

Rational design of a novel turn-on fluorescent probe for detecting hydrazine with barbituric acid as recognition group and bioimaging

Junli Du^{a,b}, Xiaolu Li^{a,b}, Songsong Ruan^{a,b}, Yingchun Li^a, Fan Ren^{a,b}, Yanjun Cao^a, Xiaoqing Wang^c, Yongmin Zhang^{a,b,d}, Shaoping Wu^{a,b*}, Jianli Li^c

^a School of Pharmacy; Key Laboratory of Resource Biology and Biotechnology in Western China (Northwest University), Ministry of Education; Biomedicine Key Laboratory of Shaanxi Province, Northwest University, Xi'an 710069, China

^b Joint International Laboratory of Glycobiology and Medicinal Chemistry, Northwest University, Xi'an, Shaanxi 710069, China

^c Key Laboratory of Synthetic and Natural Functional Molecule Chemistry of Ministry of Education, College of Chemistry & Materials Science, Northwest University, Xi'an, Shaanxi 710127, P. R. China

^d Sorbonne Université, Institut Parisien de Chimie Moléculaire, CNRS UMR 8232, 4 place Jussieu, 75005 Paris, France

* Tel.: +86 029 88304569; Fax: +86 029 88304569. E-mail: wushaoping@nwu.edu.cn

Keywords: Fluorescent probe, Hydrazine, Barbituric acid, Large Stokes shift, Bioimaging

ABSTRACT:

A novel turn-on fluorescent probe with barbituric acid as a unique recognition group has been rational designed and facile synthesized for detecting hydrazine. Probe **DPT** displays a large emission signal ratio variation (more 40-fold enhancement) in the presence of hydrazine under neutral conditions. Interestingly, a novel recognition mechanism based on the hydrazine-triggered addition-cyclisation-retro aldol was proposed and confirmed. Additionally, probe **DPT** exhibits a low detection limit (5×10^{-8} M), applicability in physiological pH range (3~12), a broad linear response range of the hydrazine concentration between 0 and 34 μ M and large Stokes shift (147 nm) for hydrazine detection in aqueous solution. Moreover, probe **DPT** was triumphantly implemented for hydrazine imaging *in vivo*.

1. Introduction

Hydrazine (NH_2NH_2) is a kind of extremely reactive base which could be used in various fields, such as chemical, pharmaceutical, industrial and agricultural [1,2]. Statistical results indicate that more than 120 000 tons of approximately 64% hydrazine hydrate solutions are produced globally per year [3]. The spread of hydrazine would inevitably generate harm to the environment and human beings due to its high solubility in water. It could drive serious impairment to protein and nucleic acid structures. Hence hydrazine may cause severe organ injuries if excess exposure, such as liver, kidneys and the central nervous system [4,5]. Additionally, the U.S. Environmental Protection Agency advised a maximum threshold limit value of 10 ppb (0.31 μM) of hydrazine in drinking water as a carcinogenic substance [6,7]. Therefore, developing a kind of concise and efficient tool for the trace amounts of hydrazine detection in vitro and environment is of great significance, and also important to protect human health as well as understand its functions in complicated biological systems.

Up to now, a lot of traditional methods for the determination of hydrazine have been exploited, such as titrimetry [8], chromatography [9,10], capillary electrophoresis [11], electrochemical analysis [12,13], chemiluminescence [14], microflow detection [15] and so on. However, these analytical methods require complicated sample pretreatment which needs long time, high instrumentation cost and professional operation. Specially, individual methods may even produce damage to cell or tissues, thus becoming not appropriate for analyzing hydrazine *in vivo* and cell imaging. These shortcomings could limit the application of these methods. Therefore, fluorescence probe [16] based on sensing of hydrazine is receiving increased attention on account of its easy execution, low-priced, unique selectivity, extremely low sensitivity and on site analysis.

So far, a variety of dyes scaffold have been designed and synthesized for detecting hydrazine based on different responsive mechanism, such as rhodamine [17], BODIPY [18,19], coumarin [20,21], resorufin [22,23], benzothiazole [24,25],

heptamethine cyanine and hemicyanine [26,27]. On the other hand, a lot of fluorescent probes for hydrazine have been reported on the strength of three different mechanisms, including selective reaction with arylidenemalononitrile [28,29], selective deprotection of levulinoyl ester or acetyl groups [30,31,32], condensation with arylaldehydes [33,34], phthalimide [35,36] and 4-bromo butyrate [37,38,39]. However, the available fluorescent probes for hydrazine detection have some shortcomings. Hence, the design of novel fluorescent probe scaffold and its unique reaction mechanism for hydrazine is still a challenge which requires fast response time and high selectivity for the detection of hydrazine.

In this article, a novel colorimetric and turn on fluorescent probe with barbituric acid as a unique recognition group has been rationally designed and facilely synthesized for detecting hydrazine (**Fig. 1**). As expected, probe **DPT** shows an emission peak at 527 nm, excitation at 380 nm and displays a large emission signal ratio variation (more 40-fold enhancement) in the presence of hydrazine under neutral conditions. Surprisingly, a novel recognition mechanism based on retro-aldol reaction was confirmed. Additionally, probe **DPT** exhibits a low detection limit (5×10^{-8} M), applicability in physiological pH range, a broad linear response range of the hydrazine concentration from 0 to 34 μ M and large Stokes shift (147 nm) for hydrazine detection at physiological conditions. Furthermore, probe **DPT** was successfully utilized for cell imaging of hydrazine in living cells.

