

Iron oxide nanoparticles for the rapeutic applications Edouard Alphandéry

▶ To cite this version:

Edouard Alphandéry. Iron oxide nanoparticles for the rapeutic applications. Drug Discovery Today, 2020, 25 (1), pp.141-149.
 10.1016/j.drudis.2019.09.020 . hal-02441707

HAL Id: hal-02441707 https://hal.sorbonne-universite.fr/hal-02441707v1

Submitted on 21 Jul2022

HAL is a multi-disciplinary open access archive for the deposit and dissemination of scientific research documents, whether they are published or not. The documents may come from teaching and research institutions in France or abroad, or from public or private research centers. L'archive ouverte pluridisciplinaire **HAL**, est destinée au dépôt et à la diffusion de documents scientifiques de niveau recherche, publiés ou non, émanant des établissements d'enseignement et de recherche français ou étrangers, des laboratoires publics ou privés.



Distributed under a Creative Commons Attribution - NonCommercial 4.0 International License

Version of Record: https://www.sciencedirect.com/science/article/pii/S135964461930380	0
Manuscript 3f8bc040cb4bb39b939576aee4ff9fea	

1 Iron oxide nanoparticles for therapeutic applications

- 2 Edouard Alphandéry
- 3

4

5 Minéralogie, de Physique des Matériaux et de Cosmochimie, IMPMC, Place Jussieu, 75005 Paris, 6 France 7 Nanobacterie SARL, 36 boulevard Flandrin, 75116, Paris, France 8 Institute of Anatomy, UZH University of Zurich, Institute of Anatomy, Winterthurerstr. 190, CH-8057, 9 Zurich, Switzerland 10 Corresponding author: Alphandéry, E. (edouardalphandery@hotmail.com). 11 12 Keywords: Iron oxide nanoparticle, medical application, cancer, magnetic hyperthermia, therapy, 13 nanotechnology, oncology, nanomedicine. 14 15 16 Teaser: Owing to their reassuring toxicity profiles and multiple mechanisms of action, iron oxide nanoparticles have been considered for a wide range of medical applications, including treatment of 17 18 iron-deficiency anemia, cancer, Parkinson's, Alzheimer's, antibacterial and antifungal diseases. 19 20

Sorbonne Université, Muséum National d'Histoire Naturelle, UMR CNRS 7590, IRD, Institut de

In nanomedicine, iron oxide nanoparticles are at an advanced stage, being commercialized for cancer treatment and iron-deficiency anemia treatment. Their therapeutic efficacy comes from their ability to target a tissue, activate a drug, locally produce reactive oxygen species or a temperature increase following (or not) the application of an external source of energy, modify genes or activate various biological materials, or replace diseased cells by stem cells. Owing to these various mechanisms of action, they can potentially be used for treating a whole range of different diseases, making them more appealing than conventional drugs that target a more limited number of indications.

29

21

31 Introduction

Nanomaterials have been developed for medical applications owing to their appealing properties. First, 32 33 their size is of the same order of magnitude as that of various biological materials such as enzymes, 34 proteins and lipids, hence promoting interactions with them, and they are larger than most individual 35 molecules used as standard drugs, resulting in better tissue interaction and larger accumulation in 36 targeted organs for nanomaterials than for standard drugs. Second, nanomaterials have a large exposed 37 surface area, which enables them to adjust their properties by changing their surface charge, 38 composition or chemistry, or by binding to their surface molecules of interest such as specific ligands or chemotherapeutic molecules. 39

40 These properties, which are stimulating from the point of view of research, make it hard to set up a well-41 established regulatory framework, hence delaying the use of nanomedicine for human treatments. In 42 addition, because each group working on a different nanomaterial tries to highlight the advantages of its 43 system compared with others, it is difficult to identify a nanosystem that is better than others. However, 44 iron oxide nanoparticles (IONPs) have paved the way among the most widely studied nanomaterials, owing to their biocompatibility and multiple ways in which they can trigger therapeutic activity. In this 45 review, after a brief presentation of IONP manufacturing methods and physicochemical properties, the 46 47 different mechanisms of action as well as treatments for specific diseases in which IONPs are involved are described. 48

49

50 **Fabrication and physicochemical properties of IONPs**

51 Methods to synthetize IONPs include: (i) co-precipitation between Fe^{2+} and Fe^{3+} in basic conditions [1]; 52 (ii) thermal decomposition of organo-iron in high boiling point organic solvents in the presence of 53 surfactants [2]; (iii) nanoparticle formation inside micelles [3]; (iv) sol-gel methods [4]; (v) 54 hydrothermal synthesis [5]; and (vi) cathodic electrochemical deposition in which anode oxidation leads 55 to the formation of metal ions that are further reduced to metals by the cathode in the presence of a 56 stabilizing material [6]. These synthesis methods enable obtaining IONPs with various sizes ~1–120

nm, size distributions, compositions [essentially consisting of Fe_3O_4 (magnetite) and γ -Fe₂O₃ 57 58 (maghemite)], crystallinity (amorphous or crystalline structure), magnetic properties (essentially 59 superparamagnetic for IONPs <10-20 nm and ferrimagnetic for IONPs >10-20 nm) and geometries (spherical, round, square, cubic, cubo-octahedric, needle, cylindrical, cigar, bullet type). To prevent 60 IONPs from aggregating, which can lead to embolism, these nanoparticles can be coated by various 61 62 materials including polymers and fatty or amino acids. Furthermore, the coating material can influence 63 nanomaterial biodistribution by controlling opsonization mechanisms; the capacity of specific cells such as macrophages to capture IONPs, for example polyethylene glycol (PEG), leads to a longer blood 64 65 circulation time than citric acid [7-10].

