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**Accumulation of TDP-43 and α -actin in an amyotrophic lateral
sclerosis patient with the K17I *ANG* mutation**

Danielle Seilhean, Cécile Cazeneuve, Valérie Thuriès, Odile Russaouen, Stéphanie Millecamps,

François Salachas, Vincent Meininger, Eric LeGuern, Charles Duyckaerts.

D. Seilhean (corresponding author), C. Duyckaerts,

UPMC Université Paris 06, AP-HP, Assistance Publique Hôpitaux de Paris, Groupe Hospitalier Pitié-Salpêtrière, Département de Neuropathologie, INSERM UMR-S 546 (DS) and UMR-S 679 (CD), 47-83 boulevard de l'Hôpital, 75651 Paris cedex 13, France

Tel: 33-1-42 16 18 81; Fax: 33-1-42 16 18 99; Email: danielle.seilhean@psl.aphp.fr

C. Cazeneuve,

AP-HP, Assistance Publique Hôpitaux de Paris, Groupe Hospitalier Pitié-Salpêtrière, Département de Génétique et Cytogénétique, U.F. de Neurogénétique Moléculaire et Cellulaire, Paris, France

V. Thuriès,

AP-HP, Assistance Publique Hôpitaux de Paris, Groupe Hospitalier Pitié-Salpêtrière, Département de Neuropathologie, Paris, France

O. Russaouen,

AP-HP, Assistance Publique Hôpitaux de Paris, Groupe Hospitalier Pitié-Salpêtrière, Département de Génétique et Cytogénétique, U.F. de Neurogénétique Moléculaire et Cellulaire, Paris, France.

S. Millecamps,

Laboratoire de Génétique Moléculaire de la Neurotransmission et des Processus Neurodégénératifs, CNRS UMR 7091, UPMC Université Paris 06, Groupe Hospitalier Pitié-Salpêtrière, Paris, France.

F. Salachas,

AP-HP, Assistance Publique Hôpitaux de Paris, Groupe Hospitalier Pitié-Salpêtrière, Fédération de Neurologie, Paris, France.

V. Meininger,

UPMC Université Paris 06, AP-HP, Assistance Publique Hôpitaux de Paris, Groupe Hospitalier Pitié-Salpêtrière, Fédération de Neurologie, Paris, France.

E. LeGuern,

UPMC Université Paris 06, INSERM, UMR-S 679 Neurologie & Thérapeutique Expérimentale, AP-HP, Assistance Publique Hôpitaux de Paris, Groupe Hospitalier Pitié-Salpêtrière, Département de Génétique et Cytogénétique, U.F. de Neurogénétique Moléculaire et Cellulaire, Paris, France

Key words : Neuropathology, motor neuron disease, ALS, hippocampus, neuronal intranuclear inclusions, actin, angiogenin, actinin, TDP-43, steatosis

Abstract.

A K17I mutation in the *ANG* gene encoding angiogenin has been identified in a case that we previously published as ALS with neuronal intranuclear protein inclusions (Seilhean et al., *Acta Neuropathol.* 2004). These inclusions were immunoreactive for smooth muscle α -actin but not for angiogenin. Moreover, they were not labeled by anti-TDP-43 antibodies, while numerous cytoplasmic inclusions immunoreactive for ubiquitin, p62 and TDP-43 were detected in both oligodendrocytes and neurons in various regions of the central nervous system. In addition, the expression of smooth muscle α -actin was increased in the liver in which a severe steatosis was observed. This is the first neuropathological description of a case with an *ANG* mutation. Angiogenin is known to interact with actin. Like other proteins involved in ALS pathogenesis, such as senataxin, TDP-43 and FUS/TLS, it plays a role in RNA maturation.

Introduction

Amyotrophic lateral sclerosis (ALS) is a fatal neurodegenerative disorder causing progressive muscle weakness due to the loss of upper and lower motor neurons. About 10% of ALS cases are familial; of these 20% are related to mutations of the Cu/Zn superoxide dismutase-1 (SOD1) gene encoding an antioxidant enzyme [50]. Other proteins implicated in the pathogenesis of ALS are the vesicle-trafficking protein VAPB [37, 44], the guanine-nucleotide exchange factor alsin [67], senataxin that is putatively involved in DNA repair and RNA processing [6], the 43 kDa TAR-DNA binding protein (TARDBP or TDP-43) [8, 16, 30, 52, 57, 60, 68] and recently another DNA/RNA-binding protein FUS/TLS [35, 61]. A relationship between angiogenesis and the development of ALS had been previously suggested since chronic vascular insufficiency and, possibly, insufficient VEGF-dependent neuroprotection led to motor neuron degeneration (MND) in mice [45]. This hypothesis was confirmed by the identification of mutations of *ANG*, encoding angiogenin, an inducer of neovascularization, in some ALS patients [14, 17, 18].

We previously described the case of a 46 year old woman with a rapidly progressive form of ALS which required mechanical ventilation nine months after onset [53]. The patient had no known familial history of ALS. She died after a four year course. Neuropathology showed severe degeneration of the spinal cord, sparing only the posterior columns (Fig. 1a-b), associated with neuronal loss in the substantia nigra and neuronal intranuclear inclusions (NIIs) in hippocampal pyramidal cells [53]. The NIIs were immunoreactive to ubiquitin, promyelocytic leukemia gene product (PML), ataxin-3, and proteasome 19S. They were not labeled by antibodies directed against expanded polyglutamine segments, neurofilaments, tau, synuclein, cystatin C, and were

not stained by ethidium bromide [53]. These results suggested an intranuclear accumulation of an unknown protein. In addition this patient presented with severe liver steatosis [12].

The principal genes responsible for the adult onset form of ALS (*SOD1*, *ANG* and *TARDBP*, encoding SOD1, angiogenin and TDP-43 proteins, respectively) were systematically analyzed in this patient. A K17I mutation of *ANG* was detected. Angiogenin is a 14.1 kDa serum protein, mainly secreted by the liver in normal conditions, that belongs to the pancreatic ribonuclease A superfamily [15, 54, 63]. First identified as an angiogenic factor in tumor-cell conditioned medium [15], angiogenin has since been implicated in various functions, including development, cell survival, and RNA maturation. Angiogenin binds to actin on the surface of endothelial cells and is internalized and transported to the nucleolus [19, 25, 26]. Once in the nucleolus, angiogenin is able to act as a stress activated ribonuclease that cleaves tRNA and inhibits protein translation [66].

