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CD8⁺T-bet⁺ cells as a predominant biomarker for inclusion body myositis

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Short running title: T-bet: biomarker of inclusion body myositis

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Abstract

Background: Myositis is a heterogeneous group of muscular auto-immune diseases with clinical and pathological criteria that allow the classification of patients into different sub-groups. Inclusion body myositis is the most frequent myositis above fifty years of age. Diagnosing inclusion body myositis requires expertise and is challenging. Little is known concerning the pathogenic mechanisms of this disease in which conventional suppressive-immune therapies are inefficacious.

Objectives: Our aim was to deepen our understanding of the immune mechanisms involved in inclusion body myositis and identify specific biomarkers.

Methods: Using a panel of thirty-six markers and mass cytometry, we performed deep immune profiling of peripheral blood cells from inclusion body myositis patients and healthy donors, divided into two cohorts: test and validation cohorts. Potential biomarkers were compared to myositis controls (anti-Jo1-, anti-3-hydroxyl-3-methylglutaryl CoA reductase-, and anti-signal recognition particle-positive patients).

Results: Unsupervised analyses revealed substantial changes only within CD8⁺ cells. We observed an increase in the frequency of CD8⁺ cells that expressed high levels of T-bet, and containing mainly both effector and terminally differentiated memory cells. The senescent marker CD57 was overexpressed in CD8⁺T-bet⁺ cells of inclusion body myositis patients. As expected, senescent CD8⁺T-bet⁺ CD57⁺ cells of both patients and healthy donors were CD28^{null}CD27^{null}CD127^{null}. Surprisingly, non-senescent CD8⁺T-bet⁺ CD57⁻ cells in inclusion body myositis patients expressed lower levels of CD28, CD27, and CD127, and expressed higher levels of CD38 and HLA-DR compared to healthy donors. Using classification and regression trees alongside receiver operating characteristics curves, we identified and validated a frequency of CD8⁺T-bet⁺ cells > 51.5% as a diagnostic biomarker specific to inclusion body myositis, compared to myositis control patients, with a sensitivity of 94.4%, a specificity of 88.5%, and an area under the curve of 0.97.

Conclusion: Using a panel of thirty-six markers by mass cytometry, we identify an activated cell population (CD8⁺T-bet⁺ CD57⁻ CD28_{low}CD27_{low}CD127_{low} CD38⁺ HLA-DR⁺) which could play a role in the physiopathology of inclusion body myositis, and identify CD8⁺T-bet⁺ cells as a predominant biomarker of this disease.

Key words: inclusion body myositis, deep immune profiling, physiopathology, CD8⁺ T-bet⁺ cells, biomarkers, myositis

Abbreviations

CART: classification and regression trees

CI: confidence interval

CTL: cytotoxic T lymphocytes

CyTOF: cytometry by time of flight (mass cytometry)

HD: healthy donors

HLADR: human leukocyte antigen-DR

HMGCR: 3-Hydroxyl-3-MethylGlutaryl CoA Reductase

IdU: iododeoxyuridine

IFN- γ : interferon-gamma

IMNM: immune mediated necrotizing myopathies

MMI: median mass intensity

PBMC: peripheral blood mononuclear cells

ROC: receiver operating characteristics

sIBM: sporadic Inclusion body myositis

SPADE: spanning-tree progression analysis of density-normalized events

SRP: signal recognition particle

T-bet: T-box expressed in T cells

TEMRA: terminally differentiated memory cells

viSNE: stochastic neighbor embedding

1. Introduction

Myositis comprises a heterogeneous group of diseases with variable clinical, pathological and serological features, involving both muscular and extra-muscular tissues [1,2]. At present, sporadic inclusion body myositis (sIBM) is the only group bereft of treatment options since there is no evidence of efficacy for corticosteroids and/or immunosuppressive therapies in the long term [3–6]. sIBM is the most common myositis subtype after age 50. Disease onset is slow and progressive, gradually leading to muscle atrophy and a reduced quality of life. Diagnosis at sIBM onset could be misleading, potentially involving administration of a treatment that causes detrimental effects [7].

CD8+ T cells have been implicated in the physiopathology of sIBM because of their aggressive and infiltrative presence within muscle fibres overexpressing major histocompatibility type 1 molecules [8–12]. Various studies have observed a diverse repertoire of CD8+ T cells in the peripheral blood of sIBM patients, suggesting the presence of antigen-driven CD8+ T cell responses [13–15].

Two major T-box transcription factors drive differentiation of naïve CD8+ T cells into cytolytic T lymphocytes (CTL): T-bet (T-box expressed in T cells) and eomesodermin (eomes) [16,17]. High levels of T-bet expression in CD8+ T cells are required for antigen-specific CD8+ response, cytotoxic function and maintenance of memory subsets [18–21].

Repeated and prolonged stimulation of T cells leads to progressive loss of co-stimulatory markers, such as CD28 and CD27. Downregulation of the interleukin-7 alpha receptor (CD127) is also observed following chronic antigen stimulation [22,23]. In addition to this immune profile, expression of CD57 defines a state of replicative senescence [24–27].

B cell implication in the pathogenicity of sIBM has been suggested by the identification of anti-cytosolic 5'-nucleotidase 1A antibodies present in ~ 30% of patients [28–30]. Plasma cells are also observed in muscle inflammatory infiltrates of sIBM [31]. Finally, muscle biopsies from sIBM patients also reveal the presence of macrophages infiltrating non-necrotic muscle fibres [32], suggesting the involvement of innate immune defence mechanisms that are still poorly investigated.

Cytometry by time of flight (CyTOF), also known as mass cytometry, is an innovative cell profiling tool based on antigen detection using metal-conjugated antibodies. CyTOF allows simultaneous screening of more than 38 markers at the single cell level [33,34]. CyTOF is of particular interest in applications of deep immune profiling and biomarker

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discovery. When using CyTOF, intra-assay variations can be reduced by barcoding cells prior to staining for markers [35–37].

Considering the detrimental effects of immunosuppressants prescribed due to misdiagnosis, there is a need to identify novel diagnostic biomarkers for sIBM and to better understand the pathogenic mechanisms involved in this disease. The aim of this study was to better characterize the phenotype of peripheral blood mononuclear cells (PBMCs) from sIBM patients, to identify cells potentially involved in the pathophysiology of the disease and to identify a biomarker that can aid in the rapid diagnosis of sIBM from other types of myositis.

2. Material and methods

2.1. Patients and samples:

Approval for this study was obtained from the *Comité Consultatif sur le Traitement de l'Information en matière de la Recherche dans le domaine de la Santé* (CCTIRS), France. Written informed consent was signed by all patients and age-matched healthy donors (HD) according to the declaration of Helsinki. Untreated sIBM patients (cohort 1, n=10 and cohort 2, n=8) were defined based on the Lloyd diagnosis criteria [38]. Control groups of active myositis patients included untreated myositis patients suffering from anti-synthetase syndrome with anti-Jo1 positive antibodies (anti-Jo1, n=10), anti-3-hydroxyl-3-methylglutaryl CoA reductase positive immune mediated necrotizing myopathy (anti-HMGCR IMNM, n=9) and anti-signal recognition particle positive IMNM (anti-SRP IMNM, n=7) defined based on published ENMC criteria [39,40]. Main characteristics of patients are described in **Supplementary Table 1**. Detection of myositis-specific antibodies was performed by immunoassays as previously described [41]. Healthy control blood samples were obtained from the *Etablissement Français du sang* (EFS, Pitié-Salpêtrière-France) donated by healthy adults age-matched with sIBM patients (cohort 1, n=8 and cohort 2, n=8).

Upon reception of blood samples, peripheral blood mononuclear cells (PBMCs) were isolated and stored at -80°C in foetal bovine serum (Life Technologies, Saint-Aubin, France, Catalogue # 10270106) supplemented with 10% dimethyl sulfoxide. Twenty-four hours later, the cells were transferred to liquid nitrogen until use.

2.2 Mass cytometry experiments:

Thirty-six metal-conjugated antibodies, viability staining, and Iododeoxyuridine (IdU) were obtained from Fluidigm (Les Ulis, France) (**Supplementary Table 2**). Immunostaining and data acquisition were performed as previously described in a technical research article [36]. Briefly, PBMCs were incubated with IdU and then stained with cisplatin. Next, cells were incubated with CCR7 and CXCR5, followed by incubation with a mix containing all other cell surface markers. Next, samples were barcoded using BD CyTOFix/Cytoperm and BD wash/perm buffer (BD, France, Catalogue # 554715). All cells from different samples were washed and pooled together in a single tube. Fixation of pooled cells was performed using BD CyTOFix/Cytoperm buffer followed by incubation with antibodies targeting intranuclear markers in BD wash/perm buffer. Finally, DNA within cells was labelled using Cell ID Intercalator-Ir (Fluidigm, les Ulis, France, Catalogue # 201192A). Cellular events

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were acquired the following day on the Helios available at the “*Plateforme de Cytométrie de la Pitié-Salpêtrière*”.

Data files were obtained in the FCS file format and analysed using different algorithms available on the Cytobank cloud based platform, R software version 3.4.0 and GraphPad Prism version 6.

2.3 Data analysis:

Beads were gated out using the following: 140/142Ce, 151/153Eu, 165Ho and 175/176 Lu. Cisplatin-positive dead cells were gated out, and automated identification of stratifying signatures in cellular populations was performed using the Citrus algorithm to identify differences between sIBM patients and healthy donors. We performed Citrus analyses using the median mass intensities (MMI) of various markers. The settings used for Citrus were as follows: population (all CD45+ cells), groups (Healthy donors and sIBM patients), clustering channels (CD19, CD56, CD11b, CD14, CD3, CD4, CD8, CD25, CD45RA, CCR7), statistics channels (HLA-DR, IgD, CD28, CD127, CD27, T-bet, FoxP3, CD38, CD57), equal event sampling (20000 each), minimum cluster size (5%), cross validation fold (5) and false discovery rate (1%).

Next, using the cytobank cloud based platform, automated cell clustering using spanning-tree progression analysis of density-normalized events (SPADE) was performed on CD45+ events. This is to allow a non-biased and uniform gating strategy of the main cell populations in our study. The SPADE tree was generated based on main phenotypic markers such as: CD3, CD4, CD8, CD14, CD11b, CD56, CD16 and CD19, with 20 nodes and a down-sampling of 10 percent. Nodes corresponding to homogenous cell populations were each exported to a new experiment.

Next, we used stochastic neighbour embedding (viSNE) to explore detail populations of interest with an unbiased approach. The settings used for the viSNE run on CD8+ T cells are as follows: equal event sampling (3615 each), channels (CD28, CD127, CD27, T-bet and CD57), iterations (1000), perplexity (50) and theta (0.3). Furthermore, we analysed the frequency of various cell populations and the MMI of various markers on the subsets obtained from the viSNE run.

