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# **Baicalein-modified hydroxyapatite nanoparticles and coatings with antibacterial and antioxidant properties**

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## **Abstract**

Aseptic loosening and bacterial infections are the two main causes of failure for metallic implants used for joint replacement. A coating that is both bioactive and possesses antimicrobial properties may address such shortcomings and improve the performance of the implant. We have sought to study the properties of combining hydroxyapatite-based nanoparticles or coatings with baicalein, a plant-extracted molecule with both antibacterial and antioxidant properties. (B-type) carbonated hydroxyapatite nanoparticles prepared by a chemical wet method could subsequently adsorb by soaking in a baicalein solution. The amount of adsorbed baicalein was determined to be  $63 \text{ mg}\cdot\text{g}^{-1}$  by thermogravimetric measurements. In a second approach, baicalein was adsorbed on a biomimetic calcium-deficient hydroxyapatite planar coating ( $12 \text{ }\mu\text{m}$  thick) deposited on Ti6Al4V alloy from an aqueous solution of calcium, phosphate, sodium and magnesium salts. Soaking of the hydroxyapatite coated on titanium alloy in a baicalein solution induced partial dissolution/remodeling of the upper surface of the coating. However, the observed remodeling of the surface was much more pronounced in the presence of a baicalein solution, compared to pure water. The presence of adsorbed baicalein on the HAp layer, although it could not be precisely quantified, was assessed by XPS and fluorescence analysis. Planar coatings exhibited significant antibacterial properties against *Staphylococcus epidermidis*. Baicalein-modified nanoparticles exhibited significant antioxidant properties. These results illustrate the potential of hydroxyapatite used as a carrier for natural biologically-active molecules and also discuss the challenges associated with their applications as antibacterial agents.

**Keywords:** Titanium alloy; Hydroxyapatite; baicalein; Antibacterial activity; Antioxidant properties

## 1. Introduction

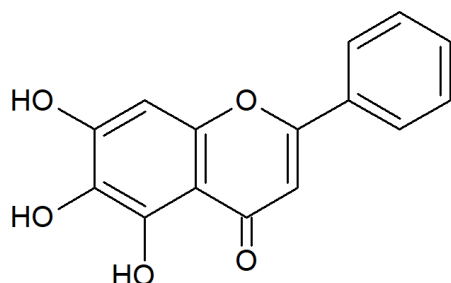
With ageing of population and recrudescence of arthrosic pathologies at early ages, the demand for biomaterials is constantly increasing, especially in the field of joint replacements (hip, knee). The requirements for physical properties do not leave many options in the choice of the material, only metals having acceptable stress and fatigue resistance. Due to the necessity of biocompatibility, titanium (Ti) and its alloys are widely used. However, several studies underline the increasing rate of revisions of arthroplasties over these last 20 years from implant failure [1], primarily caused by aseptic loosening and development of infections [2]. Aseptic loosening can occur on an implant with or without cement. The cement is used to fix the prosthesis in the implantation site, but the repetition of mechanical stress during locomotion degrades the bone-cement and cement-implant interfaces, leading to the disintegration of the cement and the production of loose pieces which can trigger an inflammatory response. In cementless arthroplasty, the loosening is related to the ability of osseointegration of the implant material into the bone.

Ti-based materials are currently the most commonly used to design the sections of implants that are in direct contact within bone [3]. However, they are not bioactive (i.e. do not favour bone growth) as such. Modifications of their surface by physical or chemical methods are possible to improve the osseointegration. Polishing techniques [4,5], or laser treatments [6] may be used to enhance the roughness and the hydrophilicity of the metallic substrate, that are known to induce better cell adhesion. Anodization of the surface to form titanium oxide nanotubes is also developed [7–9], as TiO<sub>2</sub> nanotubes enhance cell growth. Another approach is to coat the metallic surface with calcium phosphate material, due to its similarity with bone. In particular, the use of hydroxyapatite-based coatings is widespread [10,11], as they are close

in composition to the bone mineral phase and show high bioactivity [12]. Such coatings are now clinically used for cementless arthroplasty, with partially or fully coated femoral stem [13,14]. Furthermore, the possibility to incorporate active molecules or biological agents on or within hydroxyapatite coatings may provide antimicrobial activity to the surfaces of the implant [15–17]. This strategy enables higher concentrations to be achieved on the surfaces for antimicrobial activity. One possible strategy is to substitute  $\text{Ca}^{2+}$  with antibacterial metallic ions like  $\text{Ag}^+$  [18] or  $\text{Cu}^{2+}$  [19–21], but with the drawbacks of possible cytotoxicity associated with these metals. A recent approach is to synthesize  $\text{TiO}_2$  nanotubes at the surface of the titanium substrate, followed by the loading of the nanotubes by Ag-Si-Sr-substituted HAp nanoparticles, which enables an inhibition of bacterial activity while enhancing the biocompatibility [8]. Another possibility is to incorporate antibiotics directly in the hydroxyapatite coating, leading to significantly higher antibacterial effect of the material both *in vivo* and *in vitro* [15,22–26].

However, with increasing antibacterial resistance, described by the World Health Organization as “one of the biggest threats to global health, food security, and development today”, it is necessary either to find alternative antibacterial strategies or to identify drugs active against resistant strains [27,28]. In this context, natural antibiotics extracted from plants, in particular secondary metabolites such as flavonoids that are known to be involved in plant resistance mechanisms against microorganisms, appear as particularly promising [29,30]. For instance, baicalein (**Fig.1**), extracted from the root of the plant *Scutellaria baicalensis*, is known in the Chinese Pharmacopeia for centuries for its anti-inflammatory, antioxidant, antiviral, anticancer and antibacterial properties [31–34]. Its antibacterial activity was tested against sensitive *Staphylococcus aureus* and Methicillin-Resistant *S. aureus* (MRSA) [35,36]. Baicalein alone was able to eradicate a 7-days biofilm in a dose-dependent manner, for concentrations above its minimal inhibitory concentration (MIC) [35]. Used in

combination with ciprofloxacin, baicalein at concentration below MIC was able to restore the antibacterial activity of ciprofloxacin against MRSA. Additionally, a synergistic effect was observed against ciprofloxacin-resistant strains [36].



**Fig.1** Structure of baicalein (BE)

We present here the combination of baicalein and hydroxyapatite, with the goal of enhancing the osseointegration and the antibacterial properties of orthopaedic implants. The strategy was to adsorb baicalein to hydroxyapatite by soaking the material into a baicalein solution. Two hydroxyapatite shaping was used: nanoparticles that could be used to cover metallic substrates after dip or spin coating processes [37], and planar hydroxyapatite layer deposited on a titanium alloy coupon using a biomimetic method previously described in the literature [38]. The antioxidant and antibacterial activities of both modified nanoparticles and coatings after adsorption were studied. This study discusses the potential and limitations of the use of such natural drugs to confer additional biological functionalities to hydroxyapatite materials designed as metallic implants coatings.

## 2. Methods

### 2.1. Chemicals

Baicalein (BE) was purchased from TCI Europe (Zwijndrecht, Belgium) and stock solution at 0.1 M prepared in dimethylsulfoxide (DMSO) or methanol for ESI-MS analysis.

Calcium chloride dihydrate was obtained from Jessen Chemica. Sodium chloride and potassium chloride were purchased from Carlo Erba. NaHCO<sub>3</sub>, TRIS and sodium hydroxide were obtained from Sigma. Potassium phosphate monobasic, magnesium chloride hexahydrate and sodium sulfate are obtained from Prolabo. 10 mm\*10 mm Ti6Al4V titanium alloy plates with 1 mm thickness were obtained from Goodfellow Cambridge Limited (Huntingdon, UK). Before use, they were polished by #500, #800 and #1200 silicon carbide (SiC) disk, washed successively in acetone, ethanol and distilled water under ultrasound for 15 minutes, following by a hydroxylation in sodium hydroxide 5 M at 60 °C for 24 hours.

## 2.2. *Synthesis of hydroxyapatite nanoparticles*

Hydroxyapatite was synthesized by co-precipitation following the chemical wet method previously reported [39]. Briefly, 0.1 mol of Ca(OH)<sub>2</sub> (7.4 g) was dissolved in 100 mL of distilled water for 1.30 hour, and 0.06 mol (6.9 g) of NH<sub>4</sub>H<sub>2</sub>PO<sub>4</sub> was dissolved in 50 mL of water. Ammonium dihydrogen phosphate solution was added to calcium solution, and the medium was left to react for 48 h at 25 °C under stirring. Temperature was controlled by a thermostat bath. After reaction, white particles were recovered by centrifugation at 5000 rpm for 5 minutes, followed by three washes with distilled water. The powder was dried at 100 °C for 24 hours. For sorption studies, 10 mg of ground HAp particles were re-suspended in a 200 mL solution of BE in distilled water (pH ≈ 5). The BE concentration was set to 0.05 mM as it is near to the limit of solubility of BE in water, corresponding to an initial molar ratio between Ca and BE of 1. After 24 hours of contact under stirring at room temperature, particles were recovered by filtration, and dried at 100 °C for 24 hours (**Fig.2a**)

## 2.3 *Characterization of hydroxyapatite nanoparticles*

X-ray diffractograms (XRD) were recorded on a D8 Advance powder diffractometer (Bruker AXS, GmbH, Karlsruhe, Germany), equipped with a Cu source (CuK<sub>α</sub> radiation at 1.54 Å) and one-dimensional detector LynxEye in a Bragg-Brentano 2θ – 2θ geometry,

between 7 ° and 80 ° with a variable slit V12. For powder analysis, step was 0.03 °.s<sup>-1</sup> and scan speed 1 s.point<sup>-1</sup>. Scherrer equation [40] was employed to calculate the crystallite size of products (i.e. nano-HA) for each experiment:  $D = \frac{\lambda}{B \sin \theta}$ , where  $\lambda$  represents the wavelength of Cu K $\alpha$  i.e. 0.154 nm, B is the peak width at half-maximum intensity of (211) plane of HA in radians and  $\theta$  is the angle of the latter peak in degrees.