Reevaluation of this case at the histological as well as immunohistochemical level including immunolabeling of angiogenin, TDP-43, p62 and known angiogenin partners or ligands in the CNS and peripheral tissues raises new questions about the pathophysiology of the disease.

Methods

Case and controls

Human tissue samples were used after consents, according to French and European regulations. *Case* : Samples of liver and peroneal muscle were taken at autopsy. The brain was sampled as previously described [53]. Blocks of the left hippocampus, embedded in paraffin after short (12 days) fixation in formalin were also available. *Controls* : A normal brain, from a patient who died from pneumonia, without known neurological disease, was used as a control for

immunohistochemistry. Duration of the fixation was similar in the control case and in the patient. NIIs found in diseases related to a polyglutamine expansion, including Huntington disease (n=1) and spinocerebellar ataxia (SCA), (SCA1, n=1; SCA3, n=1; SCA7, n=2) were compared to the NIIs found in this case. A case with ALS unrelated to *ANG* mutation, and a case with Charcot-Marie-Tooth disease were used as controls for severe muscle atrophy related to denervation. Liver samples taken in six other cases with severe steatosis were used as controls for the changes found in the liver in this case. Causes of death were acute pancreatitis in two cases, AIDS related CMV encephalitis, lung carcinoma, alcohol encephalopathy, myocardial infarct.

Analysis of the ANG coding sequence by direct sequencing

The genetic analyses were performed on a stored blood sample that had been obtained after the patient formal consent for a genetic analysis. The coding region of the *SOD1* (5 exons), *TARDBP* (5 exons) and *ANG* (exon 1 and 2) genes were amplified in respectively 5, 6 and 3 fragments from genomic DNA, using primers containing home-made universal sequences at their 5' end (available on request), in addition to the sequence specific to the corresponding gene. The PCR products were sequenced using the universal primer pair. Forward and reverse sequence reactions were performed with the Big Dye Terminator Cycle Sequencing Ready Reaction Kit (PE Applied Biosystems®). Sequence products were run on an ABI 3730 automated sequencer (PE Applied Biosystems®). Data were analyzed with the Seqscape 2.5 software (Applied Biosystems®).

Immunohistochemistry

After deparaffinization, the slides were immunolabeled with the antibodies listed in table 1. For most antibodies, the immunostaining was performed by an automat (Benchmark® XT

Ventana® staining system), the slides being pre-treated by heat at 95°C in the CC1 buffer, a proprietary high pH (=8) retrieval buffer (Ventana Medical Systems®) (Table 1). For tau and ubiquitin immunohistochemistry, the slides were pre-treated by formic acid (FA, 80 %), for 5 to 10 minutes (Table 1). Three antibodies raised against angiogenin were tested (Table 1). For the angiogenin and p62 antibodies, the slides were pretreated by heat (450W) in a microwave oven during 2x10 min using the buffer best adapted to the pH found optimal for immunolabeling (Table 1).

The biotinylated secondary antibody was included in the detection kit (Ventana® Medical Systems Basic DAB Detection Kit 250-001®). The streptavidin-biotin-peroxidase complex was revealed by diamino-benzidine. A paraffin block of colic adenocarcinoma was used as positive control for angiogenin and fibulin-1, the former antibody labeling tumor cells [13] and the latter, elastic fibers [48]. Placenta was used as positive control for α -actinin 1, and normal striate muscle for α -actinin 2 (Fig. 5a).

Results

Direct sequencing of the coding sequence failed to detect any variants in both *SOD1* and *TARDBP* in our case. In contrast, the K17I mutation was identified in the *ANG* gene. This mutation has already been reported as pathogenic by Wu et al. (2007).

The main neuropathological findings have been described in the previous paper [53]. The anterior and lateral tracts of the spinal cord were severely degenerated (Fig. 1a, b). Additional sampling of the hippocampus showed focal cortical dysplasia in the left parahippocampal gyrus. Boundaries between cell layers were indistinct; the junction between deep cortical layers and white matter was ill defined. Dysmorphic neurons had abnormal orientation, surrounded by a

loose and fibrillary neuropil. Among them, ballooned cells had an enlarged round soma with eccentric nucleus (Fig. 2j).

The main findings of the immunohistochemistry experiments are summarized in table 2. A positive labeling was obtained in control adenocarcinoma with two of the tested angiogenin antibodies (polyclonal antibody and MANG-1 monoclonal antibody). Angiogenin was immunodetected in the choroid plexus of patient and control, as well as fibulin-1 (data not shown). Intravascular monocytes were angiogenin reactive in the normal control brain (data not shown). The MANG-1 angiogenin monoclonal antibody showed, in addition, granular intracytoplasmic immunoreactivity in neurons of the controls, in all regions examined, including anterior horns of the spinal cord (Fig. 1d). Intracytoplasmic labeling in the patient and the controls was similar (Fig. 2d) except for a weaker staining of degenerating spinal motor neurons (Fig. 1e). The MANG-1 angiogenin antibody also labeled reactive astrocytes (Fig. 1e, f). This finding was not specific to the *ANG* mutation case, as labeled astrocytes were occasionally found in control cases in regions with reactive astrogliosis. The atrophic spinal tracts were strongly labeled (Fig. 1c). MANG-1 immunohistochemistry did not detect any abnormal intracytoplasmic or intranuclear inclusion in neurons and glial cells. No signal was detected either in smooth muscle fibers or in endothelial cells.

The NIIs that were described in the previous paper, were found in the pyramidal neurons of the hippocampus (Fig. 2), and frontal cortex (medial frontal gyrus) (Fig. 3c), as well as in dystrophic neurons in the left temporal lobe (Fig. 2j). They were not immunolabeled by the antibodies raised against angiogenin but were immunoreactive to smA (smooth muscle alpha actin). No reactivity for smA was found in the NIIs of positive controls with polyglutamic inclusions. The NIIs were not immunoreactive to fibulin-1, α -actinin 1 and 2, desmin, FVIII,

CD34, vimentin and TDP-43 (Fig. 2). They were found to be immunoreactive to p62, as was expected from the previous finding of their ubiquitin positivity.

