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Preparation of unsymmetrically 2^A,3^B,6^{C(F)}-trihydroxy-per-*O*-methylated αcyclodextrin and NMR analysis

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ABSTRACT

An straightforward approach wherein an excess of diisobutylaluminum hydride (DIBAL-H, 25.0 equiv), used as a molecular scalpel, is able to strip off three methyl groups from both primary and second faces of pemethylated α -cyclodextrin **1** to provide an unsymmetric 2^A , 3^B , $6^{C(F)}$ -trihydroxy-per-*O*-methylated α -CD **6** in one step in 35% yield is first described.

Keywords:

α-Cyclodextrin; DIBAL-H; Tri-de-O-methylation; Regioselectivity.

Cyclodextrins (CDs) are a family of well-known cyclic oligosaccharides consisting of a macrocycle ring of D-gulcose subunits jointed by α -1,4-linked glycosidic bonds. A general characteristic of CDs is their ability to form inclusion complexes with both hydrophilic and hydrophobic guest molecules to increase their solubility, dissolution rate, stability, etc.¹ Recently, per-*O*-methylated CDs and their derivatives have attracted considerable attention due to their solubility in water (> 500 mg/mL) as well as in organic solvents.²⁻⁴ More importantly, inclusion complexes of methylated CDs are usually more stable than the corresponding complexes of unmodified CDs.⁵⁻⁷ There has been a growing interest in a selective chemical modification of per-*O*-methylated CDs with various functional groups.⁸⁻¹⁴ However, up till now, highly selective modification of per-*O*-methylated CDs both at primary and secondary faces remains a significant challenge for synthetic chemists.

In 2000, Sinaÿ et al ¹⁵ discovered that diisobutylaluminium hydride (DIBAL-H) is a reagent of choice for the achievement of a remarkable mono-de-*O*-benzylation at the 6^A-position or bis-de-*O*-benzylation at the 6^A- and 6^D-positions of perbenzylated α - and β -CD. These results contrast with DIBAL-H promoted a selective mono-de-*O*-methylation at the 6^A-position or bis-de-*O*-methylation at the 2^A- and 3^B-positions of per-*O*-methylated α - and β -CD.¹³ To further gain an insight into the regioselective de-*O*-methylation properties of DIBAL-H towards per-*O*-methylated α - and β -CD, we have extensively studied the selective de-*O*-methylation of per-*O*-methylated α and β -CD by DIBAL-H,¹⁶⁻²⁰ a general simple way to access to 6^A-hydroxyl-per-*O*methylated α -CD **2**, 2^A,3^B-dihydroxyl-per-*O*-methylated α -CD **3**, 2^A,3^B,2^C,3^Dtetrahydroxy-per-*O*-methylated α -CD **4** and 2^A,3^B,2^D,3^E-tetrahydroxy-per-*O*methylated α -CD **5** from per-*O*-methylated α -CD **1** was developed, respectively (Scheme 1), using DIBAL-H as a chemical "scalpel".^{13, 19}



Scheme 1. Selective de-O-methylation of per-O-methylated α -CD by DIBAL-H.^{13, 19}

In our previous study,¹⁶ a mixture of two tri-de-*O*-methylation derivatives of per-*O*-methylated β -CD promoted by DIBAL-H (50 equiv) was first observed in 17% yield and identified by FAB-MS, but was not analysed further at that stage. As a continuity to this work, we would now like to report the tri-de-*O*-methylation on both primary and secondary rims of per-*O*-methylated α -CD **1** to access to 2^A , 3^B , $6^{C(F)}$ -trihydroxy-per-*O*-methylated α -CD **6** as one isomer by this methodology.

Based on our previous studies on the de-*O*-alkylation of per-*O*-alkylated CDs,^{13,} ^{15-17, 19} we found that the reaction conditions, including the amount of DIBAL-H, the reaction temperature and the reaction time, have important effects on the selectivity of de-*O*-alkylation reaction. In addition, an amount of 6.0~40.0 equiv of DIBAL-H will helpful to the selectivity of tri-de-*O*-methyl reaction. After careful modification of the reaction conditions, we found that when per-*O*-methylated α -CD **1** was treated with 25.0 equivalent of DIBAL-H in toluene (1.0 M) at room temperature for 4 h, 2^A,3^B,6^{C(F)}-trihydroxy-per-*O*-methylated α -CD **6** was separated out in 35% yield as a major product by a regular silica-gel column chromatography (Scheme 2), along with little amount of known mixtures **4**/**5** (~20%), and an unsymmetric hexa-de-*O*-methylation derivative (~4%) as identified by ESI-HRMS. Based on the NMR analysis of the mono-hydroxy **2** and di-hydroxy **6** was achieved easily through the ESI-HRMS, 1D and 2D NMR spectra. The ESI-HR mass spectrum for **6** showed an

