



HAL
open science

Trial watch: dietary interventions for cancer therapy

Sarah Levesque, Jonathan G Pol, Gladys Ferrere, Lorenzo Galluzzi, Laurence Zitvogel, Guido Kroemer

► **To cite this version:**

Sarah Levesque, Jonathan G Pol, Gladys Ferrere, Lorenzo Galluzzi, Laurence Zitvogel, et al..
Trial watch: dietary interventions for cancer therapy. *OncoImmunology*, 2019, 8 (7), pp.1591878.
10.1080/2162402X.2019.1591878 . hal-02147212

HAL Id: hal-02147212

<https://hal.sorbonne-universite.fr/hal-02147212v1>

Submitted on 4 Jun 2019

HAL is a multi-disciplinary open access archive for the deposit and dissemination of scientific research documents, whether they are published or not. The documents may come from teaching and research institutions in France or abroad, or from public or private research centers.

L'archive ouverte pluridisciplinaire **HAL**, est destinée au dépôt et à la diffusion de documents scientifiques de niveau recherche, publiés ou non, émanant des établissements d'enseignement et de recherche français ou étrangers, des laboratoires publics ou privés.

Trial watch: dietary interventions for cancer therapy

Sarah Lévesque, Jonathan G. Pol, Gladys Ferrere, Lorenzo Galluzzi, Laurence Zitvogel & Guido Kroemer

To cite this article: Sarah Lévesque, Jonathan G. Pol, Gladys Ferrere, Lorenzo Galluzzi, Laurence Zitvogel & Guido Kroemer (2019) Trial watch: dietary interventions for cancer therapy, OncoImmunology, 8:7, 1591878, DOI: [10.1080/2162402X.2019.1591878](https://doi.org/10.1080/2162402X.2019.1591878)

To link to this article: <https://doi.org/10.1080/2162402X.2019.1591878>



© 2019 The Author(s). Published with license by Taylor & Francis Group, LLC.



Published online: 03 Apr 2019.



Submit your article to this journal [↗](#)



Article views: 300



View Crossmark data [↗](#)

REVIEW



Trial watch: dietary interventions for cancer therapy

Sarah Lévesque^{a,b,c,d,e}, Jonathan G. Pol^{a,b,c,f,g}, Gladys Ferrere^{h,i}, Lorenzo Galluzzi^{f,j,k,l}, Laurence Zitvogel^{d,h,i}, and Guido Kroemer^d 

^aEquipe 11 labellisée par la Ligue Nationale contre le Cancer, Centre de Recherche des Cordeliers, Paris, France; ^bINSERM, U1138, Paris, France; ^cMetabolomics and Cell Biology Platforms, Gustave Roussy Cancer Campus, Villejuif, France; ^dUniversité Paris-Saclay, Orsay, France; ^eFondation pour la Recherche Médicale, Paris, France; ^fUniversité Paris Descartes/Paris V, Sorbonne Paris Cité, Paris, France; ^gUniversité Pierre et Marie Curie/Paris VI, Paris, France; ^hINSERM U1015, Villejuif, France; ⁱCICBT507, Villejuif, France; ^jDepartment of Radiation Oncology, Weill Cornell Medical College, New York, NY, USA; ^kSandra and Edward Meyer Cancer Center, New York, NY, USA; ^lDepartment of Dermatology, Yale School of Medicine, New Haven, CT, USA; ^mPôle de Biologie, Hôpital Européen Georges Pompidou, AP-HP, Paris, France; ⁿDepartment of Women's and Children's Health, Karolinska University Hospital, Stockholm, Sweden

ABSTRACT

Dietary interventions have a profound impact on whole body metabolism, including oncometabolism (the metabolic features allowing cancer cells to proliferate) and immunometabolism (the catabolic and anabolic reactions that regulate immune responses). Recent preclinical studies demonstrated that multiple dietary changes can improve anticancer immunosurveillance of chemo-, radio- and immunotherapy. These findings have fostered the design of clinical trials evaluating the capacity of dietary interventions to synergize with treatment and hence limit tumor progression. Here, we discuss the scientific rationale for harnessing dietary interventions to improve the efficacy of anticancer therapy and present up-to-date information on clinical trials currently investigating this possibility.

ARTICLE HISTORY

Received 28 February 2019
Accepted 5 March 2019

KEYWORDS

Alternate-day fasting; caloric restriction mimetics; chemotherapy; fasting-mimicking diet; immune checkpoint blockers; ketone bodies

Introduction

In the Western world, overnutrition has overcome under-nutrition as a medical and societal problem.^{1–3} Beyond quantitative considerations, it appears that the consumption of ultra-processed food (rich in carbohydrates, sugars, salt, and fat) and soft drinks coupled to a relative scarcity of fruit and vegetables affects the majority of individuals even in high-income countries.^{4–9} Against this background, it is clear that the “normal diet” cannot be appreciated as a statistical norm (i.e., the diet of the average individual), but must be defined by public guidelines. Such guidelines, however, are overshadowed by political decisions and arguable observational epidemiology, meaning that they tend to differ among distinct countries.¹⁰ Moreover, much of the preclinical research done with laboratory animals (mostly mice) is based on the comparison of different types of chemically non-defined regimens, meaning that the conclusions of such studies are often based on methodologically suboptimal approaches.^{11,12} Indeed, in pharmacology, it is common practice to compare different experimental conditions that only differ with respect to the absence and the presence of a drug administered at different concentrations. This kind of rigor is absent from most nutritional studies, which ideally should be designed to test the effects of just one single macro- or micronutrient admixed as a chemically defined entity (e.g., sucrose, sodium chloride, cholesterol or specific vitamins).¹³ Notwithstanding these limitations, it has become clear that the quantity and quality of nutrition plays

a major role in determining the risk of cancer.^{14–16} Obesity is nowadays on the verge of beating tobacco as the principal avoidable risk factor for cancer.^{17–19} Along similar lines, a high variety of food that supplies all necessary micronutrients appears to be one of the principal factors that link high socioeconomic status with low disease risk.²⁰ Finally, multiple animal studies favor the idea that nutritional interventions may curb the progression of established cancers and improve the efficacy of anticancer treatments.^{21–27} These effects rely on the alteration of both oncometabolism (the anabolic and catabolic reactions that support oncogenesis, disease progression and resistance to treatment)^{28,29} and immunometabolism (the metabolic features that regulate immune responses).^{30–34}

Along the lines of our Trial Watch series,^{35,36} we discuss the rationale for harnessing nutritional interventions in support of cancer therapy and the progress of recent clinical trials testing this therapeutic paradigm in cancer patients.