In addition, immunohistochemistry of p62 and TDP-43 revealed numerous cytoplasmic inclusions in neurons and glial cells in various regions of the CNS including hippocampus, frontal cortex, striatum, medulla and spinal cord (Fig. 3). These cytoplasmic inclusions were not immunoreactive to sma, angiogenin, fibulin-1, α -actinin 1 and 2.

Intraneuronal, α -synuclein negative Lewy-like inclusions were detected with TDP-43 and phospho-TDP-43 antibodies in the striatum, mesencephalic tegmentum and anterior horns (Fig 3f, Fig 4a-c). TDP-43 positive skein-like inclusions were observed in the anterior horn of the spinal cord and inferior olive (Fig. 3 g-h). A few ring-shape inclusions were detected in the granular cells of the dentate gyrus only with phospho-TDP43 immunohistochemistry (Fig. 4d).

The labeled glial cells had the morphologic features characteristic of oligodendrocytes. The shapes of the inclusions were reminiscent either of the cap-shaped glial cytoplasmic inclusions (GCI) observed in multiple system atrophy (MSA) (Fig 3 c) or of the coiled bodies described in progressive supranuclear palsy (PSP), corticobasal degeneration (CBD) and argyrophilic grain disease (Fig 3 b, d), but these inclusions, although ubiquitin positive, were tau and α -synuclein immunonegative. They were better visualized with the p62 and TDP-43 antibodies than with the phospho-TDP-43 antibody.

Major changes in immunoreactivity were also found in the peripheral organs of our case. The peroneal muscle was severely atrophied with a spectacular decrease of α -actinin-2 reactivity specifically in the Z disk (Fig. 5c) compared to both normal (Fig. 5a) and denervated muscle (Fig. 5b). By contrast, detection of sarcomeric α -actin was remarkably unchanged in *ANG* mutation (Fig. 5f), compared to normal (Fig. 5d) and denervated (Fig. 5e) controls.

In the patient's liver, angiogenin labeling was decreased (Fig. 6a) compared to control samples with severe steatosis (Fig. 6c), fixed for various durations. SmA positive filaments surrounded the liver lobules forming an interconnecting network (Fig. 6b) in the patient and in one of the 6 control cases (which had no neurological disease history and died with acute pancreatitis). SmA positive fibers were only found in the portal and centrilobular areas in control normal liver samples and in 5 out of the six cases with severe steatosis (Fig. 6d).

Discussion

We have found that motor neuron disease associated with the K17I mutation of the *ANG* gene is characterized by three types of inclusions : NIIs, cytoplasmic inclusions either neuronal or oligodendroglial.

NIIs : ubiquitin, p62, smA positive NIIs were detected in pyramidal neurons in the hippocampus and frontal cortex. The NIIs were not immunolabeled by 1C2 antibodies [53] as are those found in polyglutamine disease, or by TDP-43 antibodies as those found in other forms of ALS-dementia or in progranulin mutations. Immunoreactivity of the intranuclear inclusions to p62 is in accordance to their immunoreactivity to ubiquitin and proteasome 19S, that we had previously reported [53]. P62 immunoreactivity contrasting with TDP-43 negativity in the NIIs observed in our case, is reminiscent of the TDP-43 negative FTLD-U recently identified [29, 39, 49] which, however, were also p62 negative [49].

Angiogenin antibodies raised against full length human did not label either the NIIs nor the cytoplasmic inclusions observed in neurons and glial cells, whatever the duration of fixation. The epitope mapping of the *ANG* antibodies is not currently available and the possibility remains that a fragment of the protein, not recognized by these antibodies, accumulates in the NIIs.

Nevertheless, the monoclonal MANG-1 antibody immunostained the cytoplasm of the neurons in all regions examined, in patient and controls, as previously described in the normal spinal cord [64].

In our case, K17I *ANG* mutation was associated with a sequestration of smA in the nucleus of susceptible neurons in the frontal cortex and the hippocampus. Remarkably, these inclusions were not found in the regions where the most severe neuronal loss was observed such as the spinal cord or the substantia nigra. This suggests that NIIs are transient and occur much earlier than cell death or that they play a protective role – a role that has been documented for the NIIs observed in SCA models [2]. In many hereditary diseases, the accumulated protein is coded by the mutated gene. In the K17I *ANG* mutation, as in *PRGN* mutations the NIIs do not contain the abnormal protein [39]. In the *PRGN* mutations the protein is not translated because the mutant mRNAs decay rapidly [1, 40].

Cytoplasmic inclusions: Many p62 and TDP-43 positive intracytoplasmic inclusions were found in neurons and oligodendrocytes throughout the CNS. These inclusions were not labeled by antibodies raised against angiogenin or other related proteins including smA, α -actinins and fibulin-1. Their shape and distribution were clearly distinct from those encountered in sporadic ALS.

Lewy-like TDP-43 and phospho-TDP-43 *neuronal cytoplasmic inclusions* were found in the striatum, mesencephalic tegmentum and anterior horns. Neuronal granular cytoplasmic inclusions with absent nuclear TDP-43 labeling, similar as those observed in the hippocampus in our case, have been previously interpreted as “pre-inclusions” in FTL-DU [4, 9, 60]. The rarity of inclusions in the dentate gyrus, detected only by the phospho-TDP43 antibody, was remarkable in our case.

The *oligodendroglial cytoplasmic inclusions* were observed in all the samples of the CNS that we examined. Some TDP-43 oligodendroglial inclusions have previously been described in the medullary tegmentum and inferior olivary nucleus [10], as well as in the cerebral white matter in FTLN-MND [21]. They may be observed in various CNS regions in sporadic ALS cases particularly when associated to TDP-43 neuronal cytoplasmic inclusions in the dentate gyrus [43]. In the present case, they were found in numerous areas of the CNS, representing one of the predominant neuropathologic features. The presence of numerous oligodendroglial inclusions of various types is characteristic of several neurodegenerative diseases such as MSA [46], PSP, CBD, argyrophilic grain disease [3, 33], and more recently in white matter tauopathy with globular glial inclusions [34]. Absence of immunoreactivity of the glial inclusions to α -synuclein and tau in the present case ruled out an association with one of these diseases.