66

67 IONP targeting methods

For IONPs to be efficient, they need to reach the organ of interest. As illustrated in Figure 1, this can be
 achieved through three different types of targeting mechanisms:

- Passive targeting also called enhanced permeability and retention (EPR) effect in which IONPs
 target tumors by passively diffusing through the holes of abnormal vessels resulting from
 angiogenesis that irrigate the tumor.
- Active targeting in the absence of magnetic field application, which relies on the use of IONPs 73 • 74 conjugated with a targeting ligand (TL), which specifically targets a specific receptor that is overexpressed at the surface of cancer cells (T). Examples of pairs (TL:T) are: (i) 75 76 antibody:antigene, such as antibody-Her-Neu:receptor-Her-2-neu [11]; (ii) aptamer:tumor-cellreceptor, such as anti-Muc-1:Muc-1 [12]; (iii) protein:tumor-cell-receptor, such as RGD: $\alpha\nu\beta_2$ 77 78 [13]; (iv) peptide:various-tumor-parts, such as F3::tumor-vasculature [14]; (v) enzyme:cellreceptor, such as uPA:uPAR [15]; (vi) vitamin:tumor-cell-receptor, such as folic-acid:folate-79 80 receptor [16]; (vii) chemical-compound:tumor-cell-receptor, such as methotrexate:folate-

- 81 receptor [17]; and (viii) hormone:tumor-cell-receptor, such as luteinizing hormone releasing 82 hormone (LHRH):LHRH-receptor [18].
- Magnetic drug targeting, in which a strong external magnetic field gradient is applied to drive
 IONPs into the area of interest.

Taking the literature as a whole, it appears that the targeting efficacy is relatively similar for different targeting, yielding a percentage of injected IONPs in the tumor of 5.10^{-4} % to 3%, 0.1% to 7% and 5.10^{-3} % to 2.6% for passive, active and magnetic targeting, respectively [19]. Next, the different mechanisms through which IONPs can generate a therapeutic activity, as summarized in Figures 1 and 2, are reviewed.

90

91 **IONP drug delivery without a mechanism of drug release**

92 Although, in several cases, drug delivery using IONPs was described without mentioning a specific 93 mechanism of drug release, this does not mean that such a mechanism did not occur. Disregarding such 94 a mechanism, the conjugation of various chemotherapeutics to IONPs can be used to deliver these drugs 95 to tumors, yielding the following advantages compared with the injection of fee drugs: (i) the possibility 96 to localize IONPs and therefore also possibly the drug conjugated to IONP, using MRI for example; (ii) 97 the increase in drug accumulation in the tumor, for example IONPs containing doxorubicin (DOX) led to a higher concentration of DOX in tumor cells compared with free DOX treatment in an ovarian 98 99 cancer model [19]; (iii) the association of drugs to IONPs can improve antitumor activity, for example 100 IONPs conjugated with vinblastine led to enhanced MCF-7 breast cancer cell inhibition compared with 101 vinblastine alone [20]; (iv) the combination of IONPs with a chemotherapeutic drug such as 102 gemcitabine or mitoxantrone and a targeting agent such as anti-CD44 or prostate-specific membrane 103 antigen (PSMA) antibodies can enhance drug targeting and antitumor efficacy on tumor cells compared 104 with free drugs [21-24]; and (v) IONPs can be combined with a specific entity to prevent DNA pro-105 tumor activity, for example DNA was captured by cytarabine attached to IONPs [25].

106

Drug delivery and activation with a mechanism of drug release

108 Delivery and activation of drugs associated to IONPs can occur via various mechanisms:

pH variation, which has been described for drugs such as 5-FU, curcumin, DOX and tamoxifen that either involve magnetic-polymer, PEG or IONPs inserted within the large pores of mesoporous hydroxyapatite-coated IONPs, or bind to monoclonal antibody (mAb)-conjugated magnetic nanocarriers, where release is more pronounced at an acidic cellular pH of 5.8 than at the blood pH of 7.4, leading to an increase of the percentage of drug release from 10–55% at pH 5.8 [26,27].

115 Temperature variation, which has been highlighted in systems containing: (i) curcumin ٠ 116 incorporated in a nanogel containing a thermosensitive polymer (PNIPAM) and IONPs, 117 releasing 35% and 70% of curcumin at 37°C and 41°C [28]; (ii) DOX bound to IONPs through 118 an Azo linker, which is destroyed above $\sim 43^{\circ}$ C, when exposed to near infrared (NIR) irradiation, leading to DOX release and inhibition of tumor growth inhibition [29]; (iii) 119 120 gemcitabine encapsulated together with IONPs inside a phospholipid bilayer to form a magneto-121 liposome, leading to an increase of drug release from 17% at 37°C without alternating magnetic 122 field (AMF) application to 70% following AMF application [30].

Magnetic field through either an indirect mechanism (i.e., the magnetic field causes nanoparticle cellular internalization of a drug bound to IONPs, such as BCNU-loaded nanocomplex in HG cells, resulting in drug release in the acidic cellular environment) [31] or a direct mechanism {i.e., the magnetic field itself causes drug release, as observed for IONP nanocapsules containing drugs surrounded by thermo-labile polymer (PEO-PPO-PEO) [32], IONP surrounded by a hydrophobic poly(vinyl alcohol) coating containing hydrophilic DOX, fluid MAG-CMX conjugated to DOX or magnetic polymerosomes loaded with DOX [33]}.

• Water or buffer surrounding IONPs that can trigger drug release as observed for 5-FU or curcumin progressively detaching from IONPs in water or different buffer solution [34],

- possibly leading to enhanced cytotoxicity against tumor cells as highlighted for F@BSA@CUR
 in MCF-7 cells [34].
- IONP shape can have an impact on drug release mechanism, as observed for 5-FU bound to
 IONPs of various shapes [35].
- Drug circulation time in the organism can influence drug release, as observed for polyplex coated magnetite (Nano-co-Plex) loaded with BCNU, which shows a time-dependent release
 efficacy with maximum release of 75% of the loaded BCNU [36].
- 139 Breaking of the bond between the drug and the IONPs, using either acidic conditions or specific 140 biological materials (proteinase), as described for: (i) L-lysine surface-coated IONPs conjugated 141 with methotrexate (MTX), where the peptide bond is cleaved in the presence of proteinase K at low pH [37]; (ii) for IONPs bound to polyethylene glycolated methotrexate (MTX-PEG) and 142 143 NIR fluorescent cyanin dye (Cy5.5) that could be hydrolyzed at intracellular pH, preventing 144 drug release in systemic circulation while allowing it inside endosomes or lysosomes [38]; (iii) 145 for IONPs linked to drugs through bounds that are cleaved by tumor enzyme-cleavable peptides 146 such as lysosomal proteases [39].