Anticancer effects of dietary interventions – a cell-autonomous rationale

Cancer cells, especially those studied in the laboratory, are characterized by an increase in anabolic reactions that give rise to the so-called Warburg effect, the fact that such cells tend to take up large amounts of glucose even in conditions in which oxidative phosphorylation can proceed in an unlimited fashion.^{37–40} This so-called ‘aerobic glycolysis’ allows glucose-derived carbon atoms to be used for biosynthetic reactions.

Cancer cells also take up large amounts of amino acids through specific transporters in the plasma membrane, acquire increased amounts of proteins by pinocytosis, and even engulf their neighbors to cannibalize them.^{41–43} In an analogous fashion, cancer cells are avid consumers of lipids.⁴⁴ Given their anabolic appetite, it is not surprising that nutritional interventions designed to reduce tumor growth involve a reduction in macronutrient uptake. Thus, it has been shown in mice that short-term starvation (STS, meaning no food supply for 1–2 days with access to drinking water *ad libitum*) and alternate-day fasting (ADF, meaning the alternation of 1-day intervals with and without access to food) can reduce tumor progression.^{25,27} Moreover, the specific depletion of proteins (as well as selected amino acids) from nutrients can be harnessed to limit cancer growth.^{45,46} One particular dietary intervention consists in a close-to-zero carbohydrate, low-protein, high-fat regimen that causes ketosis (*i.e.*, the accumulation of 3-hydroxybutyrate, acetoacetate, and acetone, commonly known as ketone bodies).⁴⁷ Ketone bodies can be used by multiple tissues in replacement of glucose for energy metabolism.⁴⁸ In mice, multiple variants of ketogenic diet slowdown the progression of some cancer types and boost the efficacy of targeted therapeutic agents,^{49,50} an effect that (at least in some setting) is linked to reduced insulin signaling.^{50,51}

One particular facet of these dietary interventions is their capacity to reduce the unwarranted side effect of genotoxic chemotherapy. For example, periodic fasting as well as the administration of a hypocaloric ‘fasting-mimicking diet’ (FMD) can enhance the efficacy of chemotherapy and, at the same time, limit chemotherapy-related weight loss and cardiotoxicity.^{27,52} It has been theorized that transient calorie deprivation enhances the ‘differential stress resistance’ between chemotherapy-treated cancer cells (that would become more susceptible to the treatment) and normal, non-neoplastic cells (that would become more resistant to the toxic side effects to chemotherapy).^{53–56}

Anticancer effects of dietary interventions – an immunological rationale

Over the past years, an ever-expanding body of evidence pleads in favor of the notion that the long-term success of chemotherapy, targeted therapy and radiotherapy requires the reestablishment of immunosurveillance.⁵⁷ In other words, the efficacy of antineoplastic treatments, which has long been thought to exclusively rely on cancer cell-autonomous effects, now turns out to require the induction of a protracted anticancer immune response to be efficient.^{58–61} Logically, the impact of dietary intervention on such immune-dependent antitumor effects has been studied in preclinical models.

In immunocompetent mice bearing transplantable tumors or carcinogen-induced breast cancer, chemotherapy with anthracyclines or oxaliplatin becomes more efficient if combined with short periods of starvation.^{22,24} These combinatorial effects of chemotherapy and dietary intervention fully rely on a T lymphocyte-mediated anticancer immune response, meaning that they are lost upon T cell depletion.^{22,24} Mechanistically, they have been linked to the

induction of heme oxygenase-1 (HO-1) in cancer cells, the stimulation of autophagy in cancer cells (which would enhance their immunogenicity),^{62–64} a decrease in circulating insulin-like growth factor-1 (IGF1), as well as an increase in the frequency of common lymphocyte precursors (which would be immunostimulatory).^{22,24,65} Whether such effects might involve major shifts in the gut microbiota has not been investigated thus far.^{66,67} Moreover, the impact of dietary interventions on immunotherapies has been poorly explored, at least to our knowledge.^{68,69} Available clinical evidence suggests that anti-melanoma immunotherapy with PD-1/PD-L1 blocking antibodies^{70,71} is more efficient in obese than lean males,⁷² casting doubts on the possibility to improve such therapies by brutal interventions designed to reduce overweight.

Published and ongoing clinical trials

Very few trials testing the ability of nutritional interventions to boost the efficacy of cancer therapy have been reported in the peer-reviewed literature so far. A series of anecdotal cases of self-imposed starvation during chemotherapy suggested an improvement of subjective well-being suggestive of a reduction of side-effects.⁷³ In the same line, fasting for 48 h prior and 24 h after platinum-based chemotherapy proved its safety and feasibility in patients treated for diverse cancer types.⁷⁴ A Phase I trial confirmed that STS for 60 h (from 36 h prior to chemotherapy to 24 h post-chemotherapy) improves quality of life and fatigue in patients with gynecological cancer.⁷⁵ In breast cancer patients treated with neoadjuvant multimodal chemotherapy,⁷⁶ a 48-h starvation period (from 24 h before to 24 h after chemotherapy) reduced hematological toxicity and accelerated recovery from DNA damage in circulating leukocytes.⁷⁷ Women with ovarian and endometrial cancer following a ketogenic diet for 12 weeks reported higher physical and energy status compared to the control group, highlighting the feasibility of this regimen.⁷⁸ A special ketogenic diet, the so-called ‘modified Atkins diet’, reportedly reduces the progression of some advanced cancer patients, especially individuals experience robust weight reduction.⁷⁹ Similarly, a ketogenic regimen has been reported to induce objective responses in 6 out of 7 patients with recurrent glioblastoma that simultaneously were treated with the antiangiogenic drug bevacizumab.^{80–82} This effect appeared particularly strong in patients with stable ketosis.⁸⁰

The website ClinicalTrials.gov informs on multiple clinical trials that are either ongoing or completed, yet generally lack published information on the outcome (Table 1). Many of these trials evaluate dietary interventions without further treatment (NCT01092247, NCT01865162, NCT02092753, NCT02286167, NCT03160599, NCT03194516, NCT03328858, NCT03785808, NCT00003367, NCT00020995, NCT00082732, NCT00444054, NCT01692587, NCT02129218, NCT02176902, NCT03221920, and NCT03679260). Such interventions include STS, intermediate fasting, FMD, multiple ketogenic and low-carbohydrate diets, low-fat/high-fiber regimens, protein-restrictive diets, low-calorie and low-glycemic regimens and a vegan diet in patients with a variety of advanced solid malignancies including glioblastoma (the most frequent indication),

Table 1. Clinical trials employing diet for cancer therapy.

