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Genetic determinants of risk in pulmonary arterial hypertension: international genome-wide association studies and meta-analysis





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Summary

Background Rare genetic variants cause pulmonary arterial hypertension, but the contribution of common genetic variation to disease risk and natural history is poorly characterised. We tested for genome-wide association for pulmonary arterial hypertension in large international cohorts and assessed the contribution of associated regions to outcomes.

Methods We did two separate genome-wide association studies (GWAS) and a meta-analysis of pulmonary arterial hypertension. These GWAS used data from four international case-control studies across 11744 individuals with European ancestry (including 2085 patients). One GWAS used genotypes from 5895 whole-genome sequences and the other GWAS used genotyping array data from an additional 5849 individuals. Cross-validation of loci reaching genome-wide significance was sought by meta-analysis. Conditional analysis corrected for the most significant variants at each locus was used to resolve signals for multiple associations. We functionally annotated associated variants and tested associations with duration of survival. All-cause mortality was the primary endpoint in survival analyses.

Findings A locus near SOX17 (rs10103692, odds ratio 1.80 [95% CI 1.55-2.08], $p=5.13\times10^{-15}$) and a second locus in HLA-DPA1 and HLA-DPB1 (collectively referred to as HLA-DPA1/DPB1 here; rs2856830, 1.56 [1.42-1.71], $p=7.65\times10^{-20}$) within the class II MHC region were associated with pulmonary arterial hypertension. The SOX17 locus had two independent signals associated with pulmonary arterial hypertension (rs13266183, 1.36 [1.25-1.48], $p=1.69\times10^{-12}$; and rs10103692). Functional and epigenomic data indicate that the risk variants near SOX17 alter gene regulation via an enhancer active in endothelial cells. Pulmonary arterial hypertension risk variants determined haplotype-specific enhancer activity, and CRISPR-mediated inhibition of the enhancer reduced SOX17 expression. The HLA-DPA1/DPB1 rs2856830 genotype was strongly associated with survival. Median survival from diagnosis in patients with pulmonary arterial hypertension with the C/C homozygous genotype was double (13.50 years [95% CI 12.07 to >13.50]) that of those with the T/T genotype (6.97 years [6.02–8.05]), despite similar baseline disease severity.

Interpretation This is the first study to report that common genetic variation at loci in an enhancer near *SOX17* and in *HLA-DPA1/DPB1* is associated with pulmonary arterial hypertension. Impairment of SOX17 function might be more common in pulmonary arterial hypertension than suggested by rare mutations in *SOX17*. Further studies are needed to confirm the association between HLA typing or rs2856830 genotyping and survival, and to determine whether HLA typing or rs2856830 genotyping improves risk stratification in clinical practice or trials.

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Research in context

Evidence before this study

We searched PubMed for research articles published in English before Aug 23, 2018, with search terms including "pulmonary arterial hypertension", "genetics", and "GWAS". Rare genetic variation, primarily in genes associated with transforming growth factor- β family members, including BMPR2, but also in the transcription factor SOX17, is known to cause pulmonary arterial hypertension. However, little is known about the contribution of common variation to this disorder. Additionally, both pulmonary vascular endothelial dysfunction and altered immune and inflammatory signalling are observed in pulmonary arterial hypertension, but the underlying genetic mechanisms are poorly characterised.

Added value of this study

To our knowledge, this is the largest genetic analysis of pulmonary arterial hypertension to date, comprising more than 2000 patients with pulmonary arterial hypertension from four international cohorts. We identified one locus near SOX17 and another in HLA-DPA1 and HLA-DPB1 that reached genome-wide significance, and we cross-validated these loci by meta-analysis. The SOX17 locus includes two independent signals, both of which identify enhancer regions that specifically regulate the expression of SOX17, which is essential for pulmonary vascular development. Allelic variation at HLA-DPB1 is associated with clinical outcomes, specifically survival, with more than two-thirds of patients harbouring genotypes associated with the poorest outcomes.

Implications of all the available evidence

Common variation near SOX17 is a risk factor for pulmonary arterial hypertension and dysregulation of SOX17 might be more common in pulmonary arterial hypertension than the occurrence of rare variants suggests. Further studies are needed to define whether HLA typing or rs2856830 genotyping improves risk stratification in clinical practice and in clinical trials.

Introduction

Pulmonary arterial hypertension refers to an uncommon but devastating disorder characterised by obliterative pulmonary vascular remodelling, leading to a progressive increase in pulmonary vascular resistance and right heart failure. Annual mortality for idiopathic and heritable pulmonary arterial hypertension remains around 10%, despite the use of modern therapies. 1,2 The high mortality partly reflects the limited effect of licensed treatments on the underlying pulmonary vascular pathology, which includes vascular smooth muscle and fibroblast hyperplasia, endothelial cell proliferation, and inflammation.3 Substantial variation between patients in their response to available treatments highlights underlying and inadequately characterised heterogeneity in the causes of pulmonary arterial hypertension.

Recent gene sequencing studies⁴⁻⁶ have revealed rare mutations in several genes, including BMPR2, genes encoding potassium channels, and most recently the transcription factor SOX17. Rare genetic variation is associated with both the risk of developing pulmonary arterial hypertension and survival, and it is found in up to 25% of patients with pulmonary arterial hypertension. In the majority of patients with pulmonary arterial hypertension, the extent of genetic contribution, including that attributable to common variation, remains largely unknown.78 Therefore, we aimed to test for genome-wide association for pulmonary arterial hypertension in large international cohorts and assess the contribution of associated regions to patient outcomes (panel). This is the first report of the associations found at SOX17 and HLA-DPA1 and HLA-DPB1 (collectively referred to as HLA-DPA1/DPB1 in this Article).

Methods

Pulmonary arterial hypertension cohorts and genotyping We did two genome-wide association studies and a metaanalysis on pulmonary arterial hypertension. Pulmonary arterial hypertension was defined by haemodynamic criteria according to international guidelines.2 Unrelated individuals with idiopathic, heritable, or anorexigenassociated pulmonary arterial hypertension were included. Individuals with evidence of other known causes of pulmonary arterial hypertension were excluded; therefore, no patients were known to have pulmonary arterial hypertension associated with clinically diagnosable autoimmune diseases (appendix pp 2-3). All enrolled individuals provided written informed consent from their respective institutions or were included as anonymous controls under the DNA databank at Vanderbilt University, BioVU, opt-out policy (appendix p 2).

