Sexual isolation with and without ecological isolation in marine isopods Jaera albifrons and J. praehirsuta

Ambre Ribardière, Elsa Pabion, Jérôme Coudret, Claire Daguin-Thiébaut, Céline Houbin, Stéphane Loisel, Sébastien Henry, Thomas Broquet

HAL Id: hal-02349248

https://hal.sorbonne-universite.fr/hal-02349248

Submitted on 5 Nov 2019

Sexual isolation with and without ecological isolation in marine isopods *Jaera albifrons* and *J. praehirsuta*.

Ambre Ribardière[1], Elsa Pabion[1], Jérôme Coudret[1], Claire Daguin-Thiébaut[1], Céline Houbin[1,2],

Stéphane Loisel[1], Sébastien Henry[1,2], Thomas Broquet[1]

1- CNRS & Sorbonne Université, UMR 7144, Station Biologique de Roscoff, 29680 Roscoff, France.

2- CNRS & Sorbonne Université, FR2424, Station Biologique de Roscoff, 29680 Roscoff, France.

Corresponding author: Thomas Broquet, Station Biologique de Roscoff, 29680 Roscoff, France. E-mail: thomas.broquet@sb-roscoff.fr, phone number +33 2 98 29 23 51

Abstract

Sexual barriers associated with mate choice are often found to be associated with some level of ecological isolation between species. The independence and relative strength of sexual isolation are thus difficult to assess. Here we take advantage of a pair of marine isopod species (*Jaera*

20   *albifrons* and *J. praehirsuta*) that show sexual isolation and coexist in populations where they share the same microhabitat or not (i.e. without or with ecological isolation). We estimated the strength of sexual isolation between *J. albifrons* and *J. praehirsuta* using no-choice trials and a multiple-choice experimental population. We found that sexual isolation is strong in both the presence and absence of ecological isolation, but that it is asymmetric and fails to prevent interspecific gene flow entirely.

25   First-generation intrinsic post-zygotic barriers were low, and there was no sexual isolation within *J. praehirsuta* across habitats. The *J. albifrons* / *J. praehirsuta* species pair thus provides an example where the role of sexual isolation as a barrier to gene flow i) does not depend upon current ecological isolation, ii) seems to have evolved independently of local ecological conditions, but iii) is insufficient to complete speciation entirely on its own.

Introduction

30

Sexual isolation resulting from divergence in mating choice is common between closely related animal species, and the evolution of this type of barrier is considered to be a major component of speciation (Coyne & Orr, 2004). However, sexual barrier effects are often found in conjunction with some level of ecological isolation, raising the following questions (discussed e.g. in Ritchie, 2007;

35 Maan & Seehausen, 2011): What are the current relative contributions of sexual and ecological barrier effects in reproductive isolation between animal species? How often, if ever, has sexual isolation initiated speciation rather than evolving secondarily to ecological isolation? And may sexual isolation be an independent driver of speciation or is the evolution of sexual barriers necessarily linked with that of ecological isolation? Similar questions extend to other isolating barriers as well,

40 but we focus here on the relationship between sexual and ecological isolation, and more precisely on the last question, pertaining to the interdependence of these two isolating barriers.

Sexual isolation may be tightly linked with ecological isolation for several reasons (see Butlin & Smadja, 2018 for a discussion of coupling mechanisms and their importance in speciation). A direct form of coupling happens when some genes or traits affect ecological and sexual barriers at once

45 (reviewed in Servedio *et al.*, 2011). In *Heliconius* butterflies, for example, wing colour patterns affect both mimicry and mating signals, resulting in a strong combination of ecological and sexual barrier effects (Jiggins *et al.*, 2001). Along the same line, sexual and ecological barriers will coevolve when they involve overlapping metabolism networks, or sets of genes that are physically linked on the genome.

50 Furthermore, when there is no such intrinsic interdependence between barrier effects, other forms of coupling can happen if environmental conditions promote ecological isolation and simultaneously affect sexual isolation mechanisms. Such situations occur when the environment has an effect on male phenotypes, mortality costs associated with sexual display or choosiness, or the transmission of sexual signals (e.g host-dependent sexual signalling in *Enchenopa* treehoppers

55 McNett & Cocroft, 2008). All these cases may lead to the simultaneous evolution of sexual and

3

ecological isolation (reviewed in Maan & Seehausen, 2011; Nosil, 2012; Safran *et al.*, 2013; Boughman & Svanback, 2017; Servedio & Boughman, 2017). This list should even be extended if one considers not only the behavioural aspects of sexual isolation but also gametic isolation. The impact of ecological differentiation appears ubiquitous and one may wonder in what conditions can sexual

60   isolation evolve independently from ecological isolation.

Sexual selection mechanisms such as the Fisher-Lande process of coevolution between arbitrary male traits and female preferences can theoretically drive reproductive isolation largely independently of environmental heterogeneity and ecological barrier effects. This is also perhaps possible in some cases when good genes or compatible genes systems drive sexual isolation between

65   populations. An objective of empirical research is thus to explore how sexual isolation mechanisms are connected to ecological conditions and preferences, and evaluate to what extent sexual isolation may act as an independent driving force in speciation.

Hybrid zones provide good opportunities to investigate the interplay between different types of isolating barriers, and the most detailed information should come from situations where one can

70   investigate the role of sexual isolation in different ecological contexts. This is possible in mosaic or otherwise replicated hybrid zones (reviewed in Harrison & Larson, 2016) where hybridizing taxa meet repeatedly in different locations. In such cases, when sexual isolation was proven to be a critical barrier to gene flow it appeared to be nonetheless strongly dependent upon ecological conditions. This is either because sexual and ecological isolation involved the same traits, or sexual isolation

75   mechanisms were linked with ecological conditions. Habitat heterogeneity may then have driven ecological isolation and sexual isolation simultaneously (e.g. speciation in Gasterosteus sticklebacks and Pundamilia cichlids driven by adaptation of female perceptual sensitivity to ambient light combined with sexual selection on male colour, Boughman, 2001; 2002; Seehausen *et al.*, 2008). In these examples, sexual isolation is likely to break down when ecological conditions are homogenized

80   (Seehausen, 2009, see also Taylor et al., 2006). There are also many other cases of mosaic or replicated hybrid zones where sexual isolation is not fully understood but where ecological isolation

or spatial segregation, regardless of other isolating barriers, appeared fundamental to the

maintenance of reproductive isolation (e.g. field crickets, Harrison & Rand, 1989; marine mussels,

Bierne *et al.*, 2003; swordtail fish, Culumber *et al.*, 2011; river herrings, Hasselman *et al.*, 2014;

85   lampreys, Rougemont *et al.*, 2015).

There are comparatively few cases of replicated hybrid zones where sexual isolation appears

to be strong and essentially independent of ecological isolation. A potential example is the hybrid

zone between *Chorthippus* grasshoppers in northern Spain, where female mate choice based on

male calling songs generates strong premating isolation that seems independent from ecological

90   differentiation, at least in some populations (Bridle *et al.*, 2001; Bridle *et al.*, 2002; Bridle *et al.*, 2006;

but see Bailey *et al.*, 2012). Other examples come from situations where sexual isolation is reinforced

by selection against hybridization. For instance, the hybrid zone between *Mus musculus musculus*

and *M. m. domesticus* subspecies of the house mouse have been studied repeatedly in distant

regions, confirming the role of sexual isolation regardless of geographic and ecological conditions

95   (Smadja *et al.*, 2004; Smadja & Ganem, 2005; Bimova *et al.*, 2011).

When sexual isolation is found in conjunction with ecological isolation, it is interesting to

understand the relative roles and interdependence between these two types of barriers. It informs

us on the mechanisms that are currently shaping species boundaries, and in some cases on the origin

and evolution of these mechanisms (Boughman, 2001; Jiggins *et al.*, 2001; Seehausen *et al.*, 2008).

100   Here we focus on isopods *Jaera albifrons* and *J. praehirsuta*, two closely related species that

show strong sexual isolation and generally occupy distinct habitats but can also be found in a region

where they coexist in the same habitat, therefore allowing us to ask whether sexual isolation persists

in a situation where ecological isolation does not.

The two species *J. albifrons* and *J. praehirsuta* belong to the *Jaera albifrons* complex. This

105   complex is composed by five species of small (2-5 mm) marine isopods that live on the shores of the

temperate and cold waters on both sides of the North-Atlantic Ocean (Bocquet, 1953; Solignac,

1978). All species of the complex are phenotypically indistinguishable except for male secondary

sexual traits that are used for tactile courtship (Bocquet, 1953; Solignac, 1981). The males of each

species have specific sets of setae and spines located at different places on their peraeopods (Fig. 1)

110 and they use these features to brush a particular region of the back of females to get them to engage

in sexual intercourse. The males of both species share the same basic courtship behaviour, whereby

they mount females in a head-to-tail position and exercise their brushes. Females accept or reject a

male based on this tactile stimulus, and behavioural isolation is thought to ensure a nearly complete

arrest of interspecific gene flow in nature (Solignac, 1978).

115   Sexual isolation is thus currently a very important barrier between these species, and it could

have initiated speciation (Solignac, 1981). However, the five species of the complex also show some

level of habitat segregation on the seashore. The marine intertidal zone is an interesting

environment to study ecological isolation as it presents contrasted ecological conditions at a small

spatial scale. The distribution of species composing the *Jaera albifrons* complex was found to vary

120 with intertidal zonation, exposure, salinity, and substrate (Naylor & Haahtela, 1966; Jones, 1972).

