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Pigeon odor varies with experimental exposure to trace metal pollution

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Abstract

Trace metals are chemical pollutants that have well-known noxious effects on wildlife and that are current major environmental issues in urban habitats. Previous studies have demonstrated their negative (e.g. lead) or positive (e.g. zinc) effects on body condition, immunity and reproductive success. Because of their effects on condition, trace metals are likely to influence the production of condition-dependent ornaments. The last decade has revealed that bird odors, like mammal odors, can convey information on individual quality and might be used as secondary sexual ornaments. Here, we used solid-phase microextraction headspace sampling with gas chromatography - mass spectrometry to investigate whether plumage scent varied with experimental supplementation in lead and/or zinc in feral pigeons. Zinc supplementation (alone or in combination with lead) changed the proportion of several volatiles, including an increase in the proportion of hydroxy-esters. The production of these esters, that most likely originate from preen gland secretions, may be costly and might thus be reduced by stress induced by zinc deficiency. Although lead is known to negatively impact pigeon condition, it did not statistically affect feather scent, despite most of the volatiles that increased with zinc exposure tended to be decreased in lead-supplemented pigeons. Further studies should evaluate the functions of plumage volatiles to predict how trace metals can impact bird fitness.

Keywords: Scent; Zinc; Lead; Birds; Dove

Introduction

Trace metals (e.g. lead, iron, copper, zinc) are normally present in very low levels in the environment and small amounts are essential part of physiology in animals and plants (Powell 2000, Prasad 2009, Plum et al. 2010). For instance, zinc is essential in regulating homeostasis, immune responses, and oxidative stress (Stefanidou et al. 2006), and zinc deficiency has serious consequences that have been recognized for many years and that include oxidative damages, diabetes and congenital malformations (Fosmire 1990; Oteiza et al. 1995; Prasad 2003). However, excessive quantities of trace metals can be toxic. The large amount of trace metals that have been released in the environment by anthropogenic activities (Nriagu 1979) have well-known noxious effects on humans (Järup 2003; Tchounwou et al. 2012) and wildlife (Demayo et al. 1982; Goutte et al. 2014; Scheuhammer et al. 2007). The binding potential (Godwin 2001; Tainer et al. 1992) and the propensity of trace metals to catalyze oxidation reactions (Koivula and Eeva 2010) are responsible for immunosuppression (Chatelain et al. 2016c; Snoeijs et al. 2005; Snoeijs et al. 2004), oxidative stress (Berglund et al. 2007) and endocrine disruptive effects (Baos et al. 2006; Meillère et al. 2016). For instance, lead, whose widespread use has caused extensive environmental contamination (Nriagu 1996), is associated with impaired learning in herring gulls (*Larus argentatus*) (Burger and Gochfeld 2005), reduced sperm quality in red deer (*Cervus elaphus*) (Castellanos et al. 2015), and higher oxidative stress in Mozambique tilapias (*Oreochromis mossambicus*) (Kaya and Akbulut 2015).

Sexual ornaments are conspicuous phenotypic traits that can take various forms such as large antlers, bright plumage, or elaborated song, and that are displayed to attract mates. Because their expression can be influenced by genotype and environment, including various stress factors, such as food availability, extreme temperatures, noise and parasites (McGraw et al. 2002; Troïanowski et al. 2017; von Schantz et al. 1999), they often honestly reveal the quality of the bearer (Zahavi and Zahavi 1999). By inducing physiological stress, trace metals can also alter ornament expression, as shown for the plumage coloration and singing behaviour of birds (Chatelain et al. 2017; Giraudeau et al. 2015; Gorissen et al. 2005; Vallverdú-Coll et al. 2015). Odor cues are also subjected to physiological and environmental influences (Ferkin et al. 1997; Johnston and Bronson 1982; Penn and Potts 1998), and are major secondary sexual traits in a wide range of species (Blaustein 1981; Havlicek et al. 2005; Martín and López 2006; Wyatt 2014). However, the effects of environmental pollution on their expression are largely unknown.

