



HAL
open science

Hirtellina lobelii DC. essential oil, its constituents, its combination with antimicrobial drugs and its mode of action

Madona Khoury, Marc El Beyrouth, Naïm Ouaini, Véronique Eparvier, Didier Stien

► To cite this version:

Madona Khoury, Marc El Beyrouth, Naïm Ouaini, Véronique Eparvier, Didier Stien. Hirtellina lobelii DC. essential oil, its constituents, its combination with antimicrobial drugs and its mode of action. *Fitoterapia*, 2019, 133, pp.130-136. 10.1016/j.fitote.2019.01.001 . hal-02168388

HAL Id: hal-02168388

<https://hal.sorbonne-universite.fr/hal-02168388>

Submitted on 28 Jun 2019

HAL is a multi-disciplinary open access archive for the deposit and dissemination of scientific research documents, whether they are published or not. The documents may come from teaching and research institutions in France or abroad, or from public or private research centers.

L'archive ouverte pluridisciplinaire **HAL**, est destinée au dépôt et à la diffusion de documents scientifiques de niveau recherche, publiés ou non, émanant des établissements d'enseignement et de recherche français ou étrangers, des laboratoires publics ou privés.

1 ***Hirtellina lobelii* DC. essential oil, its constituents, its combination with antimicrobial drugs**
2 **and its mode of action**

3

4 Madona Khoury,^{a,b,*} Marc El Beyrouthy,^b Naïm Ouaini,^b Véronique Eparvier,^a Didier Stien^{a,c,*}

5

6 ^a CNRS, Institut de Chimie des Substances Naturelles (ICSN), UPR2301, Université Paris-Sud, 1
7 avenue de la terrasse, 91198 Gif-sur-Yvette, France

8 ^b Department of Agricultural Sciences, Holy Spirit University of Kaslik, Kaslik, B.P. 446, Jounieh,
9 Lebanon

10 ^c Sorbonne Université, CNRS, Laboratoire de Biodiversité et Biotechnologie Microbienne,
11 USR3579, Observatoire Océanologique, 66650 Banyuls-sur-mer, France

12

13 * Corresponding authors.

14 *E-mail addresses:* madonakhoury@hotmail.com, didier.stien@cns.fr.

15

16 Declarations of interest: none

17 **Abstract**

18 With the goal of unravelling antimicrobial agents and mixtures inspired by plant defences, we
19 investigated the antibacterial and antifungal efficacy of *Hirtellina lobelii* DC. essential oil (EO),
20 both alone and in combination with antimicrobial drugs.

21 *Hirtellina lobelii* DC. EO was analysed by GC, GC-MS and partial fractionation/NMR. It was
22 essentially composed of oxygenated sesquiterpenes (75.2%), with α -bisabolol (34.5%), fokienol
23 (12.0%) and T-muurolol (6.8%) serving as the main components. Microbial susceptibility was
24 determined by the broth microdilution method and was expressed as minimum inhibitory
25 concentration (MIC) and minimum bactericidal or fungicidal concentration (MBC or MFC). This
26 EO was found to possess remarkable bactericidal (MBC/MIC = 2) and fungicidal (MFC/MIC = 1 -
27 4) potential, particularly against the Gram (+) bacteria *Staphylococcus aureus*, including its
28 methicillin-resistant forms, the yeast *Cryptococcus neoformans* and dermatophytes from the
29 genus *Trichophyton* (MICs 8 - 128 $\mu\text{g/ml}$). The examination of the combined effects of the EO
30 with antimicrobial drugs revealed synergisms of the EO with vancomycin against *S. aureus* and
31 of the EO with fluconazole and griseofulvin against dermatophytic fungi (FICI 0.2 - 0.5). The
32 effect of *H. lobelii* EO on the morphologies of fungal hyphae and bacteria, as determined by
33 scanning electronic microscopy (SEM), showed fungal hyphae swelling and bulging.

34 These results suggest that *H. lobelii* EO and its major constituent, α -bisabolol, have remarkable
35 antimicrobial potential. Combination therapies of this EO with antifungal drugs could offer a
36 promising alternative for treatment of human mycoses caused by filamentous dermatophytic
37 fungi.

38 **Keywords:** essential oil; antimicrobial activity; synergy; scanning electronic microscopy;

39 *Hirtellina lobelii*; α -bisabolol

40 **1. Introduction**

41 Antimicrobial resistance has become a major therapeutic challenge, as a variety of
42 multiresistant pathogenic microbes have emerged that defy commonly available treatments.
43 The decline in effectiveness of existing drugs is partly due to natural selection and partly to
44 their intensive use and misuse. As a matter of concern, the number of immunocompromised
45 patients, who frequently develop opportunistic systemic infections, has dramatically increased
46 since the 1990s [1]. Thus, infectious diseases are considered a real global threat, representing
47 26% of overall mortality in 2001 [2] and almost 30% in 2011 [3].

48 The development of microbial resistance to conventional treatments along with drug-related
49 toxicities and costs have generated a clear need for new therapeutic strategies, new
50 antimicrobial compounds and, particularly, new combination treatments and active mixtures.
51 Combinatorial therapies can be less vulnerable to the development of drug resistance and can
52 increase therapeutic efficacy [4,5]. Their potential synergistic effects provide broader
53 pharmacological windows and lower toxicities [6]. Thus, efforts toward discovering multi-agent
54 therapies that can overcome the limitations of monotherapies are highly encouraged [5–8].

55 In the quest for new antimicrobial drugs and bio-inspired mixtures, medicinal plants must not
56 be overlooked. The antimicrobial potential of aromatic plants has been recognized since
57 antiquity and is mainly attributed to their volatile oils, which contain wide chemical diversity
58 [9,10]. Essential oils (EOs) are known for their antiseptic, i.e., bactericidal, fungicidal and

59 virucidal, properties, and some of them have been claimed to cure microbial infections and
60 have been proposed for use in complementary medicine [11]. Thus, EOs could transition from
61 being used solely in traditional medicine to also being used in modern medicine.

62 Although many studies have focused on showing the antimicrobial activity of EOs, few have
63 investigated the origin of their bioactivities and their exact mechanisms of action by examining
64 their effects on the morphologies and ultrastructures of pathogenic strains.

65 *Hirtellina lobelii* DC., formerly known as *Staehelina lobelii* DC., is an herbaceous plant belonging
66 to the Asteraceae (or Compositae) family. The genus *Staehelina* (tribe Cardueae) is an
67 extremely small genus that consists of only a few species worldwide and has rarely been
68 documented. The ascription of this genus is highly problematic, and the subtribal placement of
69 *Staehelina* remains unresolved. Based on Dittrich [12], the two species of *Staehelina* with
70 hirsute pericarps (*S. fruticosa* L. and *S. lobelii* DC.) should be classified in a distinct genus,
71 *Hirtellina*. We followed the classification proposed by Dittrich, which is also accepted by most
72 databases such as Euro+Med PlantBase (<http://ww2.bgbm.org/EuroPlusMed>) and the plant list
73 (<http://www.theplantlist.org>).