Most remarkably, while *J. albifrons* and *J. praehirsuta* occupy the same narrow belt of intertidal

habitats along the American and European shores of the North-Atlantic Ocean, *J. albifrons* is

primarily found under pebbles and stones while *J. praehirsuta* is primarily found on intertidal brown

algae (at least along European coasts, Bocquet, 1953; Naylor & Haahtela, 1966; Naylor & Haahtela,

125 1967). These habitats are often in immediate proximity, and these preferences are not strict

(Solignac, 1981; Ribardière *et al.*, 2017), but they still imply that ecological isolation is strong and

thus its current relative contribution to total reproductive isolation must be important (because this

is the first barrier to occur in the life-cycle, see Coyne & Orr, 2004 p. 58).

       The *J. albifrons* / *J. praehirsuta* pair gives us an opportunity to examine the relative strength

130 and interdependence of sexual isolation and ecological isolation because these two species that

usually use distinct habitats were reported to exceptionally coexist in a population where they share

the same habitat (under stones, that is, the primary J. albifrons habitat, Solignac, 1969b; a). This

coexistence of the two species in a unique habitat was recently found to have persisted for decades

and to be more widespread than previously thought as it extends to several other sites at least in the French region Normandy and in the United Kingdom (Ribardière *et al.*, 2017; see also Mifsud 2011). Hybridization happens in these populations (Solignac, 1969a) and results in various levels of introgression (Ribardière, 2017; Ribardière *et al.*, 2017), pointing toward reduced reproductive isolation. Interestingly however, in these hybridizing populations most males bear sexual traits that are clearly identified as belonging to one or the other species and intermediate phenotypes are scarce, suggesting that reproductive isolation does not break down completely.

Ribardière *et al.* (2017) suggested that sexual isolation is one of the components allowing the persistence of bimodality in spite of introgressive hybridization. If this hypothesis is correct and sexual isolation does not disappear in absence of ecological isolation, then it would suggest that sexual isolation has evolved without a direct dependence on ecological conditions and ecological isolation.

Our main objective was to test whether sexual isolation persists in spite of introgressive hybridization in populations showing no ecological isolation. To reach this objective we quantified sexual isolation between *J. albifrons* and *J. praehirsuta* using experimental "no-choice" crosses between individuals that originated either from a region where ecological isolation is strong or a region where ecological isolation is lacking and the two taxa hybridize. Using individuals from this second region we also quantified sexual isolation in a multiple-choice experimental population where females can escape males and there is competition between individuals, unlike in no-choice crosses where mate rejection may be more constrained (e.g. Jiggins *et al.*, 2001).

In addition, no-choice crosses were also performed with individuals from across our two regions in order to test whether sexual isolation could have evolved differently in different ecological contexts. In particular, species *J. praehirsuta* is found on markedly distinct shore substrates (seaweeds vs pebbles) in our two study areas, giving us the opportunity to test for an effect of this ecological difference on sexual isolation between populations within a unique species.

Finally, we took advantage of our experimental crosses to check for potential first-generation

160   intrinsic post-zygotic barrier effects. While such isolating barriers are more likely to operate at later

generations, chromosomal differences have been reported in our two focal species (Staiger &

Bocquet, 1956; Lécher & Prunus, 1971). Thus, we took the opportunity of our experiments to check

for the possibility that such differences have an effect in first-generation hybrids.

Methods

165 *Species*

Contrary to males, females of the five species within the *Jaera albifrons* complex are morphologically indistinguishable (Bocquet, 1953; but see Khaitov *et al.*, 2007 for a discussion of potential phenotypic differences). Females follow the same reproductive cycle (total duration ca. 3 weeks) during which embryos develop in a marsupium (brood pouch) for about 12 days (Solignac,

170 1976). Development is direct, meaning that the dispersal capacity of *Jaera* species is certainly much more limited than that of other marine invertebrates with a pelagic larval phase. Offspring measure ca. 0.5 mm when they are released from the marsupium. Individuals become sexually mature and can be sexed within 4 to 5 weeks based on praeoperculum differentiation (e.g. Solignac, 1979).

175 *Experimental set-up*

Our study is based on the analysis of the reproductive output of virgin males and females used in intra- and inter-specific controlled mating experiments (set-up detailed in Fig. 2). In theory, whether or not juveniles are produced in these experiments could result not only from sexual barrier effects but also post-mating pre-zygotic or intrinsic post-zygotic barrier effects (e.g. inviability of

180 hybrid embryos). However, all past analyses of inter-specific crosses in the *Jaera albifrons* complex have shown that females either rejected hetero-specific males or produced a normal number of offspring. That is, females that produced no offspring did not mate (e.g. Solignac, 1978 p. 49), and females mated by a heterospecific male did not show any reduction in fecundity (e.g. Solignac, 1978 pp. 80-82). There is no postmating copulatory behavioural isolation or mechanical isolation (Bocquet,

185 1953 p. 297; Jones & Fordy, 1971; Veuille, 1978). The complete absence of offspring produced by a pair of individuals is thus a good indicator for sexual isolation (and most probably the behavioural component of sexual isolation, although gametic isolation has yet to be investigated in this group, see discussion).

To obtain virgin individuals of both sexes and both species, we first sampled (unidentified)

190   females in natural populations where *Jaera albifrons* and *J. praehirsuta* coexist (Fig. 2). We chose

populations where we knew from previous work (Solignac, 1978; Ribardière, 2017; Ribardière *et al.*,

2017) that the two species live on different substrates (pebbles *vs* seaweeds, populations from

Brittany) or share the same substrate (pebbles, populations from Normandy). Then we individually

reared in the lab the offspring produced by these females (which were fertilized by unknown males

195   in nature prior to sampling) until they could be sexed and males could be identified. Females were

sorted as *J. albifrons* or *J. praehirsuta* according to the sexual traits held by their brothers. At this

stage we thus had a series of *J. albifrons* and *J. praehirsuta* virgin adults originating from populations

with or without ecological isolation (region "Brittany" *vs* region "Normandy"). These individuals could

then be used in the controlled experiments described below. Female sampling and experimental

200   conditions are detailed in supplementary information.

No-choice crosses within each region

In order to understand reproductive isolation processes with or without ecological isolation,

we first ran a series of crosses where one male and one female from the same region were paired

205   and their reproductive output monitored. These crosses featured intraspecific and interspecific

crosses using either a pair of individuals from Brittany (where ecological isolation is strong) or a pair

from Normandy (where there is no ecological isolation).

We monitored 23 intraspecific and 17 interspecific crosses within each of these two conditions

(that is, 40 crosses within each region of origin; details in Table 1). These numbers were somewhat

210   constrained for three reasons. First, the number of individuals available for the experiment

depended on the (unknown) species identity of the females sampled in the wild, their survival and

fecundity in the lab, and the survival of their offspring (see preliminary steps in Fig. 2). Second, and

more importantly, crosses were designed so that the male and the female that were paired never

shared the same mother. Third, each individual was used in a unique cross, so that each cross was an

215   independent replicate.

For each cross we recorded i) if it successfully produced offspring, ii) how long it took for the

first offspring to appear, iii) how many offspring were contained in each brood produced, and iv)

offspring survival at 35 days. These data were used to estimate reproductive isolation components as

described below. All analyses were performed in R v.3.3.3 (R Core Team, 2017).

220

Reproductive isolation estimated from no-choice crosses within each region

Ecological, sexual, and first-generation intrinsic post-zygotic components of reproductive

isolation were quantified following Sobel and Chen (2014) using estimators that vary between -1

(complete disassortative mating, probability of interspecific gene flow = 1) to 1 (complete

225   reproductive isolation, probability of interspecific gene flow = 0). To compare reproductive isolation

components in the presence *vs* absence of ecological isolation, all the computations described below

were performed independently using crosses featuring individuals "from Brittany" on one hand, and

"from Normandy" on the other hand (that is, two independent sets of analyses).

First, because *J. albifrons* and *J. praehirsuta* in Brittany do not have strictly non-overlapping

230   habitats, we used survey data from Ribardière *et al.* (2017) in the same study areas to quantify

ecological isolation as

$$RI_{eco} = 1 - (S/(S + U)) \tag{1}$$

where *U* was the proportion of individuals found on the primary habitat of their species and *S*

was the proportion of individuals found on the alternative habitat (i.e. *S* is the probability that an

235   individual is in a place where it will meet the other species, that is, "shared"). This equation gives an

estimate of the reduction in interspecific gene flow that would happen if individuals would mate

randomly within each habitat (i.e. ecological isolation only).

Second, we estimated three components of sexual and intrinsic post-zygotic isolation (listed in Table 2) from our experimental crosses. The strength of each reproductive isolation barrier i was estimated as

$$RI_i = 1 - 2 \times (H_i / (H_i + C_i)) \tag{2}$$

where H_i and C_i refer to variables calculated for heterospecific and conspecific pairs (Sobel & Chen, 2014). For sexual isolation (RI_1), H and C were the proportions of inter- and intraspecific crosses that successfully produced offspring. For components of post-zygotic isolation, H and C referred to brood size (number of offspring) or survival (proportion of offspring surviving at day 35) observed from intra- and interspecific crosses (see column "parameter" in Table 2). Each of these components of reproductive isolation was thus estimated independently, giving the strength RI_i that each barrier would have if it were acting alone. Note that RI_1, RI_2, and RI_3 correspond here to different components of RI and should not be confused with similar notations in Sobel & Chen, 2014. A 95% bootstrap confidence interval was calculated for each RI estimate by resampling 10000 times the observed data if sample sizes where not too small (i.e. S and U or H_i and $C_i \geq 14$, Table 2).