Dietary intervention	Additional details (when available)	NCT	Therapeutic intervention	Cancer type	Phase	Status
Short-term starvation	24, 48, or 72 h of fasting or 48 h of FMD	NCT00936364	Platinum chemotherapy	Advanced solid tumors		Recruiting
	24, 36, or 48 h of fasting before chemotherapy	NCT01175837	Chemotherapy	Breast cancer	Pilot study	Completed
	STS 24 h before and 24 h after chemotherapy	NCT01304251	Docetaxel, Doxorubicin, Cyclophosphamide	Breast cancer	Pilot study	Completed
	STS 24 h before and 24 h after chemotherapy	NCT02379585	Doxorubicin, cyclophosphamide, paclitaxel, docetaxel, trastuzumab, pertuzumab	Breast cancer	Phase 1/2	Terminated, has results
	FMD 36 to 48 h before and 24 h after chemotherapy	NCT01954836	Chemotherapy	Gynecological	Pilot study	Completed
Fasting-mimicking diet	FMD	NCT02126449	Neoadjuvant chemotherapy	HER2-negative breast cancer	Phase 2/3	Terminated
	FMD 36 h before and 24 h after chemotherapy	NCT02710721	Chemotherapy	Prostate		Recruiting
	FMD or vegan diet 36 to 48 h before and 24 h after chemotherapy	NCT03162289	Chemotherapy	Breast and ovarian		Recruiting
	FMD (low calorie, low protein, and low carbohydrates) for 5 days	NCT03340935	Standard therapies	Any malignancy except small-cell neuroendocrine tumors		Recruiting
	5 days of FMD 13 to 15 days before or 1 month after surgery	NCT03454282	Surgery	Breast and melanoma tumors		Recruiting
	FMD for 5 days	NCT03595540	Chemo, hormone-, targeted or immuno-therapies			Recruiting
	FMD 72 h before and 24 h after chemo-immunotherapy	NCT03700437	Carboplatin/pemetrexed and pembrolizumab	NSCLC		Not yet recruiting
	FMD for 5 days	NCT03709147	Metformin	Advanced LKB1-inactive lung adenocarcinoma	Phase	Not yet recruiting
	KD	NCT00575146	Bevacizumab	Recurrent glioblastoma	Phase 1	Completed, has results
	Ketogenic diet	KD for up to one year	NCT01092247	No	High-grade glial tumors	
KD starts 2 days before chemoradiation and last at least during 5 weeks throughout the treatment		NCT01419483	Chemoradiation	Pancreatic		Terminated
KD starts 2 days before chemoradiation and last at least during 5 weeks throughout the treatment		NCT01419587	Chemoradiation	Carcinoma, non-small cell lung cancer		Terminated
Energy-restricted KD starts after surgery and continues through radio and chemotherapy, ending 6 weeks after treatments completion		NCT01535911	Surgery followed by chemo- and radiotherapy	Brain tumors		Active, not recruiting
KD		NCT01716468	No	Metastatic cancer		Completed
2 cycles of 3 days calorie-restricted KD separated by 3 days fasting		NCT01754350	Reirradiation	Recurrent glioblastoma		Active, not recruiting
KD with calorie restriction for 6 months		NCT01865162	No	Refractory/end-stage glioblastoma	Phase 1	Recruiting
KD starts 2 days before chemoradiation and last at least during 5 weeks throughout the treatment		NCT01975766	Chemoradiation	Head and neck cancer	Phase 1	Terminated (poor accrual)
KD starts after surgery and continues through radio and chemotherapy. A modified Atkins diet is implemented during the following month of chemotherapy.		NCT02046187	Chemoradiation	Glioblastoma	Phase 1/2	Terminated
KD or low glycemic and insulinemic diet for 20 weeks		NCT02092753	No	Breast cancer		Completed
Modified Atkins-based with intermittent fasting diet		NCT02286167	No	Glioblastoma		Recruiting
KD starts at the radiation initiation and continues 6 months throughout the entire period of chemoradiation		NCT02302235	Radiation and temozolomide	Glioblastoma multiforme	Phase 2	Recruiting
Ketogenic breakfast after overnight fasting and before chemoradiation or KD throughout the entire period of chemoradiation		NCT02516501	Chemoradiation	Breast ER+ cancer	Pilot presurgical study	Recruiting
Low-carbohydrate vs low-fat diet before surgery		NCT02744079	Surgery	Glioblastoma	Early Phase 1	Active, not recruiting
Modified Atkins diet		NCT02768389	Bevacizumab	Recurrent glioblastoma	Phase 1/2	Unknown
KD	NCT02939378	Salvage chemotherapy				

(Continued)

Table 1. (Continued).

Dietary intervention	Additional details (when available)	NCT	Therapeutic intervention	Cancer type	Phase	Status
KD		NCT02983942	Methotrexate	Primary central nervous system lymphoma	Phase 1/2	Not yet recruiting
Modified KD or medium-chain triglyceride KD for 12 weeks		NCT03075514	Chemo- and/or radiotherapy	Glioblastoma	Pilot study	Active, not recruiting
Restricted calorie KD for 12 weeks		NCT03160599	No	Glioblastoma multiforme		Recruiting
KD for 8 weeks		NCT03171506	No	Ovarian and endometrial cancer		Completed
Modified Atkins KD for 7 days before surgery		NCT03278249	Temozolomide and radiation	Prostate cancer		Enrolling by invitation
KD for at least 1 year		NCT03285152	Surgery	Malignant glioma		Recruiting
KD for 16 weeks throughout chemoradiation treatment		NCT03328858	No	Endometrial cancer		Recruiting
KD for 3 months		NCT03451799	Radiation and temozolomide	Brain tumors	Phase 1	Recruiting
Low-carbohydrate high-fat ketogenic-type diet vs low-fat high low-glycemic carbohydrates diet		NCT03535701	Paclitaxel	Glioblastoma		Recruiting
Low-fat high-fiber diet		NCT03785808	No	Stage IV breast cancer		Recruiting
Specific low-nutrient		NCT00003367	No	Lung cancer	Phase 3	Completed
Low-fat high-fiber diet for 3 weeks		NCT00020995	No	Prostate cancer	Phase 2	Completed
Low-fat high-fiber diet		NCT00082732	No	Hormone-refractory prostate cancer	Phase 1	Completed
Very low-carbohydrate diet for 28 days		NCT00444054	No	Advanced cancer	Pilot study	Completed
Low-fiber vs high-fiber diet		NCT01170299	Radiation	Gynecological, bladder, colorectal, or anal cancer		Completed
Protein-restrictive diet		NCT01692587	No	Prostate cancer		Completed
Low-calorie diet from 3 days before to 2 days after the 12 weeks of chemotherapy		NCT01802346	Chemotherapy	Breast, hormone-resistant, and recurrent prostate cancer	Phase 2	Recruiting
Carbohydrate-restricted diet		NCT02019979	Metformin with platinum-based chemotherapy	Non-squamous non-small cell lung cancer	Phase 2	Terminated, has results
Low- or medium-glycemic diet for 12 weeks		NCT02129218	No	Colon cancer	Pilot study	Completed
Low-fat omega-3 supplement diet for one year		NCT02176902	No	Prostate cancer	Phase 2	Recruiting
Vegetarian vs vegan diets for 6 months		NCT02437474	Prescribed therapy	Any cancer type	Pilot study	Completed
Very low-carbohydrate and high-fat diet		NCT03221920	No	Colorectal adenocarcinoma		Not yet recruiting
Low protein diet from 1 week before to 10 days after treatment		NCT03329742	Sipuleucel-T	Metastatic castrate-resistant pancreatic cancer		Recruiting
Carbohydrate restricted diet for 6 months		NCT03679260	No	Prostate cancer	Phase 2	Recruiting