Given the rarity of pulmonary arterial hypertension, four studies were used for the analyses. In the UK National Institute for Health Research BioResource (NIHRBR) Rare Diseases study, whole-genome sequencing (Illumina, San Diego, CA, USA; mean depth of around $35\times$; appendix p 2) was done in 5895 individuals of European descent, each with a rare disorder from 16 categories or their unaffected relatives, and 847 had pulmonary arterial hypertension (appendix p 4). The concept of this study was to sequence patients with rare diseases to identify genetic influences on the pathogenesis of one rare disorder using the other rare diseases as controls, assuming that distinct rare diseases are highly unlikely to share common genetic mechanisms. This assumption was tested by repeating analyses excluding each major control group (appendix p 6).

Three studies used genome-wide genotyping arrays: the US National Biological Sample and Data Repository for Pulmonary Arterial Hypertension (also known as

Q Waisfisz PhD); Mayo Clinic

PAH Biobank [PAHB]) study9 included 694 individuals with pulmonary arterial hypertension and 1560 controls ascertained for a large pharmacogenomic study at Vanderbilt University (Nashville, TN, USA); the Pulmonary Hypertension Allele-Associated Risk (PHAAR) study⁷ included 269 individuals with pulmonary arterial hypertension and 1068 population-based controls from France; and the British Heart Foundation Pulmonary Arterial Hypertension (BHFPAH) study consisted of 275 individuals with pulmonary arterial hypertension and 1983 population-based controls from several European countries (appendix p 12). All genotyping studies were imputed (appendix p 12), and single-nucleotide polymorphisms (SNPs) with good imputation quality $(r^2 \ge 0.3)$ were taken forward for testing. Individuals from NIHRBR, PHAAR, and BHFPAH were tested for relatedness to prevent inclusion of the same or related individuals across studies. Other quality-control steps are detailed in the appendix (pp 4, 5, 12).

Association analyses

We used logistic regression to test single-marker variants for genetic association with a diagnosis of pulmonary arterial hypertension assuming a log-additive genetic model and adjusting for sex, read length chemistry (NIHRBR only), and population structure using the first four (NIHRBR and PHAAR), three (PAHB), or ten (BHFPAH) principal components. We calculated the genomic inflation factor, which was verified to be between 1 and 1.05 for each study.

We used two independent sets for discovery: wholegenome sequencing data from NIHRBR (n=5895, including 847 pulmonary arterial hypertension cases); and meta-analysis of genotyping studies PAHB, PHAAR, and BHFPAH (n=5849, including 1238 pulmonary arterial hypertension cases). We cross-validated findings and confirmed loci in a meta-analysis of all four studies using the inverse variance-weighted fixed-effects approach (which maximises power for discovery studies¹⁰), implemented in the GWAMA software tool. Random-effects meta-analysis was subsequently applied to estimate generalisability of the results to different populations. We did a conditional analysis including the lead variant in each locus as a covariate to test for independent distinct signals reaching p<5×10⁻⁸.

We used LDlink to assess linkage disequilibrium of variants in all European populations from the 1000 Genomes Project. Credible sets of variants considered 99% likely to include the functional causal variants were calculated by summing ranked posterior probabilities (appendix p 6).

Annotation and functional assessment of the locus near SOX17

The locus near SOX17 was assessed against publicly available functional annotation datasets (including

Panel: Key terms

Genome-wide association study

A genetic analysis approach, typically using millions of common variants (eg, single-nucleotide polymorphisms [SNPs]) covering the genome, to test whether an allele of a genetic variant is associated with a disease or trait, or the levels of a continuous trait of interest.

Common variant

An SNP for which the frequency of the less frequent allele is at least 5% in a given population. Typically, a common variant has subtle biological effects, as opposed to rare variants (also known as mutations), which can cause diseases or extreme phenotypes.

Genetic locus

A position on the genome defined by the chromosome number and the genetic distance in centimorgans (cM) or physical distance in base pairs (bps) on the chromosome.

A locus can refer to a gene or a non-coding region of varying length (eg, from one hundred to millions of bps).

Credible set

A set of variants that is statistically likely (eg, with 99% probability) to contain the causal variant for the disease or trait of interest at a genetic locus.

ENCODE, Factorbook Motifs, and Blueprint). The locus was investigated using CRISPR-mediated repression in human pulmonary artery endothelial cells (hPAECs; PromoCell GmbH, Heidelberg, Germany) by transduction with a lentivirus containing a plasmid encoding the nuclease-deficient Cas9 (dCas9) fused to the repressor KRAB and a 20 bp guide RNA (appendix pp 6–7). Cells were harvested following blasticidin selection, and the expression of *SOX17* as well as neighbouring *MRPL15* and *TMEM68* was assessed by quantitative PCR.

In-vitro enhancer activity of the loci and variants near SOX17 was investigated using a luciferase reporter assay. Specifically, genomic DNA was isolated from endothelial progenitor cells (also known as blood outgrowth endothelial cells) derived from a patient with pulmonary arterial hypertension who was heterozygous for the lead SNP at SOX17 and used to clone 100 bp putative enhancer regions containing the SOX17 pulmonary arterial hypertension variants. The cloned products were inserted into a luciferase reporter plasmid, which was subsequently used for transformation of stable bacteria. Picking various bacterial colonies allowed for isolation of luciferase reporter plasmids containing genomic DNA inserts differing only by the allele of the SNP of interest. Reporter plasmids were transfected into hPAECs by electroporation, and luciferase activity was measured to quantify the enhancer function of the inserts with the relevant haplotype.

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See Online for appendix
For more on **BioVU** see https://
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For more on **LDlink** see https://ldlink.nci.nih.gov/

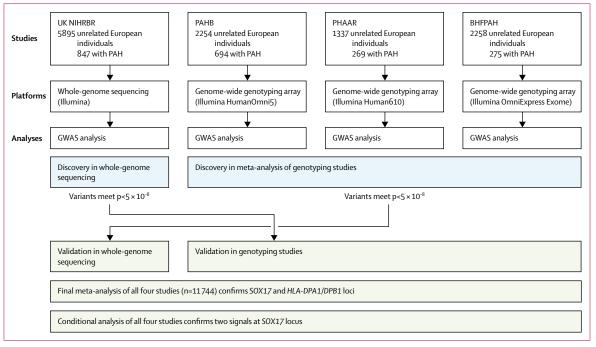


Figure 1: Study design

HLA-DPA1 and HLA-DPB1 are collectively referred to as HLA-DPA1/DPB1 in this Article. BHFPAH=British Heart Foundation Pulmonary Arterial Hypertension study. GWAS=genome-wide association study. NIHRBR=National Institute for Health Research BioResource study. PAH=pulmonary arterial hypertension. PAHB=PAH Biobank study. PHAAR=Pulmonary Hypertension Allele-Associated Risk study.