Total reproductive isolation was estimated using the product of H_i and C_i across all components of isolation (see details in supplementary information):

$$RI_{tot} = 1 - 2 \frac{S \times \prod_i H_i}{S \times \prod_i H_i + \prod_i C_i} \tag{3}$$

Finally, we estimated the absolute contribution of each individual barrier by subtracting the effect of previously acting barriers as (Sobel & Chen, 2014):

$$AC_i = RI_{[1,i]} - RI_{[1,i-1]} \tag{4}$$

With this definition, individual contributions AC_i can be seen as additive components of RI_{tot}.

No-choice crosses across regions

In addition to the 80 crosses described so far, we ran another series of 80 intraspecific and interspecific crosses (Table 1) pairing individuals from opposite regions (and thus opposite habitats in the case of *J. praehirsuta*, which lives on algae in Brittany vs. under stones in Normandy). These

crosses were useful to test for an effect of habitat on sexual isolation within and between species.

265 They also provided a direct test that *J. albifrons* (or *J. praehirsuta*) from across our two separate

regions actually belong to the same biological species, an implicit assumption of this study and most

previous investigations with this system (Solignac 1969b; Ribardière et al. 2017). The reproductive

output of these crosses was recorded as described above for no choice crosses within each region.

270 *Multiple-choice experiment*

The general aim of the multiple-choice experiment (Fig. 2) was to estimate sexual isolation

based on the reproductive output of males and females *J. albifrons* and *J. praehirsuta* freely

interacting in an experimental population (and thus experiencing intra-sexual competition and easier

male avoidance by females, unlike in no-choice experiments). This experiment focused only on the

275 situation where the two species occupy the same habitat in the wild and thus interact frequently.

Hence we used virgin adult individuals obtained from the no-choice crosses described above (region

Normandy only), so that we could mix *J. albifrons*, J*. praehirsuta*, and F1 hybrids all obtained in the

same controlled conditions and all originating from a region without ecological isolation (Fig. 2).

We used 15 females of each species, 17 males of each species, and 17 males produced by

280 inter-specific crosses. We included F1 hybrid males (and more males than females altogether) in

order to estimate sexual isolation in a situation where females would have free choice, by contrast

with the no-choice design. These numbers were constrained by several parameters, including the

fact that we avoided mixing related males and females (i.e. two males could be brothers, but we did

not use males and females from within the same family). Moreover, 4 out of 17 "hybrid" males

285 turned out to have resulted from a cross between a brother and sister *J. praehirsuta* prior to the

experiment and were removed from all analyses (see details in results and supplementary

information). All remaining F1 hybrid males came from male *J. praehirsuta* x female *J. albifrons*

crosses.

Here we outline the experimental set-up, which is presented in detail in supplementary

290   information. All adults were kept together in a small aquarium for 12 days. After that, all surviving

females were removed from the aquarium and kept individually until they produced offspring, which

were then reared individually. All adults were photographed before and after the experiment and

genotyped at 13 microsatellite loci (Ribardière *et al.*, 2015). All offspring were also genotyped at the

same loci. Photo-identification and genetic parentage assignment (using software Colony v2.0.6.1,

295   Jones & Wang, 2010) were used to identify adult females after the experiment (remember that

females of the two species cannot be distinguished otherwise) and identify the father of each

offspring.

In addition, the secondary sexual traits of all adult males (Fig. 1) were examined under a

microscope to determine their role in male mating success. This is useful in this experiment because

300   multiple-choice conditions give access to variance in male mating success with females of the two

species, and one can thus explore the link between male traits and sexual isolation. Male phenotypes

were summarized using principal component analyses (PCA) based upon 14 phenotypic variables

(see supplementary information). This approach produces linear combinations of traits that are more

efficient for investigating sexual isolation than multiple trait variables separately (Hohenlohe &

305   Arnold, 2010). We used coordinates on the first PCA axis to assess whether female mate choice

matched the distribution of male sexual traits within each species (building upon Ryan & Rand, 1993;

Arnold *et al.*, 1996).

Sexual isolation estimated from the multiple-choice experiment

310   We aimed to compare sexual isolation in these multiple-choice settings with that measured in

no-choice crosses. Hence in a first step we used the same theoretical framework (Sobel & Chen,

2014) to estimate sexual isolation RI_1 following equation (2) with H_1 and C_1 defined as the

proportions of inter-specific and intra-specific crosses that successfully produced offspring. These

proportions were calculated as the number of successful pairs divided by the number of potential

315   pairs that could possibly have formed. Note that because we initially introduced the same number of

males of each species in the experiment, using proportions (as above) or absolute numbers of

successful pairs (as in Sobel & Chen, 2014) would lead to the same result for RI_1.

It turned out that sexual isolation was very strong: only one interspecific pair and two pairs

involving F1 hybrid males produced offspring, while all other successful pairs were conspecific (see

320   results). Hence downstream barriers involving brood size and survival were not quantified because

they would be based on too few samples.

In a second step, sexual isolation was estimated using a framework described by Rolan-Alvarez

and Caballero (2000) that applies to multiple-choice experiments. Rather than estimating the effect

of sexual isolation on interspecific gene flow (as in Sobel and Chen's method), this estimation

325   procedure aims to isolate the effect of mate choice as a cause of reproductive isolation. The

computation takes into account inequalities in mating frequencies rather than assuming that the two

species have the same propensity to mate. In addition, it can detect asymmetries in sexual isolation.

Note that this method can also be applied to no-choice data (Nosil *et al.*, 2002; Coyne *et al.*, 2005) if

they are obtained from equal numbers of no-choice trials of each type (unlike in this study, see Table

330   1).

To estimate sexual isolation we counted the number of male/female pairs of each type (e.g. *J.*

albifrons/J. albifrons, J. albifrons/J. praehirsuta, etc.) that successfully reproduced. Pair sexual

isolation (*PSI*) was estimated for every pair type following Rolan-Alvarez and Caballero (2000). This

method gives a conservative view of sexual isolation as it is defined for each pair type as the number

335   of observed pairs divided by the number of expected pairs given the actual mating success observed.

We then followed these authors' recommendation to estimate I_{PSI}, a modified joint isolation index

(Merrell, 1950) based on *PSI* statistics and that varies from 0 (no isolation) to 1 (complete isolation).

Details of the computation are described in Rolan-Alvarez and Caballero (2000) and Perez-Figueroa

et al. (2005). Values of *PSI*, I_{PSI}, and their statistical significance were computed using JMating v1.0.8

340   (Carvajal-Rodriguez & Rolan-Alvarez, 2006).

Results

Reproductive isolation within each region

Estimates of reproductive isolation are presented in Table 2. In region Brittany, Ribardière *et*

345 *al.* (2017) reported 141 *J. albifrons* under pebbles *vs* 2 on algae, and 3 *J. praehirsuta* under pebbles *vs*

77 on algae. Ecological isolation can thus be estimated to $RI_{eco,Brittany}$= 98%. Because this

reproductive barrier is the first to occur in nature, all other barriers will have comparatively little

effect in natural populations in this region. This is reflected in the values for absolute contributions

(AC_i column in Table 2).

350 However, as shown in Table 2 and Figure 3, no-choice intraspecific crosses in Brittany were

more successful (probability of success=0.9) than interspecific crosses (0.14, Fisher's exact test

p<0.001). These results indicate strong sexual isolation ($RI_{1,Brittany}$= 0.73). That is, sexual isolation

would reduce interspecific gene flow by about 70% in situations were individuals of the two species

would meet, as in the case for instance when bold *J. albifrons* individuals venture on algae, or *J.*

355 *praehirsuta* on pebbles.

By contrast, there was no ecological isolation in mixed *J. albifrons* / *J. praehirsuta* populations

from Normandy (Table 2, data from Ribardière *et al.*, 2017) and there we found sexual isolation to be

half of that found in Brittany, although 95% confidence intervals were overlapping (probability of

success = 0.73 vs. 0.33 for intra- and interspecific crosses, $RI_{1,Normandy}$= 0.37, Fig. 3). Yet in this

360 region, sexual isolation was the first barrier to occur, and thus it effectively led to a strong reduction

in interspecific gene flow ($AC_{1,Normandy} = RI_{1,Normandy} = 0.37$). This stands in contrast with the

situation described above for Brittany where sexual isolation appeared stronger ($RI_{1,Brittany}$= 0.73)

but had very little effect in nature ($AC_{1,Britanny} = 0.015$, Table 2).

The time that a pair of individuals took to produce offspring (Fig. S1) did not enter calculations

365 of reproductive isolation following the framework of Sobel and Chen (2014), but it is interesting to

note that intraspecific crosses produced offspring more quickly (41.2 days on average, *n*=35) than

16

interspecific crosses (58.9 days, *n*=7, generalized linear model GLM with quasi-Poisson family *p*=0.042). The time needed to produce offspring seemed also more variable among interspecific crosses (Fig. S1). These observations further support sexual isolation in the studied populations (see discussion), but there were too few successful interspecific crosses to look at these data in each region separately.