In this study, we tested the hypothesis of the impact of trace metals exposure (zinc and lead) on plumage odor in feral pigeon (*Columbia livia*) by an experimental supplementation approach. Although birds have long been considered to rely primarily on visual and acoustic cues, the past decade has revealed that birds use olfaction in various contexts including mate choice or parental care (reviewed in Caro et al. 2015). Bird odors, like mammal odors, can vary with genotype (Leclaire et al. 2014b), sex-hormone levels (Whittaker et al. 2017) and diet (Thomas et al. 2010), and are mainly composed of alkanes, alkanols, esters and fatty acids (reviewed in Campagna et al. 2012) that may be energetically costly to produce. Using a similar experimental design, we have already described beneficial effects of zinc on body mass, immunity and female investment into eggs (Chatelain et al. 2016b; Chatelain et al. 2016c), and detrimental effects of lead on iridescent coloration, the maternal transfer of antibodies into eggs and nestling growth (Chatelain et al. 2016b; Chatelain et al. 2017). We predict therefore that if pigeon odor is condition-dependent, it should vary in opposite direction with experimental exposure to lead and zinc.

Material and methods

Capture and housing

From February 20 to March 27 2013, we captured 96 free-living adult feral pigeons in several locations in Paris. We immediately transferred them in 8 outdoor aviaries (2.20m x 2.20m) at the CEREEP field station (Centre d'Ecologie Expérimentale et Prédictive - Ecotron Ile-de-France, UMS 3194, Ecole Normal Supérieure, Saint-Pierre-lès-Nemours, France).

Metal supplementation

This study is part of a bigger study investigating the effect of zinc and lead on several life history traits of pigeons. Details on the protocol have thus been published elsewhere (Chatelain et al. 2016b; Chatelain et al. 2016c; Chatelain et al. 2017). Briefly, on April 8 2013, after an acclimation period, aviaries were divided into 4 metal treatments (2 aviaries per treatment). Birds were evenly distributed among aviaries according to their flock, sex and plumage eumelanin level (Chatelain et al. 2016b). Treatments were lead supplementation (tap water enriched with 1 ppm lead acetate), zinc supplementation (tap water enriched with 10 ppm zinc sulphate), lead and zinc supplementation (tap water enriched with 1 ppm lead acetate and 10 ppm zinc sulphate) and no supplementation, i.e. control (tap water with no metal added). We chose these concentrations based on the lead concentration measured in blood of urban birds (Roux and Marra 2007) and on the gastrointestinal absorption rate of lead in zebra finches (Dauwe et al. 2002). Zinc concentrations were approximated using the zinc/lead concentration ratio measured in the environment and in bird feathers (on average, zinc is 10 times more concentrated than lead; Azimi et al. 2005; Chatelain et al. 2014; Frantz et al. 2012). Drinking bowls and baths were filled every other day with treated water. The efficiency of our supplementation protocol to increase bird exposure to lead and zinc has been previously validated (Chatelain et al. 2016b).

Sample collection

After 20 weeks of treatment, a few feathers (~33 mg) were cut in the lower back region of 74 birds (37 males and 37 females), and stored in 4 ml PTFE-faced septum sealed glass vials at -80°C until analysis. Birds from five aviaries (two lead-supplemented aviaries, one zinc-supplemented aviary, one zinc-and-lead-supplemented aviary and one control aviary) were sampled on 27 August 2013, while birds from the remaining three aviaries (one zinc-supplemented, one zinc-and-lead supplemented and one control aviaries) were sampled on the next day. The 74 birds sampled included 20 control birds, 19 lead-supplemented birds, 17 zinc-supplemented birds and 18 zinc-and-lead-supplemented birds.