These mechanisms of selective drug release can prevent side effects as highlighted when the highly cytotoxic drug azademethylcolchicine (ICT) was conjugated to the dextran layer of the IONP ferumoxtol using a peptide linker, which was cleavable by matrix metalloproteinase-14 (MMP-14), leading to significant tumor cell apoptosis while sparing healthy cells [40].

151

152 Intracellular drug delivery with IONPs

153 If a drug is not delivered intracellularly it can lose its activity, for example by diffusing in the 154 bloodstream or by being eliminated. Drugs associated with IONPs can more easily internalize in cells 155 than free drugs because the size of the drug–IONP complex is significantly larger than that of free 156 drugs, a ligand can be bound to the IONP surface that specifically binds a cell receptor. As an example, 157 direct intracellular drug delivery of DOX by IONPs was shown to be much more efficient than that of

158 free DOX for targeting and concentrating DOX within a Hep-G2/MDR tumor site [41].

159

160 **IONP therapy working through the production of ROS**

IONPs could trigger some activity by releasing radical oxygen species (ROS) as demonstrated either 161 when IONPs are dissolved into Fe^{2+} and/or Fe^{3+} , for example following IONP cellular internalization 162 163 and degradation by lysosomes [42], or when a substance bound to IONPs produces ROS, as shown for cisplatin attached to IONPs, which activated nicotinamide adenine dinucleotide phosphate oxidase 164 165 (NOX), transforming di-oxygen (O_2) to superoxide radical ($O_2 \bullet -$) and H_2O_2 , resulting in the apparition of toxic hydroxyl radicals (•OH) through the Fenton's reaction between H_2O_2 and Fe^{2+} [43]. 166 167 Furthermore, ROS, such as •OH, have been shown to induce oxidative damage to lipids, proteins and 168 DNA in the tumor site while minimizing systemic toxicity [43].

ROS can also be produced by exposing a photosensitizing agent associated with IONPs to a laser through a technique called photodynamic therapy (PDT). In this case, IONPs are used as a platform or support for binding a large number of photosensitizing agents such as hypericin. The efficacy of such an approach was demonstrated on a complex of red blood cells, DOX, IONPs and chlorine e6 that significantly suppressed tumor growth using PDT [44].

174

175 IONP therapy based on local temperature increase (magnetic hyperthermia and photothermal176 therapy)

177 Localized heat can be produced by exposing IONPs to the following:

• An AMF using a technique called magnetic hyperthermia (MHT), where the AMF should have frequency and strength >20–100 kHz and 1-10 mT, respectively, and IONP should be sufficiently concentrated (i.e., typically above 1 mg of IONP per cm³ or ml of surrounding medium) [41], resulting in specific absorption rate (SAR) of typically 1 to 1000 W/gFe, depending on IONP properties (size, anisotropy stability of magnetic moment) and AMF parameters, leading to a series of different therapeutic activities (i.e., specific tumor cell destruction without destruction of healthy cells) [41,42]; (ii) significant or full tumor destruction on mice bearing different tumor types (e.g., subcutaneous or intracranial) [45–50]; (iii) activation of a drug such as DOX [51]; (iv) an improvement of GBM patient survival by ~7 months [44,52]; (v) significant lesion reduction by implanting a composition of IONPs and calcium phosphate cement in bone tumors [53]. A typical protocol used for MHT treatment is illustrated in Figure 3.

A laser using photothermal therapy (PTT), which produces a localized surface plasmon wave using laser wavelengths in the NIR within 650–900 nm to avoid harmful light absorption by healthy tissues, and laser power is maintained below the safe limit of tissue irradiation of 0.3–1.0 W/cm², resulting in the destruction of tumor cells (e.g., MCF-7 and MDA-MB-231) or full or partial tumor disappearance in mouse without apparent side effects, where therapeutic activity was observed in the absence and in the presence of an absorbing material such as PPy [28].

These nanoparticle-based heating techniques present the advantage of moderately heating the tumor during treatments at typically 41–55°C, which limits side effects compared with 80–90°C reached in cancer thermotherapies that do not use nanoparticles such as HIFU and radiofrequency.

Mechanisms to explain antitumor efficacy with MHT and PTT rely on: (i) structural and functional 199 200 cancer cell damage, specifically at the level of protein, membrane and cytoskeleton, which can lead to 201 apoptosis [54]; (ii) 'fragilization' of cancer cells which increases the sensitivity of these cells to other 202 treatments such as chemotherapy or radiotherapy [55]; (iii) expression of heat shock proteins at temperatures of typically 41–46°C that can attack the tumor [56], although the opposite behavior 203 204 (protection of the tumor) has also been reported for these proteins [57]; (iv) protein denaturation or 205 unfolding that can occur at high temperature; (v) pH changes, perfusion and oxygenation of the tumor 206 microenvironment that can lead to tumor necrosis [58]; and (vi) apoptosis or necrosis depending on 207 treatment parameters (temperature, duration of heating, etc.) [45].

209 Triggering gene therapy with IONPs

IONPs could potentially improve gene therapy efficacy by carrying specific nucleic acids that regulate altered gene expression resulting from carcinogenesis, essentially in the form of siRNA or plasmid DNA. An IONP–gene complex could have a larger circulation time than that of a free gene, avoid potential damages by nucleases and hence enable nucleic acids delivery to the organ of interest such as a tumor [59]. Such therapy could act against a tumor in the following ways:

- Directly, for example when IONPs conjugated with siRNA (siPLK1) act against Pololike kinase 1 (cell-cycle-specific serine/threonine kinase) and two peptides (MPAP and MUC1) were
 injected in tumor-bearing mice, the result is the accumulation of IONPs in the tumor, efficient
 PLK1 silencing and tumor suppression through increased apoptosis [60].
- Indirectly, for example when IONPs carrying the phosphatase and tensin homolog (PTEN) gene
 increased the sensitivity of A549/CDDP lung cells to cisplatin treatment indicating that PTEN
 can effectively be used against cisplatin-resistant lung cancer cells [61].