Successful broods contained from 1 to 33 offspring (mean 7.9 ind.) and on average 79.8 % of the offspring were still alive at day 35. Detailed data for intra- and interspecific crosses within each region are presented in Table 2. These data did not indicate any first-generation post-zygotic barrier effect due to reduced F1 hybrid inviability in either region (Table 2). Accordingly, pooling data from our two regions, brood size (Fig. S2) did not differ significantly between intraspecific (*n*=35 broods, mean 7.6 offspring per brood) and interspecific crosses (*n*=7 broods, mean 9.3 offspring per brood, GLM quasi-Poisson family, *p*=0.5). Although the few offspring produced by interspecific crosses gave little power for this test, we note that the point estimate for brood size was actually *higher* in interspecific crosses. The same result was obtained for survival (Fig. S3, n= 265 offspring from intraspecific crosses, 79% survival, n=65 offspring from interspecific crosses, 83% survival, GLM quasi-binomial family *p*=0.6).

No-choice crosses across regions

Experiments that crossed individuals from the same species but originating from distinct regions had a high success (28 out of 35 crosses produced offspring, Fig. 3) and this success was similar to that of intraspecific crosses within each region (Fisher's exact test *p*=0.33). Moreover, this high success was similar for *J. albifrons* pairs and *J. praehirsuta* pairs (respectively 15 out of 19 and 13 out of 16 crosses produced offspring, *p*=1). That is, there was no difference in success when a male and a female came from the same *vs* different regions, and this was true for each of the two species.

Focusing on the differences between intra- and interspecific crosses, all results obtained by crossing individuals from across two distinct regions were almost identical to the results presented

17

above for crosses within a region. Briefly, intra-specific crosses were more successful (*n*= 35 crosses,

probability of success = 0.8) than interspecific crosses (*n*=35, probability of success = 0.2, Fisher's

395   exact test *p*<0.001, Fig 3), delay to offspring production was shorter in intraspecific (*n*=28 broods,

39.2 days on average) than interspecific crosses (*n*=7, 65.3 days, GLM quasi-Poisson *p*=0.006, Fig. S1),

and there was no difference in brood size (intraspecific: *n*=219 offspring from 28 broods, 9.21

offspring per brood on average, interspecific: *n*=68 offspring from 7 broods, 11. 7 offspring per

brood, GLM quasi-Poisson *p*=0.3, Fig. S2) and survival at day 35, (intraspecific: 81% survival,

400   interspecific: 79% survival, GLM quasi-binomial *p*= 0.8, Fig. S3).

In summary, all differences in reproductive output from intra- versus inter-specific crosses

were unaffected by the region of origin of the individuals.

Results from the multiple-choice experimental population

405   We could determine the reproduction patterns for 47 males (out of 51) and 22 females (out of

30), as detailed in the supplementary information. Out of 22 females, most (16) mated with a single

male, while 5 had 2 mates and 1 had none (Fig. S4A). By contrast, the majority of males had no

reproductive success (31 out of 47), while other males mated with 1 to 4 female partners. The

distribution of reproductive success for males and females of each species is shown in Figs. S5 and

410   S6.

A total of 26 male/female pairs produced offspring. As it turned out (Fig. 4), *J. albifrons*

females reproduced only with *J. albifrons* males (*n*=18 pairs), while *J. praehirsuta* females reproduced

with the three types of males: *J. praehirsuta* males (*n*=5), "hybrid" males, (*n*=2), and *J. albifrons* males

(*n*=1). This distribution is not random (Fisher's exact test for independence between male and female

415   species composing these pairs p<0.001) and leads to an estimate for sexual isolation between *J.*

albifrons and *J. praehirsuta* equal to $RI_{1,exp.pop.}$=0.92 (CI$_{95}$ [0.75;1]), a number that can be directly

compared with $RI_{1,Normandy}$=0.37 obtained from no-choice crosses.

These results assume that all individuals had the same probability to mate. Rolan-Alvarez and Caballero's method based on the calculation of pair sexual isolation (*PSI*) is more conservative, since it takes into account the effect of differences in mating success between individuals from each species (e.g. here *J. albifrons* individuals mated more frequently than others). Figure 4 shows the number of mating pairs of each possible type expected from random mating in absence of sexual isolation but taking into account the variation in mating frequencies. Estimates of *PSI* are given in Table S1. The global modified joint isolation index I_{PSI} (Rolan-Alvarez & Caballero, 2000) was equal to 0.46, while pair-specific values were $I_{PSI}^{albi/prae}$=0.92, $I_{PSI}^{albi/hybrid}$=1, and $I_{PSI}^{prae/hybrid}$=0.38, indicating strong isolation between *J. albifrons* and *J. praehirsuta*, and lower isolation between *J. praehirsuta* and hybrids. Bootstrap-based tests performed in JMating for all *PSI* and I_{PSI} estimates showed that only $I_{PSI}^{albi/prae}$ was statistically significant (*p*=0.004).

Most of the variation in parent phenotypes (male secondary sexual traits) could be reduced to two PCA axes (61.8% and 18.3% of the variation explained, Fig. 5A). Using PCA axis 1 to summarize male sexual traits in a univariate space, we see in Figure 5B that the mates preferred by *J. albifrons* females appear to coincide with the distribution of male sexual traits of *J. albifrons*, while *J. praehirsuta* females mated with males showing a wider range of trait values.

Discussion

Sexual isolation was effective both with and without ecological isolation

We found strong sexual isolation both in a context where ecological isolation is nearly complete ($RI_{eco,Brittany}$=98%, $RI_{1,Brittany}$= 73%) and in a context where there is no ecological isolation and individuals of the two species may hybridize ($RI_{eco,Normandy}$=0, $RI_{1,Normandy}$= 37%). These RI values based on reproductive success in no-choice conditions suggest that sexual isolation is less effective when the two species co-occur in the same habitat (Normandy). However, the multiple-choice experiment showed that when the conditions allow females to escape males or choose amongst several males, then sexual isolation is in fact very efficient even in the hybridizing

populations from Normandy ($RI_{1,exp.pop.}$= 92%). The exact same value was obtained using the joint

isolation index for *J. albifrons* and *J. praehirsuta* ($I_{PSI}^{albi/prae}$=0.92), indicating that sexual isolation was

effectively due to mate choice rather than differences in mating propensity between species.

Moreover, most of our no-choice crosses were monitored for a long time (5 to 196 days,

median 22 days), meaning that females were confined with a given male in a tiny area for a long

period, and thus our estimates probably give us a lower bound on sexual isolation. We conclude that

sexual isolation remains strong in populations where the two species share the same habitat,

(despite hybridization and introgression, Ribardière, 2017; Ribardière *et al.*, 2017) and thus sexual

isolation in this system is largely independent of ecological isolation.

In addition, while males were identified as *J. albifrons* or *J. praehirsuta* based on phenotypes

that are directly relevant to reproductive isolation, this was not the case for females, which were

identified based on the phenotype of their brothers (see methods, Fig. 2, and supplementary

information). Hence in all our experiments we had no direct information on the sexual phenotype of

females (that is, preferences). This uncertainty on what really was a "*J. albifrons*" or a "*J. praehirsuta*"

female in our experiments makes it all the more remarkable that sexual isolation was found to be

strong, especially in the populations where the two species share the same habitat and hybridize.

Three potential caveats must be considered. First, we took offspring production in intra- *vs*

inter-specific crosses as an indicator of sexual isolation. This approach would be incorrect if intrinsic

post-zygotic barriers were involved. Here brood size did not differ between intraspecific and

interspecific crosses, and the abortion of an entire brood of embryos was never observed (oocytes

and developing embryos are visible in the marsupium of females). Hence, conforming to what had

been reported previously (Solignac, 1978), females in the presence of a heterospecific male either

produced no offspring because they did not mate, or produced a normal number of offspring. The

success of crosses (presence vs absence of juveniles produced) was thus a good indicator of sexual

isolation.

Second, the multiple-choice population was not replicated. Although clear-cut patterns were

470   observed, one cannot rule out that it was affected by an undetected experimental bias or that

individuals originating from other populations would behave differently. For instance, the 81

individuals mixed in the experiment were produced in the lab from 17 pairs of parents, and each

class of individuals (*J. albifrons* males, *J. albifrons* females, *J. praehirsuta* males, etc.) thus comprised

2 to 5 groups of siblings (1 to 9 siblings per family). Most notably, the fourteen *J. albifrons* females

475   that survived the experiment and reproduced came from only two families, which could have biased

the result if these 14 females were not representative of the range of female behaviour in natural

populations. No-choice crosses have shown that this is not the case (*J. albifrons* females were very

selective, both in intra-region and inter-region crosses), but repeating this experiment with more

individuals, and perhaps with individuals from other regions where the two species have been found

480   in the same habitat, would be particularly interesting (e.g. in the UK, Naylor & Haahtela, 1966;

Mifsud, 2011; Ribardière, 2017).

Did sexual isolation evolve independently of ecological contexts?

J. praehirsuta individuals in Brittany vs. Normandy dwell on different habitats (pebbles vs.

485   brown algae). However, intraspecific crosses had exactly the same success (and the same delay to

offspring production) whether or not males and females originated from the same region or different

regions (Figs. 3 and S1), and this success was the same as that of *J. albifrons* pairs (within or across

regions). Hence habitat differences did not generate any sign of sexual isolation between individuals

originating from algae vs. rock populations in species *J. praehirsuta*. Sexual isolation was unaffected

490   by the habitat of origin and thus we conclude that it did not evolve through sexual selection

mechanisms that are dependent of local ecological factors.