Thermogravimetric analysis (TGA) was performed with a SDT Q600 apparatus from TA Instruments, between 25 °C and 600 °C, with a ramp of 10 °C/min, under air. Sample weights were typically comprised between 10 and 20 mg. Surface specific area was measured with a physisorption apparatus (Belsorpmax), by measuring the adsorbed quantity of N<sub>2</sub> in function of pressure, at T = 77 K. Before measurement, vacuum degree was 3.202x10<sup>-6</sup> Pa. Sample weight was 0.9340 g. Transmission electron microscopy (TEM) images of samples were obtained with Tecnaï spirit G2 microscope with a Gatan Orius camera

Solid state NMR studies were performed on an Avance III Bruker spectrometer operating at 7.0 T (corresponding to 300.3, 121.6, 75.5 MHz for <sup>1</sup>H, <sup>31</sup>P and <sup>13</sup>C nuclei, respectively) equipped with a 4 mm Magic Angle Spinning (MAS) broadband dual probehead. Samples were inserted into zirconia rotors and spun at MAS = 12 kHz. <sup>1</sup>H experiments were acquired either with probe signal suppression (DEPTH pulse sequence)[41], or with rotor synchronised Hahn echoes [42] at 2x1 to 2x10 turns. 16 scans were recorded with a relaxation delay RD = 1 s, a 90° pulse and a Line Broadening LB = 10 Hz. For {<sup>1</sup>H}-<sup>13</sup>C experiments, Cross Polarization (CP) was used with a contact time t<sub>cp</sub> = 2 ms, 47752 scans and LB = 50 Hz. Before acquisition, the probe was cleaned with nitric acid at pH = 4.5 to avoid any residual <sup>13</sup>C pollution. Two-Dimensional (2D) {<sup>1</sup>H}-<sup>31</sup>P HETeronuclear CORrelation (HETCOR) maps [43] were obtained with a Hahn echo at 2x3 turns to allow T<sub>2</sub> transversal relaxation before CP with a contact time t<sub>cp</sub> = 1 or 10 ms, 256



scans, 180 slices, STATES-TPPI quadrature mode [44] and LB = 50 Hz in both dimensions. For both  $^{13}\text{C}$  and  $^{31}\text{P}$  CP experiments, a 95-105 % ramp was used during CP step and SPINAL64 decoupling [45] was applied during acquisition.  $^1\text{H}$  radio frequency was set at  $\nu_{1\text{H}} = 70$  kHz for  $^1\text{H}$  and  $\nu_{\text{X}} = 82$  kHz for the other nucleus.  $^1\text{H}$  and  $^{13}\text{C}$  chemical shift were referenced to tetramethylsilane, whereas  $^{31}\text{P}$  was referenced to 85% phosphoric acid.

Baicalein release from HAp particles in PBS was studied by HPLC. 10 mg of particles were suspended in 2 mL of PBS at pH 7.4, and 0.5 mL of supernatant was removed at different times for HPLC analysis, and replaced by fresh PBS. Baicalein release was studied over 6 days. HPLC analysis was performed with reverse phase on a C18 column.

#### *2.4 Synthesis of hydroxyapatite coatings on titanium alloys*

Hydroxyapatite was synthesized on titanium alloy substrates in Simulated Body Fluid (SBF) concentrated 1.5-fold (1.5-SBF) (Supplementary Material, S1 for precise composition of 1.5-SBF). Samples were vertically immersed in the 1.5-SBF solution, with volume/surface ratio  $V/S = 25$  mL.cm<sup>-2</sup>. Deposition was performed in high density polyethylene bottles (Thermo Scientific™ Nalgene™ Wide-Mouth HDPE Packaging Bottles). The solution was renewed every 48 h by freshly-prepared 1.5-SBF. Plates were withdrawn from the medium after 14 days, rinsed three times in distilled water and dried at room temperature for 24 hours. For baicalein adsorption, Ti alloy substrates coated with hydroxyapatite were then soaked in an aqueous solution of BE (0.1 mM) for 24 hours, with a volume  $V = 2$  mL for each sample (**Fig.2b**).

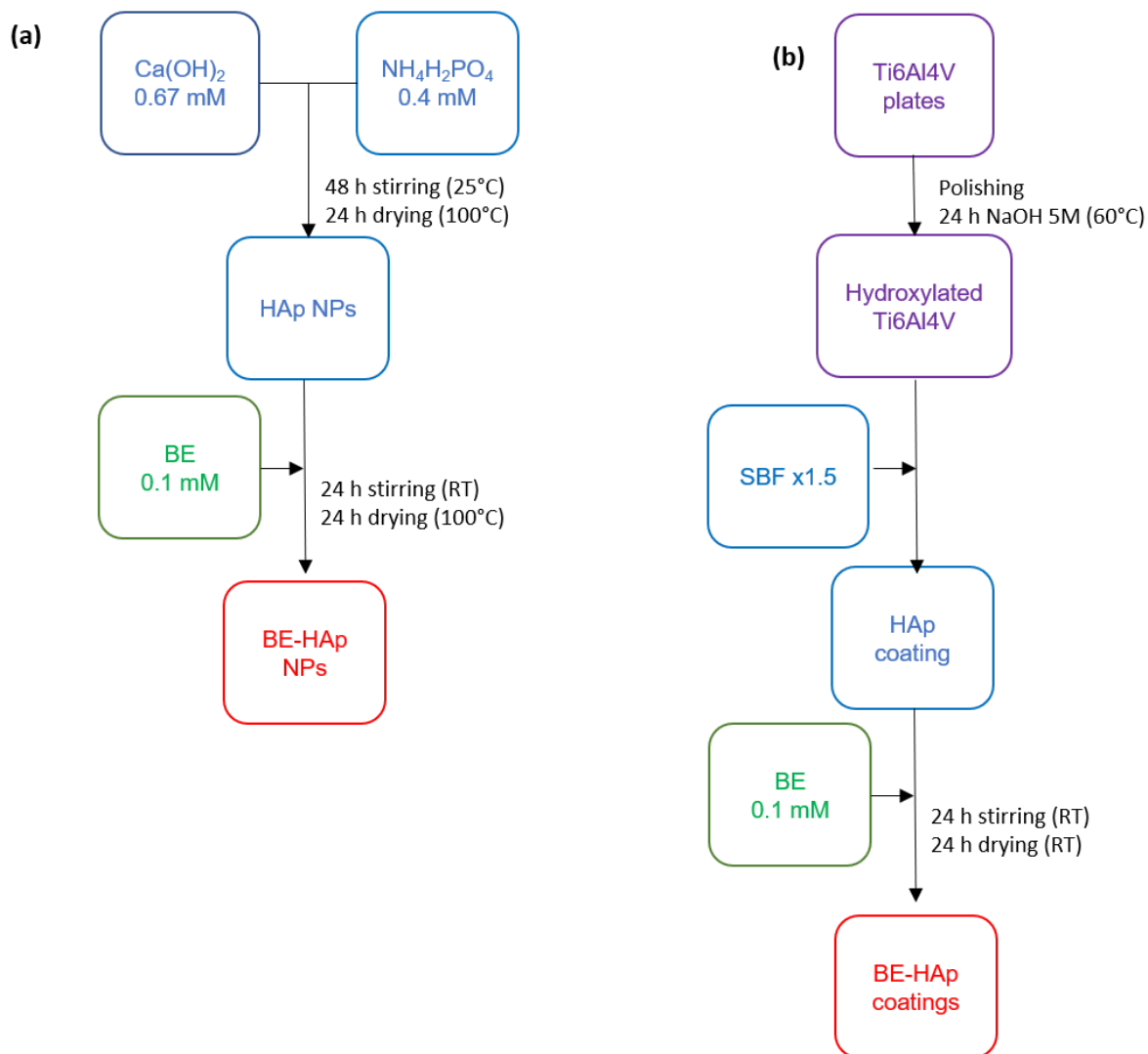
#### *2.5 Characterization of hydroxyapatite coatings*

XRD analysis of coatings was performed as previously described for nanoparticles except that the step was 0.05 °.s<sup>-1</sup> and scan speed was 0.1 s.point<sup>-1</sup>. The record was repeated 5 times to increase the signal to noise ratio. Raman spectra were collected with a Kaiser Optical

system, Raman Analyzer RXN1 microprobe equipped with a laser diode at 785 nm ( $P = 10$  mW, accumulation time: 10 s, 30 scans per spectrum), and a Leica microscope with x50 objective. Fluorescence spectra of coatings were obtained with a Fluorolog 3 apparatus from HORIBA Jobin Yvon. Excitation was set at  $\lambda_{\text{exc}} = 300$  nm with 5 nm of bandpass and emission spectrum was recorded between 320 and 650 nm with 1 nm of bandpass. Increment was set at 1.0 nm and integration time at 0.5 s. The surface was tilted from the incident beam (source Xenon, 450 W), and spectra were collected with an angle of  $22^\circ$ .

Morphology of coatings was observed by scanning electron microscopy on a Hitachi S-3400N microscope (acceleration voltage 10 kV, probe current 70). Samples were covered with carbon before imaging. Energy dispersive X-ray (EDX) spectroscopy was performed thanks to an Oxford instrument –X max (AZTEC software) and spectra were recorded at 4 different locations of the sample to check for homogeneity. Images of HAp coatings tilted with an angle of  $45^\circ$  were recorded on a SEM-FEG Hitachi SU-70 with an acceleration voltage at 1 kV. TEM experiments were performed with a Jeol JEM 1011 instrument (tungsten filament and 100 kV)

XPS spectra were collected on an Omicron Argus X-ray photoelectron spectrometer, using a monochromated Al  $K_\alpha$  ( $h\nu = 1486.6$  eV) radiation source having a 280 W electron beam power. The emission of photoelectrons from the sample was analyzed at a take-off angle of  $45^\circ$  under ultra-high vacuum conditions ( $1.10^{-7}$  Pa). XP spectra were collected at pass energy of 20 eV for  $C_{1s}$ ,  $O_{1s}$ ,  $Ti_{2p}$ ,  $Ca_{2p}$ ,  $P_{2p}$ , and  $Mg_{2s}$  core XPS levels. After data collection, the binding energies were calibrated with respect to the binding energy of the  $C_{1s}$  peak at 284.7 eV. The peak areas were determined after subtraction of a Shirley background. The atomic ratio calculations were performed after normalization using Scofield factors. Spectrum processing was carried out using the Casa XPS software package.