Chemical analyses

We extracted chemical compounds with solid phase microextraction (SPME) using a stableflex 50/30µm DVB (divinylbenzene) / CAR (carboxen) / PDMS (polydimethylsiloxan) fiber (Supelco, Sigma-Aldrich, Bellefonte, PA, USA). To clear headspace for SPME extraction, we packed feathers at the bottom of the vial, using a glass rod cleaned with dichloromethane and ethanol between samples. The sample was then placed for 20 min in a laboratory incubator maintained at 34°C. We chose 34°C to mimic the natural temperature of bird body surface during a Parisian summer. During the summer 2013, the average daily maximal temperature in Paris was 25.3°C (public data of Meteo France). At 25°C ambient temperature and low humidity, surface temperature of birds is 33.4°C (Chilgren and King 1973). The SPME fiber was then exposed to the headspace (without touching the feathers) for 25 minutes at 34°C, after which the fiber was retracted and the adsorbed chemicals were analyzed by gas chromatography-mass spectrometry (GC-MS). Only one SPME fiber was used in the study.

We analyzed the samples using a Shimadzu GCMS-QP2010plus (Shimadzu Scientific Instruments, Kyoto, Japan) equipped with an Optima®-5MS column (30 m x 0.25 mm x 0.25

μm, Macherey-Nagel, Düren, Germany) at the Plateforme d'Analyses Chimiques en Ecologie (PACE), technical facilities of the LABoratoire d'Excellence named Centre Méditerranéen de l'Environnement et de la Biodiversité (CEMEB). Helium was used as the carrier gas, at a constant 1ml/min flow rate. Fibers were exposed in a 200 °C injection port to desorb volatiles. We ran the following temperature protocol: 40° for 5 min, 40°C to 180°C ramped at 5°C/min, 180°C to 250°C ramped at 10°C/min, and finally held at 250°C for 1 min. The ion source and transfer line temperatures was 200°C and 250°C respectively. Mass were scanned from 38 - 350 m/z in electron ionization mode. Fibers were removed from the injection port after 10 min of desorption. We regularly interspersed blanks throughout the sample analyses to detect contamination. We ran an alkane standard solution (Alkanes standard solution, 04070, Sigma Aldrich®, Germany) under the same conditions for retention index calculation. The samples from 9 controls, 9 lead-supplemented pigeons, 10 zinc-supplemented birds and 11 zinc-and-lead-supplemented birds were ran in March 2015, while the other samples were ran in June 2015. Within each analysis time period (March vs. June 2015), samples were ran randomly. Consequently, there was no difference in date of analysis between zinc-supplemented and non-zinc supplemented samples or between lead-supplemented and non-lead supplemented samples (all $P > 0.15$). Ten samples had very low GCMS signal and were not included in the statistical analyses.

To confirm that the feather odor captured from captive pigeons was representative of that of wild pigeons, we compared the chromatograms with those of two samples of back feathers collected just after capture in February 2013. In addition, to shed light on the potential origin of plumage chemicals, we compared the feather profiles to those of preen secretion samples collected on two pigeons just after capture.

Chromatograms were analyzed blind to treatment using the Shimadzu GC-MS Solution software. Analytes were identified by comparison of mass spectral data using the

NIST library 2011 and cross-checking linear retention index of the spectral match found in the literature (PubChem database) with the calculated linear retention index of the analytes. Four monoterpenes were detected in pigeon feathers (α-Pinene, β-Pinene, p-Cymene and Bornyl acetate; Table 1). Most probably of plant origin (Charpentier et al. 2012), they were excluded from the analyses. One peak corresponded to a co-elution of several compounds including one contaminant (Table 1), and it was also not included in the analyses.

To ascertain that our GCMS analyses were repeatable, 5 samples were analyzed twice with a few days interval (range: 5 - 8 days). Results show that total chromatogram area was lower in the second analysis than in the first analysis (paired t-test: $t_4 = 5.32$, $P = 0.006$) but that the composition of the two samples from the same bird was repeatable (PERMANOVA analyses with bird identity as fixed factor: $F_{4,4} = 8.93$, $P = 0.001$).