222 Furthermore, to improve the efficacy of gene therapy, the IONP–gene complex can be attracted near the 223 tumor by a magnet, hence increasing gene transfer into cells [62], via a technique called magnetofection. 224 To date, clinical trials testing gene therapy have been unsuccessful owing to the existence of a multitude 225 of different genes responsible for cancer, depending on cancer type, cancer progression and type of 226 cancer patient, which can hardly all be targeted with only one gene therapeutic agent [63]. To further 227 improve the efficacy of gene therapy, it might be suggested to use IONPs to: (i) carry out a pre-228 screening of pro-tumor genes for each patient taken individually and adapt gene therapy depending on 229 the detected genes; (ii) use a combination of different genes instead of only one gene to increase the 230 probability of targeting pro-tumor genes; or (iii) combine gene therapy with another modality of 231 treatment.

232

233 Stem cell therapy using IONPs

IONPs can be used in stem cell therapy by being incorporated inside these cells, enabling them to localize, for example using MRI, and activating them with the help of magnetic actuation [64]. By using IONPs during the transplantation of oligo-dendrocyte precursor cells (OPCs), it might be possible to help in regenerating the central nervous system by increasing myelin production and hence restore the protective sheath around nerve fibers [65].

239

240 **Immunotherapy**

241 IONPs can act on the immune system in the following ways:

They modulate macrophage polarization (i.e., inducing a shift from M2-type macrophages with anti-inflammatory and pro-tumor activity to M1-type macrophages with anti-tumor and proinflammatory activity), as was shown for example for THP-1 monocytes that shifted from a M2-like phenotype before IONP exposure toward a M1-like phenotype after IONP exposure 246 [66].

• They increase the efficacy of immune-suppressive drugs used during organ transplantation, as was shown when mycophenolic acid (MPA) combined with IONPs and effectively inhibited the secretion of the cytokines interleukin (IL)-2 and tumor necrosis factor (TNF), with a concentration of MPA that is lower than that necessary to reach a similar effect in the absence of IONPs [67], hence reducing side effects observed at high MPA concentrations.

252

253 Protein and enzyme activation or removal with IONPs

It has been shown that IONPs can act like enzymes by mimicking the activity of peroxidase [60] and efficiently immobilize proteins at their surface owing to their large surface-area:volume ratio, and then separate or isolate proteins by applying an external magnetic field [68].

257

Different types of diseases treated with IONPs

A summary of the different types of diseases that can be treated with IONPs and the mechanisms of action that are involved in the treatments is provided in Figure 4 and Table 1. Cancer can be treated through the generation of ROS or localized heat by IONPs or via a chemotherapeutic drug associated with IONPs, where the IONP–drug complex can lead to the following:

- An enhanced efficacy of chemotherapeutic drug, for example: (i) IONPs associated with
 paclitaxel (IONP–PAX) are more cytotoxic than free PAX in destroying prostate tumor cells, as
 demonstrated *in vitro* on PC3 and CWR22R human prostate cancer cells [69] and *in vivo* on C4 2cell-derived xenograft tumors in athymic nude mice [69]; and (ii) IONPs associated with DOX
 are more cytotoxic toward 4T1 cells than free DOX [70].
- A reduction of toxicity of certain anticancer drugs such as docetaxel, mitoxantrone, cisplatin and carboplatin by increasing the efficacy of tumor drug delivery and by simultaneously reducing the dose necessary to reach therapeutic efficacy, as shown for IONPs conjugated to docetaxel, which led to enhanced cellular uptake ability and antiproliferative activity in PC3 prostate cancer cells, and for toxic mitoxantrone where tumor uptake was increased when it was conjugated with IONPs [69], overcoming the cytotoxicity induced by certain chemotherapeutics (cisplatin, carboplatin).
- An increase in drug-targeting efficacy through the use of: (i) stimuli-responsive drug release from IONPs, for example with a higher drug release at pH 5.8 (mimicking the tumor microenvironment) than at pH 7.4 (normal physiological condition); (ii) targeting agents that are able to target specific receptors of cancer cells as summarized above.
- Chemosensitization of tumor cells to chemotherapeutic drug, as was shown through the
 cytotoxicity enhancement of cisplatin toward cisplatin-resistant A549 tumor cells using IONPs
 loaded with cisplatin [71].

In addition to the mechanisms described above, it has been reported that IONPs can modify the tumor microenvironment by blocking an artery that irrigates the tumor with blood [72]. Alzheimer's disease (AD) could be treated by IONPs functionalized by a targeting agent that can cross the blood-brain barrier (BBB) and reach small peptides called amyloid- β (A β) and then either detect A β using MRI or affect the A β fibrillation process, responsible for AD, with an efficacy that seems to depend on IONP size, surface area, charge, concentration and level of opsonization [73]. Parkinson's disease (PD) could be treated by IONPs using the following:

Gene therapy, by conjugating IONPs with short hairpin RNA (shRNA), promoting neuronal
 uptake via nerve growth factor (NGF) receptor-mediated endocytosis or neuron apoptosis caused
 by a lower α-syn expression *in vitro* and *in vivo*, leading to effective repair in a PD model [74].

Magnetic field therapy, by applying a magnetic field on rats at the site of PD lesion 24 h after
 IONP administration, 2 h/day for 1 week, leading to an improved behavior, condition,
 mitochondrial function and attenuation of lesion volume in all treated rats, indicating the
 neuroprotective effect of this therapeutic approach [75].