breast and gynecological cancer, melanoma, head and neck cancer, non-small cell lung carcinoma, ovarian cancer, pancreatic adenocarcinoma, and prostate cancer. Several trials also aim at investigating the combination of dietary interventions with (1) chemotherapy (NCT01175837, NCT02379585, NCT02126449, NCT02710721, NCT03162289, NCT03340935, NCT03595540, NCT03700437, NCT01419483, NCT01419587, NCT01975766, NCT02046187, NCT02302235, NCT02516501, NCT02939378, NCT02983942, NCT03075514, NCT03278249, NCT03451799, NCT03535701, NCT01802346, NCT02019979, and NCT02437474), (2) radiotherapy (NCT03340935, NCT01419483, NCT01419587, NCT01754350, NCT01975766, NCT02046187, NCT02302235, NCT02516501, NCT03075514, NCT03278249, NCT03451799, NCT0170299, and NCT02437474), (3) metformin, a medication for type II diabetes with pleiotropic effects on cancer cells^{83–85} (NCT03709147 and NCT02019979), (4) targeted-therapies (NCT02379585, NCT03595540 and NCT02768389), and (5) immunotherapies such as immune checkpoint blockers targeting PD-1^{86,87} (NCT03595540 and NCT03700437) or the dendritic cells based-vaccine Sipuleucel-T^{88,89} (NCT03329742). Interestingly, one study also sets to monitor anticancer immune responses induced by an FMD (NCT03454282).

It will be interesting to see whether any of these studies will document a clinical benefit linked to a specific nutritional intervention.

Concluding remarks

Knowing the importance of nutrition and metabolism for human physiology, including the crosstalk between malignant and immune cells, it is not surprising that dietary interventions are attracting attention as safe means to limit tumor progression or restore disease control by the host immune system.^{90–92} While evidence from preclinical studies suggests that reducing total calorie intake (and perhaps specific macronutrients) may stimulate anticancer immunity, such evidence has not yet been obtained in clinical trials. Multiple trials testing these possibilities in patients with multiple types of cancer are on the way (Table 1). Unfortunately, it will be difficult to compare results from different studies for at least two reasons that add upon the usual heterogeneity of clinical trials. First, dietary interventions are quite heterogeneous in nature.¹³ Thus, the term ‘ketogenic diet’ may refer to distinct regimens differing in quantity, composition and even in the gross protein:fat ratio.⁹³ Second, the control arms of the studies, when exist, usually receive consulting on ‘healthy dietary habits’, which (1) is a non-standardized notion (with major cross-continental and cross-cultural divergences), (2) is usually not enforced, and (3) is extremely complex to monitor.

Thus, the studies listed in Table 1 might examine the differences between salutary (interventional) and poor (control) regimens, meaning that control regimens can be expected to have a negative impact on health status. For this reason, it will be important to standardize control diets, ensure compliance, and to define interventional regimens in an accurate fashion. This implies strict guidelines, their enforcement by connected objects and phone app-mediated

control, as well as monitoring of multiple metabolic parameters (such as plasma metabolome, cytokine and hormone status, stool microbiota). Moreover, it will be important to monitor immune parameters in the tumor and the peripheral blood to gain insights into therapeutically relevant anticancer immune responses. Without this information, it will be difficult to obtain any useful knowledge on the impact of nutritional interventions on cancer therapy.

Acknowledgments

SL is supported by an end of PhD grant from the Fondation pour la Recherche Médicale (FRM FDT201805005722). LG is supported by a Breakthrough Level 2 grant from the US Department of Defense (DoD), Breast Cancer Research Program (BRCP) [#BC180476P1], by a startup grant from the Dept. of Radiation Oncology at Weill Cornell Medicine (New York, US), by industrial collaborations with Lytix (Oslo, Norway) and Phosplatin (New York, US), and by donations from Phosplatin (New York, US), the Luke Heller TECPR2 Foundation (Boston, US) and Sotio a.s. (Prague, Czech Republic). GK is supported by the Ligue contre le Cancer (équipe labellisée); Agence National de la Recherche (ANR) – Projets blancs; ANR under the frame of E-Rare-2, the ERA-Net for Research on Rare Diseases; Association pour la recherche sur le cancer; Cancéropôle Ile-de-France; Chancellerie des universités de Paris (Legs Poix), Fondation pour la Recherche Médicale (FRM); a donation by Elior; European Research Area Network on Cardiovascular Diseases (ERA-CVD, MINOTAUR); the European Union Horizon 2020 Project Oncobiome; Fondation Carrefour; Institut National du Cancer (INCa); Inserm (HTE); Institut Universitaire de France; LeDucq Foundation; the LabEx Immuno-Oncology; the RHU Torino Lumière; the Seerave Foundation; the SIRIC Stratified Oncology Cell DNA Repair and Tumor Immune Elimination (SOCRATE); and the SIRIC Cancer Research and Personalized Medicine (CARPEM).

Disclosure of Potential Conflicts of Interest

LG provides remunerated consulting to OmniSEQ (Buffalo, NY, USA), Astra Zeneca (Gaithersburg, MD, USA), VL47 (New York, NY, USA) and the Luke Heller TECPR2 Foundation (Boston, MA, USA), and he is member of the Scientific Advisory Committee of OmniSEQ (Buffalo, NY, USA). GK and LZ receive a research grant by Elior.

Funding

This work was supported by the Agence Nationale de la Recherche [E-Rare-2]; Fondation pour la Recherche Médicale [FRM FDT201805005722]; H2020 European Union [Oncobiome].