Statistical analysis

Loci associated with pulmonary arterial hypertension were tested for associations with clinical variables (appendix pp 17-18). All-cause mortality was the primary endpoint in survival analyses using Kaplan-Meier estimates and Cox regression in the survival package in R, version 3.3.0.12 Survival was calculated from diagnosis to date of death or censoring (Oct 31, 2016, for NIHRBR; Aug 1, 2017, for PAHB; Sept 27, 2017, for PHAAR; Oct 12, 2017, for BHFPAH), with left truncation using date of genetic consent, and patients were censored at lung or heart-and-lung transplantation. Age and sex were covariates to correct for their association with prognosis.2 NIHRBR and PAHB were analysed separately, and PHAAR and BHFPAH were combined before analysis because of their smaller sample sizes. Cox regression results from these three analyses were then meta-analysed using the default random-effects model restricted maximum-likelihood estimator method implemented in the metafor package in R, version 3.3.0.13 All cohorts were combined for Kaplan-Meier analysis. We did sensitivity analyses excluding pathogenic BMPR2 variant carriers, all pathogenic rare variant carriers, and patients diagnosed in previous decades who might have been exposed to different treatment regimens.

HLA alleles and amino acids totalling 1873 features were determined by imputation from genotyped and high-quality imputed variants in the HLA region using the SNP2HLA software and the type 1 diabetes genetics consortium reference database. HLA alleles and amino

acids were tested for association with the novel lead variants or case-control status by χ^2 test with false discovery rate correction.

Role of the funding source

The funders of the study had no role in study design, data collection, data analysis, data interpretation, or writing of the report. CJR, KB, MB, MHa, LSo, MG, MWP, CH, AA, KBH, JHK, MK, AU, LH, JA, EMS, and SGr had access to raw data for analyses. The corresponding authors had full access to all the data in the study and had final responsibility for the decision to submit for publication.

Results

In two separate GWAS discovery analyses (figure 1), we identified two loci associated with pulmonary arterial hypertension reaching genome-wide significance (p<5×10⁻⁸; table 1; appendix p 23). One locus was 100–200 kb upstream of the transcription factor *SOX17*. A second locus was within *HLA-DPA1/DPB1*, which encodes the MHC class II DP α and β chains.

Both the SOX17 and HLA-DPA1/DPB1 loci reached genome-wide significance in the discovery analyses; our cross-validation strategy confirmed that the same alleles were more frequent in pulmonary arterial hypertension than in other disease or population controls in both analyses (table 1). The genome-wide meta-analysis of all four studies confirmed their associations with pulmonary arterial hypertension (rs2856830, odds ratio 1.56

	Chromosome and position, hg19:effect/ non-effect alleles	Effect allele frequency in non-Finnish Europeans in gnomAD	Effect allele frequency in NIHRBR controls	UK NIHRBR wh sequencing stu vs 5048 contro	dy (847 cases	Effect allele frequency in genotyping controls	Meta-analysis genotyping s PHAAR, and E (1238 cases vs 4611 controls	tudies PAHB, SHFPAH S	Meta-analysis of all cohorts (2085 cases, 9659 controls, n _{er} =6648)	
				Odds ratio (95% CI)	p value		Odds ratio (95% CI)	p value	Odds ratio (95% CI)	Meta-analysis p value
Lead SNPs										
HLA-DPA1/DPB1, rs2856830	6:33041734:C/T	0.12	0.12	1·71 (1·48–1·96)	4·41×10 ^{-14*}	0.13	1·44 (1·26-1·64)	5·35×10 ⁻⁸	1.56 (1.42-1.71)	7-65×10 ⁻²⁰ *
SOX17, signal 1 rs13266183	8:55267612:C/T	0.73	0.73	1·44 (1·26–1·64)	4·44×10 ^{-8*}	0.74	1·31 (1·17-1·46)	$4 \cdot 1 \times 10^{-6}$	1-36 (1-25-1-48)	1.69×10 ⁻¹² *
SOX17, signal 2 rs10103692	8:55258127:G/A	0.90	0.90	1·85 (1·47–2·31)	9·51×10 ⁻⁸	0.91	1·76 (1·45-2·14)	9·84×10 ^{-9*}	1.80 (1.55-2.08)	5·13×10 ⁻¹⁵ *
Other SNPs in same loc	i									
HLA-DPB1 missense SNP, rs1042140	6:33048640:G/A	0.23	0.23	1·38 (1·22-1·55)	9·21×10 ⁻⁸	0.23	1·44 (1·29–1·61)	9·73×10 ⁻¹¹ *	1.41 (1.30-1.53)	7·13×10 ⁻¹⁷ *
SOX17, genotyping lead SNP, rs28576721†	8:55265980:T/C	0.91	0.92	1·55 (1·23–1·95)	1·57×10 ⁻⁴	0.92	1·96 (1·57-2·43)	1·54×10 ⁻⁹ *	1.75 (1.50–2.05)	3·07×10 ⁻¹² *

Odds ratios are for association between effect allele and pulmonary arterial hypertension. gnomAD is the Genome Aggregation Database, which provides information including allele frequencies in different populations. HLA-DPA1 and HLA-DPB1 are collectively referred to as HLA-DPA1/DPB1 in this Article. BHFPAH=British Heart Foundation Pulmonary Arterial Hypertension study. n_{ef}=number of individuals that would make up an equally powered study with a 1:1 case:control ratio (appendix p 2). NIHRBR=National Institute for Health Research BioResource study. PAH=pulmonary arterial hypertension. PAHB=PAH Biobank study. PHAAR=Pulmonary Hypertension Allele-Associated Risk study. SNP=single-nucleotide polymorphism. *Significant. †This is the most significant SOX17 SNP after combining the three genotyping studies (not including NIHRBR) and forms part of signal 2.

