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# Validation of an automatic reference region extraction for the quantification of [<sup>18</sup>F]DPA-714 in dynamic brain PET studies.

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## Running headline

Cluster analysis in [<sup>18</sup>F]-DPA714 brain PET studies

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## Abstract (196)

There is a great need for a non-invasive methodology enabling quantification of TSPO overexpression in PET clinical imaging. [ $^{18}\text{F}$ ]DPA-714 has emerged as a promising TSPO radiotracer as it is fluorinated, highly specific and returned reliable quantification using arterial input function (AIF). Cerebellum gray matter (CRB) was proposed as reference region for simplified quantification, however this method cannot be used when inflammation involves cerebellum. Here we adapted and validated a supervised clustering (SCA) for [ $^{18}\text{F}$ ]DPA-714 analysis.

Fourteen healthy subjects genotyped for TSPO underwent an [ $^{18}\text{F}$ ]DPA-714 PET, including ten with metabolite-corrected AIF and three for a test-retest assessment. Two-tissue-compartmental modelling provided  $\text{BP}_{\text{ND}}^{\text{AIF}}$  estimates that were compared to either  $\text{BP}_{\text{ND}}^{\text{LoganSCA}}$  or  $\text{BP}_{\text{ND}}^{\text{LoganCRB}}$  generated by Logan analysis (using SCA extracted reference region or CRB).

The SCA successfully extracted a reference region with similar reliability using classes that were defined using either all subjects, or separated into HAB and MAB subjects.  $\text{BP}_{\text{ND}}^{\text{AIF}}$ ,  $\text{BP}_{\text{ND}}^{\text{LoganSCA}}$  and  $\text{BP}_{\text{ND}}^{\text{LoganCRB}}$  were highly correlated (ICC of  $0.91 \pm 0.05$ ) but  $\text{BP}_{\text{ND}}^{\text{LoganSCA}}$  were  $\sim 26\%$  higher and less variable than  $\text{BP}_{\text{ND}}^{\text{LoganCRB}}$ . Reproducibility was good with 5% variability in the test-retest study.

The clustering technique for [ $^{18}\text{F}$ ]DPA-714 provides a simple, robust and reproducible technique that can be used for all neurological diseases.

## Key words

Inflammation, Microglia, Positron Emission Tomography, Brain Imaging and Clinical trials

## Introduction

Neuroinflammation is known to play a key role in the onset and progression of chronic neurodegenerative diseases such as Alzheimer's, Parkinson's and multiple sclerosis. Activated microglia is the main cellular component that characterizes neuroinflammation in these disorders (1). The availability of a reliable imaging tool aimed at quantifying activated microglia *in vivo* might help to identify specific targets for anti-inflammatory neuroprotective strategies. The translocator protein 18 kDa (TSPO), previously named peripheral benzodiazepine receptor, is only expressed at low levels in the resting brain by quiescent microglial cells, but becomes markedly overexpressed when microglia is activated (2). Although the biological role of TSPO remains poorly understood, with putative effects on cholesterol translocation, steroid synthesis, mitochondrial functioning, and cell apoptosis, it is a promising target for molecular imaging studies of neuroinflammation (3, 4). The [ $^{11}\text{C}$ ]PK11195 radioligand was the first TSPO tracer used in humans (3) and has been investigated in several pilot imaging studies of neuroinflammation in brain diseases (5). However this tracer suffers from serious limitations, such as poor brain penetration and low specific to nonspecific binding ratio in brain and plasma, resulting in a suboptimal signal to noise ratio. Furthermore, the use of  $^{11}\text{C}$  radioactive labeling restricts its use to centers with onsite cyclotrons. This has led to the development of second-generation TSPO radiotracers with improved specificity, affinity and signal-to-noise ratio (6). However, a drawback for the quantification of second-generation TSPO tracers is related to different binding sites (7) resulting in three affinity profiles, high-, mixed- and low-affinity binders (HAB, MAB and LAB), which is indicated by the rs6971 single polymorphism contained within the TSPO gene.

Several  $^{18}\text{F}$ -labelled second-generation compounds have been investigated in human and have showed favorable properties for imaging purposes in the brain of healthy subjects (8, 9). However, among those, [ $^{18}\text{F}$ ]FEDAA1106 shows slow kinetics, [ $^{18}\text{F}$ ]FEPPA is rapidly metabolized and [ $^{18}\text{F}$ ]PBR06 produces brain-penetrant radiolabeled metabolites that could bias the *in vivo* imaging and suitable quantification (10, 11).

[ $^{18}\text{F}$ ]DPA-714 provides an improved signal to noise ratio compared to [ $^{11}\text{C}$ ]PK11195 in several preclinical models (6, 12, 13) and was identified as one of the most promising TSPO-ligands for *in vivo* imaging. The compartmental modeling of [ $^{18}\text{F}$ ]DPA-714 in the brain of healthy controls has been recently investigated and revealed that the 2-TCM best

described the regional kinetics (14, 15). Depending on the analyzed region, the  $V_T$  (mL/cm<sup>3</sup>) estimate was about 50% higher in HABs compared to MABs, supporting the need to take the genetic affinity profile into account for the quantification of this tracer in human studies (15).

For all TSPO radiotracers, there are challenges that relate to the quantification of binding parameters (5, 16). Compartmental modeling using the arterial input function is classically considered as the gold standard quantification method, and was first employed for [<sup>11</sup>C]PK11195 studies using a two tissue compartment model with a  $K_1/k_2$  value coupled to be the same across the whole brain cortex (17).

However, arterial blood sampling is an invasive procedure and therefore a number of alternatives to the standard quantification have been recently proposed. In order to simplify [<sup>18</sup>F]DPA-714 quantification, a population-based input function (PBIF) was tested on healthy subjects using one or two arterial or venous samples and compared to the arterial input function (AIF) quantification (15). A very good agreement between AIF and arterial PBIF was found but the variability increased when late venous samples were used. In addition, this technique still required an invasive extraction of at least one arterial sample as well as the analysis of the blood metabolites.

Reference region methods have also been applied in several recent [<sup>11</sup>C]PK11195 clinical studies. A reference region should be characterized by low to no specific binding but should have the same amount of free plus non-specifically bound (non-displaceable) ligand as the target region. Accordingly, the cerebellum has been the main choice as a reference region for TSPO ligands, in particular in ischemic stroke (18) and Alzheimer's disease (19-21). However, these particular properties cannot be generalized to all neurological disorders potentially affecting the cerebellum, such as for multiple sclerosis (22) or HIV (23). Furthermore, data from different studies with [<sup>3</sup>H]PK11195, [<sup>11</sup>C]PBR28 and [<sup>11</sup>C]PK11195 (7, 24, 25) showed that displaceable binding had a non-negligible contribution to the distribution volume in the cerebellum. As a result, using cerebellum as reference region would lead to an underestimation of the true specific binding.

To avoid a potential bias related to the *a priori* selection of the anatomical reference region, methods for the automatic extraction of reference region in PET images have been developed over recent years, such as the supervised clustering algorithm (SCA) that was introduced by Turkheimer et al. in 2007 (SuperPk) for [<sup>11</sup>C]PK11195 (26). The SCA was

then validated and used in a [ $^{11}\text{C}$ ]PK11195 multi-center study (27). SCA is based on the creation of a set of predefined kinetic classes that guides the algorithm to select only voxels of no or low specific binding within the brain image.