Angiogenin, which was first described as an angiogenic factor secreted by tumor cells, has several physiological roles. In culture, angiogenin promotes ribosomal RNA transcription, cellular proliferation, organization of endothelial cells into tube-like structures and supports endothelial cell migration [24, 28, 59]. A strong expression of the protein has been detected in the developing mouse nervous system as well as in post-mitotic motor neurons derived from P19 cells [58]. It is also able to modulate PI-3-kinase/Akt-1 and ERK1/2 that belong to signaling pathways involved in neuron survival [20]. Transfection of wild *ANG* but not of *ANG* K40I mutant, protects NSC34 cells against excitotoxic injury. Angiogenin treatment, even after the clinical onset of neurologic signs, increases lifespan and motor function in SOD1 mutant mice [31].

The dominant mode of inheritance of ALS related to angiogenin mutation favors a toxic gain of function. However, most *ANG* mutations identified in ALS patients are associated with

functional loss of angiogenic activity [7, 64]. Six of the seven ALS-linked mutations including K17I markedly decrease the ribonuclease activity of angiogenin [7]. In contrast with the mutations S28N, P112L [64] R31A and R31K [18] impairment of the nuclear translocation has not been observed with the K17I mutation [64].

Accumulation of actin due to the K17I *ANG* mutation is probably related to a disturbance of angiogenin-actin interaction. Actin is a ubiquitous molecule of the cytosol that plays a key role in cell structure, cell motility and contraction in both muscle and non muscle cells. Mammals produce six isoforms in a tissue specific fashion encoded by separated genes [62] and divided into three classes (α , β , γ) according to their isoelectric point. Alpha-actins are found in striate and smooth muscles whereas β and γ isoforms are prominent in non-muscle cells. Both exogenous and endogenous angiogenin is able to interact with various actin isoforms. Smooth muscle α -actin has been identified as the 42-kDa angiogenin receptor protein at the surface of endothelial cells [23, 26, 41]. Actin binding to exogenous angiogenin at the surface of the cell induces membrane actin polymerization [26]. This surface interaction also induces endocytosis of angiogenin before translocation to the nucleus [42]. Actin polymerization and angiogenin endocytosis are critical steps in the process of angiogenin-dependent neovascularization [26, 42]. In addition, angiogenin binding to α -actin promotes Akt-1 activation [31, 32]. Akt-1 belongs to the serine/threonine-specific protein kinase family and is involved in cellular survival pathways, by inhibiting apoptotic processes. It has been recently shown to play a key role in motor neuron protection *in vitro* and *in vivo* [31].

Endogenous angiogenin is able to interact with intracellular actin, a major component of the cytoskeleton by the mediation of cytosolic α -actinin-2 [27], which is a member of the spectrin/dystrophin superfamily [56]. Alpha-actinins form anti-parallel rod-shaped dimers that

bundle actin filaments in cytoskeleton frameworks [56]. In mammalian cells four α -actinin genes produce at least six different isoforms, with specific tissue type and expression profiles. In skeletal and cardiac muscle α -actinin-2 is localized in the Z disk and cross-links anti-parallel actin filaments coming from adjacent sarcomeres [56]. In the central nervous system, α -actinin-2 is a postsynaptic protein which colocalizes in dendritic spines with the NR1 and NR2B subunits of N-methyl-D-aspartate (NMDA) glutamate receptors [11, 65]. α -actinin-2 binds the receptors to the actin cytoskeleton the integrity of which is mandatory for their function [51]. In addition to neuronal degeneration, it is noteworthy that this patient presented with apparently asymptomatic cortical dysplasia in the left temporal lobe. Some dystrophic neurons contained intranuclear inclusions arguing for an effect of the mutation on neuronal migration.

Specific loss of α -actinin-2 in the striate muscle associated with Z disk disorganization is suggestive of a direct muscular toxicity of the mutation that could add its effect to the loss of motor neurons. The muscle deficiency may have played a role in the rapid course of the disease that led the patient to mechanical ventilation only a few months after onset.

Fatty liver disease observed in our patient was unusually severe compared to other ALS patients [12]. We do not exclude that steatosis was an incidental feature due to metabolic exogenous factors. Nevertheless, *ANG* mutation could influence angiogenesis and lipid metabolism in the liver. Angiogenesis plays a role in the modulation of adipogenesis [5] and angiogenin is one of the downregulated genes in the C57BL/6J mice strain characterized by a high level of hepatic triglycerides [38]. Liver is normally the site of the highest angiogenin gene expression in adult [63] although the intensity of labeling has been reported to be quite low in non tumoral liver [22]. It was particularly low in our patient compared to controls with steatosis. More striking was the pattern of expression of smA which was compatible with an increased

polymerization of α -actin. Hepatic sinusoids are normally deprived of vascular wall. They are limited by a layer of endothelial cells surrounded by stellate and Kupfer's cells, separated from hepatocytes by space of Disse. In our case there was formation of an additional actin layer located under the endothelial cells and organized in a branching network. It has been shown that some catalytically inactive angiogenin mutants were still able to induce actin polymerization [55]. Its hypothetical role in liver regeneration or steatosis remains to be explored. The liver could be another site of angiogenin dysfunction besides the CNS and muscle in K17I *ANG* mutations.