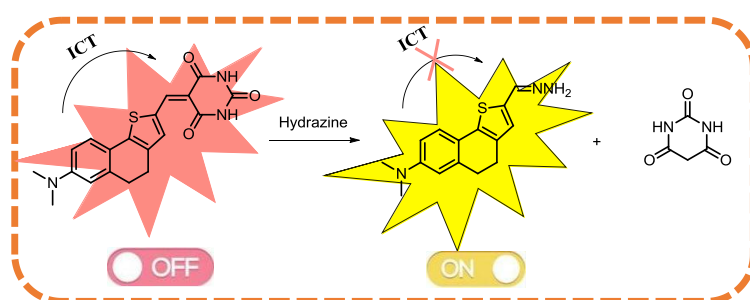


Fig. 1. Proposed mechanism of probe **DPT** towards hydrazine.

2. Materials and methods

2.1. Materials and Instrumentation

All solvents and chemical reagents for synthesis were commercially available.

6-amino-1-tetralone, hydrazine monohydrate solution, sodium sulphide and chloroacetaldehyde were purchased from J&K Shanghai Scientific Ltd in China. Stock solution of hydrazine was prepared from the commercial hydrazine monohydrate reagent. The UV-vis spectra were measured on a Shimadzu UV-2550 and the fluorescence spectra were performed *via* Hitachi F-7000 spectrophotometer. The cell imaging was collected by the FV1000 laser confocal fluorescence microscope of Olympus Company.

2.2. Synthesis of probe 5-((7-(dimethylamino)-4,5-dihydronaphtho [1,2-b] thiophen-2-yl)methylene)pyrimidine-2,4,6 (1H,3H,5H)-trione (**DPT**)

Compound **2** (50.0 mg, 0.195 mmol, 1.0 equiv.) and barbituric acid (37.4 mg, 0.29 mmol, 1.5 equiv.) in dry EtOH (5.0 mL) was blended at 25 °C, then 0.5 mL of acetic acid glacial was added as catalyst. The reaction mixture was heated for 2 h at 85 °C. The formed precipitate was filtered off by funnel, washed with ice ethanol and dried in vacuum, the pure probe **DPT** was obtained as a black powder (yield: 75.5%, $R_f = 0.74$). ^1H NMR (600 MHz, TFA- d_1) δ 11.50 (s, 1H), 11.50 (s, 1H), 8.88 (s, 1H), 7.91 (s, 1H), 7.87 (s, 1H), 7.51 (s, 2H), 3.41 (s, 6H), 3.08 (s, 2H), 2.98 (s, 2H). ^{13}C NMR (151 MHz, TFA- d_1) δ 163.62, 155.71, 151.84, 151.74, 151.11, 141.74, 140.97, 135.55, 131.98, 126.72, 126.23, 119.59, 118.94, 118.18, 117.55, 105.01, 46.27, 27.23, 20.98. HRMS ($\text{C}_{19}\text{H}_{17}\text{N}_3\text{O}_3\text{S}$): calcd. for $[\text{M}-\text{H}]^-$ 366.0912; found: $[\text{M}-\text{H}]^-$ 366.0913 (**Fig. S4-S6**).

2.3. Titration experiments of probe **DPT**

A stock solution of probe **DPT** (1.0 mM in DMF) was prepared for spectrum titrations. The 1.0 mM solution of $\text{N}_2\text{H}_4 \cdot \text{H}_2\text{O}$ was readied in pure water. Usually, test solutions were prepared by adding 30 μL of the **DPT** stock solution into a 5 mL colorimetric tube, diluting the solution with 10 mM PBS buffer solution (pH = 7.4, 20% DMSO). Then different analyte was dissolved in ultrapure water and respectively added into the test solution.

2.4. Cytotoxicity experiments

The cytotoxicity of probe **DPT** was detected using cell counting kit-8 (CCK-8). SH-SY5Y neuroblastoma cell was afforded from the ATCC Cell Bank. The cells were cultured for 24 h at 37 °C with 5% CO₂ in DMEM (supplemented with 10% fetal bovine serum (FBS)) after it was seeded in 96-well plates at a moderate density of cells per well. Then the cells were hatched with probe **DPT** at 0, 5, 10, 15, 20 μM concentrations for 24 h, next 10 μL CCK-8 solution was added to each hole of the 96-well plate for 2 h at 37 °C, the absorbance of cells were recorded by a microplate reader at 450 nm . Each treatment group was repeated five times.

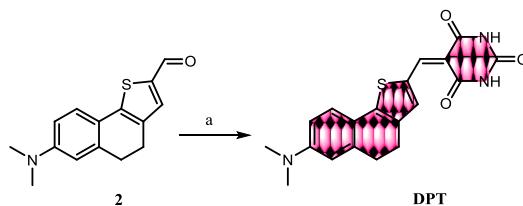
2.5. Imaging of probe DPT in living cell

For cell imaging of hydrazine, the SH-SY5Y neuroblastoma cell was put into a 10 mm diameter culture vessel and permitted to adhere for 24 h. Before cell imaging, the SH-SY5Y neuroblastoma cells were hatched with 10 μM probe **DPT** at 37 °C for 5 min, and followed by lavation three times with pH = 7.4 PBS buffer solution to move out overabundance **DPT**, next in situ treated with 0, 10, 20, 30 μM concentrations of N₂H₄·H₂O solution at 37 °C for 15 min, respectively. Then the cells were performed with a laser confocal microscope after it was laundered with PBS three times to move out excess ions.