**Fig.2** Experimental flow chart for the preparation of baicalein (BE)-hydroxyapatite (HAp) (a) nanoparticles (NPs) and (b) coatings.

## 2.6 Antibacterial activity

The minimum inhibitory concentration (MIC) of BE against *Staphylococcus epidermidis* CIP 105.777 was determined using a two-fold serial dilution method in 96-wells plates. 100  $\mu\text{L}$  of a bacterial inoculum of  $1 \times 10^6$  CFU.mL<sup>-1</sup> was prepared from an overnight culture in sterile Mueller-Hilton broth (MH) and dispensed in each of the wells of a given row 96-well microplate. 100  $\mu\text{L}$  of a BE solution 1.35 mg/mL (prepared from a 10 mg.mL<sup>-1</sup> stock solution in DMSO) was added in the first well of the row. After mixing, 100  $\mu\text{L}$  of the mixture was transferred in the second well. The series of two-fold dilutions was performed

until the last well of the microwell plate. BE final concentrations in the wells ranged from 68 to 0.033  $\mu\text{g.mL}^{-1}$ . The highest final DMSO concentration used (5%) induced no significant bacterial growth inhibition. Growth of bacterial strains was visually assessed after 24 hours incubation at 37°C. MIC was the minimal concentration of the tested compound that completely inhibited cell growth. MIC determination of ciprofloxacin (0.0625  $\mu\text{g.mL}^{-1}$  against *S. epidermidis* CIP 105.777) was performed as a control on each microplate. The accepted variance on MIC value can be estimated to a 2-fold difference due to the microdilution method used. Experiments were performed in duplicate.

Antibacterial properties of the nanoparticles were tested against *S. epidermidis* CIP 105.777. Two concentrations of (HAp + adsorbed BE) NPs were tested, 5  $\text{mg.mL}^{-1}$  and 25  $\text{mg.mL}^{-1}$ . NPs were incubated with bacterial suspension at  $5 \times 10^5$   $\text{CFU.mL}^{-1}$  in MH culture broth for 16 hours at 37 °C on a rotative shaker (30 rpm). After 16 hours, suspensions were filtrated on a sterile filtration membrane (cellulose ester, 3  $\mu\text{m}$  porosity, Whatman Millipore). The membrane was then soaked in 2 mL of phosphate buffer saline (PBS) and sonicated during 5 minutes in order to resuspend bacteria. The obtained PBS solutions were then diluted to perform plate counting on solid MH-agar medium and living bacteria were enumerated (in Colony Forming Unit) after 24 hours of incubation at 37 °C.

Antibacterial properties of the coatings were determined by dipping the BE-HAp covered Ti alloy substrates in 2 mL of *S. epidermidis* CIP 105.777 suspension ( $5 \times 10^4$   $\text{CFU.mL}^{-1}$ ) in MH at 37°C. After 16 hours, samples were rinsed carefully with PBS to remove bacteria that are weakly adsorbed to the surface. Plates were then soaked in 2 mL of PBS, and the surface was scratched before 5 min of sonication and again afterwards, in order to recover adhered bacteria. The obtained PBS solutions were then diluted to perform plate counting on solid MH-agar medium and living bacteria were enumerated (in Colony Forming Unit) after 24 hours of incubation at 37 °C.

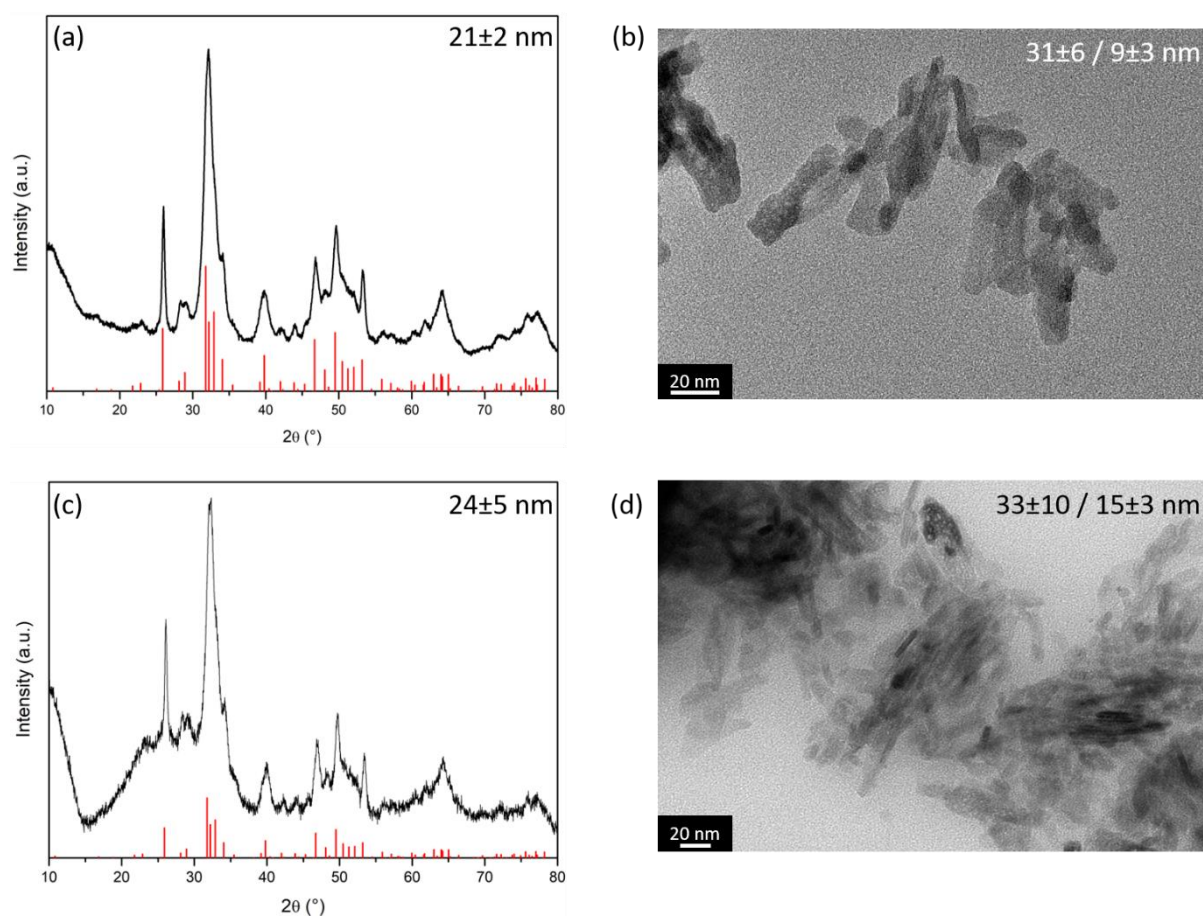
## 2.7 Antioxidant activity

Antioxidant potential of HAp NPs or coatings with adsorbed baicalein was tested using 2,2-diphenyl-1-picrylhydrazyle (DPPH) free radical scavenging method adapted from Blois [46,47]. For the NPs, the assay was performed as follows: 10 mg of NPs were suspended in 1 mL of a 1:1 (v/v) mixture of TRIS buffer at pH 7.4 and DPPH (100  $\mu$ M in ethanol). For coatings, each sample was immersed in 2 mL of the same mixture. Samples were incubated for 20 minutes in the dark and absorbance of the solution was recorded at 517 nm. DPPH absorbance reduction was calculated by the equation: %reduction =  $[1 - (A_{\text{sample}}/A_{\text{DPPH}})] \times 100$ , with,  $A_{\text{sample}}$  the absorbance after 20 min in the dark and  $A_{\text{DPPH}}$  the absorbance of DPPH solution before the addition of the solids.

### 3. Results

#### 3.1. Hydroxyapatite nanoparticles

Hydroxyapatite nanoparticles (HAp NP) were synthesized by a chemical wet method. The synthesis yield was *ca.* 60 % assuming that only stoichiometric hydroxyapatite ( $\text{Ca}_{10}(\text{PO}_4)_6(\text{OH})_2$ ) was formed. The X-ray diffractogram (**Fig.3a**) confirmed this hypothesis, by comparison to the PDF card 64-0738. TEM image as seen in **Fig.3b** showed plate-like nanoparticles  $31 \pm 6$  nm in length and  $9 \pm 3$  nm in width. Specific surface area calculated with the BET method was  $160 \text{ m}^2\cdot\text{g}^{-1}$  and the average pore size, corresponding to inter-particle distance, was 15 nm (Supplementary Material, S2).