Histograms and dot plots were used to confirm all results obtained from unsupervised analyses.

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For identification of biomarkers specific to sIBM patients (cohort 1 and 2, n=18) compared to control myositis patients (anti-Jo1, anti-SRP and anti-HMGCR, n=26), we constructed a decisional tree using classification and regression trees (CART) with R software. As a result, new subjects from different subgroups can be predicted using a ranking algorithm. We included in the CART “potential predictors” based on our previous observations: frequencies of CD8+ T-bet+ T cells, CD8+ T-bet+ CD57+ T cells, CD8+ T-bet+ CD27+ T cells, CD8+ T-bet+ CD57+ CD27+ T cells and CD8+ T-bet+ CD57- CD27+ T cells. The validity of our classification tree model and the predictors (biomarkers) were validated by cross validation on a test set.

Using GraphPad Prism, we performed receiver operating characteristic (ROC) curves to better describe the biomarkers identified. Altogether, we chose a prediction value associated with the best cross-validation prediction rate on the test set using CART and with the best sensitivity, specificity, confidence interval (CI), and area under the curve (AUC) and a p-value < 0.0001 with ROC curves.

We used Shapiro-Wilk normality test to verify the distribution of quantitative data among the groups compared. For comparison of two groups, we used either the Mann-Whitney test or the t-test. When comparing MMI of expression for a marker within a population, we used the t-test. For multiple comparisons, we used the Kruskal-Wallis test using the Geisser-Greenhouse correction. We describe in-text data values using mean \pm SD.

3. Results

3.1. Increased frequency of CD8⁺ T cells in sIBM patients expressing high levels of T-bet and presenting a senescent immune profile (CD8^{T-bet+} CD57⁺):

Mean age of sIBM patients was 63.8±2 and 61±1 for healthy donors (p=0.152). To identify biomarkers specific to sIBM, we performed automated identification of stratifying signatures in CD45⁺ cellular populations using the Citrus algorithm. This analysis aimed to identify markers that vary significantly between sIBM patients (cohort 1, n=10) and healthy donors (cohort 1, n=8). Citrus did not identify any significant differences in activation or differentiation markers within B cells, CD16±CD56^{bright} NK cells, CD16+CD56^{dim} NK cells, monocytes, CD4⁺ T cells or CD38⁺ cells (**Fig. 1**). These observations were confirmed by classical gating of individual populations. Interestingly, significant differences were observed only within CD8⁺ T cell populations regarding the expression of CD27, CD57, CD127, CD28, T-bet and CD38. We next decided to characterize in detail CD8⁺ T cells in sIBM patients.

CD3⁺ CD8⁺ events corresponding to CD8⁺ T cells from the SPADE run on all CD45⁺ events were exported for further analysis (**Supplementary Fig. 1**). First, we observed a significant increase in the frequency of CD8⁺ T-bet⁺ T cells (CD8^{T-bet+}) in sIBM patients compared to healthy donors [sIBM=75.5%±13.2, HD=57.4%±15.2, p=0.02]. Further analysis revealed a remarkable increase in the frequency of CD8⁺ terminally differentiated memory T cells (TEMRA): CD8⁺ CD45RA⁺ CCR7⁻ in patients [sIBM=51.9%±13.6, HD=19.1%±9.9, p<0.0001] (**Fig. 2**). Interestingly CD8^{T-bet+} cells contain mainly both effector memory (EM) and TEMRA cells [% EM: sIBM=27%±16.4, HD=56%±12.1, p=0.0008 and % TEMRA: sIBM=66.1%±16.8, HD=30.4%±13.3, p=0.0002] (**Supplementary Fig. 2**). Next, we compared the expression of the senescence marker CD57 within CD8^{T-bet+} cells and CD8⁺ T-bet⁻ events (CD8^{T-bet-} cells). We observed a marked increase in the expression of CD57 within CD8^{T-bet+} cells in sIBM patients, while healthy donors had very low expression of CD57 in both CD8^{T-bet+} cells and CD8^{T-bet-} cells [MMI of CD57 in CD8^{T-bet+} cells: sIBM=188, HD=6, p=0.01 and MMI of CD57 in CD8^{T-bet-} cells: sIBM=0.4, HD=0.08, p=0.09] (**Supplementary Fig. 3**).

3.2. Selective loss of CD28, CD27 and CD127 in non-senescent CD8⁺ T cells (CD8^{T-bet+} CD57⁻) in sIBM patients compared to healthy donors:

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Next, we focused on three main subsets of CD8⁺ T cells of interest: senescent CD8_{T-bet+}⁺ CD57⁺ cells, non-senescent CD8_{T-bet+}⁺ CD57⁻ cells and Naive CD8_{T-bet-}⁻ cells. We compared expression of the co-stimulatory markers CD28 and CD27 and the functional marker CD127 within these three subsets. As expected, senescent CD8_{T-bet+}⁺ CD57⁺ cells are CD28null CD27null CD127null in both patients and healthy donors (**Supplementary Fig. 4**). Unexpectedly, we observed a decrease in expression of CD28, CD27 and CD127 in non-senescent CD8_{T-bet+}⁺ CD57⁻ cells of sIBM patients compared to healthy donors. Loss of CD28, CD27 and CD127 was not observed in CD8_{T-bet-}⁻ cells of either patients or healthy donors (**Fig. 3**).

3.3. Non-senescent CD8_{T-bet+}⁺ CD57⁻ cells in sIBM patients are activated, but do not spontaneously secrete greater amounts of IFN- γ :

Observation of this particular memory immune profile in non-senescent CD8_{T-bet+}⁺ CD57⁻ cells in sIBM patients prompted us to explore their activation states in comparison to other subsets we had examined. We compared the frequency of our populations of interest expressing activation markers CD38 and HLA-DR (Human Leucocyte Antigen-DR). Spontaneous interferon-gamma (IFN- γ) was also quantified. Interestingly, we observed higher frequencies of non-senescent CD8_{T-bet+}⁺ CD57⁻ cells expressing CD38 and HLADR in sIBM patients compared to healthy donors [HLADR⁺ cells: sIBM=9.9% \pm 6.2, HD=4.6% \pm 2.2, p=0.04 and CD38⁺ cells: sIBM=24.5% \pm 17.3, HD=9% \pm 5, p=0.02]. An increase in the frequency of senescent CD8_{T-bet+}⁺ CD57⁺ cells expressing CD38 was also observed in patients [CD38⁺ cells: sIBM=34.4% \pm 9, HD=4% \pm 1.1, p=0.0005]. We did not observe changes in spontaneous IFN- γ secretion between sIBM patients and healthy donors (**Fig. 4**). Low percentages of CD8_{T-bet-}⁻ cells expressing CD8 and HLADR were similarly observed in both patients and healthy donors (**Supplementary Fig. 5**)

3.4. Confirmation of observed characteristics in a second independent cohort of sIBM patients:

In an attempt to validate the immune profiles we observed in the first cohort of sIBM patients (n=10), we stained PBMCs from a second independent cohort with a minimal set of markers using an independent experiment (cohort 2: sIBM=8 and healthy donors=8). We confirmed increased frequencies of TEMRA CD8⁺ T cells, CD8_{T-bet+}⁺ cells and activated non-senescent CD8_{T-bet+}⁺ CD57⁻ cells in sIBM patients from cohort 2 in comparison to healthy

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donors. A non-significant tendency towards higher frequencies of CD38+ senescent CD8_{T-bet+} CD57+ cells was observed. We also confirmed lower expression of CD27, CD127 and CD28 by non-senescent CD8_{T-bet+} CD57- cells in sIBM patients (**Fig. 5**). Values between sIBM patients of cohort 1 and 2 were not significantly different with respect to immune profile (**Supplementary Fig. 6**).

Taken together, these observations describe a novel increase in the frequency of senescent CD8_{T-bet+} CD57+ cells in sIBM patients compared to healthy donors. Further, sIBM patients exhibit an activated non-senescent CD8_{T-bet+} CD57- cell population characterized by lower expression of CD28, CD27 and CD127, and higher expression of CD38 and HLADR compared to healthy donors.

3.5. Identification of biomarkers specific to sIBM patients:

. Next, we aimed to identify a minimal set of biomarkers that can be used as diagnostic criteria for sIBM patients (cohort 1 and 2, n=18). Other myositis patients presenting either anti-Jo1 auto-antibodies, anti-SRP auto-antibodies or anti-HMGCR auto-antibodies were used as controls. After multiple CART runs using different predictors, such as the frequency of CD8_{T-bet+}, senescent CD8_{T-bet+} CD57+ and CD8_{T-bet+} CD27+ cells, the best predictor of classification was always the frequency of CD8_{T-bet+} cells. Given that the cut-off prediction frequency of CD8_{T-bet+} cells could vary between different CART runs, we used ROC curves to help choose the best predictive value. Altogether, using CART and ROC curves, we identified and validated a frequency of CD8_{T-bet+} cells > 51.51% as the best predictor to distinguish sIBM patients from other myositis patients (n=26), with a sensitivity of 94.4%, a specificity of 88.5%, a confidence interval of [69.9-97.6%], an area under curve of 0.97 and a p-value < 0.0001 (**Fig. 6**).

4. Discussion

Using CyTOF, we simultaneously analysed thirty-six markers on non-stimulated PBMCs from sIBM patients and healthy donors. Unsupervised analysis revealed significant changes only within CD8⁺ T cells. CD8⁺ T cells from sIBM patients overexpress T-bet. Loss of CD28, CD27 and CD127 in senescent CD8^{T-bet+} CD57⁺ cells in both sIBM patients and healthy donors was expected. Unexpectedly, we observed specific loss of CD28, CD27 and CD127 in non-senescent CD8^{T-bet+} CD57⁻ cells from sIBM patients but not from healthy subjects.

Previous studies have demonstrated that during chronic antigen stimulation, overexpression of T-bet on CD8⁺ T cells represses expression of inhibitory receptors, defining a subset of exhausted CD8⁺ T cells with proliferative capacity and effector functions (21)]. In this study we identified a subset of activated non-senescent CD8⁺ T-be⁺ cells defined by low levels of expression of CD57 and higher levels of expression of CD38 and HLA-DR in sIBM patients compared to healthy donors. This activated and non-senescent immune profile suggests continuous proliferative capacity and effector functions of these cells in sIBM patients, which could explain the progressive and destructive nature of sIBM. This suggestion is supported by the presence of not only TEMRA, but also EM cells within T-bet⁺ cells of sIBM patients.

Our results are in line with previous studies that observed increased frequencies of CD8⁺ CD28null T cells in PBMCs from sIBM patients compared to healthy individuals (15). More recently, CD8⁺ CD57⁺ large granular lymphocytes were observed in blood and muscle of sIBM patients (14)]. Here, we present a more detailed and precise phenotype of peripheral blood CD8⁺ T cells in sIBM patients, thus enhancing understanding of this disease.