3. Results and discussion

3.1. Design and synthesis of probe DPT

As detailed in **Scheme 1**, probe **DPT** was rationally designed and conveniently synthesized in one step. Starting material 6-amino-1-tetralone is commercially available. Compound **1** was generated by dimethylation of 6-amino-1-tetralone with methyl iodide and K₂CO₃ in DMF [40]. The aldehyde-chloride compound was generated which was utilized to produce compound **2** with Na₂S in chloroacetaldehyde and K₂CO₃ at 60 °C [41]. Ultimately, the compound **2** was reacted with barbituric acid in absolute ethanol solution to obtain probe **DPT** at 85 °C [42]. The elaborated synthetic protocols were delineated in the supplementary Material.



Scheme 1. Reagents and conditions: (a) Barbituric acid, Ethanol, reflux, 85 °C, 2 h, 75.5%.

3.2. UV-vis absorbance and fluorescence spectra study

The excitation and emission spectra of probe **DPT** were explored in 20% DMSO co-solution with 10.0 mM PBS buffer solution (pH 7.4) at 25 °C. As a structurally novel probe, **DPT** was assessed for its UV-vis and fluorescence properties in various kind polarity solvents including MeOH, EtOH, ACN, DMK, DMSO, DMF, TCM, DCM, EtOAc, H₂O, THF and so on. At the same time, the fluorescence quantum yield of **DPT-N₂H₄** addition compound was calculated *via* fluorescein ($\Phi_f = 0.92$, 0.1 M NaOH) as the standard compound. The detailed photophysical properties of probe **DPT** were illustrated in **Table S2**.

The absorbance spectral responses of probe **DPT** toward hydrazine have been measured in the mixture solvent of 20% DMSO and 10.0 mM PBS buffer system (pH 7.4). As depicted in **Fig. 2**, the absorbance peak at 585 nm was increased by degrees and had an excellent linear functional relationship with hydrazine between 0 and 20 μ M ($R^2 = 0.9942$) along with the hydrazine concentration increase. Meanwhile, the color change of the solution from blue purple to colorless also can be distinctly observed along with these absorption value changes under daylight lamp in **Fig. 4a**. So probe **DPT** could be used as naked eye visible hydrazine sensors in aqueous solution owing to their sensitive color sense to the solution hydrazine color change.

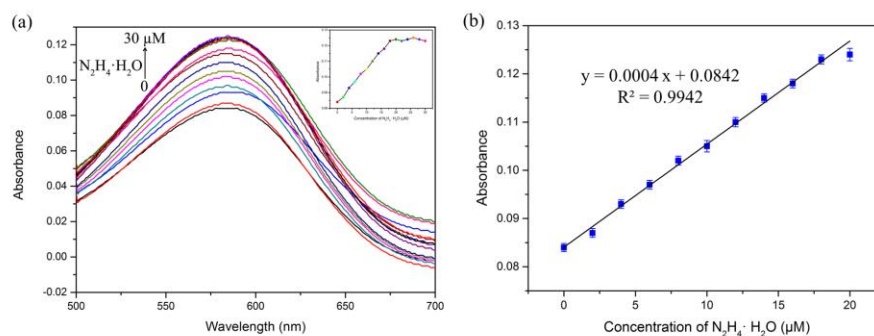


Fig. 2. (a) UV-vis absorption spectra of probe **DPT** (6 μM) upon addition of 0~30 μM $\text{N}_2\text{H}_4\cdot\text{H}_2\text{O}$ in a 20% DMSO solution and 10.0 mM PBS buffer, pH 7.4. (b) Linear relationships between UV-vis absorbance at 585 nm and concentrations of $\text{N}_2\text{H}_4\cdot\text{H}_2\text{O}$.

The fluorescent response of probe **DPT** to hydrazine was shown in **Fig. 3**, probe **DPT** itself had weak fluorescence emission at 650 nm, but the fluorescence intensity gradually increased along with enhancement in hydrazine concentration changes at 525 nm with a wide emission band from 450 nm to 650 nm. Additionally, between the fluorescence intensities with hydrazine concentration have a good linearity ($R^2 = 0.9982$) in 0~26 μM , the regression equation was $Y = 361.7531X - 66.1829$ from the linear calibration graphs with the fluorescence titration experiments. On the other hand, the minimum LOD of probe **DPT** for hydrazine was 50 nM using signal to noise ratio (S/N) = 3, it was satisfied to the detection of hydrazine. Further experiments phenomenon indicated that the fluorescent color change from light red to yellow is shown in **Fig. 4b** at 365 nm light. These data illustrated that probe **DPT** could be an excellent on-off-type fluorescent probe to detect hydrazine concentration quantitatively in complicated background sample.

