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## Nonalcoholic fatty liver disease: modulating gut microbiota to improve severity?

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### **Abstract**

Gut microbiota plays a role in the pathophysiology of metabolic diseases which also include nonalcoholic fatty liver diseases (NAFLD), through the gut-liver axis. To date, clinical

guidelines recommend a weight loss goal of 7 to 10% to improve features of NAFLD. Nevertheless, since this target is not easily achieved by all patients, alternative therapeutic options are currently being evaluated. This review focusses on therapeutics that aims to modulate the gut microbiota and the gut-liver axis. We will herein discuss how probiotics, prebiotics, synbiotic, fecal microbiota transfer, polyphenols, specific diets and exercise interventions have been shown to modify gut microbiota signatures, improve NAFLD outcomes and detail, when available, the different mechanisms by which these beneficial outcomes might occur. Apart from probiotics which have already been tested in human RCTs, most of these potential therapeutics have been studied in animals. Their efficacy still warrants confirmation in humans using appropriate design.

## Introduction

Non-alcoholic fatty liver disease (NAFLD) prevalence is increasing worldwide<sup>1</sup>, partly due metabolic disease **progression** such as insulin-resistance, type 2 diabetes (T2D) and overweight/obesity<sup>2</sup>. NAFLD also strongly relates to our current lifestyle. Diets rich in saturated-fatty acids, sugar-sweetened beverages, refined carbohydrates, fructose, Western diet and high caloric intake promote both obesity and NAFLD<sup>3</sup>. However, the pathophysiology of NAFLD is highly complex and involves numerous pathways including insulin-resistance, inflammation, lipotoxicity, increased de novo-lipogenesis and oxidative stress<sup>4,5</sup>. The contributive part and kinetic of such events in NAFLD development and progression need to be better understood, as also molecular events.

The gut microbiota (GM) is considered a novel organ involved in NAFLD pathophysiology. GM has been studied in obesity and T2D where its role was proposed based on phenotypes' transmission using fecal microbiota transfer (FMT) experiments from mice or humans into mice recipients<sup>6,7</sup>. Moreover long-term dietary habits shapes the GM composition<sup>8-10</sup>, and diet modifications strongly modulate its composition<sup>11</sup>. Noteworthy, this effect could partly explain the link between adverse diet profiles and NAFLD.

The study of the GM within NAFLD now paves the way to (i) demonstrate its causal contribution, (ii) better understand its pathophysiological mechanisms and finally (iii) decipher microbial signatures associated with the disease and its severity stages. Therefore, research evolved towards therapeutic tools to modulate the GM in order to improve NAFLD and/or limit its progression. Several means are now available and have been tested in NAFLD, mostly in animal models. This review aims to detail the different options to modulate the GM, show how these “therapeutic” actions can improve NAFLD and describe the involved mechanisms, mostly related to the gut-liver axis.

### Gut dysbiosis in metabolic diseases: impact on NAFLD?

GM has been studied deeply these past ten years, using different high-throughput technics each presenting pros and cons as reviewed<sup>12</sup> and described briefly in **Table 1**. GM changes during obesity and T2D and reveal severe dysbiosis (i.e. alteration of GM composition and function with negative effects on host metabolism). Obesity is characterized by decreased microbial gene richness (MGR)<sup>11,13</sup>. Furthermore, the prevalence of patients with low MGR increases with the severity of obesity, from 40% during mild obesity to 75% of individuals with severe obesity<sup>14</sup>. Noteworthy, low MGR is associated with a proinflammatory status<sup>15</sup> as well as with an altered clinical phenotype: worse adiposity<sup>11,13</sup> with abdominal distribution and propensity to metabolic alterations (including T2D and hypertension)<sup>14</sup>. All these factors are involved in NAFLD physio-pathology<sup>16</sup>. Furthermore, M GM undergoes drastic changes in composition during obesity with for example higher capacity to increase energy storage<sup>17</sup>, albeit this alteration was not always replicated. Obese individuals with low MGR also display a reduction in bacteria (i) producing short chain fatty acids (SCFA), (ii) involved in hydrogen and methane production and (iii) with the potential to manage oxidative stress<sup>13</sup>. Concomitantly, bacteria involved in intestinal mucus remodeling are changing as illustrated by the decrease in *Akkermansia muciniphila*, seen in some overweight, obese<sup>18</sup> and prediabetic individuals<sup>19</sup>. Patients with low MGR also display increased bacteria able to synthesize lipopolysaccharide (LPS), which was further related to insulin-resistance and adverse lipidomic profile<sup>20</sup>, both of which are also involved in NAFLD pathogenesis<sup>16</sup>. In T2D, GM similarly undergoes profound changes in terms of reduced MGR as well as composition

and functional changes<sup>21,22</sup>. Interestingly, this modified GM profile also translates in differential metabolite production between T2D, pre-diabetes and normoglycemic individuals, which is further observed in the systemic<sup>23,24</sup> and even in the portal blood<sup>24</sup>.

Thus, metabolic alteration-related GM changes could then contribute to NAFLD development. Since, NAFLD now represents the liver component of the metabolic syndrome<sup>2,25</sup>, one would anticipate that liver disease is also associated with GM dysbiosis and indeed, NAFLD-related GM signatures have recently been reported<sup>26</sup>. Several studies have analyzed and compared the GM composition of NAFLD or NASH patients to that of control subjects, summarized elsewhere<sup>27</sup>. In brief, NAFLD is frequently associated with increased Proteobacteria<sup>28–32</sup> at the phylum level, while at the family level, *Rikenellaceae*<sup>30,33</sup> and *Rumminocaceae*<sup>29–32</sup> are decreased and *Enterobacteriaceae*<sup>29,32</sup> increased. Finally, at the genera level, NAFLD is marked by increased *Escherichia*<sup>28,32</sup> and *Dorea*<sup>30,34</sup> and decreased *Anaerospobacter*<sup>33,35</sup>, *Coprococcus*<sup>28,32,33</sup>, *Eubacterium*<sup>28,32</sup>, *Faecalibacterium*<sup>32,35</sup> and *Prevotella*<sup>28,36</sup>. Likewise, microbial signatures of NAFLD-related fibrosis have been published. Compared to either healthy or low to mild fibrosis, patients with advanced fibrosis (F3-F4) display increased Gram-negative microbes, decreased Firmicutes and increased Proteobacteria abundance at the phylum level, while at the species level, *E. coli* and *B. vulgatus* were the most abundant and *E.rectale*<sup>31</sup> was decreased, a signature already observed during metabolic diseases<sup>14,37</sup>. Finally, species within the Enterobacteriaceae family<sup>38</sup> and the Streptococcus genera<sup>38,39</sup> are the most enriched in NAFLD-cirrhosis patients, the end-stage of the severity spectrum.

