

One hundred years later, resurrection of Tydemania gardineri A. Gepp & E. Gepp (Udoteaceae, Chlorophyta) based on molecular and morphological data

L. Lagourgue, H. Verbruggen, E.E. Ampou, C.E. Payri

▶ To cite this version:

L. Lagourgue, H. Verbruggen, E.E. Ampou, C.E. Payri. One hundred years later, resurrection of Tydemania gardineri A. Gepp & E. Gepp (Udoteaceae, Chlorophyta) based on molecular and morphological data. European Journal of Phycology, 2020, 55 (1), pp.89-99. 10.1080/09670262.2019.1654618 . hal-02967139

HAL Id: hal-02967139 https://hal.sorbonne-universite.fr/hal-02967139v1

Submitted on 14 Oct 2020 $\,$

HAL is a multi-disciplinary open access archive for the deposit and dissemination of scientific research documents, whether they are published or not. The documents may come from teaching and research institutions in France or abroad, or from public or private research centers. L'archive ouverte pluridisciplinaire **HAL**, est destinée au dépôt et à la diffusion de documents scientifiques de niveau recherche, publiés ou non, émanant des établissements d'enseignement et de recherche français ou étrangers, des laboratoires publics ou privés.

1	ONE HUNDRED YEARS LATER, RESURRECTION OF TYDEMANIA
2	GARDINERI A. GEPP & E. GEPP (UDOTEACEAE, CHLOROPHYTA) BASED
3	ON MOLECULAR AND MORPHOLOGICAL DATA
4	
5	Lagourgue, L. ^{1,2} , Verbruggen, H. ³ , Ampou, E.E. ⁴ , Payri, C.E. ²
6	
7	¹ Sorbonne Universités, UPMC Univ Paris 06, IFD, 4 Place Jussieu, 75252 Paris Cedex
8	05, France
9	² UMR ENTROPIE (IRD, UR, CNRS), Institut de Recherche pour le Développement,
10	B.P. A5 Nouméa Cedex, Nouvelle-Calédonie, 98848, France.
11	³ School of BioSciences, University of Melbourne, Parkville 3010, Australia
12	⁴ Institute for Marine Research and Observation, SEACORM/INDESO Center

13 Corresponding author's email address: laura.lagourgue@ird.fr

14 ABSTRACT

15 *Tydemania* Weber-van Bosse is a genus belonging to the Udoteaceae family (Bryopsidales, Chlorophyta) and currently thought to be monospecific throughout its 16 17 distribution range in the Indo-Pacific. We tested the assumption that *Tydemania* is a single species using species delimitation methods, morphological observations, and 18 phylogenetic reconstructions of large datasets. Our molecular and morphological data 19 recovered two distinct groups, which we argue are the type species T. expeditionis 20 21 Weber-van Bosse and T. gardineri A. Gepp & E. Gepp. The latter is currently 22 considered a synonym of the former and we resurrect this name from synonymy. 23 Tydemania gardineri is distinguished morphologically from T. expeditionis by the 24 systematic absence of glomeruli, whereas T. expeditionis can have glomeruli and 25 flabella, glomeruli alone, or flabella alone. The two species can also be distinguished by 26 the shape and length of the stalks at the basis of the flabella, the diameter of the main 27 axis, and of the flabella siphons at the apices. In addition, they have different geographical distributions, with T. gardineri restricted to the Western Indian Ocean, and 28 29 T. expeditionis extending from the Red sea, throughout the Indian Ocean and into the West-Pacific, including the coral triangle, the Philippines and Japan. From our dataset 30 and literature search, we hypothesize its south-western distribution limit is in northern 31 32 Madagascar. Finally, we confirm the synonymization of T. mabahithae with T. expeditionis based on specimens from the type-locality of the former ranging within the 33 T. expeditionis species and we do retrieved thalli without flabella in several other 34 specimens of T. expeditionis. 35

Keywords: macroalgae; Bryopsidales; species delimitation; phylogeny; molecular data;
morphology; Indo-Pacific.

38 INTRODUCTION

Tydemania Weber-van Bosse is a genus of calcified siphonous green algae in the family 39 Udoteaceae (Bryopsidales, Chlorophyta). The type species, Tydemania expeditionis Weber-40 41 van Bosse, was described from various places in Indonesia (Kabala Dua (Makassar Strait), Saleh bay (Sumbawa Island), de Bril, and near Fau Island) following the Siboga expedition, a 42 Dutch scientific campaign dedicated to zoological and hydrographic research in Indonesia, led 43 by Max Carl Wilhem Weber in 1899-1900 (Weber-van Bosse, 1901). The species was 44 described as composed of a cylindrical monosiphonous creeping or ascending axis, either 45 branched or unbranched. Siphonous side branches of the axis form either fans (flabella) or 46 47 dense ball-like structures (glomureli). The siphons branch out dichotomously in alternate planes to form the glomeruli, or in a single plane to form the flabella. Gepp & Gepp (1911) 48 described a second species, Tydemania gardineri A. Gepp & E. Gepp, from specimens 49 collected by the Bristish zoologist and oceanographer J. Stanley Gardiner in the central Indian 50 Ocean (Chagos archipelago and Amirante Islands) during the Sealark expedition. Tydemania 51 52 gardineri was distinguished by having only flabella and lacking glomeruli (Gepp & Gepp, 1911). Nasr (1939) proposed a third species, *Tydemania mabahithae* Nasr, to accommodate 53 specimens from the Red Sea. This species only has glomeruli and no flabella. Further studies, 54 55 based on morphological observations, assumed that the presence of either flabella or glomureli alone may be explained by environmental conditions or life-stages, and that 56 differences in siphon diameters were not significant (Gilmartin, 1966, Farghaly, 1980), and 57 Meinesz (1981) proposed the merger of T. gardineri and T. mabahithae with T. expeditionis. 58 This taxonomic decision was never confirmed by molecular data, but since then, all 59 60 *Tydemania* records have been assigned to *T. expeditionis* (Guiry & Guiry 2018). The rich collection of *Tydemania* samples housed at the herbarium of Noumea (NOU) 61

62 and at Ghent University (GENT) offered an opportunity to revisit the genus. The collection

consists of 38 specimens, collected by the authors from 13 sites during several field 63 expeditions in the Red Sea and the Indo-Pacific region, and are representative of the three 64 morphologies, including specimens with either flabella or glomeruli only, and specimens with 65 both. In addition, 42 digitalized specimens of Tydemania spp from L were examined to 66 support the study with reference collections. 67 The goal of this study was to take advantage of this large dataset, and to evaluate the 68 morphology-based synonymization of T. gardineri and T. mabahithae to T. expeditionis using 69 DNA analyses. Our approach consisted of: 1) applying species delimitation methods to 70 chloroplast and nuclear markers to evaluate species boundaries; 2) analysing whether and how 71 72 morpho-anatomical observations align with suggested species boundaries; and 3) reconstructing a multilocus phylogeny (tufA, rbcL and 18S rDNA) to assess the relationships 73 of the different samples within the genus Tydemania. 74