Statistical analyses

Because, for most samples, we did not control for the amount of feathers collected, and because an internal standard was not included in the GCMS analyses, we did not rely on the absolute abundance of chromatogram peaks for the analyses of chemical composition; rather, we quantified each peak as the proportion of the peak size relative to the total area of the chromatogram. Proportions of chemical compounds were then transformed using isometric log ratio "ilr" (Brückner and Heethoff 2017). In addition, to limit the effect of the differences in the amount of feathers on odor composition, we restricted the odor composition analyses on the peaks that each represented on average more than 0.75% of the chromatogram (n = 19 peaks representing on average 91.0 % ± 0.3 % of the total chromatogram areas, Table 1). All of these peaks, except one, were present in all birds.

To determine if the chemical composition of plumage odor was related to lead and zinc exposition, we used a PERMANOVA with 5000 permutations i.e. nonparametric multivariate analysis of variance, (Adonis2 function, "vegan" package in R; Oksanen et al. 2013), based on Euclidean distance. The two-way interaction between lead and zinc, the bird sex and the analysis time period (March vs. June 2015) were also included in the model. The date of feather sampling was used as strata, thus restricting permutations within date of sampling. In Adonis, significant differences may be caused by different within-group dispersion instead of different mean values of the groups (Warton et al. 2012). For significant effects, we therefore tested difference in dispersion between groups (Betadisper function, "vegan" package; Oksanen et al. 2013).

To determine the chemical compounds that were preferentially associated with a significant fixed effect, we used correlation indices (function "multipatt" in the "indicspecies" package in R; De Caceres et al. 2016) with the point biserial coefficient of association on proportions of chemical compounds (De Cáceres and Legendre 2009). We restricted permutations within date of sampling. This method uses permutation tests to test significance of value, but does not correct for multiple tests. Multiple testing corrections are flawed because they assume multiple tests are independent, whereas multiple volatiles can show correlated effects (Fig. S1) because of the nature of the data (relative abundance vs. absolute abundance) and because the biosynthesis pathway of these volatiles may be related. Therefore, we also performed robust principal component analyses (RPCA) on the ilr-transformed dataset (Filzmoser et al. 2009) to identify the subset of compounds that varied with treatment. RPCA axis scores were compared between zinc-supplemented and non-zinc supplemented birds using linear mixed models with zinc treatment and analysis time period as fixed effects, and date of feather sampling as a random factor.

We performed all statistical analyses using the R statistical software (R Core Team 2017) and used a significance level set at $\alpha = 0.05$.

Results

We detected a total of 45 peaks in the 63 odor profiles of pigeons, that included alkanes, aldehydes, alcohols, ketones, and esters (Table 1 and Fig. 1). Only 6 (13%) of these compounds were also detected in preen secretions (Table 1). In contrast, 39 of them (87%) were also detected in the plumage of wild Parisian pigeons, suggesting that captivity had only slight influence on the identity of feather volatiles.

Chemical composition varied with zinc supplementation ($F_{1,61} = 4.27$, $P = 0.016$; Fig. 2 and Fig. S2), but not with lead supplementation ($F_{1,60} = 0.62$, $P = 0.89$) or the interaction between zinc and lead ($F_{1,59} = 1.22$, $P = 0.56$). It did not vary with sex ($F_{1,60} = 0.14$, $P = 0.99$), but varied with the period when the samples were analyzed ($F_{1,61} = 13.20$, $P < 0.001$). Multivariate dispersion in chemical composition between zinc-supplemented and non-zinc supplemented birds was similar ($F_{1,62} = 0.59$, $P = 0.45$). Correlation indices showed that the proportions of the two hydroxy esters and 3-Hexanone were increased in zinc supplemented birds, while the proportion of Nonanal was reduced (Fig. 3). Similarly, a robust PCA showed that the second principal component, which accounted for 26% of the variance and which was positively related to the two hydroxy esters and the two ketones, was higher in zinc-supplemented birds than in non-zinc supplemented birds ($\chi^2_1 = 7.14$, $P = 0.008$; Fig. 4).

