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Solvent effects on the vibrational spectrum of 3-hydroxyflavone

Ari Paavo Seitsonen^{1,2,3,*}, Abdenacer Idrissi,⁴ Stefano Protti,⁵ Alberto Mezzetti^{6,*}

¹Institut für Chemie, Universität Zürich, Winterthurerstrasse 190, CH-8057 Zürich

²Département de Chimie, Ecole Normale Supérieure, 24 rue Lhomond, F-75005 Paris

³Sorbonne Université et Centre National du Recherche Scientifique (CNRS), F-75005 Paris

⁴LASIR UMR 8516, Université de Sciences et Technologies de Lille, Bat C5, Cité Scientifique, F-59655 Villeneuve d'Ascq cedex, France

⁵*PhotoGreen Lab, Department of Chemistry, University of Pavia, Viale Taramelli 10, I-*27100, Pavia, Italy

⁶Laboratoire de Réactivité de Surface UMR CNRS 7197, Sorbonne Université, 4 place Jussieu, F-75005 Paris, France

Abstract

3-hydroxyflavone (3HF) is a widely studied molecule that acts as a simplified prototype of biological, more complex flavonoids. Its solvation mechanism is still under investigation. Here we report a joint experimental and simulation study of the vibrational properties of 3HF in gas phase and in simple liquids tetrachloromethane, chloroform and acetonitrile using infra-red (IR) and Raman spectroscopies. We find reasonably good agreement between the static and molecular dynamics simulations employing density functional theory at the level of generalised gradient approximation (GGA) and hybrid functionals, but the agreement with the experimental spectra is only reasonable in the case of the IR spectroscopy and relatively poor in the case of Raman spectroscopy. The results can, however, be used as a starting point for discussing the solvation behaviour of the flavonoids.

Introduction

Flavonoids are natural compounds present in plants that exhibit several interesting biological properties, including, among others, radical scavenging ability [1], anti-carcinogenic [2] and antiproliferative [3]. Flavonols is a subclass of flavonoids which contain a 3-hydroxypyran-4-one ring. The chemical basis of their anti-oxidant properties have been the object of extensive research [4; 5 and refs therein]; in particular it has been found that the (micro)environment surrounding flavonols play a key role in modulating their reactivity [4; 6; 7; 8]. Interestingly, flavonol interactions with the surrounding environment deeply influence

their photophysics; for this reason, they are also widely used as fluorescent probes [9; 10]. Whereas intermolecular hydrogen bonding interactions determines flavonol emitting properties, environment-induced deprotonation of OH group leads to formation of anions whose fluorescence spectrum (band shape, color, intensity *etc*) is different from the neutral molecule [9; 11; 12].

Vibrational spectroscopy is a powerful approach to study the interaction between organic molecules and their surrounding environment, in particular proteins, at the atomic level. Vibrational spectroscopy can provide detailed information on pigment-protein / ligand-protein / cofactor-protein interactions, mechanism of ligand binding, mechanism of biochemical reactions etc [13]. Several vibrational spectroscopy techniques are widely applied in molecular biophysics to investigate these issues, such as resonant Raman spectroscopy [14], non-resonant Raman difference spectroscopy [15] and Fourier transform infra-red (FTIR) difference spectroscopy [16]. In these techniques, especially Resonance Raman, spectral contributions arising from the organic molecule can be identified, providing specific information on its interaction with the protein [14]. Molecular vibrations are sensitive to structure and intermolecular interactions (local dielectric constant, presence of hydrogen bonds etc); some vibrational marker bands are extremely sensitive probes [13]. A prerequisite to rationalise how the position of a vibrational band is influenced by the surrounding environment, is the detailed characterisation of the organic molecule in prototypical environments. To this aim organic solvents, which are characterised by precise physico-chemical parameters, are widely used. Experimental data on organic solvents can be fully exploited when calculations are used to assign vibrational modes or to rationalise solute-solvent interactions. Different approaches are used, such as density functional theory (DFT)-based calculations with an implicit solvent (see for instance [17]), DFT-based calculations with explicit solvent molecules (see for instance [18]) or QM/MM calculations (see for instance [19]).

