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1 **Title:** Mixotrophic protists display contrasted biogeographies in the global ocean

2 **Running title:** Global biogeography of marine mixotrophic protists

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18

19

20 **Abstract**

21

22 Mixotrophy, or the ability to acquire carbon from both auto- and heterotrophy, is a
23 widespread ecological trait in marine protists. Using a metabarcoding dataset of
24 marine plankton from the global ocean, 318 054 mixotrophic metabarcodes
25 represented by 89 951 866 sequences and belonging to 133 taxonomic lineages
26 were identified and classified into four mixotrophic functional types: constitutive
27 mixotrophs (CM), generalist non-constitutive mixotrophs (GNCM), endo-
28 symbiotic specialist non-constitutive mixotrophs (eSNCM) and plastidic specialist
29 non-constitutive mixotrophs (pSNCM). Mixotrophy appeared ubiquitous, and the
30 distributions of the four mixotypes were analyzed to identify the abiotic factors
31 shaping their biogeographies. Kleptoplastidic mixotrophs (GNCM & pSNCM)
32 were detected in new zones compared to previous morphological studies.
33 Constitutive and non-constitutive mixotrophs had similar ranges of distributions.
34 Most lineages were evenly found in the samples, yet some of them displayed
35 strongly contrasted distributions, both across and within mixotypes. Particularly
36 divergent biogeographies were found within endo-symbiotic mixotrophs,
37 depending on the ability to form colonies or the mode of symbiosis. We showed
38 how metabarcoding can be used in a complementary way with previous
39 morphological observations to study the biogeography of mixotrophic protists and

40 to identify key drivers of their biogeography.

41

42 **Introduction**

43

44 Marine unicellular eukaryotes, or protists, have a tremendous range of life styles,
45 sizes and forms [1], showing a taxonomic and functional diversity that remains
46 hard to define [2, 3]. This variety of organisms is having an impact on major
47 biogeochemical cycles such as carbon, oxygen, nitrogen, sulfur, silica, or iron,
48 while being at the base of marine trophic networks [4–8]. Hence, they are key
49 actors of the global functioning of the ocean.

50 Historically, marine protists have been classified into two groups depending on
51 their trophic strategy: the photosynthetic plankton (phytoplankton) and the
52 heterotrophic plankton (zooplankton). It is now clear that mixotrophy, *i.e.* the
53 ability to combine autotrophy and heterotrophy, has been largely underestimated
54 and is commonly found in planktonic protists [6, 9–13]. Instead of a dichotomy
55 between two trophic types, their trophic regime should be regarded as a continuum
56 between full phototrophy and full heterotrophy, with species from many planktonic
57 lineages lying between these two extremes [10]. Mitra et al. [11] have proposed a
58 classification of marine mixotrophic protists into four functional groups, or
59 mixotypes. The constitutive mixotrophs, or CM, are photosynthetic organisms that

60 are capable of phagotrophy, also called "phytoplankton that eat" [11]. They include
61 most mixotrophic nanoflagellates (*e.g. Pymnesium parvum, Karlodinium micrum*).
62 On the opposite, the non-constitutive mixotrophs, or "photosynthetic zooplankton",
63 are heterotrophic organisms that have developed the ability to acquire energy
64 through photosynthesis [9]. This ability can be acquired in three different ways: the
65 generalist non-constitutive mixotrophs (GNCM) steal the chloroplasts of their prey,
66 such as most plastid-retaining oligotrich ciliates (*e.g. Laboea strobila*), the plastidic
67 specialist non-constitutive mixotrophs (pSNCM) steal the chloroplasts of a specific
68 type of prey (*e.g. Mesodinium rubrum* or *Dinophysis* spp.), and finally the endo-
69 symbiotic specialist non-constitutive mixotrophs (eSNCM) are bearing
70 photosynthetically active endo-symbionts (most mixotrophic Rhizaria from
71 Collodaria, Acantharea, Polycystinea, and Foraminifera, as well as dinoflagellates
72 like *Noctiluca scintillans*).

73 As drivers of biogeochemical cycles in the global ocean, and particularly of the
74 biological carbon pump [5, 14, 15], marine protists are a key part of ocean
75 biogeochemical models [7, 16–18]. However, physiological details of mixotrophic
76 energy acquisition strategies have only been studied in a restricted number of
77 lineages [9, 19, 20]. They appear to be quite complex and greatly differ across
78 mixotypes, which makes mixotrophy hard to include in a simple model structure
79 [21–25]. Hence at this time, mixotrophy is not included in most biogeochemical

80 models, neglecting the amount of carbon fixed by non-constitutive mixotrophs
81 through photosynthesis, and missing the population dynamics of photosynthetically
82 active constitutive mixotrophs that can still grow under nutrient limitation [23, 26].
83 This is most probably skewing climatic models predictions [11, 26], as well as our
84 ability to understand and prevent future effects of global change.

85 A better understanding of the environmental diversity of marine mixotrophic
86 protists, as well as a description of the abiotic factors driving their biogeography at
87 global scale are still needed, in particular to integrate them in biogeochemical
88 models. Leles et al. [27] attempted to tackle this problem by reviewing about 110
89 000 morphological identification records of a set of more than 60 mixotrophic
90 protists species in the ocean, taken from the Ocean Biogeographic Information
91 System (OBIS) database. They found distinctive patterns in the biogeography of
92 the three different non-constitutive mixotypes (GNCM, pSNCM and eSNCM),
93 highlighting the need to better understand such diverging distributions [27].
94 Environmental molecular biodiversity surveys through metabarcoding have been
95 widely used in the past fifteen years to decipher planktonic taxonomic diversity [2,
96 28–30]. Here we exploited the global *Tara* Oceans datasets [31–33], and identified
97 133 mixotrophic lineages, that we classified into the four mixotypes defined by
98 Mitra et al. [11]. This first ever set of mixotrophic metabarcodes allowed us to
99 investigate the global biogeography of both constitutive and non-constitutive

100 mixotrophs, in relation with *in-situ* abiotic measurements. We tested (i) if new
101 information on marine mixotrophic protists distribution can be gained in
102 comparison with previous morphological identifications [27]; (ii) if the constitutive
103 mixotrophs, which are not addressed in Leles et al. [27], and the non-constitutive
104 mixotrophs diverge in terms of biogeography; (iii) if the study of diversity and
105 abundance of environmental metabarcodes could lead to the definition of key
106 environmental factors shaping mixotrophic communities.

107

108 **Materials and Methods**

109

110 *Samples collection and dataset creation*

111 Metabarcoding datasets from the worldwide *Tara* Oceans sampling campaigns that
112 took place between 2009 and 2013 [31, 33] (data published in open access at the
113 European Nucleotide Archive under project accession number PRJEB6610) were
114 investigated. We analyzed 659 samples from 122 distinct stations, and for each
115 sample, the V9-18S ribosomal DNA region was sequenced through Illumina HiSeq
116 [32]. Assembled and filtered V9 metabarcodes (cf. details in de Vargas et al. [2])
117 were assigned to the lowest taxonomic rank possible *via* the Protist Ribosomal
118 Reference (PR2) database [34]. To limit false positives, we chose to only analyze
119 the metabarcodes (*i.e.* unique versions of V9 sequences) for which the assignment

120 to a reference sequence had been achieved with a similarity of 95% or higher. This
121 represents 65% of the total dataset in terms of metabarcodes and 84% in terms of
122 total sequences. Our dataset involved 1,492,912,215 sequences, distributed into
123 4,099,567 metabarcodes assigned to 5,071 different taxonomic assignments, going
124 from species to kingdom level precision.