Recently, a similar SCA approach was successfully applied to other tracers such as [ $^{11}\text{C}$ ]PIB (28) or [ $^{11}\text{C}$ ]TMSX (29). An unsupervised clustering approach was also implemented for [ $^{18}\text{F}$ ]DPA-714 in ALS (30). However, no predefined classes were identified and employed to supervise the kinetic shape of the time activity curves (TACs) in the clusters, and the results were not validated with AIF-derived-2-TCM.

In this study, we adapted and validated the SuperPK approach (26) to automatically extract the reference region from [ $^{18}\text{F}$ ]DPA-714 images (SuperDPA) in a group of genotyped healthy subjects. The quantification using each reference region method (SCA methods or the *a priori* anatomically defined cerebellar gray matter) was compared to the AIF-derived 2-TCM quantification described in our previous [ $^{18}\text{F}$ ]DPA-714 study in the same healthy human subjects (15). A further validation of the predefined classes for the SCA was performed in three ways. Firstly the influence of the genetic affinity profile was evaluated. Secondly, the optimal set of predefined classes for SuperDPA was investigated by simulating a set of classes for a range of binding conditions based on the 2-TCM kinetic parameters. Finally, the reproducibility of the SCA method was tested with 3 patients who underwent two [ $^{18}\text{F}$ ]DPA-714 scans.

## Material and Methods

### Subjects

Fourteen healthy volunteers (mean age  $46.8 \pm 15.7$  years, 6 females) were included from two clinical protocols conducted at the Service Hospitalier Frédéric Joliot and the ICM (NCT02305264 and NCT02319382). Written informed consent was obtained from all participants and the protocols were approved by the Medical Bioethics Committee of Ile de France Region and according to French legislation and European directives. All subjects were considered healthy according to their medical history record and physical examination. They all had a normal brain MRI.

Genomic DNA from blood samples was used to genotype the rs6971 polymorphism of the TSPO gene. The analysis revealed 7 high affinity binders (HAB) (mean age  $47.4 \pm 16.0$  years, 3 females) and 7 mixed affinity binders (MAB) (mean age  $46.3 \pm 16.7$  years, 3 females). No low affinity binders were found among the 14 subjects.

### Imaging protocol

Each participant underwent a T1-weighted (T1-w) magnetic resonance image and a [ $^{18}\text{F}$ ]DPA-714 PET acquisition. More details on the imaging protocols can be found in Lavis et al 2015 (15). Three subjects (2 HABs, 1 MAB) underwent a second PET acquisition (injected activity difference  $30.2 \pm 21.6$  MBq) after 7 to 9 days to study the reproducibility of the quantification method.

T1-w imaging was performed using a turbo spin echo sequence (TSE) (TE/TR= 3/6300 ms; alpha= 10, resolution= 0.92x0.92x0.93 mm) in a 1.5T Philips Achieva (Best, The Netherlands) scanner or a MPRAGE (TE/TI/TR=2.98/900/2300, alpha=9°, resolution=1x1x1.1 mm) in a 3T Siemens Trio scanner (Erlangen, Germany).

[ $^{18}\text{F}$ ]DPA-714 was prepared according to standard conditions (31). Subjects underwent [ $^{18}\text{F}$ ]DPA-714 PET scans in a high-resolution research tomograph (HRRT, Siemens, Knoxville, TN, USA). After a transmission scan using a  $^{137}\text{Cs}$  point source, a [ $^{18}\text{F}$ ]DPA-714 bolus was intravenously injected ( $198.4 \pm 22.9$  MBq). The dynamic PET acquisition in list mode lasted 90 min.

Ten subjects (7 HABs and 3 MABs) out of the 14 included in this study were the same as those described in Lavis et. al. (15) and had AIFs corrected for metabolites.

### **Image processing**

PET acquisitions were corrected for random attenuation and scattered coincidences, and reconstructed with the iterative ordered-subset expectation maximization (Ordinary Poisson [OP]-OSEM) 3D method (4 iterations using 16 subsets) including point spread function modeling within the reconstruction (using a 3D Gaussian kernel with 2 mm full-width at half-maximum). Dynamic data were binned into 27 frames (6x1 min, 7x2 min, 14x5 min). Reconstructed dynamic PET data were realigned for motion correction using the frame-to-reference image registration in PMOD 3.5 (PMOD Technologies Ltd., Zurich, CH).

T1-w images were segmented using Freesurfer 5.3 (<http://freesurfer.net>) and regions of interest (ROIs) were selected: thalamus, hippocampus, cerebellar gray matter, white matter and occipital, parietal, frontal and cingulate cortices. A whole brain mask was also extracted. T1-w images and ROIs were resampled into the PET space using a rigid registration in order to extract time-activity curves (TACs) from each ROI. Data from the left and right hemispheres were averaged. Kinetic modeling (2-TCM) was performed using the COMKAT library and Logan graphical analysis was proceeded using in-house software in Matlab (Math Works, Natick, MA, USA).

### **Implementation of the Supervised Clustering Algorithm for [ $^{18}\text{F}$ ]DPA-714 (SuperDPA)**

As in the SuperPK (26), the SuperDPA algorithm includes three steps to extract the reference region: i) a normalization procedure to scale each frame and make the acquisition comparable across subjects; ii) the creation of a set of predefined kinetic classes adapted to [ $^{18}\text{F}$ ]DPA-714 for use in the supervised algorithm; and iii) a supervised clustering algorithm that calculates the contribution of each kinetic class to the signal of each voxel (Figure 1A).

#### *i. Normalization:*

Each frame is normalized to reduce variability across frames and subjects. The normalization step is carried out during the definition of the kinetic classes, as well as for each image analyzed using the SCA method. To normalize the scan, the mean value of the activity in the brain in each frame is subtracted from the activity of each voxel in that



frame. The resulting “centered” values are divided by the standard deviation of the values over the whole brain at the same frame.

*ii. Definition of the kinetic classes of the SuperDPA*

Because [ $^{11}\text{C}$ ]PK11195 and [ $^{18}\text{F}$ ]DPA-714 share the same target, we used the same four classes presented by Yaqub et. al. 2012 (27): blood pool class, white matter class, no to low specific binding class (reference region) and high specific binding class. The blood pool class was extracted from the carotid arteries that were manually segmented using the PET image summed over the 2 first minutes of acquisition. The white matter class was defined using the Freesurfer automatic segmentation. In healthy controls (15), the level of binding in the gray matter displayed significant variation, with the highest specific uptake being detected in the thalamus. We therefore chose the thalamus to define the high specific binding class. We chose the cerebellar gray matter (CRB<sub>GM</sub>) to define the low specific binding class (reference region class) for the following reasons. First, we have previously shown that the lowest binding was found in the CRB<sub>GM</sub>; second, TSPO mRNA expression in cortical grey matter was found the lowest across brain as derived from Allen Brain Atlas (<http://human.brain-map.org>) (32) and finally, Turkheimer et al. observed most voxels of the extracted reference region in the cerebellar gray matter, using SuperPK (26).

In each subject, the TACs for each class were then extracted from normalized images (step i.) and averaged across the subjects to create the set of classes used for the SuperDPA.

*iii. Supervised classification*

In order to extract the reference region in each PET image, the kinetics of each voxel was projected onto the kinetic classes using a non-negative least squares (NNLS) algorithm, yielding the percent contribution of the each class in that voxel. As in SuperPK (26), only voxels with a probability higher than 90% of belonging to the reference region class were averaged to create the reference TAC.