In the last decade, a large number of studies have been carried out using vibrational spectroscopy to study flavonols in different microenvironments, *eg.* binding sites of biomacromolecules (see [20; 21; 22] and refs. therein), organic nanoparticles [23], liposomes [24] have been published. Its use in the study of chemical reactivity of flavonols has also been reported (see for instance [17; 25]). Vibrational spectroscopy of flavonols has also been successfully applied to food [26] and wine analysis [27]. Despite these efforts a clear interpretation the effect of the environment – through intermolecular interactions - on the position of the vibrational bands of flavonols is still lacking.



Scheme 1 Chemical structure of 3-hydroxyflavone (3HF)

Raman and IR spectra of the simplest among flavonols, 3-hydroxyflavone (3HF), have been used to investigate its interactions with several environments, such as glassy matrices [28], polysiloxanes [29] or zeolites [30]. There is further interest in the study of the interactions between 3HF and its environment due to its dual fluorescence [31], *ie*. from its normal state N*, and from its tautomeric state T*, where the proton - after an excited-state intramolecular proton transfer (ESIPT) reaction - is localised on the carbonyl oxygen. The efficiency and speed of ESIPT (and, as a consequence, on the relative intensity of the N* and T* fluorescence bands) depend strongly on the hydrogen bond (HB) interactions of 3HF with its environment [32]. In addition, as mentioned above, environment-induced deprotonation of the OH group of 3HF can occur, leading to anion formation with a completely different fluorescence spectrum ([33; 34] and refs. therein). Also the reactivity of 3HF [4; 35] and photochemistry ([36] and refs. therein) are strongly solvent-dependent. Static and time-resolved vibrational spectroscopy have been used to follow and/or rationalise 3HF photophysics [37] and irreversible photochemistry in solution [38; 39; 40], including the effect of the solvent on reaction mechanism.

The vibrational properties of 3HF have been the subject of several studies, with contrasting results: IR spectra were obtained in solid-state ([41] and refs. therein), in gas-phase [42; 43], in Ar matrices at 10 and 12 K [44; 45], in jet-cooled 3HF-(H₂O)_n clusters [46], and in solvents [37; 47; 48]. Raman spectra were recorded in various solutions [40; 48; 49], and SERS in water solutions [41] and resonance Raman spectrum of 3HF in 3-methyl-pentane (cited in [44]) were also reported. Calculations of IR and Raman spectra of 3HF have been reported, using the AM1 method [49] or the DFT approach [40-44]. Nevertheless, some discrepancies have been reported in precise band assignments.

In this Article, we present Raman spectra of 3HF in three solvents: tetrachloromethane (CCl₄), chloroform (CHCl₃) and acetonitrile (ACN). These were chosen because they range from apolar to polar, are all aprotic yet they impose a very different dielectric environment on

the 3HF, and they do not form (strong) hydrogen bonds with the solute, so as to avoid complications due to varying coordination of hydrogen bonds. We couple the present and former experimental findings with DFT simulations with an implicit solvent, in order to better understand the solvent effects on the main vibrational bands. In addition, in order to shed light on band attribution, we have simulated the IR spectrum of 3HF in gas phase and simple explicit solvent by a dynamical computational approach, relying on Car-Parrinello molecular dynamics analysis of 3HF in vacuum, and we compare the results to those obtained for 3HF in the gas phase [42].

Methods

Experiments

3HF was purchased from Aldrich, and re-crystallised from cyclohexane before use. Solvents were of spectroscopic grade. FT-Raman spectra were recorded on a Bruker IFS 66 V spectrometer. 1064 nm radiation from a Nd:YAG laser was used for excitation. In order to obtain spectra with a high signal to noise ratio 10000-40000 scans, depending on the solvent, were averaged. Spectra of pure solvents, obtained under the same conditions, were subtracted. Spectral resolution was 4 cm⁻¹. Concentration was 500 ppm (2,1*10⁻³ M) for all samples. A preliminary set of Raman experiments ($\lambda_{exc} = 632$ nm) was carried out at lower concentrations (down to 1*10⁻³ M) showing the same spectral features, ensuring thereby the absence of solute-solute interactions. This is also in line with a series of UV-Vis spectra of 3HF in the three considered solvents, showing the same spectral features in the concentration range 5*10⁻⁵ – 3*10⁻³ M (results not shown), in agreement with a previous report [48].