However, the ecological context could still have an effect on sexual isolation via two potential

mechanisms. First, to suit our purpose of estimating reproductive isolation in different populations,

all individuals were reared in identical, artificial lab conditions (see supplementary information)

21

495   where all five species of the *Jaera albifrons* complex had been shown to have high fitness (Bocquet

1953; Solignac 1978). This means in particular that all individuals used in cross experiments were

born and raised in the exact same conditions, regardless of their population of origin. Hence we

cannot exclude that contrasted substrates (algae vs. rocks) could have some proximate effect on

mate choice mechanisms, for instance through phenotypic plasticity in cuticular hydrocarbons or

500   other chemical cues involved in sexual signalling (see e.g. Breithaupt & Hardege, 2012, for a review

of chemical cues in aquatic organisms; and Zhu *et al.*, 2012 for an example involving sexual isolation).

Such chemical signalling mechanisms have yet to be investigated in the *Jaera albifrons* complex,

although we note that their effect on reproductive isolation would necessarily be somewhat limited

because sexual isolation is already strong in uniform lab conditions.

505   Second, as mentioned above we observed from no-choice trials that sexual isolation could be

twice as strong in populations where the two species are ecologically separated ($RI_{1,Brittany}$= 73%,

$RI_{1,Normandy}$= 37%). One potential explanation is that mate choice is relaxed in introgressed

populations, but only in situations where mate options are constrained (in agreement with our

finding that sexual isolation was stronger in the multiple-choice experimental population

510   $RI_{1,exp.pop.}$= 92%). Variations in the relative abundance and spatial distribution of individuals in

natural conditions may thus modulate the likelihood of interspecific mating. It is often argued that

this could happen when dynamic habitats modulate the intensity of sexual isolation by limiting mate

choice options, perhaps in some cases approaching no-choice conditions, if for example a female of

species A finds herself isolated for some time with males of species B only. This could happen in

515   some of our populations where the quantity of sand covering pebbles and stones varies with climatic

conditions across seasons and the density of individuals is low (e.g. Fig. S1 in supplementary material

from Ribardière *et al.*, 2017). Khaitov *et al.* (2007) also observed sharp variations in the relative

frequencies of *Jaera* species in mixed populations that were presumably attributable to the

destruction of nearby habitats.

520

22

Sexual isolation is not strict, and it is asymmetric

Sexual isolation was strong, but it was not 100%. Hence sexual isolation is not sufficient to prevent interspecific gene flow entirely on its own. As a result, when ecological isolation is lacking, hybridization is expected to happen occasionally.

525 Moreover, results from the multiple-choice experimental population showed sexual isolation to be asymmetric (Figs. 4 and 5). In this experiment, *J. albifrons* females mated exclusively with *J. albifrons* males while *J. praehirsuta* females mated with *J. praehirsuta* (*n*=5), *J. albifrons* (*n*=1) and F1 hybrid males (*n*=2). Accordingly, pair sexual isolation indices for heterospecific pairs were smaller (indicating stronger isolation) when the female involved was *J. albifrons* rather than *J. praehirsuta*

530 (Table S1). Summarizing male sexual phenotypes using the first axis of a principal component analysis (Fig. 5), we found that the probability that a male mated with a *J. albifrons* female matched relatively well the density distribution of *J. albifrons* male sexual traits. By contrast, the probability that a male mated with a *J. praehirsuta* female was clearly less concordant to the density distribution of *J. praehirsuta* male sexual traits (Fig. 5B). This shows that females *J. praehirsuta* mated with our three

535 categories of males (*J. praehirsuta*, F1 hybrids, and *J. albifrons*) because they are less selective that *J. albifrons* females with regards to male sexual traits (illustrative of "type d" preference function in Fig. 5 from Ryan & Rand, 1993).

This asymmetry is also confirmed by the no-choice experiments, where inter-specific crosses were systematically more successful with *J. praehirsuta* females. Over all interspecific crosses, 4 out

540 of 36 were successful when the female was *J. albifrons*, while 10 out of 28 were successful when the female was *J. praehirsuta* (Fisher's exact test p=0.036, Fig. 6). Interestingly, the same result was obtained by M. Solignac with no-choice experiments using individuals from a population from France (Brittany) crossed with individuals from Germany (0% success for German *J. albifrons* females crossed with French *J. praehirsuta* males vs 22 to 46% success for French *J. praehirsuta* females

545 crossed with German J. *albifrons* males, depending on the method of calculation, Solignac, 1978).

The asymmetry in sexual isolation is thus observed in all conditions: large-scale allopatry (between countries, Solignac, 1978), smaller-scale allopatry (between regions, this study), sympatry with ecological isolation and no hybridization (Brittany, this study), and sympatry without ecological isolation and with introgressive hybridization (Normandy, this study). This is interesting because it

550   suggests that the asymmetry has a general cause of ancestral origin and has not evolved in response to local conditions, in agreement with our previous conclusion that sexual isolation has not evolved following mechanisms that depend on local habitat conditions.

Why do *J. albifrons* and *J. praehirsuta* coexist in hybridizing populations?

555   Individuals identified as *J. albifrons* in region Brittany (with ecological isolation) belong to the same biological species as individuals identified as *J. albifrons* in region Normandy (where there is no ecological isolation), and likewise for species *J. praehirsuta*. This was argued by taxonomists and evolutionary biologists based on the observation of secondary sexual traits (Bocquet, 1953; Prunus, 1968; Solignac, 1978) and confirmed in this study using cross experiments: conspecific crosses gave

560   the same results within and across regions (no reproductive isolation between regions).

However, we also know that reproductive isolation is not complete in populations without ecological isolation, where hybridization leads to introgression (Bocquet & Solignac, 1969; Solignac, 1969a; 1978; Ribardière *et al.*, 2017). This situation raises questions about the conditions of coexistence of the two species in spite of hybridization and ecological equivalence (e.g. Coyne & Orr,

565   1998). Ribardière *et al.* (2017) discussed the peculiar nature of the hybridizing populations, emphasizing that they seem to receive no influx of individuals from pure parental populations, and that no fine-scale ecological differentiation was observed (the two species are repeatedly found to be intermingled in a number of different sites). The authors suggested sexual and post-zygotic isolation as two potential forces somehow acting to keep the species isolated. Here we found sexual

570   isolation to be effectively strong in hybridizing populations, but no support for first-generation intrinsic post-zygotic mechanisms (AC_2 and AC_3 added to -0.04 and -0.06 in regions Brittany and

Normandy, Table 2). Moreover, this result was confirmed by between-regions crosses showing similar brood size and offspring survival for intra- and interspecific crosses. We could not quantify intrinsic post-zygotic isolation from the multiple-choice experimental population because only one

575 heterospecific mating pair and two backcrosses (F1-hybrid fathers) produced offspring. Yet these broods showed no sign of reduced fitness in any way (brood size and offspring survival, data not shown). Post-zygotic barrier effects need to be investigated further (especially in later generations), but so far, sexual processes (mate choice and/or intra-sexual interactions) constitute the only force identified that could maintain the two species separated in spite of extensive introgression. This

580 alone is unusual, and the fact that the two species are found to coexist in different places and on the long term despite ecological equivalence poses another conundrum that deserves further investigations.

Conclusion & open questions

585   In this study we looked at two closely related species that are reproductively isolated by ecological and sexual barriers, asking first if sexual isolation could maintain species integrity on its own or is only acting secondarily alongside ecological isolation, and second if sexual isolation may have evolved independently of local ecological context. We found that sexual isolation is a strong barrier that does not break down when ecological isolation is absent and most probably evolved

590 independently of local ecological conditions. The tactile courtship of *J. albifrons* and *J. praehirsuta* therefore probably exemplifies a situation where the set of genes and traits involved in male phenotypes and female preferences have no simultaneous role in local adaptation to different habitats and are not constrained by natural selection (no environmental effect on sexual traits, courtship behaviour, transmission of sexual signals, or choosiness). Under these conditions one can

595 expect sexual isolation to be largely independent of ecological isolation, as observed for instance in some *Chorthippus* grasshoppers (Bridle *et al.*, 2001; Bridle *et al.*, 2002) and *Laupala* crickets (Mendelson & Shaw, 2005) in which female mate choice is based on male calling songs. In some

25

Drosophila species, laboratory lines reared in artificial conditions provide another example where

mate choice based on male courtship songs and pheromones (reviewed in Coyne & Orr, 1997) has no

600 obvious ecological component (although this will not be the case of all *Drosophila* species, see e.g.

Stennett & Etges, 1997; Etges, 2002). A few other potential examples have been reviewed by

Svensson (2012). Will reproductive isolation be generally weaker in these conditions? Empirical

examples reported in the literature show that reproductive isolation can be particularly strong when

sexual isolation is inherently coupled with ecological isolation (Servedio *et al.*, 2011). Whether this is

605 a general rule remains to be clarified.

Local ecological conditions could still affect the strength of sexual isolation if habitat dynamics

modulate mate choice possibilities through stochastic effects on population densities. Whether such

effects could constrain mate choice sufficiently to bypass sexual isolation and affect the likelihood of

speciation completion remains an open question. More importantly, we found that reproductive

610 isolation was not complete without ecological isolation. Both *J. albifrons* and *J. praehirsuta* are found

in variable ecological contexts along the shores of the temperate and cold waters on both sides of

the North-Atlantic Ocean and nearby seas. Whether or not there is niche differentiation may

influence the local coexistence and hybridization of these species.

