract of *Inula racemosa* Hook.f. by preparative separation with silver ion  
93 chromatography technique in combination with online heart-cutting LC-LC technique.

## 94 **2. Experimental**

### 95 **2.1. Materials and reagents**

96 The dried roots of *Inula racemosa* Hook.f. were purchased from Bai Ding (Tibet, China).  
97 Alantolactone and isoalantolactone were purchased from National Institute of Food and Drug  
98 Control for the quantification and qualitative study (Beijing, China). Silver nitrate was of chemical  
99 grade (Zhengzhou Kaidi Chemical Products Co., Ltd, Henan, China). Silica gel (5 ~ 10  $\mu\text{m}$  particles  
100 and 10 ~ 40  $\mu\text{m}$  particles) was purchased from Qingdao Yida Silica Reagent Factory (Fujian, China).  
101 Ethyl acetate, n-hexane, 95% ethanol and acetonitrile reagents were of analytical grade and  
102 purchased from Xi'an Chemical Reagent Factory (Shaanxi, China).

### 103 **2.2. Sample preparation**

104 The dried roots of *Inula racemosa* Hook.f. were pulverized and sieved through a screen (100  
105 ~ 200 mesh). 100 g of this powder was extracted with 1000 ml of 95% ethanol solution by  
106 ultrasonic treatment for 30 min at 45 °C and 100 kHz/450 W (Bandelin Sonorex, Germany) [33]  
107 and repeated three times. The extracts were concentrated on a rotavapor at 45 °C (REC32E,  
108 Shanghai Yarong Instrument, China). The residues were dissolved in n-hexane/ethyl acetate (100  
109 mL: 70/30, v/v) solution to get sample solutions for the preparative separation, and the contents of  
110 AL and IS in this sample solution were 25.62 mg/mL and 31.32 mg/mL, respectively.

111 The stock solution for the analysis of AL and IS was prepared by dissolving AL and IS into  
112 n-hexane/ethyl acetate (70/30, v/v) and mixing at concentration containing AL of 25.86 mg/mL  
113 and IS of 30.26 mg/mL.

### 114 **2.3. Column packing**

115 The laboratory-made silver ion column, utilizing silica gel impregnated with silver nitrate as

116 column packing materials, was prepared for the preparative separation in heart-cutting LC-LC.  
117 The silica gel impregnated with silver nitrate was prepared by the method reported previously with  
118 slightly modification [34] (Figure.2). To avoid waste and the risk of blindness, the silver ion content  
119 in silica gel impregnated with silver nitrate for the impact on the separation of these two isomers  
120 was first observed on silver ion TLC plate [34]. Then the silver ion columns (Stainless steel tube,  
121 Jiangsu Hanbang Co. Ltd., China) were slurry packed with ethyl acetate using a CST  
122 chromatographic column packing machine (KeSheng Experimental Equipment Co., Ltd., Suzhou,  
123 China) according to the operating instructions, and the performance of the prepared silver ion  
124 column for the preparative separation of these two isomers was investigated and compared with  
125 C<sub>18</sub> column on preparative HPLC instrument (Two Alltech-627 high-pressure pumps equipped  
126 with a Model-500 UV-VIS detector, Alltech, USA).

#### 127 **2.4. Connections of the heart-cutting LC-LC system**

128 The connections of the heart-cutting LC-LC system are illustrated in Figure.3. The column  
129 temperatures were controlled at 30 °C using an independent QT-330 chromatographic column  
130 thermostat (Henan Jiuhegongchuang co., Ltd., China). Two NP7000-C high-pressure pumps  
131 equipped with a NU3010-C UV-VIS detector (Jiangsu Hanbang Technology Co., Ltd., Jiangsu,  
132 China) were involved in the first dimension, and the UV detector was set at 260 nm. The system  
133 control and data collection were carried out using an EasyChrom-1000 chromatographic  
134 workstation. A silica gel column (22 mm i.d. × 250 mm, 5 μm, Grace, USA) as a preparative  
135 column was used for the first-dimension separation, while a 22 mm i.d. × 20 mm silica gel column  
136 as pre-column was equipped between the sample loop and the preparative column. In addition,  
137 two Alltech-627 high-pressure pumps equipped with a Model-500 UV-VIS detector (Alltech, USA)  
138 were used in the second dimension, and the UV detector was set at 260 nm. The system control and  
139 data acquisition were performed by using an AllChrom Plus server workstation. The  
140 second-dimension separation was performed on a laboratory-made silver ion column (22 mm i.d. ×  
141 250 mm).