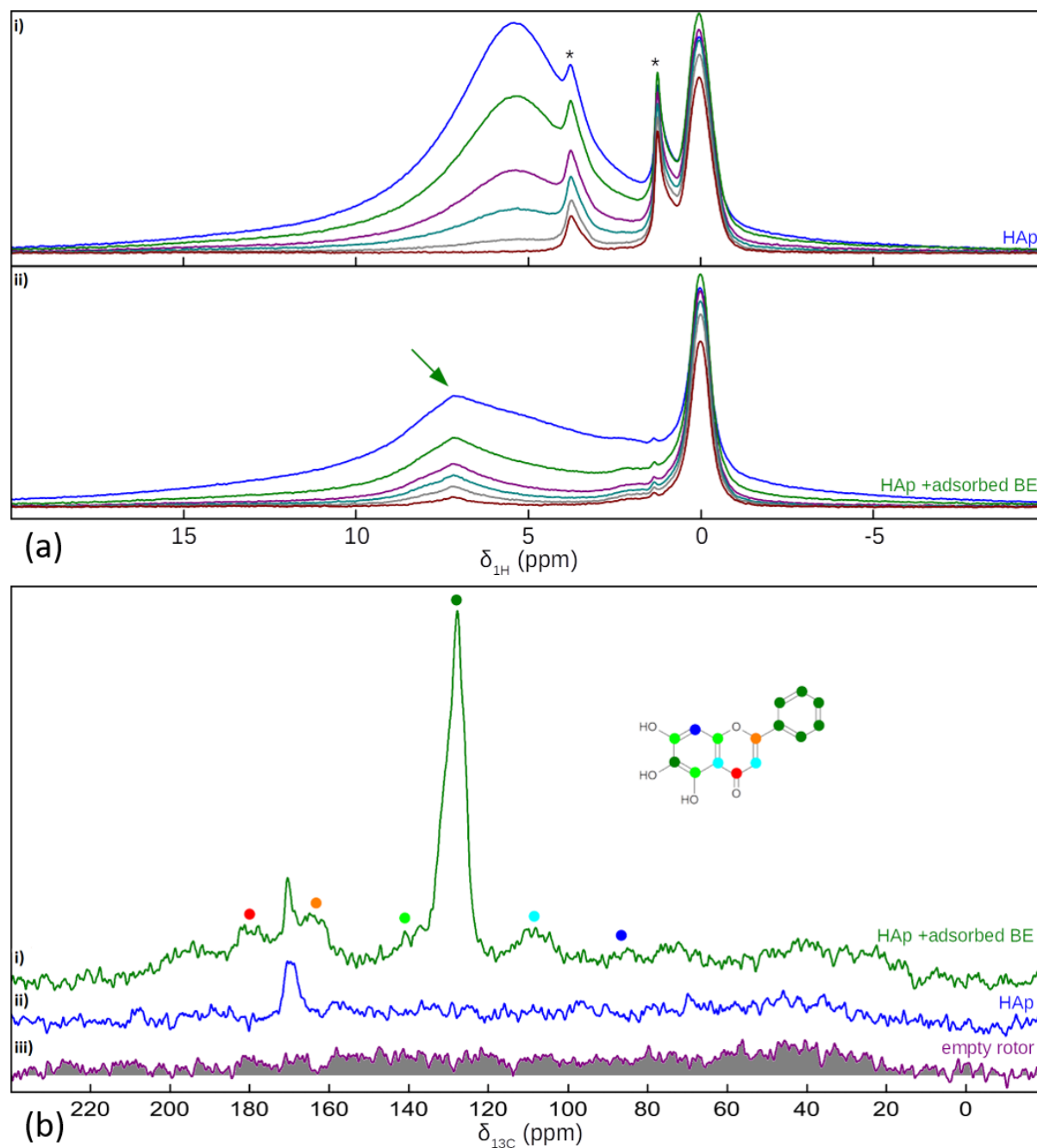
**Fig.3** X-ray diffractogram with crystallite size (a,c) and TEM image with average particle dimensions (length/width) (b,d) of hydroxyapatite nanoparticles before (a,b) and after (c,d) baicalein adsorption.

Baicalein adsorption on the HAp NP was performed over 24 h in distilled water, at the limit of solubility of BE (0.05 mM). The amount of baicalein on hydroxyapatite nanoparticles was measured by thermogravimetric analysis (TGA), leading to 63 mg.g<sup>-1</sup> of baicalein (Supplementary Material, **S3**). Particle dimensions as observed by TEM and crystallite size as estimated by XRD were not significantly modified by the adsorption process, but particles were more prone to aggregate as observed by TEM (**Fig.3b,d**).

For further characterization, <sup>1</sup>H solid-state NMR experiments were performed on hydroxyapatite nanoparticles before (**Fig.4a i**) and after adsorption of baicalein (**Fig.4a ii**). Both figures present a superimposition of <sup>1</sup>H spectra with probe background suppression (in blue) and with increasing Hahn echo delay (green to brown). This helps to discriminate species by filtering broad peaks having short T<sub>2</sub> transversal relaxation. On the spectrum of pure HAp NP, typical signals of protons were evidenced, with a narrow peak at 0 ppm characteristic of isolated protons of hydroxyl group in the crystal lattice, a broader peak at 5.5 ppm characteristic of adsorbed water and a tailing edge up to 17 ppm corresponding to disordered (di)hydrogenophosphates at the HAp surface [48]. After BE adsorption, an asymmetric peak at 7.2 ppm was observed, indicated by a green arrow on **Fig.4a ii**, corresponding to the aromatic protons of baicalein. Noticeably, water and ethanol peaks strongly decreased, highlighting surface polarity changes.

The <sup>1</sup>{<sup>1</sup>H}-<sup>13</sup>C CP spectrum of HAp NP (**Fig.4b ii**) was characterized by a broad peak of weak intensity at 170 ppm corresponding to both type A and B carbonates, *i.e.* substitution of hydroxide and phosphate ions, respectively [49]. Carbonates were also visible on the spectrum after adsorption of baicalein (**Fig.4b i**). However, the peak is narrower, with a more intense part above 170 ppm, which would suggest a type B substitution. In addition, an intense peak at 128 ppm is seen on the spectrum after adsorption of BE. This peak can be assigned to aromatic carbons of baicalein, and confirms the presence of baicalein on the HAp

NP. Smaller peaks were tentatively attributed to other carbons of the molecule, according to SDBS card 51630 of the parent baicalin molecule. It should be noted that a residual  $^{13}\text{C}$  probe signal ranging from 0 to 230 ppm was observed after 14 h of CP acquisition of an empty rotor (highlighted in grey on **Fig.4b iii**)).

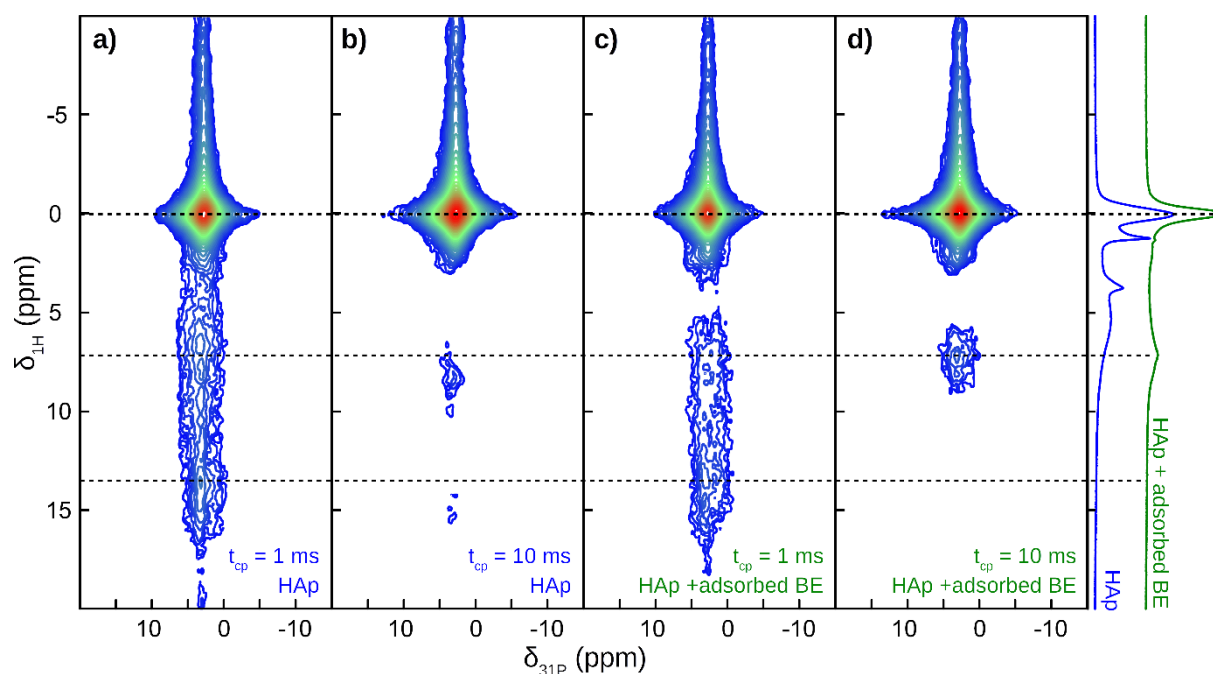


**Fig.4** (a)  $^1\text{H}$  solid-state NMR spectra of pure hydroxyapatite (i) and after adsorption of baicalein (initial concentration for adsorption = 0.05 mM) (ii). (\* indicates residual peaks of washing ethanol); (b)  $\{^1\text{H}\}$ - $^{13}\text{C}$  CP



solid-state NMR spectra of (i) hydroxyapatite after adsorption of baicalein (initial concentration for adsorption = 0.05 mM), (ii) pure hydroxyapatite, (iii) empty rotor (positive signal is colored in grey).

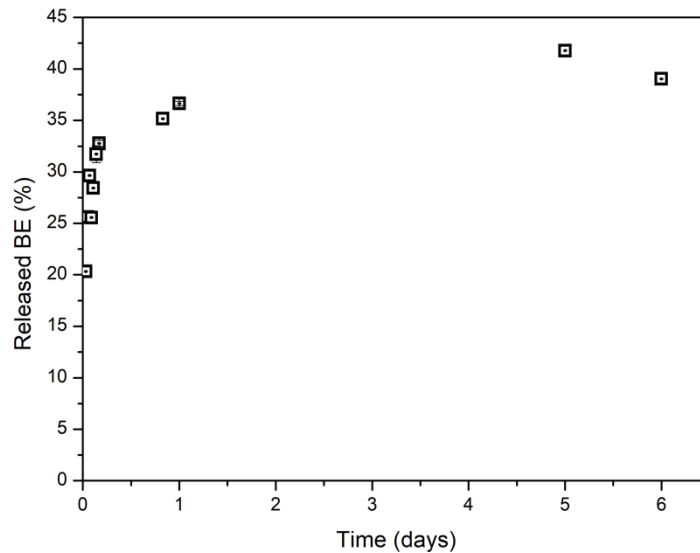
2D correlation maps between  $^1\text{H}$  and  $^{31}\text{P}$  of HAp NP were recorded. A contact time of 1 ms allowed for the observation of both crystalline HAp at 0 ppm for  $^1\text{H}$  and disordered (di)hydrogenophosphates at the HAp surface up to 17 ppm for  $^1\text{H}$  (**Fig.5a**). However, at 10 ms, the combined filtering of  $T_2$  relaxation during  $^1\text{H}$  Hahn echo and  $T_{1\rho}$  relaxation during CP almost completely suppressed the surface peak, keeping only a small residue at 8.3 ppm (**Fig.5b**). The same correlation maps were recorded after adsorption of BE. At a contact time of 1 ms, water correlation at 5.5 ppm decreased as compared to HAp NP (**Fig.5c**). At 10 ms a new correlation was visible at 7.1 ppm for  $^1\text{H}$  and 2.7 ppm for  $^{31}\text{P}$ , corresponding to an interaction between aromatic protons of BE and crystalline phosphates (**Fig.5d**). This is the sign of the vicinity of BE with HAp NP.