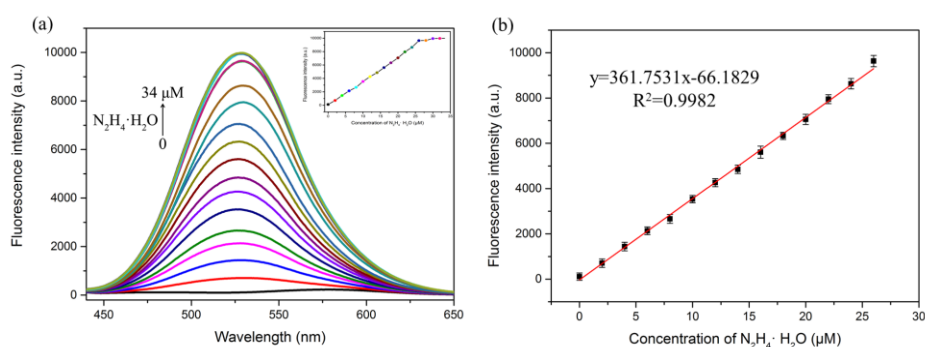


Fig. 3. (a) Fluorescence titration spectra of probe **DPT** (6 μM) upon addition of 0~34 μM $\text{N}_2\text{H}_4\cdot\text{H}_2\text{O}$ in 10 mM PBS (pH 7.4, containing 20% DMSO). $\lambda_{\text{ex}} = 380$ nm, $\lambda_{\text{em}} = 525$ nm. slit: 5.0 nm/5.0 nm. (b) Linear relationship between probe **DPT** (6 μM) at 525 nm and $\text{N}_2\text{H}_4\cdot\text{H}_2\text{O}$ concentration (0~26 μM).

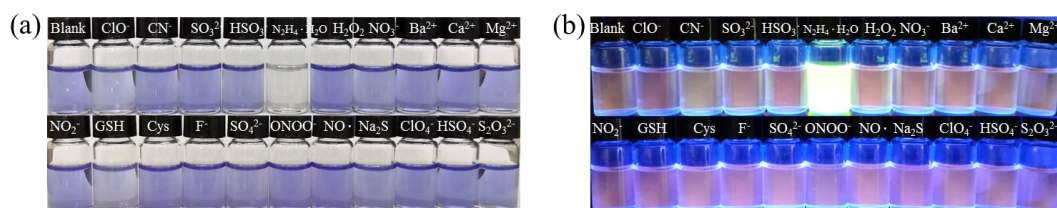


Fig. 4. Color change of probe **DPT** solution upon addition various ions under daylight lamp (a) and ultraviolet lamp (b). Fluorescence ($\lambda_{\text{ex}} = 365$ nm) change upon addition of different metal ions

and anions in PBS buffer (pH 7.4, 10 mM, containing 20% DMSO) solution. (From left to right: Blank, ClO^- , CN^- , SO_3^{2-} , HSO_3^- , H_2O_2 , NO_3^- , Ba^{2+} , Ca^{2+} , Mg^{2+} , NO_2^- , GSH, Cys, F^- , SO_4^{2-} , ONOO^- , $\text{NO}\cdot$, Na_2S , ClO_4^- , HSO_4^- , $\text{S}_2\text{O}_3^{2-}$, $\text{N}_2\text{H}_4\cdot\text{H}_2\text{O}$).

3.3. Selectivity and competitive experiment

The selectivity of probe **DPT** toward $\text{N}_2\text{H}_4\cdot\text{H}_2\text{O}$ were investigated by various biologically relevant analytes including anions, cations and ROS. As displayed in **Fig. 5a**, only the fluorescence intensity of probe **DPT** markedly raised along with the addition of $\text{N}_2\text{H}_4\cdot\text{H}_2\text{O}$, the fluorescence intensity was no obvious changed along with addition of other analytes in the same condition.

Next the anti-interference property of probe **DPT** with various analytes has been also studied as shown in **Fig. 5b**, the detection of $\text{N}_2\text{H}_4\cdot\text{H}_2\text{O}$ did not interfere in present the of all the competing analytes, these outcomes suggested that probe **DPT** has unique selectivity for $\text{N}_2\text{H}_4\cdot\text{H}_2\text{O}$ over multifarious interfering species that could be exist in living cells and environment. In a word, it was capacitated to monitor $\text{N}_2\text{H}_4\cdot\text{H}_2\text{O}$ in sophisticated biological sample.

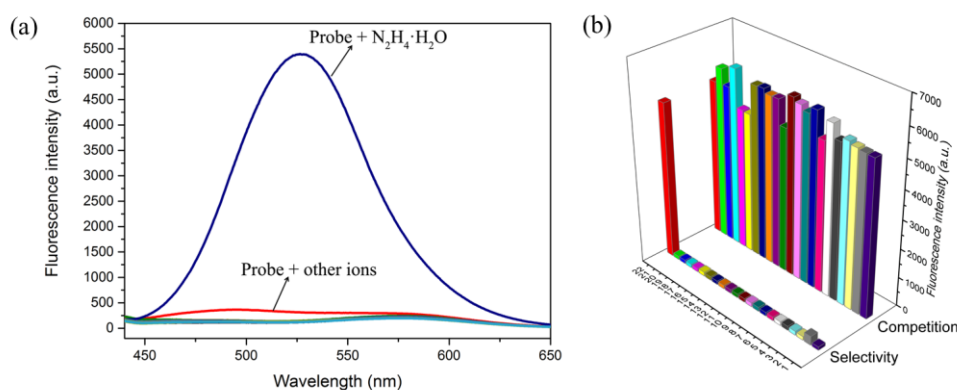


Fig. 5. (a) Fluorescence spectra of probe **DPT** (6 μM) upon addition 20 μM of different ions in PBS buffer (pH 7.4, 10 mM, containing 20% DMSO) solution ($\lambda_{\text{ex}} = 380 \text{ nm}$, slit = 5.0 nm/5.0 nm). (b) The pillars in the front row represent fluorescence response of the probe to the metal ions of interest. The pillars in the back row represent the subsequent addition 20 μM $\text{N}_2\text{H}_4\cdot\text{H}_2\text{O}$ to the solution containing probe and the metal ions, respectively. (1. Blank, 2. ClO^- , 3. CN^- , 4. SO_3^{2-} , 5. HSO_3^- , 6. H_2O_2 , 7. NO_3^- , 8. Ba^{2+} , 9. Ca^{2+} , 10. Mg^{2+} , 11. NO_2^- , 12. GSH, 13. Cys, 14. F^- , 15. SO_4^{2-} , 16. ONOO^- , 17. $\text{NO}\cdot$, 18. Na_2S , 19. ClO_4^- , 20. HSO_4^- , 21. $\text{S}_2\text{O}_3^{2-}$, 22. $\text{N}_2\text{H}_4\cdot\text{H}_2\text{O}$).