125

126 *Defining a set of mixotrophic organisms*

127 Among these 5,071 taxonomic assignments, we searched for mixotrophic protist
128 lineages, taking into account the 4 mixotypes described by Mitra et al. [11]:
129 constitutive mixotrophs (CM), generalist non-constitutive mixotrophs (GNCM),
130 endo-symbiotic specialist non-constitutive mixotrophs (eSNCM), and plastidic
131 specialist non-constitutive mixotrophs (pSNCM). We used the table S2 from Leles
132 et al. [27] which is referencing 71 species or genera belonging to three non-
133 constitutive mixotypes (GNCM, pSNCM and eSNCM), as well as multiple other
134 sources coming from the recent literature on mixotrophy [6, 9–12, 35–47], and
135 inputs from mixotrophic protists' taxonomy specialists (cf. Acknowledgments
136 section). Within the 5,071 taxonomic assignments of variable precisions, we
137 identified 5 GNCM, 9 pSNCM, 77 eSNCM, and 42 CM lineages (detailed list
138 available publicly under the DOI 10.6084/m9.figshare.6715754, and all
139 metabarcodes were tagged with their mixotypes in the PR2 database). Among these

140 133 taxonomic assignments that we will call “lineages”, 92 were defined at the
141 species level, 119 at the genus level, and the last 14 at higher taxonomic levels
142 where mixotrophy is always present (mostly eSNCM groups like Collodaria). In
143 the Chrysophyceae family, metabarcodes assigned to clades B2, E, G, H and I were
144 included even though we couldn’t find a general proof that all species included in
145 these clades have mixotrophic capabilities. However, if we exclude the
146 photolithophilic Synurophyceae and genera like *Paraphysomonas* and *Spumella*,
147 which we did, a vast majority of Chrysophyceae are considered mixotrophic [10].
148 The final dataset included 318 054 metabarcodes assigned to the 133 mixotrophic
149 lineages selected, as well as their sequence abundance in 659 samples (table
150 available publicly under the DOI 10.6084/m9.figshare.6715754).

151

152 *Environmental dataset*

153 We built a corresponding contextual dataset using the environmental variables
154 available in the PANGAEA repository from the *Tara* Oceans expeditions [33, 49].
155 The set of 235 environmental variables was reduced to 57 due to several selection
156 steps (Data available publicly under the DOI 10.6084/m9.figshare.6715754; see the
157 details of variable selection in section 1 of Supp. Mat.).

158

159 *Distribution and diversity of mixotrophic protists*

160 For each mixotype, the number of metabarcodes, the total sequence abundance and
161 the mean sequence abundance by metabarcode was computed (Table 1). Also, we
162 measured each metabarcode's station occupancy, *i.e.* the number of stations in
163 which it was found, and station evenness, *i.e.* the homogeneity of its distribution
164 among the stations in which it was detected (Figure 2). Diversity of mixotrophic
165 protists was investigated through mixotype-specific metabarcode richness per
166 station (Table 1). As the number of samples taken per station can impact the
167 abundance and diversity of detected metabarcodes, richness was computed only at
168 stations for which the maximum number of 8 samples were available (40 stations
169 over 122).

170

171 *Global biogeography of mixotrophic protists*

172 Two statistical analyses were performed to investigate mixotrophic protists
173 biogeography. One at the metabarcode level, and one at the lineage level, *i.e.*
174 merging the sequence abundance of metabarcodes sharing the same taxonomical
175 assignation. The metabarcodes abundance table was composed of 318054
176 rows/metabarcodes, and 659 columns/samples, whereas the lineage abundance
177 table was composed of 133 rows/lineages and 659 columns/samples (both datasets
178 are available publicly under the DOI 10.6084/m9.figshare.6715754). The two
179 analyses led to very similar conclusions, but the biogeography of lineages appeared

180 easier to visually represent and interpret than the one of metabarcodes. Hence, we
181 only present here the results of the lineage-based analysis (See section 3 of Sup.
182 Mat. for metabarcode-level analysis results and discussion).

183 Our statistical model was designed to identify lineages (or metabarcodes) with
184 contrasted biogeographies, and relate their presence to the environmental context.

185 We normalized the sequence counts from the lineage abundance matrix using a
186 Hellinger transformation [51]. We used the environmental dataset and the
187 mixotrophic lineages' abundance matrix as explanatory and response matrices,
188 respectively, to conduct a redundancy analysis (RDA) [51]. For that, we made a
189 species pre-selection using Escoufier's vectors [52] which allowed to keep only the
190 62 most significant mixotrophic lineages. This method selects lineages according to
191 a principal component analysis (PCA), sorting them based on their correlation to
192 the principal axes. We then used a maximum model ($Y \sim X$) and a null model ($Y \sim 1$)
193 to conduct a two directional stepwise model selection based on the Akaike
194 information criterion (AIC) [53]. The resulting model contained 28 environmental
195 response variables. More details about statistical analyses are available in section 2
196 and 3 of the supplementary materials. Analyses and graphs were realized with the
197 R software version 3.4.3 [54]. All scripts are available on GitHub platform
198 (<https://github.com/upmcgenomics/MixobioGeo>).

199

200 **Results**

201

202 *Global distribution and diversity of marine mixotrophic protists*

203 Mixotrophic protists metabarcodes were detected in all the 659 samples with a total
204 sequence abundance of 89 951 866, representing 12.56% of the total sequence
205 abundance in the 659 samples studied. They represented a mean of 12.64% of the
206 total sequence abundance per sample, with a maximum of 96.96% and a minimum
207 of 0.01%. To avoid any potential overestimation of mixotrophic lineages presence
208 in the following results, we marked all records of less than a hundred sequences as
209 questionable. We found both eSNCM and CM in each of the 122 stations studied
210 (Table 1, Figure 1). In only two occasions the number of sequences belonging to
211 CM was questionable, at stations for which only one sample was sequenced.
212 GNCM were found absent in only 2 stations and their presence was questionable in
213 39 stations (Figure 1). pSNCM were absent at 5 stations (3 in the Indian Ocean, and
214 2 in the Pacific Ocean) and detected with questionable presence in 54 additional
215 stations, which were mostly located in the central Pacific and the Indian Ocean
216 (Figure 1). We found significant amounts of sequences corresponding to GNCM in
217 the Central Pacific, Southern subtropical Atlantic, and Indian Ocean. The presence
218 of GNCM in these areas has not yet been recorded through morphological
219 identifications during field expeditions [27]. Also, we detected more than 100

220 sequences of pSNCM metabarcodes at 11 stations belonging to biogeographical
221 provinces in which no morphological identifications had been published [27, 55],
222 mostly in offshore areas of the Atlantic and Pacific Ocean (Figure 1).

223
224 The mean evenness of mixotrophic metabarcodes across stations was of 0.87, and
225 82.3% of the metabarcodes had a station evenness above 0.5 (Figure 2). Station
226 occupancy varied a lot depending on the metabarcodes, with a high density of rare
227 metabarcodes leading to a mean of 5.14 stations over a maximum of 122, and a
228 standard deviation of 7.7. However, three eSNCM metabarcodes were found in all
229 the 122 stations, and three CM metabarcodes were detected in 121 stations. The
230 maximum occupancy for a GNCM metabarcode was of 111 stations, while 92
231 stations was the maximum for a pSNCM metabarcode. CM and GNCM
232 metabarcodes showed a strong tendency towards high evenness values (Figure 2,
233 means of 0.90 and 0.95, respectively), even for the most sequence abundant
234 metabarcodes. Many eSNCM metabarcodes had high evenness values, but below
235 average values were detected for the most abundant ones (Figure 2, global mean of
236 0.87). pSNCM metabarcodes had a similar mean of evenness values (0.87), but a
237 different distribution compared to other mixotypes (Figure 2). Among the 50 most
238 abundant metabarcodes, 43 corresponded to Collodaria lineages, 47 were eSNCM

239 and 3 were CM, all three assigned to *Gonyaulax polygramma*. GNCCM and pSNCCM
240 metabarcodes had homogeneously low sequence abundances (Figure 2, Table 1).

241

242 *Main factors affecting the biogeography of mixotrophic protists*

243 The redundancy analysis helped to investigate further the environmental variables
244 responsible for the mixotrophic protists' biogeography. The 62 lineages selected
245 with the Escoufier's vector method corresponded to 20 CM, 34 eSNCCM, 3 GNCCM
246 and 5 pSNCCM. Even after selection, a significant part of the lineages did not show
247 any response to environmental data in their distribution (Figure 3, *e.g.* 19 of the 62
248 lineages were found between -0.01 and 0.01 on both RDA1 and RDA2). The
249 adjusted R-squared of the RDA was of 34.89% (41.43% unadjusted), with 24.01%
250 of variance explained on the two first axes (Figure 3). The first RDA axis (14.96%)
251 marks an opposition between samples from oligotrophic waters with low
252 productivity (RDA1>0) and samples from eutrophic and productive water masses
253 (RDA1<0). This axis is negatively correlated to chlorophyll concentration, particles
254 density, ammonium concentration, absorption coefficient of colored dissolved
255 organic matter (acCDOM), duration of daylight, silica, CO₃, oxygen, and PO₄
256 concentration, as well as longitude. It is positively correlated to bathymetry, deep
257 euphotic zone, deep oxygen maximum, deep mixed layer, as well as to the distance
258 to coast. The second RDA axis (9.05%) is opposing offshore and subpolar samples

259 (RDA2>0) to coastal and subtropical ones (RDA2<0). The axis is positively
260 correlated to the depth of the mixed layer, the distance to coast, the bathymetry,
261 high maximum Lyapunov exponents as well as high concentrations of PO₄,
262 oxygen, CO₃ and silica. It is negatively correlated to temperature, salinity and
263 photosynthetically active radiations (PAR).