**Validation of kinetic classes definition**

Based on SuperDPA methodology, we developed two different approaches to validate the predefined classes. The first approach assumed that the kinetics of [ $^{18}\text{F}$ ]DPA-714 depend on the TSPO affinity profile of each given subject. Classes were therefore independently created for HAB and MAB subjects in a so-called 'affinity study'. The second approach, a 'validation study', used simulated predefined classes with different characteristics to

investigate whether using classes with higher or lower specific binding would yield a more appropriate reference curve (Figure 1-B).

*i. Affinity study*

We investigated the impact of the genetic binding profile of the subjects on the shape of the classes and consequently on the extracted reference curves and estimated parameters. We therefore defined separate sets of classes based on the genetic affinity profiles: classes were created using either the 7 MABs, yielding the SuperDPA<sub>MAB</sub> method to process the MAB subjects or the 7 HABs subjects, yielding the SuperDPA<sub>HAB</sub> to process the HABs only. For comparison, we also generated a single set of classes from both HAB and MAB subjects pooled together (n=14), yielding the SuperDPA<sub>ALL</sub> applied on all subjects, whatever their genetic profile.

*ii. Validation of the classes*

We generated 2 types of simulated classes: 1) a simulated reference region class with less specific binding than in the initial CRB<sub>GM</sub> TAC and 2) a simulated high specific binding class with higher specific binding (from the thalamus TAC). To create the simulated classes, we modified the  $k_3$  parameter previously determined in Lavis et al. (15) using the 2-TCM for the 7 HAB subjects. To generate the simulated CRB<sub>GM</sub> class with lower specific binding for each subject, the initial  $k_3$  of the cerebellum gray matter was decreased by - 40% while all other parameters ( $K_1$ ,  $k_2$ ,  $k_4$ ) remained unchanged. Similarly, to generate a simulated higher specific binding class, the initial  $k_3$  was increased by +40%. Again, this was done for each subject. The resulting simulated TACs were used to create the classes for the reference region class and the high specific binding class using the method described earlier. For each simulation, the white matter and the blood classes were kept the same as in SuperDPA<sub>HAB</sub>. The simulations required all subjects to have AIF available for the estimation of the parameters and curve generation. To avoid the impact of the genetic affinity, only the HAB group (n=7) was used for these simulations.

In total, three SuperDPA methods were defined in this 'validation study': the SuperDPA<sub>C40</sub> method with a 40% reduction in the specific binding of the CRB<sub>GM</sub> (2) the SuperDPA<sub>T40</sub> with a 40% increase in the specific binding of the thalamus and (3) SuperDPA<sub>CT40</sub> method with both 40% decrease in the CRB<sub>GM</sub> and 40% increase in the thalamus.

### Binding parameter estimation

We previously showed (15) that the model that best described the kinetics of [<sup>18</sup>F]DPA-714 was the 2-TCM that was therefore used as ground truth in this study. For each ROI, BP<sub>ND</sub> was indirectly estimated using the AIF as in previous studies (26, 27) to obtain BP<sub>ND</sub><sup>AIF</sup> with:

$$BP_{ND}^{AIF} = (V_{TROI} - V_{TREF}) / V_{TREF}$$

where V<sub>TROI</sub> and V<sub>TREF</sub> are the total volumes of distribution computed with the 2-TCM for the ROI and the extracted reference region of each method, respectively.

In parallel, [<sup>18</sup>F]DPA-714 binding in all ROIs was estimated using the Logan reference graphical method for all SuperDPA approaches and CRB<sub>GM</sub> (33), as previously described (34). The Logan graphical analysis provided the distribution volume ratio (DVR<sup>LOGAN</sup>) that was converted into BP<sub>ND</sub><sup>LOGAN</sup> (BP<sub>ND</sub><sup>LOGAN</sup> = DVR<sup>LOGAN</sup> - 1).

BP<sub>ND</sub><sup>LOGAN</sup> parametric images were computed using the extracted reference region of the SuperDPA methods to show the distribution of specific binding across the brain for each method.

### Method evaluation and validation (Figure 1B)

#### *i. Affinity study*

BP<sub>ND</sub><sup>LOGAN</sup> values estimated from reference regions extracted using SuperDPA<sub>ALL</sub> and SuperDPA<sub>HAB/MAB</sub> were cross-compared for each subject (n=14) and compared to the BP<sub>ND</sub> from subjects with arterial sampling (n=10; BP<sub>ND</sub><sup>AIF</sup>). In addition, BP<sub>ND</sub><sup>LOGAN</sup> from these two methods were also correlated to the BP<sub>ND</sub><sup>LOGAN</sup> obtained for the CRB<sub>GM</sub>. Comparisons were assessed using the Pearson correlation, intra-class correlation (ICC) and relative error coefficients. Relative error was calculated based on absolute BP<sub>ND</sub><sup>LOGAN</sup> and BP<sub>ND</sub><sup>AIF</sup> values.

#### *ii. Validation of the classes*

BP<sub>ND</sub><sup>LOGAN</sup> provided by the SuperDPA<sub>HAB</sub> and the three sets optimized SuperDPA methods (superDPA<sub>C40</sub>, superDPA<sub>T40</sub> and superDPA<sub>TC40</sub>) were compared to BP<sub>ND</sub><sup>AIF</sup> from the 2-TCM-AIF-quantification. The agreement between the methods was measured for each ROI

using correlation coefficient, intra-class correlation (ICC) and the relative error between  $BP_{ND}^{LOGAN}$  and  $BP_{ND}^{AIF}$ .

*iii. Test-retest study*

The reproducibility of the  $SuperDPA_{ALL}$ ,  $SuperDPA_{HAB/MAB}$  and  $CRB_{GM}$  was evaluated using the three subjects who had two PET acquisitions within 7-9 days. Each reference region method was applied for both the first and the second PET exams independently, yielding  $BP_{ND}^{LOGAN}$  estimates. The relative error was computed between the  $BP_{ND}^{LOGAN}$  measurements from the scan and rescan images.

## **Statistics**

For each of the SCA methods investigated, the set of predefined classes were computed using a leave-one-out approach. Then, the reference region for the remaining subject was extracted using the classes created without that particular subject.

All statistics were computed using the freely distributed software R ([www.r-project.org](http://www.r-project.org)). Pearson correlation was used as correlation coefficient. The ICC measured the absolute agreement between the parameters derived when using the AIF and the reference region quantifications, according to the Shrout and Fleiss convention (35). Coefficient of variation was defined as the standard deviation of the  $BP_{ND}^{LOGAN}$  estimates of all group subjects divided by their mean. Pairwise comparisons were performed using Mann-Whitney or paired Mann-Whitney, when appropriate.  $BP_{ND}^{LOGAN}$  obtained from all methods were compared using a one-way paired ANOVA followed by post hoc Bonferroni corrected Fisher tests. Significance for all tests was set to  $p < 0.05$ .

## RESULTS

### *Reference region extraction using the SuperDPA*

Weighting maps were extracted from dynamic [ $^{18}\text{F}$ ]DPA-714 scans of HAB and MAB subjects, using the SuperDPA<sub>ALL</sub> and the SuperDPA<sub>HAB/MAB</sub> approaches. The coefficient maps for blood and white matter kinetic classes displayed expected distributions (*data not shown*). Maps representing high specific binding showed voxels mainly located in the thalamus region. Interestingly, the majority of voxels assigned to the reference region (probability higher than 90% of belonging to the reference region class) were located in the cerebellar gray matter and caudate and to some extent throughout the cortical gray matter. Representative probability maps from SuperDPA<sub>ALL</sub> and SuperDPA<sub>HAB</sub> are displayed in Figure 2A for one HAB subject.