**Fig.5** 2D  $T_2$ -filtered hetero correlation map  $^1\text{H}$ - $^{31}\text{P}$  of HAp (a-b) and HAp-BE (c-d) with a contact time of 1 ms (a,c) and 10 ms (b,d). Horizontal dashed lines highlight peak positions.

Baicalein release from HAp NPs after adsorption was studied by HPLC. Cumulative release profile of baicalein from NPs, as percentage of the total adsorbed amount, is presented

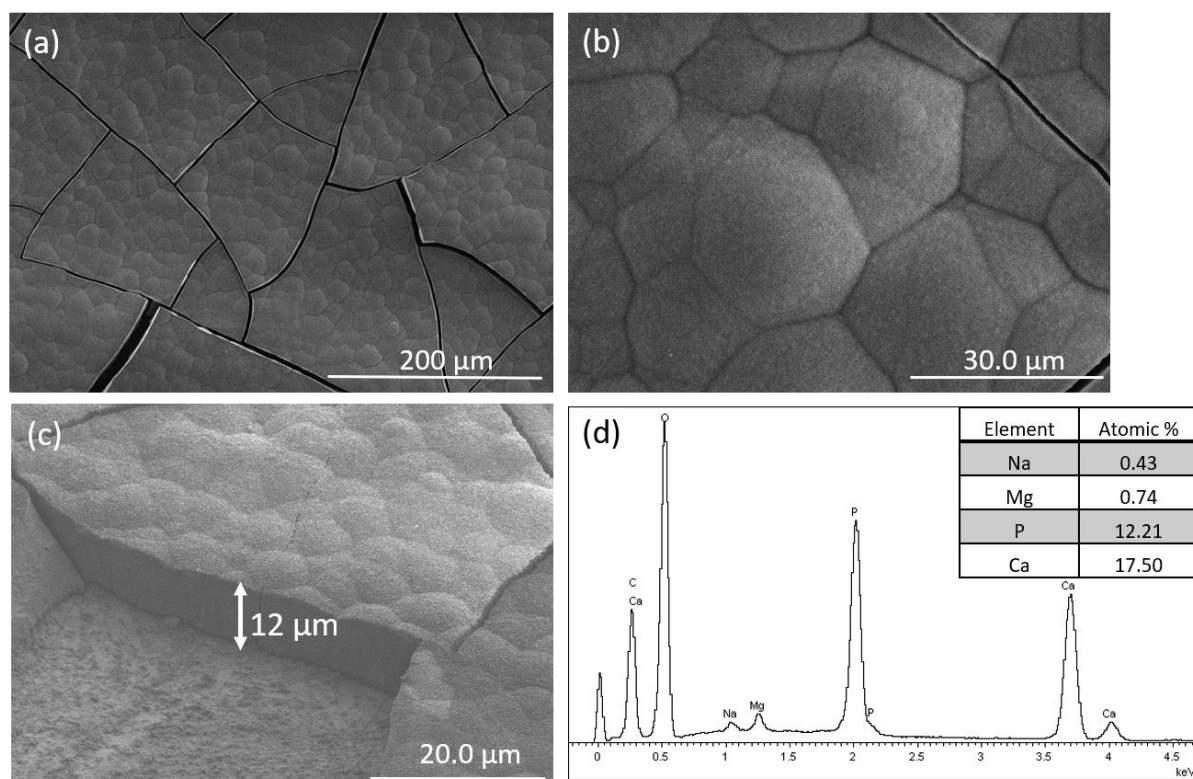
in **Fig.6**. Release profile is characterized by an initial burst, with 20 % of the total amount of adsorbed baicalein released in the first hour. Thereafter, release rate is slower during 24 hours and then a plateau is reached. After 6 days, 40 % of baicalein is released from the hydroxyapatite particles. This suggests that a noticeable amount of baicalein strongly binds to hydroxyapatite.



**Fig.6** Cumulative release of baicalein with time from nanoparticles.

### 3.2 Hydroxyapatite coatings

A hydroxyapatite coating was deposited on alkaline-treated Ti6Al4V alloy in 1.5-fold concentrated Simulated Body Fluid for 14 days [38,50]. The resulting coating can be described as a dense packing of spheroid-like particles, as observed by SEM on **Fig.7**. This morphology is typical of HAp coatings deposited from supersaturated solutions [50]. The thickness of the coating, measured after tilting of the sample, is about 12  $\mu\text{m}$  (**Fig.7c**).

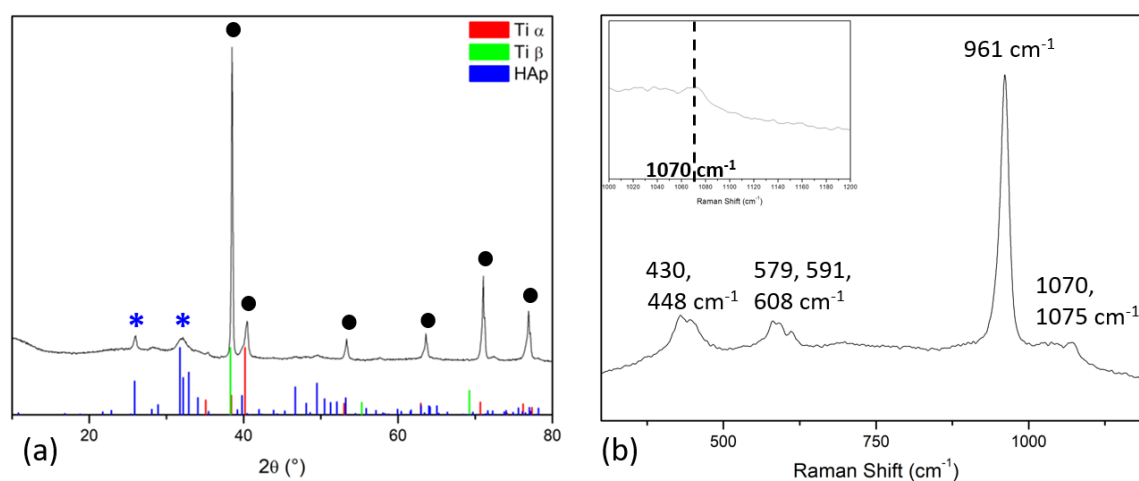


**Fig.7** SEM images of Ti6Al4V substrate after 14 days of deposition in 1.5-SBF with tilt = 0 ° (a,b) and 45 ° (c); (d) Corresponding EDX spectrum and elemental analysis.

The chemical composition of the HAp layer was assessed by EDX spectroscopy, evidencing the presence of Ca, P, Mg and Na (**Fig.7d**). The presence of Na and Mg peaks suggests that the HAp layer is substituted. Elemental content was homogeneous over the four analysed zones; the corresponding Ca/P ratio is  $1.44 \pm 0.02$  and the  $(Ca+Na+Mg)/P$  ratio is  $1.54 \pm 0.02$ . A stoichiometric hydroxyapatite has a typical Ca/P ratio of 1.67, and, in biological HAp, this ratio is closer to 1.5 due to cationic substitutions [51]. This suggests that the deposited HAp coating is a biomimetic calcium-deficient hydroxyapatite.

Coatings were also characterized by Raman analysis and XRD. As expected, based on the thickness of the HAp coating measured by SEM, the diffraction peaks representative of Ti  $\alpha$  and  $\beta$  forms (**Fig.8a**) were the most visible ones on the diffractogram of the coating. However, diffraction peaks representative of hydroxyapatite at 26° and 32° were clearly visible on the X-ray diffractogram. The presence of hydroxyapatite was also confirmed by

Raman analysis (**Fig.8b**), with the presence of the vibrations representative of phosphate in HAp:  $\nu_1$  at  $961\text{ cm}^{-1}$ ,  $\nu_2$  at  $430\text{ cm}^{-1}$ ,  $\nu_3$  at  $1075\text{ cm}^{-1}$  and  $\nu_4$  at  $591\text{ cm}^{-1}$  [52]. The weak shoulder at  $1070\text{ cm}^{-1}$  may indicate the presence of carbonates in the hydroxyapatite layer [53,54]. Several studies have correlated the full width at maximum half (FWMH) of  $\nu_1$  peak at  $961\text{ cm}^{-1}$  with the presence of carbonates in the hydroxyapatite structure: the more the hydroxyapatite is carbonated, the larger the peak is [55,56]. In our case, the FWHM is *ca.*  $16\text{ cm}^{-1}$ . Knowing that non-carbonated crystalline HAp particles have a FWHM below  $10\text{ cm}^{-1}$  [56], this result could be a clue of the presence of carbonated HAp as a coating. However, the broadening of the peak can also be related to the crystallinity of the material [54] and to the substitution of  $\text{Ca}^{2+}$  by  $\text{Mg}^{2+}$  [57], so that no conclusion can be unambiguously drawn about the coating carbonatation from Raman analysis.