264
265 Among the 20 CM lineages, 7 clearly emerged from the redundancy analysis
266 (Figure 3) and showed distinct biogeographies related to environmental variables.
267 *Gonyaulax polygramma*, *Alexandrium tamarense* and *Fragilidium mexicanum*,
268 three Dinophyceae belonging to the Gonyaulacales order, were mainly found in
269 oligotrophic waters with a deep euphotic zone, warm temperature, high salinity and
270 PAR (RDA1>0, RDA2<0). The four other CMs (involving all the Chrysophyceae
271 included in the analysis as well as one Dinophyceae from the Kareniaceae family,
272 *Karlodinium micrum*) were found mostly in productive water masses (RDA1<0).

273
274 eSNCMs can be divided in three groups in the RDA space. The first group
275 (RDA1<0) corresponds to eSNCM species dominating rich and productive
276 environments. It includes mainly Acantharia and Spumellaria species. The second
277 group (RDA1>0) dominates oligotrophic environments, and includes multiple
278 Collodaria as well as one Dinophyceae genus (*Ornithocercus*). Within this group,

279 *Ornithocercus* spp. is found mainly in coastal subtropical environments (RDA2<0),
280 as opposed to *Sphaerozoum punctatum* that is found mainly in offshore subpolar
281 regions (RDA2>0). *Siphonosphaera cyathina* lies between these two trends as it is
282 found only in oligotrophic samples, but isn't influenced by temperature or
283 bathymetry (Figure 3 and 4). The third group corresponds to the eSNCM lineages
284 that can be interpreted as distributed homogeneously in regards of the
285 environmental data we are using (e.g. lineages with the shortest arrows in Figure
286 3). These notably include the 12 Foraminifera lineages present in the RDA.
287 Looking at filters centroids in the RDA space (Figure 3), we can suppose that
288 eSNCM lineages dominating eutrophic systems (RDA1<0) are smaller in size than
289 those dominating oligotrophic ones (RDA1>0).

290
291 Out of the five pSNCM included in the RDA, only *Mesodinium rubrum*, the most
292 abundant one, is distinctively represented in the RDA space. This suggests that the
293 other pSNCM have homogeneous distributions in response to our environmental
294 variables. *Mesodinium rubrum* dominates eutrophic environments, independently
295 from the bathymetry or the temperature (RDA1<0, RDA2 ≈ 0). We find a similar
296 pattern for GNCM, with only *Pseudotontonia simplicidens* well represented in the
297 RDA space out of the three species included in the analysis. Like *M. rubrum*,

298 *Pseudotontonia simplicidens* is the most abundant species in its group and it is
299 mainly found in eutrophic waters (RDA1<0).

300

301 **Discussion**

302

303 *Mixotrophy occurs everywhere in the global ocean*

304 Our metabarcoding survey confirms that marine mixotrophic protists are ubiquitous
305 in the global ocean [27], possibly extending the known range of distribution of two
306 mixotypes (Figure 1 and 2). Mixotrophic organisms represented more than 12% of
307 the sequences in the complete *Tara* Oceans metabarcoding dataset, showing that
308 they should not be understated. We found contrasted biogeographies among
309 metabarcodes and their corresponding lineages, both within and across mixotypes
310 (Figure 2, 3, 4 and S1, Sup. Mat. section 3). We found constitutive mixotrophs
311 (CM) and endo-symbiotic specialist non-constitutive mixotrophs (eSNCM)
312 metabarcodes at all the 122 stations included in this global study (Table 1 and
313 Figure 2), verifying that these two mixotypes are the most abundant in the ocean
314 [27, 47, 54, 55]. This dominance of eSNCM and CM in our data is also linked to
315 the relatively high number of metabarcodes available for these two mixotypes in
316 databases. Using 1 360 generalist non-constitutive mixotrophs (GNCM)
317 metabarcodes corresponding to only 5 lineages, we detected them in 10

318 biogeographical provinces [55] where no morphological identification had been
319 recorded before [27]. GNCM metabarcodes had consistently high evenness values,
320 and some had station occupancy records comparable to the most abundant eSNCM
321 and CM metabarcodes (Figure 2). These results support the hypothesis of a globally
322 ubiquitous distribution of GNCM. Plastidic specialist non-constitutive mixotrophs
323 (pSNCM) were found in 5 provinces in which no record existed so far from
324 morphological identification field studies [27]. However, these observations were
325 often in a questionable range in terms of sequence abundance (Figure 1), and the
326 overall distribution of pSNCM in our data appears as very concordant with
327 morphological observations [27]. pSNCM metabarcodes had dominantly low
328 station evenness values, which again supports the conclusions of Leles et al. [27]
329 that identified pSNCM as highly seasonal and spatially restricted in their
330 distribution.

331 While building our set of mixotrophic lineages, some widespread and potentially
332 mixotrophic genera did not appear, such as *Ceratium* spp., *Tontonia* spp.,
333 *Amphisolenia* spp., *Triposolenia* spp. or *Citharistes* spp., mainly because of a poor
334 representation in the PR2 database. Also, we decided to only consider
335 metabarcodes with more than 95% similarity to a reference sequence. This
336 threshold could be too selective for some species and not enough for some others,
337 as single similarity threshold are hardly efficient when studying whole eukaryotic

338 populations [56, 57]. For example, some species appeared with low sequence
339 abundance in the data even though they couldn't have been sampled, such as three
340 lacustrine species, *e.g.* *Poterospumella lacustris*. Considering these biases and the
341 sometimes relatively low sequence counts (marked as questionable in Figure 1),
342 some of the new GNCM and pSNCM records we observed should be considered
343 with care, as they could be over-estimated or even sometimes artefactual. However,
344 the low number of lineages found for these two mixotypes in PR2 and in our
345 dataset are leading us to think that we were unable to capture the whole GNCM and
346 pSNCM communities. This supposes a global underestimation of GNCM and
347 pSNCM abundances in our results.

348 *Tara* Oceans metabarcoding dataset is built on snapshot samples taken irregularly
349 during a three-year cruise, hence allowing no proper seasonal variations
350 investigations. However, morphological identifications of mixotrophic protists
351 revealed seasonal variations in their abundance, with *Mesodinium* biomass
352 blooming in spring in coastal seas for example [27]. As metabarcoding datasets
353 have been successfully applied on time series to detect species successions across
354 gradients of time and space [58–60], it would be interesting to similarly investigate
355 seasonal trends in mixotrophic communities. Our set of mixotrophic lineages and
356 metabarcodes being publicly available, our method will be applicable to any other

357 metabarcoding dataset, including time-series. It will also be open to inputs and
358 updates from the global scientific community.

359

360 *The contrasted biogeographies of marine mixotypes*

361 • *Constitutive Mixotrophs*

362 Constitutive mixotrophs (CM) have very diverse feeding behaviours, with some
363 species requiring phototrophy to grow, others phagotrophy, and some being
364 obligate mixotrophs [9]. They were described in all waters of the global ocean [61–
365 65]. We found them distributed in a range of conditions almost as wide as non-
366 constitutive mixotrophs (Figure 1 and 3). Among highly abundant lineages, most
367 were dominantly found in eutrophic and shallow habitats. However, a few
368 dinoflagellates were found to be highly dominant in oligotrophic, subtropical
369 waters, showing how wide of a range of conditions constitutive mixotrophs can
370 grow in (Figure 3). This illustrates how mixotrophy can allow organisms to
371 dominate ecosystems even when environmental conditions are poorly adapted to
372 purely phototrophic or heterotrophic organisms. When taken explicitly into account
373 in biogeochemical models, marine mixotrophs increase carbon export by up to 30%
374 [22]. Hence, their global ubiquity supposes that the carbon export of the biological
375 carbon pump could be underestimated in both oligotrophic and eutrophic areas
376 [26].