Finally, the conditions of coexistence of these two species in hybridizing populations are

615 another open question. For that, further examination of post-zygotic barrier effects, fine-scale tests

for cryptic habitat specialization, and interactions favouring density-dependence mechanisms (such

as species-specific pathogen effects) may provide some answers.

26

platform at Station Biologique de Roscoff and was funded by the French Agence Nationale de la

625   Recherche (grant ANR-13-JSV7-0001-01 to T.B.).

Author contributions

Conceptualization and methodology: AR and TB. Field sampling and species identification: AR,

EP, JC, CH, SL and TB. Crossing experiments and maintenance of the individuals in the laboratory: AR,

630   EP, JC, CH, SH, and TB. Genotyping and phenotyping: AR, EP, JC, CDT, CH and TB. Analyses and

writing: AR, EP, and TB. Supervision, project administration and funding acquisition: TB.

Conflict of interest disclosure

The authors of this preprint declare that they have no financial conflict of interest with the

635   content of this article

Data accessibility

The data associated with this study are available on the Dryad digital repository at

https://doi.org/10.5061/dryad.dv41ns1t8

640   .

References

Arnold, S.J., Verrell, P.A. & Tilley, S.G. 1996. The evolution of asymmetry in sexual isolation: A model and a test case. *Evolution* **50:** 1024-1033.

Bailey, R.I., Saldamando-Benjumea, C.I., Tatsuta, H. & Butlin, R.K. 2012. Habitat Association and Seasonality in a Mosaic and Bimodal Hybrid Zone between Chorthippus brunneus and C. jacobsi (Orthoptera: Acrididae). *Plos One* **7**.

Bierne, N., Bonhomme, F. & David, P. 2003. Habitat preference and the marine-speciation paradox. *Proceedings of the Royal Society B-Biological Sciences* **270:** 1399-1406.

Bimova, B.V., Macholan, M., Baird, S.J.E., Munclinger, P., Dufkova, P., Laukaitis, C.M., Karn, R.C., Luzynski, K., Tucker, P.K. & Pialek, J. 2011. Reinforcement selection acting on the European house mouse hybrid zone. *Molecular Ecology* **20:** 2403-2424.

Bocquet, C. 1953. Recherches sur le polymorphisme naturel des *Jaera Marina* (Fabr.)(Isopodes Asellotes) : Essai de systématique évolutive, Centre national de la recherche scientifique.

Bocquet, C. & Solignac, M. 1969. Etude morphologique des hybrides expérimentaux entre *Jaera* (*albifrons*) *albifrons* et *Jaera* (*albifrons*) *praehirsuta* (isopodes asellotes). *Archives de zoologie expérimentale et générale* **110:** 435-452.

Boughman, J.W. 2001. Divergent sexual selection enhances reproductive isolation in sticklebacks. *Nature* **411:** 944-948.

Boughman, J.W. 2002. How sensory drive can promote speciation. *Trends in Ecology & Evolution* **17:** 571-577.

Boughman, J.W. & Svanback, R. 2017. Synergistic selection between ecological niche and mate preference primes diversification. *Evolution* **71:** 6-22.

Breithaupt, T. & Hardege, J.D. 2012. Pheromones mediating sex and dominance in aquatic animals. *Chemical Ecology in Aquatic Systems***:** 39-56.

Bridle, J.R., Baird, S.J.E. & Butlin, R.K. 2001. Spatial structure and habitat variation in a grasshopper hybrid zone. *Evolution* **55:** 1832-1843.

Bridle, J.R., Vass-De-Zomba, J. & Butlin, R.K. 2002. Fine-scale ecological and genetic variation in a

Chorthippus grasshopper hybrid zone. *Ecological Entomology* **27:** 499-504.

Bridle, J.R., Saldamando, C.I., Koning, W. & Butlin, R.K. 2006. Assortative preferences and

discrimination by females against hybrid male song in the grasshoppers Chorthippus

brunneus and Chorthippus jacobsi (Orthoptera : Acrididae). *Journal of Evolutionary Biology*

19: 1248-1256.

Butlin, R.K. & Smadja, C.M. 2018. Coupling, Reinforcement, and Speciation. *The American Naturalist*

191: 155-172.

Carvajal-Rodriguez, A. & Rolan-Alvarez, E. 2006. JMATING: a software for the analysis of sexual

selection and sexual isolation effects from mating frequency data. *Bmc Evolutionary Biology*

6.

Coyne, J.A. & Orr, H.A. 1997. ''Patterns of speciation in Drosophila'' revisited. *Evolution* **51:** 295-303.

Coyne, J.A. & Orr, H.A. 1998. The evolutionary genetics of speciation. *Philosophical Transactions of

the Royal Society of London Series B-Biological Sciences* **353:** 287-305.

Coyne, J.A. & Orr, H.A. 2004. *Speciation*. Sinauer associates, Sunderland.

Coyne, J.A., Elwyn, S. & Rolan-Alvarez, E.L. 2005. Impact of experimental design on Drosophila sexual

isolation studies: Direct effects and comparison to field hybridization data. *Evolution* **59:**

2588-2601.

Culumber, Z.W., Fisher, H.S., Tobler, M., Mateos, M., Barber, P.H., Sorenson, M.D. & Rosenthal, G.G.

2011. Replicated hybrid zones of Xiphophorus swordtails along an elevational gradient.

Molecular Ecology **20:** 342-356.

Etges, W.J. 2002. Divergence in mate choice systems: does evolution play by rules? *Genetica* **116:**

151-166.

Harrison, R.G. & Rand, D.M. 1989. Mosaic hybrid zones and the nature of species boundaries. In:

Speciation and its consequences (D. Otte & J. A. Endler, eds), pp. 111-133. Sinauer Associates,

Inc.

Harrison, R.G. & Larson, E.L. 2016. Heterogeneous genome divergence, differential introgression, and the origin and structure of hybrid zones. *Molecular Ecology* **25:** 2454-2466.

Hasselman, D.J., Argo, E.E., Mcbride, M.C., Bentzen, P., Schultz, T.F., Perez-Umphrey, A.A. & Palkovacs, E.P. 2014. Human disturbance causes the formation of a hybrid swarm between two naturally sympatric fish species. *Molecular Ecology* **23:** 1137-1152.

Hohenlohe, P.A. & Arnold, S.J. 2010. Dimensionality of mate choice, sexual isolation, and speciation. *Proceedings of the National Academy of Sciences of the United States of America* **107:** 16583-16588.

Jiggins, C.D., Naisbit, R.E., Coe, R.L. & Mallet, J. 2001. Reproductive isolation caused by colour pattern mimicry. *Nature* **411:** 302-305.

Jones, M. 1972. Effects of salinity on the survival of the Jaera albifrons Leach group of species (Crustacea: Isopoda). *Journal of Experimental Marine Biology and Ecology* **9:** 231-237.

Jones, M.B. & Fordy, M.R. 1971. Stereoscan Electron Microscope Study of Male Reproductive Characters in Jaera-Albifrons Group of Species (Crustacea-Isopoda). *Marine Biology* **10:** 265-&.

Jones, O.R. & Wang, J.L. 2010. COLONY: a program for parentage and sibship inference from multilocus genotype data. *Molecular Ecology Resources* **10:** 551-555.

Khaitov, V.M., Kuzmin, A.A. & Terovskaya, E.V. 2007. Morphological differences between females of different Jaera species (Crustacea : Asellota : Isopoda) in the White Sea: a possible solution to an old problem. *Marine Biology* **150:** 1205-1214.

Lécher, P. & Prunus, G. 1971. Caryologie et taxinomie de *Jaera albifrons* (crustacé, isopode), populations des côtes Bretonnes. *Archives de Zoologie expérimentale et générale* **112:** 715-730.

Maan, M.E. & Seehausen, O. 2011. Ecology, sexual selection and speciation. *Ecology Letters* **14:** 591-602.

McNett, G.D. & Cocroft, R.B. 2008. Host shifts favor vibrational signal divergence in Enchenopa binotata treehoppers. *Behavioral Ecology* **19:** 650-656.

Mendelson, T.C. & Shaw, K.L. 2005. Sexual behaviour: Rapid speciation in an arthropod. *Nature* **433:** 375-376.

Merrell, D.J. 1950. Measurement of Sexual Isolation and Selective Mating. *Evolution* **4:** 326-331.

Mifsud, D.V. 2011. The genetic basis of speciation in the *Jaera albifrons* species group of intertidal isopods, University of Aberdeen. **PhD:** 201.

Naylor, E. & Haahtela, I. 1966. Habitat Preferences and Interspersion of Species within Superspecies Jaera Albifrons Leach (Crustacea . Isopoda). *Journal of Animal Ecology* **35:** 209-&.

Naylor, E. & Haahtela, I. 1967. Quantitative ecological distribution of the Jaera albifrons group of species in the Baltic. *Ophelia* **4:** 19-27.

Nosil, P., Crespi, B.J. & Sandoval, C.P. 2002. Host-plant adaptation drives the parallel evolution of reproductive isolation. *Nature* **417:** 440-443.

Nosil, P. 2012. Ecological Speciation. *Ecological Speciation***: 1-280.

Perez-Figueroa, A., Caballero, A. & Rolan-Alvarez, E. 2005. Comparing the estimation properties of different statistics for measuring sexual isolation from mating frequencies. *Biological Journal of the Linnean Society* **85:** 307-318.