In this study, we did not observe an increase in spontaneous IFN- γ secretion by CD8^{T-bet+} cells from sIBM patients. This could be because cells were not stimulated, since our aim was to gain insight into the immune profile of PBMCs from sIBM patients without stimulation. We previously showed an increase in IFN- γ secretion by PBMC in sIBM patients after stimulation (13)].

In this study, we were also able to identify a novel diagnostic biomarker for IBM. Various diagnostic criteria for sIBM have been described, with sensitivities ranging from 11% to 90% and specificities above 96% (38)]. Compared to other myositis subtypes (anti-Jo1 positive antibodies, IMNM with anti-HMGCR positive and anti-SRP positive antibodies), we

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obtained the highest sensitivity described thus far (sensitivity of 94.4%), and 88.5% specificity. All previous diagnostic criteria combined clinical and pathological analyses, both of which require specific expertise. The biomarker we present herein is based on two biological markers (CD8 and T-bet), and could be complementary to the actual diagnostic approaches used for sIBM patients. This biomarker could aid in categorizing patients for clinical trials and research, especially when diagnosis remains uncertain. Further prospective analyses are needed to compare this novel biological biomarker to pathological criteria in patients with IBM suspicion.

5. Funding source

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6. Competing interests

All the authors have no conflicts of interest to disclose.

7. Supplementary material

Supplementary Table 1: Clinical characteristics of active sIBM and myositis control patients

Supplementary Table 2: List of antibodies

Supplementary Fig 1: Automated cell clustering using the SPADE algorithm

Supplementary Fig 2: Distribution of CD8 cell populations within CD8+ T-bet+ cells

Supplementary Fig 3: Expression of CD57 in CD8+ T cells

Supplementary Fig 4: Expression of CD28, CD27 and CD127 in senescent CD8 cells

Supplementary Fig 5: Frequency of cells expressing activation markers

Supplementary Fig 6: Close values for the immune profiles of sIBM patients from cohort 1 and cohort 2

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9. Figure captions

Fig 1: Automated identification of stratifying signatures in cellular populations between patients and healthy donors (use of median mass intensities)

Cells from sIBM patients (cohort 1, n=10) and age-matched healthy donors (n=8) were labeled individually with antibodies targeting cell surface antigens. Next, the cells were barcoded, pooled and labelled with antibodies targeting intranuclear antigens. Beads and dead cells were gated out. Automated identification of stratifying signatures in cellular populations was performed using the Citrus algorithm available on the Cytobank cloud-based platform. The settings used for the Citrus runs on total cells were as follow: population (all CD45+ cells), groups (Healthy donors and sIBM patients), clustering channels (CD3, CD4, CD8, CD11b, CD56, CD19, CD25, CD45RA and CCR7), statistics channels (HLA-DR, IgD, CD28, CD127, FoxP3, CD27, CD38, T-bet and CD57), equal event sampling (20.000 events each), minimum cluster size (5%), cross validation fold (5) and false discovery rate (1%). **a)** Clustering channels, **b)** shows the model error rate of the output classification models and **c)** markers which vary significantly between sIBM patients and HD within the colored nodes.

Fig 2: Increase frequency of CD8+ T cells in sIBM patients expressing high levels of T-bet and presenting a senescent immune profile (T-bet+ CD57+)

Cells from sIBM patients (cohort 1, n=10) and healthy donors (n=8) were labeled individually with antibodies targeting cell surface antigens. Next, the cells were barcoded, pooled and labelled with antibodies targeting intranuclear antigens. Automated cell clustering was performed using the SPADE and viSNE algorithms available on the cytobank cloud-based platform. Here, we describe detail observations on CD8+ T cells: **a)** frequency of T-bet+ cells in a patient and a healthy individual, **b)** visualization of different CD8+ T cell populations, **c)** and **d)** comparison of the percentages of T-bet+ and TEMRA cells respectively, between sIBM patients and healthy donors. Two-tailed statistical analyses were performed using Mann-Whitney test (*p<0.05, **p<0.001, ***p<0.0001 and ****p<0.00001). The data shown are representative of an independent experiment and represent median with interquartile.

Fig 3: Selective loss of CD28, CD27 and CD127 in non-senescent CD8+ T cells (CD8^{T-bet+} CD57⁻) in sIBM patients compared to healthy donors

Data correspond to sIBM patients and healthy donors of cohort 1. We defined three populations of CD8+ T cells based on the expression of T-bet and CD57: naive (CD8+ T-bet⁻), senescent (CD8+ T-bet⁺ CD57⁺) and non-senescent (CD8+ T-bet⁺ CD57⁻) CD8 cells. **a)**

9. Figure captions

dot plot showing the expression of CD57 amongst CD8+ T-bet+ cells of an individual, **b)** heat maps showing the median mass intensity (MMI) of CD28, CD27 and CD127 and **c)** comparison of the MMI of CD28, CD27 and CD127 between sIBM patients and HD. Two-tailed statistical analyses were performed using t-test (* $p < 0.05$, ** $p < 0.001$, *** $p < 0.0001$ and **** $p < 0.00001$). The data shown are representative of an independent experiment and represent median with interquartile.

Fig 4: Non-senescent CD8+ cells in sIBM patients show an activated immune profile; nevertheless they do not spontaneously secrete greater amounts of IFN- γ

Data correspond to sIBM patients and healthy donors of cohort 1. Automated cell clustering was performed using the viSNE algorithm available on the cytobank cloud-based platform. The settings used for the viSNE run on CD8+ T cells were as follow: equal event sampling (3615 events each), channels (CD28, CD127, CD27, T-bet and CD57), iterations (1000), perplexity (50) and theta (0.3). Each dot represents a cell and each viSNE map is representative of a single subject. ViSNE maps show the single cell expression of **a)** CD38, **b)** HLADR and **c)** IFN γ . Next, we compared the frequencies of cells expressing activation markers in different populations between sIBM patients and HD: **d)** comparison of the percentages of CD38+ cells, **e)** HLA-DR+ cells and **f)** IFN γ + cells. Two-tailed statistical analyses were performed using Mann-Whitney test (* $p < 0.05$, ** $p < 0.001$, *** $p < 0.0001$ and **** $p < 0.00001$). The data shown are representative of an independent experiment and represent median with interquartile.

Fig 5: Confirmation of the immune profile identified using an independent cohort of sIBM patients and healthy donors

In order to confirm the immune profile of sIBM in comparison to healthy donors, we used an independent mass cytometry experiment and different subjects. Cells from sIBM patients (cohort 2, $n=8$) and healthy donors ($n=8$) were labeled individually with antibodies targeting cell surface antigens. Next, the cells were barcoded, pooled and labelled with antibodies targeting intranuclear antigens. Automated cell clustering was performed using the SPADE algorithm available on the cytobank cloud-based platform. CD8+ T cells were exported for downstream analyses. **a)** frequency of T-bet+ cells, **b)** frequency of TEMRA cells, **c)** frequency of CD38+ and HLADR+ non-senescent CD8 cells and **e)** MMI of CD28, CD27 and CD127 in non-senescent CD8 cells. Two-tailed statistical analyses were performed using either t-test or Mann-Whitney (depending on normality test) to compare either the percentages

9. Figure captions

or the MMI (* $p \leq 0.05$). The data shown are representative of an independent experiment and represent median with interquartile.

Fig 6: Identification of biomarkers specific to sIBM patients in comparison to control myositis patients

In order to identify biomarkers specific to sIBM patients (cohort 1 and 2, $n=18$) compared to other sub-groups of myositis (anti-Jo1: $n=10$, anti-SRP: $n=7$ and anti-HMGCR: $n=9$), we built and validated a decisional algorithm tree with classification and regression trees (CART) using the R software version 3.4.0. Potential biomarkers included in the CART were the percentages of CD8+ T-bet+ cells, CD8+ T-bet+ CD57+ cells, CD8+ T-bet+ CD27+ cells, CD8+ T-bet+ CD57+ CD27+ cells and CD8+ T-bet+ CD57- CD27+ cells. **a)** Decisional tree. CV was performed by dividing the data set into 10 parts. We randomly selected 70% of the data for the training set ($n=25$), with the remainder assigned to the test set ($n=19$), **b)** ROC curve based on the percentages of CD8+ T-bet+ cells of all myositis patients, showing sensitivity (Se), specificity (Sp), confidence interval (CI), area under curve (AUC) and p value for a cut-off value of CD8+ T-be+ cells $> 51.51\%$ and **c)** percentages of T-bet+ cells of all myositis patients within total CD8 cells.