74 *Hirtellina lobelii* grows on rock crevices in large clumps and is a locally important element of
75 Mediterranean chasmophytic vegetation. Its distribution is restricted to Lebanon, Cyprus, Syria
76 and Asiatic Turkey [13, Euro+Med PlantBase (<http://ww2.bgbm.org/EuroPlusMed>)].

77 We evaluated the chemical composition of the EO of *H. lobelii*, its antimicrobial potential alone
78 and in combination with antibiotics and antifungal drugs as well as its mechanism of action by
79 observing the morphological alterations to pathogen structures that it caused by scanning

80 electron microscopy (SEM). To the best of our knowledge, this is the first description of the
81 chemical composition and the antimicrobial potential of *Hirtellina lobelii* DC. EO. In addition, no
82 volatile organic compound has ever been described from species of the *Hirtellina* or *Staelina*
83 genera.

84

85 **2. Experimental part**

86 **2.1. Plant material and essential oil extraction**

87 The aerial part of the plant (fresh leaves and stems) was collected in June 2012 from Qartaba,
88 Mount Lebanon (34°05'58.70" N 35°48'46.39" E) at an altitude of 1250 m. A voucher specimen
89 was deposited at the Herbarium of the Department of Botany and Medicinal plants, Holy Spirit
90 University, Faculty of Agricultural and Food Sciences (USEK-Lebanon) under the registry number
91 MNV446a. Hydrodistillation of the plant was performed for 3 h using a Clevenger-type
92 apparatus according to the European Pharmacopoeia, 1997. The EO was obtained with a yield
93 of 0.1%.

94 **2.2. Essential oils analyses**

95 *2.2.1. GC analyses*

96 Analytical gas chromatography was performed using a Thermo Electron Corporation gas
97 chromatograph fitted with a DB-5 MS capillary column (30 m × 0.25 mm, 0.1 µm film thickness)
98 or a fused silica HP Innowax polyethylene glycol capillary column (50 m × 0.20 mm, 0.20 µm film
99 thickness). Helium was the carrier gas (0.7 ml/min). The column temperature was initially set to

100 35 °C and was gradually increased to 85 °C at 5 °C/min. It was held at 85 °C for 20 min and then
101 raised to 300 °C at 10 °C/min. Finally, it was held at 300 °C for 5 min. Diluted 1 µl samples
102 (1/100, vol/vol) were manually injected at 250 °C in the splitless mode. Flame ionisation
103 detection (FID) was performed at 310 °C.

104 2.2.2. GC/MS analyses

105 The GC/MS analyses were performed using an Agilent gas chromatograph 6890 coupled with a
106 Mass Detector 5975. The 7683 B autosampler injected 1 µL of each oil sample. A fused silica
107 capillary column DB-5 MS (30 m × 0.25 mm internal diameter, 0.1 µm film thickness) or a fused
108 silica HP Innowax polyethylene glycol capillary column (50 m × 0.20 mm, 0.20 µm film
109 thickness) was used. Helium was the carrier gas (0.7 ml/min). The oven temperature program
110 was identical to that described in 2.2.1. The mass spectra were recorded at 70 eV with an ion
111 source temperature of 310 °C and a transfer line heated to 320 °C. The acquisition was
112 recorded in full scan mode (50 – 400 amu).

113 2.2.3. Identifications and quantifications

114 Most constituents were identified by GC and GC/MS by comparing their retention indices (RI)
115 with those from the literature [14,15] or those of authentic compounds obtained from Sigma-
116 Aldrich (Lebanon). The retention indices were determined relative to a homologous series of *n*-
117 alkanes (C8 to C24) that had been analysed under the same operating conditions. Their mass
118 spectra using both columns were compared with those provided in the NIST and Wiley 275
119 libraries, our home-made library constructed with pure compounds and EOs of known
120 compositions or mass spectra from the literature [14,16]. The relative concentrations of the

121 components were calculated based on the GC peak areas without correction and are reported
122 in Table 1.

123 *2.2.4. Essential oil fractionation by HPLC*

124 Analytical and preparative HPLCs were conducted using a Gilson system equipped with a 322
125 pumping device, GX-271 fraction collector, 171 diode array detector, and ELSII preparative
126 electrospray nebulizer detector. Phenomenex Luna C18 columns of two sizes were used for
127 these experiments: a 4.6 × 250 mm column with 5 μm film thickness for analytical HPLC and a
128 21.2 × 250 mm column with 5 μm film thickness for preparative HPLC. The flow rates were set
129 to 1 and 21 ml/min for analytical and preparative HPLC, respectively, using a linear gradient of
130 water mixed with an increasing proportion of acetonitrile (30/70 to 0/100 over 35 min and then
131 100% CH₃CN for 24 min). The EO was diluted in acetonitrile at 10 mg/ml for analytical HPLC. The
132 EO was also diluted in acetonitrile for prep-HPLC (60 mg EO in 300 μL CH₃CN), and 250 μL of the
133 diluted solution was injected.

134 Forty-nine 25 ml fractions were collected between 2 and 59 min and were combined into 22
135 fractions according to their HPLC profiles. These 22 fractions were analysed by GC/MS. Fraction
136 23/24 contained 4 compounds, among which the unidentified component was the major one
137 and accounted for 47% of the mixture according to GC/MS integration. The volatile organic
138 compounds were collected from the fraction as follows: the fraction was separated between
139 ether (250 ml) and water (250 ml). The organic layer was washed with water (3 × 50 ml), dried
140 with Na₂SO₄, and evaporated without heating.

141 *2.2.5. NMR spectroscopy*

142 The nuclear magnetic resonance (NMR) spectra (^1H -NMR, ^{13}C -NMR, ^1H - ^1H COSY, HSQC and
143 HMBC) of fraction 23/24 allowed us to confirm the identification of one of the main EO
144 constituents (fokienol, **2**, Fig. 1) by comparison with NMR data reported in the literature
145 [17,18]. The NMR spectra were recorded using a Bruker 500 MHz spectrometer equipped with
146 a 5 mm inverse detection probe. Chemical shifts (δ) are reported as ppm based on the TMS
147 signal, with s, d, t, and br standing for singlet, doublet, triplet and broad, respectively. The NMR
148 signals of fokienol as extracted from the NMR analysis of fraction 23/24 are as follows:
149 Fokienol (**2**): ^1H NMR (500 MHz, CDCl_3) δ 1.29 (3H, s, H-15), 1.61 (3H, s, H-14), 1.56-1.62 (2H, m,
150 H-4), 1.84 (3H, s, H-12), 2.02-2.08 (2H, m, H-5), 2.76 (2H, br d, $J = 7.2$ Hz, H-8), 4.89 (2H, br s, H-
151 13), 5.19 (1H, m, H-6), 5.07 (1H, dd, $J = 11.0$; 1.2 Hz, H-1a), 5.19 (1H, m, H-6), 5.23 (1H, dd, $J =$
152 17.1; 1.2 Hz, H-1b), 5.62 (1H, dt, $J = 15.9$; 7.2 Hz, H-9), 5.93 (1H, dd, $J = 17.1$; 11.0 Hz, H-2), 6.14
153 (1H, br d, $J = 15.9$ Hz, H-10). ^{13}C -NMR (125 MHz, CDCl_3) δ 16.0 (C-14), 18.5 (C-12), 22.6 (C-5),
154 27.7 (C-15), 41.8 (C-4), 42.8 (C-8), 73.2 (C-3), 111.6 (C-1), 114.4 (C-13), 125.0 (C-6), 128.5 (C-9),
155 133.9 (C-10), 134.1(C-7), 141.9 (C-11), 145.0 (C-2).