Calculations

We performed the dynamical DFT simulations using the QuickStep module [50] of the CP2K package [https://www.CP2K.org/]. We employed the generalised gradient approximation BLYP [51; 52] to the exchange-correlation functional is the Kohn-Sham equations, and the DFT+D3 dispersion correction in order to be consistent with simulations where an explicit solvent molecules might be used. We used the TZV2P-GTH Gaussian basis set for the orbitals and a plane wave basis set up to a cut-off energy of 400 Ry for density together with Goedecker-Teter-Hutter pseudo potentials [53] in the Gaussian plane wave (GPW) scheme [54]. We ran Born-Oppenheimer molecular dynamics with a time step of 0.5 fs

in a canonical ensemble, where the average temperature was kept at 298.15 K using a thermostat chain. 35 ps of trajectory was considered in the analysis, and the initial trajectory of 94 ps with the smaller basis set TZVP-GTH was discarded. A similar molecular dynamics run was also performed on the explicitly solvated 3HF in CCl₄ and ACN, with trajectories of at least 50 ps and the first 15 ps neglected. 128 solvent molecules were included in the simulation cell [55].

The static calculations with and without the implicit solvent were performed with the code TurboMole [56] within the harmonic analysis of the vibrations. We used the same BLYP+D3 approach as in the dynamical simulations together with simulations employing the B3LYP hybrid functional [57] with the D3 term added, and def2-TZVPP basis sets. The model for the implicit solvent was COSMO [58].

The infra-red (IR) spectra from the static calculations were simulated by Gaussian broadening at the calculated harmonic frequencies ω_i , marked in the graphs with circles. The IR intensity is calculated from the change in the dipole moment *D* along a displacement Δ_i the corresponding normal mode *i*, $I_{IR} \propto \sum_i |\partial D/\partial \Delta_i| \delta(\omega - \omega_i)$. Similarly the Raman intensity is evaluated from the differential of the polarisability along each normal mode.

Results

Vibrations in gas phase (isolated molecule)

We start by discussing the results in the gas phase, or isolated, molecule. In Figure 1 we have the computed IR and isotropic Raman spectra both from the dynamical and static calculations, the latter broadened by convoluting with a Gaussian function. Overall the agreement is very good, with an additional shoulder in the dynamical spectrum of IR at about 1585 cm⁻¹. In the spectra the strongest intensity is at around 1630 cm⁻¹. This feature is red-shifted in the dynamical simulations, most likely due to anharmonic effects that are not included in the static approach. The two methods yield similar spectra, so in the following we assume that the harmonic, static approach works relatively well.

[Figure 1 comes here]

Solvated 3HF

Next we discuss the experimental and calculated harmonic, static IR and Raman spectra in the gas-phase molecule and the simple solvents CCl₄, CHCl₃ and ACN. In the calculations we used both BLYP+D3 and B3LYP+D3 approaches to the exchange-correlation term, and in the simulation of solvated 3HF the implicit solvent. The results in Figure 2 require a careful analysis as the errors in the calculated spectra do not allow a straightforward assignment of the computed bands to the experimental peaks. We have fitted the scaling value of the computed frequencies so that best overall agreement is obtained between the experimental peak positions and computed frequencies leading to a value of 1.040 with BLYP+D3 and 0.991 with B3LYP+D3. Both the unscaled and scaled Frequencies are shown in Table 1. There are clear discrepancies between the calculated and measured spectra in the peak positions and the intensities. For example in the infra-red spectrum in ACN the calculated frequency of the highest-intensity peak is underestimated. In the FT-Raman spectra only one peak or frequency appears in the simulations at 1560-1610 cm⁻¹, whereas two peaks are present in the experimental spectra, possibly reflecting two different situations (*ie.* solute-solvent interaction scenarios). In ACN the agreement in the position of the computational main peak is worse than in the other (implicit) solvents. This might be due to the complicated equilibrium of solute-solvent HBs of 3HF in ACN, a solvent endowed with both hydrogen bond accepting basicity and also with some HB donating acidity. On the one hand, hydrogen bonding between the methyl group of ACN (HB donor) and the O_{CO} is possible, but on the other hand the intra-molecular hydrogen bond can also be distorted if the OH group forms HB as a donor with the solvent molecules acting as acceptors via the nitrogen atom.