296 Neuro-AIDS, which refers to a reservoir of AIDS viruses in the brain, could potentially be treated by 297 attaching conventional treatments (tenofovir and vorinostat) to IONPs to enable these drugs to cross the 298 BBB and to efficiently eradicate AIDS viruses in the brain, as demonstrated by IONP antiviral activity 299 *in vitro* over a period of 5 days after HIV infection in primary human astrocytes [76]. Iron-deficiency 300 anemia, which cannot be efficiently cured with oral iron treatment owing to drug gastrointestinal 301 digestion, could be treated with intravenous injection of IONPs, improving iron biodisponibility; and this has been commercialized in formulations such as Dexferrum[®], Feraheme[®], Ferrisat[®], Ferriecit[®], 302 Monofer[®] and Venofer[®] [19]. 303

Fungi or bacterial contamination can be eradicated by inhibiting the growth of fungi such as *Trichophyton verrecosum, Trichophyton mentagrophytes, Dermatophilus sp., Trichothecium roseum, Cladosporium herbarum, Penicillium chrysogenum, Alternaria alternata and Aspergillus niger* [77] or bacteria such as *Escherichia coli, Bacillus subtilis, Candida albicans, Aspergillus niger and Fusarium solani* [78]. Growth inhibition can be the result of leakage of the cell contents leading to cell death, inhibition of spore germination, membrane destruction, production of ROS with lipid peroxidation, DNA damage, release of metal ions and/or the presence of antibacterial or antifungal substances

associated with IONPs such as vanilla essential oil, antibiotics and chlorhexidine (CHX). Furthermore, 311 312 IONPs can penetrate inside biofilms and hence yield efficient biofilm destruction, a property that seems 313 to depend upon IONP physicochemical characteristics, such as IONP surface charge, hydrophobicity 314 and high surface-area:volume ratio [79]. Blood contamination can be treated by attaching IONP 315 molecules that specifically bind to contaminant material, such as pathogens or toxic compounds, as 316 shown for IONPs colliding with Staphylococcus aureus, internalizing inside these bacteria and then 317 being removed under magnetic field application [80]; or for IONPs surrounded by a specific coating that 318 can bind radioactive material or heavy metal.

319

320 Concluding remarks and future perspectives

321 Despite their huge potential, IONPs are currently only commercialized for treatment of iron-deficient anemia (e.g., Venofer[®] commercialized by American Regent) and cancer treatment (e.g., Nanotherm[®] 322 323 sold by Magforce). To foster IONP commercialization for other indications, new developments can be carried out such as: (i) IONP synthesis methods that do not contain toxic products and yield sufficient 324 325 production of highly pure and stable IONPs; (ii) IONPs that can be activated on demand by applying an 326 external source of energy, only when they are located in the organ to be treated, hence avoiding IONP 327 side effects on healthy tissues; and (iii) an energy source that is affordable and can easily be used by 328 doctors, especially within the hospital environment.

329

330 Acknowledgments

We would like to thank the BPI (banque publique d'investissement), the region of Paris (Paris Région Entreprise), the French Research Tax Credit program (crédit d'impôt recherche), the incubator Paris Biotech Santé, the ANRT (CIFRE 2014/0359, CIFRE 2016/0747, CIFRE 2013/0364, CIFRE 2015/976), the Eurostars programs (Nanoneck-2 E9309 and Nanoglioma E11778), the AIR program (aide à l'innovation responsable) from the region of Paris (A1401025Q), the ANR (Agence Nationale de la Recherche) Méfisto, as well as the Universities Paris 6 and Paris 11. We would also like to thank the

- 337 Nomis Foundation and Markus Reinhard for their support. Edouard Alphandéry wrote the article and
- 338 directed the research presented in this article.
- 339

340 **Conflicts of interest**

341 Edouard Alphandéry has been working at the start-up Nanobacterie.

342 Figure legends

Figure 1. Schematic representation of iron oxide nanoparticle (IONP) therapeutic mechanisms of action through chemotherapeutic drug release and activation, heat and reactive oxygen species (ROS) production, under application (or not) of an external source of energy. Schematic illustration of the three targeting mechanisms (passive, active, magnetic targeting).

Figure 2. Iron oxide nanoparticle (IONP) mechanisms of action can be subdivided between: (i) IONP local temperature increase (PTT, MHT); (ii) gene therapy (intentionally or non-intentionally triggered); (iii) reactive oxygen species (ROS) production (PDT or due to IONP themselves); (iv) drug delivery caused by the release of drugs initially attached to IONPs; (v) IONP targeting of specific cells or biological entities through active, passive or magnetic targeting.

Figure 3. Example of iron oxide nanoparticle (IONP) treatment using magnetic hyperthermia by administering IONPs to a tumor and then heating IONPs under alternating magnetic field application. The mechanisms of action involve moderate local heating at typically 42–50°C followed by death of cancerous cells.

Figure 4. Iron oxide nanoparticle (IONP) therapeutic applications including treatment of neuro-AIDS, cancer, blood contamination, Alzheimer's disease and Parkinson's disease, where the treatments rely on antifungal, antibacterial, antitumoral, nanozyme or immunotherapy.

	360	References
--	-----	------------

361 Lagrow, A.P. et al. (2019) Unravelling the growth mechanism of the coprecipitation of iron 1. 362 oxide nanoparticles with the aid of synchrotron X-Ray diffraction in solution, *Nanoscale* 11, 6620–6628 363 2. Dos Santos Monteiro, D. and Oliveira da Guarda Souza, M. (2016) Thermal decomposition of 364 precursors and iron oxide properties. J. Therm. Anal. Calorim. 123, 955–963 365 3. Groult, H. et al. (2018) Micellar iron oxide nanoparticles coated with anti-tumor glycosides. 366 Nanomaterials 8, 567 367 4. Sharafi, Z. et al. (2018) Synthesis of silica-coated iron oxide nanoparticles: preventing 368 aggregation without using additives or seed pretreatment. Iran. J. Pharm. Res. 17, 386-395 369 5. Ge, S. et al. (2009) Facile hydrothermal synthesis of iron oxide nanoparticles with tunable 370 magnetic properties. J. Phys. Chem. C 113, 13593-13599 371 6. Karimzadeh, I. et al. (2017) Superparamagnetic iron oxide (Fe3O4) nanoparticles coated with 372 PEG/PEI for biomedical applications: a facile and scalable preparation route based on the cathodic 373 electrochemical deposition method. Adv. Phys. Chem. 2017, 9437487 374 7. Arias, L.S. et al. (2018) Iron oxide nanoparticles for biomedical applications: a perspective on 375 synthesis, drugs, antimicrobial activity, and toxicity. Antibiotics 7, 1–32 376 8. Ghazanfari, M.R. et al. (2016) Perspective of Fe3O4 nanoparticles role in biomedical 377 applications. Biochem. Res. Int. 2016, 7840161 378 9. Hu, Y. et al. (2018) Construction of iron oxide nanoparticle-based hybrid platforms for tumour 379 imaging and therapy. Chem. Soc. Rev. 47, 1874–1900 380 10. Mosayebi, J. et al. (2017) Functionalization, and design of magnetic nanoparticles for 381 theranostic applications. Adv. Healthcare Mater. 6, 170030 382 11. Artemov, D. et al. (2003) MR molecular imaging of the Her-2/neu receptor in breast cancer cells 383 using targeted iron oxide nanoparticles. Magn. Res. Med. 49, 403–408