156 **2.3. Antimicrobial activity**

157 *2.3.1. Microorganisms*

158 The antimicrobial activity of the EOs against the following microbial strains was investigated:
159 Gram (–) bacterial strain, *Escherichia coli* ATCC 25922; Gram (+) bacterial strains,
160 *Staphylococcus aureus* ATCC 29213 and methicillin-resistant *Staphylococcus aureus* (also called
161 oxacillin-resistant *Staphylococcus aureus*) ATCC 33591; yeasts, *Candida albicans* ATCC 10231,
162 *Candida parapsilosis* ATCC 22019 and *Cryptococcus neoformans* SNB-CN1; and filamentous
163 fungi, *Trichophyton rubrum* SNB-TR1, *Trichophyton violaceum* SNB-TV1, *Trichophyton*

164 *soudanense* SNB-TS1, *Trichophyton tonsurans* SNB-TT1, *Trichophyton mentagrophytes* SNB-TM1
165 and *Aspergillus fumigatus* SNB-AF1. The ATCC strains were purchased, while the other strains
166 were clinical isolates kindly provided by Prof. Philippe Loiseau, Université Paris Sud. These
167 strains were identified by Prof. Philippe Loiseau and Christian Bories; the molecular analyses
168 were conducted by BACTUP. The ITS sequences were deposited in the NCBI GenBank database
169 under the following registry numbers: KF360235 (*C. neoformans* SNB-CN1), KC692746 (*T.*
170 *rubrum* SNB-TR1), KF360236 (*T. violaceum* SNB-TV1), KF360237 (*T. soudanense* SNB-TS1),
171 KF360238 (*T. tonsurans* SNB-TT1), KF360239 (*T. mentagrophytes* SNB-TM1) and KC692747 (*A.*
172 *fumigatus* SNB-AF1).

173 2.3.2. Microdilution method

174 The broth microdilution method was used to determine the minimal inhibitory concentration
175 (MIC) of the EOs according to the Clinical and Laboratory Standards Institute guidelines [19–22].
176 The essential oil and its major components diluted in DMSO were tested at concentrations
177 ranging from 512 to 1 µg/ml. Oxacillin, vancomycin and gentamicin (16 - 0.03 µg/ml) were used
178 as reference antibiotics, while itraconazole (16 - 0.03 µg/ml) and fluconazole (64 - 0.125 µg/ml)
179 were used as positive controls for antifungal activity. These antimicrobial standard drugs were
180 purchased from Molekula, and pure terpene was purchased from Sigma-Aldrich. The
181 microplates were incubated at 37 °C for 24 h for bacteria, 48 h for yeasts and *A. fumigatus*, and
182 five days for the other filamentous fungi. The MIC values corresponding to the lowest
183 concentration that prevented visible microbial growth are reported in Table 2.

184 2.3.3. Minimum bactericidal and fungicidal concentrations (MBCs and MFCs)

185 After MIC determination, the bactericidal and fungicidal activities of the EO were determined as
186 previously described [23–25]. From each well with no detected microbial growth, 20 µl of the
187 culture medium of bacterial and fungal cultures were subcultured on Mueller-Hinton and
188 Sabouraud dextrose agar plates, respectively; the contents of the wells were not agitated prior
189 to removal of the specified volume. The growth control wells of the microdilution plate were
190 used as the growth control, and oxacillin, itraconazole and fluconazole were used as positive
191 controls. The plates were incubated at 35 °C until growth was observed in the growth control
192 subculture (24 h for bacteria, 48 h for yeasts and 5 days for dermatophytic fungi). The MBC and
193 MFC was defined as the lowest concentration that resulted in no visible bacterial or fungal
194 growth on agar plates, respectively, and those values are reported in Table 2.

195 2.3.4. Synergy test by Microdilution Checkerboard assay

196 This test is based on the microdilution method, and the protocol was described by Shin and Lim
197 [26] and Houël et al. [27]. EO activity was tested in combination with the following drugs
198 against the following microorganisms: vancomycin, *S. aureus*; fluconazole, *C. neoformans*; and
199 fluconazole and griseofulvin, dermatophytic fungi *Trichophyton* sp. The drug solutions were
200 diluted along the horizontal orientation so that the plates contained final concentrations of
201 vancomycin and griseofulvin ranging from 16 to 0.03 µg/ml and final concentrations of
202 fluconazole ranging from 64 to 0.125 µg/ml. The EO solution was diluted along the vertical
203 orientation so that the final concentrations ranged from 64 to 1 µg/ml for bacteria and
204 dermatophytes and from 256 to 4 µg/ml for yeast.

205 Fractional inhibitory concentrations (FIC), which represent the gain in activity of individual
206 components of the mixture, were determined for each point by dividing the MIC of the

207 combinations of the two products by the MIC of the essential oil or the drug alone. The FIC
208 index (FICI), which is a measure of synergy, was obtained by adding both FICs. The FICI was
209 interpreted as follows: values ≤ 0.5 indicated a synergistic effect, values > 0.5 and ≤ 2.0
210 indicated an indifferent effect, and values > 2.0 indicated an antagonistic effect [26,28].
211 The results were also analysed by the isobologram method (see supplementary material, S1).

212 **2.4. Cytotoxicity Assays**

213 Cytotoxicity assays were conducted with MRC5 (human foetal lung fibroblast) and MDA435
214 (melanoma) cell lines according to the procedure described by Rochais et al. [29].

215 **2.5. Scanning electron microscopy observations**

216 To examine the effect of *H. lobelii* EO on the surface topography of *S. aureus*, *T. rubrum* and *T.*
217 *soudanense*, samples for SEM observation were prepared as follows. For bacteria, a
218 conventional broth microdilution assay was performed in a 96-well microtitre plate; then, 40
219 μL of the suspension was sampled and deposited on sterilized glass squares distributed in a 24-
220 well polystyrene plate. Sedimentation lasted three hours. Fungi were directly grown on the
221 same system.

222 Samples treated with a sub-inhibitory concentration were fixed with 2.5% glutaraldehyde in 0.1
223 M sodium cacodylate buffer at pH 7.4 for 1 hour at RT. The untreated control microorganisms
224 were processed in parallel for the SEM analyses. After three washes of 10 min with the same
225 buffer, the samples were post-fixed with 1% osmium tetroxide in 0.1 M sodium cacodylate
226 buffer at pH 7.4 for 1 hour at RT and then washed in sterile water three times for 10 min. The
227 cells were dehydrated with increasing concentrations of ethanol (50%, 70%, 90%, 3x100%) at

228 RT for 10 min for each bath. The samples were critical-point dried at 75 bar and 37 °C with
229 liquid CO₂ as the transition fluid and were then depressurized slowly (400 cm³/min) in a
230 Quorum Technologies K850 device (Elexience, France). Then, the samples were sputter-
231 coated in Argon plasma with Platinum (thickness ≈ 30 nm) in a Polaron SC7640 device
232 (Elexience, France) at 10 mA and 0.8 kV for 200 s. Observations were performed using a FE-SEM
233 Hitachi S4500 (Hitachi, Japan) in a high vacuum with a sample holder tilted at 45° and a low SE
234 detector at 2 kV and 21 mm WD. The experiments were performed using the MIMA2
235 microscopy platform (<http://www6.jouy.inra.fr/mima2>).