377 • *Plastidic-Specialist and Generalist Non-Constitutive mixotrophs*
378 (*pSNCM & GNCM*)

379 Like Leles et al. [27], we found pSNCM and GNCM to have quite similar
380 biogeographies (Figure 3, section 3 of Sup. Mat.). Sequence abundance of most of
381 the metabarcodes for these two mixotypes was homogeneously low (Table 1), but
382 the two most abundant species, *Mesodinium rubrum* (pSNCM) and *Pseudotontonia*
383 *simplicidens* (GNCM), were found mostly in coastal and eutrophic waters,
384 consistently with Leles et al. [27]’s morphological observations (Figure 3, section 3
385 of Sup. Mat.). No species-level barcode is available in the PR2 database for the
386 *Tontonia* genus, and only one can be found for *Pseudotontonia* and *Laboea* genera,
387 even though morphological records of these GNCM are numerous [27].
388 Experiments using meso- and microcosms combined with individual counts and
389 morphological identification have found that GNCM ciliates can represent up to
390 half of the individuals in ciliate communities of the photic zone [11, 66, 67]. A
391 proportion we would have trouble to reach with the 5 lineages we were able to
392 consider, knowing that there are 8,686 different ciliate lineages available in PR2.
393 This highlights the urgent need for supplementing 18S reference databases for
394 mixotrophic ciliates.

395 • *Endo-symbiotic Specialist Non-Constitutive Mixotrophs (eSNCM)*

396 Endo-symbiotic specialist non-constitutive mixotrophs (eSNCM) is by far the most
397 widespread and abundant non-constitutive mixotype in the global ocean (Figure 1
398 and 2) [27, 47, 54]. Their biogeography stands out, with a lot of highly abundant
399 ubiquitous lineages, and some other specialized towards certain types of
400 ecosystems (Figure 3). They represent 95.7% of the sequence counts in our study
401 and correspond to 90.7% of the metabarcodes (Table 1), which highlights their
402 abundance and diversity. The very high number of rDNA copies present in Rhizaria
403 orders such as Collodaria [47] might lead the eSNCM to appear more abundant in
404 metabarcoding datasets than they ecologically are. However, in oligotrophic open
405 oceans the Rhizaria biomass is estimated to be equivalent to that of all other
406 mesozooplankton [68], and positively correlated to the carbon export [15], showing
407 how ecologically important they can be.

408

409 *Investigating the divergent biogeographies of Collodaria and Acantharia*

410 Collodaria are living either as solitary large cells or as colonies [47], which
411 explains why they are predominantly found in macro-sized (180-2000 μm) filter
412 samples (Figure 3). All described Collodaria species so far harbour photosynthetic
413 endo-symbionts, mostly identified as the dinoflagellate species *Brandtodinium*
414 *nutricula* [47, 69]. These dinoflagellates are able to get in and out of their
415 symbiotic state, which implies a light and/or reversible effect of the Collodarian

416 host on its symbiont metabolism [69]. Based on the same metabarcoding dataset,
417 Collodaria were described as particularly abundant and diverse in the oligotrophic
418 open ocean [47]. In our results, Collodaria dominate oligotrophic, relatively deep
419 waters (Figure 3 and 4a). These Collodaria appear opposed to another set of
420 Rhizaria (Acantharia and Spumellaria) linked to eutrophic and shallow waters
421 (Figure 3 and 4b, section 3 of Sup. Mat.). Acantharia are found ubiquitously in the
422 global ocean, but display particularly high sequence abundances in some specific
423 regions [54]. Mixotrophic Acantharia live in symbiosis with the cosmopolitan
424 haptophyte *Phaeocystis*, which is highly abundant and ecologically active in its
425 free-living phase [54]. Unlike the one of Collodaria, this symbiosis is irreversible :
426 an algal symbiont can not go back to its free-living phase [54]. Our results suppose
427 that these specific symbiotic modes could enable Acantharia and Collodaria to
428 dominate different ecosystems (Figure 3 and 4). Moreover, living in colonies as
429 Collodaria could help to dominate oligotrophic systems, *e.g.* by accumulating more
430 food and nutrients through their gelatinous extra-cellular matrix [47]. Experiments
431 and modeling studies should help to investigate the contribution of this assumption,
432 comparing food acquisition capacity and growth rates of free individuals *versus* in
433 colony.

434

435 *Towards an integration of mixotrophic diversity into marine ecosystem models*

436 The future of marine communities' modeling lies in the integration of omics
437 datasets into modeling frameworks [18, 70–73]. The use of metabolic networks and
438 gene-centric methods has already shown very promising results in modeling
439 prokaryotic ecological dynamics [18, 73]. However, eukaryotic metabolic
440 complexity makes these methods hard to apply on protists for now, and we still
441 lack a universal theoretical framework on how to integrate such methods into
442 concrete modeling [70]. Mixotrophic protists are physiologically complex, and
443 their feeding behaviour can vary drastically on short time scales [9]. It will then
444 take a few more years of comparative genomics and transcriptomics studies before
445 being able to model their physiology with purely gene-based approaches. Still,
446 mechanistic models of mixotrophy exist and are quite complex [21, 23], even if the
447 one from Ghyoot et al. [23] could be implemented in a global biogeochemical
448 model [74]. Most models make the choice to represent either one or two (NCM and
449 CM) types of organisms able to play the role of all mixotypes depending on
450 parameterization. However, no global agreement has been reached on to what
451 extent the different mixotypes should be modeled. This is mainly due to a lack of
452 quantitative and comparative data on the global impact of grazing and carbon
453 fixation by the different mixotypes [75]. With our study, we show how meta-omics
454 data can be used to identify groups of organisms distributed differently in response
455 to the environment. It also allows the identification of ecological traits and

456 environmental factors potentially responsible for these divergences. This
457 information can be used to identify key species or lineages, and design controlled
458 experiments with variations of targeted environmental factors to produce the
459 quantitative data needed by modelers. Considering our results, we propose that
460 host-symbiont dynamics of eSNCM should be investigated as a trait playing a
461 potential role on Rhizaria ability to thrive in oligotrophic conditions. Particularly,
462 the mechanisms behind holobiont formation and its potential reversibility could
463 play major roles on eSNCM carbon fixation in various nutrient conditions. Future
464 experiments comparing responses of Collodaria and Acantharia holobionts to
465 different stresses in terms of grazing and carbon fixation could lead to a better
466 understanding of the physiological differences between their two modes of
467 symbiosis. Also, our results suggest that the metabolic flexibility of CM should
468 allow this mixotype to grow in almost any conditions, with individual species
469 probably spanning continuously between complete autotrophy and complete
470 heterotrophy. The risk is then to create a "perfect beast" mixotroph dominating all
471 systems [21]. To avoid that, we need more comparative data on grazing and carbon
472 fixation of obligate phototrophs *versus* obligate heterotrophs in response to nutrient
473 depletion and environmental fluctuation. Here again, meta-omics data could help to
474 identify candidates for efficient experiment designs. Finally, the small number of
475 lineages of GNCM and pSNCM in our study makes it hard to come up with

476 strongly supported conclusions on whether they should be differentiated in models
477 or not. They seem to share similar biogeographies using snapshot data (Figure 3,
478 section 3 of Sup. Mat.), but considering that they have different abilities for
479 conserving stolen chloroplasts over time, it might not be the case when looking at a
480 time series analysis [20, 76, 77].

481
482 Our study uses meta-omics data to investigate the global distribution and
483 biogeography of mixotrophic protists in the ocean. Our results, currently based on
484 metabarcoding data, complement morphological records and will be complemented
485 in the near future by metagenomics and metatranscriptomics studies. The latter will
486 allow to distinguish the protists with mixotrophic capabilities from the ones with
487 ongoing mixotrophic activity. This could lead to quantitative estimations of
488 mixotrophic rates in environmental samples, allowing a sharpened study of
489 mixotrophic protists ecology in the global ocean. It could also lead to a metabolic
490 description of complex processes like kleptoplasty and endo-symbiosis, hence
491 facilitating the modeling of mixotrophic behaviours and its incorporation in ocean
492 biogeochemical models.