**Fig.8** X-ray diffractogram (\*: HAp diffraction peaks, ●: substrate diffraction peaks) (a) and Raman spectrum (b) of Ti6Al4V substrate after 14 days of deposition in 1.5-SBF (inset: Raman spectrum between  $1000$  and  $1100\text{ cm}^{-1}$  showing the peak of  $\text{CO}_3^{2-}$  at  $1070\text{ cm}^{-1}$ )

HAp coatings before and after BE adsorption were also studied by XPS (**Fig.9a, b**). Oxygen, carbon, calcium, phosphorous, sodium and magnesium were detected on the XPS survey spectrum of bare HAp coating (**Fig.9a**). Energy and atomic percentages of elements found in the HAp coating are reported in **Table 1** and compared to literature data for

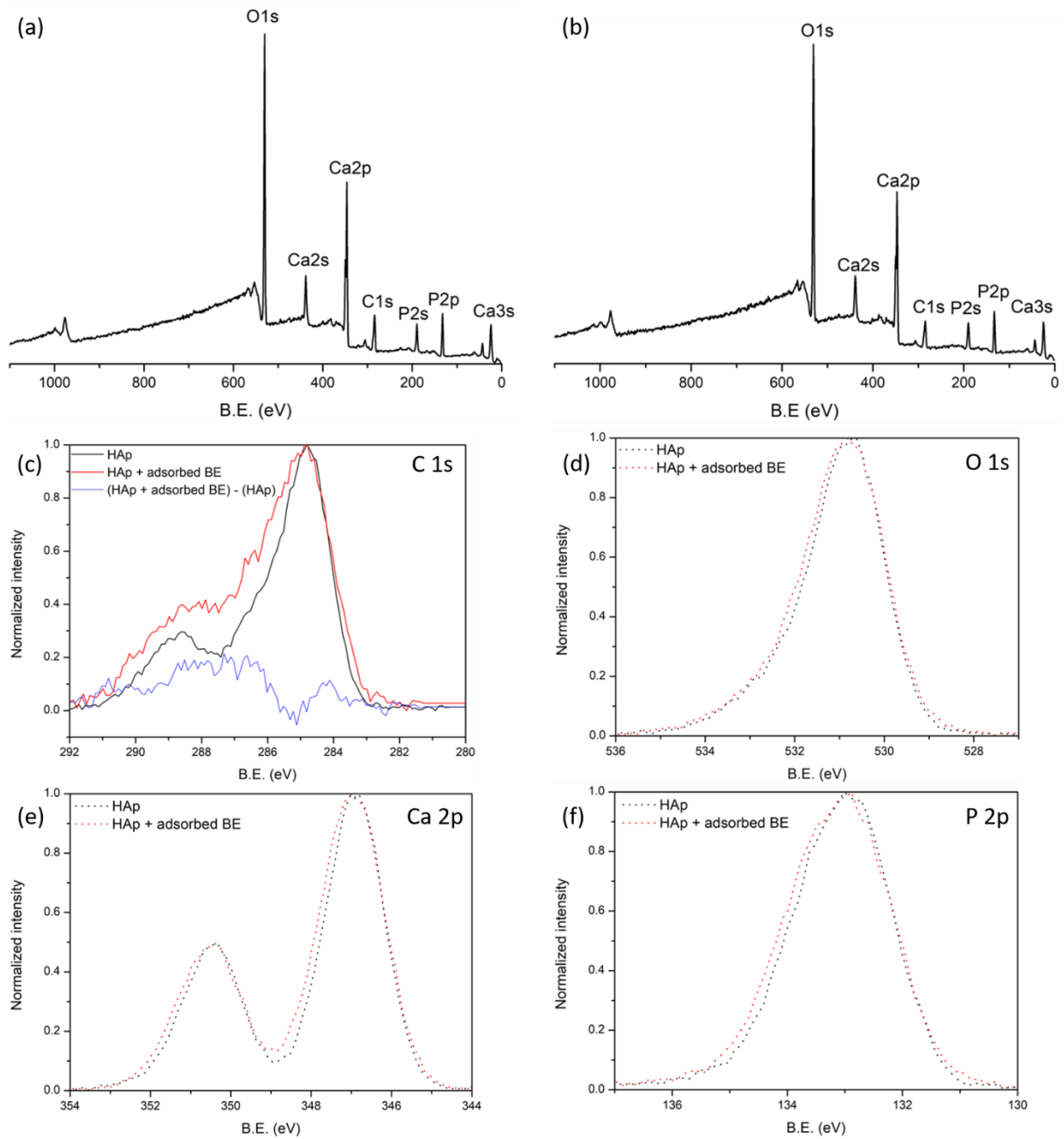
stoichiometric HAp (StHAp) [58]. Ca/P ratio in HAp coating was comprised between  $1.3 \pm 0.12$ , thus lower than that of stoichiometric hydroxyapatite (1.67), confirming that the layer is calcium-deficient. More, variable proportions of magnesium and sodium up to 0.4 % were found in the five samples analyzed in this work, thus being consistent with the possible substitution of calcium ions in the upper layer of hydroxyapatite coating.  $C_{1s}$ ,  $O_{1s}$ ,  $Ca_{2p}$  and  $P_{2p}$  peaks are presented in **Fig.9** c-f and their decomposition as Supplementary Material (**S4**).  $C_{1s}$  accounts for *ca.* 15 % of the total atomic percentage of the coating and results from organic adventitious contamination of the inorganic surface [59]. However, the  $C_{1s}$  decomposition reveals a contribution –accounting for 9.1 % of the total  $C_{1s}$  one- at 290.3 eV that can be assigned to metal carbonate [60], thus confirming the carbonation of the hydroxyapatite coating synthesized in 1.5-SBF.  $Ca_{2p}$  shows a characteristic doublet of Ca(II) at 352 eV and at 347 eV due to  $Ca_{2p_{1/2}}$  and  $Ca_{2p_{3/2}}$ , respectively.  $P_{2p}$  has closely spaced spin-orbit components, resulting in an asymmetric peak envelope. The decomposition clearly shows the  $P_{2p_{1/2}}$  and  $P_{2p_{3/2}}$  contributions at 134.2 eV and 133 eV, respectively. Binding energies for all peaks were similar to those reported for stoichiometric hydroxyapatite [58].

**Table 1.** Binding energy (eV), atomic percentages (At%) of carbon, oxygen, calcium, phosphorous, magnesium and sodium and calculated Ca/P ratio for bare hydroxyapatite coatings (HAp) and after baicalein sorption (HAp+BE) and comparison with stoichiometric hydroxyapatite (StHAp) [58].

	Sample	$C_{1s}$	$O_{1s}$	$Ca_{2p_{3/2}}$	$P_{2p}$	$Mg_{1s}$	$Na_{1s}$	Ca/P
<b>Binding energy (eV)</b>	HAp	284.9*	530.9	346.9	132.9	1304.2	1070.9	
	StHAp[58]	285.4	531.8	347.8	133.8	-	-	
	HAp	17±1	48.4±1.2	18.5±1.2	14.5±1.5	0-0.4	0-0.4	1.3±0.1
<b>At%</b>	HAp + BE	17±1	46.1±0.3	19.6±0.2	16.5±0.3	<i>b.d.</i>	<i>b.d.</i>	1.2±0.1
	StHAp[58]	-	59.1	22.7	13.6	-	-	1.67

\*carbon from adventitious organic contamination; *b.d.*: below detection limit

XPS analysis of the coating after baicalein adsorption showed unchanged element proportions compared to the one before adsorption (**Fig. 9b** and **Table 1**). Comparison of C<sub>1s</sub>, O<sub>1s</sub>, Ca<sub>2p</sub> and P<sub>2p</sub> signal detected in HAp coating before and after BE adsorption are presented in **Fig.9** c-f. Only the C<sub>1s</sub> spectrum showed a clear envelop modification after BE adsorption. As evidenced on the difference spectrum (**Fig.9c**), the contributions at 286.4 eV, corresponding to C-O-C and/or C-O-H bonds and the one at 288.5 eV, corresponding to C=O, are higher after adsorption of baicalein: 10.5 % to 23.6 % and 13.3 % to 16.8 %, respectively.



**Fig.9** XPS survey spectrum of HAp coating before (a) and after (b) adsorption of baicalein; comparison of XPS spectra of C<sub>1s</sub> (c), O<sub>1s</sub> (d), Ca<sub>2p</sub> (e) and P<sub>2p</sub> (f) of HAp coating before and after adsorption of baicalein.

This result could indicate the presence of BE in the HAp coating. However, it is worth noticing that a potential adsorption of BE on HAp cannot unambiguously be evidenced from XPS data since BE is composed of only C, O and H atoms and all these elements are already

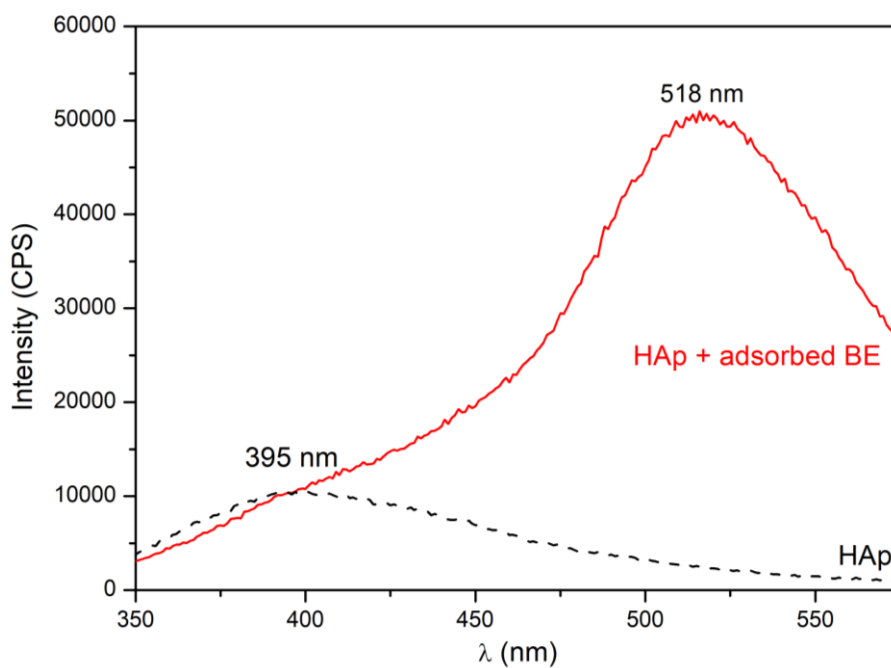
present on bare HAp XP spectrum because of the presence of adventitious contamination whose proportion is indeed uncontrolled.