#### 142 **2.5. Procedure for preparative separation**

143 *First-dimension unit separation*– N-hexane (A)/ethyl acetate (B) (v/v) were employed as the  
144 mobile phase. 10 mL of the sample solution was injected into a 20 mL sample loop while valve A

145 was set at position B. Then the elution program was carried out by the EasyChrom-1000  
146 chromatographic workstation: 0–15 min (Valve A at position A; Flow rate, 12 mL/min; 85%  
147 A/15% B; When the peak of the fraction containing AL and IS started to emerge, valve B and  
148 valve A were all switched to position B: the fraction containing AL and IS was cut and transferred  
149 onto the silver ion column via the automatic FC-AS-312 component collector), and 15.1–25 min  
150 (Valve A at position B; flow rate, 15 mL/min; 0% A/100% B) and 25.1–30 min (Valve A at  
151 position B; flow rate, 15 mL/min; 85% A/15% B).

152 *Second-dimension unit separation*– When all of the fraction containing AL and IS from the  
153 first-dimension column had been completely transferred onto the second-dimension column, the  
154 following elution program was run with a mobile phase composition of (A) n-hexane/(B) ethyl  
155 acetate: 0–22 min (Valve B at position A; Flow rate, 10 mL/min; 75% A/25% B; AL and IS were  
156 automatically collected by the component collector on the second-dimension unit), 22.1–25 min  
157 (Valve B at position B; Flow rate, 15 mL/min; 0% A/100% B) and 27.1–32 min (Valve B at  
158 position B; Flow rate, 15 mL min<sup>-1</sup>; 75% A/25% B).

## 159 **2.6. Quantitative HPLC analysis**

160 The crude *Inula racemosa* Hook.f. extract and the prepared compounds were analyzed by  
161 a commercially available silver ion column (4.6 mm i.d. × 250 mm, ChromSpher Lipids CP28313,  
162 Varian, USA) in the analytical HPLC instrument (Waters 2695 HPLC system equipped with a  
163 Waters 2487 UV-VIS detector controlled through an Empower chromatographic workstation,  
164 Waters, USA). The concentration range of calibration solutions obtained by diluting the stock  
165 solutions at the range of 5172.0-323.25 µg/mL for AL and 6052.0-378.25 µg/mL for IS.  
166 Chromatographic separation was performed using an isocratic elution at a flow rate of 1.0  
167 mL/min with mobile phase 75/25 (n-hexane/ethyl acetate). The column temperature was controlled  
168 at 30 °C, and the effluents were monitored at 260 nm by a UV detector.

## 169 **3. Results and discussion**

### 170 **3.1. Silver ion content in the improvement of separation**

171 For the preparative separation of AL and IS, the goal in the present study was to obtain them  
172 with high efficiency. Since low sample load reduces the efficiency in preparative separation of AL  
173 and IS, we were concerned about the increase of the maximal sample load. It is well known that

174 the resolution has great influence on the maximal sample load, as well as run times, equilibration  
175 times and solvent consumption. With increase of sample load, the resolution decreases [35]. The  
176 decreased resolution limits the increase of the maximal sample load and throughput per unit time.  
177 Therefore, the enhancement of the resolution between AL and IS may improve the maximal  
178 sample load.

179 The amount of silver ion in the silica gel impregnated with silver nitrate is vital for the  
180 improvement of the resolution. Therefore, the investigation on the content of silver nitrate may be  
181 necessary. TLC technique in the evaluation of the effect of silver ion content is easy and  
182 convenient in this study. In the TLC separation, the developing solvent was composed of  
183 n-hexane/ethyl acetate (80/20). The TLC plates were sprayed with an ethanol solution containing  
184 10% sulfuric acid and then heated in an IR drier until obvious colour appeared. As shown in Fig.4,  
185 AL and IS could not be isolated completely on the TLC plates when no silver nitrate added into  
186 silica gel (Figure.4-1 and Figure.4-3), whereas these two isomers on TLC plates by using silica gel  
187 (10 ~ 40  $\mu\text{m}$  particles) impregnated with silver nitrate showed a good separation (Figure.4-2 and  
188 Figure.4-4 - Figure.4-9). Since the level of silver nitrate impregnation recommended has varied  
189 from 0.5-40% [36] and the case where we have found 10% of impregnation is more effective for  
190 separations of isomers in column separation [37], 6%, 10% and 15% silver nitrate to improve the  
191 separation were investigated in the present study. From the obtained thin-layer chromatograms  
192 (Figure.4), with the increase of the silver nitrate content, the amount of sample load increased with  
193 complete separation. Since 15% silver nitrate facilitated the isolation more effectively than other  
194 ratios with the increase of sample load (Figure.4-3 - Figure.4-9), silica gel (5 ~ 10  $\mu\text{m}$  particles)  
195 impregnated with 15% silver nitrate as column packing materials was prepared in the following  
196 study. Whether more than 15% silver nitrate is more effective in preparative separation of these  
197 two isomers needs to be explored.