493

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495

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522

523 **Competing interests**

524

525 The authors declare that they have no competing interests.

526

527 *Supplementary information is available at ISME’s journal website.*

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714

715

716 **FIGURES & TABLES LEGENDS**

717

718 *Figure 1:*

719 Global distribution of mixotypes from metabarcoding data. Maps showing for each
720 station the proportion of sequences (in %) belonging to each mixotype over the
721 total number of mixotrophic sequences. Stations in which no sequence was found
722 were marked as absent, ones with less than 100 sequences marked as questionable.
723 Each Longhurst biogeographical provinces [53] is coloured in the background if
724 more than 100 sequences are detected in at least one of its stations.

725

726 *Figure 2:*

727 Sequence abundance, occupancy and spatial evenness of each mixotrophic
728 metabarcode across sampled stations. Each metabarcode is plotted as a bubble, with
729 its station occupancy, *i.e.* the number of stations in which it was found, and its
730 station evenness, *i.e.* the homogeneity of its distribution among the stations in
731 which it was detected, as coordinates. Violin plots were drawn for each mixotype
732 on both the x and y axes. The size of each bubble is scaled to the sequence
733 abundance found globally for the corresponding metabarcode.

734

735 *Figure 3:*

736 Impact of environmental variables on the distribution of marine mixotrophs. Triplot
737 of the redundancy analysis (RDA) computed on the 62 Escoufier-selected lineages,
738 after model selection. The adjusted R-squared of the analysis is of 34.89% (41.43%
739 unadjusted). Each grey dot corresponds to a sample, *i.e.* one filter at one depth at
740 one station. The blue dashed arrows correspond to the quantitative environmental
741 variables. Abbreviations are as follows: *MLD* = mixed layer depth, *O2MaxD* = O2
742 maximum depth, *EuphzoneD* = euphotic zone depth, *PAR* = photosynthetically
743 active radiations, *Calcite Sat. St.* = Calcite Saturation State, *c_660* = optical beam
744 attenuation coefficient at 660 nm, *c_part* = beam attenuation coefficient of
745 particles, *acCDOM* = absorption coefficient of colored dissolved organic matter.
746 Plain arrows correspond to mixotrophic lineages, colors indicating mixotypes. For
747 more readability, we do not represent all qualitative variables included in the
748 model. That is why only the filter centroids are appearing, even though the
749 sampling depth, season, season moment, *i.e.* early, middle or late, and
750 biogeographical province were used in the RDA calculation.

751

752 *Figure 4:*

753 Contrasted global distributions of metabarcodes corresponding to two eSNCM
754 lineages. Maps of Hellinger-transformed sequence count abundances for
755 metabarcodes assigned to the Collodaria *Siphonosphaera cyathina* (A) and the

756 Acantharia Acanthrometridae F3 spp. (B). These two lineages are opposed on the
757 first RDA axis (Figure 3 and S1). Size and color both illustrate abundance for better
758 readability. Ellipses were drawn to highlight high abundance zones, and reveal the
759 differences in lineages distribution.

760

761 *Table 1:*

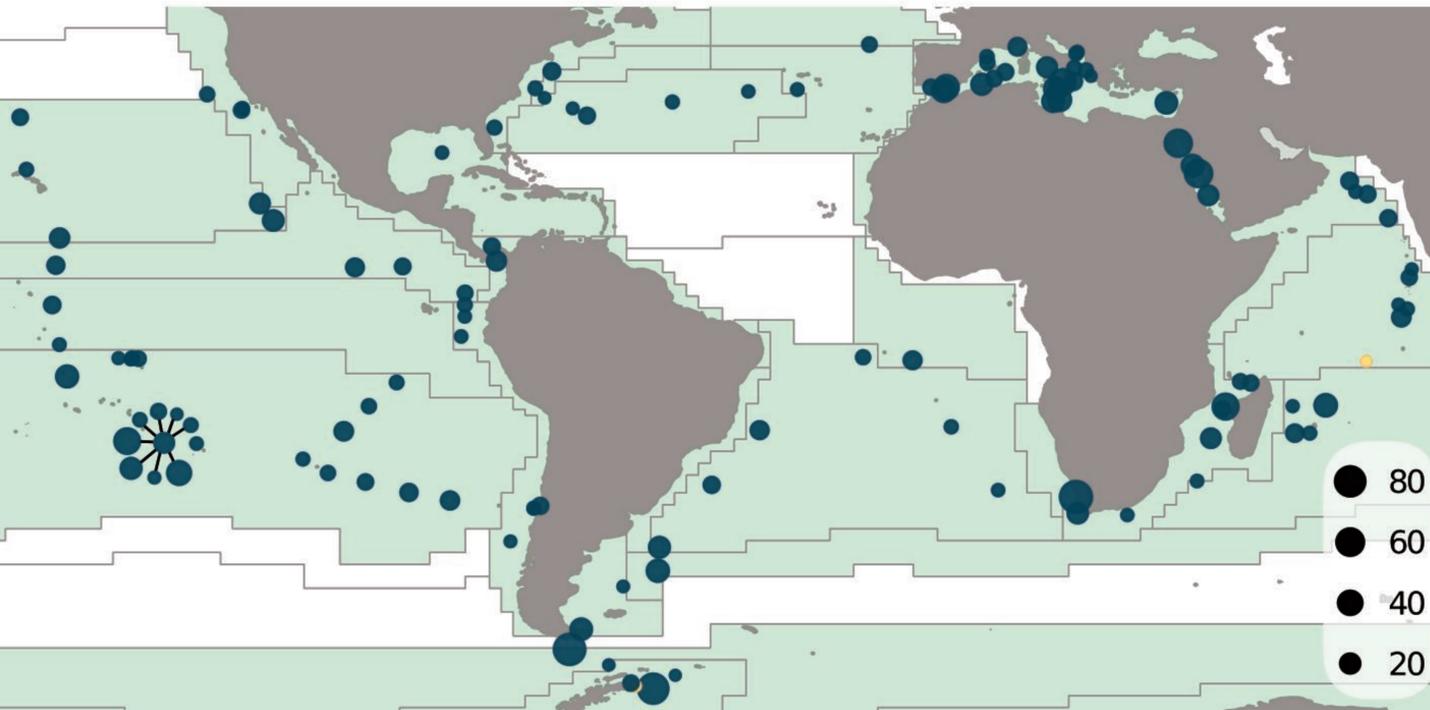
762 Detailed number of lineages found for each mixotype, as well as the number of
763 metabarcodes, the corresponding total sequence counts over all stations, the mean
764 sequence abundance by metabarcode, and mean metabarcode richness. The
765 richness was computed as the number of different metabarcodes present at each
766 station. It was calculated for each mixotype and means are indicated in the fifth
767 column. Absences correspond to the number of stations in which no sequences
768 were detected for the corresponding mixotype.

769 *The mean indicated here was calculated using only stations having the maximum
770 number of samples (see main text).