Fluorescence spectra of the coating were also studied. Under excitation at 300 nm, HAp coating on titanium alloy exhibited an emission spectrum with a band with a maximum at 395 nm, as seen on **Fig.10**. The latter signal was also observed as a slight shoulder on the spectrum of HAp + BE. Because the HAp band gap ranges between 5.58 and 5.78 eV [61], this material is transparent for wavelengths higher than 222 nm. Consequently, the signal cannot be assigned to HAp. However, a broad photoluminescence spectrum has already been reported, with emitted light wavelength in the range of 400-450 nm as a result of defects in the crystal lattice and/or of the presence of some impurities as  $\text{CO}_2^-$  radicals resulting from the bond cleavages and trapped by the already formed hydroxyapatite lattice or interstitial positions [62,63].  $\text{CO}_3^{2-}$  substituting for OH groups and its decomposition could also contribute to the appearance of additional energy levels [61]. The observed emission peak at ~395 eV can also be due to the recombination of photogenerated electron-hole pairs as it corresponds to optical band gap of anatase titania (~3.2 eV) formed as a surface layer after titanium alloy basic treatment. In the presence of BE, an additional broad peak is evidenced on the HAp coated alloy with a maximum at 518 nm. As a comparison, the fluorescence spectra of baicalein in solution measured in DMSO at the same excitation wavelength exhibits a peak 540 nm (Supplementary Material, **S5**). It therefore seems reasonable to attribute the 518 nm peak of the HAP+BE coating to baicalein, the wavelength shift being attributed to the modification of the environment of the molecule on the hydroxyapatite surface compared to DMSO.

Attempts were made to determine the amount of adsorbed BE on the HAp coating by microthermogravimetry analysis. Measurements have been carried out in a microthermogravimetric apparatus (Hiden Isochema IGA-003) enabling the mass changes

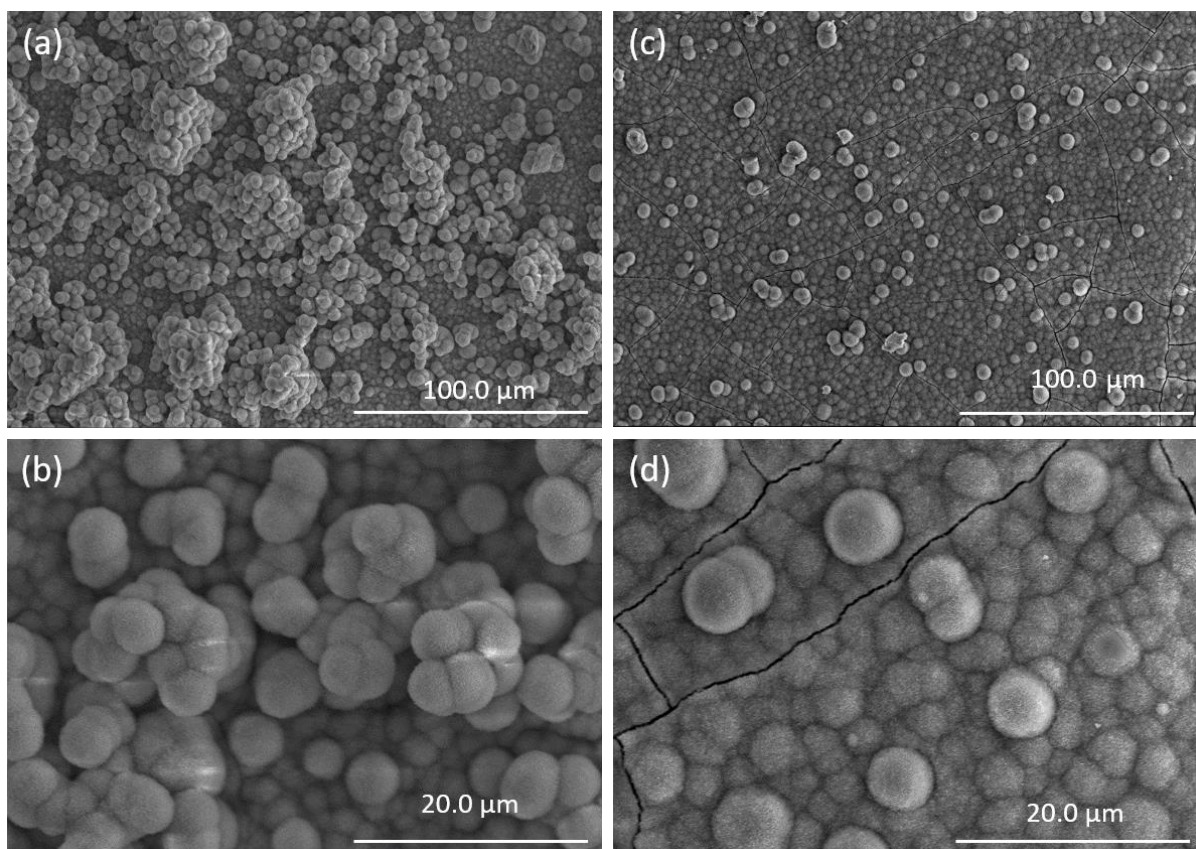


samples to be determined as a function of time with the accuracy lower than  $2 \cdot 10^{-7}$  g. Assuming that all the BE present in the soaking solution has been adsorbed, the HAp coating could contain up to  $5.5 \times 10^{-5}$  g of baicalein, that is higher than the TG device sensitivity. It could therefore be expected to detect some BE on the HAp. However, no significant variation in the mass loss curve could be detected when increasing the temperature of a HAp+BE sample from 20° C to 500 °C (Supplementary Material, **S6**). The low amount of adsorbed BE is also confirmed by the absence of peaks attributable to baicalein on the FTIR spectrum of the HAp+BE coating, only the bands at  $1031 \text{ cm}^{-1}$  and  $1062 \text{ cm}^{-1}$ , indicative of HAp formation, being observed before and after adsorption (Supplementary Material, **S7**).



**Fig.10** Fluorescence spectra of hydroxyapatite coatings before and after adsorption of baicalein at  $\lambda_{\text{ex}} = 300 \text{ nm}$

SEM images of hydroxyapatite coating after adsorption are presented in **Fig.11a**. Two different layers could be evidenced: a lower dense and homogeneous one that looks like flattened and joined spheroids and an upper one with a cauliflower structure non-uniformly covering the lower layer. This would suggest that part of the hydroxyapatite coating was dissolved and/or remodeled during the adsorption process. To check whether this modification of the upper surface morphology is linked to the presence of BE or to the immersion of the coating, a HAp coating was dipped in water for 24 hours. The resulting SEM images show a dense layer of spheroids (**Fig. 11b**), reminiscent of the layer before immersion, as seen on **Fig.6**, but less dense and homogeneous. Therefore, it seems that both water immersion and BE adsorption can impact on the surface morphology of the coatings, with a greater effect in the case of BE adsorption.



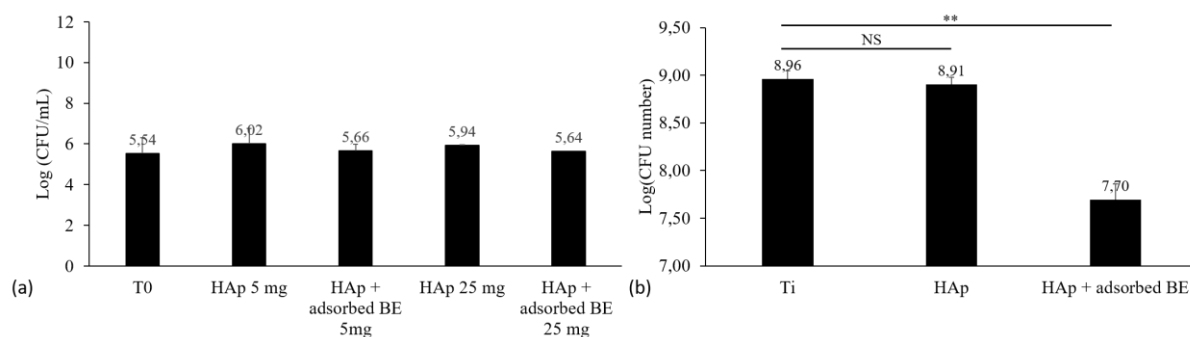
**Fig.11** SEM images of HAp coating after baicalein adsorption (a,b) and after immersion in water for 24 hours (c,d). Magnification: x500 (a,c) and x2.5 k (b,d)

### 3.3 Antibacterial and antioxidant activities

#### 3.3.1. Antibacterial activity

Antibacterial activity of baicalein in solution was tested against *S. epidermidis* CIP 105.777, a strain being capable of forming biofilms. The minimum inhibitory concentration (MIC) of baicalein was  $34 \mu\text{g.mL}^{-1}$ , which corresponds to  $126 \mu\text{mol.L}^{-1}$ .

The antibacterial activity of HAp NPs was tested in liquid medium against *S. epidermidis* CIP 105.777 in MH culture broth for 16 hours at  $37^\circ\text{C}$  (**Fig.12a**). Two concentrations of HAp NPs were tested:  $5 \text{ mg.mL}^{-1}$  and  $25 \text{ mg.mL}^{-1}$ . The number of living bacteria in contact with particles counted after 24 hours was identical (within the experimental error) for NP and NP+BE.



**Fig.12.** (a) Antibacterial activity of pure HAp nanoparticles and particles after adsorption of baicalein against *S. epidermidis* CIP 105.777 (T0 is the bacterial density at the beginning of the experiment); (b) Antibacterial activity of pure HAp coatings and coatings after adsorption of baicalein against *S. epidermidis* CIP 105.777 (NS: non-significant, \*\*:  $p \leq 0.01$ )

The antibacterial activity of hydroxyapatite coating was tested against *S. epidermidis* CIP 105.777, MH culture broth for 16 hours at 37 °C. The number of bacteria alive after 16 hours incubation is shown in **Fig. 12b**. In the presence of bare Ti substrate and HAp coated Ti, these numbers were identical, *i.e.*  $9.1 \times 10^8$  CFU.mL<sup>-1</sup>. However, the bacterial growth was reduced by one order of magnitude on the HAp + BE surface compared to HAp coating with  $5.0 \times 10^7$  CFU.mL<sup>-1</sup>. This decrease of bacterial growth is significant, with a *p* value calculated with Student's t-test of 0.008.

The comparison of the results obtained with NPs and coating probably highlights the effect of different surface properties. Indeed, while bacteria are expected to have only local interactions with nanoscale particles, they can firmly adhere on the extended surface of the coatings, all the more as SEM shows that they exhibit a significant roughness. Such a tight interaction could favour the antibacterial effect of adsorbed BE.