Fig 1

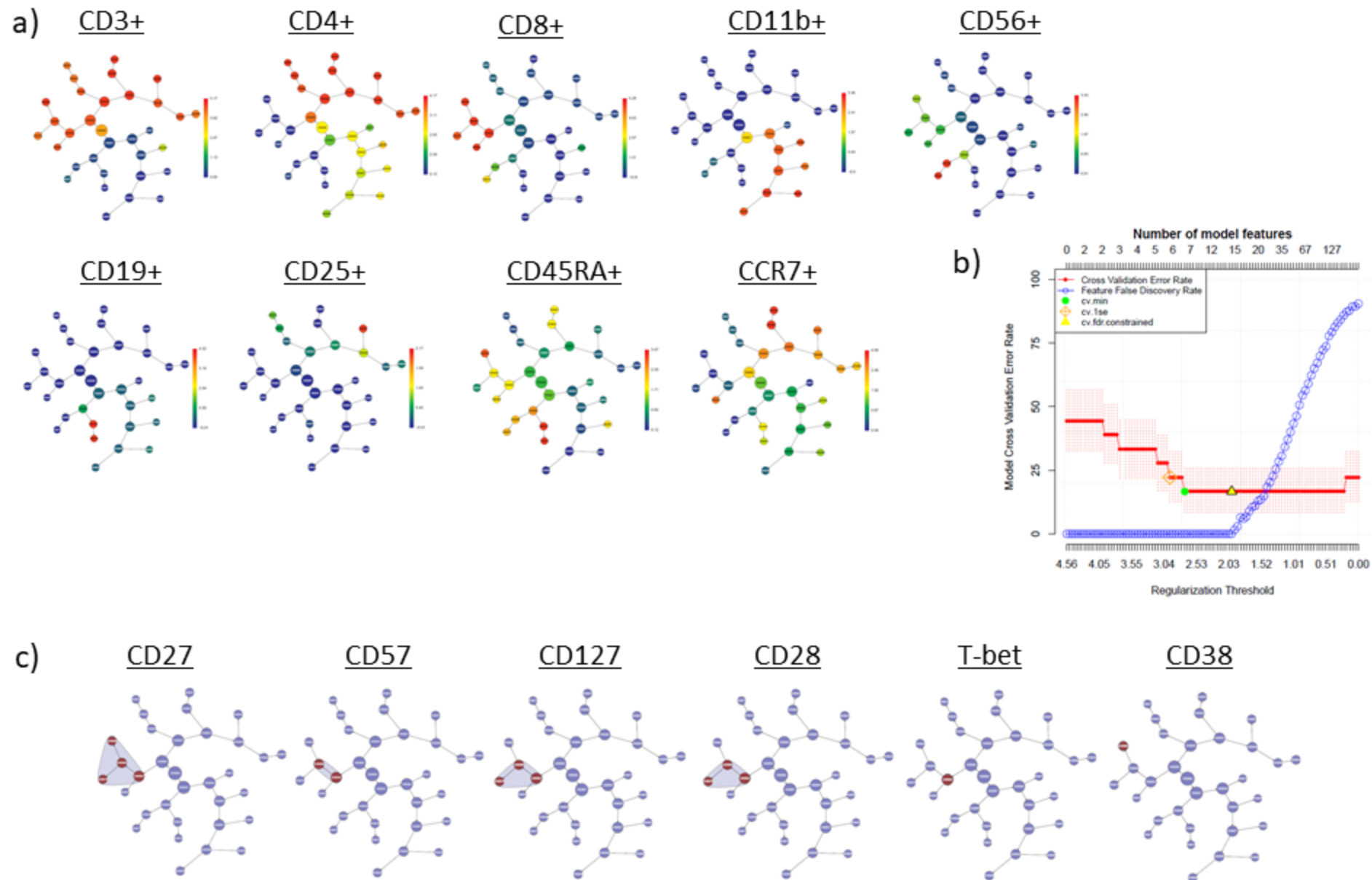


Fig 1

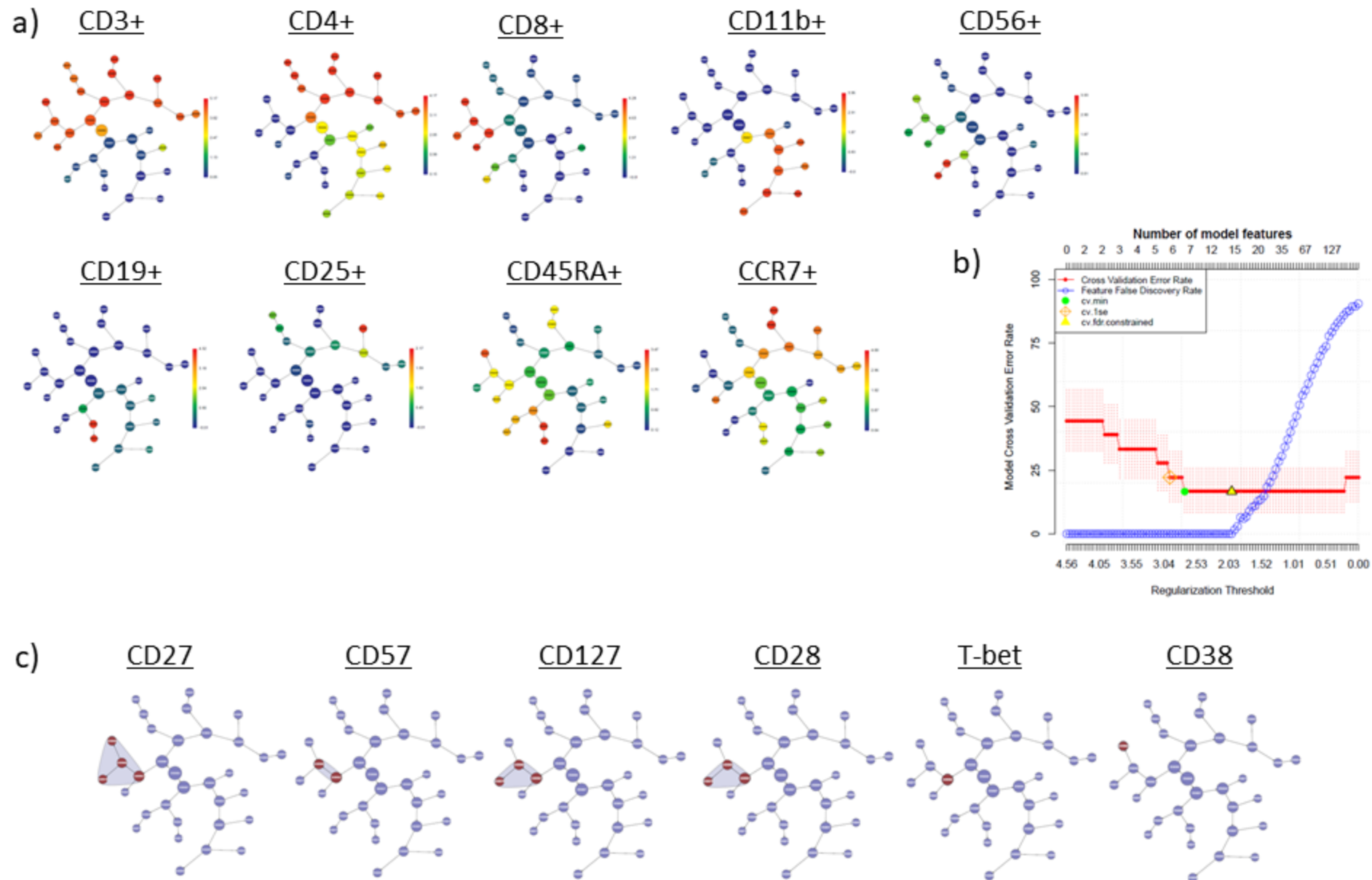


Fig 2

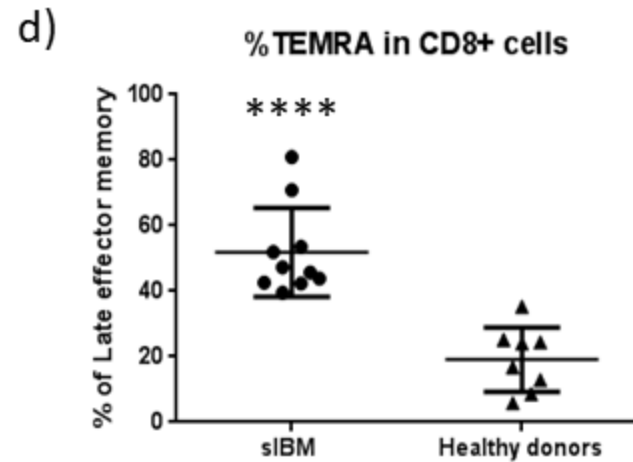
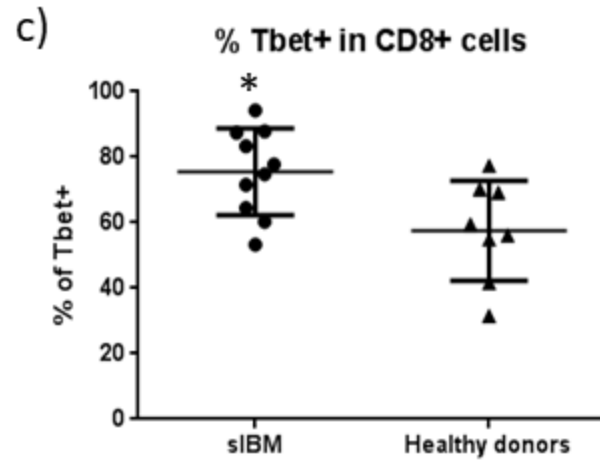
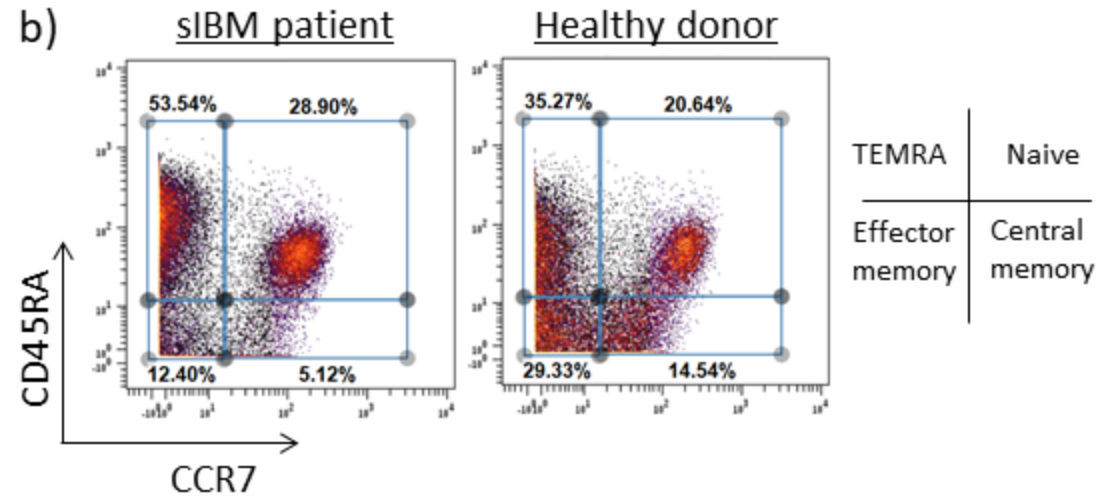
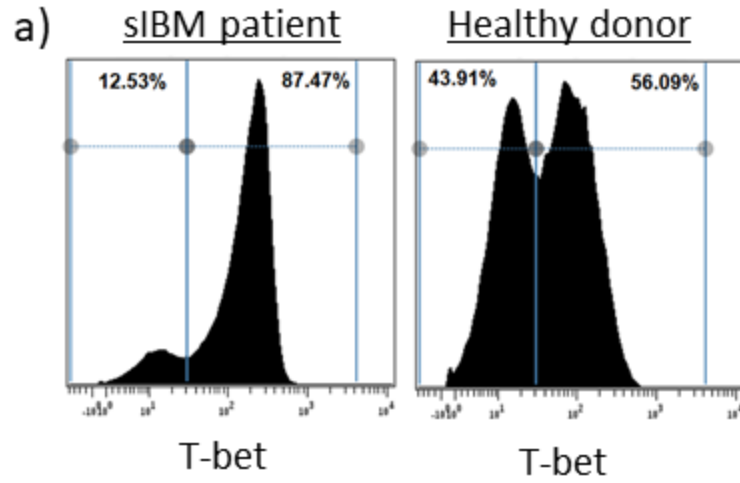