 $[M+Na]^+$ ion at m/z 1205.5422 Da (Calcd for C₅₁H₉₀NaO₃₀, 1205.5409), indicating that it is a tri-de-O-methyl derivative of 1. Due to random substitution of the methyl group in the narrom and wide rims of the molecule, the ¹H NMR spectrum of 6 is much complicated, except the lower-field and high field regions relatively well resolved. The signal at $\delta = 5.10-5.00$ ppm and 3.21-3.15 ppm, referring to 6H and 5H, respectively, were assigned to $6 \times H_1$ and $5 \times H_2$, and six carbons appearing at 99.65 (2C), 99.89, 99.93, 100.38 and 102.30 was assigned to C_1 (Figure 1). With the aid of ¹H-¹³C HSQC spectrum (Figure S1) and the Jmod spectra of compounds 2, 3 and 6 (Figure S2), some key signals such as H_2^A , H_3^B and $H_6^{C(F)}$ are unambiguously isolated. The triplet at $\delta = 4.12$ ppm (J = 9.4 Hz) and 3.59 ppm (overlap with others), each referring to 1H, was assigned to H_3^B and H_2^A , respectively, and two carbons appearing at 71.75 ppm and 73.47 ppm were assigned to C_3^B and C_2^A , respectively. The multiple signals at $\delta = 4.01$ -3.95 ppm, referring to 2H, was assigned to 2 × H₆^{C(F)}, and one carbon appearing at 62.37 ppm was assigned to $C_6^{C(F)}$. The structure of compound **6** was further confirmed using tandem mass spectrometry (MS/MS) techniques (Figures S3-S5). These data clearly shows the cleavage of mono-hydroxy monosaccharide (m/z 191.10), di-hydroxy disaccharide (m/z 381.20) and tri-hydroxy trisaccharide (m/z 571.30) fragments from compound 6, which indicate the C6-OH should be located at C or F glucose subunit. To our knowledge, the further determination of the position of C6-OH, which marked by an asterisk (Scheme 2), would be still difficult since there is no accumulation of spectral data for such systems.



Scheme 2. Reagents and conditions: (a) DIBAL-H (25.0 equiv), RT, 4 h, 35%; (b) Ac₂O, pyridine, DMAP, RT, 18 h, 90%. Asterisks indicated that the position of hydroxyl or oxyacetyl group locate at C or F glucose subunit.



Figure 1. 400 MHz Jmod spectrum (CDCl₃, 298K) of the tri-hydroxy **6**. Asterisk indicated that the position of hydroxyl locate at C or F glucose subunit.

The structure of tri-hydroxy 6 was further confirmed by its acetylated derivative 7. By comparing their spectral data of 7 with those of 6^A-O-acetyl-per-O-methylated α -CD 8 and 2^A,3^B-di-O-acetyl-per-O-methylated α -CD 9 (Supporting Information, S13-S14), which were prepared according to the procedure described by du Roizel ¹³ with minor modifications,²⁰ several deshielded signals were assigned conveniently. The low-field doublet of doublets at 4.69 ppm ($J_{2,3} = 10.4$ Hz, $J_{1,2} = 2.8$ Hz) and at 5.45 ppm ($J_{2,3} = 10.4$ Hz, $J_{3,4} = 9.2$ Hz), each referring to 1H, which, according to their ¹H-¹H coupling constants and their ¹H-¹³C correlation spectra, could be assigned to H₂^A and H₃^B, respectively. From the ¹H-¹³C correlation spectrum, two carbons appearing at 74.35 and 71.67 ppm, could be assigned to C_2^A and C_3^B , respectively (Figure S6). The broad doublet at $\delta = 4.58$ ppm ($J_{6a,6b} = 12.4$ Hz) and doublet of doublets at 4.36 ppm ($J_{6a,6b} = 12.4$ Hz, $J_{5,6a} = 3.2$ Hz), each referring to 1H, was assigned to $2 \times H_6^{C(F)}$, and one carbon appearing at 63.53 ppm was assigned to $C_6^{C(F)}$, due to the acetylation of the hydroxyl group. Clearly, the high-field signal at 2.21, 2.19 and 2.08 ppm, each referring to 3H, was assigned to three acetyl groups, and six carbons appearing at 170.54, 170.42, 170.37 and 21.68, 21.44, 20.89 ppm could be assigned to three carbonyl and methyl groups of acetates, respectively. The NMR spectra of 7 further confirms the presence of a free C2-OH and a freee C3-OH which located at two adjacent sugars and a free C6-OH in tri-hydroxy 6.