ORCID

Laurence Zitvogel  <http://orcid.org/0000-0003-1596-0998>

Guido Kroemer  <http://orcid.org/0000-0002-9334-4405>

References

1. NCD Risk Factor Collaboration (NCD-RisC). Trends in adult body-mass index in 200 countries from 1975 to 2014: a pooled analysis of 1698 population-based measurement studies with 19.2 million participants. *Lancet Lond Engl.* 2016;387(10026):1377–1396. doi:10.1016/S0140-6736(16)30054-X.
2. Hossain P, Kawar B, El Nahas M. Obesity and diabetes in the developing world—a growing challenge. *N Engl J Med.* 2007;356(3):213–215. doi:10.1056/NEJMp068177.

3. Bianchini F, Kaaks R, Vainio H. Overweight, obesity, and cancer risk. *Lancet Oncol.* 2002;3(9):565–574. doi:10.1016/S1470-2045(02)00849-5.
4. Dehghan M, Mente A, Zhang X, Swaminathan S, Li W, Mohan V, Iqbal R, Kumar R, Wentzel-Viljoen E, Rosengren A, et al. Associations of fats and carbohydrate intake with cardiovascular disease and mortality in 18 countries from five continents (PURE): a prospective cohort study. *The Lancet.* 2017;390(10107):2050–2062. doi:10.1016/S0140-6736(17)32252-3.
5. Myles IA. Fast food fever: reviewing the impacts of the Western diet on immunity. *Nutr J.* 2014;13:61. doi:10.1186/1475-2891-13-61.
6. Mozaffarian D, Hao T, Rimm EB, Willett WC, Hu FB. Changes in diet and lifestyle and long-term weight gain in women and men. *N Engl J Med.* 2011;364(25):2392–2404. doi:10.1056/NEJMoa1014296.
7. Kroemer G, López-Otín C, Madeo F, de Cabo R. Carbotoxicity-noxious effects of carbohydrates. *Cell.* 2018;175(3):605–614. doi:10.1016/j.cell.2018.07.044.
8. Lock K, Pomerleau J, Causer L, Altmann DR, McKee M. The global burden of disease attributable to low consumption of fruit and vegetables: implications for the global strategy on diet. *Bull World Health Organ.* 2005;83:100–108.
9. Bazzano LA, He J, Ogden LG, Loria CM, Vupputuri S, Myers L, Whelton PK. Fruit and vegetable intake and risk of cardiovascular disease in US adults: the first national health and nutrition examination survey epidemiologic follow-up study. *Am J Clin Nutr.* 2002;76(1):93–99. doi:10.1093/ajcn/76.1.93.
10. World Health Organization WHO. Global nutrition policy review 2016–2017. 2018. [accessed February 15, 2019]. <https://apps.who.int/iris/bitstream/handle/10665/275990/9789241514873-eng.pdf?ua=1>
11. Pellizzon M. Choice of laboratory animal diet influences intestinal health. *Lab Anim.* 2016;45:238–239. doi:10.1038/labana.1014.
12. Warden CH, Fisler JS. Comparisons of diets used in animal models of high-fat feeding. *Cell Metab.* 2008;7(4):277. doi:10.1016/j.cmet.2008.03.014.
13. Yao CK, Gibson PR, Shepherd SJ. Design of clinical trials evaluating dietary interventions in patients with functional gastrointestinal disorders. *Am J Gastroenterol.* 2013;108(5):748–758. doi:10.1038/ajg.2013.77.
14. Deschasaux M, Huybrechts I, Murphy N, Julia C, Hercberg S, Srouf B, Kesse-Guyot E, Latino-Martel P, Biessy C, Casagrande C, et al. Nutritional quality of food as represented by the FSAM-NPS nutrient profiling system underlying the nutri-score label and cancer risk in Europe: results from the EPIC prospective cohort study. *PLoS Med.* 2018;15(9):e1002651. doi:10.1371/journal.pmed.1002651.
15. Key TJ, Allen NE, Spencer EA, Travis RC. The effect of diet on risk of cancer. *Lancet Lond Engl.* 2002;360(9336):861–868. doi:10.1016/S0140-6736(02)09958-0.
16. Jansen MCJF, Bueno-de-Mesquita HB, Feskens EJM, Streppel MT, Kok FJ, Kromhout D. Quantity and variety of fruit and vegetable consumption and cancer risk. *Nutr Cancer.* 2004;48(2):142–148. doi:10.1207/s15327914nc4802_3.
17. Lauby-Secretan B, Scoccianti C, Loomis D, Grosse Y, Bianchini F, Straif K; International Agency for Research on Cancer Handbook Working Group. Body fatness and cancer—viewpoint of the IARC working group. *N Engl J Med.* 2016;375(8):794–798. doi:10.1056/NEJMs1606602.
18. Calle EE, Kaaks R. Overweight, obesity and cancer: epidemiological evidence and proposed mechanisms. *Nat Rev Cancer.* 2004;4(8):579–591. doi:10.1038/nrc1408.
19. Font-Burgada J, Sun B, Karin M. Obesity and cancer: the oil that feeds the flame. *Cell Metab.* 2016;23(1):48–62. doi:10.1016/j.cmet.2015.12.015.