### 198 **3.2. Silver ion column for the separation of two sesquiterpenes positional isomers**

199 Even though these two positional isomers achieved good separation on silver ion TLC plates  
200 with 15% of impregnation, whether the preparative separation in HPLC, utilizing silver ion  
201 column packed with this impregnation (15% silver nitrate), could acquire improvement was  
202 unknown. Based on the TLC analysis, the investigation was first performed on a single



203 laboratory-made 4.6 mm i.d. × 250 mm silver ion column with mobile phase 80/20  
204 (n-hexane/ethyl acetate, v/v) at the flow rate of 1 mL/min, where it obtained a good peak  
205 resolution, and a satisfactory peak shape and separation times within 20 μL of the stock solution  
206 (Figure.5a), whereas peak resolution decreased when the sample load increased to 80 μL of the  
207 stock solution (Figure.5b). Since the mobile phase with n-hexane/ethyl acetate (80/20, v/v)  
208 shortened their separation times and reduction of ethyl acetate concentration (Such as reduction of  
209 5% ethyl acetate) in mobile phase has a great influence on the retention times, the optimization of  
210 the mobile phase composition for the increase of sample load was not carried out. Furthermore,  
211 considering that reduction of the flow rate may improve the peak resolution and then increase the  
212 sample load [38], the mobile phase 80/20 (n-hexane/ethyl acetate, v/v) at the flow rate of 0.8  
213 mL/min and 0.6 mL/min were evaluated in the following experiment, respectively. When the flow  
214 rate decreased to 0.8 mL/min, a complete resolution could be obtained within 180 μL of the stock  
215 solution (Figure.6), while the retention times of these two isomers slightly delayed but there was  
216 no obvious changes in the peak shape. When the sample load continued increasing, bad resolution  
217 appeared (Figure.6). In a view of this, the flow rate reduced to 0.6 mL/min, in which the maximal  
218 sample load increased to 480 μL of the stock solution with a complete resolution (Figure.5c and  
219 Figure.6). However, the peaks seem to be flat and trailing, and the retention times delayed  
220 markedly (Figure.5c). The reason may be the decrease in the elution strength of solvent at the low  
221 flow rate. Therefore, the mobile phase was optimized for n-hexane/ethyl acetate (75/25), which  
222 reduced peaks trailing and retention times (Figure.5d) and given a complete resolution.

223 Fig.6 shows the relationship among the resolution, sample load and flow rate. With the  
224 increase of sample load, the peak resolution decreased. The increased flow rate also reduced the  
225 peak resolution and led to the decrease of the peak height. Although the decrease of the flow  
226 rate made peak width increase slightly, it obtains a good resolution ( $\geq 1.5$ ) with the increase of  
227 sample load. When the sample volume increased to 480 μL of the stock solution, the resolution  
228 between the AL and IS nearly equals to 1.5 at the flow rate of 0.6 mL/min, so the sample load  
229 didn't increase any more.

230 In order to validate the recovery and repeatability under the developed conditions using the  
231 prepared silver column, 480 μL of the stock solution was separated on the 4.6 mm i.d. × 250 mm

232 column with n-hexane/ethyl acetate (75/25) at the flow rate of 0.6 mL/min. By six replicate  
233 experiments, the recovery of AL was 95.3% with RSD of 1.3% and that of IS was 95.1% with  
234 RSD of 1.6%, while the repeatability was determined with the values of RSD of 1.09% and 1.23%.  
235 The results showed a satisfactory recovery and repeatability.