Fig 2

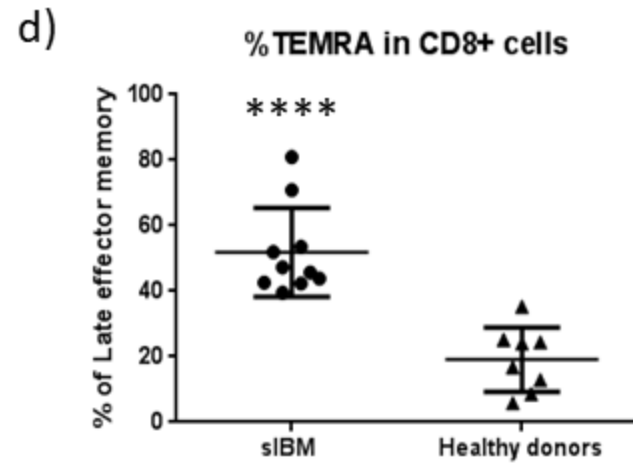
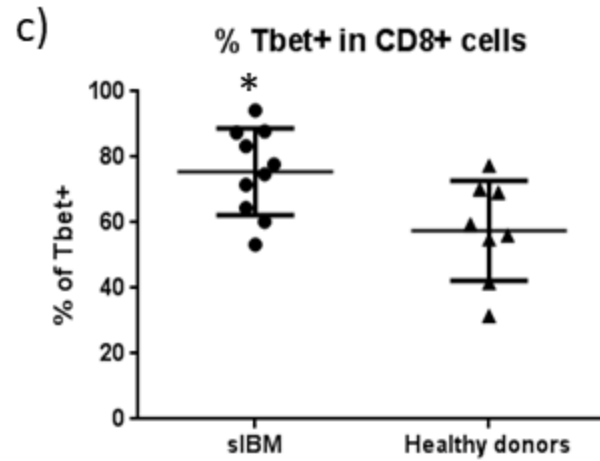
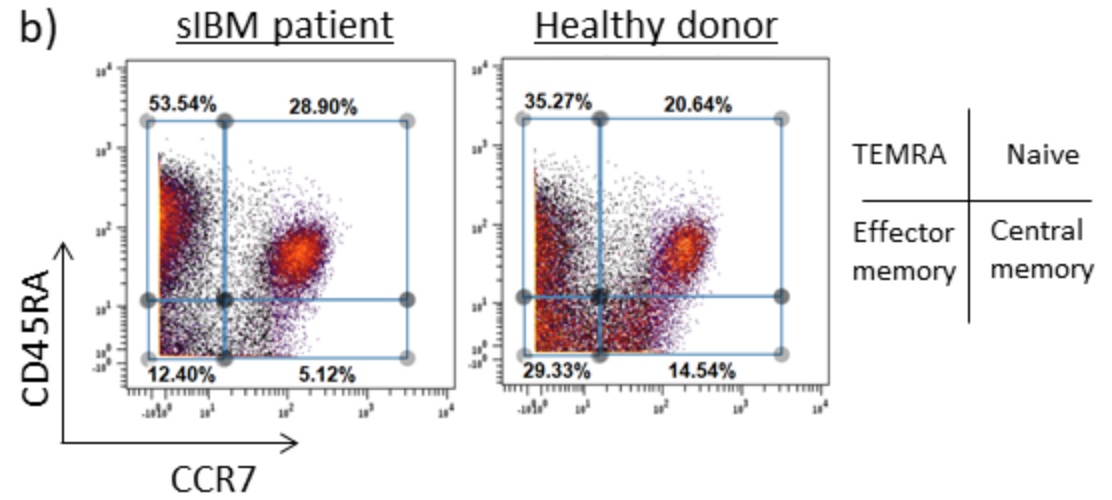
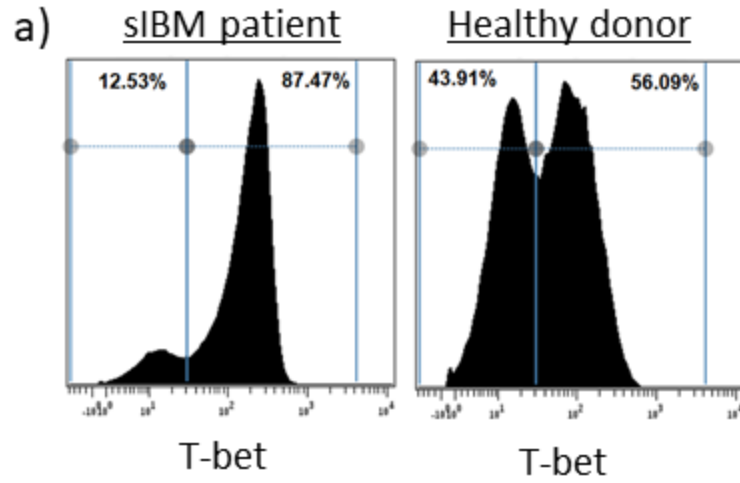