3.4. Time kinetic and pH-tolerability of probe DPT

The reaction kinetics is a vital factor for the optical signal changing between probe **DPT** and hydrazine concentration, so the response kinetics of probe **DPT** to

hydrazine was also investigated. As displayed in **Fig. 6a**, a plateau of fluorescence intensity changes could be achieved within 10 min upon addition of 20 μM of hydrazine in the probe aqueous solution. Therefore, we selected 10 min as the optimum reaction time in the all experiments.

For the sake of the complex biological sample applications, the optimal pH range of probe **DPT** is crucial to be found. The fluorescence intensity of individual probe **DPT** and probe **DPT** with $\text{N}_2\text{H}_4\cdot\text{H}_2\text{O}$ were investigated in 20% DMSO aqueous solution under different pH values, respectively. It was found that probe **DPT** did not display any remarkable fluorescence intensity change in the wide pH range of 3~12. Furthermore, the mixture solution of probe **DPT** with hydrazine showed a outstanding fluorescence increase at 525 nm and the fluorescence intensity was a very little change in the wide pH range of 5~10. These results show that probe **DPT** and **DPT-N₂H₄** were very stable in the wide pH range of 3~12, which indicates probe **DPT** is suitable for detection of hydrazine concentration *in vivo* and *in vitro* sample under physiological conditions.

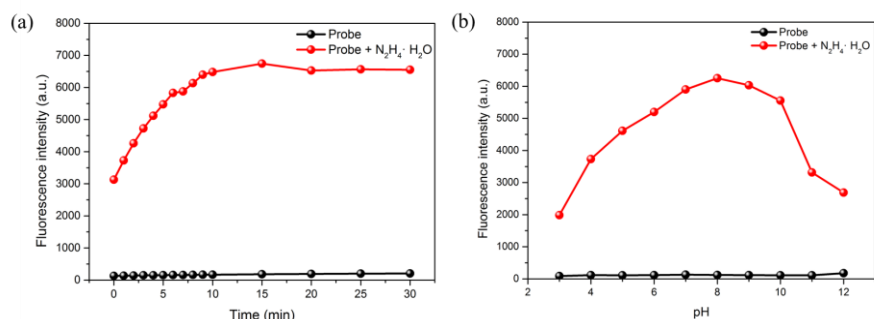
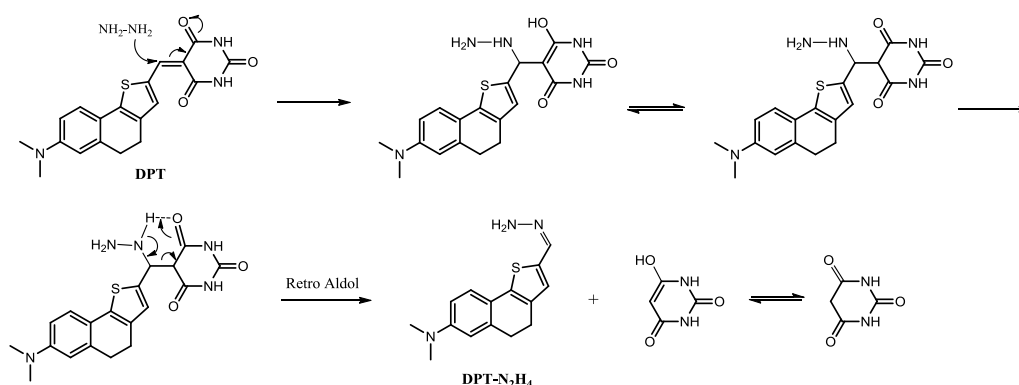


Fig. 6. (a) Time-kinetic fluorescence changes of probe **DPT** (6 μM) with $\text{N}_2\text{H}_4\cdot\text{H}_2\text{O}$ (20 μM) in 10 mM PBS (pH 7.4, 20% DMSO). (b) Changes in fluorescence spectra of probe **DPT** (6 μM) in the absence and present of $\text{N}_2\text{H}_4\cdot\text{H}_2\text{O}$ (20 μM) as a function of pH. $\lambda_{\text{ex}} = 380$ nm, slits: 5.0 nm/5.0 nm.

3.5 Recognition mechanism and theoretical computation

On the basis of these spectral results, the proposed recognition mechanism of probe **DPT** toward hydrazine is speculated as follows: in the presence of hydrazine, the barbituric acid moiety bearing on probe **DPT** could react with hydrazine by addition reaction, and then the intermediate alcohol would turn into keto by transformation of enol. Then the barbituric acid moiety would be released by retro-aldol reaction to provide the elimination product **DPT-N₂H₄** (**Scheme 2**). The

proposed recognition mechanism was confirmed by MS spectrum, **DPT** (50 μM) and hydrazine (60 μM) was reacted each other for 5 min in THF solution at 25 $^{\circ}\text{C}$, the obtained solution was timely monitored to high resolution mass spectroscopy after suitably dilution, an unexpected peak was checked at m/z 272.1228 in accord with **DPT-N₂H₄** ($\text{C}_{15}\text{H}_{18}\text{N}_3\text{S}$, Exact Mass: $[\text{M}+\text{H}]^+$ 272.1221) (**Fig. S7**). Therefore, all these data supports the hydrazine-triggered addition-cyclisation-retro aldol recognition mechanism.