[Figure 2 comes here]

[Table 1 comes here]

We performed a further comparison between the dynamical and static calculations with experiments by also computing the IR spectrum in the explicit solvent. These results are collected into Figure 3. The statistics is quite poor in the case of ACN, leading to very strong background signal, but we try to discern the signal from the weak maxima of the intensity. In the case of ACN we include also the experimental spectrum from Ref. [37], as this spectrum covers a wider spectral range than the spectra of solvated 3HF from Ref. [48]. This helps to compare the agreement of the lowest computational frequency at ~1580 cm⁻¹. Further experimental data are given in Table 2 as the peak positions. The harmonic, static and the

dynamical simulation again agree well in the case of gas-phase 3HF and solvated in CCl₄. In ACN the comparison is more difficult, possibly due to formation of hydrogen bonds between the solute and solvent molecules.

[Figure 3 comes here]

[Table 2 comes here]

In order to understand the character of the different vibrational modes we analysed the computational vibrational frequencies and the IR spectrum of two isotopically labelled 3HF molecules in gas phase with the harmonic-B3LYP+D3 approach, and from the dynamical properties of the C=O unit in the BLYP+D3 dynamical simulations. The latter were analysed using the Fourier transform of the bond length C=O, $d_{C=O}(t)$, shown in Figure 4, and the Fourier transform of the auto-correlation function of the velocity of the O_{C=O}. Thus the two quantities, results of which are given in Figure 5, indicate the contribution of the C=O stretching vibration to the power spectrum at different frequencies.

[Figures 4 and 5 come here]

The isotopic substitution of the H_{OH} to D leads to a large red shift of the highest frequency by about 10 cm⁻¹, and smaller lowering of the second-highest frequency. Thus the highest frequency has a component on the H_{OH} . The replacement of the ¹⁶O_{CO} by ¹⁸O_{CO} leads to larger changes in the IR intensity and the frequencies. Again the highest frequency decreases, indicating a contribution on the O_{CO}. On the other hand the peak with the highest intensity is the third highest in the ¹⁶O_{CO}-3HF, whereas in fourth highest in ¹⁸O_{CO}-3HF. Thus also this peak has a large weight on the O_{CO}.

Conclusions

We have reported computer simulations and experiments on the vibrational properties of 3HF in the gas phase and in different organic solution. Nearly all 3HF vibrational bands have shown to be very sensitive to the surrounding solvent. Interestingly, the behaviour is not the same for all bands when moving from 3HF solution in the apolar CCl₄ to the solution in the weakly polar CHCl₃ and to the polar ACN. This strongly suggests that specific solute-solvent interactions (hydrogen bonds) occur, especially in ACN. This is also the conclusion of a recent study based on UV-Vis and fluorescence spectroscopy in a large series of solvents, which underline that hydrogen bonds are relevant also in ACN, an organic solvent normally considered as aprotic [59]. Possibly the partial disruption of the intramolecular HB in 3HF plays an important role in modifying spectral properties. The effect of isotope substitutions on calculated spectra suggests that it is very difficult to assign a given band to a single localised vibration (*eg.* C=O stretching). A larger set of experimental vibrational data on isotopically-labelled 3HF in several solvents is needed to better understand this issue.

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[1] B.H. Havsteen, Pharmacol. Ther. 96 (2002) 67-202

[2] F.L. Juana, P.A.J. Angel, V.M. Manuel, in: R. Atta ur (ed)., Studies in Natural Products Chemistry, Elsevier (2012), pp. 115-134

[3] E. Middleton Jr, C. Kandaswami, T.C. Theoharides, Pharmacol. Rev. 52 (2000) 673-751

[4] M. Musialik, R. Kuzmicz, T.S. Pawlowski, G. Litwinienko, J. Org. Chem. 74 (2009) 2699-2709

[5] B. Butun, G. Topcu, T. Ozturk, Mini Rev Med Chem 18 (2018) 98-103

[6] M. Foti, G. Ruberto, G. J. Agric. Food Chem. 49 (2001) 342-348

[7] K. Sak, Mini Rev. Med. Chem. 14 (2014) 494-504

[8] C. Aliaga, M.C. Rezende, A. Arenas A, Food Chem. 113, (2009) 1083-87

[9] S. Protti, A. Mezzetti Photochemistry, 40 (2012) 295-322

[10] P.K. Sengupta, in Rev. Fluoresc. Geddes, C.D. (Ed), Springer UK 2016, pp. 45-70