Nevertheless, discrepant results are also seen across studies<sup>35,40–42</sup> in terms of NAFLD- and fibrosis-related microbial signatures, being dependent or not on the existence of metabolic disorders. These aspects have been discussed in length in (Aron-wisnewsky et al Nature Review hepatol and gastro: in press) and GM signature variability appears to depend on study designs that involved different types of control, severity of obesity, the presence and severity of other related-metabolic alterations and their specific treatment, ethnicity, the stage of NAFLD and the methods used for NAFLD diagnosis. Nonetheless, models combining several microbial species and a few clinical parameters accurately predict patients with NAFLD-related advanced fibrosis<sup>31</sup> or NAFLD-cirrhosis<sup>38,39</sup>. These models have been

validated in several independent cohorts<sup>31,38</sup>, suggesting that non-invasive microbiota-related biomarkers could be useful to identify diseased patients, but probably combined with other information. Interestingly, since (i) first-degree relatives of patients diagnosed with NAFLD-cirrhosis have a 12-fold increased risk of advanced fibrosis<sup>43</sup> and (ii) shared-housing individuals also share a large similarity in their microbiome<sup>38,44</sup>, it might, in the future, be interesting to screen patient's family members after the proband has been diagnosed with NAFLD-cirrhosis using these non-invasive microbiota models. Nevertheless, this should be further validated in independent cohorts and cost effectiveness should be evaluated.

### **Rationale for a role of the gut microbiota in NAFLD physio-pathogenesis**

Although, deciphering a NAFLD-related microbial signature has some interest for biomarker development and future use in routine care, demonstrating the causal role of GM in NAFLD development remains critical to improve understanding of its pathophysiology, identify new pathways and subsequent adequate therapeutic interventions targeting identified novel pathways.

GM role in NAFLD originates from mouse studies using FMT or co-housing experiments<sup>45</sup>. FMT from metabolically-altered mice into germ-free mice reproduced some NAFLD histology features, but not all. Specifically, FMT from conventional mice upon high-fat diet (HFD) that developed metabolic alterations including steatosis, into germ-free mice, translated into a 3-fold increase in liver triglyceride content and increased expression of liver lipogenesis genes in the recipients<sup>6,46</sup>. Conversely, germ-free mice receiving FMT from obese weight-matched mice without steatosis maintained their healthy liver, thus showing that some NAFLD-related microbiota alterations are, at least partly, involved in liver injury<sup>46</sup>. These findings were confirmed in humans<sup>47</sup>. FMT from NAFLD patients or healthy individuals into gnotobiotic recipient mice induced in the former group a 4.6-fold increase in liver triglyceride contents, authentic liver histological alterations (i.e. steatosis and inflammation), increased expression of liver genes involved in inflammatory pathways resulting in increased systemic inflammation and LPS concentration (endotoxemia). These alterations were even further exacerbated in mice fed a HFD<sup>47</sup>. Overall, these reports propose a contribution of the GM in NAFLD pathophysiology.

## Gut microbiota-related mechanism involved in NAFLD

If NAFLD **complex** pathogenesis include many factors<sup>48</sup>, several mechanisms, now termed the gut-liver axis, involve the GM as reviewed herein<sup>49–52</sup>. In brief, they include<sup>50,53,54</sup> the enterohepatic circulation of bile acids<sup>55</sup>, increased intestinal permeability, systemic inflammation *per-se* and altered immunity as well as the role of microbial-related metabolites<sup>56,57</sup>. Frequently cited metabolites include choline metabolite<sup>58</sup>, phospholipids<sup>54</sup>, microbial-associated molecular patterns, SCFA, microbially-produced ethanol<sup>59,60</sup> and 3-(4-hydroxyphenyl) lactate<sup>61</sup> reviewed in (Aron-wisnewsky et al Nature Review hepatol and gastro: in press).

Since this review focuses on potential therapeutics involving GM modulation to improve NAFLD, we chose to **solely** detail **pathways demonstrated to be ameliorated** after **GM-targeted** interventions associated with a **proven** beneficial effect on NAFLD.

### Intestinal permeability / endotoxemia

Human studies have demonstrated increased intestinal permeability in biopsy-proven NAFLD patients compared to healthy controls<sup>62</sup>. Intestinal permeability can be measured using a lactulose/mannitol test<sup>60,63</sup> or the urinary excretion of Cr-ethylene diamine tetraacetate<sup>64</sup>. Increased intestinal permeability leads to translocation of total or parts of bacteria membrane, subsequently leading to increased concentration of LPS<sup>65,66</sup>. Endotoxemia rises in humans with increasing liver severity from NAFLD towards NASH as seen with the direct measure of systemic LPS concentration<sup>63,67,68</sup> or the indirect measure of LBP<sup>69</sup>, which is the LPS-binding protein. Interestingly, intestinal permeability increases **as early as 1-week after a HFD in mice** (as seen with decreased ZO-1 intestinal protein expression and FITC-dextran fluorescent probe associated with increased bacterial translocation to the lamina propria)<sup>70</sup>. A longer HFD duration translated into similar increased intestinal permeability yet associated with liver alterations (i.e. steatosis, fibrosis and inflammation) and the presence of intra-liver parenchyma bacteria, thus confirming bacterial translocation. These results suggest that gut barrier dysfunction **represents** an early event in NAFLD pathogenesis that is further followed by bacterial translocation and liver alterations.