Mean TACs from the reference region extracted using SuperDPA<sub>ALL</sub> and SuperDPA<sub>HAB/MAB</sub> together with TACs of the CRB<sub>GM</sub> and thalamus are shown in Figure 2C. TACs using both SuperDPA<sub>ALL</sub> and SuperDPA<sub>HAB/MAB</sub> showed a higher peak and a faster wash-out than that obtained in the cerebellar gray matter and reached the same residual activity at 90 minutes.

### *Quantification of PET data*

Representative BP<sub>ND</sub><sup>LOGAN</sup> parametric images obtained using the voxel-wise Logan method in one HAB subject (SuperDPA<sub>ALL</sub>) are shown in Figure 2B. As expected, the highest specific binding was found in the thalamus (15). BP<sub>ND</sub><sup>LOGAN</sup> obtained using the SuperDPA<sub>ALL</sub>, the SuperDPA<sub>HAB/MAB</sub> and the CRB<sub>GM</sub> are presented in Figure 3. Among studied ROIs, the highest BP<sub>ND</sub><sup>LOGAN</sup> estimates were found in the thalamus, the parietal, cingulate and frontal cortices. The lowest binding was seen in the white matter and hippocampus. To validate the SuperDPA approach, these BP<sub>ND</sub><sup>LOGAN</sup> estimates were compared to BP<sub>ND</sub><sup>AIF</sup>: correlation coefficients  $r$  and ICC values between both parameters were found higher than 0.92 in all ROIs except in the white matter ( $r=0.85$ ,  $\text{ICC}=0.86$ ) using SuperDPA<sub>ALL</sub> (Table 1).

### *Influence of the genetic affinity profiles*

For all methods (SuperDPA<sub>ALL</sub>, SuperDPA<sub>HAB/MAB</sub> and CRB<sub>GM</sub>), the average TACs of the extracted reference regions showed a slightly different shape between the HAB and the MAB groups (Figure 2C). Similar to the CRB<sub>GM</sub> TACs, both SuperDPA<sub>ALL</sub> and

SuperDPA<sub>HAB/MAB</sub> could identify TSPO affinity based on the TAC shapes, with a wider peak and a slower wash-out for the HAB group.

As shown in Figure 3, the mean BP<sub>ND</sub><sup>LOGAN</sup> value was higher in the HAB group compared to the MAB group in the four regions with high specific binding (thalamus and cingulate, frontal and parietal cortices), whichever reference region was used. The difference reached significance in the thalamus and the cingulate cortex when using the supervised clustering approaches only: respectively, +24.4% and +27.8% for SuperDPA<sub>ALL</sub> ( $p=0.04$ ) and +24.0% and +27.0% for SuperDPA<sub>HAB/MAB</sub> ( $p=0.02$ ). As illustrated in Figure 3, the BP<sub>ND</sub><sup>LOGAN</sup> using both SuperDPA methods showed much lower standard deviation compared to those obtained with CRB<sub>GM</sub> in all regions: the inter-subject variability measured by the coefficient of variation was 0.2 and 0.19 (for HABs and MABs, respectively) using SuperDPA<sub>HAB/MAB</sub> and 0.17 and 0.21 using SuperDPA<sub>ALL</sub> compared to 0.61 and 0.49 for the CRB<sub>GM</sub> reference region. The CRB<sub>GM</sub> approach did not allow the difference between HABs and MABs to become significant in any region.

For all methods, the BP<sub>ND</sub><sup>AIF</sup> and BP<sub>ND</sub><sup>LOGAN</sup> were found to be highly correlated (Table 1 and Figure 4):  $r$  values were above 0.9 for gray matter ROIs and above 0.85 for the white matter. ICC were also high, slightly higher for CRB<sub>GM</sub> (mean  $0.99 \pm 0.1$ ) followed by the SuperDPA<sub>ALL</sub> (mean  $0.91 \pm 0.05$ ) and SuperDPA<sub>HAB/MAB</sub> (mean  $0.84 \pm 0.05$ ).

According to affinity group, the mean BP<sub>ND</sub><sup>LOGAN</sup> estimates were significantly higher with SuperDPA<sub>ALL</sub> and with SuperDPA<sub>HAB/MAB</sub> compared to those obtained with CRB<sub>GM</sub> method in each ROI (averaged over the 7 ROIs : respectively  $+25.6 \pm 10.6\%$ ;  $p < 0.0002$  and  $+20.4 \pm 10.6\%$ ;  $p < 0.006$  compared to CRB<sub>GM</sub>). Furthermore, the mean BP<sub>ND</sub><sup>LOGAN</sup> estimates obtained with all SuperDPA methods overestimated the BP<sub>ND</sub><sup>AIF</sup> values ( $+15.2\%$  and  $+45.4\%$  respectively). Interestingly, this relative error was lower in ROIs characterized by a high specific binding such as the thalamus ( $-0.5\%$  and  $+6.4\%$ ), the frontal cortex ( $+7.2\%$  and  $+16.9\%$ ), and the cingulate cortex ( $7.5\%$  and  $15.9\%$ ) for SuperDPA<sub>ALL</sub> and SuperDPA<sub>HAB/MAB</sub>, respectively. Using the CRB<sub>GM</sub> as reference region resulted in a wider range of relative error from  $-106.3\%$  to  $+34.3\%$  with overall higher absolute values. The standard deviation of the relative error was lower for SuperDPA<sub>ALL</sub> ( $+38.7\%$ ) compared to other methods (Table 1).

### *Validation of the predefined classes*

Correlations and ICC between the  $BP_{ND}^{AIF}$  and the  $BP_{ND}^{LOGAN}$  estimates were very high for the three simulations tested (SuperDPA<sub>T40</sub>, SuperDPA<sub>CT40</sub>, SuperDPA<sub>C40</sub>- Table 2). The three simulations did not differ ( $p>0.2$ ) with SuperDPA<sub>HAB</sub> regarding the strength of their correlations with  $BP_{ND}^{AIF}$ . ICC between the  $BP_{ND}^{AIF}$  and  $BP_{ND}^{LOGAN}$  was slightly but not significantly lower for the SuperDPA<sub>C40</sub> ( $0.86\pm0.03$ ) when compared to ICC between  $BP_{ND}^{AIF}$  and  $BP_{ND}^{LOGAN}$  estimates from SuperDPA<sub>HAB</sub> ( $0.93\pm0.03$ ). Likewise, increasing the specific binding in the high binding class (SuperDPA<sub>T40</sub> and SuperDPA<sub>CT40</sub>) resulted in a similar ICC compared to the SuperDPA<sub>HAB</sub> ( $0.93\pm0.03$  and  $0.90\pm0.03$ ). Overall, very high agreement between  $BP_{ND}^{LOGAN}$  and  $BP_{ND}^{AIF}$  was found in all ROIs.

### *Test-retest study*

With respect to the scan-rescan reproducibility, the SuperDPA methods showed a lower average absolute variability compared to the CRB<sub>GM</sub> (CRB<sub>GM</sub>: 6.3%, SuperDPA<sub>HAB/MAB</sub>: and SuperDPA<sub>ALL</sub>: 4.5% - Figure 5) with similar standard deviation around 3.5 % for all methods. Interestingly, regions with high specific binding (thalamus, cingulate, frontal and parietal cortices) displayed better reproducibility measures when using SuperDPA methods compared to the CRB<sub>GM</sub>.