### 236 **3.3. Conventional separation methods**

237 C<sub>18</sub> and silica gel packing materials are commonly used for preparative separation. Evidently,  
238 compared with silica gel, silica gel impregnated with silver nitrate achieved good separation with  
239 the increase of sample load (Figure.4). To further evaluate the performance of the laboratory-made  
240 silver ion column (4.6 mm i.d. × 250 mm), a same size C<sub>18</sub> column (4.6 mm i.d. × 250 mm) for  
241 preparative separation of AL and IS was compared with the silver ion column (Chromatographic  
242 conditions were presented in supplementary material). Although a satisfactory separation achieved  
243 on C<sub>18</sub> column using a higher concentration of water in the mobile phase, it sacrificed a lot of time  
244 and still had limited sample treatment capacity because of the limitation of the peak resolution.  
245 However, the silver ion column reduced the retention times and increased the maximal sample  
246 load with good resolution, compared to C<sub>18</sub> column (Table. 1 and Figure.7).

### 247 **3.4. Application of heart-cutting LC-LC system in the preparative separation**

248 The online heart-cutting LC-LC technique was applied to the preparative separation of these  
249 two positional isomers from this natural plant. Although AL and IS could not be separated  
250 completely with silica gel materials, the fraction of AL and IS achieved good separation  
251 (Figure.4-1 and Figure.8a). Therefore, a silica gel columns was equipped on the first-dimension  
252 separation to isolate the fraction of AL and IS from the crude extract, while a laboratory-made 22  
253 mm i.d. × 250 mm silver ion column was selected for the second-dimension separation to further  
254 separate AL and IS. The elution conditions both in the first dimension and the second dimension  
255 were developed based on the above analysis. The maximal sample load and the flow rate for the  
256 selected silver ion column in LC-LC separation were estimated by applying the linear  
257 amplification analysis (About 10.9 mL of the stock solution and flow rate of 14 ml/min) [39], and  
258 then the diameter of the silica gel column used in first-dimension separation was chosen based on  
259 the requirement for making full use of the selected silver ion column in the advantage of the high  
260 sample load. For this reason, if 11 mL of the crude extract solution can be treated by silica gel

261 column equipped on the first dimension, the sample load amount loaded on the selected silver ion  
262 column transferred from the first dimension may be close to its maximal sample load in LC-LC  
263 preparative separation. Therefore, two silica gel columns, with different diameters (10 mm i.d. ×  
264 250 mm, 22 mm i.d. × 250 mm), were tested respectively. The results showed that the 22 mm i.d.  
265 × 250 mm column was superior to another one, in which it could treat 10 mL of the crude extract  
266 solution with n-hexane/ethyl acetate (85/15, v/v) at the flow of 12 mL/ min (Figure.8b-d). Hence,  
267 a 22 mm i.d. × 250 mm silica gel column with n-hexane/ethyl acetate (85/15, v/v) as eluent at the  
268 flow of 12 mL/ min was used for the first-dimension separation.

269 Although 10 mL of the crude extract solution could be treated in the first-dimension  
270 separation (Figure.9a), a bad separation appeared in the second-dimension separation with  
271 n-hexane/ethyl acetate (75/25, v/v) at the flow of 14 mL/ min. However, when the flow rate in the  
272 second-dimension separation decreased to 10 mL/min, the peaks of these two isomers obtained a  
273 baseline separation (Figure.9b). In addition, to maintain the column separation performance per  
274 unit time, the columns restored by elution using 100% ethyl acetate at the end of separation each  
275 time. Using multicycle and consecutive separation, 2.3 g of AL with a purity of 97.8% and 2.8 g  
276 of IS with a purity of 98.8% were obtained from of the crude extract within 8.5 hours. In an  
277 LC-LC separation model, the preparative separation achieved high-efficiency compared with  
278 conventional methods.

279 Moreover, the crystals obtained from 90% ethanol were analysed by X-ray diffraction. Full  
280 crystallographic details for the compounds reported in this paper were deposited in the Cambridge  
281 Crystallographic Data Centre as Supplementary Publication No. CCDC 1438651 for AL (1) [40]  
282 and No. CCDC 1438652 for IS (2) [41]. Copies of these data can be obtained upon application to the  
283 CCDC, 12 Union Road, Cambridge, CB2 1EZ, UK [<http://www.ccdc.cam.ac.uk>].