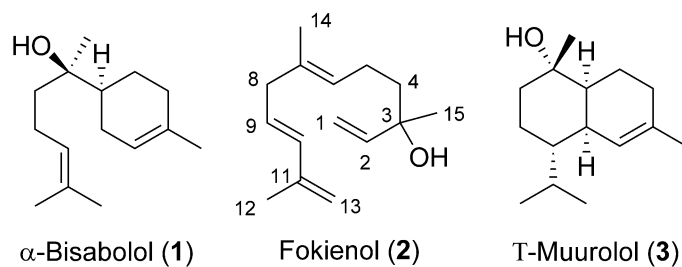
236 **3. Results and Discussion**

237 **3.1. Essential oils analyses**

238 The essential oil of the fresh aerial parts of *Hirtellina lobelii* was obtained by hydrodistillation
239 with a yield of 0.1% (vol/wt, relative to dry weight material). The chemical composition of the
240 EO and the relative proportions of the components are reported in Table 1.

241 GC, GC-MS, and fractionation/NMR led to the identification of 25 components representing
242 86.9% of the EO. The EO was essentially composed of oxygenated sesquiterpenes (75.2%), with
243 α-bisabolol (**1**, fig. 1; 34.5%), fokienol (**2**, fig. 1; 12.0%) and T-muurolol (**3**, fig. 1; 6.8%) as the
244 main components. We used NMR to identify the second major component, fokienol, which
245 could not be identified by GC/MS, or by comparison with Kovats Index (KI) and MS databases.

246



248 **Fig. 1.** Major compounds identified in the *H. lobelii* EO

249

250 **Table 1.** Composition of the essential oil of *Hirtellina lobeli*

251

R_i^a	R_i^b	Identification ^c	Compound ID	Leaves EO
1013	1188	R_i , MS, CoGC	α -Phellandrene	t
1030	1203	R_i , MS, CoGC	Limonene	t
1098	1553	R_i , MS, CoGC	Linalool	0.1
1115	1584	R_i , MS	β -Fenchyl alcohol	0.1
1352	1466	R_i , MS	α -Cubebene	0.1
1377	1497	R_i , MS	α -Copaene	0.4
1382	1838	R_i , MS	(E)- β -Damascenone	0.2
1415	1612	R_i , MS, CoGC	β -Caryophyllene	0.7
1455	1689	R_i , MS	α -Humulene	0.3
1483	1784	R_i , MS	α -Curcumene	1.7
1500	1740	R_i , MS	α -Muurolene	1.3
1515	1716	R_i , MS, CoGC	γ -Cadinene	3.0
1526	1773	R_i , MS	δ -Cadinene	3.2
1566	2050	R_i , MS	Nerolidol	4.0
1577	2008	R_i , MS, CoGC	Caryophyllene oxide	2.6
1585	2182	NMR	Fokienol	12.0
1625	2088	R_i , MS	Epi-cubanol	2.4
1640	2188	R_i , MS	T-Cadinol	3.3
1642	2209	R_i , MS	T-Muurolol	6.8
1649	2256	R_i , MS	α -Cadinol	4.3
1669	2229	R_i , MS, CoGC	α -Bisabolol	34.5
1702	2323	R_i , MS	8-Cedren-13-ol	1.2
1758	2355	R_i , MS	Nuciferol	4.1
1750	2655	R_i , MS	Benzyl benzoate	0.3
2118	2603	R_i , MS	Phytol	0.2
			Monoterpene hydrocarbons	0.1
			Oxygenated monoterpenes	0.2
			Sesquiterpenes hydrocarbons	10.9
			Oxygenated sesquiterpenes	75.2
			Other	0.5
			Total identified	86.9

252 Notes: t = trace, less than 0.05%. ^a Retention index on a HP-5MS column; ^b Retention index on an

253 Innovax column; ^c R_i Retention index identical to bibliography.

254 MS: identification based on comparison of mass spectra. Co-GC: retention time identical to authentic
255 compounds; NMR: comparison of NMR spectra with those reported in the literature.

256

257 **3.2. Antimicrobial activity**

258 The minimum inhibitory concentrations (MICs) of *H. lobelii* EO and its main compound, α -
259 bisabolol, as well as the minimum bactericidal and fungicidal concentrations (MBCs and MFCs)
260 of the oil are reported in Table 2. The oil was considered active if the minimal inhibitory
261 concentration was 128 $\mu\text{g/ml}$ or below [30]. Most of the tested pathogens were sensitive to *H.*
262 *lobelii* EO. The only resistant strains were the bacterium *E. coli*, the *Candida* yeasts, and the
263 clinical isolate *A. fumigatus*. The EO was very active against the five dermatophytic
264 *Trichophyton* species, *T. rubrum*, *T. mentagrophyte*, *T. violaceum*, *T. soudanense* and *T.*
265 *tonsurans* (MIC values ranging from 8 to 64 $\mu\text{g/ml}$) and against the Gram (+) bacterium *S.*
266 *aureus* with a MIC value of 32 $\mu\text{g/ml}$. It was also moderately active against the encapsulated
267 yeast *C. neoformans* with a MIC value of 128 $\mu\text{g/ml}$. Interestingly, the growth of methicillin-
268 resistant *S. aureus* (MRSA) was also inhibited by the EO, with a MIC value of 128 $\mu\text{g/ml}$.
269 Although this value is higher than that for the non-resistant *S. aureus*, this result supports the
270 use of combination therapies for MRSA infections that include *H. lobelii* EO.

271

272 **Table 2.** Antimicrobial activity (MIC, MBC or MFC in $\mu\text{g/ml}$) of *Hirtellina lobelii* essential oil and its major
273 compound.