Combined with our previous studies,^{13, 19} the DIBAL-H promoted regioselective

de-*O*-methylation of **1** may include the following steps (Scheme 3): the first step would be a mono-de-*O*-methylation at C₆ of one sugar and bis-de-*O*-methylation at C₂ and C₃ of two adjacent sugar units to afford the mono-hydroxy **2** and the di-hydroxy **3**, respectively.¹³ The second step would be a bis-de-*O*-methylation at C₂ and C₃ of two adjacent sugar units of the mono-hydroxy **2** or a mono-de-*O*-methylation at C₆ of the di-hydroxy **3** to afford the tri-hydroxy **6**; or a duplication of bis-de-*O*-methylation at C₂ and C₃ at other two adjacent sugar units of di-hydroxy **3** to afford the tri-hydroxy **6**; or a duplication of bis-de-*O*-methylation at C₂ and C₃ at other two adjacent sugar units of di-hydroxy **3** to afford the tetra-hydroxys **4** and **5**. Theoritically, further careful modification of the reaction conditions by using a higher DIBAL-H concentration and/or a prolonged period of reaction time should allow to hexa-de-*O*-methylation of **1** to access to $2^A, 3^B, 2^C, 3^D, 2^E, 3^F$ -hexahydroxy-per-*O*-methylated α -CD **10**. However, in spite of our efforts, we were unable to get it.



Scheme 3. Proposed pathway for the formation of 2^A , 3^B , $6^{C(F)}$ -tri-hydroxy **6** from per-*O*-methylated α -CD **1**. Asterisk indicated that the position of hydroxyl locate at C or

F glucose subunit.

As already observed in our previous studies,^{13, 16, 17, 19} no simultaneous de-*O*methylation on both primary and secondary faces at the same molecule of per-*O*methylated α - and β -CD has been isolated and characeterized. It is generally believed that once mono-de-*O*-methylation on the primay face or bis-de-*O*-methylation on the secondary face happened, no further de-*O*-methylation on the other face takes place.¹³ It is the first time for us to isolate tri-de-*O*-methylation on both faces of per-*O*methylated α -CD **1** as a signle isomer in only one chemical operation using DIBAL-H as a chemical "scalpel". Statistical calculations indicate that for tri-de-*O*methylation of per-*O*-methylated α -CD **1** 109 regioisomers are possible, of which, we obtain only one single isomer as a major product, indicating the remarkable regioselectivity of DIBAL-H promoted tri-de-*O*-methylation of per-*O*-methylated α -CD **1**. As the nucleophilicity/reactivity of the three hydroxys are different, which provide a potential method for unsymmetrical introduction of three functional groups into per-*O*-methylated α -CD derivatives directly.

Experimantal section

General procedures

Optical rotations were measured at 20 ± 2 °C with a Perkin Elmer Model 343 digital polarimeter, using a 10 cm, 1 mL cell. High Resolution Mass Spectrometry (HRMS) were obtained with an APEX IV FT-MS (7.0 T) spectrometer (Bruker). The collision-activated dissociation (CID) tandem mass spectrometry (MS/MS) were obtained with Waters Synapt G2-Si spectrometer. NMR spectra were recorded on a Bruker DRX 400 spectrometer at ambient temperature. ¹H NMR chemical shifts are referenced to internal standard tetramethylsilane (TMS) ($\delta_{\rm H} = 0.00$ ppm). ¹³C NMR chemical shifts are referenced to the solvent signal ($\delta_{\rm C} = 77.00$ for the central line of CDCl₃). Reactions were monitored by thin-layer chromatography (TLC) on a precoated silica gel 60 F₂₅₄ plate (layer thickness 0.2 mm; E. Merck, Darmstadt, Germany) and detected by charring with a 10% solution of sulphuric acid in ethanol. Flash column chromatography was performed on silica gel 60 (200-300 mesh, Qingdao Haiyang Chemical Co. Ltd).