Discussion

By analyzing feather volatiles of feral pigeons chronically exposed to zinc and/or lead, we provide evidence for an effect of zinc on pigeon odor composition. In particular, we found that zinc mainly increased the relative abundance of the two hydroxy esters and 3-Hexanone. All the compounds affected by zinc supplementation may potentially be produced endogenously by the bird. However, in contrast to several species, where most plumage chemicals originate from the preen gland (Leclaire et al. 2011; Mardon et al. 2011; Zhang et al. 2009), only the two hydroxy-esters were detected in pigeons' preen secretions. Although the sample size used to study the difference between preen secretion and feather volatiles was low (n = 2 birds sampled for preen secretions), this result is consistent with a study on common wood pigeons (*Columba palumbus*), where 93% of the whole plumage lipids do not originate from preen secretions (Jacob and Grimmer 1975).

The other compounds affected by zinc exposure were not detected in preen secretions, and may rather originate from other lipid-producing areas in the integument (Jacob and Ziswiler 1982) or from feather bacteria metabolism (Whittaker and Theis 2016). The role of bacteria in animal olfactory signaling has recently received renewed interests (Archie and Theis 2011; Douglas and Dobson 2013; Ezenwa and Williams 2014) and a few experimental studies in mice, elephants (*Loxodonta africana*), Indian mongooses (*Herpestes auropunctatus*) and European hoopoes (*Upupa epops*) have shown that some major odorants require commensal bacteria for their production (Goodwin et al. 2016; Gorman et al. 1974; Li et al. 2013; Martin-Vivaldi et al. 2010). Methyl-branched alkanes, methyl ketones, and aldehydes can all be released by bacterial degradation of fatty acids (Schulz and Dickschat 2007). Feathers of zinc-supplemented pigeons have lower bacterial load and different bacterial assemblage than feathers of birds non-exposed to zinc (Chatelain et al. 2016a), which may have led to differences in the composition of feather volatiles between birds. Further studies on the mechanisms of avian odor production are clearly needed to understand how environment can affect feather volatiles.

We found that zinc increased the proportion of preen-gland-related compounds, which might suggest higher production of preen oil in zinc-exposed birds. Several studies have suggested that birds in higher condition are able to produce higher quantity of preen oil. For instance, experimental infections and immune challenge reduce the size of the preen gland in house sparrows (*Passer domesticus*) and tawny owls (*Strix aluco*) (Moreno‐Rueda 2015; Pap et al. 2013; Piault et al. 2008). Zinc provided at low dose is essential for many biological processes (Stefanidou et al. 2006), and accordingly, zinc supplementation has various beneficial effects on pigeon condition (Chatelain et al. 2016b; Chatelain et al. 2016c). Therefore, zinc, through its positive effects on bird condition, may increased the ability of bird to produce wax esters and other costly volatiles. Although avian odors have reapeatedly been shown to vary with genetics (i.e., Krause et al. 2012; Leclaire et al. 2017) and physiological status (Douglas et al. 2008; Whittaker et al. 2018), a very few studies have shown the effect of environmental stressors on their expression (Bombail et al. 2018). For instance, infection influences the wax ester profiles of preen oil in song sparrows (*Melospiza melodia*) and the volatile profiles of feces in mallards (*Anas platyrhynchos*) (Grieves et al. 2018; Kimball et al. 2013). Our study, which shows that the volatile profiles of feather is related to zinc levels in the environment, is therefore consistent with the stress-induced effects on odors demonstrated in other birds.