TARDBP or TDP-43 is a DNA and RNA-binding protein which regulates transcription and splicing. An abnormal form of the protein, hyperphosphorylated and truncated, has been found to be accumulated in the cytoplasm, and sometimes nucleus, of neurons and glial cells in many cases of sporadic ALS and in FTLD-U. TDP-43 immunoreactivity is absent from the nucleus of cells containing cytoplasmic inclusions suggesting that pathogenesis of these proteinopathies could be due to the loss of normal TDP-43 function in the nucleus [36]. In *PGRN* gene mutations, causing familial FTLD-U with TDP-43 intranuclear and intracytoplasmic aggregates, genetic variability of microRNAs (miRNA) that bind mRNAs, can significantly modulate the translation of the protein, increasing the risk to develop the disease [47]. Two other genes coding for proteins involved in RNA processing have been found to be mutated in familial ALS. One is *SETX* coding for senataxin, which belongs to the helicase family that are involved in DNA repair and production of RNA. The other is the *FUS/TLS* (*fusing in sarcoma/translated in liposarcoma*) gene coding for another DNA/RNA binding protein [35, 61]. Analysis of the brain and spinal cords of ALS patients carrying the *FUS/TLS* mutations showed intracytoplasmic neuronal aggregates reminiscent of TDP-43 aggregates [35, 61]. Detection of TDP-43 in the cytoplasm of neurons and glial cells in this case of *ANG* K17I mutation and recent discovery of

angiogenin role in tRNA cleavage and inhibition of protein translation [66] support the hypothesis that angiogenin-related ALS pathogenesis involves also a defect in RNA maturation.

However, in contrast to *TARDBP* and *FUS/TLS* mutations, it is not the mutated protein that accumulates in the K171 *ANG* mutation, but smA, probably a partner of angiogenin. Additional accumulation of TDP-43 in neurons and oligodendrocytes in cells different from those with smA positive NIIs suggest associated pathways of toxicity.

The present case, which is, to the best of our knowledge, the first neuropathology report of an *ANG* mutation, illustrates in several ways the toxicity related to this mutation, including alteration of neuronal and muscle fibers survival, and possible associated impairment of neuronal cell migration and lipid metabolism. Neuropathologically, the salient characteristics of the case were smA and ubiquitin positive NIIs, and TDP-43 positive oligodendroglial and neuronal cytoplasmic inclusions. The anti-smooth muscle α -actin antibody can be a useful tool for the neuropathological analysis of ALS. *ANG* mutations should be looked for in patients presenting with rapidly progressing ALS, as well as in patients with cortical dysplasia or unexplained severe steatosis.

It is important to recognize that RNA maturation related to angiogenin, TDP-43 and *FUS/TLS* plays a key role in the pathogenesis of ALS.

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Legends of the figures

Figure 1. Spinal cord pathology

a-c Low magnification of spinal cord sections stained with 2F11 anti-neurofilament antibody (a) and with Bodian silver impregnation coupled with Luxol fast blue (b), both showing severe degeneration of the anterior and lateral tracts. By contrast, MANG-1 angiogenin immunohistochemistry (c) strongly labels the degenerated tracts. Bars = 0,5 cm.

d-f. Angiogenin immunohistochemistry in the anterior horn of the spinal cord in a control (d) and in the patient (e, f). The anti-angiogenin MANG-1 monoclonal antibody stains the cytoplasm of normal motor neurons (d). In the patient, severe neuronal loss is associated with a decrease in angiogenin cytoplasmic labeling (e : arrow head). Reactive astrocytes are strongly labeled with the MANG-1 antibody (e, f : arrows). Bars = 10 μ m (d), 20 μ m (e, f).

Figure 2. Immunoreactivity pattern of hippocampal NIIs.

NII immunoreactivity for p62 (a) and smooth muscle actin (b). Absence of immunoreactivity for : α -actinin-2 (c : arrow), angiogenin (d), fibulin-1 (e : arrow), α -actinin-1 (f : arrow), β -actin (g), TDP-43 (h : arrow), sarcomeric α -actin (k : arrow) and desmin (l : arrow). Note normal cytoplasmic labeling of pyramidal neurons in the hippocampus with the MANG-1 monoclonal antibody, even in neurons containing a NII (d). Likewise, NIIs could be observed together with a normal TDP-43 nuclear labeling (h : arrow head). Diffuse granular cytoplasmic immunoreactivity (arrow), and loss of normal nuclear TDP-43 positivity (arrow head), without NII, in a pyramidal cell of the hippocampus (i). Bars = 10 μ m.

Cortical dysplasia in the left hippocampus (j): ubiquitin positive NII in a ballooned neuron. Bar = 40 μm .

Figure 3. Oligodendroglial and neuronal cytoplasmic inclusions.

a-f : Oligodendroglial inclusions. P62 immunoreactive oligodendroglial inclusions in the medulla oblongata (a), the hippocampus (b), and midfrontal gyrus (c : arrow). Note in c, a NII (arrow head) similar to those described in the hippocampus (Fig 2a). TDP-43 immunoreactive oligodendroglial inclusions in the hippocampus (d : arrow) and in the striatum (e, f : arrow). Note the absence of nuclear labeling in cells bearing cytoplasmic inclusions compared to normal nuclear labeling in neighboring oligodendrocytes (d, arrow head).

f-h Neuronal intracytoplasmic inclusions. In the striatum, “Lewy like” TDP-43 positive intraneuronal inclusions (f, arrow head). “Skein-like” TDP-43-positive intraneuronal inclusions in the spinal cord (g) and the inferior olive (h). Bars = 8 μm (b), 10 μm (a, c-e, g, h), and 40 μm (f) respectively.

Figure 4. pTDP43 immunoreactivity

Neuronal intracytoplasmic pTDP-43-positive inclusions in mesencephalon (a), striatum (b), anterior horns (c), dentate gyrus (d). Bars = 10 μm .

Figure 5. Patterns of immunoreactivity in the muscle

a-c : α -actinin-2; d-f : sarcomeric α -actin. Patterns of staining in normal muscle (a, d), in denervated muscle (b, e) and in *ANG* mutation (c, f). In *ANG* mutation, sparing of sarcomeric α -actin is noticeable, compared to the severe decrease of α -actinin-2. Bars = 100 μm .