Prunus, G. 1968. Etude de systématique des populations chez l'isopode *Jaera* (*albifrons*) *albifrons* Forsman. *Archives de zoologie expérimentale et générale* **109:** 643-702.

R Core Team 2017. R: A language and environment for statistical computing, R Foundation for Statistical Computing. Vienna, Austria. URL http://www.R-project.org.

Ribardière, A., Broquet, T. & Daguin-Thiébaut, C. 2015. Microsatellite markers for the *Jaera albifrons* species complex (marine isopods). *BMC Research Notes* **8:** 632.

Ribardière, A. 2017. Isolement reproductif et architecture génomique de la différenciation chez deux espèces du complexe *Jaera albifrons* (isopodes marins) - Etude de populations mixtes

présentant des niveaux d'isolement interspécifique contrastés, Pierre & Marie Curie University. **PhD:** 276.

Ribardière, A., Daguin-Thiébaut, C., Houbin, C., Coudret, J., Broudin, C., Timsit, O. & Broquet, T. 2017. Geographically distinct patterns of reproductive isolation and hybridization in two sympatric species of the Jaera albifrons complex (marine isopods). *Ecology and Evolution* **7:** 5352-5365.

Ritchie, M.G. 2007. Sexual selection and speciation. *Annual Review of Ecology Evolution and Systematics* **38:** 79-102.

Rolan-Alvarez, E. & Caballero, M. 2000. Estimating sexual selection and sexual isolation effects from mating frequencies. *Evolution* **54:** 30-36.

Rougemont, Q., Gaigher, A., Lasne, E., Cote, J., Coke, M., Besnard, A.L., Launey, S. & Evanno, G. 2015. Low reproductive isolation and highly variable levels of gene flow reveal limited progress towards speciation between European river and brook lampreys. *Journal of Evolutionary Biology* **28:** 2248-2263.

Ryan, M.J. & Rand, A.S. 1993. Species recognition and sexual selection as a unitary problem in animal communication. *Evolution* **47:** 647-657.

Safran, R.J., Scordato, E.S.C., Symes, L.B., Rodriguez, R.L. & Mendelson, T.C. 2013. Contributions of natural and sexual selection to the evolution of premating reproductive isolation: a research agenda. *Trends in Ecology & Evolution* **28:** 643-650.

Seehausen, O., Terai, Y., Magalhaes, I.S., Carleton, K.L., Mrosso, H.D.J., Miyagi, R., van der Sluijs, I., Schneider, M.V., Maan, M.E., Tachida, H., Imai, H. & Okada, N. 2008. Speciation through sensory drive in cichlid fish. *Nature* **455:** 620-U623.

Seehausen, O. 2009. Progressive levels of trait divergence along a 'speciation transect' in the lake Victoria cichlid fish *Pundamilia*. In: *Speciation and patterns of diversity* (R. Butlin, J. Bridle & D. Schluter, eds), pp. 155-176. Cambridge University Press.

Servedio, M.R., Van Doorn, G.S., Kopp, M., Frame, A.M. & Nosil, P. 2011. Magic traits in speciation: 'magic' but not rare? *Trends in Ecology & Evolution* **26:** 389-397.

Servedio, M.R. & Boughman, J.W. 2017. The Role of Sexual Selection in Local Adaptation and Speciation. *Annual Review of Ecology, Evolution, and Systematics* **48:** 85-109.

Smadja, C., Catalan, J. & Ganem, G. 2004. Strong premating divergence in a unimodal hybrid zone between two subspecies of the house mouse. *Journal of Evolutionary Biology* **17:** 165-176.

Smadja, C. & Ganem, G. 2005. Asymmetrical reproductive character displacement in the house mouse. *Journal of Evolutionary Biology* **18:** 1485-1493.

Sobel, J.M. & Chen, G.F. 2014. Unification of Methods for Estimating the Strength of Reproductive Isolation. *Evolution* **68:** 1511-1522.

Solignac, M. 1969a. Hybridation introgressive dans la population complexe des *Jaera albifrons* de Luc-sur-Mer. *Archives de zoologie expérimentale et générale* **110:** 629.

Solignac, M. 1969b. Contiguous Distributions Sympatry and Natural Hybridization in a Super-Species Jaera Albifrons Leach (Isopods Asellidae). *Comptes Rendus Hebdomadaires Des Seances De L Academie Des Sciences Serie D* **268:** 1610-&.

Solignac, M. 1976. Demographic aspects of interspecific hybridization - A study of the Jaera albifrons species complex (Crustacea, Isopoda, Asellota). *Oecologia* **26:** 33-52.

Solignac, M. 1978. Nature, déterminisme et origine des mécanismes d'isolement dans le complexe Jaera albifrons (Isopodes, Asellotes). Phd Thesis., PhD thesis.

Solignac, M. 1979. Experimental-Analysis and Variability of Interspecific Hybridization in the Species Complex Jaera-Albifrons (Crustacea, Isopoda). *Archives De Zoologie Experimentale Et Generale* **120:** 353-378.

Solignac, M. 1981. Isolating mechanisms and modalities of speciation in the *Jaera albifrons* species complex (*Crustacea*, *Isopoda*). *Systematic Zoology* **30:** 387-405.

Staiger, H. & Bocquet, C. 1956. Les chromosomes de la super-espèce Jaera marina (F.) et de quelques autres Janiridae (Isopodes Asellotes). *Bull. Biol. Fr. Belg* **90:** 1-32.

Stennett, M.D. & Etges, W.J. 1997. Premating isolation is determined by larval rearing substrates in cactophilic Drosophila mojavensis. III. Epicuticular hydrocarbon variation is determined by

use of different host plants in Drosophila mojavensis and Drosophila arizonae. *Journal of Chemical Ecology* **23:** 2803-2824.

Svensson, E.I. 2012. Non-ecological speciation, niche conservatism and thermal adaptation: how are they connected? *Organisms Diversity & Evolution* **12:** 229-240.

Taylor, E.B., Boughman, J.W., Groenenboom, M., Sniatynski, M., Schluter, D. & Gow, J.L. 2006. Speciation in reverse: morphological and genetic evidence of the collapse of a three-spined stickleback (Gasterosteus aculeatus) species pair. *Molecular Ecology* **15:** 343-355.

Veuille, M. 1978. Biology of Reproduction in Jaera .1. Structure and Functionment of Males Copulatory Appendages. *Cahiers De Biologie Marine* **19:** 299-308.

Zhu, J., Zhang, D. & Lin, J.D. 2012. Asymmetric Mating Isolation between Two Caridean Shrimp: Lysmata Wurdemanni and Lysmata Boggessi. *Journal of Shellfish Research* **31:** 195-199.

Table 1 - Number of experimental "no-choice" crosses performed in order to estimate reproductive isolation

between marine isopods *J. albifrons* and *J. praehirsuta* in two French regions where they are ecologically isolated

(Brittany) or not (Normandy). Intra- and interspecific crosses were performed using males and females from the

same region (intra-region) and different regions (inter-region). Inter-region crosses involved either females from

Brittany (F. Britt) crossed to males from Normandy (M. Norm) or the opposite (F. Norm crossed to M. Britt).

Cross type	Female	Male	Intra-region		Inter-region	
			Brittany	Normandy	F. Britt. x M. Norm.	F. Norm. x M. Britt.
Intraspecific						
	J. albifrons	*J. albifrons*	17	6	10	10
	J. praehirsuta	*J. praehirsuta*	6	17	10	10
Interspecific						
	J. albifrons	*J. praehirsuta*	10	10	10	10
	J. praehirsuta	*J. albifrons*	7	7	10	10

Table 2 - Components of reproductive isolation between marine isopods *Jaera albifrons* and *J. praehirsuta* in two regions with contrasting levels of isolation. Reproductive isolation (RI) was calculated following Sobel & Chen (2014). Bootstrap confidence intervals (CI$_{95}$) based on 10000 resampling of observed data are given whenever sample sizes were not too small (e.g. sample sizes for survival of interspecific offspring in Brittany were particularly small since interspecific crosses were rarely successful). RI gives the reduction of interspecific gene flow that would be caused by each specific barrier acting alone. AC gives the actual absolute contribution of each barrier given that other barriers are acting earlier in life cycle. The sum of AC over all barriers is equal to the total strength of reproductive isolation. Sexual isolation contains a strong behavioural component, but could also include a (so far untested) gametic component. RI due to F1 hybrid inviability is based on brood size (number of offspring produced by a mother after intra-marsupial development) and survival of these offspring after 35 days.