## Discussion

In this study, we adapted a supervised clustering approach to automatically extract reference regions within the brain from dynamic [ $^{18}\text{F}$ ]DPA-714 PET scans of healthy volunteers. This method provides a non-invasive quantification that has the potential to greatly simplify the use of this tracer in the clinical setting. The quantification performed using SuperDPA showed a very high agreement with the gold standard AIF quantification, and a very low inter-and intra-subject variability. We found that the creation of the set of predefined classes using both HAB and MAB subjects is adequate to extract a reference region, as it provided results with the same level of reliability than with the creation of two separate sets of predefined classes.

The SuperDPA method was found to be robust for identification of voxels belonging to the reference. This was further validated when the input regions to the SuperDPA classes were compared with optimized set of classes. Although these new classes were based on simulations from the 2-TCM quantification with lower or higher specific binding, they did not impact the correlation with  $\text{BP}_{\text{ND}}^{\text{AIF}}$  estimates.

We found that the SuperDPA-based-quantification had excellent agreement with the AIF-based-quantification. Interestingly, correlations of the SuperDPA method with AIF quantification and reproducibility measures were both better in ROIs characterized by high specific binding compared to regions with low binding. This same finding was observed by Collste et al. in healthy volunteers (36) and by Park et al. in multiple sclerosis patients compared to healthy controls (37). This suggests that the accuracy and reproducibility of the quantification using SuperDPA should be optimal in pathological conditions associated with a high level of TSPO expression.

The use of  $\text{CRB}_{\text{GM}}$  as pseudo reference region has previously provided consistent findings for differentiating healthy subjects and Alzheimer patients, at prodromal and dementia stages (17, 19). This method also showed a strong correlation with AIF quantification among healthy volunteers (38) as we found in this study. However, only the SuperDPA method should enable to discriminate subtle microglial activation in brain diseases where the location and amplitude of neuroinflammation is unpredictable and may include the cerebellum. The reference region extracted from the SuperDPA methods showed a faster wash-out kinetics than the  $\text{CRB}_{\text{GM}}$  and a lower  $V_{\text{T}}$  (mean  $V_{\text{T}} = 2.84 \pm 1.44$  and  $V_{\text{T}} = 3.41 \pm 2.15$ , respectively). This suggests that SuperDPA method could provide reference region TAC



closer to the shape of non-displaceable binding than that of the  $CRB_{GM}$ . Consequently, SuperDPA provided significantly higher  $BP_{ND}^{LOGAN}$  estimates than for  $CRB_{GM}$  ( $+25.6 \pm 10.6\%$  and  $+20.4 \pm 10.6\%$  for SuperDPA<sub>ALL</sub> and SuperDPA<sub>HAB/MAB</sub> compared to  $CRB_{GM}$ ).

Furthermore, inter-subject variability was found to be lower for SuperDPA than for  $CRB_{GM}$  (Figure 3). For both SuperDPA methods, the coefficient of variation of the  $BP_{ND}^{Logan}$  estimates in the HAB group was of about 33%, which is the same order of magnitude as the  $V_T$  estimates in HABs calculated in (15) using the AIF quantification in the same subjects. A similar variability was described by Kreisl et al. (39) for the  $[^{11}C]PBR28$  (29-36% for the  $V_T$  in HABs) and by Guo et al. for the  $[^{11}C]PBR111$  (~35% in HABs) (8). However, all these results were obtained using  $V_T$  parameter which is related to the global uptake, whereas the  $BP_{ND}$  parameter used in the present study should reflect the specific binding more accurately.

The test-retest analysis performed here ( $n=3$ ) provided a very low  $BP_{ND}$  coefficient of variation, less than 7%, for all quantification methods. This coefficient was lower for the SuperDPA approach (4.5%) than for the  $CRB_{GM}$  (6.3%), although not significantly. In comparison, poor reproducibility was found using the  $[^{11}C]DPA-713$  in healthy controls by Coughlin et al. (40) according to a regional  $V_T$  systematically increasing from test to retest. For the  $[^{11}C]PBR28$ , Collste et al. (36), showed a mean  $V_T$  absolute variability of 18 % in gray matter and of 48% in white matter in healthy subjects, while Park et al. (37) described a test-retest variability between 7 and 9% in healthy volunteers and MS patients. Using the SCA approach and  $[^{11}C]PK11195$ , this variation was found of 10.6% in four AD patients (26).

Another goal of this study was to take into account the impact of genetic polymorphism related to the affinity of  $[^{18}F]DPA-714$  to TSPO binding site on the SCA quantification. We confirmed here that the  $[^{18}F]DPA-714$  binding parameters estimated are affected by the affinity binding status of each subject, similarly to the other second generation TSPO tracers (8). In particular, the difference between HABs and MABs was particularly high in TSPO-rich regions:  $BP_{ND}^{LOGAN}$  was significantly higher of ~26% in thalamus and cingulate in HABs compared to MABs. This difference is comparable to previous studies: using the 2-TCM in the same subjects, Lavissee et al. found approximately +30% difference in  $[^{18}F]DPA-714$   $V_T$  in the thalamus (15). The  $[^{18}F]FEPPA$  tracer was shown to induce either a 27% difference in  $V_T$  (with the highest difference in TSPO-rich ROIs) (41) or a 15%

difference (non-significant) in white matter (42). For [ $^{11}\text{C}$ ]PBR28, Kreisl et al. (39) found SUV  $\sim 40\%$  greater in HAB subjects than in MAB subjects.

The ability of the SuperDPA method to detect the impact of the TSPO polymorphism on [ $^{18}\text{F}$ ]DPA-714 uptake finally confirms the superiority of this method over the CRB<sub>GM</sub> which failed to show a significant difference in binding parameters between HABs and MABs.

Compared to SuperDPA<sub>HAB/MAB</sub>, the BP<sub>ND</sub><sup>LOGAN</sup> values obtained with the SuperDPA<sub>ALL</sub> were found closer to those obtained with AIF with a lower relative error, indicating that SuperDPA<sub>ALL</sub> provides binding parameter estimation that is as accurate as SuperDPA<sub>HAB/MAB</sub>, if not even better. Therefore, the same supervised clustering procedure can be applied for reference voxels extraction in HAB and MAB subjects. Running SCA does not need the prerequisite of TSPO polymorphism knowledge but we confirm here that TSPO genotype correction is required for cross-sectional comparisons.

One limitation of the study is that our method was only applied to a healthy group of volunteers. However, healthy subjects have a low expression of TSPO throughout brain regions and the quantification is expected to be more challenging than in patients where higher levels of TSPO expression are observed and therefore statistical differences should be more easily detected. Simulating pathological condition here by increasing the specific binding in the corresponding class (SuperDPAT<sub>40</sub>) did not change the extracted reference region and resulting binding parameters. This suggests that our method, based on kinetic classes defined in a population of healthy volunteers, can be applied in patients to accurately detect and quantify neuroinflammation.