Figure 6. Patterns of immunoreactivity in the liver

a-c : decreased angiogenin immunostaining in the liver of the patient with K17I *ANG* mutation (a) compared to a control with severe steatosis (c).

b-d : increased smA staining in the liver of the patient with K17I *ANG* mutation (b) compared to a control with severe steatosis (d). Bars = 100 μ m

Antigen	Poly/monoclonal	Producer	Immunogen	Clone	Pretreatment	Dilution	Incubation time
α -actin (smooth muscle)	Monoclonal (mouse)	Dako®	N-terminal of the α smooth cell actin	1A4	CC1® 8min at 95°C	1:500	20 min +4°C
α -actin (sarcomeric)	Monoclonal (mouse)	Dako®	α -skeletal and α -cardiac muscle actins	Alpha-Sr-1	CC1®	1:50	32 min +37°C
β -actin	Monoclonal (mouse)	Abcam®	N-terminal of the β isoform of actin	AC-15	CC1® 30min	1:5000	32 min +37°C
α -actinin (H-2)	Monoclonal (mouse)	Santa Cruz Biotechnology, Inc. ®	C-terminal amino acids 593-892 of human α -actinin-1	H-2	CC1® 30min at 95°C	1:500	28 min +37°C
α -actinin 2 (ACTN2)	Polyclonal (rabbit)	Lifespan biosciences®	Human α -actinin-2, skeletal muscle isoform	-	CC1® 30 min at 95°C	1:100	32 min +37°C
Angiogenin	Polyclonal (goat) IgG	R&D systems®	Full length recombinant human angiogenin	-	DakoCytomation® Buffer pH=9 microwave	1:20	Overnight +4°C
Angiogenin	Monoclonal (mouse) IgG	Sigma®	Full length recombinant human angiogenin	14017.7	DakoCytomation® Buffer pH=9 microwave	1:500	Overnight +4°C
Angiogenin	Monoclonal (mouse) IgM	AbD serotec ®	Full length recombinant human angiogenin	MAN G-1	Ventana ® Protease 1 Buffer pH=7.2 microwave	1:200	60 min +37°C
CD34	Monoclonal (mouse)	Dako®	Human endothelial venules	QBEend10	CC1® 30 min at 95°C	1:50	20 min +37°C
Desmin	Monoclonal (mouse)	Dako®	Desmin purified from human muscle	D33	CC1® 30 min at 95°C	1:50	20 min +37°C
Factor VIII	Polyclonal (rabbit)	Dako®	Human plasma	-	CC1®	1:400	20 min +37°C
Fibulin-1	Polyclonal (rabbit)	Atlas antibodies®	Recombinant protein epitope signature tag (PrEST)	-	CC1® 30 min at 95°C	1:50	32 min +37°C
Neurofilament	Monoclonal (mouse)	Dako®	70 kDa subunit of neurofilament (axons)	2F11	CC1®	1:2000	32 min +37°C
p62 lck ligand	Monoclonal (mouse)	BD Biosciences®	Human p62 lck ligand aa. 257-437	257-437	Citrate Buffer pH=6 microwave	1:1000	Overnight +4°C
α -Synuclein	Monoclonal (mouse)	Novocastra®	Full length recombinant alpha-synuclein	KM51	CC1® 30 min at 95°C	1:20	56 min +37°C

Phospho-TDP-43 pS409/410-2	Polyclonal (rabbit)	Cosmo Bio Co. Ltd®	Phosphorylated full length TDP-43 CMDSKS(p)S(p)G WGM,S(p): phosphoserine 409/410	-	CC1®	1 : 5000	60 min +37°C
TARDBP (TDP-43)	Polyclonal (rabbit)	Protein Tech Group®	Human recombinant protein	-	CC1® 60 min at 95°C	1:1000	80 min +20°C
Tau	Polyclonal (rabbit)	Dako®	C terminal of human tau protein expressed in E.coli	-	CC1® 8min at 95°C + FA 5 min	1:500	32 min +37°C
Ubiquitin	Polyclonal (rabbit)	Dako®	Ubiquitin isolated from cow erythrocytes	-	CC1® at 95°C 30 min + FA 10 min	1:500	32 min +4°C
Vimentin	Monoclonal (mouse)	Dako®	Purified vimentin from porcine eye lens	V9	CC1®	1:50	32 min +4°C

Table 1: Immunohistochemical methods.

CC1, a proprietary high pH (=8) buffer; **FA** : formic acid.

	CNS			Striated muscle	Liver
	Hippocampal and frontal NII	Neuronal intracytoplasmic inclusions	Oligodendroglial intracytoplasmic inclusions		
Angiogenin	0	0	0	0	-
Smooth muscle α -actin	+	0	0	0	++
Sarcomeric α -actin	0	0	0	+	0
β -actin	0	0	0	0	0
α -actinin-1	0	0	0	0	0
α -actinin-2	0	0	0	-	0
Fibulin-1	0	0	0	0	0
p-62	+	+	+	0	0
Phospho-TDP-43	0	+	+	0	0
TDP-43	0	+	+	0	0
Ubiquitin	+	+	+	0	0

Table 2 : Main immunohistochemistry findings in K171 ANG mutation.

0 = no significant change compared to normal; +/++ = positive signal of low (+) or high (++) intensity; - = abnormally low detection.