Table 1: Novel loci associated with pulmonary arterial hypertension in sequenced and genotyped cohorts

[95% CI 1.42–1.71], p= 7.65×10^{-20} for *HLA-DPA1/DPB1*; rs10103692, 1.80 [1.55–2.08], p= 5.13×10^{-15} for *SOX17*; table 1; figure 2; appendix p 13). We detected no further loci at genome-wide significance. Allele frequencies in the different control groups were similar between studies and to non-Finnish Europeans in the public database gnomAD (table 1).

The conditional analysis confirmed that the HLA-DPA1/ DPB1 locus contained a single signal of association, but showed that the SOX17 locus was composed of two independent signals; signal 1 is 100-103 kb upstream of SOX17 ($p_{conditional}$ = 9.82×10^{-9}) and signal 2 is 106–200 kb upstream of SOX17 ($p_{conditional}=4\cdot16\times10^{-11}$; figure 2; appendix p 15). The lead SNPs for the two signals in the SOX17 locus were rs13266183 (for signal 1, odds ratio 1.36 [1.25-1.48], p=1.69×10⁻¹²) and rs10103692 (signal 2). A Bayesian credible set analysis to narrow the variants in these loci to those 99% likely to be causal (appendix p 15) showed that the HLA-DPA1/DPB1 locus included nine SNPs (all p<9·1×10-18), SOX17 signal 1 included four SNPs 100-103 kb upstream of SOX17 (all p<3·3×10⁻⁸), and SOX17 signal 2 included 31 SNPs 106-142 kb upstream of SOX17 (all p<5.7×10⁻¹⁰).

Previous studies have reported the association of variants near *CBLN2*, and *PDE1A* and *DNAJC10* with pulmonary arterial hypertension.⁷⁸ These common variant signals showed no association with pulmonary arterial hypertension in the combined NIHRBR, PAHB, and BHFPAH cohorts (p=0·17 for *CBLN2* and p=0·24 for *PDE1A* and *DNAJC10*; appendix p 14). Sensitivity analyses excluding pathogenic *BMPR2* variant carriers, all pathogenic rare variant carriers, and controls from

different disease groups yielded similar results to the main analyses (appendix p 9).

To search for evidence of regulatory elements in relevant tissues at *SOX17* signal 1 and signal 2, we examined publicly available epigenomic data (including histone modifications; figure 3; appendix p 24). We identified several putative enhancer elements active in both lung tissue and endothelial cells (figure 3). One of these elements (around hg19-chr8:55·270 Mb) contains a cluster of three of four credible variants from *SOX17* signal 1 (figure 3). Another (around hg19-chr8:55·252 Mb) contains one credible variant from *SOX17* signal 2. Of these variants, rs10958403 in signal 1 and rs765727 in signal 2 overlap a DNase I hypersensitivity signal, which indicates accessible chromatin (allowing binding of transcription factors), detected in hPAECs (figure 3).

To study the effects of the pulmonary arterial hypertension risk variants on the putative enhancers defined by the epigenomic signals, we developed reporter constructs containing 100 bp of the regions containing either the risk allele or non-risk alleles at each of the four SNPs using genomic DNA from a patient heterozygous for both SOX17 signals. A haplotype-specific reporter assay in hPAECs confirmed that the regions containing either rs10958403 or rs765727 exhibited enhancer activity (between threefold and sixfold induction of luciferase: Renilla ratio, p<0.0001), whereas constructs containing rs12674755 or rs12677277 had no effect compared with the empty vector control (figure 4). We also observed haplotype-specific activity with the active constructs, which differed only by the alleles at pulmonary arterial hypertension-associated risk variants rs10958403 or rs765727 (both p<0.05; figure 4).

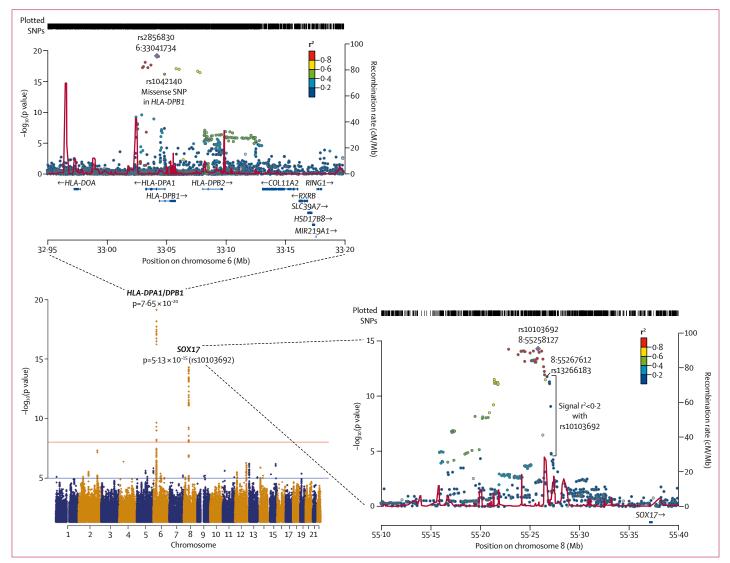


Figure 2: A meta-analysis of all cohorts and regional plots of novel loci
The regional plots indicate variant location at the HLA-DPA1 and HLA-DPB1 (collectively referred to as HLA-DPA1/DPB1 in this Article) locus and linkage disequilibrium structure at SOX17 locus. At the SOX17 locus, several variants associated with pulmonary arterial hypertension are in very weak or no linkage disequilibrium (r²<0·2) with the lead single-nucleotide polymorphism (SNP), rs10103692. We refer to these variants as SOX17 signal 1 and the most significant variant, rs13266183, is indicated. The variants coloured as in linkage disequilibrium with rs10103692 comprise signal 2.

DNA folding patterns determined by Hi-C data from lung tissue and endothelial cells (human umbilical vein endothelial cells [figure 3] and human microvascular endothelial cells [data not shown]) indicate that the *SOX17* pulmonary arterial hypertension locus resides in a defined topologically associated domain in which the only gene found, and thus likely target of any regulatory elements in this region, is *SOX17*. CRISPR-mediated inhibition of the *SOX17* signal 1 region in hPAECs resulted in selective downregulation of *SOX17* expression but not the expression of neighbouring genes *MRPL15* and *TMEM68*, suggesting that the enhancers in this locus specifically regulate *SOX17* (figure 4; appendix p 26).