#### 284 **4. Conclusion**

285 Silver ion chromatography was demonstrated to be an effective technique for improvement of  
286 the separation of AL and IS in preparative scale. Online heart-cutting LC-LC technique by  
287 combination of the silica gel column and silver ion column was successfully applied to preparative  
288 separation of AL and IS from a crude extract of *Inula racemosa* Hook.f. This study offers a rapid  
289 and effective approach for the preparative separation of these two positional isomers from natural

290 product.

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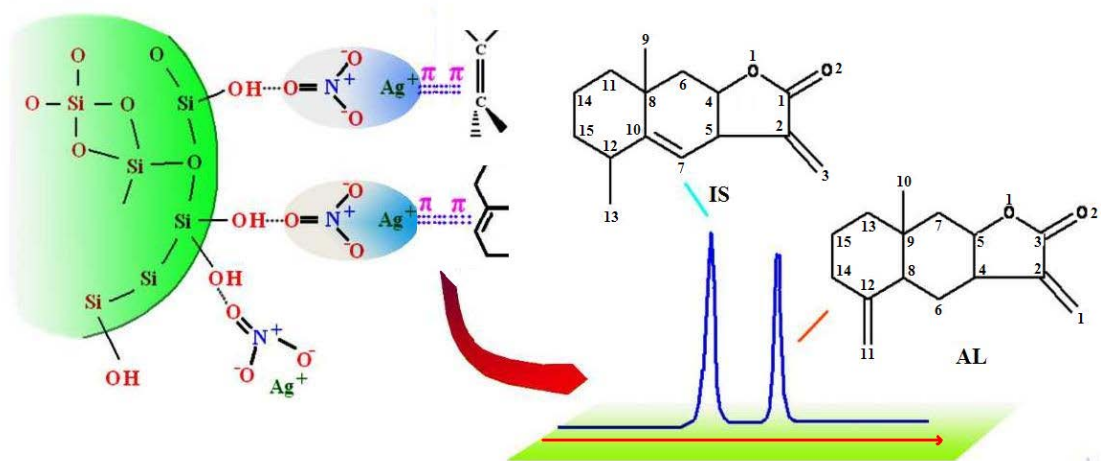


Figure.1 Schematic illustration of silver ion chromatography for the separation of alantolactone (AL) and isoalantolactone (IS), and their chemical structures.

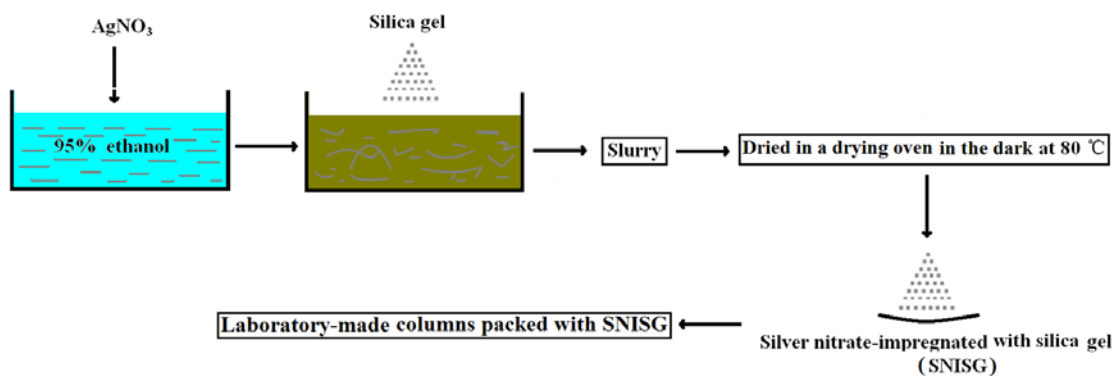


Figure.2. Schematic illustration of the preparation of silver ion columns packed with silica gel impregnated with silver nitrate (SNISG).