[11] A. Mezzetti, S. Protti, C. Lapouge, J.P. Cornard, Phys. Chem. Chem. Phys. 13 (2011) 6858-6864

[12] S. Hofener, P.C. Kooijman, J. Groen, F. Ariese, L. Visscher Phys. Chem. Chem. Phys. 15 (2013) 12572-12581

[13] A. Barth, C. Zscherp, Q. Rev. Biophys. 35 (2002) 369-430

[14] B. Robert, Photosynth. Res. 101 (2009) 147-155

[15] R. Callender, H. Deng Ann. Rev. Biophys. Biomol. Struct. 23 (1994) 215-245

[16] A. Barth, Biochim. Biophys. Acta Bioenerg. 1767 (2007) 1073-1101

[17] Z. Jurasekova, C. Domingo, J.V. Garcia-Ramos, S. Sanchez-Cortes, Phys. Chem. Chem. Phys. 16 (2014) 12802-12811

[18] C. Marrassini, A. Idrissi, I. De Waele, K. Smail, N. Tchouar, M. Moreau, A. Mezzetti, J. Mol. Liq. 205 (2015) 2-8

[19] D. Bovi, A. Mezzetti, R. Vuilleumier, M.P. Gaigeot, B. Chazallon, R. Spezia, L. Guidoni, Phys. Chem. Chem. Phys. 13 (2011) 20954-64

[20] F.T. Liu, S.G. Agrawal, Z. Movasaghi, P.B. Wyatt, I.U. Rehman, J.G. Gribben, A.C. Newland, L. Jia, Blood 112 (2008) 3835-3846

[21] B. Sengupta, B. Pahari, L. Blackmon, P.K. Sengupta, Plos One 8 (2013) e65383

[22] S. Ionescu, I. Matei, C. Tablet, M. Hillebrand, Curr. Drug. Metab. 14 (2013) 474-490

[23] A. Barras, A. Mezzetti, A. Richard, S. Lazzaroni, S. Roux, P. Melnyk, D. Betbeder, N. Montfillette-Dupont Int. J. Pharm. 379 (2009) 270-277

[24] M. Huang, E. Su, F. Zheng, C. Tan, Food Funct. 8 (2017) 3198-3208

[25] Z. Jurasekova, A. Torreggiani, M. Tamba, S. Sanchez-Cortes, J.V. Garcia Ramos, J. Mol. Struct. 918 (2009) 129-137

[26] O. Abbas, G. Compère, Y. Larondelle, D. Pompeu, H. Rogez, V. Baeten, Vibr. Spectr. 92 (2017) 111-118

[27] C.A. Teixeira dos Santos, R.N.M.J. Pascoa, J.A. Lopes, Trends Anal Chem. 88 (2017) 100-118

[28] S. Protti, K. Raulin, O. Cristini, C. Kinowski, S. Turrell, A. Mezzetti, J. Mol. Struct. 993 (2011), 485-490

[29] M. Buffa, S Carturan, A. Quaranta, G. Maggioni, G. Della Mea, Opt. Mater. 34 (2012) 1219-1224

[30] A. Moissette, M. Hureau, A. Kokaislova, A. Le Person, J.P. Cornard, I. De Waele, I. Batonneau-Gener, Phys. Chem. Chem. Phys. 17 (2015) 26207-26219

[31] P.K. Sengupta, M. Kasha, Chem Phys Lett 68 (1979) 382-5

[32] A.J.G. Strandjord, P.F. Barbara, J. Chem. Phys. 89 (1985) 2355-61

[33] S. Protti, A. Mezzetti, J.P. Cornard, C. Lapouge, M. Fagnoni, Chem Phys Lett 467 (2008) 88-93

[34] D. Loco, N. Gelfand, S. Jurinovich, S. Protti, A. Mezzetti, B. Mennucci, J. Phys. Chem A 122 (2018) 390-397

[35] S.L. Studer, W.E. Brewer, M.L. Martinez, P.-T. Chou, J. Am. Chem. Soc. 111 (1989) 7643-7644

[36] S. Protti, A. Mezzetti, J. Mol. Liq. 205 (2015) 110-114

[37] K. Chevalier, M.M.N. Wolf, A. Funk, M. Andres, M. Gerhards, R. Diller, Phys. Chem. Chem. Phys. 14 (2012) 15007-15020