- 384 12. Aghanejad, A. et al. (2018) Mucin-1 aptamer-armed superparamagnetic iron oxide nanoparticles
- for targeted delivery of doxorubicin to breast cancer cells. *BioImpacts* 8, 117–127

- 386 13. Zhang, C. *et al.* (2007) Specific targeting of tumor angiogenesis by RGD-conjugated ultrasmall
 superparamagnetic iron oxide particles using a clinical 1.5-T magnetic resonance scanner. *Cancer Res.*67, 1555–1562
- Reddy, G.R. *et al.* (2006) Vascular targeted nanoparticles for imaging and treatment of brain
 tumors. *Clin. Cancer Res.* 12, 6677–6686
- 391 15. Yang, L. *et al.* (2008) Development of receptor targeted magnetic iron oxide nanoparticles for
- efficient drug delivery and tumor imaging. J. Biomed. Nanotechnol. 4, 439–449
- 393 16. Bonvin, D. et al. (2017) Folic acid on iron oxide nanoparticles: platform with high potential for
- 394 simultaneous targeting, MRI detection and hyperthermia treatment of lymph node metastases of prostate
- 395 cancer. Dalton Trans. 46, 12692–12704
- 396 17. Kohler, N. et al. (2004) A bifunctional poly(ethylene glycol) silane immobilized on metallic
- 397 oxide-based nanoparticles for conjugation with cell targeting agents. J. Am. Chem. Soc. 126, 7206–7211
- 18. Leuschner, C. *et al.* (2006) LHRH-conjugated magnetic iron oxide nanoparticles for detection of
- 399 breast cancer metastases. *Breast Cancer Res. Treat.* 99, 163–176
- 400 19. Alphandéry, E. (2019) Biodistribution and targeting properties of iron oxide nanoparticles for
 401 treatments of cancer and iron anemia disease. *Nanotoxicol.* 2, 1–24
- 402 20. Javid, A. *et al.* (2013) Chitosan-coated superparamagnetic iron oxide nanoparticles for
 403 doxorubicin delivery: synthesis and anticancer effect against human ovarian cancer cells. *Chem. Biol.*404 *Drug Des.* 82, 296–306
- 405 21. Albermani, M.S.K. *et al.* (2017) Vinblastine based iron oxide nano drug delivery system. *J.*406 *Global Pharma Tech.* 8, 90–96
- 407 22. Nagesh, P.K.B. *et al.* (2016) PSMA targeted docetaxel-loaded superparamagnetic iron
 408 oxidenanoparticles for prostate cancer. *Colloids Surf. B Biointerface*. 144, 8–20
- 409 23. Barar, J. et al. (2015) Multifunctional mitoxantrone-conjugated magnetic nanosystem for
- 410 targeted therapy of folate receptor-overexpressing malignant cells. J. Nanobiotechnol. 13, 26

- 411 24. Zaloga, J. *et al.* (2016) Pharmaceutical formulation of HSA hybrid coated iron oxide
 412 nanoparticles for magnetic drug targeting. *Eur. J. Pharm. Biopharm.* 101, 152–162
- 413 25. Shahabadi, N. *et al.* (2016) Improving antiproliferative effect of the anticancer drug cytarabine
 414 on human promyelocytic leukemia cells by coating on Fe3O4@SiO2 nanoparticles. *Colloids Surf. B*
- 415 *Biointerface*. 141, 213–222
- 416 26. Asadi, N. *et al.* (2018) Synthesis, characterization and *in vitro* evaluation of magnetic
 417 nanoparticles modified with PCL–PEG–PCL for controlled delivery of 5FU. *Art. Cell. Nanomed.*418 *Biotechnol.* doi: 10.1080/21691401.2018.1439839
- 419 27. Prabha, G, and Raj, V. (2016) Preparation and characterization of polymer nanocomposites
- 420 coated magnetic nanoparticles for drug delivery applications. J. Magnetism Magnetic Mater. 408, 26–34
- 421 28. Estelrich, J. and Busquets, M.A. (2018) Iron oxide nanoparticles in photothermal therapy.
 422 *Molecules* 23, 1567
- 423 29. Wu, L. *et al.* (2017) Remotely controlled drug release based on iron oxide nanoparticles for
 424 specific therapy of cancer. *Colloids Surf. B Biointerface.* 152, 440–448
- 30. Ferreira, R.V. *et al.* (2016) Thermosensitive gemcitabine magnetoliposomes for combined
 hyperthermia and chemotherapy. *Nanotechnol.* 27, 085105
- 427 31. Akilo, O.D. et al. (2016) An in vitro evaluation of a carmustine-loaded Nano-co-Plex for
- 428 potential magnetic-targeted intranasal delivery to the brain. Int. J. Pharm. 500, 196–209
- 429 32. Liu, J. et al. (2015) Design of hybrid nanovehicles for remotely triggered drug release: an
- 430 overview. J. Mater. Chem. B 3, 6117–6147
- 431 33. Nadeem, M. *et al.* (2016) Magnetic properties of polyvinyl alcohol and doxorubicine loaded iron
- 432 oxide nanoparticles for anticancer drug delivery applications. *PLoS One* 11, e0158084
- 433 34. Nosrati, H. et al. (2018) Bovine serum albumin (BSA) coated iron oxide magnetic nanoparticles
- as biocompatible carriers for curcumin-anticancer drug. *Bioorg. Chem.* 76, 501–509