Another possible confounding factor that remains to be assessed for [ $^{18}\text{F}$ ]DPA-714 quantification is the influence of endothelial binding on SCA-based estimates. It has been suggested that the endothelial cells express non negligible TSPO for binding with [ $^{18}\text{F}$ ]DPA-714. The introduction of an extra irreversible compartment representing this endothelial cell binding on blood brain barrier in the model (2-TCM-1K) has been showed to enhance the estimation accuracy of the [ $^{11}\text{C}$ ]PBR28 binding parameters(16, 43, 44). In this study, the validation of the SCA-based parameter estimation methods was done using estimates from our previous quantification study of [ $^{18}\text{F}$ ]DPA-714 which describes the 2-TCM as an appropriate model. The question of whether it is necessary to include endothelial binding into the AIF-based quantification for [ $^{18}\text{F}$ ]DPA-714 and whether it needs to be accounted for in simplified modeling methods as for [ $^{11}\text{C}$ ]PK11195 (27, 45) is still under investigation.

## Conclusion

In this study, we adapted a supervised clustering approach to automatically extract reference regions within the brain from dynamic [ $^{18}\text{F}$ ]DPA-714 PET images of healthy volunteers. The creation of a unique set of predefined classes from both HAB and MAB subjects was shown to be adequate to extract a reference region as it provided results with the same level of reliability than when creating two separate sets of predefined classes. Thus, the knowledge of TSPO genetic status is not a prerequisite to run the SCA method but the 30% difference in  $\text{BP}_{\text{ND}}^{\text{LOGAN}}$  between HABs and MABs highlights the need to allocate subjects in their genetic affinity status to allow clinical interpretation. The SuperDPA method was validated through 1) simulated kinetic classes, 2) high correlation with results obtained using quantification based on invasive AIF and 3) by showing very low intra-subject variability. Regarding those criteria, the SCA method appeared to be more robust and accurate than the  $\text{CRB}_{\text{GM}}$  reference region method. Moreover, the SCA allows the possibility of a non-invasive quantification that should greatly simplify the use of this tracer in any neurological disorders. The very good reproducibility and low inter-subject variability suggest that this method might be able to detect subtle changes in TSPO binding on [ $^{18}\text{F}$ ]DPA-714 images of patients and to measure longitudinal changes in neuroinflammation.

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No other potential conflict of interest relevant to this article was reported.

## Disclosures / Conflicts of interest

Authors do not have any conflicts of interest with respect the publication of this paper.

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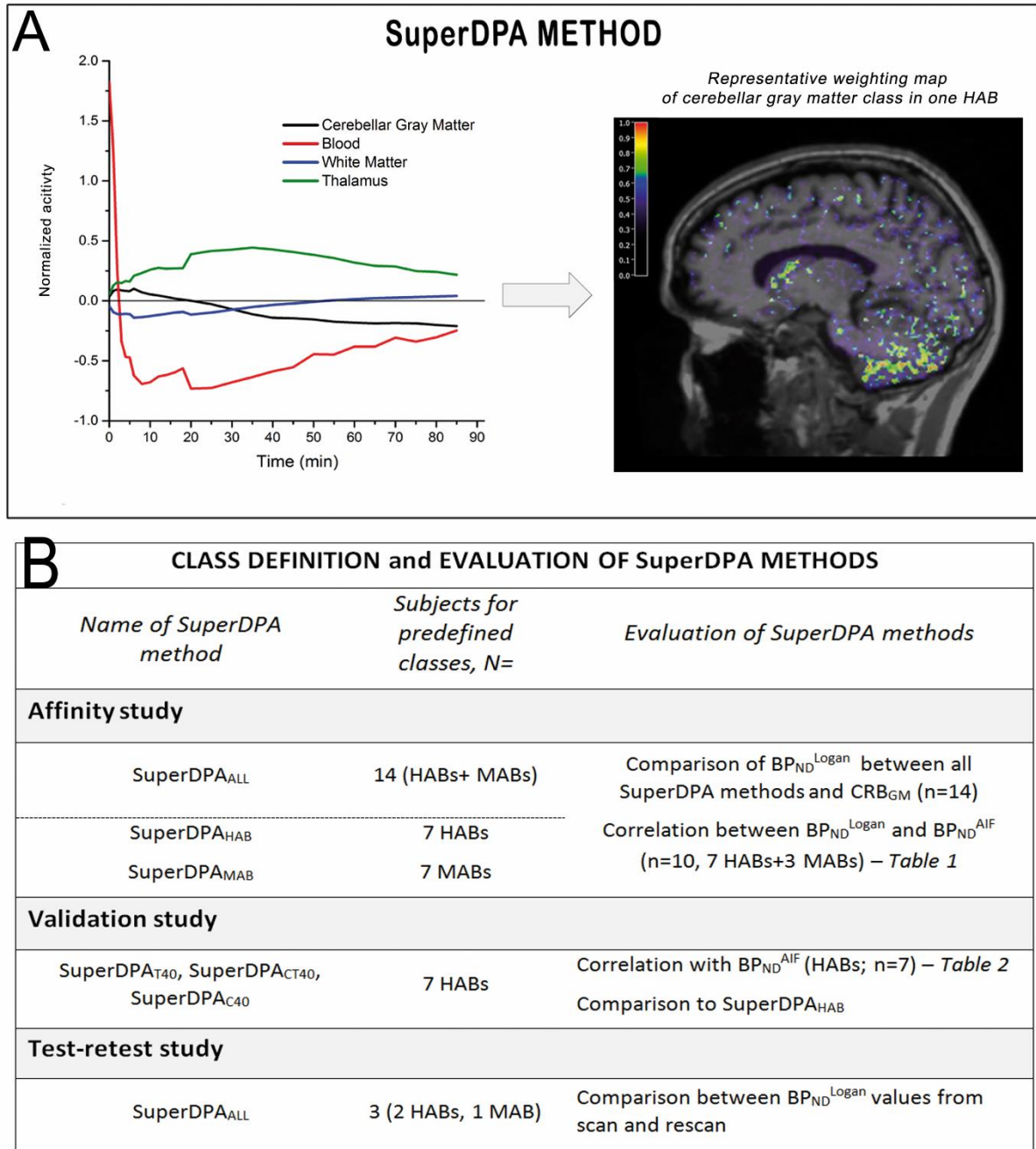
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## Figures :