Scheme 2. Proposed mechanism for the reaction of probe **DPT** with $\text{N}_2\text{H}_4 \cdot \text{H}_2\text{O}$.

To roundly comprehend the spectral changes of probe **DPT** responding to hydrazine, theoretical calculations were executed to use TD-DFT (time dependent density functional theory) with the PBE0/6-31+G(d, p) in a suite of the Gaussian 09 program. The solvent effects were thought about in DMSO by the polarizable continuum model (PCM) [43]. The optimized geometries and the HOMO and LUMO of probe **DPT** and **DPT-N₂H₄** are presented in **Fig. S8**. Molecular orbital surfaces of the HOMO and LUMO for probe **DPT** and additive product **DPT-N₂H₄** were shown in **Fig. 7**. For probe **DPT**, the electron density of the HOMO was spread across the whole probe scaffold. On the contrary, that of the LUMO was concentrated on the thiofuran part and barbituric acid moiety, respectively. As for **DPT-N₂H₄**, the electron in HOMO and LUMO orbital was chiefly spread over the naphthalene skeleton. Remarkably, ICT process didn't occur in the **DPT-N₂H₄**, in that a sp^2 hybridized carbon atom existed in thiofuran. The disruption of the ICT course generated in the raised HOMO-LUMO energy gap of **DPT-N₂H₄** (3.598 eV)

compared with that of the **DPT** (2.671 eV). These above calculations result confirmed that $\text{N}_2\text{H}_4\cdot\text{H}_2\text{O}$ was reacted to the barbituric acid moiety and are identical with the spectra data.

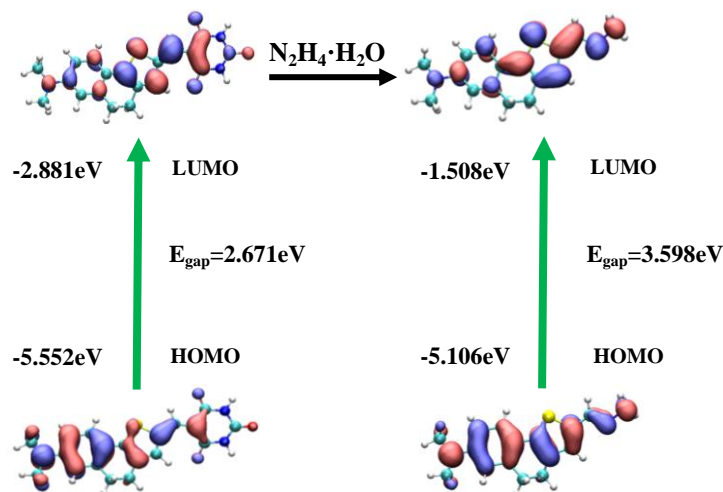


Fig. 7. HOMO-LUMO energy levels and frontier molecular orbital of probe **DPT** and the adduct **DPT-N₂H₄**.

3.6. Cytotoxicity and bioimaging in living cells

It is great concern to safety of probe **DPT** on living SH-SY5Y neuroblastoma cells. The cellular viability of probe **DPT** was estimated to be greater than 90% after 1 h at 10 μM concentration, and the cellular viability was 75% at the concentration of 25 μM by CCK-8 assay using standard cell viability protocols (**Fig. S9**). Briefly, probe **DPT** was the low cytotoxicity in SH-SY5Y neuroblastoma cells, these results suggest that probe **DPT** could be applied for detecting the $\text{N}_2\text{H}_4\cdot\text{H}_2\text{O}$ level in living cells.

Next, the ability of probe **DPT** to visualize hydrazine in live cell was also studied by using fluorescence microscopic imaging (**Fig. 8**). SH-SY5Y neuroblastoma cell was incubated with 10 μM probe **DPT** and incubated with **DPT** (10 μM) followed by hydrazine (20 μM) in DMEM at 37 $^\circ\text{C}$, respectively. The incubation time was 10 min. Weak fluorescence was detected at blue channel in the cytoplasm. After addition of hydrazine (20 μM), strong fluorescence signal was observed at the blue channel, suggesting the reaction of probe **DPT** with hydrazine. The high stability, low toxicity and excellent bioimaging make probe **DPT** a successful imaging tool for hydrazine.