2^{A} , 3^{B} , $6^{C(F)}$ -trihydroxyl per-*O*-methylated α -CD (6)

To a solution of dried hexakis (2,3,6-tri-O-methyl)-α-CD 1 (271 mg, 0.22 mmol) was added 5.5 mL (5.5 mmol, 25.0 equiv) of DIBAL-H (1.0 M in toluene) at room temperature under nitrogen. The mixture was stirred for 4 h. Aqueous HCl (1.0 M) was carefully added dropwise to quench the reaction and the mixture was stirred vigorously at room temperature for 30 min. The toluene phase was separated and the water phase was extracted by ethyl acetate $(3 \times 20 \text{ mL})$. The combined organic phase was washed with brine, dried with MgSO4, filtrated and the solvent was removed in vacuo. The residue was subjected to flash chromatography (eluent: CH₂Cl₂/CH₃OH = 50:1-30:1) to give tri-hydroxy 6 as a white foam in 35% yield. $R_f = 0.16$ $(CH_2Cl_2/CH_3OH = 10:1); [\alpha]_D + 146 (c = 1.0 in CHCl_3); {}^{1}H NMR (400 MHz, CDCl_3):$ δ 3.15-3.21 (m, 5H, 5×H₂), 3.40 (br s, 9H, 3×OCH₃ (C₆)), 3.41 (s, 3H, OCH₃ (C₆)), 3.42 (s, 3H, OCH₃ (C₆)), 3.47 (s, 3H, OCH₃ (C₂)), 3.49 (s, 3H, OCH₃ (C₂)), 3.50 (s, 6H, 2×OCH₃ (C₂)), 3.51 (s, 3H, OCH₃ (C₂)), 3.60 (m, 1H, H₂^A), 3.61 (s, 3H, OCH₃ (C₃)), 3.63 (s, 6H, 2×OCH₃ (C₃)), 3.64 (s, 3H, OCH₃ (C₃)), 3.68 (m, 1H, H₄^B), 3.74 (s, 3H, OCH₃ (C₃)), 3.95-4.01 (m, 2H, 2×H_{6a}^{C(F)}), 3.47-4.01 (m, 27H, 5×H₃, 5×H₄, $6 \times H_5$, $5 \times H_{6a}$, $6 \times H_{6b}$), 4.12 (t, 1H, $J_{2,3} = J_{3,4} = 9.4$ Hz, H_3^B), 5.01 (d, 1H, $J_{1,2} = 2.8$ Hz, H_1^A), 5.03 (d, 1H, $J_{1,2} = 3.2$ Hz, H_1), 5.04-5.05 (m, 2H, 2×H₁), 5.09-5.10 (m, 2H, $2 \times H_1$); ¹³C NMR (100 MHz, CDCl₃): δ 57.66, 57.69, 57.78, 57.86, 58.10 (5C, 5×OCH₃(C₂)), 59.02, 59.05, 59.07, 59.11, 59.15 (5C, 5×OCH₃(C₆)), 61.68, 61.71, 61.82, 62.41 (5C, 5×OCH₃ (C₃)), 62.37 (C₆^{C(F)}), 70.38, 71.37, 72.09, 72.34 (6C, 6×C₅), 71.13, 71.49, 71.57 (5C, 5×C₆), 71.75 (C₃^B), 73.47 (C₂^A), 81.01, 81.20, 81.27, 81.28, 81.38, 81.95, 82.06, 82.32, 82.35, 82.42, 82.46, 82.96 (16C, 5×C₂, 5×C₃, 6×C₄), 99.65, 99.89, 99.93, 100.38 (5C, 5×C₁), 102.30 (C₁^A); ESI-HRMS (m/z) Calcd for C₅₁H₉₀O₃₀Na [M+Na]⁺: 1205.5415. Found 1205.5422.

2^{A} , 3^{B} , $6^{C(F)}$ -tri-*O*-acetyl per-*O*-methylated α -CD (7)

To a solution of **6** (25 mg, 0.021 mmol) in dry pyridine (2 mL) was added 2.6 mg of DMAP (0.021 mmol, 1.0 equiv) and 1 mL Ac₂O at room temperature. The reaction mixture was stirred for 18 h under nitrogen. The solvent was removed in vacuo. The residue was subjected to flash chromatography (eluent: CH_2Cl_2/CH_3OH