20. Merletti F, Galassi C, Spadea T. The socioeconomic determinants of cancer. *Environ Health Glob Access Sci Source.* 2011;10(Suppl 1):S7. doi:10.1186/1476-069X-10-S1-S7.
21. Galluzzi L, Vitale I, Senovilla L, Olaussen KA, Pinna G, Eisenberg T, Goubar A, Martins I, Michels J, Kratassiouk G, et al. Prognostic impact of vitamin B6 metabolism in lung cancer. *Cell Rep.* 2012;2(2):257–269. doi:10.1016/j.celrep.2012.06.017.
22. Di Biase S, Lee C, Brandhorst S, Manes B, Buono R, Cheng C-W, Cacciottolo M, Martin-Montalvo A, de Cabo R, Wei M, et al. Fasting-mimicking diet reduces HO-1 to promote T cell-mediated tumor cytotoxicity. *Cancer Cell.* 2016;30(1):136–146. doi:10.1016/j.ccell.2016.06.005.
23. Di Biase S, Longo VD. Fasting-induced differential stress sensitization in cancer treatment. *Mol Cell Oncol.* 2016;3(3):e1117701. doi:10.1080/23723556.2015.1117701.
24. Pietrocola F, Pol J, Vacchelli E, Rao S, Enot DP, Baracco EE, Levesque S, Castoldi F, Jacquelin N, Yamazaki T, et al. Caloric restriction mimetics enhance anticancer immunosurveillance. *Cancer Cell.* 2016;30(1):147–160. doi:10.1016/j.ccell.2016.05.016.
25. Safdie F, Brandhorst S, Wei M, Wang W, Lee C, Hwang S, Conti PS, Chen TC, Longo VD. Fasting enhances the response of glioma to chemo- and radiotherapy. *PLoS One.* 2012;7(9):e44603. doi:10.1371/journal.pone.0044603.
26. Klement RJ, Champ CE. Calories, carbohydrates, and cancer therapy with radiation: exploiting the five R's through dietary manipulation. *Cancer Metastasis Rev.* 2014;33(1):217–229. doi:10.1007/s10555-014-9495-3.
27. Lee C, Raffaghello L, Brandhorst S, Safdie FM, Bianchi G, Martin-Montalvo A, Pistoia V, Wei M, Hwang S, Merlino A, et al. Fasting cycles retard growth of tumors and sensitize a range of cancer cell types to chemotherapy. *Sci Transl Med.* 2012;4(124):124ra27. doi:10.1126/scitranslmed.3003293.
28. Vander Heiden MG. Targeting cancer metabolism: a therapeutic window opens. *Nat Rev Drug Discov.* 2011;10(9):671–684. doi:10.1038/nrd3504.
29. Galluzzi L, Kepp O, Vander Heiden MG, Kroemer G. Metabolic targets for cancer therapy. *Nat Rev Drug Discov.* 2013;12(11):829–846. doi:10.1038/nrd4145.
30. O'Neill LAJ, Kishton RJ, Rathmell J. A guide to immunometabolism for immunologists. *Nat Rev Immunol.* 2016;16(9):553–565. doi:10.1038/nri.2016.70.
31. Gruenbacher G, Thurnher M. Mevalonate metabolism governs cancer immune surveillance. *Oncoimmunology.* 2017;6(10):e1342917. doi:10.1080/2162402X.2017.1342917.
32. Bantug GR, Galluzzi L, Kroemer G, Hess C. The spectrum of T cell metabolism in health and disease. *Nat Rev Immunol.* 2018;18(1):19–34. doi:10.1038/nri.2017.99.
33. Al-Khami AA, Zheng L, Del Valle L, Hossain F, Wyczzechowska D, Zabaleta J, Sanchez MD, Dean MJ, Rodriguez PC, Ochoa AC. Exogenous lipid uptake induces metabolic and functional reprogramming of tumor-associated myeloid-derived suppressor cells. *Oncoimmunology.* 2017;6(10):e1344804. doi:10.1080/2162402X.2017.1344804.
34. Buck MD, Sowell RT, Kaech SM, Pearce EL. Metabolic instruction of immunity. *Cell.* 2017;169(4):570–586. doi:10.1016/j.cell.2017.04.004.
35. Vacchelli E, Eggermont A, Sautès-Fridman C, Galon J, Zitvogel L, Kroemer G, Galluzzi L. Trial watch: oncolytic viruses for cancer therapy. *Oncoimmunology.* 2013;2(6):e24612. doi:10.4161/onci.24612.
36. Aranda F, Vacchelli E, Eggermont A, Galon J, Fridman WH, Zitvogel L, Kroemer G, Galluzzi L. Trial watch: immunostimulatory monoclonal antibodies in cancer therapy. *Oncoimmunology.* 2014;3(1):e27297. doi:10.4161/onci.27297.
37. Hsu PP, Sabatini DM. Cancer cell metabolism: warburg and beyond. *Cell.* 2008;134(5):703–707. doi:10.1016/j.cell.2008.08.021.
38. Kroemer G, Pouyssegur J. Tumor cell metabolism: cancer's Achilles' heel. *Cancer Cell.* 2008;13(6):472–482. doi:10.1016/j.ccr.2008.05.005.
39. Starkova J, Hermanova I, Hlozkova K, Hararova A, Trka J. altered metabolism of leukemic cells: new therapeutic opportunity. *Int Rev Cell Mol Biol.* 2018;336:93–147. doi:10.1016/bs.ircmb.2017.07.012.
40. Costa ASH, Frezza C. Metabolic reprogramming and oncogenesis: one hallmark, many organelles. *Int Rev Cell Mol Biol.* 2017;332:213–231. doi:10.1016/bs.ircmb.2017.01.001.