- 435 35. Tuncelli, G. et al. (2015) 5-Fluorouracil intercalated iron oxide@layered double hydroxide core-
- 436 shell nano-composites with isotropic and anisotropic architectures for shape-selective drug delivery
- 437 applications. Mater. Sci. Eng. C 55, 562–568
- 438 36. Akilo, O.D. *et al.* (2016) An *in vitro* evaluation of a carmustine-loaded Nano-co-Plex for 439 potential magnetic-targeted intranasal delivery to the brain. *Int. J. Pharm.* 500, 196–209
- 440 37. Nosrati, H. et al. (2018) Methotrexate-conjugated L-lysine coated iron oxide magnetic
- nanoparticles for inhibition of mcf-7 breast cancer cells. *Drug Dev. Ind. Pharm.* 44, 886–894
- 442 38. Lin, J. et al. (2015) Drug/dye-loaded, multifunctional PEG-chitosan-iron oxide nanocomposites
- for methotraxate synergistically self-targeted cancer therapy and dual model imaging. *ACS Appl. Mater.*
- 444 Interface. 7, 11908–11920
- 445 39. Anderson, C.F. and Cui, H. (2017) Protease-sensitive nanomaterials for cancer therapeutics and 446 imaging. *Ind. Eng. Chem. Res.* 56, 5761–5777
- 447 40. Ansari, C. *et al.* (2014) Development of novel tumour-targeted theranostic nanoparticles 448 activated by membrane-type matrix metalloproteinases for combined cancer magnetic resonance 449 imaging and therapy. *Small* 10, 566–417
- 450 41. Wang, Y. et al. (2016) In vivo dual-targeted chemotherapy of drug resistant cancer by rationally
- 451 designed nanocarrier. *Biomater*. 75, 71–81
- 452 42. Saikia, C. *et al.* (2016) Effect of crosslinker on drug delivery properties of curcumin loaded 453 starch coated iron oxide nanoparticles. *Int. J. Biol. Macromol.* 93, 1121–1132
- 454 43. Ma, P. *et al.* (2017) Enhanced cisplatin chemotherapy by iron oxide nanocarrier-mediated 455 generation of highly toxic reactive oxygen species. *Nano Lett.* 17, 928–937
- 456 44. Li, K. *et al.* (2017) Next generation superparamagnetic iron oxide nanoparticles for cancer 457 theranostics. *Drug Discov. Today* 22, 1421–1429
- 458 45. Alphandéry, E. et al. (2017) Development of non-pyrogenic magnetosome minerals coated with
- 459 poly-L-lysine leading to full disappearance of intracranial U87-Luc glioblastoma in 100% of treated
- 460 mice using magnetic hyperthermia. *Biomaterials* 141, 210–222

- 461 46. Alphandéry, E. *et al.* (2017) Chains of magnetosomes with controlled endotoxin release and 462 partial tumour occupation induce full destruction of intracranial U87-Luc glioma in mice under the 463 application of an alternating magnetic field. *J. Control. Release* 261, 259–272
- 464 47. Le Fèvre, R. *et al.* (2017) Enhanced antitumour efficacy of biocompatible magnetosomes for the
 465 magnetic hyperthermia treatment of glioblastoma. *Theranostics* 7, 4618–4631
- 466 48. Hamdous, Y. *et al.* (2017) Biocompatible coated magnetosome minerals with various 467 organization and cellular interaction properties induce cytotoxicity towards RG-2 and GL-261 glioma 468 cells in the presence of an alternating magnetic field. *J. Nanobiotechnol.* 15, 74
- 469 49. Mandawala, C. et al. (2017) Biocompatible and stable magnetosome minerals coated with poly-
- 470 L-lysine, citric acid, oleic acid, and carboxy-methyl-dextran for application in the magnetic
- 471 hyperthermia treatment of tumours. J. Mater. Chem. B 5, 7644–7660
- 472 50. Hilger, I. (2013) *In vivo* applications of magnetic nanoparticle hyperthermia. *Int. J.*473 *Hyperthermia* 29, 828–834
- El Hajj Diab, D. *et al.* (2018) Combined treatments of magnetic intra-lysosomal hyperthermia
 with doxorubicin promotes synergistic anti-tumoral activity. *Nanomaterials* 8, 468
- 476 52. Maier-Hauff, K. et al. (2011) Efficacy and safety of intratumoural thermotherapy using magnetic
- iron-oxide nanoparticles combined with external beam radiotherapy on patients with recurrent
 glioblastoma multiforme. *J. Neurooncol.* 103, 317–324
- 479 53. Cortajarena, A.L. *et al.* (2014) Engineering iron oxide nanoparticles for clinical settings.
 480 *Nanobiomedicine* doi: 10.5772/58841
- 481 54. Master, A.M. *et al.* (2016) Remote actuation of magnetic nanoparticles for cancer cell selective
 482 treatment through cytoskeletal disruption. *Sci. Rep.* 6, 33560
- 483 55. Giustini, A.J. *et al.* (2011) Magnetic nanoparticle hyperthermia in cancer treatment. *Nanolife*484 doi: 10.1142/S1793984410000067
- 485 56. Kobayashi, T. (2011) Cancer hyperthermia using magnetic nanoparticles. Biotechnology J. 6,
- 486 1342-1347

- 487 57. Calderwood, S.K. (2016) Heat shock proteins promote cancer: it's a protection racket. *Trends*
- 488 Biochem. Sci. 41, 311–323
- 489 58. Patel, A. and Sant, S. (2016) Hypoxic tumour microenvironment: opportunities to develop
 490 targeted therapies. *Biotechnol. Adv.* 34, 803–812
- 491 59. Kievit, F.M. and Zhang, M. (2011) Surface engineering of iron oxide nanoparticles for targeted
 492 cancer therapy. *Acc. Chem. Res.* 44, 853–862
- 493 60. Mahajan, U.M. *et al.* (2016) Tumour-specific delivery of siRNA-coupled superparamagnetic
 494 iron oxide nanoparticles, targeted against PLK1, stops progression of pancreatic cancer. *Gut* 65, 1838–
- 495 **1849**
- 496 61. Min, L.F. *et al.* (2012) Magnetic iron oxide nanoparticles carrying PTEN gene to reverse
 497 cisplatin-resistance of A549/CDDP cell lines. *J. Central South University* 19, 331–339
- 498 62. Son, S. *et al.* (2015) Magnetofection mediated transient NANOG overexpression
 499 enhances proliferation and myogenic differentiation of human hair follicle derived mesenchymal stem
 500 cells. *Bioconjug. Chem.* 26, 1314–1327
- 501 63. Amer, M.H. (2014) Gene therapy for cancer: present status and future perspective. *Mol. Cell* 502 *Ther.* 2, 27
- 503 64. Zhao, M. *et al.* (2018) A GPC3-specific aptamer-mediated magnetic resonance probe for 504 hepatocellular carcinoma. *Int. J. Nanomed.* 13, 4433–4443
- 505 65. Connell, J.J. et al. (2015) Advanced cell therapies: targeting, tracking and actuation of cells with
- 506 magnetic particles. Regen. Med. 10, 757–772
- 507 66. Laskar, A. *et al.* (2013) SPION primes THP1 derived M2 macrophages towards M1-like 508 macrophages. *Biochem. Biophys. Res. Commun.* 441, 737–742
- 509 67. Zanganeh, S. et al. (2016) Iron oxide nanoparticles inhibit tumour growth by inducing pro-
- 510 inflammatory macrophage polarization in tumour tissues. Nat. Nanotechnol. 11, 986–994
- 511 68. Gao, L. et al. (2017) Iron oxide nanozyme: a multifunctional enzyme mimetic for biomedical
- 512 applications. *Theranostics* 7, 3207–3277