To assess the role of GM therein, FMT from mice submitted to 1-week HFD or chow diet were performed in specific-pathogen free (SPF) mice fed a chow diet. **Increased intestinal permeability was only observed in SPF mice receiving the FMT of mice fed high-fat diet, suggesting the major impact of the diet on the GM that subsequently transfer in clinical adverse outcomes.** Disruption of gut vascular barrier (GVB) is mandatory to induce NASH in the recipient mice (as demonstrated with a cre-lox model of mice on the WNT/ $\beta$ -catenin system, involved in GVB)<sup>70</sup>. These experiments in mice demonstrate that altered GVB is an early event, necessary to induce NASH and is linked to the HFD-induced GM dysbiosis. These recent results confirm a previous study in mice genetically deficient for intestinal junctional adhesion molecule A (JAM-A), which developed more severe NAFLD and NASH upon HFD than the control mice<sup>71</sup>.

#### **TLR4/ NLRP3**

*In-vitro* and *in-vivo* studies have demonstrated that LPS increases intestinal permeability through TLR4/MyD88 pathways, and can be prevented in TLR4-KO mice<sup>72</sup> or using small interfering TLR4 silencing<sup>73</sup>. LPS activation increases TLR4 and CD14 intestinal expression<sup>73,74</sup>, as well as liver TLR4 expression<sup>67</sup>. Importantly, NLRP3 inflammasome, which plays a role in intestinal homeostasis **and** GM composition<sup>75</sup>, is involved in this pathway<sup>76,77</sup>. NLRP3 inflammasome-deficient mice display GM dysbiosis, increased portal TLR4 agonist, increased TNF $\alpha$  liver expression (a downstream inflammatory mediator of TLR pathway), which further leads to exacerbated liver alterations<sup>45</sup>. This phenotype can be recapitulated using co-housing experiments with NLRP3-deficient and wild-type mice<sup>45</sup>. Noteworthy, increased TNF $\alpha$  liver expression was confirmed in humans with NASH compared to those with simple steatosis<sup>69</sup>. Furthermore, NLRP3-deficient mice upon high-fructose diet develop more severe NASH than wild-type mice and up-regulation of genes involved in fatty acid uptake and de novo-lipogenesis. **Moreover, the worsened liverd phenotype observed in NLRP3-deficient mice upon high-fructose diet** is associated with increased intestinal permeability and bacterial translocation, lower levels of intestinal Angiogenin-4 (an anti-microbial peptide involved in intestine homeostasis) **and increased** liver TLR4 expression and macrophage infiltration<sup>78</sup>. The role of NLRP3-dependant GM dysbiosis was further confirmed since antibiotics treatment prevented liver damage. Overall



NAFLD pathogenesis involves GM dysbiosis, bacterial translocation, TLR4 activation leading to increased liver lipid uptake and lipogenesis as well as increased inflammation (**Figure 1**), suggesting that targeting these pathways through GM modulation could be beneficial to NAFLD development.

## Modulations of the gut microbiota to improve NAFLD

European clinical guidelines recommend lifestyle intervention<sup>79</sup> as the best therapeutic option for human NAFLD<sup>80</sup>. A randomized control trial (RCT)<sup>81</sup> has indeed demonstrated that  $\geq 7\%$  weight-loss significantly improves steatosis, lobular inflammation and ballooning, thus resulting in a significant decrease in NAFLD activity score (NAS<sup>82</sup>, a validated biopsy-based semi-quantitative system to evaluate NAFLD severity<sup>80</sup>). However, only 40% of the patients in the intensive arm achieved this goal and improved NAFLD at 48 months, demonstrating the complexity of reaching this target in everyday life<sup>81</sup>. Noteworthy, since diet impacts the GM composition<sup>10,44</sup>, weight-loss, through diet intervention thus associated with both quantitative and qualitative changes in food intake, has indeed been shown to induce GM modulation<sup>83</sup>. However, this is beyond the topic of this review, where we will focus only on new therapeutic interventions aiming at modulating the GM to improve NAFLD, without targeting weight loss.

### Probiotics

International expert consensus define probiotics, as "live microorganisms which when administered in adequate amounts confer host health benefits"<sup>84</sup>. Murine studies have accumulated large pieces of evidence regarding their beneficial effects on NAFLD and deciphered the underlying mechanistic pathways, reviewed in<sup>85</sup>. Herein, we have chosen to select only human studies with RCT design (displayed in **Table 2**), testing probiotics vs. placebo without the adjunction of a diet intervention, including mostly biopsy-proven NAFLD. Noteworthy, no intervention study performed a second liver biopsy after the intervention, thus limiting the final conclusion regarding the potential beneficial effects of probiotics on liver histology. RCT generally assessed steatosis changes using non-invasive tools, yet with