We investigated whether the *HLA-DPA1/DPB1* and *SOX17* variants affect clinical outcomes in pulmonary

arterial hypertension, specifically all-cause mortality. The *HLA-DPA1/DPB1* rs2856830 genotype, but not the *SOX17* locus, was strongly associated with survival (figure 5). Median survival from diagnosis in patients with pulmonary arterial hypertension with the C/C homozygous genotype was double (13 · 50 years [95% CI $12 \cdot 07$ to $>13 \cdot 50$]) that of those with the T/T genotype (6 · 97 years [6 · 02 – 8 · 05]). Cox regression survival analyses showed that the rs2856830 T/T genotype conferred an increased annual risk of death in pulmonary arterial hypertension (hazard ratio [HR] $1 \cdot 94$ [95% CI $1 \cdot 08 - 3 \cdot 51$]; figure 5).

Sensitivity analyses excluding pathogenic *BMPR2* variant carriers, all pathogenic rare variant carriers, and patients diagnosed in previous decades who might have

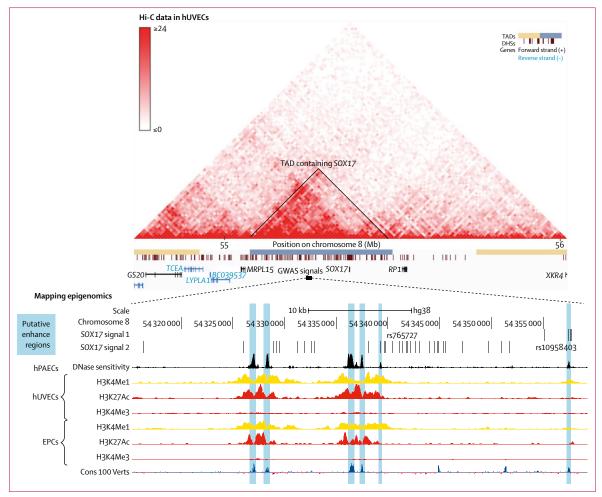


Figure 3: In-silico analysis of SOX17 locus

Hi-C data from human umbilical vein endothelial cells (hUVECs) indicate regions of DNA found in close proximity in the three-dimensional structure. The genomic region containing the significant variants identified by the genome-wide association study (GWAS) analysis is indicated by a black box, overlapping a topologically associated domain (TAD) indicated in blue, which contains only SOX17. Mapping of SOX17 locus variants associated with pulmonary arterial hypertension with public epigenomic data is underneath Hi-C data. The credible set indicates positions of variants 99% likely to contain the causal variants. Auxiliary hidden Markov models, which summarise epigenomic data to predict the functional status of genomic regions in different tissues or cells, are shown. Epigenomic data in endothelial cells including hUVECs, human pulmonary artery endothelial cells (hPAECs), and endothelial progenitor cells (EPCs), indicate areas likely to contain active regulatory regions and promoters. Markers include histone H3 lysine 4 monomethylation (H3K4Me1; often found in enhancers) and trimethylation (H3K4Me3; strongly observed in promoters) and H3 lysine 27 acetylation (H3K27Ac; often found in active regulatory regions). The blue vertical blocks indicate where epigenomic data suggest a putative enhancer region, some overlapped by variants associated with pulmonary arterial hypertension. These regions were cloned for the luciferase reporter experiments (figure 4B). DHSs=DNase I hypersenstivity sites.

been exposed to different treatment regimens gave results similar to the main analyses (appendix p 27).

We tested both loci for association with other clinical variables, including disease severity measures and comorbidities (appendix pp 17–18). The C allele at *HLA-DPA1/DPB1* lead SNP rs2856830 was associated with younger age at diagnosis (figure 5), with C/C homozygotes presenting a decade earlier than T/T homozygotes (appendix p 17). The rs2856830 genotype was not associated with vasoresponder status.

The *HLA-DPA1/DPB1* locus included a missense variant rs1042140 in *HLA-DPB1* reaching genome-wide significance (table 1) in partial linkage disequilibrium

(r^2 =0·45 with lead rs2856830 in Europeans). The SNP rs1042140 determines a glutamic acid (Glu⁶⁹) or a lysine at amino acid residue 69. To determine specific HLA alleles associated with the lead variant, rs2856830, we imputed HLA types from the genotype data. These types are represented by digit codes, where the first two digits represent related groups of similar alleles (eg, DPB1*02), and four digits represent specific proteins with distinct amino acid sequences (eg, DPB1*02:01). We found that the pulmonary arterial hypertension-enriched C allele of rs2856830 was associated with HLA-DPB1*02:01/02:02/16:01 (all p<1×10-9 after false discovery rate correction; table 2; appendix p 19), which

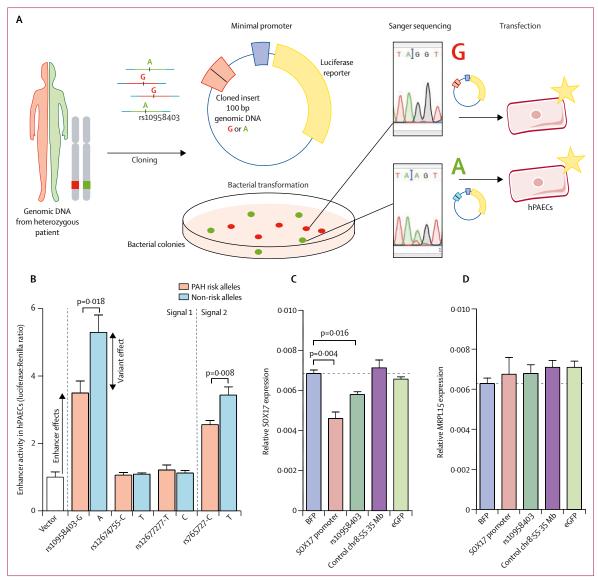


Figure 4: In-vitro analysis of SOX17 locus

(Å) Process for haplotype-specific reporter construct derivation. 100 bp genomic DNA inserts containing SOX17 single-nucleotide polymorphisms (SNPs) are isolated from endothelial progenitor cells derived from a patient with pulmonary arterial hypertension (PAH) heterozygous for the SOX17 SNPs. Colonies of transformed bacteria can be sequenced to determine alleles present in the product. Transfection of luciferase reporter constructs containing inserts into human pulmonary artery endothelial cells (hPAECs) allows for determination of luciferase activity. (B) Luciferase reporter assay results. Luciferase:Renilla ratios relative to the empty vector demonstrate haplotype-dependent enhancement of promoter activity. Enhancer effects were tested by one-way analysis of variance followed by Dunnett's post-hoc tests: rs10958403-G/A and rs765727-C/T were both p<0.0001 significant versus empty vector; variant effects of these two SNPs were tested by t test. The mean (SEM; error bars) of five experiments is shown. (C) Relative expression of SOX17:ACTB in hPAECs on CRISPR-mediated repression of the near SOX17 genome-wide association study (GWAS) locus. The mean (SEM; error bars) of four measurements in a representative experiment is shown. Three further experiments showed consistent results. Blue fluorescent protein (BFP), enhanced green fluorescent protein (eGFP), and control, which refers to a region between the enhancer region and the SOX17 gene that is negative for regulatory markers, are used as negative controls. The SOX17 promoter was targeted as a positive control of repression. Significance shown versus BFP by Dunnett's post-hoc analysis. (D) Relative expression of MRPL15:ACTB in hPAECs on CRISPR-mediated repression of the GWAS locus.