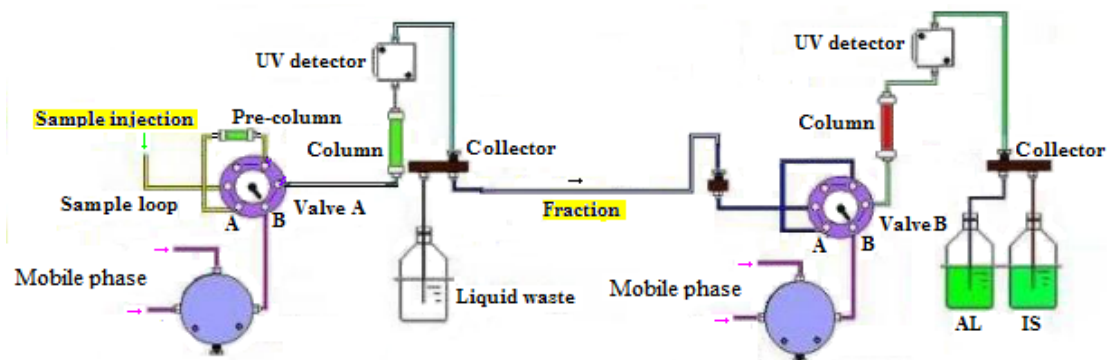


Figure.3. Schematic illustration of the instrumental configurations used for online heart-cutting LC-LC preparative separation.



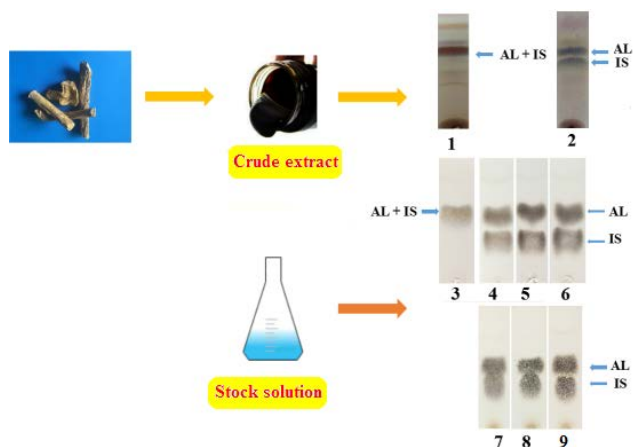


Figure.4. 0% (1, 3), 6% (4, 7), 10% (5, 8) and 15% (6, 9) silver nitrate as additives: (3-6), 10  $\mu$ L of the stock solution was separated on silver ion TLC plates; (7-9), 20  $\mu$ L of the stock solution was separated on silver ion TLC plates; (1), the crude extract was separated on silica gel TLC plate; (2), the crude extract was separated on silver ion TLC plate (Impregnation with 6% silver nitrate).