= 50:1) to give 35.1 mg (90%) of compound 7 as a white foam. $R_f = 0.37$ $(CH_2Cl_2/CH_3OH = 20:1); [\alpha]_D + 139 (c = 1.0 in CHCl_3); {}^{1}H NMR (400 MHz, CDCl_3):$ δ 2.08 (s, 3H, CH₃CO (C₆^{C(F)})), 2.19, 2.21 (2×s, 6H, 2×CH₃CO (C₂^A, C₃^B)), 3.13-3.19 (m, 4H, 4×H₂), 3.21 (dd, 1H, $J_{1,2}$ = 3.2 Hz, $J_{2,3}$ = 9.2 Hz, H_2^B), 3.38 (s, 3H, OCH₃ (C₆)), 3.39 (2×s, 9H, 3×OCH₃(C₆)), 3.40 (s, 3H, OCH₃(C₆)), 3.43 (s, 3H, OCH₃(C₂)), 3.46 (s, 3H, OCH₃ (C₂)), 3.47 (s, 3H, OCH₃ (C₂)), 3.51 (2×s, 6H, 2×OCH₃ (C₂)), 3.56 (s, 3H, OCH₃ (C₃)), 3.57 (s, 3H, OCH₃ (C₃)), 3.62 (s, 3H, OCH₃ (C₃)), 3.63 (m, 1H, H₃^A), 3.64 (s, 3H, OCH₃ (C₃)), 3.69 (s, 3H, OCH₃ (C₃)), 3.77 (m, 1H, H₄^B), 3.95 (m, 1H, H₅^{C(F)}), 3.43-4.07 (m, 24H, 4×H₃, 5×H₄, 5×H₅, 5×H_{6a}, 5×H_{6b}), 4.36 (dd, 1H, J_{5.6a} = 3.2 Hz, $J_{6a,6b}$ = 12.4 Hz, $H_{6a}^{C(F)}$), 4.58 (br d, 1H, $J_{6a,6b}$ = 12.4 Hz, $H_{6b}^{C(F)}$), 4.69 (dd, 1H, $J_{1,2} = 2.8$ Hz, $J_{2,3} = 10.4$ Hz, H_2^A), 4.98 (d, 1H, $J_{1,2} = 2.8$ Hz, H_1^A), 5.03-5.06 (m, 4H, $4 \times H_1$), 5.11 (d, 1H, $J_{1,2} = 3.2$ Hz, H_1^B), 5.45 (t, $J_{2,3} = 10.4$ Hz, $J_{3,4} = 9.2$ Hz, H_3^B); ¹³C NMR (100 MHz, CDCl₃): δ 20.89 (<u>C</u>H₃CO (C₆^{C(F)})), 21.43, 21.68 (2C, 2×<u>C</u>H₃CO (C2^A, C3^B)), 57.49, 57.82, 57.96, 58.01, 58.27 (5C, 5×OCH3 (C2)), 58.81, 58.94, 59.03, 59.07 (5C, 5×OCH₃ (C₆)), 61.41, 61.54, 61.74, 61.83, 62.00 (5C, 5×OCH₃ (C₃)), 63.53 (C₆^{C(F)}), 69.61 (C₅^{C(F)}), 70.65, 70.75, 71.33, 71.78 (5C, 5×C₆), 70.86, 71.17, 71.26, 71.40 (5C, 5×C₅), 71.67 (C₃^B), 74.35 (C₂^A), 78.43, 78.99, 79.86, 80.77, 80.93, 81.02, 82.08, 82.17, 82.23, 82.31, 82.52, 82.64, 82.68, 82.82, 82.89 (16C, 5×C₂, 5×C₃, 6×C₄), 98.42 (C₁^A), 99.68, 99.82, 99.89, 100.41, 100.48 (5C, 5×C₁), 170.37, 170.42 (2C, $2 \times CH_3 \underline{C}O$ (C_2^A , C_3^B)), 170.54 ($CH_3 \underline{C}O$ ($C_6^{C(F)}$)); ESI-HRMS (m/z) Calcd for C₅₇H₁₀₀NO₃₃ [M+NH₄]⁺: 1326.6172. Found 1326.6201. Calcd for C₅₇H₉₆O₃₃Na [M+Na]⁺: 1331.5726. Found 1331.5738. Calcd for C₅₇H₉₆KO₃₃ [M+K]⁺: 1347.5471. Found 1347.5517.

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Supplementary data

Supplementary data (copies of ¹H NMR, ¹³C NMR, HRMS spectra for compounds **6-9** and MS/MS spectra of compound **6**) associated with this article can be found, in the online version, at

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