References

1. Baker M, Mackenzie I, Pickering-Brown S et al (2006) Mutations in progranulin cause tau-negative frontotemporal dementia linked to chromosome 17. *Nature* 442:916-919
2. Bowman A, Yoo S, Dantuma N, Zoghbi H (2005) Neuronal dysfunction in a polyglutamine disease model occurs in the absence of ubiquitin-proteasome system impairment and inversely correlates with the degree of nuclear inclusion formation. *Hum Mol Genet* 14:679-691
3. Braak H, Braak E (1989) Cortical and subcortical argyrophilic grains characterize a disease associated with adult onset dementia. *Neuropathol Appl Neurobiol* 15:13-26
4. Cairns N, Neumann M, Bigio E et al (2007) TDP-43 in familial and sporadic frontotemporal lobar degeneration with ubiquitin inclusions. *Am J Pathol* 171:227-240
5. Cao Y (2007) Angiogenesis modulates adipogenesis and obesity. *J Clin Invest* 117:2362-2368
6. Chen Y, Bennett C, Huynh H et al (2004) DNA/RNA helicase gene mutations in a form of juvenile amyotrophic lateral sclerosis (ALS4). *Am J Hum Genet* 74:1128-1135
7. Crabtree B, Thiagarajan N, Prior S et al (2007) Characterization of human angiogenin variants implicated in amyotrophic lateral sclerosis. *Biochemistry* 46:11810-11818
8. Daoud H, Valdmanis P, Kabashi E et al (2008) Contribution of TARDBP mutations to sporadic amyotrophic lateral sclerosis. *J Med Genet*: Epub ahead of print
9. Davidson Y, Kelley T, Mackenzie I et al (2007) Ubiquitinated pathological lesions in frontotemporal lobar degeneration contain the TAR DNA-binding protein, TDP-43. *Acta Neuropathol* 113:521-533
10. Dickson D, Josephs K, Amador-Ortiz C (2007) TDP-43 in differential diagnosis of motor neuron disorders. *Acta Neuropathol* 114:71-79
11. Dunah A, Wyszynski M, Martin D, Sheng M, Standaert D (2000) α -actinin-2 in rat striatum: localization and interaction with NMDA glutamate receptor subunits. *Brain Res Mol Brain Res* 79:77-87
12. Dupuis L, Corcia P, Fergani A et al (2008) Dyslipidemia is a protective factor in amyotrophic lateral sclerosis. *Neurology* 70:988-989
13. Etoh T, Shibuta K, Barnard G, Keitano S, Mori M (2000) Angiogenin expression in human colorectal cancer : the role of focal macrophage infiltration. *Clin Cancer Res* 6:3545-3551
14. Fernández-Santiago R, Hoenig S, Lichtner P et al (2009) Identification of novel Angiogenin (ANG) gene missense variants in German patients with amyotrophic lateral sclerosis. *J Neurol* 2009: Epub ahead of print
15. Fett J, Strydom D, Lobb R et al (1985) Isolation and characterization of angiogenin, an angiogenic protein from human carcinoma cells. *Biochemistry* 24:5480-5486
16. Gitcho M, Baloh R, Chakraverty S et al (2008) TDP-43 A315T mutation in familial motor neuron disease. *Ann Neurol* 63:535-538

17. Greenway M, Alexander M, Ennis S et al (2004) A novel candidate region for ALS on chromosome 14q11.2. *Neurology* 63:1936-1938
18. Greenway M, Andersen P, Russ C et al (2006) *ANG* mutations segregate with familial and "sporadic" amyotrophic lateral sclerosis. *Nat Genet* 30:411-413
19. Hatzi E, Badet J (1999) Expression of receptors for human angiogenin in vascular smooth muscle cells. *Eur J Biochem* 260:825-832
20. Hetman M, Gozdz A (2004) Role of extracellular signal regulated kinases 1 and 2 in neuronal survival. *Eur J Biochem* 271:2050-2055
21. Hiji M, Takahashi T, Fukuba H, Yamashita H, Kohriyama T, Matsumoto M (2008) White matter lesions in the brain with frontotemporal lobar degeneration with motor neuron disease: TDP-43-immunopositive inclusions co-localize with p62, but not ubiquitin. *Acta Neuropathol* 116:183-191
22. Hisai H, Kato J, Kobune M et al (2003) Increased expression of angiogenin in hepatocellular carcinoma in correlation with tumor vascularity. *Clin Cancer Res* 9:4852-4859
23. Hu G, Chang S, Riordan J, Vallee B (1991) An angiogenin-binding protein from endothelial cells. *Proc Natl Acad Sci USA* 88:2227-2231
24. Hu G, Riordan J, Vallee B (1994) Angiogenin promotes invasiveness of cultured endothelial cells by stimulation of cell-associated proteolytic activities. *Proc Natl Acad Sci USA* 91:12096-12100
25. Hu G, Riordan J, Vallee B (1997) A putative angiogenin receptor in angiogenin-responsive human endothelial cells. *Proc Natl Acad Sci U S A* 94:2204-2209
26. Hu G, Strydom D, Fett J, Riordan J, Vallee B (1993) Actin is a binding protein for angiogenin. *Proc Natl Acad Sci USA* 90:1217-1221
27. Hu H, Gao X, Sun Y, Zhou J, Yang M, Xu Z (2005) α -actinin-2, a cytoskeletal protein, binds to angiogenin. *Biochem Biophys Res Commun* 329:661-667
28. Jimi S, Ito K, Kohno K, Kuwano M, Itagaki Y, Ishikawa H (1995) Modulation by bovine angiogenin of tubular morphogenesis and expression of plasminogen activator in bovine endothelial cells. *Biochem Biophys Res Commun* 211:476-483
29. Josephs K, Lin W, Ahmed Z, Stroh D, Graff-Radford N, Dickson D (2008) Frontotemporal lobar degeneration with ubiquitin-positive, but TDP-43-negative inclusions. *Acta Neuropathol* 116:159-167
30. Kabashi E, Valdmanis P, Dion P et al (2008) TARDBP mutations in individuals with sporadic and familial amyotrophic lateral sclerosis. *Nat Genet* 40:572-574
31. Kieran D, Sebastia J, Greenway M et al (2008) Control of motoneuron survival by angiogenin. *J Neurosci* 28:14056-14061
32. Kim H, Kang D, Kim H, Kang S, Chang S (2007) Angiogenin-induced protein kinase B/Akt activation is necessary for angiogenesis but is independent of nuclear translocation of angiogenin in HUVE cells. *Biochem Biophys Res Commun* 352:509-513
33. Komori T (1999) Tau-positive glial inclusions in progressive supranuclear palsy, corticobasal degeneration and Pick's disease. *Brain Pathol* 9:663-679
34. Kovacs G, Majtenyi K, Spina S et al (2008) White matter tauopathy with globular glial inclusions: a distinct sporadic frontotemporal lobar degeneration. *J Neuropathol Exp Neurol* 67:963-975