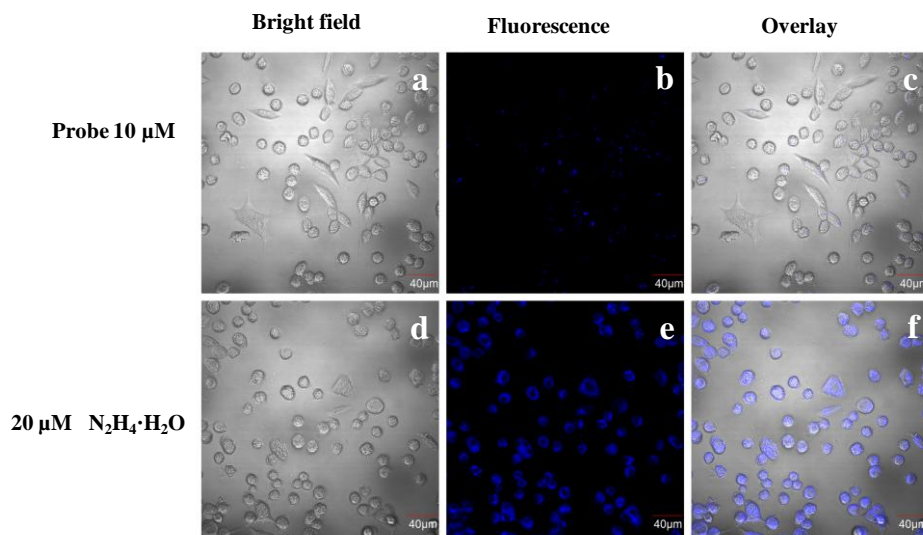


Fig. 8. Confocal fluorescence microscopic images of SH-SY5Y neuroblastoma cells. that cells incubated by probe **DPT** (10 μM) for 10 min at 37 $^{\circ}\text{C}$ and observed under bright field (a), blue channel (b), overlay (c), then further incubation with $\text{N}_2\text{H}_4\cdot\text{H}_2\text{O}$ (20 μM) for 15 min at 37 $^{\circ}\text{C}$ and observed under bright field, blue channel, overlay.

4. Conclusion

In conclusion, a novel on-off fluorescent probe was concise synthesized with barbituric acid as unique recognition group for detection of $\text{N}_2\text{H}_4\cdot\text{H}_2\text{O}$ *via* simply three steps. A novel recognition mechanism based on the hydrazine-triggered addition-cyclisation-retro aldol process was proposed and confirmed by HRMS. Probe **DPT** exhibits low limit of detection, large Stokes shift and broad pH tolerability range in practical applications. Additionally, probe **DPT** can be successfully implemented to indicate exogenous bioimaging of $\text{N}_2\text{H}_4\cdot\text{H}_2\text{O}$ in SH-SY5Y neuroblastoma cells with negligible cytotoxicity and excellent biocompatibility. Moreover, we desire that a novel probe **DPT** could be utilized to advance disclose some necessary information of hydrazine in extensive samples.

Acknowledgements

Thank for the Program for Changjiang Scholars and Innovative Research Team in University (No. IRT-15R55), Scientific Research Program Funded by Shaanxi Provincial Education Department (No. 18JK0774), the International Science & Technology Cooperation Program of Shaanxi Province (No. 2019KWZ-001), Program of Innovation for Undergraduates of Northwest University (No. 2019253),

Program of Graduate Quality Improvement of Northwest University (No. YZZ17169).

Appendix A. Supplementary data

Supplementary data associated with this article can be found, in the online version, at <http://>

References

- [1] J. Sanabria-Chinchilla, K. Asazawa, T. Sakamoto, K. Yamada, H. Tanaka, P. Strasser, *J. Am. Chem. Soc.*, 2011, **133**, 5425-5431.
- [2] A. Serov, M. Padilla, A. J. Roy, P. Atanassov, T. Sakamoto, K. Asazawa, H. Tanaka, *Angew. Chem., Int. Ed.*, 2014, **53**, 10336-10339.
- [3] J. P. Schirmann, P. Bourdauducq, Ullmann's Encyclopedia of Industrial Chemistry, Wiley-VCH, Weinheim, 2002.
- [4] C. A. Reilly, S. D. Aust, *Chem. Res. Toxicol.*, 1997, **10**, 328-334.
- [5] G. Wang, C. Zhang, X. He, Z. Li, X. Zhang, L. Wang, B. Fang, *Electrochim. Acta*, 2010, **55**, 7204-7210.
- [6] U. S. E. P. A., Integrated Risk Information System (IRIS) on Hydrazine/Hydrazine Sulfate. National Center for Environmental Assessment, Office of Research and Development Washington, D. C., 1999.
- [7] U. S. E. P. A., *Hydrazine Hazard Summary-Created in April 1992; Revised in January 2000*, Office of Research and Development Washington, D. C., 2008.
- [8] S. D. Zelnick, D. R. Mattie, P. C. Stepaniak, *Aviat. Space. Env. Med.*, 2003, **74**, 1285-1291.
- [9] M. J. Sun, L. Bai, D. Q. Liu, *J. Pharm. Biomed. Anal.*, 2009, **49**, 529-533.
- [10] A. Umar, M. M. Rahman, S. H. Kim, Y. B. Hahn, *Chem. Commun.*, 2008, 166-168.
- [11] J. Liu, W. Zhou, T. You, F. Li, E. Wang, S. Dong, *Anal. Chem.*, 1996, **68**, 3350-3353.
- [12] A. Benvidi, P. Kakoolaki, H. R. Zare, R. Vafazadeh, *Electrochim. Acta*, 2011, **56**, 2045-2050.
- [13] J. Cheng, E. Yang, P. Ding, J. Tang, D. Zhang, Y. Zhao, Y. Ye, *Sens. Actuators, B*, 2015, **221**, 688-93.
- [14] A. Safavi, M. A. Karimi, *Talanta*, 2002, **58**, 785-92.
- [15] Z. He, B. Fuhrmann, U. Spohn, *Anal. Chim. Acta*, 2000, **409**, 83-91.
- [16] (a) W. Y. Feng, G.Q. Feng, *Dyes Pigm.*, 2019, **164**, 174-181; (b) J. X. Hong, W. Y. Feng, G. Q. Feng, *Sens. Actuators, B*, 2018, **262**, 837-844; (c) W. Y. Feng, S. Y. Gong, E. B. Zhou, X. Y. Yin, G. Q. Feng, *Anal. Chim. Acta*, 2018, **1029**, 97-103.
- [17] S. Nandi, A. Sahana, S. Mandal, A. Sengupta, A. Chatterjee, D. A. Safin, M. G. Babashkina, N. A. Tumanov, Y. Filinchuk, D. Das, *Anal. Chim. Acta*, 2015, **893**, 84-90.
- [18] Y. D. Lin, T. J. Chow, *RSC Adv.*, 2013, **3**, 17924-17929.
- [19] S. Goswami, S. Paul, A. Manna, *RSC Adv.*, 2013, **3**, 18872-18877.