Compounds

		<i>H. lobelii</i> EO	α -bisabolol	Oxacillin	Vancomycin	Gentamicin	Itraconazole	Fluconazole	
Bacteria	<i>S. aureus</i> ATCC 29213	MIC	32	32	0.5	1	-	-	
		MBC	64	-	0.5	-	-	-	
		MBC/MIC	2	-	1	-	-	-	
	MRSA ATCC 33591	MIC	128	-	>16	-	-	-	
	<i>E. coli</i> ATCC 25922	MIC	> 512	-	-	8	-	-	
Yeasts	<i>C. albicans</i> ATCC 10231	MIC	512	-	-	-	4	16	
	<i>C. parapsilosis</i> ATCC 22019	MIC	512	-	-	-	0.5	2	
	<i>C. neoformans</i> SNB-CN1	MIC	128	64	-	-	1	8	
Filamentous fungi	<i>A. fumigatus</i> SNB-AF1	MIC	>512	-	-	-	0.5	>512	
	<i>T. rubrum</i> SNB-TR1	MIC	64	32	-	-	-	<0.03	2
		MFC	64	-	-	-	-	4	>64
		MFC/MIC	1	-	-	-	-	>128	>32
	<i>T. mentagrophytes</i> SNB-TM1	MIC	32	32	-	-	-	0.125	64
		MFC	32	-	-	-	-	16	>64
		MFC/MIC	1	-	-	-	-	128	>1
	<i>T. violaceum</i> SNB-TV1	MIC	16	32	-	-	-	<0.03	4
		MFC	16	-	-	-	-	0.5	64
		MFC/MIC	1	-	-	-	-	>16	16
	<i>T. soudanense</i> SNB-TS1	MIC	16	16	-	-	-	<0.03	4
		MFC	32	-	-	-	-	4	64
		MFC/MIC	2	-	-	-	-	>128	16
	<i>T. tonsurans</i> SNB-TT1	MIC	8	8	-	-	-	0.25	16
		MFC	32	-	-	-	-	1	64
MFC/MIC		4	-	-	-	-	4	4	

274

275 Next, we investigated the source of the antimicrobial potential of this EO. The major
276 constituent of *H. lobelii* oil, α -bisabolol, represented 34.5% of the total oil, and thus, it was
277 tested on the microorganisms that showed the greatest susceptibility to the EO. The activity of
278 α -bisabolol was roughly equivalent to that of the crude oil, with MIC values ranging from 8 to
279 64 μ g/ml (Table 2). This result suggests a crucial role of α -bisabolol in the observed
280 antimicrobial activity of this EO.

281 In the context of this study, it was interesting to discern whether the EO possesses bactericidal
282 and fungicidal properties capable of destroying bacterial and fungal cells or simple growth
283 inhibition effects (bacteriostatic and fungistatic activities). The bactericidal and fungicidal
284 activities of the oil were evaluated on the most sensitive pathogens (MIC < 128 µg/ml) and
285 were compared to those of the positive control drugs, oxacillin, itraconazole and fluconazole
286 (Table 2). The MBC/MIC and MFC/MIC ratios were calculated for each microorganism;
287 compounds are considered bactericidal or fungicidal when the MBC/MIC or MFC/MIC ratio is ≤
288 4 [31]. *H. lobelii* EO was found to be bactericidal against *S. aureus* (MBC/MIC = 2) and fungicidal
289 against all tested *Trichophyton* spp. (MFC/MIC = 1 to 4). Additionally, the EO was more effective
290 than the positive control antifungal drugs (MFC/MIC = 4 to > 128). Even itraconazole, which
291 inhibited the growth of the dermatophytic fungi at much lower concentrations (MIC 0.25 to <
292 0.03 µg/ml), was essentially fungistatic. The development of fungicidal therapies is crucial
293 because the prophylactic use of fungistatic drugs has been shown to be associated with an
294 increased frequency of acquired drug resistance in clinical isolates [32]; thus, these results add
295 more value to the *H. lobelii* EO.

296 **3.3. Combined effects of *H. lobelii* oil and antimicrobial drugs**

297 Based on the above results, it was relevant to evaluate the combined antimicrobial effects of *H.*
298 *lobelii* EO with various antimicrobial drugs. Synergies between EOs and antimicrobial drugs can
299 shorten the duration of therapies, decelerate the emergence of drug resistance and reduce the
300 possible side effects of current therapies by decreasing the necessary doses of the current
301 drugs and EO. The results of the checkerboard assay (FIC and FICI) are reported in Table 3, and

302 the isobolograms are presented in Fig. S1 (Supplementary Material). Most of the tested
 303 combinations of the oil and the drugs showed synergistic activity.

304

305 **Table 3.** Combined effects of *Hirtellina lobelii* essential oil and antimicrobial drugs

Path	Combination	EO			Drug				MRC5		MDA435		
		MIC _a	MIC _c	FIC	MIC _a	MIC _c	FIC	FICI	MIC _M	IC ₅₀	SI	IC ₅₀	SI
<i>S. a</i> ^a	EO:Vancomycin (1:0.5)	32	1	0.03	1	0.5	0.5	0.5	1.5	8	5.3	9	6
<i>C. n</i> ^b	EO:Fluconazole (1:0.125)	128	64	0.5	8	8	1	1.5					
	EO:Itraconazole(1:0.016)	128	64	0.5	1	1	1	1.5					
<i>T. r</i> ^c	EO:Fluconazole (1:0.5)	64	1	0.01	2	0.5	0.25	0.3					
	EO:Griseofulvin* (1:0.25)	64	1	0.01	1	0.25	0.25	0.3	1.25	7	5.6	8.5	6.8
<i>T. m</i> ^c	EO:Fluconazole (1:8)	32	2	0.06	64	16	0.25	0.3					
	EO:Griseofulvin* (1:0.25)	32	1	0.03	0.5	0.25	0.5	0.5	1.25	7	5.6	8.5	6.8
<i>T. v</i> ^c	EO:Fluconazole (1:0.5)	16	1	0.06	4	0.5	0.13	0.2					
	EO:Griseofulvin* (1:0.03)	16	4	0.25	0.5	0.13	0.25	0.5	4.125	7.5	1.8	8.5	2.1
<i>T. s</i> ^c	EO:Fluconazole (1:0.5)	16	2	0.125	4	1	0.25	0.4					
	EO:Griseofulvin*(1:0.125)	16	1	0.06	0.5	0.13	0.25	0.3	1.125	7.2	6.4	8.5	7.5
<i>T. t</i> ^c	EO:Fluconazole (1:8)	8	1	0.125	16	8	0.5	0.6					
	EO:Griseofulvin*(1:0.125)	8	2	0.25	1	0.25	0.25	0.5	2.25	7.2	3.2	8.5	3.8

306 ^aBacteria: *S.a.* = *Staphylococcus aureus* (ATCC 29213), ^bYeast: *C.n.* = *Cryptococcus neoformans* (SNB-CN1),

307 ^cFilamentous fungi: *T.r.* = *Trichophyton rubrum* (SNB-TR1), *T.m.* = *Trichophyton mentagrophytes* (SNB-TM1), *T.v.* =

308 *Trichophyton violaceum* (SNB-TV1), *T.s.* = *Trichophyton soudanense* (SNB-TS1), *T.t.* = *Trichophyton tonsurans* (SNB-

309 TT1).

310 MIC_a: MIC of the product alone (in µg/ml); MIC_c: MIC of the drug or the essential oil representing the highest

311 synergy or antagonism (in µg/ml); MIC_M: MIC of the mixture (MIC_c EO + MIC_c drug); FIC: fractional inhibitory

312 concentration; FICI: FIC index; SI: selectivity index (SI = IC₅₀/MIC).

313 **H. lobelii* EO: itraconazole combinations could not be tested on dermatophytic fungi because itraconazole has

314 very low MIC values. Instead, *H. lobelii* EO: griseofulvin combinations were tested, given that griseofulvin is

315 frequently prescribed for dermatophytosis.