- 513 69. Chowdhury, P. *et al.* (2017) Magnetic nanoformulations for prostate cancer. *Drug Discov. Today*
- 514 22, 1233–1241
- 515 70. El-Zahabya, S.A. *et al.* (2019) Reviewing two decades of nanomedicine implementations in 516 targeted treatment and diagnosis of pancreatic cancer: an emphasis on state of art. *J. Control. Release* 517 293, 21–35
- 518 71. Srikanth Vallabani, N.V. and Singh, S. (2018) Recent advances and future prospects of iron 519 oxide nanoparticles in biomedicine and diagnostics. *3 Biotech.* 8, 279
- 520 72. Wang, L. *et al.* (2018) Tumour microenvironment-enabled nanotherapy. *Adv. Healthcare Mater.*521 7, 1701156
- 522 73. Brambilla, D. et al. (2011) Nanotechnologies for Alzheimer's disease: diagnosis, therapy, and
- 523 safety issues. Nanomed. Nanotechnol. Biol. Med. 7, 521-540
- 524 74. Niu, S. *et al.* (2017) Inhibition by multifunctional magnetic nanoparticles loaded with alpha525 synuclein RNAi plasmid in a Parkinson's disease model. *Theranostics* 7, 344–356
- 526 75. Umarao, P. *et al.* (2016) Neuroprotective potential of superparamagnetic iron oxide 527 nanoparticles along with exposure to electromagnetic field in 6-OHDA rat model of Parkinson's
- 528 disease. J. Nanosci. Nanotechnol. 16, 261–269
- 529 76. Sagar, V. *et al.* (2016) Magnetic nanotherapeutics for dysregulated synaptic plasticity during 530 neuroAIDS and drug abuse. *Molecular Brain* 9, 57
- 531 77. Parveen, S. et al. (2018) Preparation, characterization and antifungal activity of iron oxide
- nanoparticles. *Microbial Pathogenesis* 115, 287–292
- 533 78. Arakha, M. et al. (2015) Antimicrobial activity of iron oxide nanoparticle upon modulation of
- nanoparticle-bacteria interface. Sci. Rep. 5, 14813
- 535 79. Wang, L. *et al.* (2017) The antimicrobial activity of nanoparticles: present situation and 536 prospects for the future. *Int. J. Nanomed.* 12, 1227–1249
- 537 80. Kang, J.H. et al. (2015) Optimization of pathogen capture in flowing fluids with magnetic
- 538 nanoparticles. Small 11, 5657–5666









Types of treated diseases	Mechanisms involved in treatment	Experimental conditions (Celluar/animal/human)
	Enhanced chemotherapy	 IONP-PAX more efficient than free PAX: <i>in vitro</i> on PC3/CWR22R cells [69] <i>in vivo</i> on C4-2 mouse tumors [69]
Cancer	Reduction of toxicity of anticancer drugs	Tumor uptake of mitoxantrone increased with IONP by reducing therapeutically efficient dose of mitoxantrone shown <i>in</i> <i>vitro</i> and <i>in vivo</i> [69]
	Increase in drug-targeting efficacy	 Stimuli drug release as a function of: pH variation (<i>in vitro</i> [26]) Temperature variation (<i>in vivo</i> [27]) Magnetic field (<i>in vitro</i> [30)] Water/buffer (<i>in vitro</i> [34])
	ROS	 IONPs produce ROS: by themselves (Fenton reaction) (<i>in vitro</i> and <i>in vivo</i> [43]) following laser excitation (PDT) (<i>in vivo</i> [44])
	Localized heat	 MHT (human [52]) PTT (<i>in vitro</i> [28])
	Prevent tumor growth	Blocking the blood artery that irrigates the tumor and promotes tumor growth (<i>in vivo</i> [72])
Alzheimer's	Acts on fibillation process	Therapy/diagnosis (in vitro, in vivo [73])
Darkinson's	Gene therapy	Inhibition by IONP-RNAi (<i>in vitro, in vivo</i> [74])
Parkinson s	Magnetic field therapy	Neuroprotective ability of IONPs under magnetic field exposure (<i>in vivo</i> [75])
Neuro-AIDS	Enables AIDS drugs to cross blood–brain barrier	Proposed model for <i>in vitro/in vivo/</i> human configurations [76]
Iron-deficiency anemia	Enhances iron bioavailability	Commercially available to treat IAD [19]
Fungi/bacteria contamination	Destroys fungi/bacteria through ROS production/DNA damages	IONP activity demonstrated directly on fungi/bacteria [77–79]
Blood contamination	Removal of blood contaminants using IONP	Capture of blood bacteria by IONP demonstrated <i>in vitro</i> [80]

Table 1. Types of diseases treated with IONPs; mechanisms involved in the treatment; experimental demonstration;associated references

Abbreviations: PAX, paclitaxel; PDT, phtodynamic therapy; MHT, magnetic hyperthermia; PTT, photothermal therapy; RNAi, RNA interference; IAD, iron anemia disease.