proven efficacy to evaluate liver fat evolution as an end-point in NASH clinical trials<sup>86</sup>. A small pilot study, using proton magnetic resonance spectroscopy found a major and significant reduction in intra-hepatic triglyceride content, 6 months after a probiotic cocktail<sup>87</sup>. In confirmation, another RCT randomized individuals with steatosis (defined by MRI-PDFF value $\geq$ 5.0) to a cocktail of 6 bacteria or placebo for 12 weeks and evaluated the changes in their MRI-assessed intra-hepatic fat<sup>88</sup>. Probiotics induced a significant yet small reduction in liver fat which differed from the placebo group for whom liver fat displayed a small non-significant increase. Importantly, the response displayed major variability among individuals within the probiotic group, with 40% of good responders, 47% of patients with stable evolution, while the remaining subjects had higher liver fat content<sup>88</sup>. The probiotic cocktail induced GM composition changes and a beneficial clinical response (i.e. the increase in *Agathobaculum*, *Dorea*, *Blautia*, *Ruminococcus* were associated with steatosis reduction). Similarly, a Chinese RCT where NAFLD patients received *Bifidobacterium*, *Lactobacillus* and *Enterococcus* capsules or placebo for 3 months, enabled a significant reduction in liver fat measured with ultrasound, concomitantly with improved liver enzymes and reduced endotoxemia<sup>89</sup> in the treatment arm. Likewise, a RCT performed in children submitted to a 4-months probiotic cocktail (i.e. VSL#3) displayed significant improvement in liver steatosis (measured by ultrasound)<sup>90</sup> in the treated groups as compared to a worsening in the placebo arm. In adults, a 3 months intervention with VSL#3 also resulted in improved liver enzymes, albeit this being an indirect NAFLD severity marker<sup>91</sup>. The mechanistic effects of VSL#3 have been investigated in murine models and involve improvement in insulin signaling, insulin-resistance and gut barrier permeability<sup>92</sup> as well as reduced liver collagen accumulation<sup>93</sup> or reduced lipid peroxidation in humans<sup>91</sup>. Other trials observed an improvement of indirect NAFLD-biomarkers after probiotic intervention, specifically *Lactobacillus bulgaricus* and *Streptococcus thermophilus* for 3 months significantly decreased ALT<sup>94</sup> whereas multiprobiotic “Symbiter” decreased FLI yet with major variability in individual responses<sup>95</sup>.

Overall, these RCTs show heterogeneity in length of treatment or in the type of probiotic cocktail composition or dosage. Despite that, a short duration probiotic cocktail

intervention is safe and appears able to slightly improve liver steatosis or at least prevent further worsening as compared to placebo. Nevertheless, most studies commonly describe einter-individual response variability, yet no predictor of good response has been studied and should probably be the focus of future research. Overall, steatosis improvement involves a series of pathways (Figure 1): modification of GM dysbiosis<sup>88,96</sup>, reduced endotoxemia<sup>89</sup> probably suggesting improved intestinal barrier function, minor yet significant reduction in BMI<sup>88,90,97</sup> (yet not with all the tested cocktail) and improvement in insulin-sensitivity parameters<sup>97</sup>. Importantly, short-term intervention trials do not seem to impact liver fibrosis evolution generally measured by transient elastography<sup>88</sup> in humans. Intervention studies are thus needed to evaluate whether longer-term probiotic supplementation could induce more important liver fat reduction and eventually translate in slowing down fibrosis evolution, which represents the critical prognosis parameter in NAFLD<sup>98,99</sup>. Interestingly, an on-going RCT will evaluate whether 24-weeks of a probiotic mix efficiently reduces liver fibrosis by transient elastography in NAFLD patients.<sup>100</sup>.

### Next-generation probiotics

The GM field rapidly evolves and has moved towards Next-Generation Probiotics (NGPs), defined as “live commensal microorganisms, identified upon comparative microbiota analyses, that when administered in adequate amounts, confer a host health benefit”<sup>101</sup>. NGPs are seen as disease specific. Their safety and effects need to be demonstrated and their mechanism of action understood<sup>102</sup>. Several candidates (some more advanced than others) are rising within the metabolic field<sup>102</sup> and should be further tested in NAFLD since they have shown beneficial effects in NAFLD-related mechanisms. They include *Akkermansia muciniphila*<sup>103</sup> with a possible beneficial role in glucose metabolism or insulin-resistance<sup>104–106</sup>, or *Christensenella minuta* and *Parabacteroides goldsteinii*. *C.minuta* abundance is lower in obese compared to lean individuals in independent cohorts<sup>107</sup>. FMT from an obese human donor into a recipient mice supplemented with *C.minuta* (10<sup>8</sup> *C.minuta* cells/day during 21 days) induced lower weight and adiposity than non-supplemented recipient, concomitant with modified GM composition, suggesting that *C.minuta* could induce weight-loss through gut modulation<sup>107</sup>. More translational research is warranted in particular

in the NAFLD field. Another example is *P.goldsteinii* which is increased after prebiotic supplementation in mice (i.e. water extract of Ganoderma lucidum mycelium=WEGL). In mice upon HFD, WEGL prevented weight gain, reduced liver inflammation and systemic endotoxemia and decreased the expression of genes involved in liver lipogenesis<sup>108,109</sup>. Therefore, *P.goldsteinii* was further tested as a NGP. Mice supplemented with *P. goldsteinii* ( $4 \times 10^7$  colony-forming units/day for 8 weeks), submitted to HFD display weight-loss, fat mass reduction, improved gut barrier integrity, insulin-resistance and inflammation<sup>110</sup>, all of which are involved in NAFLD pathophysiology. Therefore, testing whether *P.goldsteinii* could be beneficial in NAFLD improvement is now warranted.

*A.muciniphila* represents the most studied bacteria within the metabolic field with advanced translational research. *A.muciniphila* is a commensal bacteria involved in intestinal mucus remodeling thus playing a role in gut barrier integrity<sup>111</sup>. Several recent mice studies have tested the effects of different forms of *A.muciniphila* (live, pasteurized or through the infusion of its immunomodulatory outer-membrane protein 'Amuc\_1100')<sup>112</sup>. This probiotic enables reduction in body weight and fat mass, improvement in insulin-resistance and liver insulin signaling pathways as well as improved intestinal permeability and reduced endotoxemia<sup>112-114</sup>. Surprisingly, the pasteurized form, a safer galenic where bacteria is treated 30 min at 70 °C, thus limiting the denaturation of its cellular components, provides the stronger clinical benefit<sup>112</sup>. Live ( $10^{10}$  bacteria/day<sup>112</sup>) and pasteurized ( $10^{10}$  bacteria/day<sup>112</sup>) *A.muciniphila* were further safely tested during three months, in humans with obesity and T2D (thus individuals at risk of NAFLD, although not assessed at baseline) and recapitulated several beneficial effects observed in mice, but did not show any beneficial effect on insulin-sensitivity in the subjects treated with alive or pasteurized strains<sup>115</sup>. However, pasteurized *A.muciniphila* significantly reduced AST, GGT and endotoxemia as well as ALT (approaching significance)<sup>115</sup>. Noteworthy, Liraglutide, a GLP-1 agonist, tested in murine models of NAFLD, reduced hepatic fat content and liver inflammatory cell infiltration and amongst GM modification, associates with an 346%-fold increase in *A.muciniphila* and a 9% reduction in *Proteobacteria*<sup>116</sup>. Liraglutide also improved intestinal epithelium (increased number of goblet cells which has already been linked to *A.muciniphila*<sup>113</sup>), thus again suggesting the potential