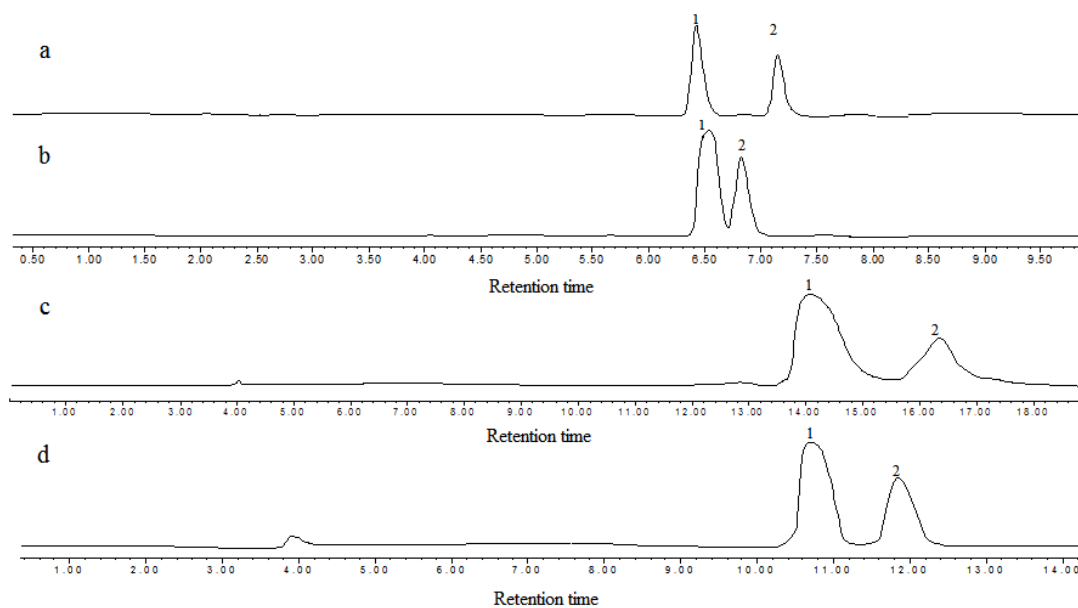


Figure.5. (1) AL and (2) IS. (a) 20  $\mu$ L of the stock solution was separated with mobile phase 80/20 (v/v) for n-hexane/ethyl acetate at the flow of 1 mL/min; (b) 80  $\mu$ L of the stock solution was separated with mobile phase 80/20 (v/v) for n-hexane/ethyl acetate at the flow of 1 mL/min; (c) 480  $\mu$ L of the stock solution was separated with mobile phase 80/20 (v/v) for n-hexane/ethyl acetate at the flow of 0.6 mL/min; (d) 480  $\mu$ L of the stock solution was separated with mobile phase 75/25 (v/v) for n-hexane/ethyl acetate at the flow of 0.6 mL/min. Detection: UV 260 nm. Column: Laboratory-made 4.6 mm i.d.  $\times$  250 mm silver ion column.

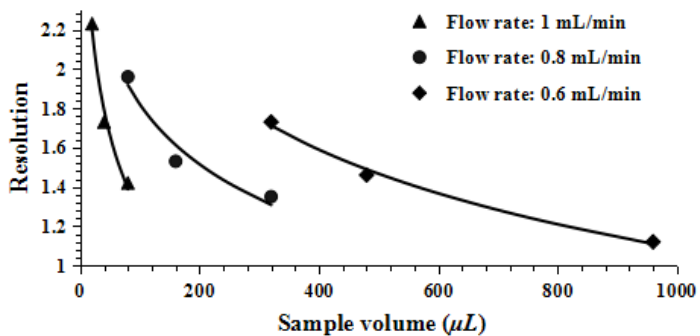


Figure. 6. The relationship among the resolution, sample load and flow rate. Column: Laboratory-made 4.6 mm i.d. × 250 mm sliver ion column. Mobile phase: n-hexane/ethyl acetate. Detection: UV 260 nm. Sample: the stock solution.

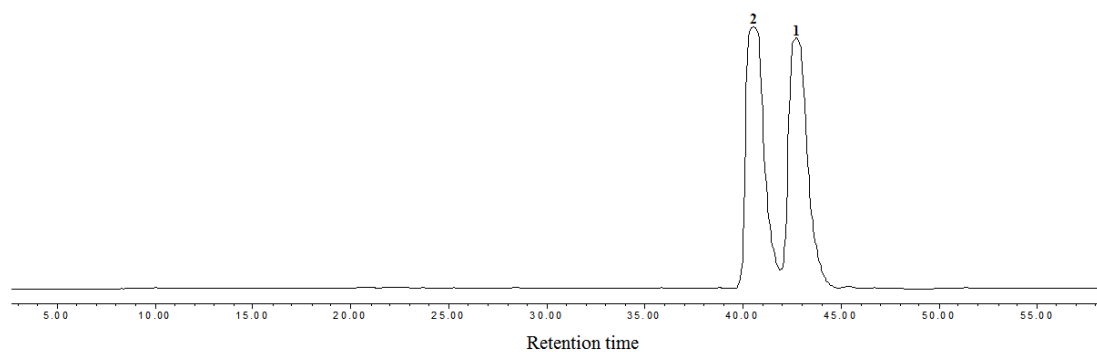


Figure.7. (1) AL and (2) IS. Chromatogram: Separation of AL and IS with mobile phase 65:35 (v/v) for acetonitrile:water. Flow rate: 0.6 mL/min. Detection: UV 220 nm. Column: Commercially available C18 column. Injection volume: 80 μL of the stock solution.