75

76 MATERIAL AND METHODS

77 Specimen sampling:

A total of 38 newly collected specimens of *Tydemania* were included in this study. The 78 samples were collected by SCUBA between 2005 and 2017 from many locations in the Indian 79 80 Ocean (Madagascar, Mayotte, Banc du Geyser, Tanzania), the Pacific Ocean (Indonesia (Bunaken); Papua New Guinea (Kavieng & Madang); the Solomon Islands; Vanuatu; New 81 Caledonia (Grande Terre and Loyalty islands); Fiji), and the Red Sea. Most specimens were 82 photographed in situ prior to sampling, labeled and subsampled in both 95% ethanol and silica 83 gel for later DNA analyses. Exceptions were specimens from the Solomon Islands, Vanuatu, 84 85 and Fiji for which DNA extractions were done on herbarium material. Specimens were then pressed as herbarium vouchers and are currently housed at NOU and GENT. Herbarium 86 abbreviations follow Thiers (2016). 87

88

89

90 Molecular analyses:

DNA was extracted using a cetyltrimethylammonium bromide (CTAB; Doyle & Doyle, 1987) 91 based method with modifications (available on request). Sequences from three markers 92 93 (chloroplast: *tuf*A and *rbc*L; nuclear: 18S rDNA) were obtained using primers previously 94 published for the Udoteaceae family (Händeler et al., 2010; Kooistra, 2002; Lam and Zechman, 2006; Verbruggen et al., 2009) (Supplementary information, Table 1). PCR 95 96 amplifications were conducted in a volume of 25 µl containing 12.5 µl AmpliTaq Gold 360 Master Mix (Applied Biosystems), 0.75 µl dimethylsulfoxyde (DMSO), 1 µl bovine serum 97 albumin (BSA), 1 µl of each F and R primers (10 µM), 2.5 µl DNA extract, and 6.25 µl ultra-98 pure water. Marker-specific PCR programs follow Lagourgue et al. (2018), and Sanger 99 100 sequencing was carried out by Genoscreen using 20 µl of PCR product (Lille, France). Sequences were quality-trimmed and aligned with Geneious version 7.1.9 101 (http://www.geneious.com, Kearse et al., 2012) and the MUSCLE algorithm (Edgar, 2004) in 102 Geneious, along with Tydemania expeditionis sequences downloaded from Genbank and 103 sequences for three outgroups: Udotea flabellum (J. Ellis & Solander) M. Howe, Udotea dotyi 104 105 D.S. Littler & Littler and Udotea geppiorum Yamada. All the specimens used in this study along with Genbank and BOLD accession numbers are listed in Supplementary Information 106 107 (Table 2).

108 Phylogenetic reconstructions:

109For the species delimitation analyses, gene trees were built using Maximum

110 Likelihood (ML) in RAXML (Stamatakis, 2014). This was done through the CIPRES portal

111 (Miller et al., 2010) using the GTR+G evolution model, the best scoring ML tree algorithm,

and 1,000 bootstrap iterations (Stamatakis et al., 2008). Data were partitioned by codon
position for the *tuf*A and *rbc*L datasets.

For phylogenetic reconstructions, the *tuf*A, *rbc*L and 18S rDNA alignments were 114 concatenated. PartitionFinder v.1.1.0 (Lanfear, et al., 2012) was used to identify the best 115 partitioning scheme for the dataset under the BIC criterion as follows: by gene and codon for 116 tufA (F81; HKY; GTR) and rbcL (JC; GTR+G; JC; F81; JC; HKY), and as one partition only 117 118 for the 18S rDNA (HKY+I). A Bayesian phylogenetic tree was obtained using MrBayes v.3.2. (Ronquist and Huelsenbeck, 2003) through the CIPRES portal, starting from a random 119 tree and with 10 million generations of MCMC. The convergence of the runs and ESS values 120 121 were checked with Tracer 1.5. (Rambaut and Drummond, 2007). The consensus topology and the posterior probabilities were computed after removing the first 10% generations as burn-in. 122 The ML tree was built using RAXML (Stamatakis, 2014) through the CIPRES portal. The 123 analysis was run on the dataset partitioned by gene and codon, under a GTR+G+I nucleotide 124 model, 1,000 rapid bootstrap replicates and search for best-scoring ML tree algorithm 125 126 (Stamatakis et al., 2008).

127 Species delimitation:

128 Species delimitation was assessed with the Automatic Barcode Gap Discovery method

129 (ABGD, Puillandre et al., 2012), and the Poisson Tree Process method (PTP, Zhang et al.,

130 2013). These two methods were chosen and combined for the complementarity of their

underlying assumptions: ABGD is a distance-based method and attempts to identify a barcode

132 gap between intra- and interspecific distances in the distribution of genetic distances between

samples; PTP is a tree-based method and relies on the number of substitutions along branches

to identify a shift between speciation and coalescence events.

ABGD was performed through the website <u>http://wwwabi.snv.jussieu.fr/public/abgd/</u>
 <u>abgdweb.html</u> for each gene. The analysis was run under the simple distance method with

137 default values for all parameters. PTP was run through the Exelixis Lab web server

138 (http://sco.h-its.org/exelixis/web/software/PTP/index.html) on ML rooted gene trees, for

- 139 100,000 generations, a thinning value of 100, and a burn in of 10%.
- 140

141 Morphological observations:

Morpho-anatomical observations were made on herbarium material, decalcified in HCl 142 143 solution (5%), using a binocular (WILD M3Z, Heerbrugg Switzerland) and a light microscope (Imager.A2, Zeiss, AXIO) equipped with a Canon EOS 100D digital camera. They followed 144 the descriptions and observations from previous studies (Gepp & Gepp 1911, Weber-van 145 146 Bosse 1901, and Meinesz 1981) and mainly focused on the external habit, the presence of glomeruli and flabella, the stalk (also called stipites by Gepp & Gepp 1911) and flabellum 147 shape, the type of attachment of the stalks to the main axis of either flabella or glomeruli (Fig. 148 1). Measurements of diameters were made for the main axis, and the stalk siphons for both 149 flabella and glomeruli. Morphoanatomical observations and measurements are detailed in 150 151 Supplementary Information (Table 3). We also made observations on the digitized herbarium specimens available online from L. 152

153

154 **RESULTS:**

155 Molecular results and species delimitation analyses

156 Among the 38 *Tydemania* specimens available for molecular analysis, 35 were successfully

- sequenced and a total of 72 sequences were obtained, including 32 *tuf*A sequences (841bp),
- 158 25 *rbc*L sequences (1306 bp), and 15 18S rDNA sequences (1243 bp), and an additional 4
- *tufA*, 2 *rbcL*, and one 18S rDNA sequences from Genbank were added. No sequences could
- 160 be obtained from the pressed herbarium sheets from Fiji, the Solomon Islands, and Vanuatu.