35. Kwiatkowski T, Bosco D, Leclerc A et al (2009) Mutations in the FUS/TLS gene on chromosome 16 cause familial amyotrophic lateral sclerosis. *Science* 323:1205-1208
36. Lagier-Tourenne C, Cleveland D (2009) Rethinking ALS: the FUS about TDP-43. *Cell* 136:1001-1004
37. Landers J, Leclerc A, Shi L et al (2008) New VAPB deletion variant and exclusion of VAPB mutations in familial ALS. *Neurology* 70:1179-1185
38. Lin X, Yue P, Chen Z, Schonfeld G (2005) Hepatic triglyceride contents are genetically determined in mice: results of a strain survey. *Am J Physiol Gastrointest Liver Physiol* 288:G1179-G1189
39. Mackenzie I, Baker M, Pickering-Brown S et al (2006) The neuropathology of frontotemporal lobar degeneration caused by mutations in the progranulin gene. *Brain* 129:3081-3090
40. Maquat L (2004) Nonsense-mediated mRNA decay: splicing, translation and mRNP dynamics. *Nat Rev Mol Cell Biol* 5:89-99
41. Moroianu J, Fett J, Riordan J, Vallee B (1993) Actin is a surface component of calf pulmonary artery endothelial cells in culture. *Proc Natl Acad Sci USA* 90:3815-3819
42. Moroianu J, Riordan J (1994) Nuclear translocation of angiogenin in proliferating endothelial cells is essential to its angiogenic activity. *Proc Natl Acad Sci USA* 91:1677-1681
43. Nishihira Y, Tan C, Onodera O et al (2008) Sporadic amyotrophic lateral sclerosis: two pathological patterns shown by analysis of distribution of TDP-43-immunoreactive neuronal and glial cytoplasmic inclusions. *Acta Neuropathol* 116:169-182
44. Nishimura A, Mitne-Neto M, Silva H et al (2004) A mutation in the vesicle-trafficking protein VAPB causes late-onset spinal muscular atrophy and amyotrophic lateral sclerosis. *Am J Hum Genet* 75:822-831
45. Oosthuyse B, Moons L, Storkebaum E et al (2001) Deletion of the hypoxia-response element in the vascular endothelial growth factor promoter causes motor neuron degeneration. *Nat Genet* 28:131-138
46. Papp M, Kahn J, Lantos P (1989) Glial cytoplasmic inclusions in the CNS of patients with multiple system atrophy (striatonigral degeneration, olivopontocerebellar atrophy and Shy-Drager syndrome). *J Neurol Sci* 94:79-100
47. Rademakers R, Eriksen J, Baker M et al (2008) Common variation in the miR-659 binding-site of GRN is a major risk factor for TDP43-positive frontotemporal dementia. *Hum Mol Genet* 17:3631-3642
48. Roark E, Keene D, Haudenschild C, Godyna S, Little C, Argraves W (1995) The association of human fibulin-1 with elastic fibers: an immunohistological, ultrastructural, and RNA study. *J Histochem Cytochem* 43:401-411
49. Roeber S, Mackenzie I, Kretschmar H, Neumann M (2008) TDP-43-negative FTL-D is a significant new clinico-pathological subtype of FTL *Acta Neuropathol* 116:147-157
50. Rosen D, Siddique T, Patterson D et al (1993) Mutations in Cu/Zn superoxide dismutase gene are associated with familial amyotrophic lateral sclerosis. *Nature* 362:59-62
51. Rosenmund C, Westbrook G (1993) Calcium-induced actin depolymerization reduces NMDA channel activity. *Neuron* 10:805-814
52. Rutherford N, Zhang Y, Baker M et al (2008) Novel mutations in TARDBP (TDP-43) in patients with familial amyotrophic lateral sclerosis. *PLoS Genet* 4:e1000193

53. Seilhean D, Takahashi J, El Achimi K et al (2004) Amyotrophic lateral sclerosis with neuronal intranuclear protein inclusions. *Acta Neuropathol* 108:81-87
54. Shapiro R, Strydom D, Olson K, Vallee B (1987) Isolation of angiogenin from normal human plasma. *Biochemistry* 26:5141-5146
55. Shapiro R, Vallee B (1989) Site-directed mutagenesis of histidine-13 and histidine-114 of human angiogenin. Alanine derivatives inhibit angiogenin-induced angiogenesis. *Biochemistry* 28:7401-7408
56. Sjöblom B, Salmazo A, Djinovic-Carugo K (2008) α -actinin structure and regulation. *Cell Mol Life Sci* 65:2688-2701
57. Sreedharan J, Blair I, Tripathi V et al (2008) TDP-43 mutations in familial and sporadic amyotrophic lateral sclerosis. *Science* 319:1668-1672
58. Subramanian V, Feng Y (2007) A new role for angiogenin in neurite growth and pathfinding: implications for amyotrophic lateral sclerosis. *Hum Mol Genet* 16:1445-1453
59. Tsuji T, Sun Y, Kishimoto K et al (2005) Angiogenin is translocated to the nucleus of HeLa cells and is involved in ribosomal RNA transcription and cell proliferation. *Cancer Res* 65:1352-1360
60. Van Deerlin V, Leverenz J, Bekris L et al (2008) TARDBP mutations in amyotrophic lateral sclerosis with TDP-43 neuropathology: a genetic and histopathological analysis. *Lancet Neurol* 7:409-416
61. Vance C, Rogelj B, Hortobagyi T et al (2009) Mutations in FUS, an RNA processing protein, cause familial amyotrophic lateral sclerosis type 6. *Science* 323:1208-1211
62. Vandekerckhove J, Weber K (1978) At least six different actins are expressed in a higher mammal: an analysis based on the amino acid sequence of the amino-terminal tryptic peptide. *J Mol Biol* 126:783-802
63. Weiner H, Weiner L, Swain J (1987) Tissue distribution and developmental expression of the messenger RNA encoding angiogenin. *Science* 237:280-282
64. Wu D, Yu W, Kishikawa H et al (2007) Angiogenin loss-of-function mutations in amyotrophic lateral sclerosis. *Ann Neurol* 62:609-617
65. Wyszynski M, Lin J, Rao A et al (1997) Competitive binding of α -actinin and calmodulin to the NMDA receptor. *Nature* 385:439-442
66. Yamasaki S, Ivanov P, Hu G, Anderson P (2009) Angiogenin cleaves tRNA and promotes stress-induced translational repression. *J Cell Biol*
67. Yang Y, Hentati A, Deng H-X et al (2001) The gene encoding alsin, a protein with three guanine-nucleotide exchange factor domains, is mutated in a form of recessive amyotrophic lateral sclerosis. *Nat Genet* 29:160-165
68. Yokoseki A, Shiga A, Tan C et al (2008) TDP-43 mutation in familial amyotrophic lateral sclerosis. *Ann Neurol* 63:538-542