all contain the Glu⁶⁹ residue. The most numerous *DPB1**02:01 and *DPB1**04:01 alleles were associated with survival in patients with pulmonary arterial hypertension (HR 0.70 [95% CI 0.49-1.00] for *DPB1**02:01 and 1.33 [1.04-1.70] for *DPB1**04:01; appendix p 21).

The risk alleles at both signals within the SOX17 locus are common (risk allele frequencies are 74% for

rs13266183-C and 92% for rs9298503-C), such that 1230 (59%) of 2085 patients with pulmonary arterial hypertension were homozygous for the risk allele at both *SOX17* SNPs, compared with 4443 (46%) of 9659 controls.

The alleles at *HLA-DPB1* associated with the poorest outcomes are also common (risk allele frequency of 86% for rs2856830-T), such that 1432 (69%) of

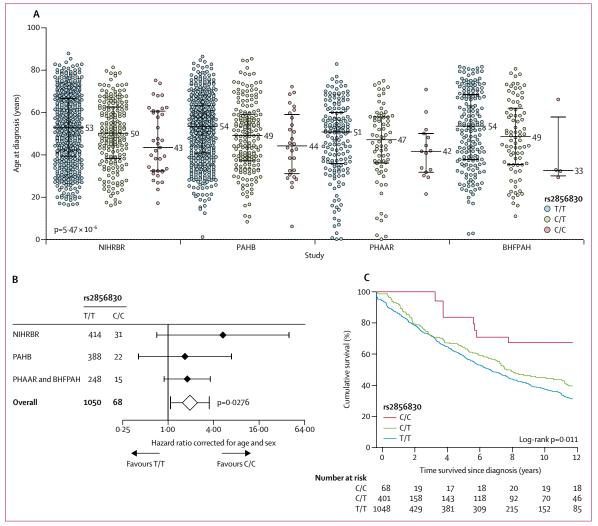


Figure 5: Clinical impact of HLA-DPB1 rs2856830

(A) Age at diagnosis by genotype in four cohorts of patients with pulmonary arterial hypertension (PAH). Bars indicate medians (IQRs); numbers given are median values in subgroups. The p value shown is from a linear regression model correcting for cohort differences. (B) Forest plot showing hazard ratios for the rs2856830 6:33041734 T/T versus C/C genotypes, corrected for age and sex in Cox regression survival analyses in each cohort, individually and with meta-analysis results. Error bars indicate 95% CIs. (C) Kaplan-Meier survival plot in patients with pulmonary arterial hypertension divided into groups on the basis of the genotype of HLA-DPA1 and HLA-DPB1 single-nucleotide polymorphism (SNP) rs2856830 in all cohorts. Numbers at risk indicates numbers at risk in each time period, which increases as truncated patients are recruited into the study after diagnosis and decreases as patient follow-up ends. Significance from the log rank test is given. BHFPAH-British Heart Foundation Pulmonary Arterial Hypertension study. NIHRBR=National Institute for Health Research BioResource study. PAHB=PAH Biobank study. PHAAR=Pulmonary Hypertension Allele-Associated Risk study.

2085 patients with pulmonary arterial hypertension had the T/T genotype associated with the poorest outcomes and 1975 (95%) of 2085 patients had at least one T allele.

Discussion

Through a meta-analysis of 11744 individuals, we have established loci at an enhancer upstream of *SOX17* and at *HLA-DPA1/DPB1* associated with pulmonary arterial hypertension disease risk. Common genetic variants in the enhancer region of *SOX17* are biologically plausible candidates for susceptibility to pulmonary vascular disease. Polymorphic variation at the *HLA-DPA1/DPB1* locus is strongly associated with both the

age at diagnosis and prognosis in pulmonary arterial hypertension.

Both in-silico and experimental analyses of the common variants upstream of the *SOX17* gene suggest that they affect susceptibility to pulmonary arterial hypertension through regulation of *SOX17* expression. We provided direct evidence that inhibition of *SOX17* signal 1 reduced *SOX17* expression, and luciferase activity experiments showed a functional variant in both signals. Combined with the in-silico data, it seems highly likely that signal 2 would also be targeting *SOX17* because no other gene is present in the topologically associated domain and promoter capture HiC data show that this area associates

Table 2: Associations of HLA-DPB1 alleles with the lead SNP rs285683

	Amino acid residues in HLA-DPB1 alleles															Frequencies by GWAS SNP rs2856830			q after FDR correction				
	8	9	11	33	35	36	55	56	57	65	69	76	84	85	86	87	96	178	194	T/T	T/C	C/C	
DPB1*02:01†	L	F	G	E	F	V†	D†	E†	E	ı	E†	M	G	G	Р	М	R	L	R	244/8870 (3%)	1182/2682 (44%)	211/234 (90%)	<5×10 ⁻²⁴⁷
DPB1*02:02†	L	F	G	E	L	V†	E	Α	E	I	E†	M	G	G	Р	M				2/8870 (<1%)	93/2682 (3%)	16/234 (7%)	2·77×10 ⁻⁸⁷
DPB1*16:01†	L	F	G	E	F	V†	D†	E†	E	I	E†	M	D	E	Α	V				4/8870 (<1%)	64/2682 (2%)	5/234 (2%)	7·08×10 ⁻⁴¹
DPB1*03:01‡	V	Υ	L	E	F	V†	D†	E†	D	L	K‡	V	D	E	Α	V	K	L	R	1094/8870 (12%)	152/2682 (6%)	0/234 (0%)	5·50 × 10 ⁻²³
DPB1*04:01‡	L	F	G	E	F	A‡	A‡	A‡	E	I	K‡	M	G	G	Р	M	R	L	R	4251/8870 (48%)	697/2682 (26%)	1/234 (<1%)	2·40×10 ⁻¹³⁸
DPB1*04:02‡	L	F	G	E	F	V†	D†	E†	E	I	K‡	M	G	G	Р	M	R	M	R	1153/8870 (13%)	166/2682 (6%)	0/234 (0%)	2·08 × 10 ⁻²³
DPB1*01:01‡	V	Υ	G	E	Υ	A‡	A‡	A‡	Е	I	K‡	V	D	E	Α	V	K	L	Q	611/8870 (7%)	99/2682 (4%)	0/234 (0%)	1·17×10 ⁻⁸
Data are n/N (%) unless otherwise stated. FDR=false discovery rate. GWAS=genome-wide association study. SNP=single-nucleotide polymorphism. †Alleles and residues depleted in pulmonary arterial hypertension cases. ‡Alleles and residues enriched in pulmonary arterial hypertension cases.																							