The ABGD and PTP species delimitation methods applied to the chloroplast and 161 162 nuclear markers led to similar results with samples splitting into two groups (i.e., putative species) (see supplementary information: Figure 1, Data 1 and 2). The first group was 163 composed of samples from the Red Sea (Egypt, Sudan), the Western Indian Ocean 164 (Madagascar, the Maldives), Indonesia (Bunaken), the Philippines (Siquijor and Balicasag 165 166 islands), Papua New Guinea (Madang), Japan (Kerama Islands, Okinawa) and New 167 Caledonia. This group also included Genbank sequences of *Tydemania expeditionis* from Japan, Guam, Philippines, and Maldives. The second putative species included samples from 168 Madagascar, Mayotte, Tanzania and the Banc du Geyser (EPI). 169

170

171 Morpho-anatomical observations

Photos of relevant morphological features are shown in Figures 2-24. The morphological 172 traits of specimens from the first group matched the original description of the species 173 Tydemania expeditionis (Weber-van Bosse 1901). We observed specimens with a creeping 174 monosiphonous axis from which both glomeruli and flabella were arising (Fig. 2), while other 175 specimens had glomeruli only (Fig. 3), or flabella only (Fig. 4). Nevertheless, common 176 morpho-anatomical features are shared among all these specimens. In situ, thalli are from 177 light to dark green with axes creeping over the substratum and, when present, glomeruli are 178 successively stacked in an upright pompon-like structure (Fig. 3). When present, flabella are 179 180 found below a series of glomeruli (Fig. 5), or arising alone along the main siphon. In both cases, the main siphon, ranging from 360 to 780 μ m in diameter (mean size = 518 μ m ± 100), 181 becomes monoliform (i.e. with more constrictions) near the secondary structures (glomeruli or 182 flabella) (Fig. 6). Flabella are generally large, monostromatic and composed of aligned and 183 dichotomously divided siphons (Fig. 7). Some dichotomies are anisomorphic (Fig. 8). The 184 constrictions above dichotomies show thickened cell walls. Both glomeruli and flabella are 185

supported by short and thick stalks, with few dichotomies (Fig. 9 and 10), which are attached in whorls to the main axis (Fig. 11 and 12). Glomeruli siphons range from 240 to 460 μ m at the bases, and 80 to 180 μ m (mean size= 110 μ m ± 22) at the apex. Flabella siphons range from 200 to 450 μ m at the base, and 60 to 130 (mean size= 79 μ m ± 13) at the margin. The main axis protrudes from the top of flabella or glomeruli, continuing the main growth axis of the plant (Fig. 13 and 14). A porous calcified sheath is present around the siphons.

192 The second group was characterized by specimens with only flabella arising from the main axis, while glomeruli were absent from all the specimens (Fig. 15 and 16). In situ, thalli 193 form dense tufts or patches of numerous flabella, greenish-grey in color. The size of the main 194 195 siphon ranges from 285 to 570 μ m (mean size= 426 μ m \pm 94). The flabella are supported by long stalks, with strongly marked constrictions giving a beaded necklace-like aspect (Fig. 17, 196 18 and 19). The stalks are highly dichotomously divided in alternating planes, leading to a 197 dense and stacked mass of siphons at the basis of the flabella (Fig. 20, 21 and 22). Flabella 198 can be of different sizes but generally small, monostromatic, and composed of dichotomously 199 200 dividing siphons arranged in one plane (Fig. 24). Siphons show thickened cell walls at the strongly constricted areas above dichotomies as well as along the constrictions seen in the 201 202 stalks (Fig. 23). The siphons of the flabella measure between 160 and 340 µm at the base, and 203 from 30 to 60 μ m (mean size= 44.5 μ m \pm 7) at the apices. The calcified sheath of the siphons is porous. These observations correspond to the diagnosis of *Tydemania gardineri* A. Gepp & 204 E. Gepp. The digitized specimens L.3958458 housed in Leiden and collected in the Amirante 205 Islands (Seychelles) by Coppejans et al. (2001) also matched the habit of the second group. 206

The siphon diameter data, acquired from a large collection of samples, highlighted that the main axis and the flabella siphons at the apices are thinner in *T. gardineri* than in *T. expeditionis*, a useful trait for diagnosis (Table 1). The general habit of the thalli is also useful for distinguishing between the two species, with the *T. gardineri* specimens lacking glomeruli and having numerous flabella arising from the main axis, and *T. expeditionis* having only a
few flabella if any. Therefore, specimens of the *T. gardineri* group appear like dense tufts or
patches of flabella (Fig. 15 and 16). In *T. gardineri*, the main axis is usually creeping along
the substrate rather than growing upright as in *T. expeditionis*. The form and the length of
stalks supporting the flabella are also distinctive, being long, monoliform (beaded aspect), and
highly dichotomously divided in *T. gardineri* (Fig. 17, 18 and 19), and shorter, stockier, and
less divided in *T. expeditionis*. Morphological differences are summarized in Table 1.

218

219 Phylogenetic reconstruction

The topologies of the phylogenetic trees were congruent between the markers (tufA, rbcL, and 220 18S rDNA) when analyzed individually or concatenated. The topology as well as nodes 221 support were similar for ML and Bayesian trees, although ML tree appeared somewhat better 222 223 resolved towards the terminal branches. We considered nodes with bootstrap values (bs) > 90and Posterior Probability (PP) > 0.95 as highly supported. Both ML and BI trees show a 224 well-supported dichotomy (bs: 100; PP: 1) separating the Tydemania specimens into two 225 fully-supported clades (bs: 100; PP: 1) (Fig. 25). The first clade grouped sequences of 226 specimens from the Indo-Pacific and morphologically identified as *Tydemania expeditionis*, 227 along with sequences downloaded from Genbank. Within this clade, sequences for two 228 specimens collected in New Caledonia grouped together in a fully supported subclade (bs: 99; 229 230 PP: 0.98). The second clade included only samples collected from the Western Indian Ocean region and morphologically identified as T. gardineri. This clade was not subdivided in any 231 meaningful way. 232

233

234 **DISCUSSION**

235 Molecular data and morphological observations support the resurrection of Tydemania 236 gardineri

237 Both the molecular and morpho-anatomical analyses recovered two distinct groups of 238 specimens among our Tydemania dataset. We assigned one to T. expeditionis based on its morpho-anatomical similarities to the species' original description by Weber-van Bosse 239 (1901), and the second was considered to be Tydemania gardineri based on its morpho-240 241 anatomical correspondence with the diagnosis of Gepp &Gepp (1911), even though the species is currently recognized as a synonym of T. expeditionis (Guiry & Guiry 2019). 242 243 The origin of the synonymy is due to Meinesz (1981) who based his proposition on previous studies (Srinivasan, 1954; Gilmartin, 1966) as well as his own observations. 244 According to Meinesz (1981), the differences in siphon diameters between T. expeditionis and 245 246 T. gardineri were not significant, and the author hypothesized that flabelliform thalli were juveniles of T. expeditionis, or restricted forms in unfavorable environmental conditions. Our 247 248 study, however, does not support the conclusions of Meinesz (1981), and instead shows clear 249 molecular and morphological evidence that T. gardineri is a distinct species. We also confirm the diagnostic characters (habit, aspect of stalks, and siphon diameters) described by Gepp & 250 Gepp (1911) to discriminate both species. 251

We were unable to compare our specimens with the holotypes of T. gardineri or T. 252 expeditionis, since none were designated by Gepp & Gepp (1911) or Weber-van Bosse 253 254 (1901), and no lectotypes have been assigned since. Likewise, except for one specimen from the Amirante Islands, Seychelles (L.3958458, Coppejans et al. 2001), we have not been 255 successful at finding other herbarium specimens previously identified as T. gardineri, either 256 257 because there is no information about where they are housed, or because they have been merged with collections of *T. expeditionis*. Similarly, it was difficult to locate specimens of *T*. 258 expeditionis but we have found the collection of Weber-van Bosse in L. None of these 259

specimens were designated as holotype at the time of description, but Weber-van Bosse 260 261 described the species from this Indonesian collection, which leads us to consider these specimens as syntypes. We also were not able to sequence type specimens, or newly collected 262 specimens from the syntype localities of T. gardineri (Chagos Archipelagos or Amirante 263 islands) or *T. expeditionis* (various localities in Indonesia) in our study, but we have examined 264 specimens from neighboring localities including Mayotte and northern Madagascar for T. 265 gardineri, and localities in Indonesia for T. expeditionis. This, combined with the clear 266 morpho-anatomical differences described above give us confidence to conclude that our two 267 genetic lineages correspond to T. expeditionis and T. gardineri. 268