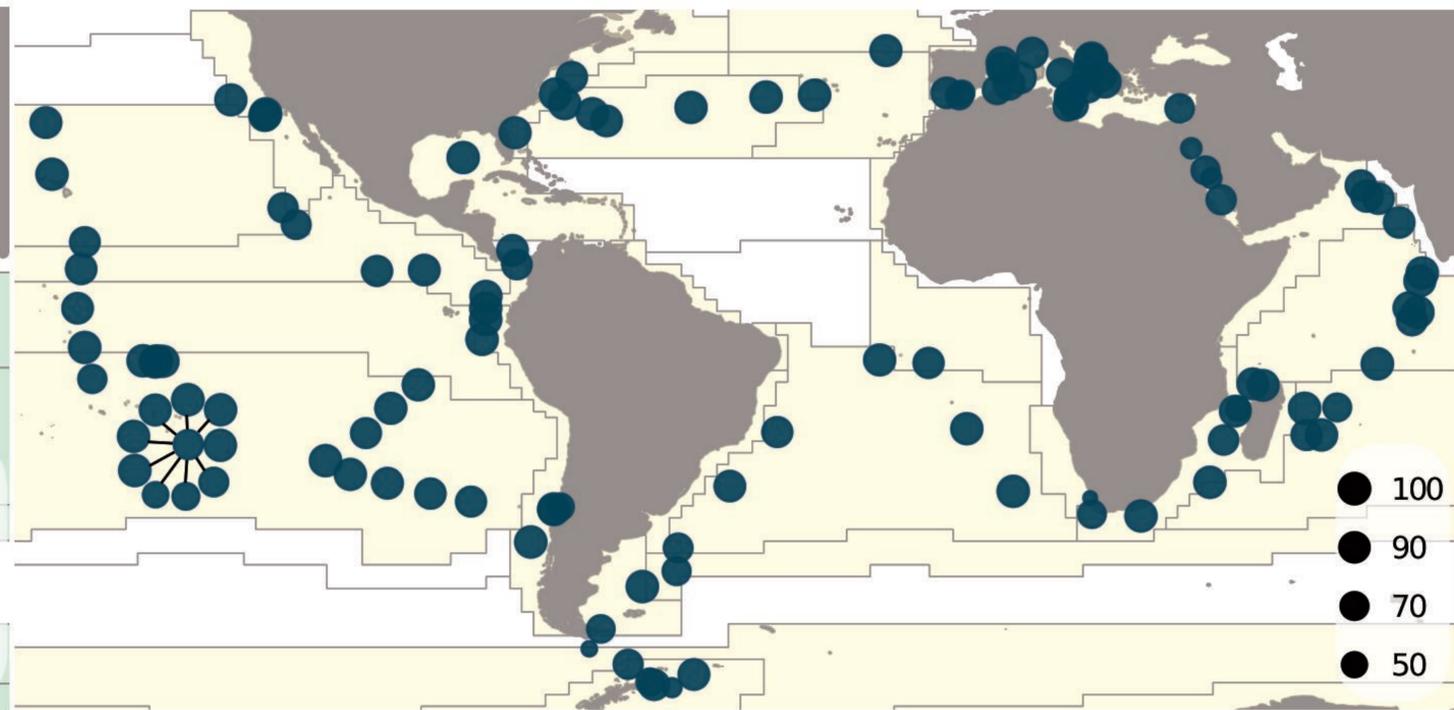
771

Mixotypes	CM	eSNCM	pSNCM	GNCM
Number of lineages used in this study	42	77	9	5
Number of V9 metabarcodes	26015	288536	2143	1360
Total sequence abundance	3,581,751	86,098,397	208,096	63,622
Mean sequence counts per metabarcode	137.7	298.4	97.1	46.8
Mean metabarcode richness per station* (Std Dev)	2162 (1115)	18502 (9238)	67 (102)	84 (111)
Number of absences/station	0/122	0/122	5/122	3/122