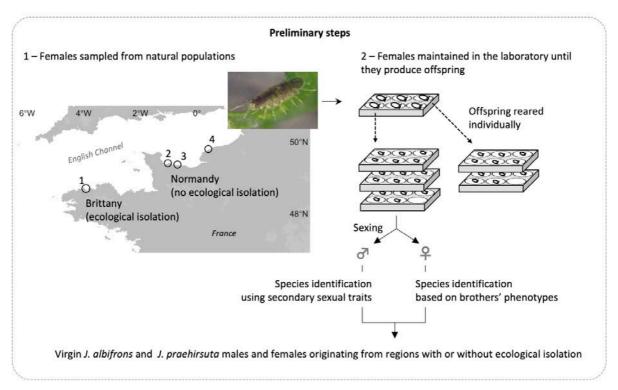| Barrier	Notation	Parameter	Brittany		RI [CI$_{95}$]	AC	Normandy		RI [CI$_{95}$]	AC
			Data				Data			
Ecological isolation	RI_{eco}	Probability of heterospecific encounter	5/222 ind. in shared habitat	217/222 ind. in unshared habitat	0.98 [0.955 ; 0.995]	0.977	80/80 ind. in shared habitat	0/80 ind. in unshared habitat	0.00	0.00
Sexual isolation	RI_1	Proportion of successful crosses	19/21 successful intrasp. crosses	2/14 successful intersp. crosses	0.73 [0.45 ; 1]	0.015	16/22 successful intrasp. crosses	5/15 successful intersp. crosses	0.37 [0.06 ; 0.73]	0.37
F1 hybrid inviability	RI_2	Number of offspring released per brood	6.2 ind. (*n*=19 intrasp. broods)	5.5 ind. (*n*=2 intersp. broods)	0.06	0.001	9.3 ind. (*n*=16 intrasp. broods)	10.8 ind. (*n*=5 intersp. broods)	-0.08	-0.07
	RI_3	Survival probability at 35 days	0.74 (*n*=117 intrasp. offsp.)	0.91 (*n*=11 intersp. offsp.)	-0.10	-0.001	0.83 (*n*=148 intrasp. offsp.)	0.81 (*n*=54 intersp. offsp.)	0.01 [-0.06 ; 0.09]	0.01
Total reproductive isolation	RI_{tot}				0.99	0.99			0.31	0.31

Figure 1

Sexual traits used for tactile courtship by male *Jaera albifrons* and *J. praehirsuta*. In male *J. albifrons*, the second segment (carpus) of peraeopods 6 and 7 extends as a lobe bearing a patch of setae (indicated by the black arrow on the left). Male *J. praehirsuta* instead have curved setae distributed on the first three segments (propus, carpus, merus) of peraeopods 1-4 (right black arrow), and one or two spines on the carpus of peraeopods 6 and 7. The photo on the left shows a 4mm-long adult female (photo credit to Guillaume Evanno & Thomas Broquet). The drawings are reproduced from Solignac (1981) with authorization. After fluorescent labelling of dissected appendices, close-up pictures were obtained with a confocal laser scanning microscope and processed using software Fiji and IMARIS (photo credit to Sébastien Colin).
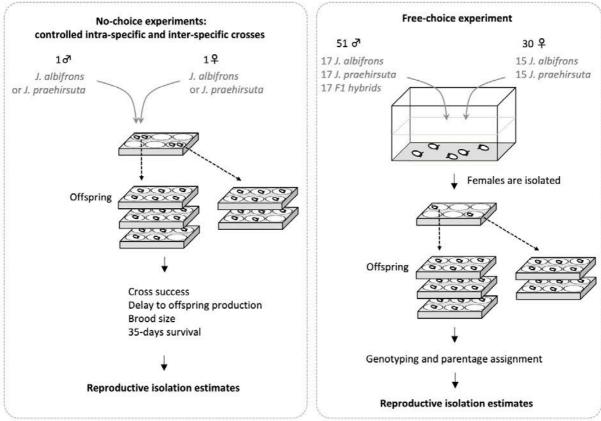
Figure 2

Figure 2 – Outline of experimental protocols. The upper panel presents the preliminary steps that

were taken to obtain virgin males and females of each species, which could then be used in

controlled experiments. Adult females were sampled from natural populations as shown on the map. We knew from previous genetic analyses that local sympatric populations of *J. albifrons* and *J. praehirsuta* were reproductively isolated (region Brittany) or not (introgressive hybridization, region Normandy). Adult females were kept in the laboratory until they produced offspring that were then raised until the male offspring could be identified. Virgin individuals born in the lab were then chosen to be used in no-choice cross experiments. Individuals born and reared in the lab following these no-choice experiments were then used in a multiple-choice experimental population.
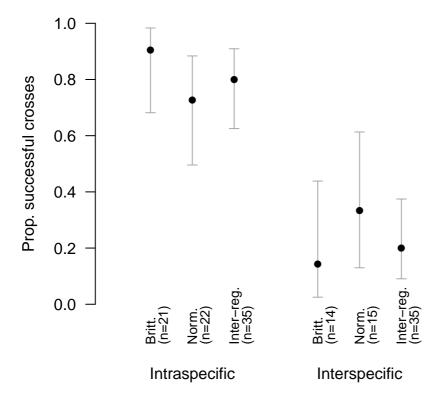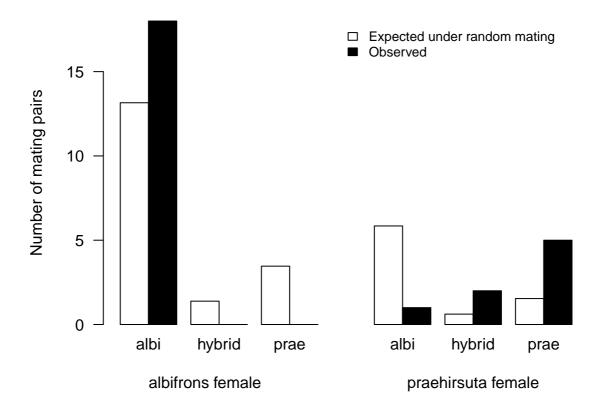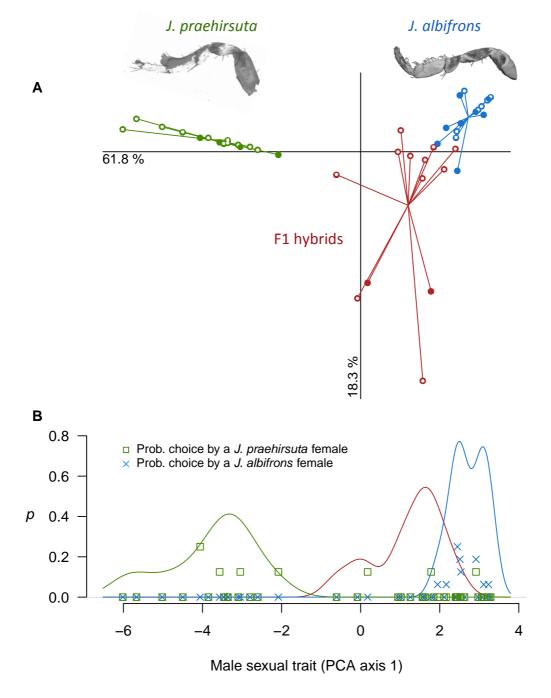
Figure 3

Figure 3 - Proportion of successful crosses in no-choice experiments involving intraspecific crosses (either *Jaera albifrons* or *J. praehirsuta*) and inter-specific crosses. The male and female of a given cross could come from the same region (Brittany or Normandy, see text) or from a different region (inter-reg. crosses). The sample sizes (number of experimental crosses) are given within brackets, and the bars give 95% confidence intervals around each observed proportion. A cross was successful if it produced at least one offspring. Interspecific crosses were consistently less successful than intraspecific ones, pointing towards sexual isolation between *J. albifrons* and *J. praehirsuta*.

Figure 4

Figure 4

Number of mating pairs of each possible type expected from random mating in absence of sexual

isolation (white bars) and observed from an experimental population (black bars). The expected

numbers take into account not only the number of males and females of each type that entered the

experiment but also their propensity to mate (see text). We see that *J. albifrons* females mated

successfully only with *J. albifrons* males, while *J. praehirsuta* females produced offspring with males

of the three different types, although not in equal proportions. These results point to strong but

imperfect, asymmetric sexual isolation.

Figure 5

Figure 5

Asymmetric sexual isolation between *Jaera albifrons* and *J. praehirsuta*. Panel A) shows the first two components of a principal component analysis of male phenotypes (potential parents in the multiple-choice experimental population) grouped by types. We see that individuals of the *albifrons* or *praehirsuta* types are phenotypically differentiated, while individuals of the hybrid type (i.e.

produced by an "interspecific" cross) show more phenotypic variability, including phenotypes

indistinguishable from the parental morphs. In this PCA plot, empty circles represent the males that

did not sire any offspring, while solid dots show the males that successfully reproduced. In panel B),

the solid curves show the density distribution of male sexual trait values in univariate space (PCA axis

1 only). The probability that a female mated with a male showing a particular sexual trait value is

shown by green squares (*J. praehirsuta* females) and blue crosses (*J. albifrons* females). The

preferred mates of *J. albifrons* females coincide with the distribution of *J. albifrons* male sexual trait.

By contrast, this concordance is much relaxed for *J. praehirsuta* females, which mated with males

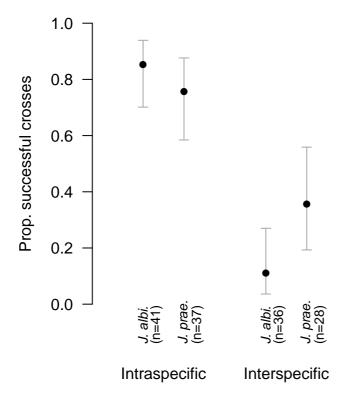showing a wide range of trait values.

Figure 6

Figure 6 - Proportion of successful intra- and interspecific crosses in no-choice experiments involving either *J. albifrons* or *J. praehirsuta* females. The sample sizes (number of experimental crosses) are given within brackets, and the bars give 95% confidence intervals around each observed proportion. A cross was successful if it produced at least one offspring. Sexual isolation between species is stronger in one direction (*J. albifrons* female).