269 The reappraisal of T. gardineri raises the question of the status of a third species, T. mabahithae Nasr, which was described by Nasr (1939) as having glomeruli only. The species 270 was also synonymized with T. expeditionis by Meinesz (1981) based on morphological 271 analysis of several samples collected from the type locality (Red Sea) similar to those 272 described by Nasr. Meinesz could not confirm differences between T. mabahithae and T. 273 274 expeditionis based on siphon diameters or other morpho-anatomical characters (Meinesz 1981). Our molecular analysis agrees with Meinesz (1981), as our sequences of Red Sea 275 276 samples (Egypt, Sudan), all showing a strict glomeruli form, clearly fell in the *T. expeditionis* 277 clade (Supplementary Information, Figure 1). Our results confirm that T. mabahithae should be considered a synonym of *T. expeditionis* as proposed by Meinesz (1981). 278

279

280 Geographical distribution of Tydemania species

Based on our data, the distribution of *Tydemania gardineri* appears to be restricted to the

282 Western Indian Ocean whereas *T. expeditionis* is found across the Indo-Pacific region and the

Red Sea (Fig. 25). From our data, it is possible that the southwestern boundary of *T*.

expeditionis distribution be the northern coast of Madagascar, and that its easternmost 284 285 distribution be Guam and Fiji. Northern Madagascar was the only locality where we found the two species in sympatry. Several records of T. gardineri are also reported for Taiwan (Shao 286 287 2003-2014) and New Caledonia (Farghaly 1980) but these are doubtful because the identifications were based on the presence of flabella only, which is a character that can be 288 found in T. expeditionis. Some New Caledonian specimens of our own collections (e.g., 289 290 NOU087203 and NOU087205) are similar in only having flabella, but grouped within the T. expeditionis clade based on DNA sequences. In the absence of DNA data, microscopic 291 observations, which were not made in previous studies, would be needed to confidently assess 292 293 identification based on siphon diameters, flabella shape and aspect of the stalk.

294

295 Taxonomical treatment:

296 Based on our results, we resurrect Tydemania gardineri, and provide a taxonomical description of the genus Tydemania and its two species: Tydemania expeditionis Weber-van 297 Bosse and Tydemania gardineri A. Gepp &E. Gepp. Despite intensive searches for specimens 298 of Tydemania in various herbaria, no holotype was found. However, we found a large 299 collection of Tydemania expeditionis specimens in L, including in the Weber-van Bosse 300 Herbarium. Six of these specimens were collected during the Siboga expedition and very 301 likely represent all or part of the specimens studied by Weber-van Bosse when writing the 302 303 diagnoses of the genus and its type species T. expeditionis (L.3998896, L.3998892, L.3998897, L.3998894, L.3998895, L.3998893). We consider these specimens as syntype 304 collections of Tydemania expeditionis. We expected to find the collection of Gepp & Gepp in 305 306 BM, but were not able to find any relevant specimens there. We have been unable to identify any other herbarium housing their collection and assume that their syntype of T. gardineri is 307 lost. Only one specimen of T. gardineri collected by E. Coppejans from near one of the 308

309	syntype localities was found in L (L.3958458, St. Joseph Atoll, Amirante Islands, Seychelles)
310	We propose to designate this specimen as a neotype for <i>T. gardineri</i> . We further list reference
311	DNA sequences of specimens of our own collection, collected near the type localities of both
312	species. For each species, the list of specimens studied for DNA analysis and morphology is
313	provided in Supplementary Information (Data 3)

314

315 *Tydemania*, Weber-van Bosse, 1901

Type species: *Tydemania expeditionis* Weber-van Bosse, 1901 Études sur les algues de

317 l'Archipel Malaisien. (III). Annales du Jardin Botanique de Buitenzorg 17: 126-141, pls XVII-

318 XIX

319 Morpho-anatomical description (adapted from the original description of Weber-van Bosse
320 (1901) and new observations made here):

Calcareous thallus composed of a monosiphonous cylindrical axis, simple or ramified, bearing cylindrical branches with rhizoids below, and branches assembled in stacked glomeruli and/or arranged in flabella above. In the glomeruli structures, the siphons are repeatedly divided dichotomously in alternate planes and interwoven to form a dense pompon-like structure. In the flabella, siphons are divided dichotomously in one plane and are aligned and adhere to form a fan. The siphon diameter decreases towards the tips of both structures. Growth is horizontal, and in some cases vertical. Reproductive structures are unknown.

328

Tydemania expeditionis Weber-van Bosse, 1901. Études sur les algues de l'Archipel
Malaisien. (III). Annales du Jardin Botanique de Buitenzorg 17: 39.

331	Syntypes: L.3998896 (De Bril, Herb. Weber-van Bosse); L.3998893 (Banda rif, Herb.
332	Weber-van Bosse); L.3998894 (Muaras rif; Herb. Weber-van Bosse); L.3998895 (Saleyer rif,
333	Herb. Weber-van Bosse); L.3998892, L.3998897 (unknown locality, Herb. Weber-van Bosse)
334	[digitized specimens available from L].

335

336 Syntype localities: Kabala dua reefs (Makassar Strait); Saleh bay (Sumbawa Island); de Bril,
337 and near Fau Island (Weber-van Bosse, 1901).

338 Morpho-anatomical description (adapted from the original description of Weber-van Bosse
339 (1901) and new observations made here):

340 Thallus calcified, glaucous green, with upright structures. Main axis cylindrical to moniliform 341 or septate-like near structures, 360-780 µm thick, with branches below bearing rhizoids, and branches above bearing glomeruli arranged in series and/or flabella. When both structures are 342 343 present, flabella are found closer to the base than glomeruli. Glomeruli are composed of 344 interwoven siphons divided dichotomously in alternate planes and form a sphere, or pomponlike structure. Glomeruli siphons measure 240-460 µm at the base and 80-180 µm at the 345 apices. Flabella are monostromatic and composed of aligned siphons dichotomously divided 346 347 in one plane, with symmetrical constrictions with thickened cell walls above dichotomies. Flabella are generally big and form loose groups of one to four. Flabella siphons taper from 348 349 200-450 µm at the base to 60-130 µm at the apices. Both glomeruli and flabella structures are borne on short and stocky stalks, dividing a few times, and arising in whorls from the main 350 351 axis. Growth is both horizontal and vertical, with main axes both creeping and ascending, and 352 apices protruding at the top of flabella and/or glomeruli series.