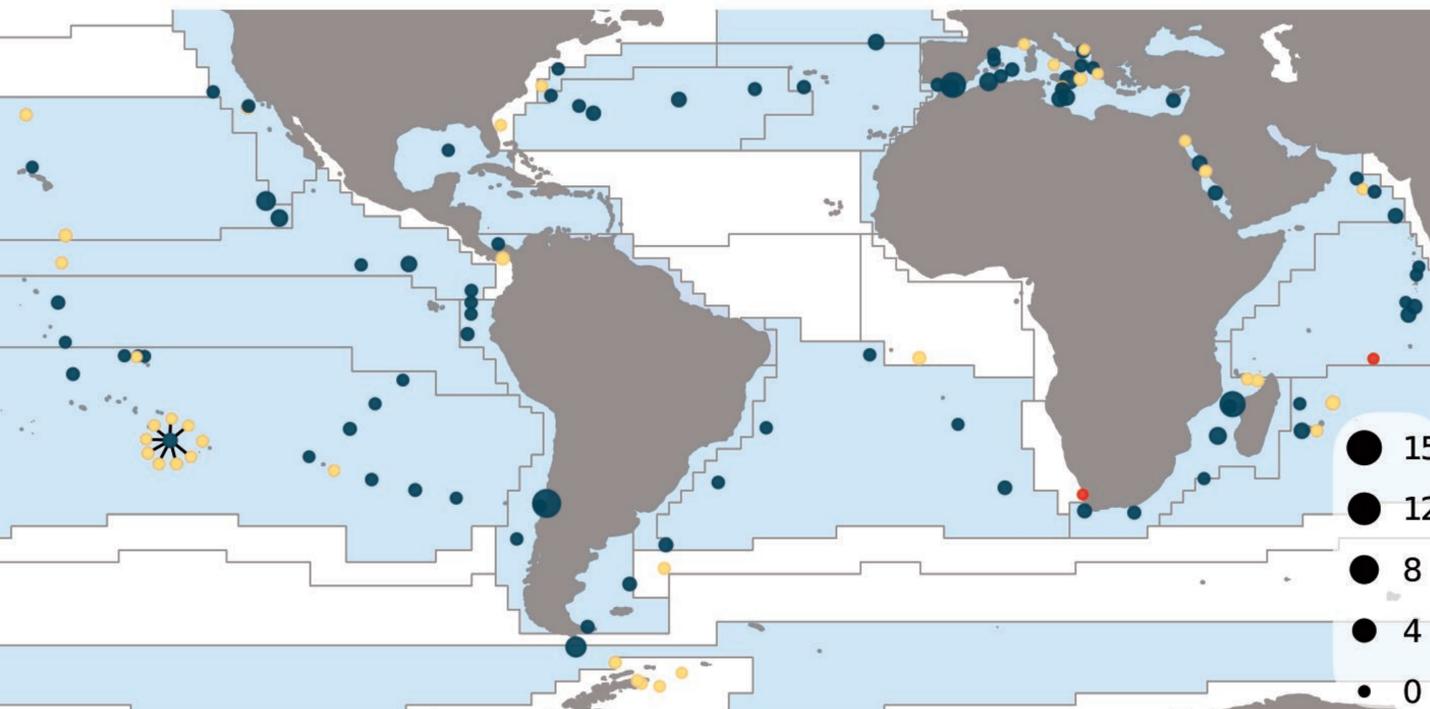
CM



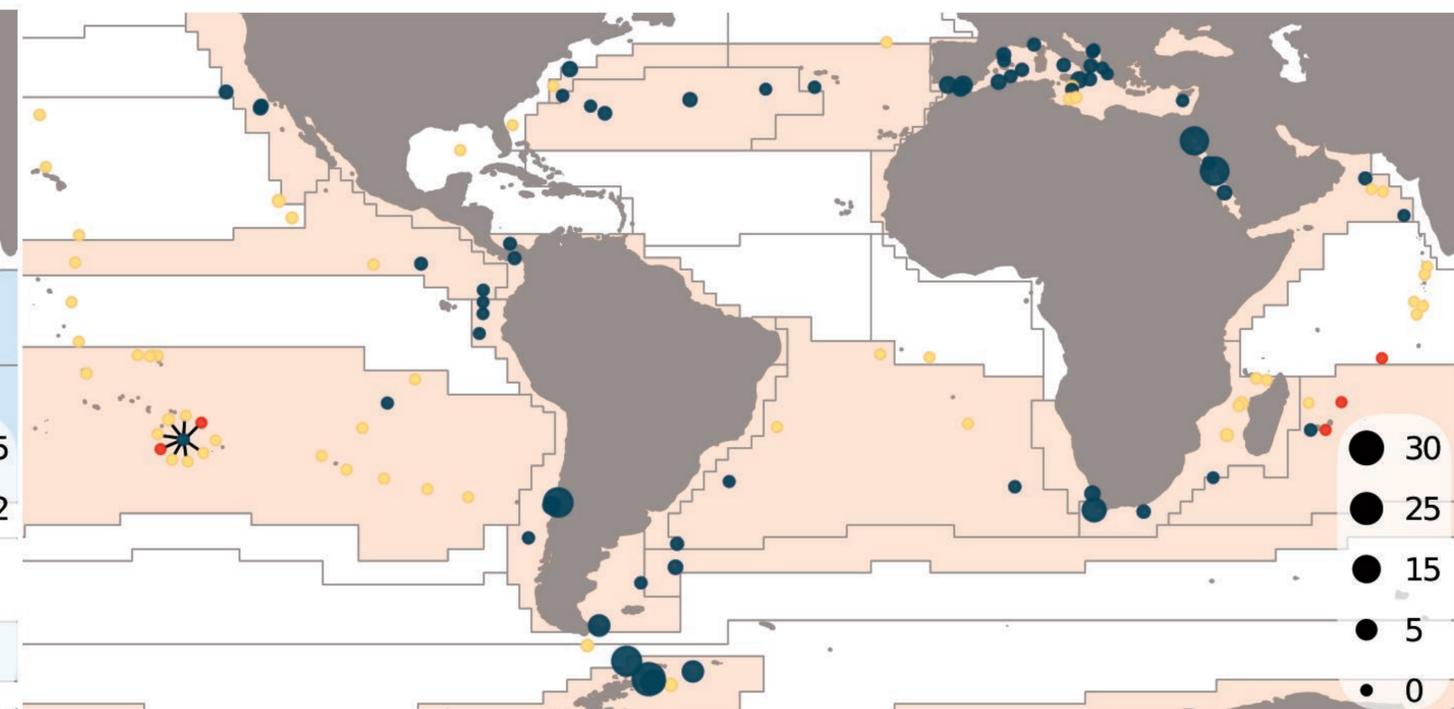
eSNCM



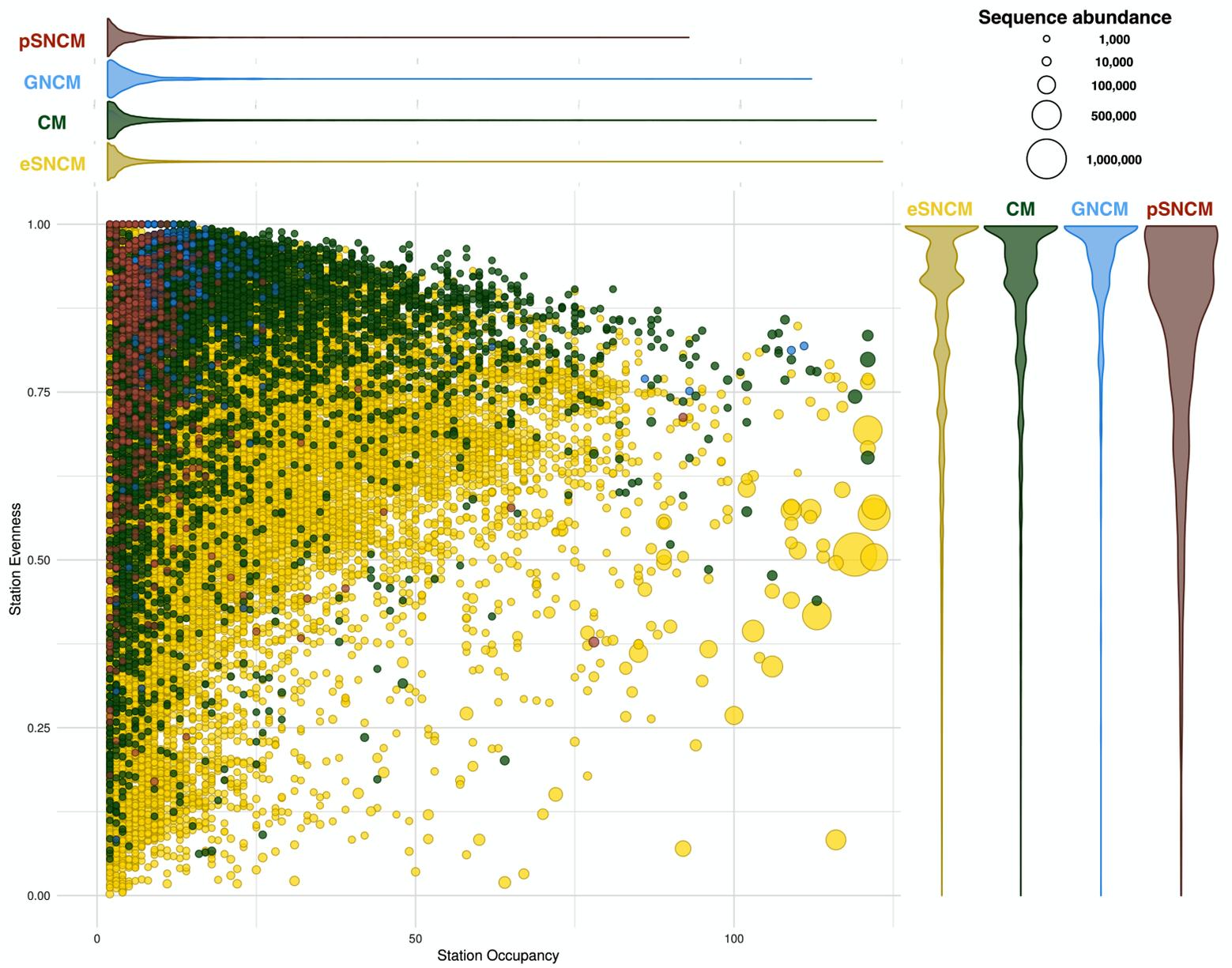
GNCM

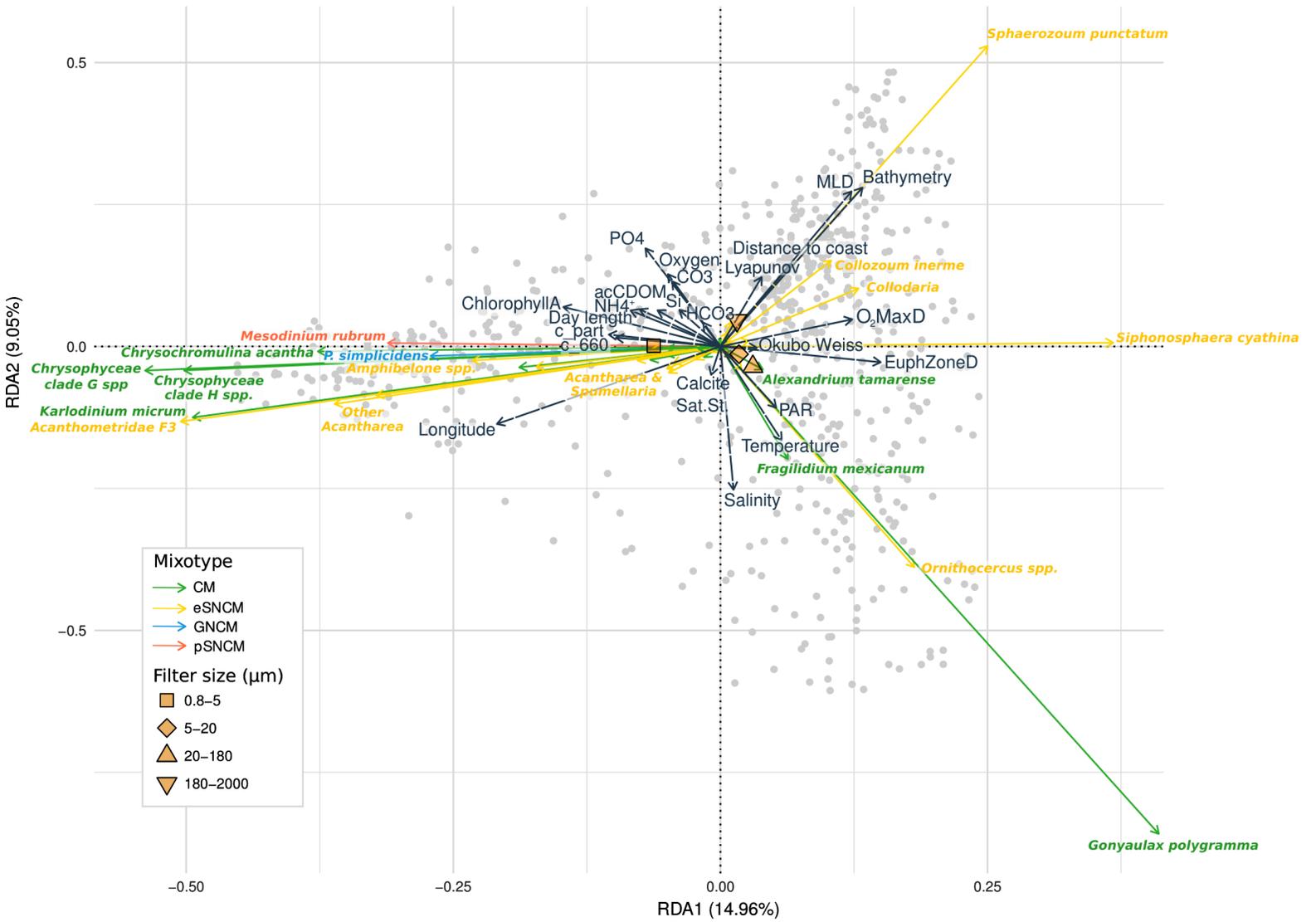


pSNCM

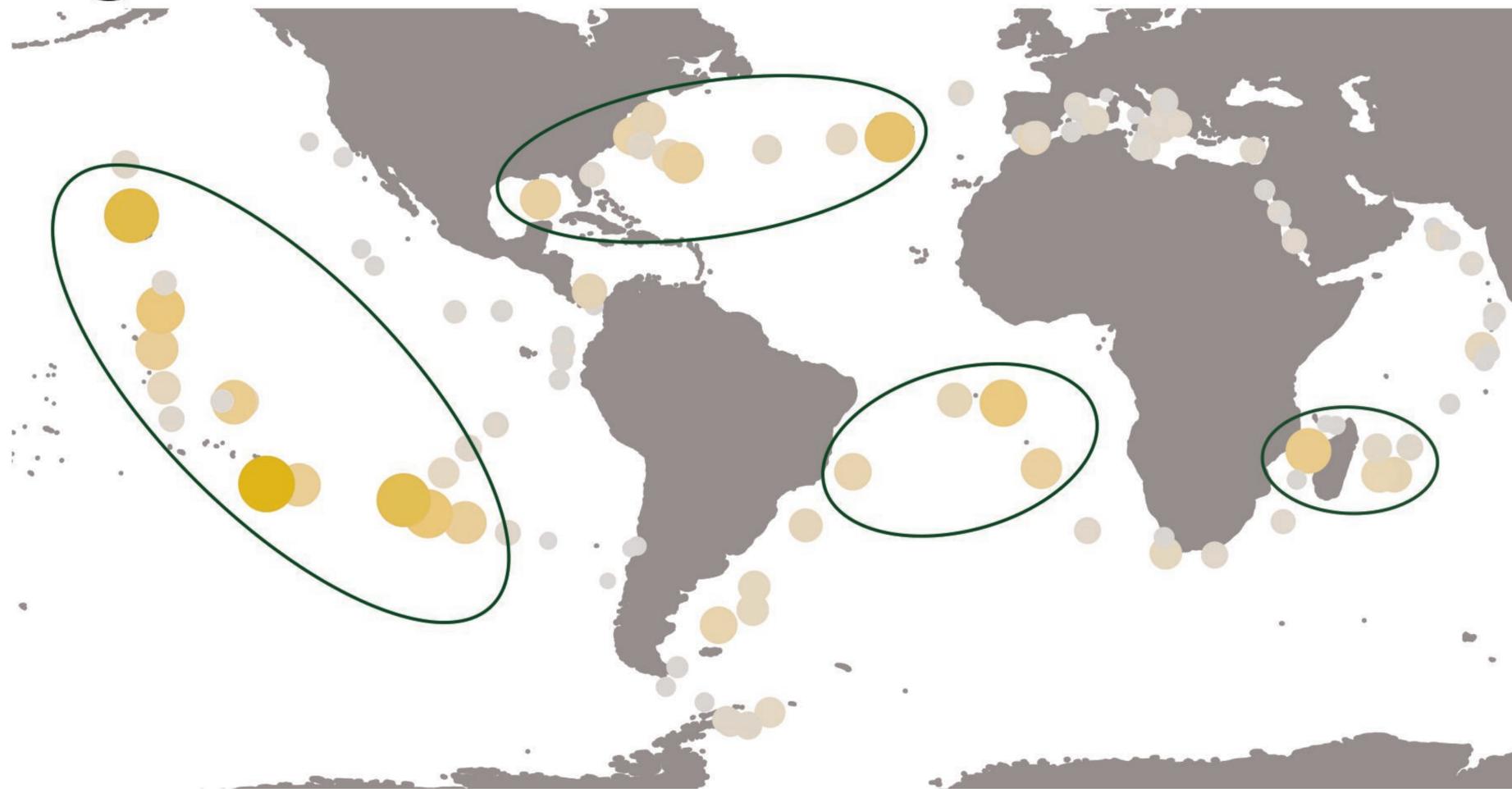


- Absent
- Present (>100 sequences)