Fig 3

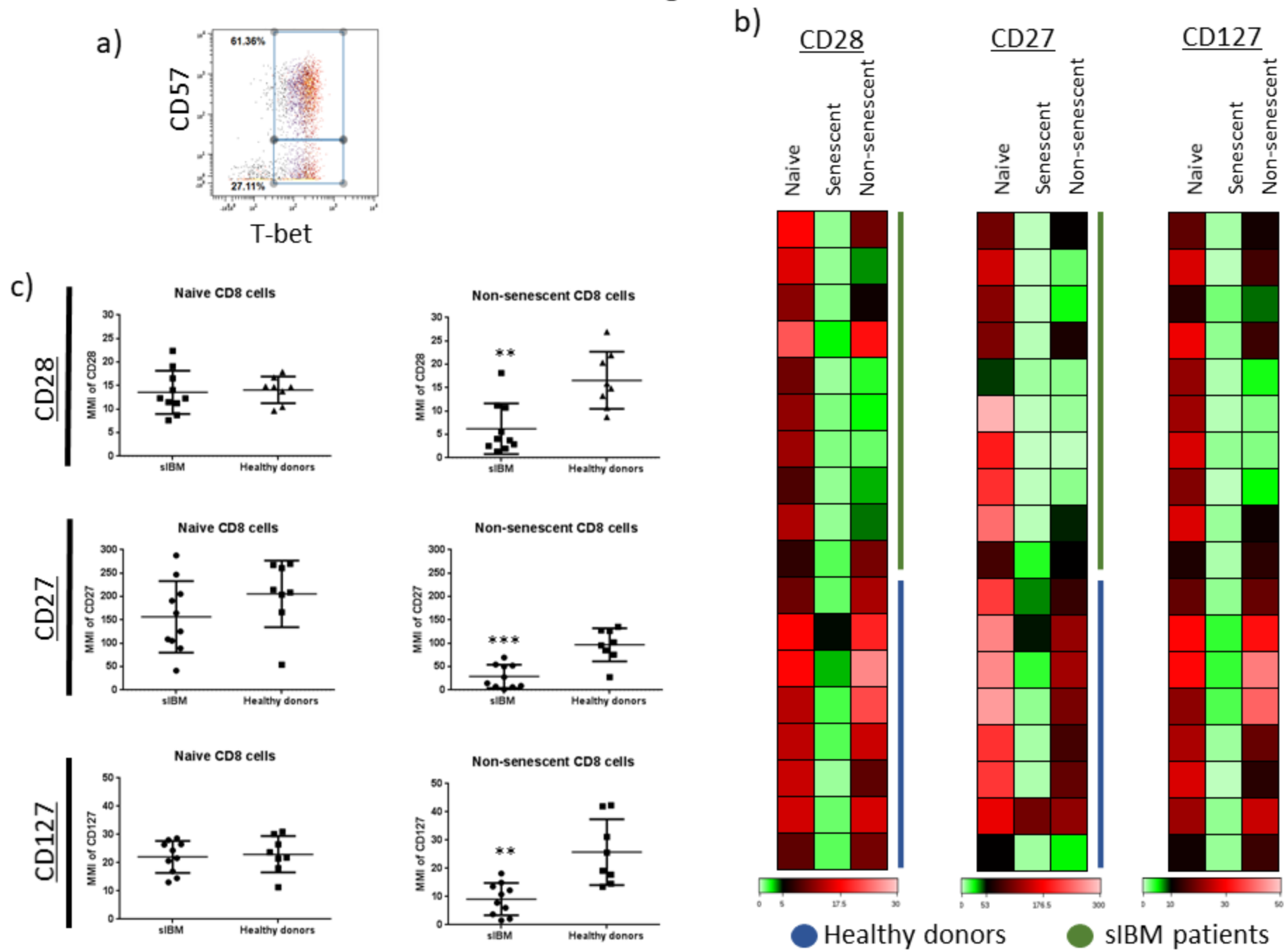


Fig 3

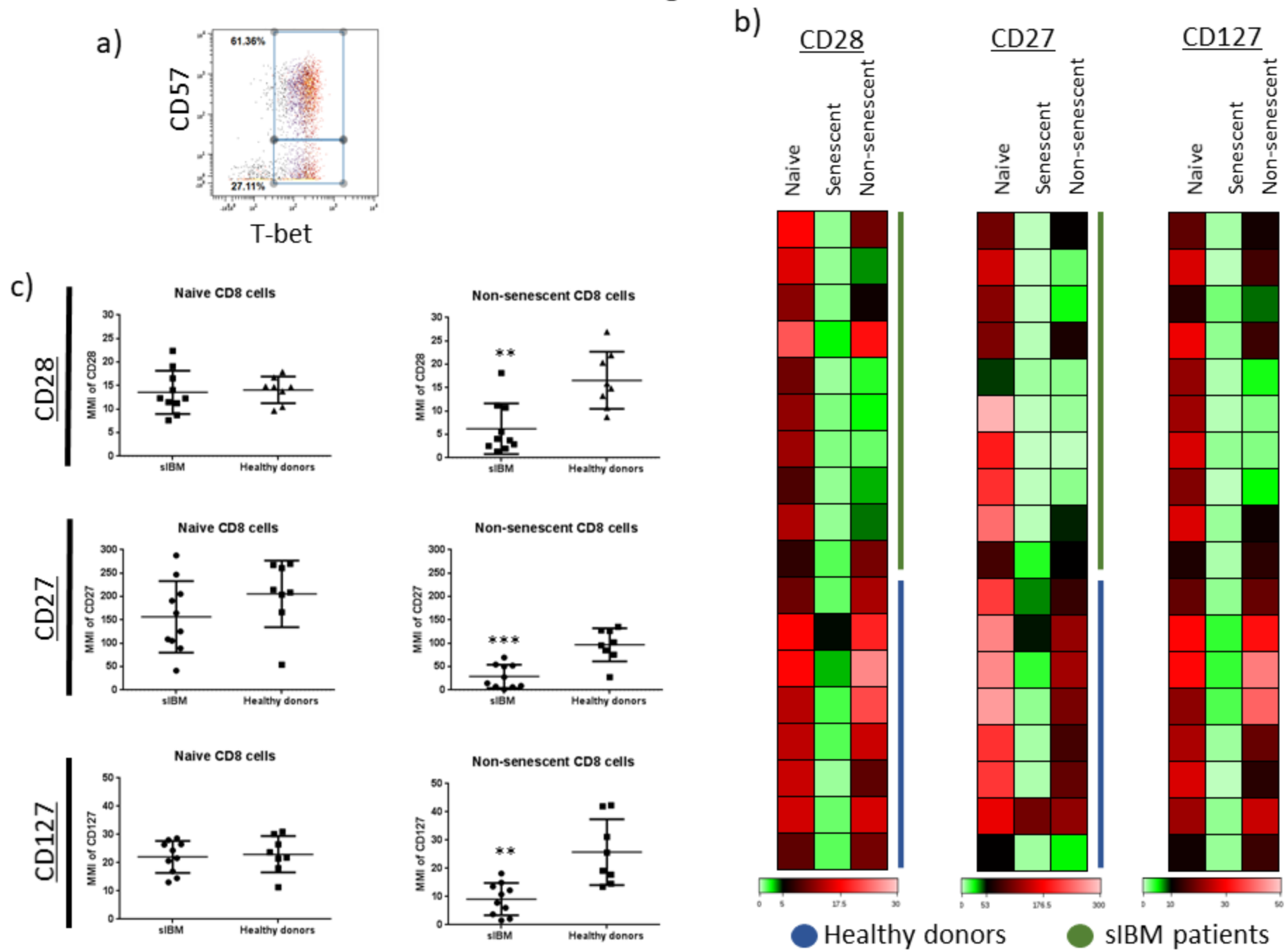


Fig 4

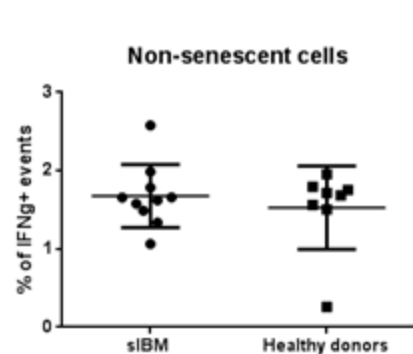
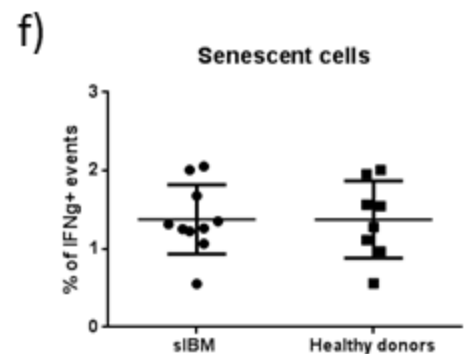
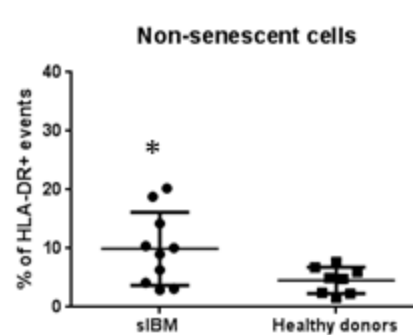
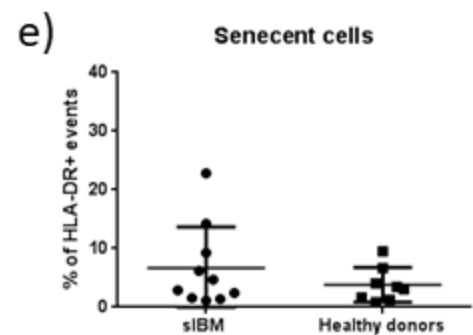
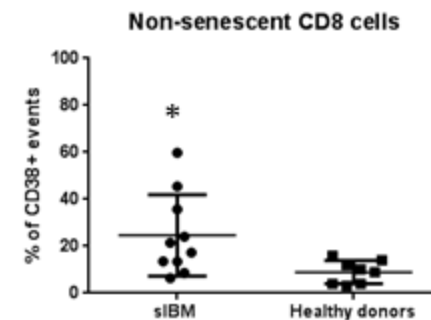
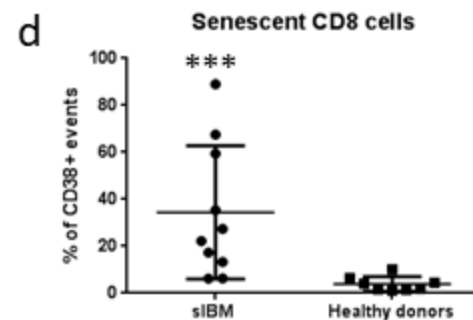
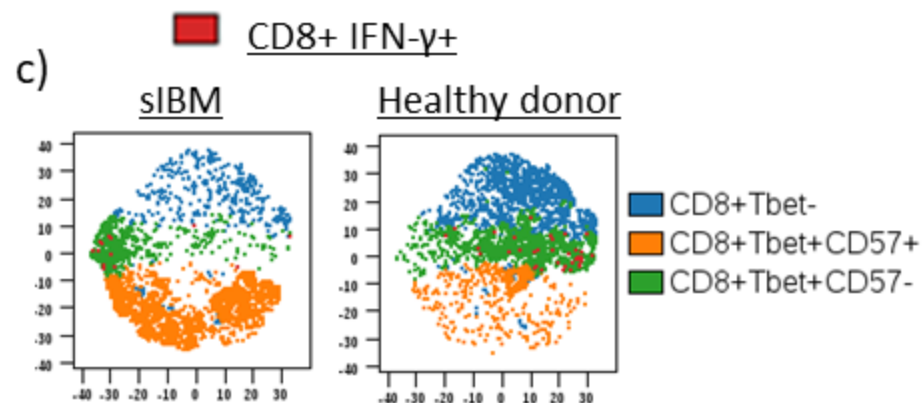
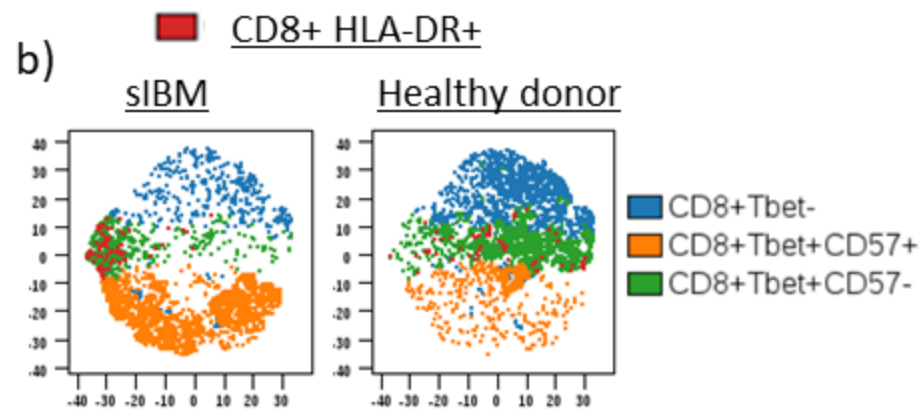
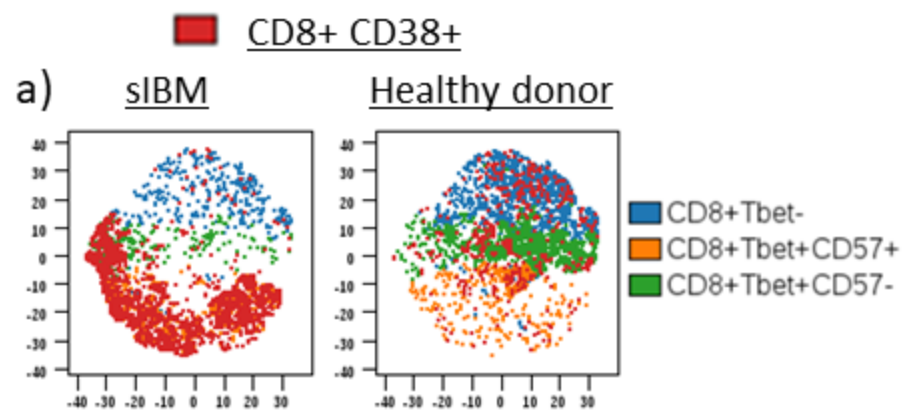