[38] P-T. Chou, S.L.Studer, M.L. Martinez, E. Orton, M. Young, Photochem. Photobiol. 53 (1991) 587-593

[39] M. Wang, T. Spataru, J.R. Lombardi, R.L. Birke, J. Phys. Chem. C.111 (2007) 3044-3052

[40] S Protti, A Mezzetti, C Lapouge, JP Cornard Photochem. Photobiol. Sci. 7 (2008) 109-119 [41] M. Wang, T. Teslova, F. Xu, T. Spataru, J.R. Lombardi, R.L. Birke, M. Leona, J. Phys. Chem. C 111 (2007) 3038-3043

[42] A. Vavra, R. Linder, K. Kleinermanns, Chem. Phys. Lett. 463 (2008) 349-352

[43] K. Bartl, A. Funk, K. Schwing, H. Fricke, G. Kock, H-D. Martin, M. Gerhards Phys. Chem. Chem. Phys. 11 (2009) 1173-1179

[44] J.M. Petroski, C. De Sa Valente, E.P. Kelson, S. Collins, J. Phys. Chem. A 106 (2002) 11714-11718

[45] W.E. Brewer, S. L. Studer, P T. Chou, Chem. Phys. Lett. 158 (1989) 345-350

[46] K. Bartl, A. Funk, M. Gerhards, J. Chem. Phys. 129 (2008) 234306

[47] N.A. Tyukavkina, N.N. Pogodaeva, E.I. Brodskaya, Y.M. Sapozhnikoy, Chem. Nat. Compd. 11 (1975) 613-616

[48] L. Vrielynck, PhD Thesis (1996) Université de Sciences et Technologies de Lille

[49] J.P. Cornard, L. Vrielynck, J.C. Merlin, J.C. Wallet Spectrochim. Acta. A. 51 (1995) 913-923

[50] J. VandeVondele, M. Krack, F. Mohamed, M. Parrinello, T. Chassaing, J. Hutter, Quickstep: Fast and accurate density functional calculations using a mixed Gaussian and plane waves approach, Comput. Phys. Commun. 167 (2005) 103

[51] A. D. Becke, Density-functional exchange-energy approximation with correct asymptotic behavior, Phys. Rev. A 38 (1988) 3098

[52] C. Lee, W. Yang, and R. G. Parr, Development of the Colle-Salvetti correlation-energy formula into a functional of the electron density, Phys. Rev. B 37 (1988) 785

[53] S. Goedecker, M. Teter, and J. Hutter, Separable dual-space Gaussian pseudopotentials, Phys. Rev. B 54 (1996) 1703

[54] G. Lippert, J. Hutter and M. Parrinello, The Gaussian and augmented-plane-wave density functional method for ab initio molecular dynamics simulations, Theor. Chem. Acc. 103 (1999) 124

[55] A. P. Seitsonen, A. Idrissi, S. Protti, A. Mezzetti, in preparation

[56] TURBOMOLE V7.1 2016, a development of University of Karlsruhe and Forschungszentrum Karlsruhe GmbH, 1989-2007, TURBOMOLE GmbH, since 2007; available from http://www.turbomole.com/

[57] P.J. Stephens, F. J. Devlin, C. F. Chabalowski and M. J. Frisch, Ab Initio Calculation of Vibrational Absorption and Circular Dichroism Spectra Using Density Functional Force Fields, J. Phys. Chem. 98 (1994) 11623–11627

[58] A. Klamt; G. Schüürmann. COSMO: A new approach to dielectric screening in solvents with explicit expressions for the screening energy and its gradient. J. Chem. Soc. Perkin Trans. 2 (1993) 799–805. *doi:10.1039/P29930000799*

[59] S. Lazzaroni, D. Dondi, A. Mezzetti, S. Protti, Role of solute-solvent hydrogen bonds on the ground state and the excited state proton transfer in 3-hydroxyflavone. A systematic spectrophotometric study, Photochem. Photobiol. Sci. 17 (2018) 923-933. *doi:10.1039/C8PP00053K*

Table 1. The computed six vibrational frequencies of 3HF in different environments using an implicit solvent in the frequency range of interest here; both unscaled, *ie* original, and scaled frequencies are shown