In several avian species, odor plays a crucial role in social interactions (Balthazart and Schoffeniels 1979; Balthazart and Taziaux 2009; Hirao et al. 2009). Zinc-induced changes in pigeon scent might therefore influence the behavior of conspecifics, with, for instance, females avoiding the scent of zinc-deficient males. However, although pigeons are well-known to use their olfactory abilities for homing (reviewed in Wallraff 2005), it is unknown whether they use also olfaction for social communication. Feather chemicals may play also a role in social communication by affecting feather coloration or condition (Moreno‐Rueda 2017). For instance, in house finches (*Carpodacus mexicanus*), preen oil enhances the coloration of red feathers (López-Rull et al. 2010). In pigeons, changes in the coloration of melanic feathers have been induced by zinc supplementation (Chatelain et al. 2017), and might therefore be mediated by changes in the chemical composition of feather volatiles.

Feather chemicals do not only play a role in social communication; they can protect feathers from wear and act as chemical defenses against parasites (Douglas 2013; Hagelin and Jones 2007). For instance, in rock pigeons, preen oil has positive effects on plumage condition (Moyer et al. 2003). 3-Hexanone, which was increased in zinc-exposed birds, is a common urine component of mammals, including humans (Burger et al. 2006; delBarco‐Trillo et al. 2011; Raymer et al. 1986; Smith et al. 2008), and has been detected in the feathers of white Leghorn chickens (*Gallus gallus domesticus L.)* (Bernier et al. 2008) and king penguins (Gabirot et al. 2018). It belongs to the volatile methyl-ketone family, whose some compounds can provide protection against parasites (Borges et al. 2015; Kirillov et al. 2017). According to the parasite-repellent function of volatile ketones, volatile methyl-ketones of dark-eyed juncos (*Junco hyemalis*) are increased during the breeding season (Soini et al. 2007), when the need for protection against nest parasites is high. Zinc, by inducing higher production of hydroxy-esters and 3-hexanone, might thus enhance plumage condition and the anti-parasite properties of feathers, thus adding to the numerous positive effects of zinc on pigeon (Chatelain et al. 2016b; Chatelain et al. 2016c).

Lead is a toxic trace metal that can impair various biological processes, leading to illness and mortality (Papanikolaou et al. 2005). In feral pigeons, lead decreases iridescent coloration, the maternal transfer of antibodies into eggs, and nestling growth (Chatelain et al. 2016b; Chatelain et al. 2017). Despite these detrimental effects on pigeon condition, we did not detect effects of lead on pigeon scent. As our experimental exposure to lead underestimated the natural range (lead was 1.5 times less concentrated in the feathers of lead-supplemented pigeons than in those of wild feral pigeons; Chatelain et al. 2016b), we cannot exclude that, in urban areas, the high concentration of lead negatively impacts odor production in birds.

We did not find evidence for a sex signature in the feather odor of wild-caught pigeons kept in captivity. Similarly, no sex-differences in preen secretion composition have been detected in non-breeding rock pigeons (Montalti et al. 2005). Although sexual dimorphism in chemical cues from feathers or preen secretions has been reported in several bird species (Amo et al. 2012; Leclaire et al. 2011; Whittaker et al. 2010; Zhang et al. 2010), it is not ubiquitous. For instance, king penguins (*Aptenodites patagonicus*) and Cory's shearwaters (*Calonectris borealis*) seem to exhibit no sex-differences in scent (Gabirot et al. 2018; Gabirot et al. 2016). Pigeons display sexual polymorphism in iridescent coloration of neck feathers (Chatelain et al. 2017; Leclaire et al. 2014a), and might, therefore, use this other trait to discriminate between sexes. Alternatively, because the molecules detected using GC-MS depend on the analysis strategy (e.g., extraction protocol and GC column), the profile of chemical composition presented here is not exhaustive. For instance, carboxylic acids, which are better resolved using a polar column (Whelan et al. 2010), might encode sex information.