353

Distribution records: Africa: Egypt (Marconi et al., 2011; this study), Tanzania* (incl. 355 356 Zanzibar) (Coppejans et al., 2000), Sudan (this study). Indian Ocean Islands: Amirante Islands* (Silva et al., 1996), Andaman Islands * (Silva et al., 1996), Chagos Archipelago * 357 358 (Silva et al., 1996), Maldives (Silva et al., 1996; this study), Nicobar Islands (Silva et al., 1996), Seychelles * (Silva et al., 1996). South-west Asia: India (Silva et al., 1996; Sahoo et 359 al., 2001; Gupta 2012; Rao & Gupta, 2015). Asia: China (Tseng, 1984; Liu, 2008), Japan 360 361 (Okamura 1936; Segawa 1981, Yoshida et al., 1990; Yoshida 1998; Yoshida et al., 2015), South China Sea (Phang et al., 2016), Taiwan (Shao, 2003-2014). South-east Asia: Indonesia 362 (Verheij & Prud'homme van Reine, 1993; Atmadja & Prud'homme van Reine, 2014), 363 Philippines (Silva et al., 987; Ang et al., 2014), Vietnam (Tien, 2007; Nguyen et al., 2013). 364 Australia and New Zealand: Lord Howe Island (Lewis, 1987), Papua New Guinea (Coppejans 365 et al., 2001), Queensland (Lewis, 1987; Phillips, 1997; Phillips, 2002; Bostock & Holland, 366 2010). Pacific Islands: American Samoa (Skelton et al., 2004), Central Polynesia (Tsuda & 367 Walsh, 2013), Federated States of Micronesia (Lobban & Tsuda, 2003; Tsuda 2006), Fiji 368 (N'Yeurt et al., 1996; South & Skelton, 2003; Littler & Littler 2003; this study), Guam 369 370 (Lobban & Tsuda, 2003; Lam & Zechman, 2006), Mariana Islands (Tsuda, 2003), Marshall Islands (Taylor, 1950), New Caledonia (Payri, 2007; this study), Republic of Palau (Ohba et 371 372 al., 2007), Solomon Islands (Womersley & Bailey, 1970; this study), Vanuatu (this study). N.B.: "*" indicates localities which we suspect may be erroneous records resulting from 373 misleading species identification (referring to T. gardineri rather than T. expeditionis) and 374 should be confirmed with DNA sequencing or detailed anatomical analyses. 375 376

Tydemania gardineri A. Gepp & E. Gepp, 1911. The Codiaceae of the Siboga Expedition
including a monograph of Flabellarieae and Udoteae Siboga-Expeditie Monographie LXII:
67-68, 141, pl. XVIII: fig. 155

380 Neotype: L.3958458 from Seychelles, Amirante Islands, St. Joseph Atoll [digitized specimen
381 available from L].

382

We have been unable to locate any of the specimens from the Gepp & Gepp collection and assume that their syntype collection of *T. gardineri* is lost. Following art. 9.8 of the CIN, we have selected the specimen above because it is from Amirante Islands (Seychelles), one of the syntype localities of *T. gardineri*.

Syntype localities: Chagos Archipelago and Amirante Islands (Gepp & Gepp, 1911)

388

389 Morpho-anatomical description (adapted from the original description of Gepp & Gepp
390 (1911) and new observations made here):

Thallus calcified, laxly caespitose, greenish-gray, lacking glomeruli and composed of flabella 391 only. Creeping moniliform main axis, 250-570 µm in diameter, with sparse cylindrical 392 branches bearing rhizoids below and branches with flabella above. Flabella are borne from 393 long and moniliform stalks with a beaded necklace-like aspect, that are highly dichotomously 394 395 divided and arise from the main axis singly or in pairs. Stalks are densely arranged and superimposed at the basis of the flabella. Flabella are small, numerous, monostromatic, and 396 397 composed of aligned siphons dichotomously branched and tapering from 160-340 µm 398 diameter at the base to 30- 60 µm diameter at the apices. Constrictions above dichotomies are symmetric with a cell-wall thickening. Growth is horizontal. 399

400

401 Distribution: Western Indian Ocean: Chagos archipelago and Amirante Islands (Gepp &
402 Gepp, 1911); northern Madagascar (this study), Mayotte (this study), Banc du Geyser (this

- 403 study), Tanzania (this study), Seychelles, Amirante Islands (Coppejans et al., 2001); Asia:
- 404 Taiwan * (Shao 2003-2004).
- **N.B.:** the locality indicated with "*" is suspected to be an erroneous record based on a
- 406 misidentification of *T. expeditionis* specimens.

407 **AKNOWLEDGEMENTS**

408 This work was mainly supported by LabEx Corail grant.

- 409 Samples were collected during several scientific campaigns and by various collectors who we
- 410 could like to acknowledge here : Madang, 2012 : <u>http://dx.doi.org/10.17600/12100070</u>; New
- 411 Caledonia, 2012 : <u>http://dx.doi.org/10.17600/12100060</u> ; 2017 : N/O Alis PostBlanco1 ;
- 412 Kavieng, 2014 : <u>http://dx.doi.org/10.17600/14004400</u> ; Madagascar, 2016 : MAD
- 413 <u>http://dx.doi.org/10.17600/16004700</u>; Fiji, 2007 : BSM-Fidji, 10.17600/7100030 ; Vanuatu,
- 414 2006 : Santo, http://dx.doi.org/10.17600/6100100; Solomon Islands, 2004: BSM-Salomon,
- 415 2004; Mayotte 2016: SIREME. Sample collection in Bunaken and DNA analyses were made
- 416 possible thanks to the INDESO project, under the research permit 133/SIP/FRP/SM/V/2015
- 417 and 918/BLITBANKKP/II/2016 issued by the Indonesian government and under a material
- 418 transfer agreement between BALITBANG KP (now BRSDM KP, Ministry of Maritime
- 419 Affairs and Fisheries) and the IRD. The authors would like to thank Thierry Mulochau and
- 420 Lionel Bigot (Epicure project: TAAF, Ifremer and CUFR Mayotte) for samples collected
- 421 from the Banc du Geyser.

HV thanks Sofie D'Hondt, Joana Costa and Chiela Cremen for their molecular work on the
Bryopsidales. Part of the funding for this study was provided by the Australian Biological
Resources Study (project RFL213-08 to HV).

425 SUPPLEMENTARY INFORMATION

- 426 Table 1: list of molecular markers and corresponding primers
- 427 Table 2: List of vouchers, Bold sample IDs and Genbank accession numbers
- 428 Table 3: Morphological observations
- 429 Figure 1: Maximum clade credibility tree obtained from a Beast analysis of the *tuf*A collapsed
- 430 dataset. The number of identical haplotypes is indicated between brackets. The origin of each
- 431 sample is indicated by colored circles and correspond to the map on the right. Results of the
- 432 ABGD and PTP species delimitation methods are shown and the presence of flabella (F) and
- 433 glomeruli (G) is indicated.

- 435 Data 1: Overview of the ABGD and PTP species delimitation results for *rbc*L
- 436 Data 2: Overview of the species ABGD and PTP delimitation results for 18S rDNA
- 437 Data 3: List of specimens studied through molecular and morphological approaches for *T*.
- 438 *expeditionis* and *T. gardineri*

439 AUTHORS CONTRIBUTIONS

440 L. Lagourgue acquired and analyzed most molecular and morphological data, drafted and

edited the manuscript; H. Verbruggen collected samples from the Red Sea and Tanzania,

- 442 produced molecular data for these localities and contributed to writing and editing the
- 443 manuscript; E. Ampou collected samples from Indonesia and contributed to editing the
- 444 manuscript; C. Payri collected samples from various localities, and contributed to editing the
- 445 manuscript.