beneficial role of this bacteria in the NAFLD context. Furthermore, obese individuals with metabolic syndrome who display low feces abundance of *A.muciniphila* have poorer liver status, as seen with altered liver enzyme, compared to healthy obese patients with high *A.muciniphila*<sup>18</sup>. Whether the individual abundance of *A.muciniphila* is important in the response to its therapeutic administration still needs to be explored. Finally, *A.muciniphila* abundance decreases with the severity of alcohol steatohepatitis (ASH) in humans and mice and its supplementation in mice both prevents the appearance of ASH as well as therapeutically improve pre-existing ASH and protect gut barrier integrity<sup>17</sup>. These pioneering results are in favor of testing this future NGP in NAFLD, in mouse models prior to transfer them to human clinical trials.

### Polyphenols

Polyphenols are plant-derived components found in fruit and vegetable that constitute a large group of bioactive phytochemicals with proven health benefits in some chronic non-communicable diseases<sup>18</sup>. While a certain percent is absorbed in the small intestine, a large amount is found in the colon where the GM process them into metabolites, potentially acting on the host<sup>19</sup>. Polyphenols also modulates GM thus further impacting host health<sup>20</sup>. Animal studies mostly, now suggests that different types of polyphenols can reverse or improve features of NAFLD, through GM modifications and modulation of the gut-liver axis. The most studied polyphenols in NAFLD treatment are flavonoids' compounds. For example, mice upon HFD submitted to high dose of raw bowl tea (containing 7 polyphenol compounds) reduce their body weight, liver steatosis and triglyceride content, liver enzyme and systemic inflammation to levels similar to that of control mice on chow diet<sup>21</sup>. These mouse data confirm previous reports in HFD-fed Zucker rats<sup>22</sup>. Furthermore, raw bowl tea reversed the HFD-induced small intestine alterations (namely, reduced immune cell infiltration within the lamina propria and increased tight-junction together with reduced endotoxemia, thus suggesting reduced intestinal permeability)<sup>21</sup>. Another murine study confirmed the benefice of polyphenols (found in green tea) on HFD-induced NAFLD histologic alterations and liver triglyceride content<sup>23</sup>. Green tea induced GM modifications with a major increase in Verrucomicrobia phylum among which *A.muciniphila*<sup>23</sup>, and a reduction in Proteobacteria<sup>23</sup>

(often found increased in NAFLD<sup>28–31</sup>). These observations also corroborate similar results observed with a tea polyphenol extract: epigallocatechin-3-gallate<sup>124</sup>. Green tea corrected HFD-induced altered bile acid profile, which was associated with some of the modified bacteria, suggesting that polyphenol improves NAFLD also through bile acid modulation. A 16-weeks Quercetin (i.e. a well-known flavonoid), supplementation also partly reverse HFD-induced NAFLD liver alterations (i.e. steatosis and ballooning), liver triglyceride and insulin-resistance<sup>125</sup>. Quercetin also induces GM modification, specifically, by decreasing HFD-induced increase in Proteobacteria and normalizes HFD-induced altered intestinal SCFA production, gut barrier and endotoxemia<sup>125</sup>. As discussed above, whereas HFD increased liver TLR4 gene expression and NLRP3 inflammasome<sup>45</sup>, Quercetin restored their expression to levels observed in controls<sup>125</sup>. Interestingly, HFD fed germ-free mice receiving FMT from mice upon HFD+quercetin were protected against liver alterations compared to those receiving FMT from mice solely upon HFD, suggesting that quercetin-induced GM modification is partially responsible for the beneficial liver clinical outcomes obtained after polyphenol supplementation<sup>126</sup>. Improved steatosis and gut barrier function, reduced endotoxemia and GM modulation were also obtained in a rat model upon HFD supplemented with 4-weeks of curcumin (i.e. a polyphenolic compound)<sup>127</sup>. Loquat fruit extract containing several polyphenols, also reduces steatosis, endotoxemia, improves gut barrier function and partially restores GM dysbiosis in rat fed a high-fructose diet<sup>128</sup>. Another fruit containing polyphenol (Red pitaya betacyanins) also improves HFD-induced NAFLD histologic liver alteration, insulin-resistance, systemic inflammation together with the increase in *A.muciniphila*<sup>129</sup>. Overall, these results highlight how several polyphenols can counteract NAFLD alterations via modulating the GM and the gut-liver axis albeit mostly in rodents (Figure 2). Whether these studies have translational significance in humans need serious investigations before their use can be recommended in patients. In humans with NAFLD, resveratrol has been the most investigated polyphenol, yet using indirect biomarkers of NAFLD severity rather than diagnostic criteria (i.e. liver enzymes). By contrast to data obtained in rodents, meta-analysis of human RCT using resveratrol in NAFLD failed to observe a positive effect<sup>130,131</sup>.