In conclusion, our study showed that exposure to a moderate amount of zinc induced changes in the feather volatile composition of feral pigeons. Future studies should now evaluate the role of these volatiles in social communication, protection against parasites or feather wear to further predict how trace metals, which are chemical pollutants of prime concern in cities, can impact bird fitness.

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Conflict of interest

The authors declare that they have no conflict of interest.

Compliance with Ethical Standards

All experiments were carried out in strict accordance with the recommendations of the "European Convention for the Protection of vertebrate Animals used for Experimental and Other Scientific Purposes" and were conducted under the authorizations of the "Ministère de l’éducation nationale, de l’enseignement supérieur et de la recherche” (authorization N_00093.02) and the "Direction Departementale des Services Veterinaires de Seine-et-Marne" (authorization N_ 77-05). The authors declare no conflict of interest.

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Figure captions

Figure 1: Representative chromatogram of feather volatiles in captive pigeons. Letters refer to compounds listed in table 1.

Figure 2: Partial unconstrained distance-based redundancy analysis plot showing separation between pigeons by zinc supplementation. The effect of the period when the samples were ran on the GCMS was partialled out to better show separation.

Figure 3: Mean ± SE proportion of the four chemical compounds that varied with zinc exposure in zinc-supplemented birds and non zinc-supplemented birds. P values were calculated using a mixed model with date of sampling as a random effect and analyses time period as covariate.

Figure 4: Robust principal component analysis plot showing separation between pigeons by zinc supplementation. Vectors representing chemical compounds with high loading on axis 2 are highlighted.

Table 1: Chemical compounds putatively identified in the feathers of feral pigeons. Compounds in bold are those that were > 0.75 % of the total chromatogram and that were used in statistical analyses.

Name	**Letter (see figure 1)**	**Retention index calculated**	**Detected in preen secretions**	**Detected on the plumage of wild Parisian pigeons**
Ketones				
Hexan-3-one	a	n.d.		x
6-methyl-5-Hepten-2-one	b	982		x
Aldehydes				
Hexanal	c	n.d.		x
Heptanal	d	896		x
Nonanal	e	1101	x	x
Decanal		1202	x	x
Alkanes				
Octane	f	n.d.		x
2, 4-dimethyl-Heptane		817		x
Nonane	g	899		x
Branched alkane #1	h	1060		x
Branched alkane #2	o	1065		x
Branched alkane #3	i	1104		x
Branched alkane #4		1109		x
Dodecane		1200		x

Branched alkane #5		1277		x
Branched alkane #6	j	1283		x
Branched alkane #7		1305		x
Branched alkane #8		1322		x
Branched alkane #9	k	1329		x
Branched alkane #10		1337		x
Branched alkane #11		1345		x
Branched alkane #12		1348		x
Tridecane		1300		x
Pentadecane		1500		x
Alcohols				
Pentanol		n.d.		x
2-butoxy-Ethanol		903		x
1-butoxy-2-Propanol		940		x
1-Octen-3-ol		978		x
2-ethyl-Hexan-1-ol	l	1028		x
2, 6-dimethyl-7-Octen-2-ol		1071		x
Esters				
Butanoic acid, methyl ester		n.d.		x
Propanoic acid, propyl ester		804		
Acetic acid, butyl ester	m	809		x
Hexanoic acid, methyl ester		922		x
Hydroxy ester #1	n	1357	x	x
Hydroxy ester #2	o	1376	x	x

Fatty acids				
Nonanoic acid		1273		x
Unknown				
Unknown #1	p	884		x
Unknown #2	q	910		
Unknown #4		1143		
Unknown #5		1232		
Unknown #6	r	1295		
Unknown #7		1307		x
Unknown #8		1316		
2, 2, 4, 6, 6-pentamethylheptane + hexanoic acid [a]	s	993	x	x

n.d.: not determined

[a]: These two compounds were eluted together.