41. Pavlova NN, Thompson CB. The emerging hallmarks of cancer metabolism. *Cell Metab.* 2016;23(1):27–47. doi:10.1016/j.cmet.2015.12.006.
42. Commisso C, Davidson SM, Soydaner-Azeloglu RG, Parker SJ, Kamphorst JJ, Hackett S, Grabocka E, Nofal M, Drebin JA, Thompson CB, et al. Macropinocytosis of protein is an amino acid supply route in Ras-transformed cells. *Nature.* 2013;497(7451):633–637. doi:10.1038/nature12138.
43. Fais S. Cannibalism: a way to feed on metastatic tumors. *Cancer Lett.* 2007;258(2):155–164. doi:10.1016/j.canlet.2007.09.014.
44. Beloribi-Djefafila S, Vasseur S, Guillaumond F. Lipid metabolic reprogramming in cancer cells. *Oncogenesis.* 2016;5(1):e189. doi:10.1038/oncsis.2015.49.
45. Rubio-Patiño C, Bossowski JP, De Donatis GM, Mondragón L, Villa E, Aira LE, Chiche J, Mhaidly R, Lebeauin C, Marchetti S, et al. Low-protein diet induces IRE1 α -dependent anticancer immunosurveillance. *Cell Metab.* 2018;27(4):828–842.e7. doi:10.1016/j.cmet.2018.02.009.
46. Maddocks ODK, Athineos D, Cheung EC, Lee P, Zhang T, van Den Broek NJF, Mackay GM, Labuschagne CF, Gay D, Kruiswijk F, et al. Modulating the therapeutic response of tumours to dietary serine and glycine starvation. *Nature.* 2017;544(7650):372–376. doi:10.1038/nature22056.
47. Paoli A, Rubini A, Volek JS, Grimaldi KA. Beyond weight loss: a review of the therapeutic uses of very-low-carbohydrate (ketogenic) diets. *Eur J Clin Nutr.* 2013;67(8):789–796. doi:10.1038/ejcn.2013.116.
48. Puchalska P, Crawford PA. Multi-dimensional roles of ketone bodies in fuel metabolism, signaling, and therapeutics. *Cell Metab.* 2017;25(2):262–284. doi:10.1016/j.cmet.2016.12.022.
49. Klement RJ, Champ CE, Otto C, Kämmerer U. Anti-tumor effects of ketogenic diets in mice: a meta-analysis. *PLoS One.* 2016;11(5):e0155050. doi:10.1371/journal.pone.0155050.
50. Hopkins BD, Pauli C, Du X, Wang DG, Li X, Wu D, Amadiume SC, Goncalves MD, Hodakoski C, Lundquist MR, et al. Suppression of insulin feedback enhances the efficacy of PI3K inhibitors. *Nature.* 2018;560(7719):499–503. doi:10.1038/s41586-018-0343-4.
51. Crudden C, Shibano T, Song D, Suleymanova N, Girnita A, Girnita L. Blurring boundaries: receptor tyrosine kinases as functional G protein-coupled receptors. *Int Rev Cell Mol Biol.* 2018;339:1–40. doi:10.1016/bs.ircmb.2018.02.006.
52. Bonora M, Wieckowski MR, Sinclair DA, Kroemer G, Pinton P, Galluzzi L. Targeting mitochondria for cardiovascular disorders: therapeutic potential and obstacles. *Nat Rev Cardiol.* 2019;16(1):33–55. doi:10.1038/s41569-018-0074-0.
53. Raffaghello L, Lee C, Safdie FM, Wei M, Madia F, Bianchi G, Longo VD. Starvation-dependent differential stress resistance protects normal but not cancer cells against high-dose chemotherapy. *Proc Natl Acad Sci USA.* 2008;105(24):8215–8220. doi:10.1073/pnas.0708100105.
54. Raffaghello L, Longo V. Metabolic alterations at the crossroad of aging and oncogenesis. *Int Rev Cell Mol Biol.* 2017;332:1–42. doi:10.1016/bs.ircmb.2017.01.003.
55. Lee C, Safdie FM, Raffaghello L, Wei M, Madia F, Parrella E, Hwang D, Cohen P, Bianchi G, Longo VD. Reduced levels of IGF-I mediate differential protection of normal and cancer cells in response to fasting and improve chemotherapeutic index. *Cancer Res.* 2010;70(4):1564–1572. doi:10.1158/0008-5472.CAN-09-3228.
56. Buono R, Longo VD. Starvation, stress resistance, and cancer. *Trends Endocrinol Metab.* 2018;29(4):271–280. doi:10.1016/j.tem.2018.01.008.
57. Galluzzi L, Buqué A, Kepp O, Zitvogel L, Kroemer G. Immunological effects of conventional chemotherapy and targeted anticancer agents. *Cancer Cell.* 2015;28(6):690–714. doi:10.1016/j.ccell.2015.10.012.
58. Stoll G, Pol J, Soumelis V, Zitvogel L, Kroemer G. Impact of chemotactic factors and receptors on the cancer immune infiltrate: a bioinformatics study revealing homogeneity and heterogeneity among patient cohorts. *Oncoimmunology.* 2018;7(10):e1484980. doi:10.1080/2162402X.2018.1484980.
59. Krysko DV, Garg AD, Kaczmarek A, Krysko O, Agostinis P, Vandenabeele P. Immunogenic cell death and DAMPs in cancer therapy. *Nat Rev Cancer.* 2012;12(12):860–875. doi:10.1038/nrc3380.
60. Garg AD, More S, Rufo N, Mece O, Sassano ML, Agostinis P, Zitvogel L, Kroemer G, Galluzzi L. Trial watch: immunogenic cell death induction by anticancer chemotherapeutics. *Oncoimmunology.* 2017;6(12):e1386829. doi:10.1080/2162402X.2017.1386829.
61. Galluzzi L, Kroemer G. An epigenetic modifier triggers therapeutic immune responses against breast cancer. *Oncoimmunology.* 2017;6(5):e1313376. doi:10.1080/2162402X.2017.1313376.
62. Michaud M, Martins I, Sukkurwala AQ, Adjemian S, Ma Y, Pellegatti P, Shen S, Kepp O, Scoazec M, Mignot G, et al. Autophagy-dependent anticancer immune responses induced by chemotherapeutic agents in mice. *Science.* 2011;334(6062):1573–1577. doi:10.1126/science.1208347.
63. Galluzzi L, Baehrecke EH, Ballabio A, Boya P, Bravo-San Pedro JM, Cecconi F, Choi AM, Chu CT, Codogno P, Colombo MI, et al. Molecular definitions of autophagy and related processes. *Embo J.* 2017;36(13):1811–1836. doi:10.15252/emboj.201796697.
64. Galluzzi L, Pietrocola F, Bravo-San Pedro JM, Amaravadi RK, Baehrecke EH, Cecconi F, Codogno P, Debnath J, Gewirtz DA, Karantza V, et al. Autophagy in malignant transformation and cancer progression. *Embo J.* 2015;34(7):856–880. doi:10.15252/emboj.201490784.
65. Burger T, Molnár L, Schmelzer M, Tóvari E, Szabó A, Paál M, Királyfalvi L. Changes in T lymphocyte subgroups and their effect in chronic B-lymphoid leukemia. *Orv Hetil.* 1988;129:2189–2193.
66. Routy B, Gopalakrishnan V, Daillère R, Zitvogel L, Wargo JA, Kroemer G. The gut microbiota influences anticancer immunosurveillance and general health. *Nat Rev Clin Oncol.* 2018;15(6):382–396. doi:10.1038/s41571-018-0006-2.
67. Derosa L, Routy B, Kroemer G, Zitvogel L. The intestinal microbiota determines the clinical efficacy of immune checkpoint blockers targeting PD-1/PD-L1. *Oncoimmunology.* 2018;7(6):e1434468. doi:10.1080/2162402X.2018.1434468.
68. Orillion A, Damayanti NP, Shen L, Adelaiye-Ogala R, Affronti H, Elbanna M, Chintala S, Ciesielski M, Fontana L, Kao C, et al. Dietary protein restriction reprograms tumor-associated macrophages and enhances immunotherapy. *Clin Cancer Res Off J Am Assoc Cancer Res.* 2018;24(24):6383–6395. doi:10.1158/1078-0432.CCR-18-0980.
69. Soldati L, Di Renzo L, Jirillo E, Ascierto PA, Marincola FM, De Lorenzo A. The influence of diet on anti-cancer immune responsiveness. *J Transl Med.* 2018;16. doi:10.1186/s12967-018-1448-0.
70. Vanpouille-Box C, Lhuillier C, Bezu L, Aranda F, Yamazaki T, Kepp O, Fucikova J, Spisek R, Demaria S, Formenti SC, et al. Trial watch: immune checkpoint blockers for cancer therapy. *Oncoimmunology.* 2017;6(11):e1373237. doi:10.1080/2162402X.2017.1373237.
71. Galluzzi L, Chan TA, Kroemer G, Wolchok JD, López-Soto A. The hallmarks of successful anticancer immunotherapy. *Sci Transl Med.* 2018;10:459. doi:10.1126/scitranslmed.aat7807.
72. Wang Z, Aguilar EG, Luna JI, Dunai C, Khuat LT, Le CT, Mirsoian A, Minnar CM, Stoffel KM, Sturgill IR, et al. Paradoxical effects of obesity on T cell function during tumor progression and PD-1 checkpoint blockade. *Nat Med.* 2019;25(1):141–151. doi:10.1038/s41591-018-0221-5.
73. Safdie FM, Dorff T, Quinn D, Fontana L, Wei M, Lee C, Cohen P, Longo VD. Fasting and cancer treatment in humans: a case series report. *Aging.* 2009;1(12):988–1007. doi:10.18632/aging.100114.
74. Dorff TB, Groshen S, Garcia A, Shah M, Tsao-Wei D, Pham H, Cheng C-W, Brandhorst S, Cohen P, Wei M, et al. Safety and feasibility of fasting in combination with platinum-based chemotherapy. *BMC Cancer.* 2016;16. doi:10.1186/s12885-016-2370-6.
75. Bauersfeld SP, Kessler CS, Wischnowsky M, Jaensch A, Steckhan N, Stange R, Kunz B, Brückner B, Sehoul J,