Fig 4

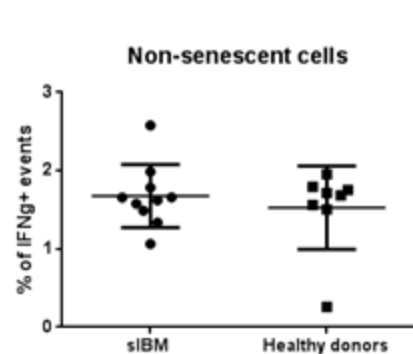
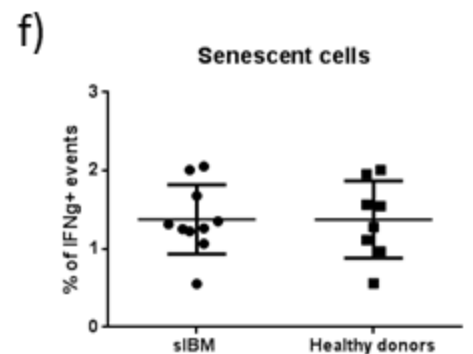
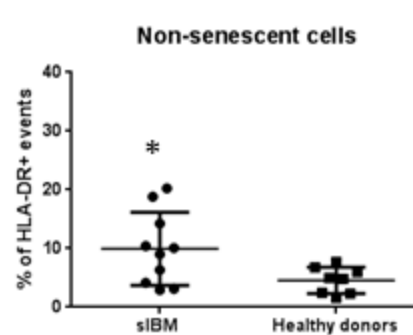
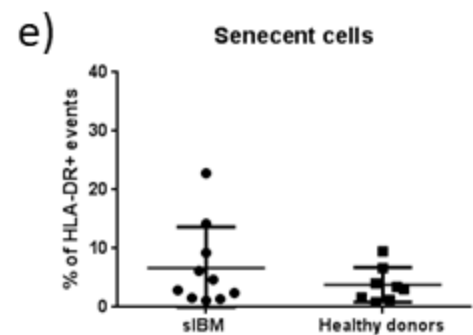
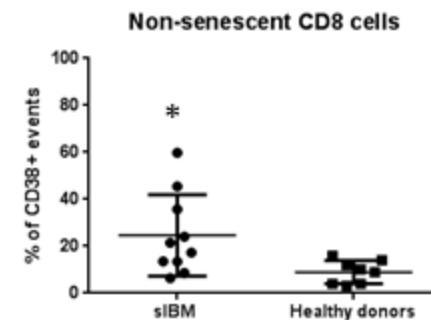
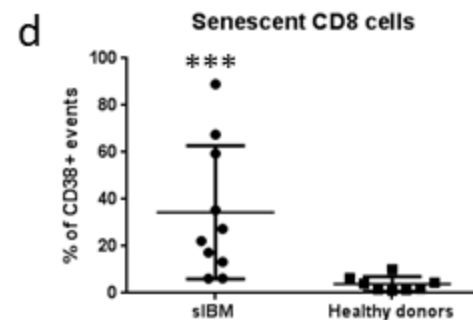
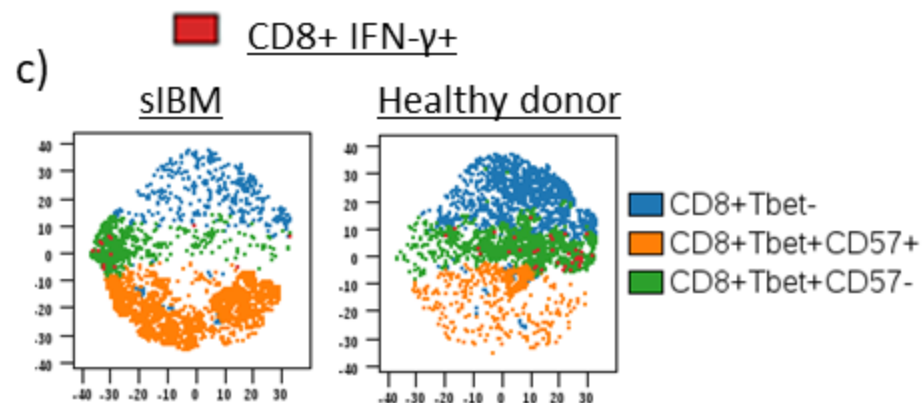
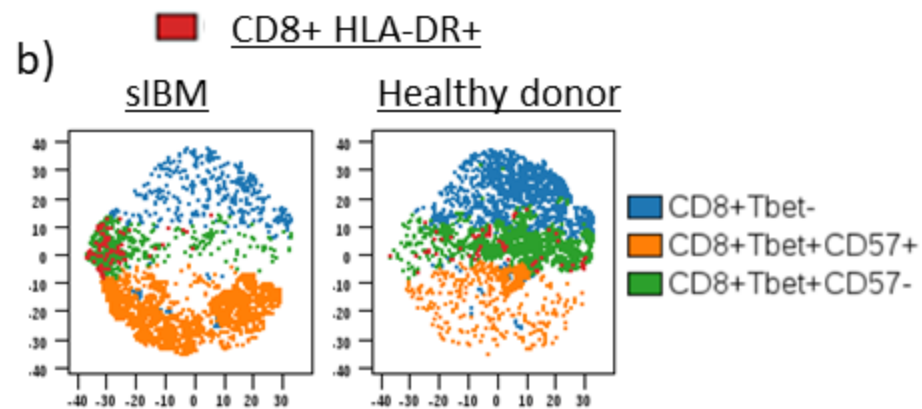
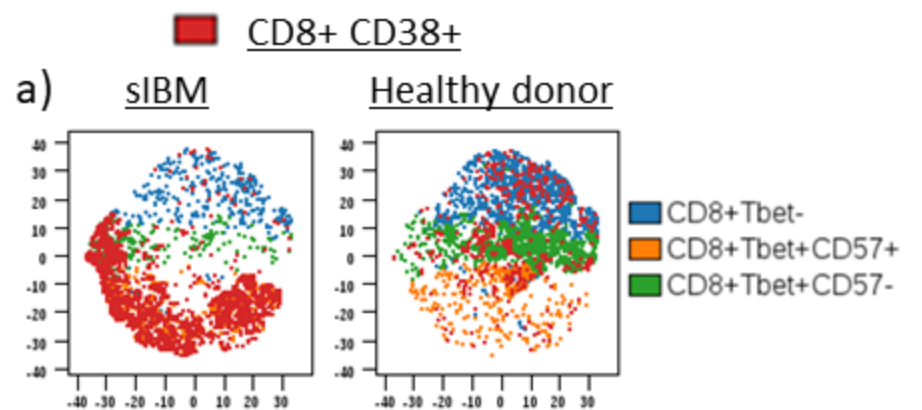


Fig 5

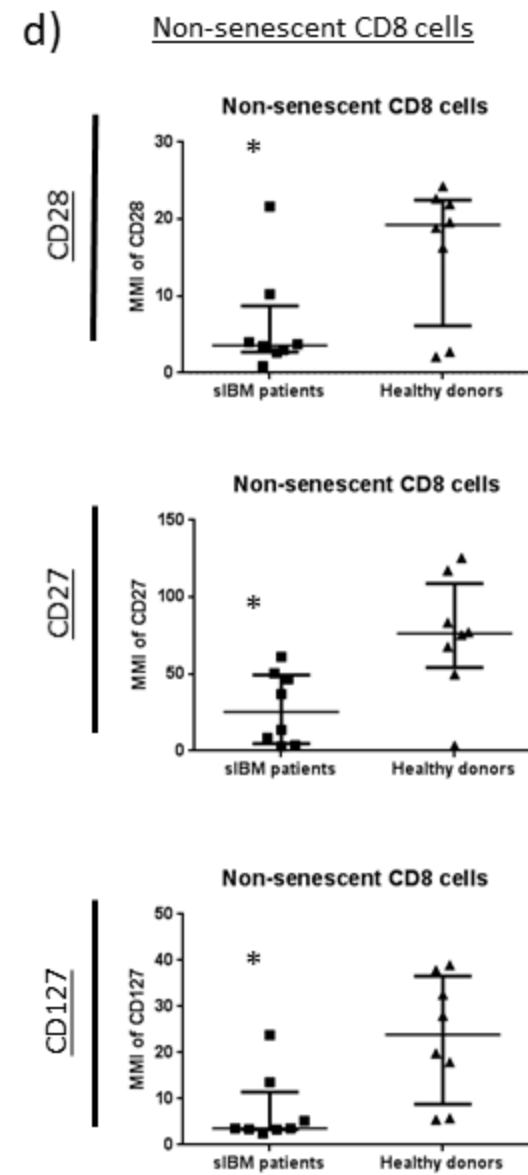
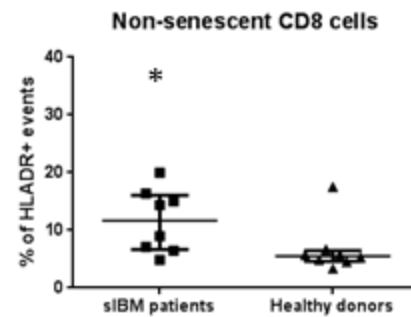
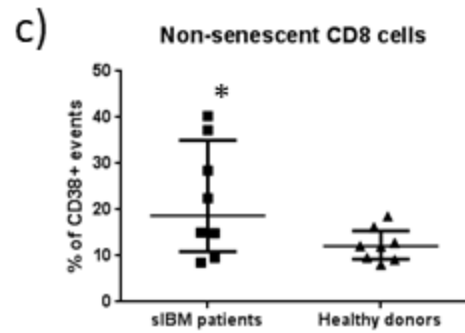
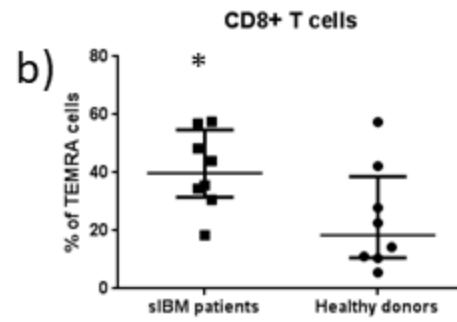
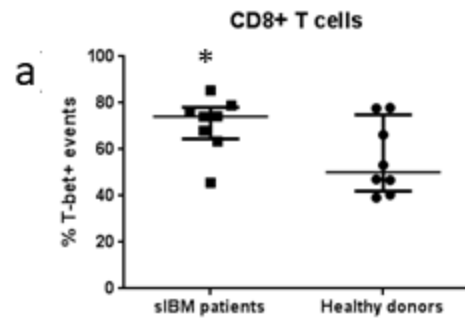