## Prebiotics

Prebiotics defined as a “substrate that is selectively utilized by host microorganisms conferring a health benefit”<sup>132</sup>, indeed modulates the GM both in mice<sup>66,105</sup> and humans<sup>37,133</sup>. In general, prebiotics improve mouse metabolic health by reducing weight, insulin-resistance, endotoxemia and improving gut barrier function<sup>134</sup>. Human studies are still controversial regarding their effects on metabolic health. While some show beneficial outcomes<sup>37</sup>, others do not observe any difference between the treated or placebo arm<sup>133</sup>, despite similar intervention duration, yet with different type of prebiotics possibly explaining the discrepant results. Prebiotics interventions in humans with NAFLD are still scarce, but some promising effects on pathways involved in NAFLD pathogenesis are already available in animal studies. *In-vivo* studies demonstrated that oligofructose (OFS)<sup>135</sup> (a prebiotic composed of nondigestible/fermentable fructo-oligosaccharides<sup>136</sup>) as well as inulin<sup>137</sup> (a fructan dietary fiber) decreased de novo-lipogenesis in HFD rats, by modulating the expression of lipogenic enzyme gene<sup>136</sup>. An 8-weeks fructan supplementation in Zucker rats translated in reduced steatosis both measured with NMR spectroscopy and histology concomitantly with reduced levels of portal propionate<sup>138</sup>. These results were confirmed with only 3-weeks inulin supplementation to HFD-fed rats<sup>137</sup>. Further *in-vitro* study demonstrated that reduced liver triglyceride content originated from decreased de novo-lipogenesis due to higher portal propionate levels<sup>138</sup>. Likewise, OFS supplementation for 3-weeks to rats fed a high-sucrose diet<sup>139</sup>, or for 4-weeks in rats fed a high-fructose diet<sup>140</sup>, translated in reduced liver weight and reduced FAS enzyme activity<sup>139</sup> thus suggesting reduced steatosis due to decreased lipogenesis<sup>139,140</sup>. Finally, WEGL, composed of polysaccharides, thus acting as a prebiotic, given to HFD mice enables major metabolic health improvement. Specifically, it reduces liver weight and steatosis as well as liver lipogenic gene expression, hepatic inflammation, endotoxemia and TLR4 signaling in the liver. Furthermore, it restores intestinal permeability and reverses HFD-induced GM dysbiosis. Moreover, FMT from WEGL treated mice into recipient mice upon HFD replicated the reduced liver weight, thus suggesting that the beneficial effects of this prebiotic originates from GM modulation<sup>108</sup>.

Turning to humans, several small studies have been performed. An 8-weeks RCT with OFS or placebo in 7 biopsy-proven NAFLD translated in a minor yet significant reduction in AST, **yet no** significant change in **ultrasound-measured** steatosis<sup>141</sup>. The absence of significant results could originate **either** from the small number of individuals **or** the heterogeneity of baseline liver lesions severity. By contrast, a RCT including 14 biopsy-proven NAFLD individuals randomized to a 9-months OFS or placebo intervention demonstrated a significant improvement in steatosis and inflammation, confirmed on follow-up biopsies. OFS induced changes in GM composition, specifically an increase in *Bifidobacterium*<sup>142</sup>, which has since been tested in most probiotic studies<sup>87–91,95</sup>. On-going RCTs including NAFLD patients (diagnosed upon ALT and Ultrasound) will evaluate the effects of a 6-months combined oligofructose+inulin associated with weight loss intervention on liver fibrosis (assessed by Fibroscan® and Fibrotest®) and on steatosis (measured by MRI)<sup>143</sup>. Prebiotics have proven efficient to improve metabolic diseases in mice (**Figure 2**), nevertheless few data have yet focused on specific NAFLD outcomes. More research is warranted specifically in humans to gain more insights into the importance of the effects and their underlying mechanisms.

### **Synbiotic**

Synbiotics, defined as a combination of both pro and prebiotics, recently emerged and some have been evaluated in NAFLD. **Although**, studies **are still** scarce and endpoints of published data are often indirect markers of NAFLD (liver enzymes or GGT<sup>144</sup>), **some** have quantified the change in steatosis or liver stiffness. Ultrasound-proven NAFLD patients randomized to a 24-weeks synbiotic cocktail of *Bifidobacterium animalis* and inulin or placebo, observed a significant reduction in **ultrasound-assessed** steatosis (with a major effect for individuals with the most severe grade), concomitant **with improved** liver enzymes<sup>145</sup>. Another 28-weeks RCT with placebo or a probiotic cocktail and OFS displayed significantly reduced ALT **and** liver fibrosis measured by transient elastography<sup>146</sup>. An on-going RCT will evaluate the effect of a 1-year treatment with placebo or synbiotic (OFS+*Bifidobacterium animalis* lactis) on liver fat content (assessed with MRI and NMR spectroscopy) and fibrosis (assessed by non-invasive algorithm and transient elastography)



in 104 biopsy-proven NAFLD individuals<sup>147</sup>. Overall, although promising, this field warrants larger studies in human with outcomes focusing on validated markers of NAFLD or using a second biopsy.

## FMT

FMT modulates the GM composition and function<sup>148</sup> and represents a novel therapeutic tool in NAFLD context. Firstly described for its ability to cure antibiotic-resistant *Clostridium Difficile* infection<sup>149</sup>, it is now widely tested in other diseases<sup>150</sup> including metabolic diseases<sup>151</sup>. FMT can also be safely performed using oral capsule instead of more invasive routes with similar beneficial effects at least in *Clostridium Difficile* infection<sup>152,153</sup>. Within the metabolic field, two studies performed in humans with metabolic syndrome showed that FMT from lean healthy donors significantly improved peripheral insulin-sensitivity, yet with major inter-individual responses<sup>154,155</sup>. Beneficial response was only observed in individuals with low MGR before the allogenic transfer, which significantly increased MGR six weeks after<sup>154</sup>. Yet, this beneficial effect is not prolonged in time<sup>154</sup> and suggests that repeated FMTs might be necessary in chronic diseases. No human studies have however observed any effect of FMT in weight reduction in overweight<sup>154,155</sup> or obese individuals<sup>156</sup>. In none of these studies, due to logistic reasons, stratification on GM profile was done prior to FMT.