- Michalsen A. The effects of short-term fasting on quality of life and tolerance to chemotherapy in patients with breast and ovarian cancer: a randomized cross-over pilot study. *BMC Cancer*. 2018;18(1):476. doi:10.1186/s12885-018-4353-2.
76. de la Cruz-Merino L, Chiesa M, Caballero R, Rojo F, Palazón N, Carrasco FH, Sánchez-Margalet V. Breast cancer immunology and immunotherapy: current status and future perspectives. *Int Rev Cell Mol Biol*. 2017;331:1–53. doi:10.1016/bs.ircmb.2016.09.008.
77. de Groot S, Vreeswijk MPG, Welters MJP, Gravesteyn G, Boei JJWA, Jochems A, Houtsmá D, Putter H, van der Hoeven JJM, Nortier JWR, et al. The effects of short-term fasting on tolerance to (neo) adjuvant chemotherapy in HER2-negative breast cancer patients: a randomized pilot study. *BMC Cancer*. 2015;15:652. doi:10.1186/s12885-015-1663-5.
78. Cohen CW, Fontaine KR, Arend RC, Soleymani T, Gower BA. Favorable effects of a ketogenic diet on physical function, perceived energy, and food cravings in women with ovarian or endometrial cancer: a randomized, controlled trial. *Nutrients*. 2018;10:9. doi:10.3390/nu10091187.
79. Tan-Shalaby JL, Carrick J, Edinger K, Genovese D, Liman AD, Passero VA, Shah RB. Modified Atkins diet in advanced malignancies - final results of a safety and feasibility trial within the veterans affairs Pittsburgh healthcare system. *Nutr Metab*. 2016;13:52. doi:10.1186/s12986-016-0113-y.
80. Rieger J, Bähr O, Maurer GD, Hattingen E, Franz K, Brucker D, Walenta S, Kämmerer U, Coy JF, Weller M, et al. ERGO: a pilot study of ketogenic diet in recurrent glioblastoma. *Int J Oncol*. 2014;44(6):1843–1852. doi:10.3892/ijo.2014.2382.
81. Lapeyre-Prost A, Terme M, Pernot S, Pointet A-L, Voron T, Tartour E, Taieb J. Immunomodulatory activity of VEGF in cancer. *Int Rev Cell Mol Biol*. 2017;330:295–342. doi:10.1016/bs.ircmb.2016.09.007.
82. Jabeen S, Zucknick M, Nome M, Dannenfelser R, Fleischer T, Kumar S, Lüders T, von der Lippe Gythfeldt H, Troyanskaya O, Kyte JA, et al. Serum cytokine levels in breast cancer patients during neoadjuvant treatment with bevacizumab. *Oncoimmunology*. 2018;7(11):e1457598. doi:10.1080/2162402X.2018.1457598.
83. Kasznicki J, Sliwinska A, Drzewoski J. Metformin in cancer prevention and therapy. *Ann Transl Med*. 2014;2:6. doi:10.3978/j.issn.2305-5839.2014.06.01.
84. Cha J-H, Yang W-H, Xia W, Wei Y, Chan L-C, Lim S-O, Li C-W, Kim T, Chang -S-S, Lee -H-H, et al. Metformin promotes antitumor immunity via endoplasmic-reticulum-associated degradation of PD-L1. *Mol Cell*. 2018;71(4):606–620.e7. doi:10.1016/j.molcel.2018.07.030.
85. Qin G, Lian J, Huang L, Zhao Q, Liu S, Zhang Z, Chen X, Yue D, Li L, Li F, et al. Metformin blocks myeloid-derived suppressor cell accumulation through AMPK-DACH1-CXCL1 axis. *Oncoimmunology*. 2018;7(7):e1442167. doi:10.1080/2162402X.2018.1442167.
86. Pardoll DM. The blockade of immune checkpoints in cancer immunotherapy. *Nat Rev Cancer*. 2012;12(4):252–264. doi:10.1038/nrc3239.
87. Tang J, Yu JX, Hubbard-Lucey VM, Nefitelinov ST, Hodge JP, Lin Y. Trial watch: the clinical trial landscape for PD1/PDL1 immune checkpoint inhibitors. *Nat Rev Drug Discov*. 2018;17:854–855. doi:10.1038/nrd.2018.210.
88. Kantoff PW, Higano CS, Shore ND, Berger ER, Small EJ, Penson DF, Redfern CH, Ferrari AC, Dreicer R, Sims RB, et al. Sipuleucel-T immunotherapy for castration-resistant prostate cancer. *N Engl J Med*. 2010;363(5):411–422. doi:10.1056/NEJMoa1001294.
89. Hagihara K, Chan S, Zhang L, Oh DY, Wei XX, Simko J, Fong L. Neoadjuvant sipuleucel-T induces both Th1 activation and immune regulation in localized prostate cancer. *Oncoimmunology*. 2019;8(1):e1486953. doi:10.1080/2162402X.2018.1486953.
90. Wang A, Luan HH, Medzhitov R. An evolutionary perspective on immunometabolism. *Science*. 2019;363:6423. doi:10.1126/science.aar3932.
91. Renner K, Singer K, Koehl GE, Geissler EK, Peter K, Siska PJ, Kreutz M. Metabolic hallmarks of tumor and immune cells in the tumor microenvironment. *Front Immunol*. 2017;8. doi:10.3389/fimmu.2017.00248.
92. Catalán V, Gómez-Ambrosi J, Rodríguez A, Ramírez B, Ortega VA, Hernández-Lizoain JL, Baigauli J, Becerril S, Rotellar F, Valenti V, et al. IL-32 α -induced inflammation constitutes a link between obesity and colon cancer. *Oncoimmunology*. 2017;6(7):e1328338. doi:10.1080/2162402X.2017.1328338.
93. Zilberter T, Zilberter Y. Ketogenic ratio determines metabolic effects of macronutrients and prevents interpretive bias. *Front Nutr*. 2018;5. doi:10.3389/fnut.2018.00075.