Method							
Unscaled							
Gas phase	BLYP +D3	1534	1551	1568	1581	1588	1598
	B3LYP +D3	1604	1613	1641	1648	1655	1678
CCl4	BLYP +D3	1529	1550	1559	1579	1588	1592
	B3LYP +D3	1601	1613	1635	1643	1655	1668
CHCl3	BLYP +D3	1524	1551	1555	1577	1588	1590
	B3LYP +D3	1598	1613	1631	1642	1654	1664
ACN	BLYP +D3	1519	1551	1554	1577	1587	1590
	B3LYP +D3	1595	1614	1626	1642	1656	1661
Scaled							
Gas phase	BLYP +D3	1596	1613	1630	1644	1651	1662
	B3LYP +D3	1589	1598	1626	1633	1640	1663
CCl4	BLYP +D3	1590	1612	1621	1642	1652	1656
	B3LYP +D3	1587	1598	1620	1628	1640	1653
CHCl3	BLYP +D3	1585	1613	1618	1640	1651	1654
	B3LYP +D3	1583	1599	1616	1628	1640	1649
ACN	BLYP +D3	1579	1613	1616	1640	1651	1654
	B3LYP +D3	1581	1599	1612	1627	1641	1646

Table 2. Experimental IR and Raman spectra of 3HF in different environments

Normal mode	C=O stretch;	Ring B quinoid	C=O stretch;		C=O stretch;
according to Ref [39]	OH bending; C=C stretch	stretch; OH bend; C=C	C=C stretch		OH bending; C=C stretch
		stretch			
IR Bands					
Solid state [41]	1562s	1602s	1610	1627w	1647 vw
Ar matrix 10 K [44]				1628	1652w
Ar matrix 12 K [45]	1578		1618 sh	1630	1653 ; 1659sh
gas phase [42]	1570		1618	1638	(1656w)
gas phase [43]			1624	1630	1646/1653
CCl4 [47]			1622		1652
CCl4 [48]			1620		1650
CHCl3 [48]			1613		1643
CD ₃ CN [48]			1620		1641
Raman bands					
Solid state [41]	1566vs	1594vs	1619vs		
CCl4 [49]					1648
MeOH [40]	1569	1600	1614	1622	
MeOH/H2O 10% [40]	1566	1600	1614	1622	
Nanoporous SiO ₂ [28]	1568, 1558w	1604	1615	1627w; 1639	1651
Methylpentane Res Raman [44]			1625		1650
CCl4 present work	1574	1600	1620	1632w	1648
CHCl3 present work	1569	1599	1613		1639 (broad)
CH ₃ CN present work	1571	1600	1621		1639 (broad)
Normal mode according to [39]	C=O stretch; OH bending; C=C stretch	Ring B quinoid stretch; OH bend; C=C stretch	C=O stretch; C=C stretch		C=O stretch; OH bending; C=C stretch

Figure 1 Computed vibrational a) IR and b) Raman spectra of an isolated 3HF molecule from CPMD trajectories (solid lines) and harmonic analysis (dashed lines).



Figure 2 Experimental (black, dashed line) and simulated (BLYP+D3: solid lines and multiplecoloured circles; B3LYP+D3: dot-dashed lines and green circles) a) IR and b) Raman spectra of 3HF in gas phase (gp) and solvated in CCl₄, CHCl₃ and ACN. The intensities of the gasphase calculations were used in Panel b). The experimental data are from Ref. [42] (IR, gasphase), Refs. [37,48] (IR, solvated) and present work (Raman).

Figure 3 Experimental (black dashed lines) and simulated BLYP+D3 dynamical (solid lines) and harmonic (dot-dashed lines, circles) IR spectra. The dashed lines are from the experimental spectrum from Ref. [42] (gas-phase 3HF) and Ref. [48] (solvated 3HF) and the purple dot-dashed line in ACN from Ref. [37].



Figure 4 IR spectra of different isotopically–labelled 3HF in the gas phase calculated with the harmonic-B3LYP+D3 approach.



Figure 5 Power spectrum of the C=O units from the Fourier transform of the C=O bond (solid lines) and the auto-correlation function of velocity of the O_{CO} (dashed lines) in different solvents from the dynamical simulations. The harmonic frequencies are given at the top.