446 **REFERENCES**

- Ang, P.O, Jr., Sin Man Leung & Mei Mei Choi (2014). A verification of reports of marine algal species
 from the Philippines. *Philippine Journal of Science, Special Issue* 142
- 449 Atmadja, W.S. & Prud'homme van Reine, W.F. (2014). *Checklist of the seaweed species biodiversity of*
- 450 Indonesia with their distribution and classification: green algae (Chlorophyta) and brown algae
- 451 (Phaeophyceae, Ochrophyta). pp. [2], i-v, 1-59. Leiden & Indonesia: Naturalis Biodiversity Centre,
- 452 Indonesian Institute of Sciences (LIPI)
- 453 Bostock, P.D. & Holland, A.E. (2010). *Census of the Queensland Flora*. pp. 1-320. Brisbane:
- 454 Queensland Herbarium Biodiversity and Ecosystem Sciences, Department of Environment and455 Resource Management.
- Coppejans, E., De Clerck, O. & Leliaert, F. (2001). Marine brown algae (Phaeophyta) from the north
 coast of Papua New Guinea, with a description of Dictyota magneana sp.nov. *Cryptogamie, Algologie,*
- 458 22:15–40.
- 459 Coppejans, E., Leliaert, F. & De Clerck, O. (2000). Annotated list of new records of marine macroalgae
- for Kenya and Tanzania, since Isaac's and Jaasund's publications. *Biologisch Jaarbuch Dodonaea* 67:
 31-93, 1 map, 28 figs.
- 462 Doyle, J.J. & Doyle, J.L. (1987). A Rapid DNA Isolation Procedure for Small Quantities of Fresh Leaf
 463 Tissue. *Phytochemical Bulletin* 19, 11-15
- Drummond, A.J., Ho, S.Y.W., Phillips, M.J. & Rambaut, A. (2006). Relaxed Phylogenetics and Dating
 with Confidence. *PLoS Biology* 4(5): e88. DOI: 10.1371/journal.pbio.0040088
- Edgar, R.C. (2004). MUSCLE: multiple sequence alignment with high accuracy and high throughput.
 Nucleic Acids Research. 32:1792–7.
- Farghaly, M. (1980). Algues Benthiques de la Mer Rouge et du bassin occidental de l'océan Indien.
 Université des sciences et techniques du Languedoc, Montpellier, 274 pp.
- 470 Gepp, A. & Gepp, E.S. (1911). The codiaceae of the Siboga Expedition, including a monograph of
 471 Flabellarieae and Udoteaceae. *Siboga-Expeditie*. 62:1–150.
- Gilmartin, M. (1966). Ecology and morphology of *Tydemania expeditions*, a tropical deep-water
 siphonous green alga. *Journal of Phycology*. 2:100–5.
- 474 Guiry, M.D. & Guiry, G.M. (2019). *AlgaeBase*. World-wide electronic publication, National University
 475 of Ireland, Galway. http://www.algaebase.org
- 476 Gupta, R.K. (2012). Algae of India Volume 2. A checklist of Chlorophyceae, Xanthophyceae.
- 477 *Chrysophyceae and Euglenophyceae.* pp. [i]-iii, [1]-428, 8 pls. Salt Lake, Kolkata: Botanical Survey of
 478 India, Ministry of Environment & Forests.
- 479 Handeler, K., Wagele, H., Wahrmund, U., Rudinger, M. & Knoop, V. (2010). Slugs' last meals:
- 480 Molecular identification of sequestered chloroplasts from different algal origins in Sacoglossa
- 481 (Opisthobranchia, Gastropoda). *Molecular Ecology Resources*. 10:968–78.
- 482 Kearse, M., Moir, R., Wilson, A., Stones-Havas, S., Cheung, M., Sturrock, S., Buxton, S., Cooper, A.,
- 483 Markowitz, S., Duran, C., Thierer, T., Ashton, B., Meintjes, P. & Drummond, A. (2012). Geneious Basic:
 484 An integrated and extendable desktop software platform for the organization and analysis of
- 485 sequence data. *Bioinformatics*. 28:1647–9.
- 486 Kooistra, W.H.C.F. (2002). Molecular phylogenies of Udoteaceae (Bryopsidales, Chlorophyta) reveal

- 487 nonmonophyly for Udotea, Penicillus and Chlorodesmis. *Phycologia*. 41:453–62.
- 488 Lagourgue, L., Puillandre, N. & Payri, C.E. (2018). Exploring the Udoteaceae diversity (Bryopsidales,
- Chlorophyta) in the Caribbean region based on molecular and morphological data. *Molecular Phylogenetics and Evolution* 127: 758-769.

Lam, D.W. & Zechman, F.W. (2006). Phylogenetic analyses of the Bryopsidales (Ulvophyceae,
Chlorophyta) based on Rubisco large subunit gene sequences. *Journal of Phycology*. 42:669–78.

Lanfear, R., Calcott, B., Ho, S.Y.W. & Guindon, S. (2012). PartitionFinder: Combined Selection of
Partitioning Schemes and Substitution Models for Phylogenetic Analyses. *Molecular Biology and Evolution*. 29:1695–701.

- Lewis, J.A. (1987). *Checklist and bibliography of benthic marine macroalgae recorded from northern Australia III. Chlorophyta.* pp. 1- 56.: Department of Defense. Defense Science and Technology
 Organisation. Materials Research Laboratories, Melbourne, Victoria, Report MRL-R-1063
- Littler, D.S. & Littler, M.M. (2003). *South Pacific Reef Plants. A diver's guide to the plant life of the South Pacific Coral Reefs*. pp. 1-331. Washington, DC: OffShore Graphics, Inc.
- Liu, R.Y. (Ed.) (2008). *Checklist of biota of Chinese seas*. pp. 1-1267. Beijing: Science Press, Academia Sinica
- Lobban, C.S. & Tsuda, R.T. (2003). Revised checklist of benthic marine macroalgae and seagrasses of
 Guam and Micronesia. *Micronesica* 35/36: 54-99
- 505 Marconi, M., Giordano, M. & Raven, J.A. (2011). Impact of taxonomy, geography, and depth on
- delta13C and delta15N variation in a large collection of macroalgae. *Journal of Phycology* 47(5):
 1023-1035
- Meinesz, A. (1981). Sur le genre *Tydemania* (Udotéacée, Caulerpale) et sa présence en Mer Rouge. *Cryptogamie, Algologie*. 2:57–66.
- 510 Miller, M.A., Pfeiffer, W. & Schwartz, T. (2010). Creating the CIPRES Science Gateway for inference of 511 large phylogenetic trees. In: *Gateway Computing Environments Workshop (GCE)*. IEEE, pp. 1–8.
- N'Yeurt, A.D.R., South, G.R. & Keats, D.W. (1996). A revised checklist of the benthic marine algae of
 the Fiji Islands, South Pacific (including the island of Rotuma). *Micronesica*. 29:49–98.
- Nasr, A.H. (1939). Algae. *Publications of the Marine Biological Station, Ghardaqa* 1: 47-76, 17 figs, 1
 plate
- Nguyen, T.V., Le, N,H., Lin, S.-M., Steen, F. & De Clerck, O. (2013). Checklist of the marine macroalgae
 of Vietnam. *Botanica Marina* 56(3): 207-227
- 518 Ohba, H., Victor, S., Golbuu, Y & Yukihara, H. (2007). *Tropical marine plants of Palau*. pp. [1]-153,
- Numerous colour photographs; 8 figs. Koror: Palau International Coral Reef Center; JapanInternational Cooperation Agency.
- 521 Okamura, K. (1936). *Nippon kaisô shi [Descriptions of Japanese algae]*. pp. [4], [1]-964, [1]-11, 522 frontispiece portrait, 1-427 figs. Tokyo: Uchida Rokakuho.
- 523 Payri, C.E. (2007). Revised checklist of marine algae (Chlorophyta, Rhodophyta and Ochrophyta) and
- 524 seagrasses (Marine Angiosperma) of New Caledonia. In: *Compendium of marine species from New*
- 525 Caledonia. Documents Scientifique et Techniques. II7. Sciences de la Mer. Ed. 2. (Payri, C.E. & Richer
- de Forges, B. Eds), pp. 95-112. Nouvelle-Calédonie: Centre IRD de Nouméa
- 527 Phang, S.-M., Yeong, H.-Y., Ganzon-Fortes, E.T., Lewmanomont, K., Prathep, A., Hau, L.N., Gerung,
- 528 G.S, & Tan, K.S. (2016). Marine algae of the South China Sea bordered by Indonesia, Malaysia,