Interestingly, allogenic FMT induced a significant yet minor change in fecal acetate level and fecal bile acid pool<sup>154,156,157</sup> and a reduced expression of inflammatory genes within the adipose tissue<sup>157</sup>. In line, metabolic characteristics of FMT donors (either post bariatric surgery or metabolic syndrome allogenic donors) also majorly affects insulin-sensitivity as well as bile acid metabolism<sup>157</sup>, both epathways being involved in NAFLD development but to date princeps FMT studies have only been performed in murine models. As such, FMT originating from mice fed a chow diet, into HFD-mice reduces liver triglyceride content and improves liver histological alterations as well as circulating liver enzymes, independently of insulin-resistance status. FMT also improves intestinal permeability as seen with increased gene and protein ZO-1 expression, potentially explaining the observed reduced systemic endotoxemia. FMT also partially corrects HFD-induced GM dysbiosis and increases fecal

butyrate concentration<sup>158</sup>. These first results observed in mice seem promising and warrant further confirmation in humans with NAFLD/NASH. Two studies, not yet recruiting, are registered on clinical trial (NCT03803540 and NCT02469272) to evaluate the potential benefits of FMT on liver histological alterations (NCT03803540) and the reduction of MRI-assessed steatosis (NCT02469272). FMT is now being tested in phase 1 trials in patients with end-stage liver disease and liver-related complications. In patients with cirrhosis and hepatic encephalopathy, enema<sup>159</sup> or oral capsulized<sup>160</sup> FMTs from healthy donors were well tolerated and accompanied by improvement in EncephalApp score<sup>159,160</sup> concomitant with a reduced inflammatory tone (i.e. reduced LBP)<sup>160</sup>. Although promising, further FMT studies are still needed in earlier stages of NAFLD to evaluate its efficacy on liver histological alterations and then test whether it could slow its progression.

### **Exercise:**

Physical activity is encouraged and prescribed to individuals with overweight/obesity<sup>161</sup> to improve cardiovascular diseases<sup>162</sup> and T2D<sup>163</sup>. An 8-week individualized training program, including combined endurance and strength exercise 3-5 times/week in patients with biopsy-proven NAFLD significantly improves NAFLD-related non-invasive biomarkers. Indeed, exercise lowers markers of systemic inflammation, steatosis (FLI), fibrosis-related parameters (FIB-4, and transient elastography) and liver enzymes displayed a 15%-fold reduction, suggesting the overall benefit of exercise in NAFLD patients<sup>164</sup>. Thus, both aerobic exercise and resistance training are now recommended in the latest European guidelines and its prescription should be personalized according to patients' profile<sup>80</sup>. Literature now shows that physical activity modulates the GM.

Indeed, endurance training drastically modifies GM composition and microbial-related circulating metabolites in healthy individuals running the half-marathon<sup>165</sup>. Whether such changes also occur in individuals with metabolic alteration still needs to be deciphered, yet this was assessed in several animal models and can therefore improve our understanding as to how physical activity could be beneficial for NAFLD through the microbiome. A recent study<sup>166</sup> performed in juvenile rats upon HFD, explored the effects of a 5-weeks combined aerobic and resistance training protocol on NAFLD evolution. Compared to the control group

(i.e. sedentary), the exercise group improved NAFLD (i.e. reduced triglyceride liver content and micro and macro-vesicular steatosis, both of which became similar to rats upon chow diet)<sup>166</sup>. These histological improvements were associated with decreased expression of genes involved in lipid metabolism (SREBP-1c, FAT/Cd36 and C/EBP $\alpha$ ) compared to the control group (HFD+sedentary)<sup>166</sup>. Going further, whereas HFD induced a well-known GM dysbiosis, the exercise corrected the microbiota imbalanced composition (i.e. the abundance of some bacteria went back to levels observed in sedentary chow-fed rats)<sup>166</sup>. This later result thus partly confirms results found in mice upon HFD or chow diet with or without exercise, where indeed exercise counteracted the HFD-induced GM dysbiosis for some specific bacteria. Nevertheless, the picture appears more complex, since for some other microbiome features, adding exercise to HFD subsequently shifted the microbiome to yet another state that even differs significantly from the controls fed a chow diet with or without exercise<sup>167</sup>. However, another mice study evaluating the effects of intensive interval training after a HFD found that exercise reversed some of the HFD-induced microbial alterations<sup>168</sup>. Specifically, exercise increased MGR in the most distal part of the intestine. This last finding was further confirmed in a human study, where 8-weeks training significantly increased MGR<sup>164</sup>. The changes in some phyla after the intervention correlated with the importance of clinical response<sup>164</sup>. More important than microbiota composition, exercise acts on several pathways involved in NAFLD pathophysiology. While HFD impairs GM functions, specifically related to metabolism and TCA cycle, exercise restores these genetic capacities to the level of control mice on chow diet<sup>168</sup>, possibly contributing to the improved metabolic alterations including NAFLD. Furthermore, exercise restores HFD-induced gut barrier dysfunction and altered mucosa alteration and reduces endotoxemia to levels observed in controls<sup>166</sup>. Likewise, mouse submitted to moderate swimming session, display maintained intestinal integrity, improved RRS-induced intestinal permeability and reduced bacterial translocation<sup>169</sup>. Exercise improves HFD-induced activation of the gut-liver axis, by decreasing TLR-4 (within the intestine and liver) and its subsequent inflammatory response in both organs<sup>166</sup>. Physical activity improves many features involved in NAFLD pathogenesis in mice (**Figure 2**), nevertheless, its effect on microbiota composition and function warrants further well-controlled studies and confirmation in humans with metabolic disease and

NAFLD. Finally, disentangling potential exercise-induced GM switch from that of dietary action is still needed..

## Diet

Specific diet interventions contribute to the treatment of NAFLD<sup>2</sup>, by modulating the gut-liver axis, with or without targeting weight-loss. Especially, Crete, Italian or Greek<sup>170–172</sup> Mediterranean diets (detailed composition<sup>170</sup> in **Table 3**) have demonstrated their ability to improve metabolic parameters and reduce weight. Furthermore, compared to caloric restriction, a 2-weeks low-carbohydrate diet intervention is more efficient in reducing excessive intrahepatic triglycerides, one of the hallmarks of NAFLD<sup>173</sup>. Therefore, studies have tested the effects of each or the combination of these approaches (Mediterranean diet and/or low-carbohydrate) on NAFLD. A 2-year Mediterranean or a low-carbohydrate diet intervention, in moderately obese subjects resulted in greater weight-loss and decreased ALT compared to a low-fat diet<sup>174</sup>. The association of Mediterranean diet and low-carbohydrate intake, decreased hepatic fat content and cardiometabolic risk parameters more than a low-fat diet<sup>175</sup>. Finally, a 6-months intervention with Mediterranean diet or its association with mid-day rest and exercise both resulted in more significant reduction in body weight and liver stiffness than the control group<sup>176</sup>. Interestingly, solely the combined exercise and Mediterranean diet group significantly decreased ALT<sup>176</sup>.