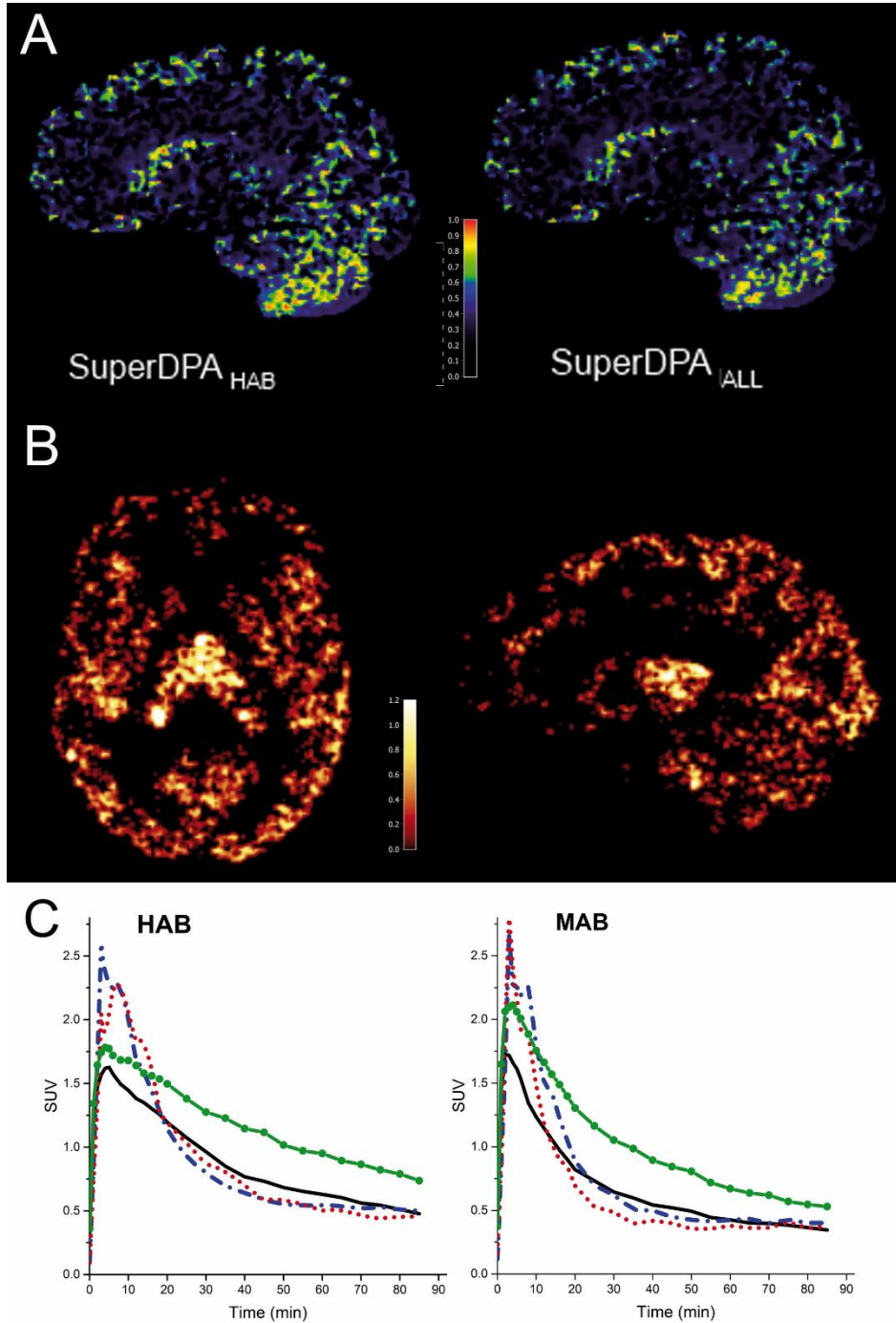
**Figure 1 : Design and evaluation of the SuperDPA method**



**Figure 2 : A:** Weighting maps of low/non-specific class (used for reference region selection) for one representative HAB subject using SuperDPA<sub>HAB</sub> (left) and SuperDPA<sub>ALL</sub> (right).

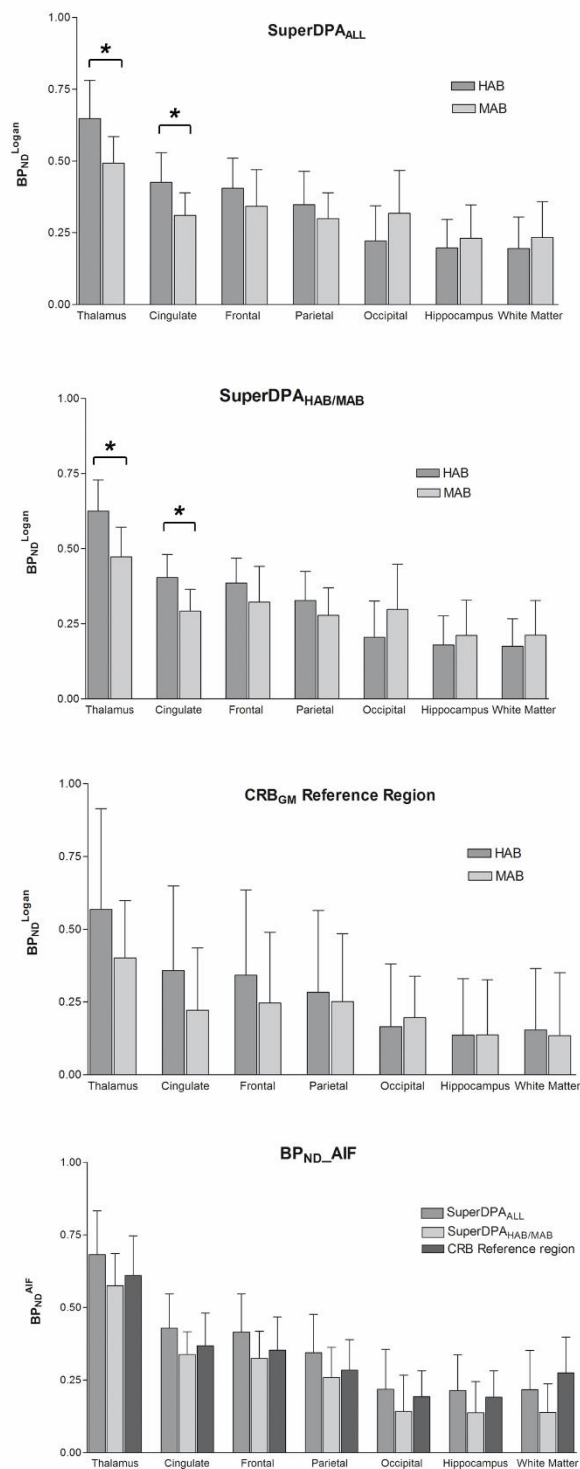
**B:** BP<sub>ND</sub><sup>LOGAN</sup> parametric map (axial and sagittal views) from one HAB subject using SuperDPA<sub>ALL</sub>.

**C:** Averaged TACs of thalamus (green line with solid circle), cerebellar gray matter (solid black line) and SCA-based-reference-regions obtained with SuperDPA<sub>ALL</sub> (dashed blue line) and SuperDPA<sub>HAB/MAB</sub> (dotted red line) methods. TACs are average SUV from HABs (left, n=7) and MABs (right, n=7).

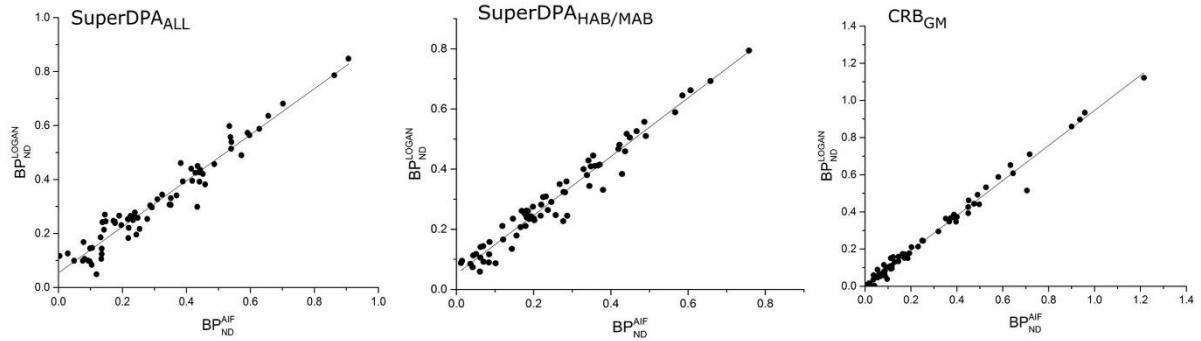




**Figure 3 :**  $BP_{ND}^{LOGAN}$  estimates of each ROI using SuperDPA<sub>ALL</sub> (top), SuperDPA<sub>HAB/MAB</sub> (middle) and the CRB<sub>GM</sub> (bottom) methods in HAB (n=7) and MAB (n=7) subjects. Error bars indicate the standard deviation. \* Significant difference,  $p < 0.05$