with the SOX17 promoter. We have recently reported4 enrichment and familial segregation in pulmonary arterial hypertension of causal rare deleterious variation in SOX17, implicating this gene in the pathogenesis of pulmonary arterial hypertension. SOX17 is involved in the development of the endoderm, 15-17 vascular endothelium, haemopoietic cells,18 and cardiomyocytes.19,20 SOX17 also determines the endothelial fate of CD34 progenitor cells de-differentiated from fibroblasts.21 Deletion in the mouse leads to abnormal pulmonary vascular development, poor distal lung perfusion and biventricular hypertrophy.22 SOX17 is a pro-angiogenic transcription factor and interacts with well established endothelial molecular mediators;23,24 reduction of SOX17 in endothelial cells through Notch activation (which is associated with BMPR2 signalling²⁵) restricts angiogenesis.²³ Conversely, vascular endothelial growth factor (VEGF) upregulates SOX17 and, as part of a positive-feedback loop, SOX17 promotes expression of VEGF receptor 2.24

We report that HLA-DPB1 alleles are associated with pulmonary arterial hypertension and have a pivotal role in determining disease progression. The beneficial effect of the C/C genotype at rs2856830 on survival is clinically significant, extending average survival from about 7 to about 14 years, despite no apparent difference in baseline disease severity by standard clinical measures, including haemodynamics and exercise capacity. Patients with the C allele at rs2856830 presented at a significantly younger age than those with the T allele, but the association of the HLA-DPB1 SNP with survival remains significant after correction for both age and sex. The somewhat conflicting observation that the C/C genotype is associated with earlier, more frequent presentation but improved survival compared with the C/T or T/T genotypes perhaps suggests that there could be two different mechanisms involved; one that affects initial disease pathogenesis and another

that alters the adaptation to the established disease state. A parallel in pulmonary arterial hypertension is the paradox of female prevalence contrasted with poorer outcomes for male patients,² although the mechanisms for this still remain unclear. Further evaluation of this survival association in independent datasets would help to define how clinical HLA typing or rs2856830 genotyping could improve risk stratification in clinical practice and in clinical trials, in which over-representation of the C/C genotype in one treatment group could significantly affect outcomes.

The mechanism of rs2856830 involvement in pulmonary arterial hypertension is probably through its association with specific HLA-DPB1 alleles. Class II (HLA-DRB1, HLA-DQB1, and HLA-DPB1) antigen-presenting proteins have crucial roles in the adaptive immune response.^{26,27} The HLA-DPB1 alleles associated with rs2856830 (HLA-DPB1*02:01/02:02/16:01) in the current study have also previously been linked to susceptibility to hard metal lung diseases, such as berylliosis. 28,29 A number of individual amino acid residues in the peptide-binding pockets of the HLA-DPB1 molecule affect its function and T-cell recognition, either by changing peptide antigen binding or the conformation of the peptide-binding groove.30 HLA-DPB1*02:01/ 02:02/16:01 all contain a glutamate at position 69 and a valine at position 36 that reduce the risk of clinical deterioration. These same residues are essential for T-cell activation and cytokine production in berylliosis.31,32 The potential role of this modification in antigen binding, autoimmune response, and vascular damage in pulmonary arterial hypertension demands further investigation.

To examine whether the associations observed were driven by *trans* effects of known rare pathogenic variants in pulmonary arterial hypertension, we did sensitivity analyses that demonstrated that the associations were independent of *BMPR2* and other rare pathogenic variants.

Although the sequencing and array platforms used in this study might perform differently across the genome, the signals detected for each platform remained strong.

This study has some limitations. The majority of patients studied were prevalent cases, and the association with survival is only marginally significant and based on a relatively small sample size, from multiple studies with different ascertainment criteria. Thus, confirmation of the survival analysis in an independent sample of patients recruited at diagnosis would greatly increase confidence in this finding. Some variants displayed heterogeneity of effects between studies, which is most likely due to the limited sample size in the smaller genotyping studies. Variants in *CBLN2* and other loci previously associated with pulmonary arterial hypertension? were not replicated by this study, suggesting that these previous findings were either false positives or only relevant to the specific subpopulations studied.

We have shown in a rare disorder that common variation can drive significant clinical differences in presentation and outcomes. Furthermore, a common non-coding variant can regulate expression of a gene linked by rare, deleterious mutations to the same disease. *HLA-DPB1*, and wider immune regulatory pathways, should be considered a priority for patient stratification and investigation of new treatments in pulmonary arterial hypertension. *SOX17* is a key endothelial regulator and its dysfunction in pulmonary arterial hypertension might be more common than suggested by the occurrence of rare pathogenic variants in heritable cases.

Contributors

All authors were involved in study design, data collection, and constructive criticism of the manuscript. CJR, KB, MB, MHa, LSo, MG, MWP, JA, LH, IP, RK, SGr, WCN, RCT, AAD, NWM, and MRW were involved in data interpretation and writing of the manuscript. Data were analysed by CJR, KB, MB, MHa, LSo, MG, MWP, CH, JA, AA, KBH, JHK, MK, AU, LH, EMS, and SGr.