Fig 5

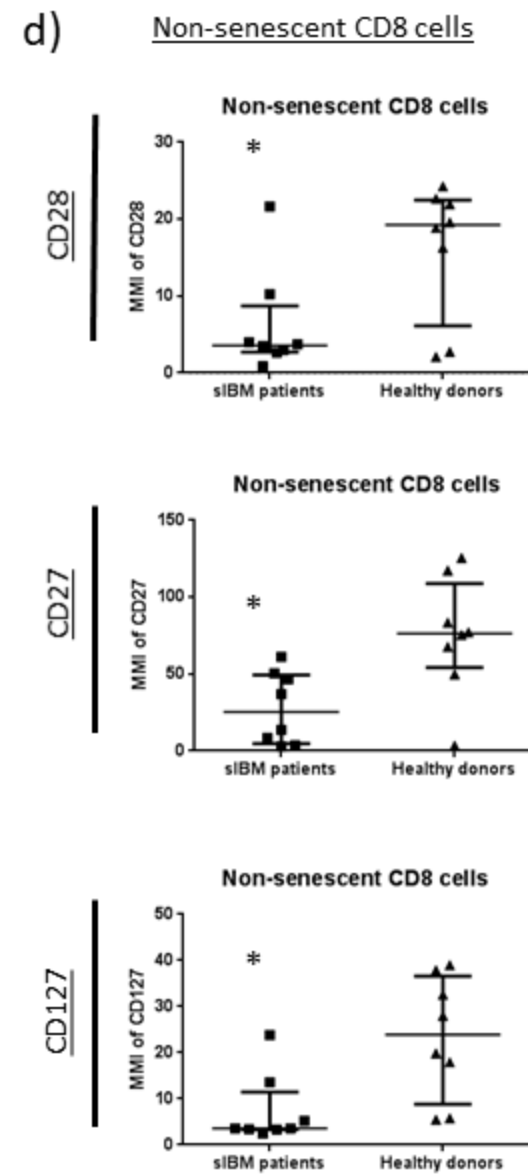
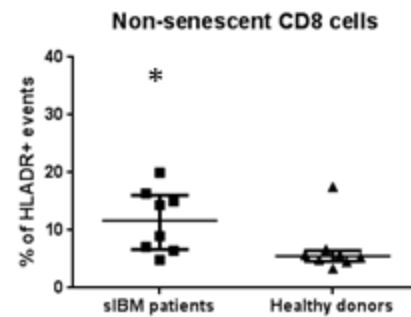
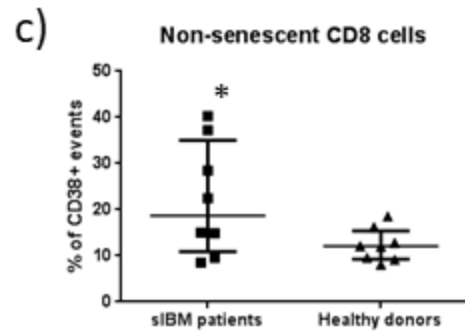
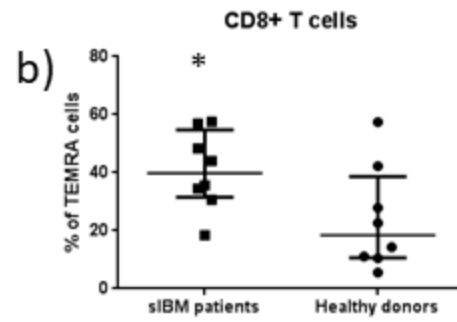
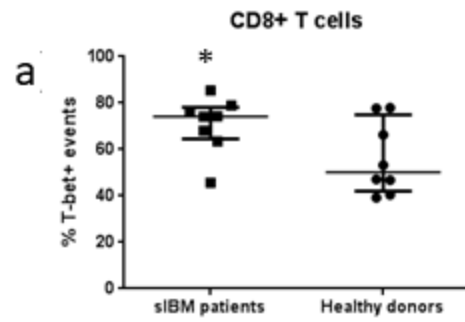
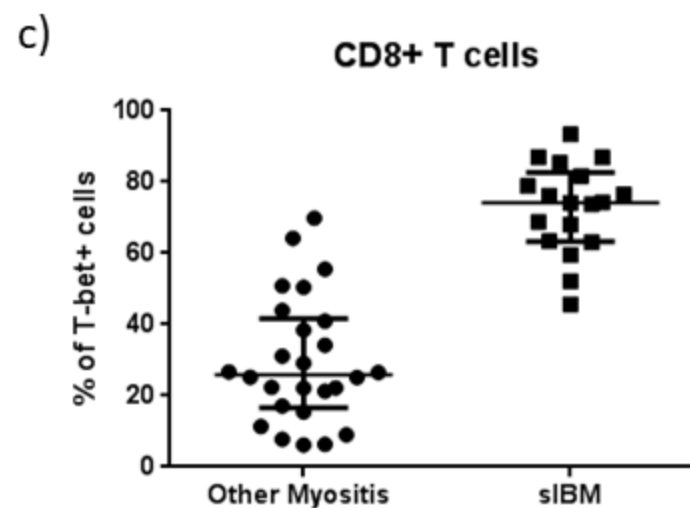
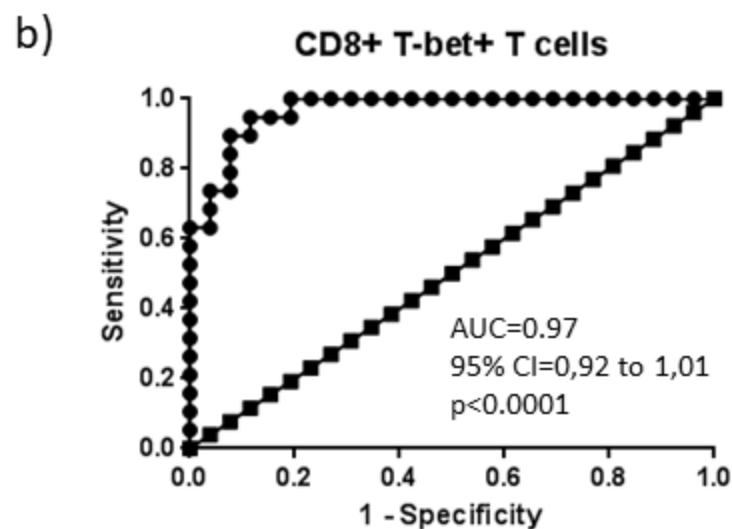
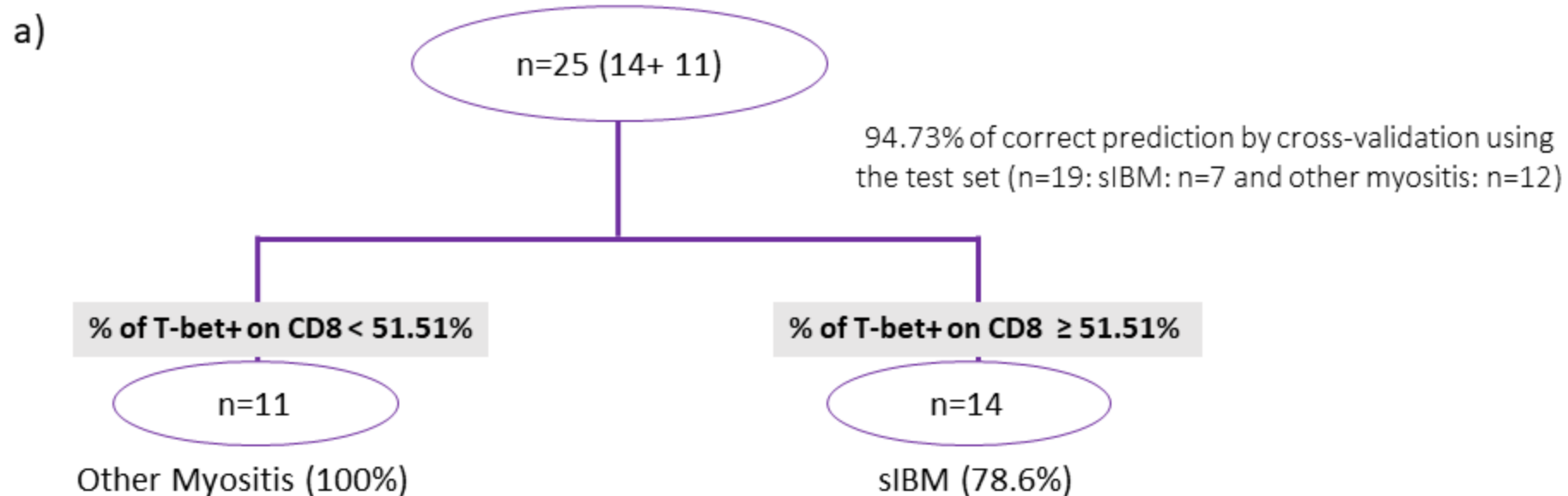
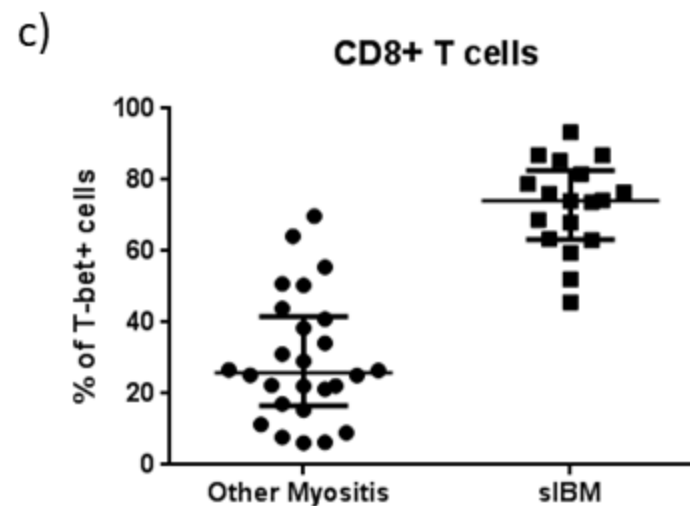
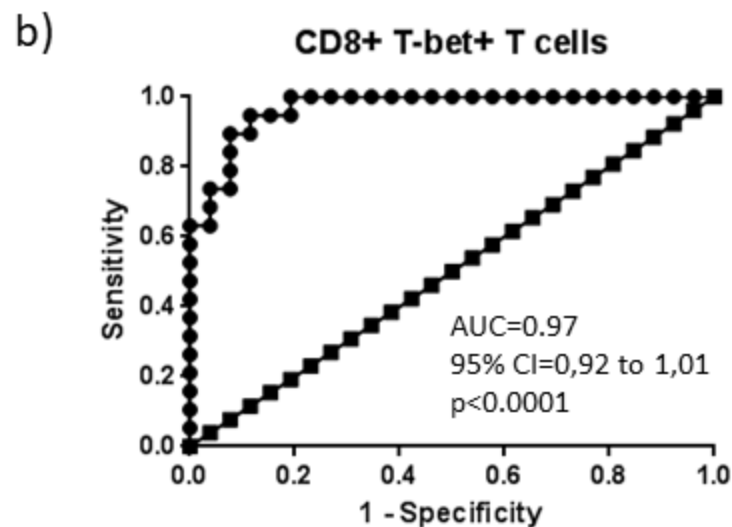
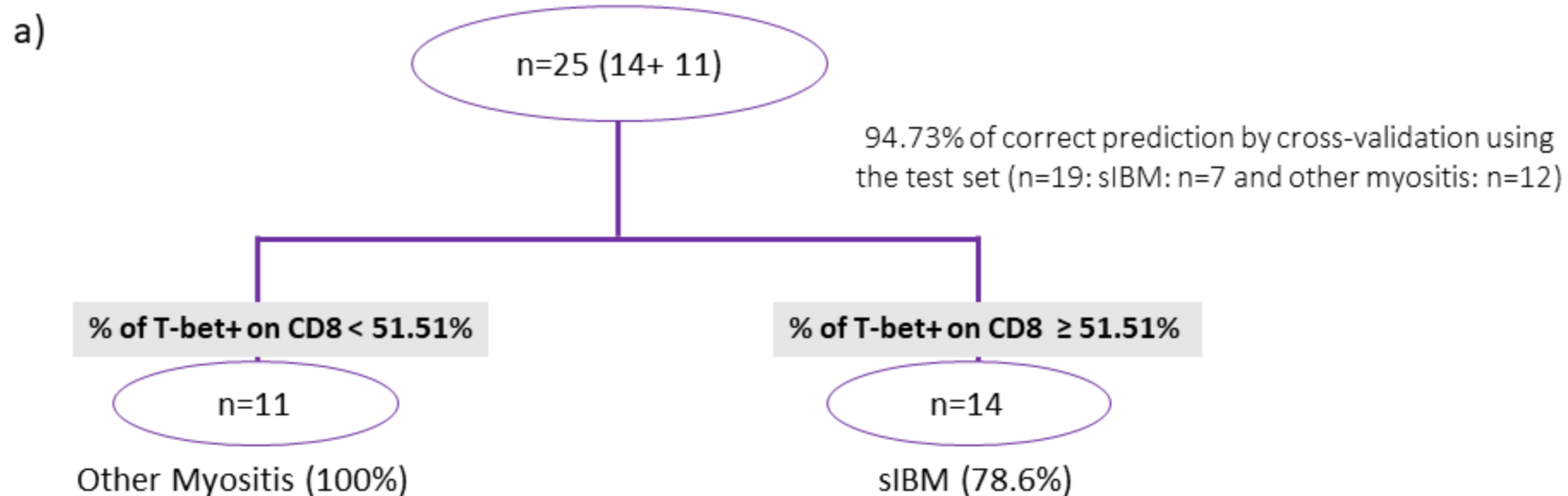


Fig 6



For frequencies of CD8+ T-bet+ T cells > 51.51%, **Se=94.44%**, **Sp=88.46**

Fig 6



For frequencies of CD8+ T-bet+ T cells > 51.51%, **Se=94.44%**, **Sp=88.46**