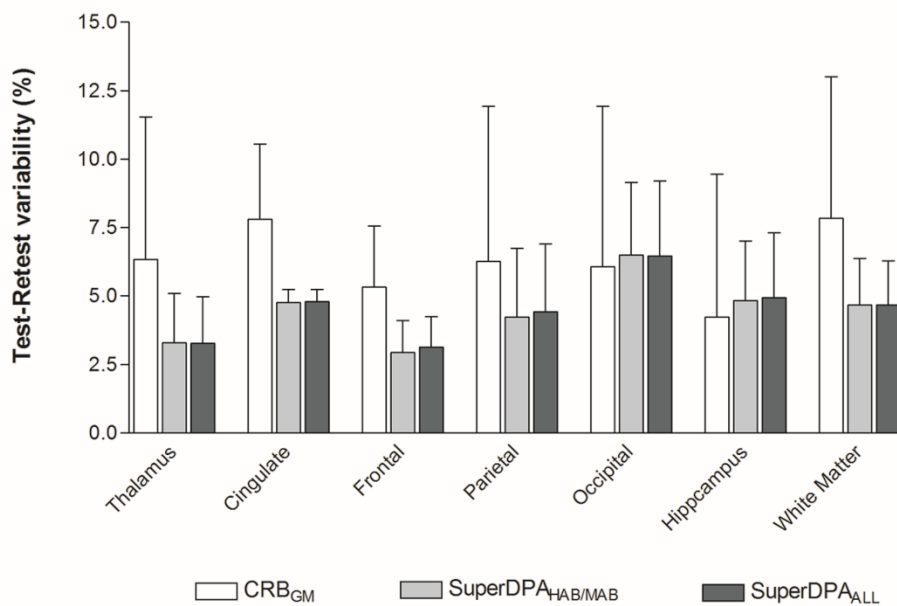


**Figure 4 :** Relationship between  $BP_{ND}$  estimates with the arterial input function analysis ( $BP_{ND}^{AIF}$ ) and reference input Logan graphical analysis ( $BP_{ND}^{LOGAN}$ ) using the SuperDPA<sub>ALL</sub> (left), SuperDPA<sub>HAB/MAB</sub> (middle) extracted reference region and the CRB<sub>GM</sub> (right)



**Figure 5 :** Evaluation of intrasubject-variability (test-retest) of the [ $^{18}F$ ]DPA-714 scan measures of each ROI using CRB<sub>GM</sub>, SuperDPA<sub>ALL</sub> and SuperDPA<sub>HAB/MAB</sub> methods. Test-retest variability is calculated as the absolute value of the difference as follows:

$$\text{Variability (\%)} = 100 \times |(DVR_{\text{Test}} - DVR_{\text{ReTest}})| / \text{Mean} (DVR_{\text{Test}}, DVR_{\text{ReTest}})$$



**Table 1**

**Table 1 :** BP<sub>ND</sub><sup>AIF</sup> and BP<sub>ND</sub><sup>Logan</sup> correlations using the SuperDPA methods

	CORRELATION			ICC			% RELATIVE ERROR			RELATIVE ERROR SD (%)			REGRESSION COEFF		
	ALL	HAB MAB	CRB <sub>GM</sub>	ALL	HAB MAB	CRB <sub>GM</sub>	ALL	HAB MAB	CRB <sub>GM</sub>	ALL	HAB MAB	CRB <sub>GM</sub>	ALL	HAB MAB	CRB <sub>GM</sub>
<b>Thalamus</b>	0.96	0.96	1.00	0.94	0.92	0.99	-0.53	6.37	-6.73	10.27	7.50	5.86	0.79	0.95	1.07
<b>Cingulate</b>	0.97	0.94	1.00	0.95	0.83	1.00	7.47	15.91	-5.67	19.28	13.69	8.97	0.81	0.97	1.13
<b>Frontal</b>	0.98	0.92	1.00	0.96	0.83	1.00	7.16	16.89	-3.19	19.61	15.08	5.72	0.73	0.97	0.89
<b>Parietal</b>	0.97	0.95	1.00	0.93	0.8	1.00	15.32	27.03	1.22	27.25	17.14	12.72	0.76	1.00	0.96
<b>Occipital</b>	0.90	0.96	1.00	0.88	0.83	1.00	21.95	112.52	106.30	33.86	191.23	294.73	0.79	1.00	0.96
<b>Hipp</b>	0.90	0.96	0.99	0.88	0.88	0.98	15.12	41.41	-6.75	42.20	34.66	47.91	0.69	0.88	0.90
<b>WM</b>	0.87	0.85	0.98	0.83	0.78	0.96	38.66	97.46	-34.32	118.40	170.58	37.28	0.60	0.79	0.78
<b>Mean</b>	0.94	0.93	0.99	0.91	0.84	0.99	15.17	45.37	23.46	38.70	64.27	59.03	0.74	0.94	0.96
<b>SD</b>	0.05	0.04	0.01	0.05	0.05	0.01	12.70	42.38	45.17	36.67	80.33	105.25	0.07	0.08	0.11

Correlation coefficients (r), ICC values, relative error (%), relative error standard deviation (%) and regression coefficients between BP<sub>ND</sub><sup>AIF</sup> and BP<sub>ND</sub><sup>Logan</sup>. The latter are estimated using SuperDPA<sub>ALL</sub>, SuperDPA<sub>HAB/MAB</sub> and CRB<sub>GM</sub> (n=10, 7 HABs+ 3 MABs). Relative error is calculated against BP<sub>ND</sub><sup>AIF</sup>.

**TABLE 2**  $BP_{ND}^{AIF}$  estimates and  $BP_{ND}^{LOGAN}$  correlations using simulated classes.

	<b>SuperDPA<sub>ALL</sub></b>	<b>T<sub>40</sub></b>	<b>C<sub>40</sub></b>	<b>CT<sub>40</sub></b>
Thalamus	0.94 (1.0)	0.95 (3.3)	0.93 (6.9)	0.96 (7.0)
Cingulate	0.93 (4.9)	0.92 (2.4)	0.94 (14.2)	0.97 (13.9)
Frontal	0.94 (6.0)	0.96 (2.8)	0.92 (14.8)	0.96 (14.5)
Parietal	0.95 (4.7)	0.96 (0.7)	0.95 (15.0)	0.97 (14.7)
Occipital	0.97 (13.3)	0.97 (0.2)	0.94 (23.3)	0.97 (21.6)
Hippocampus	0.97 (6.7)	0.93 (22.5)	0.93 (11.0)	0.95 (10.5)
White Matter	0.85 (10.7)	0.85 (13.1)	0.89 (12.7)	0.93 (11.5)

Correlation coefficients (and relative errors in %) between  $BP_{ND}^{AIF}$  estimates and  $BP_{ND}^{LOGAN}$  obtained with SuperDPA<sub>ALL</sub>, SuperDPA<sub>T40</sub>, SuperDPA<sub>C40</sub>, SuperDPA<sub>CT40</sub> (n= 7 HABs) in all studied ROIs.