Declaration of interests

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References

- McGoon MD, Benza RL, Escribano-Subias P, et al. Pulmonary arterial hypertension: epidemiology and registries. J Am Coll Cardiol 2013; 62 (25 suppl): D51–59.
- 2 Galie N, Humbert M, Vachiery JL, et al. 2015 ESC/ERS guidelines for the diagnosis and treatment of pulmonary hypertension: the Joint Task Force for the Diagnosis and Treatment of Pulmonary Hypertension of the European Society of Cardiology (ESC) and the European Respiratory Society (ERS): Endorsed by: Association for European Paediatric and Congenital Cardiology (AEPC), International Society for Heart and Lung Transplantation (ISHLT). Eur Respir J 2015; 46: 903–75.
- 3 Stacher E, Graham BB, Hunt JM, et al. Modern age pathology of pulmonary arterial hypertension. Am J Respir Crit Care Med 2012; 186: 261–72.
- 4 Gräf S, Haimel M, Bleda M, et al. Identification of rare sequence variation underlying heritable pulmonary arterial hypertension. *Nat Commun* 2018; 9: 1416.
- 5 Hiraide T, Kataoka M, Suzuki H, et al. SOX17 mutations in Japanese patients with pulmonary arterial hypertension.
 Am J Respir Crit Care Med 2018; 198: 1231–33.
- 6 Zhu N, Welch CL, Wang J, et al. Rare variants in SOX17 are associated with pulmonary arterial hypertension with congenital heart disease. Genome Med 2018; 10: 56.
- 7 Germain M, Eyries M, Montani D, et al. Genome-wide association analysis identifies a susceptibility locus for pulmonary arterial hypertension. *Nat Genet* 2013; 45: 518–21.
- 8 Kimura M, Tamura Y, Guignabert C, et al. A genome-wide association analysis identifies PDE1A|DNAJC10 locus on chromosome 2 associated with idiopathic pulmonary arterial hypertension in a Japanese population. Oncotarget 2017; 8: 74917–26.

- 9 Bowton E, Field JR, Wang S, et al. Biobanks and electronic medical records: enabling cost-effective research. Sci Transl Med 2014; 6: 234cm3.
- 10 Pereira TV, Patsopoulos NA, Salanti G, Ioannidis JP. Discovery properties of genome-wide association signals from cumulatively combined data sets. Am J Epidemiol 2009; 170: 1197–206.
- Magi R, Morris AP. GWAMA: software for genome-wide association meta-analysis. BMC Bioinformatics 2010; 11: 288.
- 12 Therneau T. A package for survival analysis in S version 2.38. 2015. https://CRAN.R-project.org/package=survival (accessed Oct 31, 2016).
- 13 Viechtbauer W. Conducting meta-analyses in R with the metafor package. J Stat Soft 2010; 36: 48.
- 14 Jia X, Han B, Onengut-Gumuscu S, et al. Imputing amino acid polymorphisms in human leukocyte antigens. PLoS One 2013; 8: e64683.
- 15 Hudson C, Clements D, Friday RV, Stott D, Woodland HR. Xsox17α and -β mediate endoderm formation in Xenopus. Cell 1997; 91: 397–405.
- 16 Alexander J, Stainier DY. A molecular pathway leading to endoderm formation in zebrafish. Curr Biol 1999; 9: 1147–57.
- 17 Kanai-Azuma M, Kanai Y, Gad JM, et al. Depletion of definitive gut endoderm in Sox17-null mutant mice. Development 2002; 129: 2367–79.
- 18 Kim I, Saunders TL, Morrison SJ. Sox17 dependence distinguishes the transcriptional regulation of fetal from adult hematopoietic stem cells. Cell 2007; 130: 470–83.
- 19 Zhang C, Basta T, Klymkowsky MW. SOX7 and SOX18 are essential for cardiogenesis in Xenopus. Dev Dyn 2005; 234: 878–91.
- 20 Liu Y, Asakura M, Inoue H, et al. Sox17 is essential for the specification of cardiac mesoderm in embryonic stem cells. Proc Natl Acad Sci USA 2007; 104: 3859–64.
- Zhang L, Jambusaria A, Hong Z, et al. SOX17 regulates conversion of human fibroblasts into endothelial cells and erythroblasts by dedifferentiation into CD34+ progenitor cells. *Circulation* 2017; 135: 2505–23.
- 22 Lange AW, Haitchi HM, LeCras TD, et al. Sox17 is required for normal pulmonary vascular morphogenesis. *Dev Biol* 2014; 387: 109–20.
- 23 Lee SH, Lee S, Yang H, et al. Notch pathway targets proangiogenic regulator Sox17 to restrict angiogenesis. Circ Res 2014; 115: 215–26.
- 24 Kim K, Kim IK, Yang JM, et al. SoxF transcription factors are positive feedback regulators of VEGF signaling. Circ Res 2016; 119: 839–52.
- 25 Hurst LA, Dunmore BJ, Long L, et al. TNFα drives pulmonary arterial hypertension by suppressing the BMP type-II receptor and altering NOTCH signalling. Nat Commun 2017; 8: 14079.
- 26 Shiina T, Hosomichi K, Inoko H, Kulski JK. The HLA genomic loci map: expression, interaction, diversity and disease. J Hum Genet 2009; 54: 15–39.
- 27 Trowsdale J, Knight JC. Major histocompatibility complex genomics and human disease. Annu Rev Genomics Hum Genet 2013; 14: 301–23.
- 28 Richeldi L, Sorrentino R, Saltini C. HLA-DPB1 glutamate 69: a genetic marker of beryllium disease. Science 1993; 262: 242–44.
- 29 Potolicchio I, Mosconi G, Forni A, Nemery B, Seghizzi P, Sorrentino R. Susceptibility to hard metal lung disease is strongly associated with the presence of glutamate 69 in HLA-DP β chain. Eur J Immunol 1997; 27: 2741–43.
- 30 Diaz G, Amicosante M, Jaraquemada D, et al. Functional analysis of HLA-DP polymorphism: a crucial role for DPβ residues 9, 11, 35, 55, 56, 69 and 84–87 in T cell allorecognition and peptide binding. Int Immunol 2003; 15: 565–76.
- 31 Fontenot AP, Torres M, Marshall WH, Newman LS, Kotzin BL. Beryllium presentation to CD4+ T cells underlies disease-susceptibility HLA-DP alleles in chronic beryllium disease. Proc Natl Acad Sci USA 2000; 97: 12717–22.
- 32 Lombardi G, Germain C, Uren J, et al. HLA-DP allele-specific T cell responses to beryllium account for DP-associated susceptibility to chronic beryllium disease. J Immunol 2001; 166: 3549–55.