- 529 Philippines, Singapore, Thailand and Vietnam. *Raffles Bulletin of Zoology Supplement* 40: 13-59
- Phillips, J.A. (1997). Algae. In: *Queensland Plants: Names and Distribution*. (Henderson, R.J.F. Eds), pp.
 223-240. Indooroopilly, Queensland: Queensland Herbarium, Department of Environment.
- 532 Phillips, J.A. (2002). Algae. In: *Names and distribution of Queensland plants, algae and lichens*.
- (Henderson, R.J.F. Eds), pp. 228-244. Brisbane: Queensland Government Environmental ProtectionAgency.
- Puillandre, N., Lambert, A., Brouillet, S. & Achaz, G. (2012). ABGD , Automatic Barcode Gap Discovery
 for primary species delimitation. *Molecular Biology and Evolution*. 1864–77.
- 537 Rambaut, A. & Drummond, A. (2007). Tracer version 1.5.
- 538 Rao, P.S.N. & Gupta, R.K. (2015). Algae of India Volume 3. A checklist of Indian marine algae
- *(excluding diatoms & dinoflagellates).* pp. [i]-xviii, [1]-93, 11 pls. Salt Lake, Kolkata: Botanical Survey
 of India Ministry of Environment, Forests & Climate Change Government of India
- Ronquist, F. & Huelsenbeck, J.P. (2003). MrBayes 3: Bayesian phylogenetic inference under mixed
 models. *Bioinformatics (Oxford, England)*. 19:1572–4.
- Sahoo, D., Nivedita & Debasish (2001). *Seaweeds of Indian coast.* pp. xxi + 283. New Delhi: A.P.H.
 Publishing
- Segawa, S. (1981). *Genshoku Nihon kaiso zukan. Colored illustrations of the seaweeds of Japan.*Fourth printing. pp. [i]-xviii, 1-195, pls 1-72. Osaka: Hoikusha Publishing Co. Ltd
- 547 Shao, K.T (2003-2014). TaiBNET (Catalogue of Life in Taiwan). http://taibnet.sinica.edu.tw. Taiwan
- Silva, P.C., Menez, E.G. & Moe, R.L. (1987). *Catalog of the benthic marine algae of the Philippines.*Smithsonian Institution Press, Washington, D.C. 1-179 pp.
- Silva, P.C., Basson, P.W. & Moe, R.L. (1996). *Catalogue of the benthic marine algae of the Indian Ocean*. University of California Publications in Botany 79: 1-1259
- 552 Skelton, P.A. & South, G.R. (2004). New records and notes on marine benthic algae of American
 553 Samoa Chlorophyta & Phaeophyta. *Cryptogamie Algologie* 25: 291-312
- South, G.R. & Skelton, P.A. (2003). Catalogue of the marine benthic macroalgae of the Fiji Islands,
 South Pacific. *Australian Systematic Botany* 16: 699-758
- 556 Srinivasan, K.S. (1954). On a rare and little known alga (*Tydemania expeditionis* Weber-van Bosse) 557 new to Nancowry. *Phytomorphology* 4: 247-255.
- 558 Stamatakis, A. (2014). RAxML version 8: a tool for phylogenetic analysis and post-analysis of large 559 phylogenies. *Bioinformatics*. 30:1312–3.
- 560 Stamatakis, A., Hoover, P., Rougemont, J. & Renner, S. (2008). A Rapid Bootstrap Algorithm for the 561 RAxML Web Servers. *Systematic Biology*. 57:758–71.
- Taylor, W.R. (1950). *Plants of Bikini and other northern Marshall Islands*. pp. [i]-xv, [1]- 227, 79 pls.
 Ann Arbor: University of Michigan Press
- Thiers, B. (2016). Index Herbariorum: A Global Directory of Public Herbaria and Associated Staff. New
 York Botanical Garden's Virtual Herbarium. http://sweetgum.nybg.org/science/ih/
- 566 Tien, N.V. (2007). Thuc Vât Chí Viêt Nam. Flora of Vietnam. Volume 10: Nganh Rong Luc -
- 567 Chlorophyta Pascher (Cac Taxon Vung Bien). pp. 1-279, 162 fig. Hà Nôi [Hanoi]: Science and Technic
 568 Publishing House

- 569 Tseng, C.K. (1984). Common Seaweeds of China. pp. 1-318, 149 pls. Beijing: Science Press
- 570 Tsuda, R.T. (2003). Checklist and bibliography of the marine benthic algae from the Mariana Islands
- 571 (Guam and CNMI). *Technical Report. University of Guam Marine Laboratory* 107: i-v, 1-49
- 572 Tsuda, R.T. (2006). Checklist and bibliography of the marine benthic algae within Chuuk, Pohnpei, and
- 573 *Kosrae States, Federated States of Micronesia*. pp. 1-43. Honolulu: Bishop Museum Press 1525
 574 Bernice Street, Honolulu, Hawai
- Tsuda, R.T. & Walsh, S.K. (2013). Bibliographic checklist of the marine benthic algae of Central
 Polynesia in the Pacific Ocean (excluding Hawai'i and French Polynesia). *Micronesica*. 2013-02: 1-91
- Verbruggen, H., Ashworth, M., LoDuca, S.T., Vlaeminck, C., Cocquyt, E., Sauvage, T., Zechman, F.W.,
 Littler, D.S., Littler, M.M., Leliaert, F. & De Clerck, O. (2009). A multi-locus time-calibrated phylogeny
 of the siphonous green algae. *Molecular Phylogenetics and Evolution*. 50:642–53.
- Verheij, E. & Prud'homme van Reine, W.F. (1993). Seaweeds of the Spermonde Archipelago, SW
 Sulawesi, Indonesia. *Blumea* 37: 385-510, 6 figs, 1 table, 19 plates
- Weber-van Bosse, A. (1901). Etudes sur les algues de l'archipel malaisien. Annales du Jardin
 Botanique de Buitenzorg. 17: 126-141, pls XVII-XIX..
- Womersley, H.B.S. & Bailey, A. (1970). Marine algae of the Solomon Islands. *Philosophical Transactions of the Royal Society, London.Biological Series B*. 259:257–352.
- Yoshida, T. (1998). *Marine algae of Japan.* pp. [1-2], 1-25, 1-1222. Tokyo: Uchida Rokakuho Publishing
 Co., Ltd.
- Yoshida, T., Nakajima, Y. & Nakata, Y. (1990). Check-list of marine algae of Japan (revised in 1990).
 Japanese Journal of Phycology 38: 269-320
- Yoshida, T., Suzuki, M. & Yoshinaga, K (2015). Checklist of marine algae of Japan (Revised in 2015).
 Japanese Journal of Phycology 63: 129-189
- 592 Zhang, J., Kapli, P., Pavlidis, P. & Stamatakis, A. (2013). A general species delimitation method with
- applications to phylogenetic placements. *Bioinformatics*. 29:2869–76.