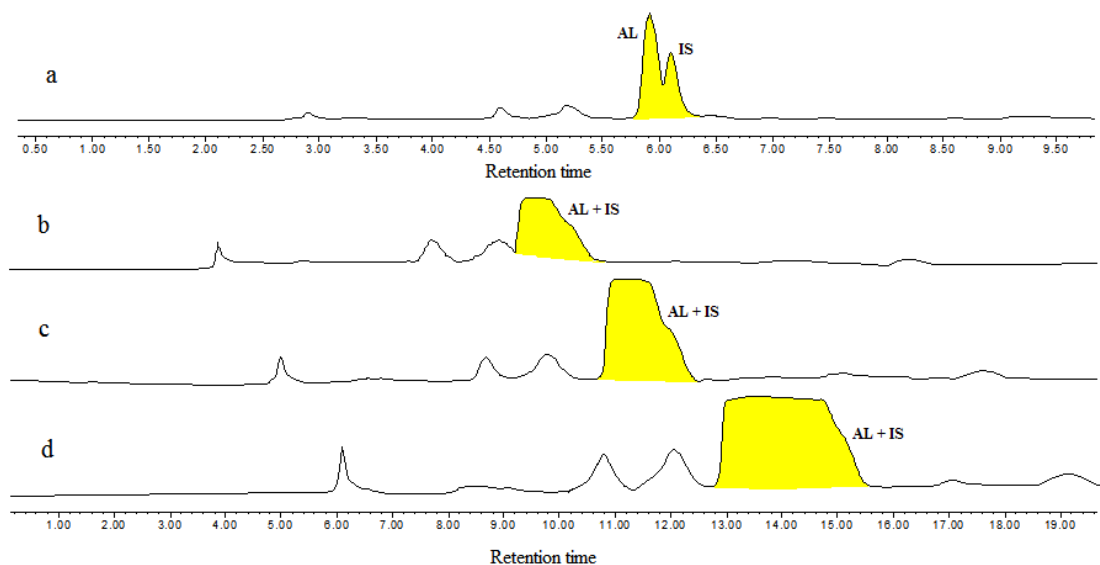


Figure.8. (a) 10  $\mu$ L of the crude extract solution was separated on a 4.6 mm i.d.  $\times$  250 mm column with mobile phase 80/20 (v/v) for n-hexane/ethyl acetate at the flow of 1 mL/min. (b) 6 mL of the crude extract solution was separated on a 10 mm i.d.  $\times$  250 mm column with mobile phase 80/20 (v/v) for n-hexane/ethyl acetate at the flow of 4 mL/min; (c) 6 mL of the crude extract solution was separated on a 22 mm i.d.  $\times$  250 mm column with mobile phase 80/20 (v/v) for n-hexane/ethyl acetate at the flow of 15 mL/min; (d) 10 mL of the crude extract solution was separated on a 22 mm i.d.  $\times$  250 mm column with mobile phase 85/15 (v/v) for n-hexane/ethyl acetate at the flow of 12 mL/min. Detection: UV 260 nm. Column: Commercially available silica gel column.

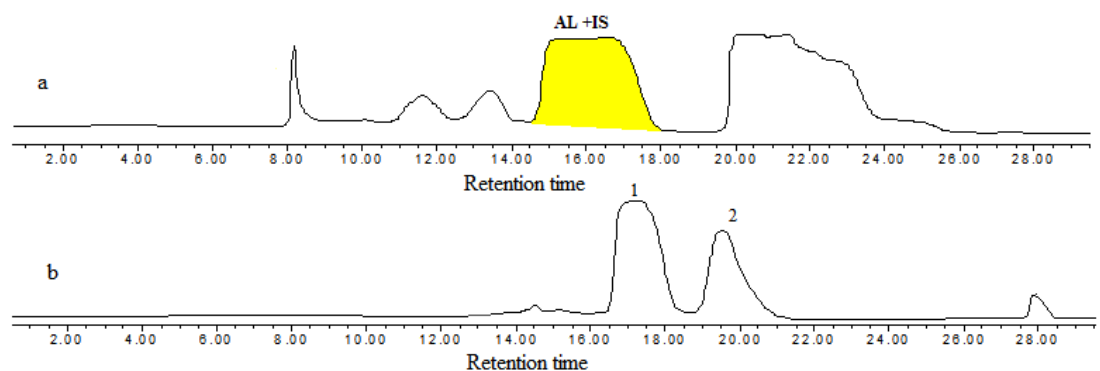


Figure.9. (1) AL and (2) IS. Chromatogram of online heart-cutting LC-LC separation: (a) the first-dimension separation was carried out on a silica gel column (22 mm i.d.  $\times$  250 mm) with mobile phase 85/15 (v/v) for n-hexane/ethyl acetate at the flow of 12 mL/min, and (b) the second-dimension separation was performed on a laboratory-made sliver ion column (22 mm i.d.  $\times$  250 mm) with mobile phase 75/25 (v/v) for n-hexane/ethyl acetate at the flow of 10 mL/min. Detection: UV 260 nm..