316

317 We observed a synergistic interaction between vancomycin and *H. lobelii* EO against *S. aureus*
318 (FICI = 0.5). For the yeast *C. neoformans*, an additive effect was only detected for the
319 combination of EO and azole drugs (FICI = 1.5). However, the most interesting results of the
320 combination treatments were observed for the dermatophytic fungi. *H. lobelii* EO was
321 synergistic with fluconazole and griseofulvin against all tested *Trichophyton* spp. (FICI values
322 from 0.2 to 0.5), with one exception of an indifferent interaction between the EO and
323 fluconazole against *T. tonsurans* (FICI = 0.6). The greatest synergism was observed for the
324 combination treatment of EO with fluconazole against *T. violaceum*, with a FICI value of 0.2.
325 Overall, the combinations tested in this study would allow for a notable decrease in the
326 necessary concentrations of commercial drugs due to a 2- to 8-fold decrease in MIC values.
327 Likewise, the oil concentration required to inhibit the growth of the pathogens was reduced to
328 very small amounts (MIC_C 1 – 2 µg/ml), thus, limiting the potential cytotoxic effect of the EO
329 (Table 3, see also Fig. S1). Indeed, the selectivity indexes (SI) of the synergetic mixtures at the
330 FICI values showed that the antimicrobial activity did not exceed the cytotoxicity (with the
331 exception of *T. violaceum*). For *S. aureus*, *T. rubrum*, *T. mentagrophytes* and *T. soudanense*, the
332 selectivity indexes of the combination treatments ranged from 5.3 to 7.5, indicating relatively
333 low cytotoxicities. The selectivity indexes were calculated based on the cytotoxicity
334 measurements of the drugs in the skin cancer cell line MDA435, which is a relevant cell line in
335 the context of the topical application of antimicrobial agents.

336 The mechanism by which some EOs have synergistic interactions with antimicrobial drugs is not
337 yet clear. It has been postulated that some terpenes may act as solvents for the antimicrobial
338 drug, facilitating its passage across cell membranes [33]. However, *H. lobelii* EO is strongly

339 antimicrobial on its own. This may indicate a specific mode of action and could explain why
340 combination treatments with this EO are so effective.

341

342 **3.4. Scanning electron microscopy analysis of in vitro effects of *H. lobelii* EO**

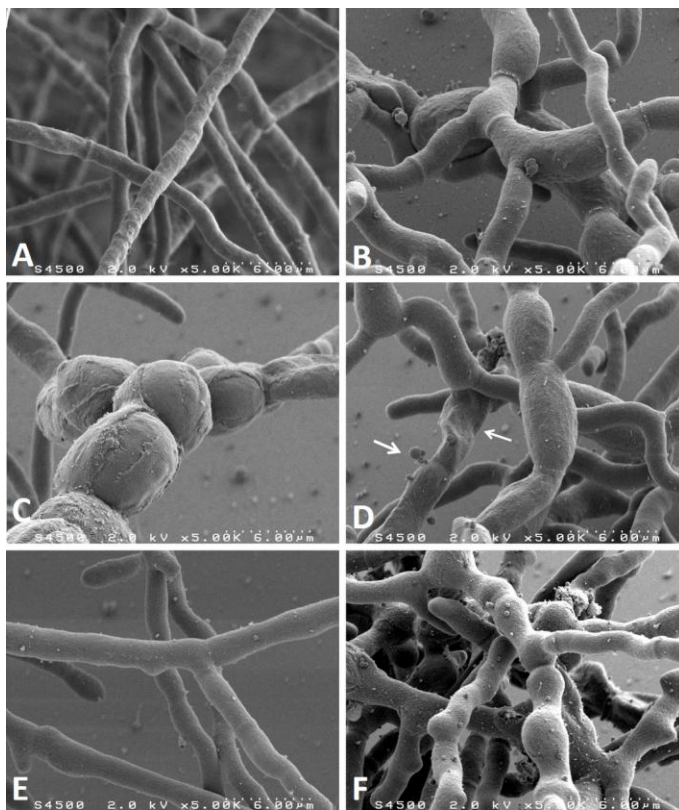
343 Changes in morphology of bacteria and filamentous fungi caused by antimicrobial compounds
344 have often provided insight into their mechanism of action [34]. To investigate the effect of *H.*
345 *lobelii* EO on the morphological characteristics of *S. aureus*, *T. rubrum* and *T. soudanense*,
346 bacterial and fungal samples treated with sub-inhibitory concentrations (sub-MICs) of the oil
347 were observed by SEM. At sub-MICs, microbial growth is not severely affected while
348 morphological alterations can sometimes be detected [35,36].

349 The morphological characteristics of *S. aureus* bacteria were not affected by EO treatment. No
350 morphological changes were detectable by SEM (data not shown).

351 Untreated *T. rubrum* and *T. soudanense* showed typical structures of healthy hyphal elements,
352 including rod-shaped filaments of uniform width with lines of separation (septa) and a smooth
353 surface. Hyphae showed regular branching (fig. 2a, e). In contrast, an unusual pattern of hyphal
354 growth, including alterations in cell shape and size were evident in both *T. rubrum* and *T.*
355 *soudanense* hyphae in response to the EO (fig. 2b, c, d, f). The treated hyphae showed
356 aberrant morphologies, such as a loss of linearity with bulging, swollen cells and anomalous
357 branching. A portion of the *T. rubrum* mycelia seemed to be particularly inflated with a rough
358 surface, probably because they were covered by extruded cell material (fig. 2c). Interestingly,
359 fig. 2d shows a flattened and shrivelled hyphal element with partial distortion and a few small
360 vesicles on the surface. At this time point, hyphal or cytoplasmic debris were dispersed on the

361 surface of the hyphae. These morphological alterations have been previously observed in
362 dermatophyte hyphae treated with terbinafine [37]. These different morphological
363 characteristics could give us a glance on the sequence of events during the exposure of fungi to
364 the EO, starting with cell swelling and cellular leakage, leading to the breakage and collapse of
365 hyphal cells.

366



367

368 **Fig. 2.** Scanning electron micrographs of dermatophytic hyphae (5000x magnification)
369 (A) Untreated control *T. rubrum* hyphae. (B), (C) and (D) *T. rubrum* hyphae treated with 32 µg/ml *H. lobelii* EO; (B)
370 showing swollen cells and anomalous branching, (C) showing highly inflated cells with a rough surface, (D) arrow
371 indicates flattened and shriveled hyphae with vesicles on the surface. (E) Untreated control of *T. soudanense*
372 hyphae. (F) *T. soudanense* hyphae treated with 8 µg/ml *H. lobelii* EO, showing swollen cells and anomalous
373 branching.