### 3.3.2 Antioxidant activity

Due to its phenolic structure, baicalein develops antioxidant activity that was studied by Blois method [46,47]. Antioxidant activity of HAp NP and HAp-coated Ti was expressed as a percentage of the DPPH absorption reduction, after 20 min of reaction in the dark. As a

comparison, the activity of a 0.1 mM solution of BE in TRIS buffer was also tested, and results are gathered in **Table 2**. The antioxidant activity of 0.1 mM baicalein in solution is 36 % at pH 7.4. Bare HAp NPs did not exhibit any antioxidant activity. (BE + HAp) NPs exhibited 70 % of antioxidant activity. After 1 and 5 days in suspension, the activity was *ca.* 50 %, which can be related to the concomitant release of *ca.* 40 % of BE over this period (**Fig.6**). In contrast, HAp-coated Ti with adsorbed baicalein had a non-zero but very small one (2 %). Such a low antioxidant activity is however in line with what could be expected on the basis of the maximal amount of adsorbed BE, *i.e.*  $2.10^{-7}$  moles.

**Table 2.** Antioxidant activity expressed as DPPH bleaching percentage of baicalein (0.1 mM in Tris buffer), hydroxyapatite nanoparticles (10 mg) and coatings before and after adsorption of baicalein.

Sample		DPPH absorbance reduction (%)
0.1 mM baicalein		36
HAp alone	Nanoparticles	0
	Coatings	0
HAp + BE	Nanoparticles	$70 \pm 11$
	Coatings	$2 \pm 1$

## 4. Discussion

### 4.1. Adsorption of baicalein on hydroxyapatite nanoparticles and coatings

Baicalein was successfully adsorbed on hydroxyapatite nanoparticles to an extent of  $63 \text{ mg.g}^{-1}$ . The presence of baicalein was confirmed by  $^1\text{H}$ ,  $^{13}\text{C}$  and  $^{31}\text{P}$  solid state NMR spectroscopy. Adsorption of baicalein did not modify the morphology nor crystallinity of HAp NPs, as observed by TEM and XRD. In case of coating of Ti alloy plates with HAp synthesized in 1.5-SBF, the presence of adsorbed BE was evidenced by fluorescence measurements and, to some extent, by a modification of the XPS  $\text{C}_{1s}$  peak. Moreover, the fact that significant antioxidant activity is observed for (BE+HAp)-coated plates and not for HAp-

coated ones support baicalein adsorption. However, the amount of adsorbed BE was too low to be quantified by thermogravimetry, *i.e.* below 1.4 mg per g of HAp on the basis of the detection limit of the used thermobalance.

Three kinds of interactions are likely to take place between adsorbed BE and the hydroxyapatite surface: (1) electrostatic, (2) hydrophilic (hydrogen bond)/hydrophobic and (3) complexation with the calcium ions [64–66]. The first two are highly correlated to the surface charge of HAp: at pH below its isoelectric point (IEP), the surface is positively-charged and has good affinity for anionic species, while the mirror situation occurs at pH above IEP. Near the IEP, the HAp surface has a marked hydrophobic character. The IEP of hydroxyapatite can range between 7 and 8 [67], being therefore positively-charged in the sorption conditions in this work (pH 5), whereas baicalein is neutral at this pH. Here the significant affinity of BE for the hydroxyapatite surface can be interpreted in light of a study by Chirdon *et al.*[68] showing that catechol and pyrogallol adsorbed readily and strongly on HAp surfaces. The proposed explanation is based on cooperative hydrogen-bond interactions thanks to the presence of two or three close phenol groups. It is well-documented that organic molecules, such as catechols, that have strong affinity for mineral surfaces can accelerate their dissolution [69,70]. Thus, it can be suggested that BE is able to complex calcium on the HAp surface and dissolve it to release Ca-BE species. This would increase the relative amount of P, in agreement with the XPS data. The reason why this would occur for coatings and not for nanoparticles may be related to the difference in crystallinity of the samples.

#### *4.2. Antioxidant and antibacterial activities*

Antioxidant properties of baicalein, as characterized by the DPPH radical scavenging assay, was 36 % at a concentration of 0.1 mM in Tris buffer. After adsorption on HAp nanoparticles, the molecule kept its activity: HAp NP with adsorbed BE had an antioxidant

activity of 70 %. It is important to point out that, in the conditions of the DPPH test with 10 mg of HAp+BE nanoparticles in 1 mL, baicalein concentration is *ca.* 2 mM. This value is higher than that tested for baicalein in solution (0.1 mM), which can contribute to the highest antioxidant activity. As a matter of fact, such a 2 mM concentration is larger than the solubility limit of BE in water, evidencing the benefit of its association with the HAp nanoparticles. Recently, hydroxyapatite nanoparticles modified with catechin, another flavonoid, also demonstrated higher antioxidant activity for the surface-immobilized molecules compared to the soluble form, attributed to the high surface density of the active molecules [71]. In contrast to nanoparticles, the activity of here-obtained coatings after adsorption was very low. This result may be explained by an adsorbed quantity of baicalein on coatings that is not sufficient to exhibit a measurable antioxidant activity against DPPH radical.

As far as the antibacterial activity is concerned, (HAp NP + BE) were not bactericidal against *S. epidermidis* CIP 105.777. This contrasts with other drug-release hydroxyapatite nanoparticles reported in the literature [72–76]. For instance, porous hydroxyapatite nanoparticles loaded with tetracycline, a broad-spectrum antibiotic, was shown to inhibit *Escherichia coli* growth in 2 hours, and remained active for several days [74]. Antibacterial activity was tested following the disk diffusion method, with particles shaped as pellets. This antimicrobial testing method was also used to evaluate the activity of hydroxyapatite nanoparticles loaded with vancomycin, against laboratory strains of *S. aureus* and *E. coli*, and MRSA [73]. Particles were active for 7 days against MRSA, and for more than 20 days against the other strains. In the first example, a drug loading similar to our system was achieved, while in the latter one, much higher amounts of antibiotics ( $160 \text{ mg.g}^{-1}$ ) were immobilized. Thus, the difference of activity between these systems and ours should be strongly linked to the intrinsic antibacterial activity of the used molecules. These studies

described hydroxyapatite loading with classic broad-spectrum antibiotic (vancomycin and tetracyclin), with MIC much below that of baicalein (MIC < 1  $\mu\text{g.mL}^{-1}$  compared to 34  $\mu\text{g.mL}^{-1}$  for BE).

Quite surprisingly, a small but significant antibacterial activity was measured for hydroxyapatite coatings with adsorbed baicalein against *S. epidermidis* CIP 105.777 in rich medium, despite the low drug loading. The bacterial growth was reduced by one order of magnitude in presence of baicalein compared to titanium alloy substrate or pure hydroxyapatite coatings. Noticeably, preliminary cytotoxicity assays using fibroblasts did not evidence any harmful effect for baicalein solution of concentrations up to 50  $\mu\text{g.mL}^{-1}$  (Supplementary Material, **S8**). Such a concentration is very unlikely to be reached *in vivo* considering that BE loading on coatings is maximum 25  $\mu\text{g}$  per  $\text{cm}^2$  of coating. This suggests that the presence of BE should not impact the biocompatibility of the implant. Again, this activity is lower than the activity of other calcium phosphate coatings incorporating antibiotics reported in the literature. For example hydroxyapatite coatings with adsorbed tobramycin or cephalothin are effective against *S. aureus* in 16 h [15]. Tobramycin was also adsorbed on phosphate calcium coating on fixation pins, with a sustained antibacterial activity for 7 days, despite an initial burst release in 15 min [23]. However, the comparison of antibacterial activity results was not obvious, because the method of testing, in solid medium on agar plates, was different from the one we used (test in solution). Yet, the major difference between these coatings and ours was again the antibacterial activity of the antibiotics alone.

The question is raised why, despite a lower BE loading, coatings showed some antibacterial activity whereas nanoparticles did not. One plausible reason is that the area of contact between the bacteria and the modified surface is larger on the flat coating compared to the curved nanoparticles, which should be more favorable towards biofilm-forming bacteria.



## 5. Conclusions

With the aim of developing new coatings for metallic implants that would combine bioactivity and antibacterial properties, we have associated hydroxyapatite, in a nanoparticulate and a coating form, with baicalein, a natural antibacterial compound. Nanoparticles have the advantages of easy preparation and high antibiotic loading but showed no antibacterial activity in the conditions of this work. Nevertheless, it is possible to take advantage of another property of baicalein, its antioxidant activity, to design nanoparticles with higher efficiency than the molecule alone. As a comparison, hydroxyapatite coatings exhibiting a biomimetic composition, *i.e.* calcium-deficient carbonated hydroxyapatite, showed a significant although low antibacterial activity, despite a low amount of adsorbed baicalein (below 1 wt%).

Overall, the antibacterial performances of the material studied in this work are below that of related systems in the literature using current synthetic antibiotics. One of the main reasons is that such synthetic antibiotics are at least 100 times more active (*i.e.* have MIC two orders of magnitude smaller) than natural antibacterial molecules on susceptible strains. This issue is more problematic here as the concentration of the solution that can be used for baicalein adsorption is limited by the poor solubility of BE in water (0.1 mM). Nevertheless, the choice of active molecules from traditional medicine remains a promising solution to address current concern raised by the rapidly developing phenomenon of antibiotic resistance. Improvement of baicalein loading may be possible if porous hydroxyapatite nanoparticles or coatings are used. Moreover, in traditional medicine, plant extracts containing several antibacterial molecules acting synergistically are used instead of single molecules. Thus, associating more complex natural formulations, such as essential oils, with hydroxyapatite could be a highly relevant strategy for the future.

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## **CRedit author statement**

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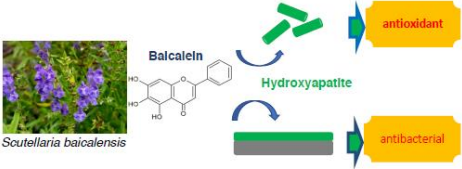


## Declaration of interests

The authors declare that they have no known competing financial interests or personal relationships that could have appeared to influence the work reported in this paper.

The authors declare the following financial interests/personal relationships which may be considered as potential competing interests:

Graphical abstract



## **Highlights**

-Hydroxyapatite and baicalein were associated to design biologically-active coatings

-Baicalein-Hydroxyapatite coatings on Ti6AlV4 exhibit antibacterial properties

-Baicalein- Hydroxyapatite nanoparticles are efficient antioxidant agents

-Benefits and limitations of baicalein to functionalize hydroxyapatite are discussed