- [20] M. G. Choi, J. Hwang, J. O. Moon, J. Sung, S. K. Chang, *Org. Lett.*, 2011, **13**, 5260-5263.
- [21] H. Tse, Q. H. Li, S. M. Chan, Q. H. You, W. M. Lee, W. H. Chan, *RSC Adv.*, 2016, **6**, 14678-14681.
- [22] X. D. Jiang, J. Guan, H. Bian, Y. Xiao, *Tetrahedron Lett.*, 2017, **58**, 2351-2354.
- [23] T. Tang, Y. Chen, B. S. Fu, Z. Y. He, H. Xiao, F. Wu, J. Q. Wang, S. R. Wang, X. Zhou, *Chin. Chem. Lett.*, 2016, **27**, 540-544.
- [24] S. Goswami, S. Das, K. Aich, B. Pakhira, S. Panja, S. K. Mukherjee, S. Sarkar, *Org. Lett.*, 2013, **15**, 5412-5415.
- [25] J. Zhou, R. Y. Shi, J. X. Liu, R. Wang, Y. F. Xu, X. H. Qian, *Org. Biomol. Chem.*, 2015, **13**, 5344-5348.
- [26] C. Hu, W. Sun, J. F. Cao, P. Gao, J. Y. Wang, J. L. Fan, F. L. Song, S. G. Sun, X. J. Peng, *Org. Lett.*, 2013, **15**, 4022-4025.
- [27] Z. L. Lu, W. L. Fan, X. M. Shi, Y. N. Lu, C. H. Fan, *Anal. Chem.*, 2017, **89**, 9918-9925.
- [28] J. Fan, W. Sun, M. Hu, J. Cao, G. Cheng, H. Dong, K. Song, Y. Liu, S. Sun, X. Peng, *Chem. Commun.*, 2012, **48**, 8117-8119.
- [29] Y. Tan, J. Yu, J. Gao, Y. Cui, Y. Yang, G. Qian, *Dyes Pigm.*, 2013, **99**, 966-971.
- [30] Y. Sun, D. Zhao, S. Fan, L. Duan, *Sens. Actuators, B*, 2015, **208**, 512-517.
- [31] (a) K. Li, H. R. Xu, K. K. Yu, J. T. Hou, X. Q. Yu, *Anal. Methods*, 2013, **5**, 2653-2656; (b) Q. S. Zhai, W. Y. Feng, G. Q. Feng, *Anal. Methods*, 2016, **8**, 5832-5837.
- [32] B. Liu, Q. Liu, M. Shah, J. Wang, G. Zhang, Y. Pang, *Sens. Actuators, B*, 2014, **202**, 194-200.
- [33] L. Xiao, J. Tu, S. Sun, Z. Pei, Y. Pei, Y. Pang, Y. Xu, *RSC Adv.*, 2014, **4**, 41807-41811.
- [34] X. Chen, X. Yu, Z. Li, A. Tong, *Anal. Chim. Acta*, 2008, **625**, 41-46.
- [35] L. Cui, Z. Peng, C. Ji, J. Huang, D. Huang, J. Ma, S. Zhang, X. Qian, Y. Xu, *Chem. Commun.*, 2014, **50**, 1485-1487.
- [36] L. Cui, C. Ji, Z. Peng, L. Zhong, C. Zhou, L. Yan, *Anal. Chem.*, 2014, **86**, 4611-4617.
- [37] S. Goswami, S. Das, K. Aich, B. Pakhira, S. Panja, S. K. Mukherjee, S. Sarkar, *Org. Lett.*, 2013, **15**, 5412-5415.
- [38] Y. Qian, J. Lin, L. Han, L. Lin, H. Zhu, *Biosens. Bioelectron.*, 2014, **58**, 282-286.
- [39] X. Jin, C. Liu, X. Wang, H. Huang, X. Zhang, H. Zhu, *Sens. Actuators, B*, 2015, **216**, 141-149.
- [40] L. Q. Cui, Z. L. Dong, K. Liu, C. Zhang, *Org. Lett.*, 2011, **13**, 6488-6491.
- [41] C. Herbivo, A. Comel, G. Kirsch, M. Manuela, M. Raposo, *Tetrahedron*, 2009, **65**, 2079-2086.
- [42] T. Sun, Q. F. Niu, Y. Li, T. D. Li, T. T. Hu, E. H. Wang, H. X. Liu, *Sens. Actuators, B*, 2018, **258**, 64-71.
- [43] J. Tomasi, B. Mennucci, E. Cancès, *J. Mol. Struct. (Theochem)*, 1999, **464**, 211-226.