374

375 In previous studies, similar cell damaging effects were also observed in hyphae exposed to
376 terpenes or plant extracts [38,39]. It has been postulated that terpenoids interfere with the
377 phospholipid bilayers of membranes and increase their permeability [40]. Our SEM
378 observations validate the disruption of the fungal membrane in response to *H. lobelii* EO.
379 Interestingly, our results with *H. lobelii* EO do not necessarily correlates with those obtained
380 with closely related antifungal sesquiterpenes. For example, nerolidol did not induced
381 membrane swelling in *Trichophyton mentagrophytes* [41]. At high concentration (0.11 mg/mL
382 and above), internal vesicles appeared in the cells, ultimately leading to membrane breakdown
383 and abnormal mitochondria structure.
384 Our results are in agreement with the observed synergetic combinations because alteration in
385 membrane permeability by the EO would facilitate the infiltration of the cell cytoplasm by more
386 hydrophilic molecules, including a wide range of antibiotics and antifungal drugs. However, in
387 this case, *H. lobelii* EO is active at low concentration, lower than that of many terpenes
388 described in the literature. This might indicate an additional specific interaction of EO
389 constituents against cell membrane components, most probably α -bisabolol which was active
390 on its own.

391 **4. Conclusions**

392 Our data demonstrated that *Hirtellina lobelii* EO and its major component, α -bisabolol, have
393 potent antimicrobial activities, as manifested by drastic morphological changes, particularly in
394 dermatophytic fungal cells. Although the mechanism of action of this EO and α -bisabolol

395 against *S. aureus* remains unresolved, SEM observation on EO-treated fungi revealed alteration
396 of cell membrane permeability, presumably associated with a specific antifungal potential
397 linked to the presence of α -bisabolol. Altogether, these results demonstrate that both *H. lobelii*
398 EO and α -bisabolol could be used in antifungal mixtures of either the EO or the terpene in
399 combination with antifungal drugs, such as fluconazole and griseofulvin. These combinations
400 should improve the efficiency of the drugs in local applications, in particular, when the drugs
401 alone are essentially fungistatic. Combination treatments will also reduce the amount of drug
402 used and restrict the use of newly developed drugs to serious clinical cases, therefore, slowing
403 the development of chemoresistant strains. Additional combinations of this EO with other
404 antifungal drugs are worth testing.

405

406 **Acknowledgments**

407 This work was supported by an "Investissement d'Avenir" grant managed by the Agence
408 Nationale de la Recherche (CEBA, ref. ANR-10-LABX-25-01).

409 The authors gratefully acknowledge Pr. Soizic Pardo (MNHN) for fruitful discussions and Alexis
410 Acanette (MIMA2 microscopy platform) for his technical support and expertise in electron
411 microscopy.

412 **References**

- 413 [1] P.K. Linden, Approach to the immunocompromised host with infection in the intensive care unit,
414 *Infect. Dis. Clin. North Am.* 23 (2009) 535–556. doi:10.1016/j.idc.2009.04.014.
- 415 [2] K. Becker, Y. Hu, N. Biller-Andorno, Infectious diseases - a global challenge, *Int. J. Med. Microbiol.*
416 296 (2006) 179–185. doi:10.1016/j.ijmm.2005.12.015.
- 417 [3] World Health Organization, *World health statistics 2013*, 2013.
- 418 [4] A.H. Groll, T.J. Walsh, Antifungal chemotherapy: Advances and perspectives, *Swiss Med. Wkly.*
419 132 (2002) 303–311. doi:2002/23/smw-09729.
- 420 [5] G.R. Zimmermann, J. Lehár, C.T. Keith, Multi-target therapeutics: when the whole is greater than
421 the sum of the parts, *Drug Discov. Today.* 12 (2007) 34–42. doi:10.1016/j.drudis.2006.11.008.
- 422 [6] J.B. Fitzgerald, B. Schoeberl, U.B. Nielsen, P.K. Sorger, Systems biology and combination therapy
423 in the quest for clinical efficacy, *Nat. Chem. Biol.* 2 (2006) 458–466. doi:10.1038/nchembio817.
- 424 [7] A.A. Borisy, P.J. Elliott, N.W. Hurst, M.S. Lee, J. Lehar, E.R. Price, G. Serbedzija, G.R. Zimmermann,
425 M.A. Foley, B.R. Stockwell, C.T. Keith, Systematic discovery of multicomponent therapeutics,
426 *Proc. Natl. Acad. Sci.* 100 (2003) 7977–7982. doi:10.1073/pnas.1337088100.
- 427 [8] C. Onyewu, J. Heitman, Unique applications of novel antifungal drug combinations, *Antiinfect.*
428 *Agents Med. Chem.* 6 (2007) 3–15. doi:10.2174/187152107779314142.
- 429 [9] F. Bakkali, S. Averbeck, D. Averbeck, M. Idaomar, Biological effects of essential oils – a review,
430 *Food Chem. Toxicol.* 46 (2008) 446–475. doi:10.1016/J.FCT.2007.09.106.
- 431 [10] A.E. Edris, Pharmaceutical and therapeutic potentials of essential oils and their individual volatile
432 constituents: a review, *Phyther. Res.* 21 (2007) 308–323. doi:10.1002/ptr.2072.
- 433 [11] S. Stea, A. Beraudi, D. De Pasquale, Essential oils for complementary treatment of surgical
434 patients: state of the art, *Evidence-Based Complement. Altern. Med.* 2014 (2014) 726341.
435 doi:10.1155/2014/726341.

- 436 [12] M. Dittrich, Die bedeutung morphologischer und anatomischer achänen-merkmale für die
437 systematik der tribus Echinopeae Cass. und Carlineae Cass., Boissiera. 51 (1996) 9–102.
438 <https://baselbern.swissbib.ch/Record/27247035X>.
- 439 [13] P. Mouterde, Nouvelle flore du Liban et de la Syrie ; Tome troisième, Librairie Orientale,
440 Beyrouth, 1983. <https://www.frantiq.fr/fr/opac/26605/show?searchResultNb=41> (accessed
441 November 26, 2018).
- 442 [14] W.G. Jennings, Qualitative analysis of flavor and fragrance volatiles by glass capillary gas
443 chromatography, Academic Press, New York, 1980.
- 444 [15] N.W. Davies, Gas chromatographic retention indices of monoterpenes and sesquiterpenes on
445 methyl silicon and Carbowax 20M phases, J. Chromatogr. A. 503 (1990) 1–24. doi:10.1016/S0021-
446 9673(01)81487-4.
- 447 [16] R.P. Adams, Identification of essential components by gas chromatography/mass spectroscopy,
448 Allured Pub Corp, Carol Stream, IL, 2009.
- 449 [17] J. De Pascual-T, S. Vicente, M.S. González, I.S. Bellido, Nerolidol-5,8-oxides from the essential oil
450 of *santolina oblongifolia*, Phytochemistry. 22 (1983) 2235–2238. doi:10.1016/S0031-
451 9422(00)80154-5.
- 452 [18] A. Stoessl, J.B. Stothers, E.W.B. Ward, The structures of some stress metabolites from *Solanum*
453 *melongena*, Can. J. Chem. 53 (1975) 3351–3358.
- 454 [19] CLSI, Methods for dilution antimicrobial susceptibility tests for bacteria that grow aerobically;
455 Approved standard-ninth edition. CLSI document M07-A9, Clinical and Laboratory Standards
456 Institute, Wayne, PA, 2012.
- 457 [20] CLSI, Reference method for broth dilution antifungal susceptibility testing of yeasts; third
458 informational supplement. CLSI document M27-S3, Clinical and Laboratory Standards Institute,
459 Wayne, PA, 2008.