- Questionable (<100 sequences)





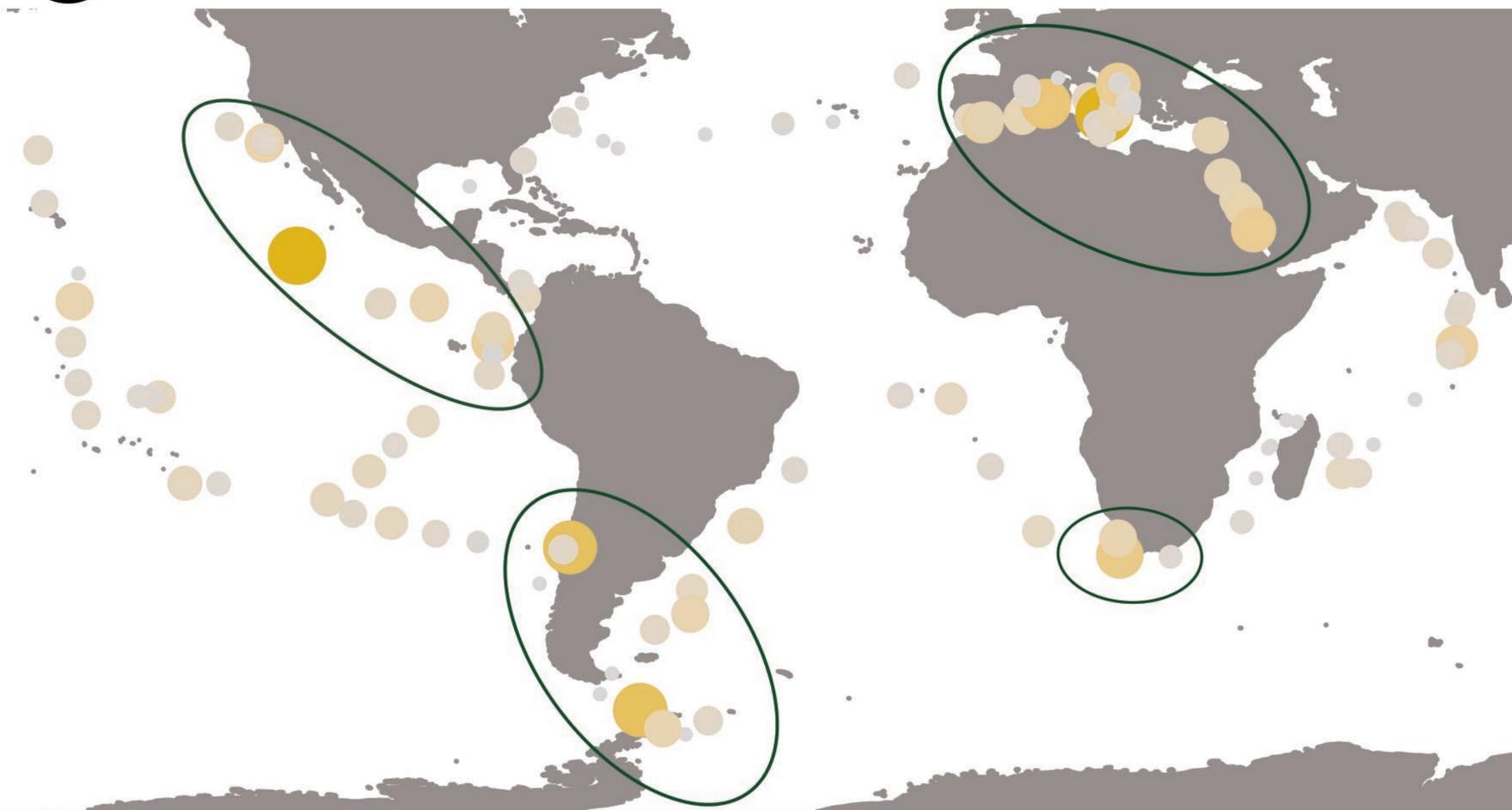
(A) Distribution of the *Siphonosphaera cyathina* barcodes



Sequence abundance
(Hellinger transformed)



(B) Distribution of the Acanthrometridae F3 spp. barcodes



Sequence abundance
(Hellinger transformed)