594 TABLE

Table 1: Morpho-anatomical characters and geographical distribution of *Tydemania*

596 *expeditionis* and *Tydemania gardineri* resulting from the present study, and values in bold

from the descriptions of Gepp &Gepp (1911). Values (average \pm standart deviation)

Morpho-		
anatomical	Tydemania expeditionis	Tydemania gardineri
characters		
	Upright structures formed by a	Caespitose patch of flabella
Habit	succession of glomeruli and/or	arising from a creeping main
	flabella	axis
Clomonuli	Predominant when present/	Always absent
Glomeruli	Lacking in strict flabella form	
Fish alla	Sometimes absent (strict	Always present
riadena	glomeruli form)	
Diameter of the	360 - 780 μm (518 ±100)	285 - 570 μ m (426 ± 94)
Diameter of the main axis	360 - 780 μm (518 ±100) (450-500 μm)	285 - 570 μm (426 ± 94) (250-400 μm)
Diameter of the main axis Diameter of flabella	360 - 780 μm (518 ±100) (450-500 μm) 60-130 μm (79 ± 13)	285 - 570 μm (426 ± 94) (250-400 μm) 30-60 μm (44.5 ±7)
Diameter of the main axis Diameter of flabella siphons at apices	360 - 780 μm (518 ±100) (450-500 μm) 60-130 μm (79 ± 13) (Min. 63 μm)	285 - 570 μm (426 ± 94) (250-400 μm) 30-60 μm (44.5 ±7) (50-40 μm)
Diameter of the main axis Diameter of flabella siphons at apices	360 - 780 μm (518 ±100) (450-500 μm) 60-130 μm (79 ± 13) (Min. 63 μm) Flabella generally big and in small	$285 - 570 \ \mu m \ (426 \pm 94)$ $(250-400 \ \mu m)$ $30-60 \ \mu m \ (44.5 \pm 7)$ $(50-40 \ \mu m)$ Flabella generally small but
Diameter of the main axis Diameter of flabella siphons at apices Flabella shape and	360 - 780 μm (518 ±100) (450-500 μm) 60-130 μm (79 ± 13) (Min. 63 μm) Flabella generally big and in small number (1 to 4), arising one by	$285 - 570 \mu m (426 \pm 94)$ (250-400 μm) 30-60 $\mu m (44.5 \pm 7)$ (50-40 μm) Flabella generally small but numerous (several dozens),
Diameter of the main axis Diameter of flabella siphons at apices Flabella shape and number	$360 - 780 \ \mu m \ (518 \pm 100)$ (450-500 \ \ \ m) $60-130 \ \mu m \ (79 \pm 13)$ (Min. 63 \ \ m) Flabella generally big and in small number (1 to 4), arising one by one or in pairs, and forming loose	$285 - 570 \ \mu m \ (426 \pm 94)$ (250-400 \ \ \ m) 30-60 \ \ \ m \ (44.5 \ \pm 7) (50-40 \ \ \ m) Flabella generally small but numerous (several dozens), assembled in dense groups
Diameter of the main axis Diameter of flabella siphons at apices Flabella shape and number	$360 - 780 \ \mu m \ (518 \pm 100)$ (450-500 \ \ \ m) $60-130 \ \mu m \ (79 \pm 13)$ (Min. 63 \ \ m) Flabella generally big and in small number (1 to 4), arising one by one or in pairs, and forming loose groups	$285 - 570 \mu m (426 \pm 94)$ (250-400 μm) 30-60 $\mu m (44.5 \pm 7)$ (50-40 μm) Flabella generally small but numerous (several dozens), assembled in dense groups
Diameter of the main axis Diameter of flabella siphons at apices Flabella shape and number	$360 - 780 \mu m (518 \pm 100)$ $(450-500 \mu m)$ $60-130 \mu m (79 \pm 13)$ $(Min. 63 \mu m)$ Flabella generally big and in small number (1 to 4), arising one by one or in pairs, and forming loose groups Short, stocky and fewly divided	$285 - 570 \mu m (426 \pm 94)$ $(250-400 \mu m)$ $30-60 \mu m (44.5 \pm 7)$ $(50-40 \mu m)$ Flabella generally small but numerous (several dozens), assembled in dense groups Long, highly dichotomously

		strong constrictions (beaded
		siphons), crowded and
		superimposed at the basis of
		the flabella
Stalk aspect	Monoliform to septate	Monoliform
	In whorls, arising in groups of two	Simple, arising from the main
Stalk attachment in	or four, on the same section of the	axis
	main axis.	
	Horizontal and vertical growth	Horizontal growth (creeping
Growth	(main axis both creeping and	main axis only)
Growth	(main axis both creeping and ascending)	main axis only)
Growth Distribution	(main axis both creeping and ascending) Indo-Pacific and Red Sea	main axis only) Western Indian Ocean only

600 FIGURE LEGENDS

Figure 1: Diagram showing the different morphological structures of *Tydemania:* specimenwith both glomeruli and flabella (left), and with flabella only (right).

603

Figures 2-14: Tydemania expeditionis. Fig. 2: In situ habit of the thalli with flabella and 604 glomeruli; Fig. 3: In situ habit of the thalli with glomeruli only; Fig. 4: In situ habit of the 605 606 thalli with flabella only; Fig. 5: Succession of flabella and glomeruli arising from the main axis; Fig. 6: Monoliform main axis near glomeruli and flabella; Fig. 7: Siphons branching 607 608 dichotomously; Fig. 8: Anisomorphic dichotomy; Fig. 9: Short and monoliform stalks with a few dichotomous divisions (flabella form); Fig. 10: Short and monoliform stalks with few 609 dichotomous divisions (glomeruli form); Fig. 11: Verticillate stalks at the basis of a flabella; 610 Fig. 12: Verticillate stalks at the basis of a glomeruli; Fig. 13: Apex of the main axis 611 612 protruding at the top of a flabella; Fig. 14: Apex of main axis protruding at the top of a glomeruli. Scale bars: Fig. 2: 0.8 cm; Fig. 3: 3.2 cm; Fig. 4: 2 cm; Fig. 5: 3.5 mm; Fig. 6: 613 600 μm; Fig. 7: 320 μm; Fig. 8: 160 μm; Fig. 9: 760 μm; Fig. 10: 820 μm; Fig. 11: 475 μm; 614 Fig. 12: 800 µm; Fig. 13: 875 µm ; Fig. 14: 500 µm. 615

616



623 Fig. 23: 350 μm; Fig. 24: 250 μm.

624	Figure 25: From left to right: (1) Maximum likelihood tree reconstruction from the
625	concatenated matrix (tufA, rbcL and 18S rDNA). Circles at each node represent ML bootstrap
626	values (left) and posterior probabilities (right). Black circles indicate high support (bs> 90 and
627	PP >95), grey circles indicate low support (bs<90 and PP <95), and white circles indicate
628	incongruence between maximum likelihood and Bayesian trees. (2) Habit information for
629	each specimen: presence of flabella (F) or glomeruli (G). (3) Grouping of sequences per
630	marker according to the results of the species delimitation analyses (stripes indicate missing
631	sequences). (4) Global geographical distribution. (5) Drawings showing the external habits
632	observed for specimens in each of the two main clades. (6) Map showing the distribution of
633	specimens from the two main clades