- 460 [21] CLSI, Reference method for broth dilution antifungal susceptibility testing of filamentous fungi;
461 approved standard—second edition. CLSI document M38-A2, Clinical and Laboratory Standards
462 Institute, Wayne, PA, 2008.
- 463 [22] A.M.S. Rodrigues, P.N.E.T. Theodoro, V. Eparvier, C. Basset, M.R.R. Silva, J. Beauchêne, L.S.
464 Espíndola, D. Stien, Search for antifungal compounds from the wood of durable tropical trees, J.
465 Nat. Prod. 73 (2010) 1706–1707. doi:10.1021/np1001412.
- 466 [23] CLSI, Methods for Determining Bactericidal Activity of Antimicrobial Agents; Approved Guideline
467 Volume 19 Number 18 Methods for Determining Bactericidal Activity of Antimicrobial Agents;
468 Approved Guideline. CLSI document M26-A, Clinical and Laboratory Standards Institute, Wayne,
469 PA, 1999.
- 470 [24] A. Espinel-Ingroff, A. Fothergill, J. Peter, M.G. Rinaldi, T.J. Walsh, Testing conditions for
471 determination of minimum fungicidal concentrations of new and established antifungal agents
472 for *Aspergillus* spp.: NCCLS collaborative study, J. Clin. Microbiol. 40 (2002) 3204–3208.
473 doi:10.1128/JCM.40.9.3204-3208.2002.
- 474 [25] M.R. Prado, E.H.S. Brito, R.S.N. Brilhante, R.A. Cordeiro, J.J.G. Leite, J.J.C. Sidrim, M.F.G. Rocha,
475 Subculture on potato dextrose agar as a complement to the broth microdilution assay for
476 *Malassezia pachydermatis*, J. Microbiol. Methods. 75 (2008) 341–343.
477 doi:10.1016/J.MIMET.2008.05.022.
- 478 [26] S. Shin, S. Lim, Antifungal effects of herbal essential oils alone and in combination with
479 ketoconazole against *Trichophyton* spp., J. Appl. Microbiol. 97 (2004) 1289–1296.
480 doi:10.1111/j.1365-2672.2004.02417.x.
- 481 [27] E. Houël, A.M.S. Rodrigues, A. Jahn-Oyac, J.-M. Bessière, V. Eparvier, E. Deharo, D. Stien, In vitro
482 antidermatophytic activity of *Otacanthus azureus* (Linden) Ronse essential oil alone and in
483 combination with azoles, J. Appl. Microbiol. 116 (2014) 288–294. doi:10.1111/jam.12377.

- 484 [28] M.-S. Pyun, S. Shin, Antifungal effects of the volatile oils from *Allium* plants against *Trichophyton*
485 species and synergism of the oils with ketoconazole, *Phytomedicine*. 13 (2006) 394–400.
486 doi:10.1016/J.PHYMED.2005.03.011.
- 487 [29] C. Rochais, T. Cresteil, V. Perri, M. Jouanne, A. Lesnard, S. Rault, P. Dallemagne, MR22388, a
488 novel anti-cancer agent with a strong FLT-3 ITD kinase affinity, *Cancer Lett.* 331 (2013) 92–98.
489 doi:10.1016/J.CANLET.2012.12.017.
- 490 [30] P. Cos, A.J. Vlietinck, D. Vanden Berghe, L. Maes, Anti-infective potential of natural products: how
491 to develop a stronger in vitro ‘proof-of-concept,’ *J. Ethnopharmacol.* 106 (2006) 290–302.
492 doi:10.1016/J.JEP.2006.04.003.
- 493 [31] J. Meletiadis, C. Antachopoulos, T. Stergiopoulou, S. Pournaras, E. Roilides, T.J. Walsh, Differential
494 fungicidal activities of amphotericin B and voriconazole against *Aspergillus* species determined by
495 microbroth methodology, *Antimicrob. Agents Chemother.* 51 (2007) 3329–3337.
496 doi:10.1128/AAC.00345-07.
- 497 [32] B.C. Monk, A. Goffeau, Outwitting multidrug resistance to antifungals, *Science*. 321 (2008) 367–
498 369. doi:10.1126/science.1159746.
- 499 [33] J. Gershenzon, N. Dudareva, The function of terpene natural products in the natural world, *Nat.*
500 *Chem. Biol.* 3 (2007) 408–414. doi:10.1038/nchembio.2007.5.
- 501 [34] S. Gunji, K. Arima, T. Beppu, Screening of antifungal antibiotics according to activities inducing
502 morphological abnormalities, *Agric. Biol. Chem.* 47 (1983) 2061–2069.
503 doi:10.1080/00021369.1983.10865911.
- 504 [35] V. Lorian, Antibiotiques à concentrations sudinhibitrices. Effet sur la morphologie et la
505 croissance, *Pathol. Biol.* 25 (1977) 291–298.
- 506 [36] D.J. Frost, K.D. Brandt, D. Cugier, R. Goldman, A whole-cell *Candida albicans* assay for the
507 detection of inhibitors towards fungal cell wall synthesis and assembly, *J. Antibiot.* 48 (1995)

508 306–310. doi:10.7164/antibiotics.48.306.

509 [37] M. Ghannoum, N. Isham, W. Henry, H.-A. Kroon, S. Yurdakul, Evaluation of the morphological
510 effects of TDT 067 (terbinafine in Transfersome) and conventional terbinafine on dermatophyte
511 hyphae *in vitro* and *in vivo*, *Antimicrob. Agents Chemother.* 56 (2012) 2530–2534.
512 doi:10.1128/AAC.05998-11.

513 [38] A. Escalante, M. Gattuso, P. Pérez, S. Zacchino, Evidence for the mechanism of action of the
514 antifungal phytolaccoside B isolated from *Phytolacca tetramera* Hauman, *J. Nat. Prod.* 71 (2008)
515 1720–1725. doi:10.1021/np070660i.

516 [39] C. Romagnoli, R. Bruni, E. Andreotti, M.K. Rai, C.B. Vicentini, D. Mares, Chemical characterization
517 and antifungal activity of essential oil of capitula from wild Indian *Tagetes patula* L., *Protoplasma.*
518 225 (2005) 57–65. doi:10.1007/s00709-005-0084-8.

519 [40] R. Vila, B. Freixa, S. Cañigüeral, Antifungal compounds from plants, in: D. Muñoz-Torrero, A.
520 Cortés, E.L. Mariño (Eds.), *Recent Adv. Pharm. Sci. III*, Transworld Research Network, Trivandrum,
521 India, 2013: pp. 23–43.

522 [41] M.J. Park, K.S. Gwak, I. Yang, K.W. Kim, E.B. Jeung, J.W. Chang, I.G. Choi, Effect of citral, eugenol,
523 nerolidol and α -terpineol on the ultrastructural changes of *Trichophyton mentagrophytes*,
524 *Fitoterapia.* 80 (2009) 290–296. doi:10.1016/J.FITOTE.2009.03.007.

525