Some data are now available suggesting that high-adherence to the Mediterranean diet modulates the GM<sup>177–180</sup> towards a healthier state as seen with lower *Escherichia coli* counts<sup>179</sup>, higher bifidobacterial/*E. coli* ratio<sup>179</sup>, increased fecal SCFA<sup>177,179,180</sup> and increased *Prevotella*, also associated with a healthier status<sup>15</sup>. Several compounds of the Mediterranean diet such as whole grains and monounsaturated fatty acids (MUFA), which influence GM composition have been attributed to NAFLD improvement<sup>178</sup>. Other compounds are reviewed in detail elsewhere<sup>181</sup>. The beneficial effects of whole grains are generally related to specific phytochemicals (including fiber and polyphenols). They reduce energy intake and modulate GM (i.e. increase *Bifidobacteria* and *Lactobacilli*), both used in

several probiotic studies with proven beneficial effects on NAFLD<sup>87–91,95</sup>. Whole grains increase *Clostridium leptum*<sup>182–185</sup>, which is involved in butyrate production from fibers<sup>186</sup>. Therefore, one can hypothesize that whole grains improves NAFLD through GM-related butyrate production, which has been shown to decrease insulin-resistance and have anti-inflammatory effects in animal models<sup>187</sup>. In accordance, butyrate producing probiotics reduce NAFLD and endotoxemia in rats<sup>188</sup>. Furthermore, as discussed in the probiotic section, the administration of *Bifidobacteria* and *Lactobacilli* improves liver transaminases levels and histological lesions<sup>189</sup>. They also improve some NAFLD-features (i.e. improved glucose-induced insulin secretion, glucose tolerance and inflammatory status in mice<sup>190</sup>). Thus, proposing diet that enriches those bacteria in NAFLD patient needs evaluation, specifically on their effects on liver outcomes.

MUFA enrichment represent another characteristic of the Mediterranean diet. An enriched-MUFA diet is more efficient in reducing hepatic liver fat than a high-carbohydrate/high-fiber/low-glycemic index diet in subjects with TD2, thus at risk of NAFLD<sup>191</sup>. It has been hypothesized that a MUFA-enriched diet decreases hepatic steatosis through increased fat oxidation<sup>192</sup>, since plasma  $\beta$ -hydroxybutyrate, a surrogate marker for  $\beta$ -oxidation, was increased in the MUFA groups<sup>192</sup>. n MUFA supplementation during high-fat feeding also restored intestinal HFD-induced GM dysbiosis (i.e. significant decrease in *Enterobacteriales* and increase in *Bifidobacterium spp.*) in mice<sup>193</sup>. Nevertheless, available data on the effects of MUFA on the GM are still scarce and somewhat discrepant<sup>194</sup> thus warranting further studies in humans. Overall, the current literature suggests that interventions involving a high-degree adherence to a Mediterranean diet with lifestyle changes contribute to improved NAFLD outcomes<sup>80</sup> together with its well-known reduction in metabolic and cardiovascular risk. This might be due to caloric restriction, improvement of GM composition, higher fiber intake or a combination of the aforementioned factors. However, more research is needed to confirm the current findings in humans and optimize the effect of diet interventions targeting NAFLD treatment.

## Conclusions

Available NAFLD therapeutic options are scarce apart from efficient weight-loss, which is not easily achieved by all patients even in RCT, let alone in real life. Since GM is involved in NAFLD pathogenesis, targeting the GM to improve liver alterations seem an ideal alternative to weight management. We have reviewed several therapeutics or interventions acting on the GM with proven efficacy in improving NAFLD-related parameters. To date, the probiotic field is the most advanced with human RCT showing small yet significant improvement in NAFLD mostly assessed with non-invasive yet effective methods<sup>86</sup>. Murine studies have already deciphered mechanistic pathways to explain the observed beneficial outcomes. Most importantly, (single strain) probiotics seem at least able to prevent time-related NAFLD exacerbation. Nevertheless, longer-term studies are warranted to evaluate their efficacy on fibrosis, which represent the adverse liver alteration associated with NAFLD-related complication and progression. Regarding prebiotics, synbiotic, polyphenols or FMT, although data obtained *in-vitro* or *in-vivo* seem promising, there is a need for human translational research with harmonized and validated NAFLD methods to diagnose patients at inclusion. In the future, combining several of these options, as already tested with synbiotic, with specific lifestyle and diet intervention could prove even more efficient but should be formerly evaluated. Finally, since NAFLD pathophysiology involves multiple pathways with major inter-individual variability among patients, deciphering microbial-related or other biomarkers of response in these interventions is warranted to subsequently propose personalized approaches to modulate the gut microbiome in order to improve NAFLD.

**Table 1: Differences in sequencing methodologies: focus on pros and cons**

	<b>16S-pyrosequencing</b>	<b>Shotgun metagenomic sequencing</b>
<b>Method</b>	16S pyrosequencing approach amplifies and further sequences the highly conserved 16S-rDNA gene, present in every bacterium thus enabling the design of universal primer	Untargeted shotgun sequencing breaks the complete DNA material from a sample, into small and defined-size pieces that are subsequently sequenced
<b>Pros</b>		
<b>Use</b>	widely used	Increasingly used
<b>Costs</b>	Cheaper than shot gun	Costly yet with regular decrease

<b>Taxonomic Results</b>	Allows to examine the taxonomic composition of a sample	Allows to <b>examine the complete taxonomic composition</b> of a sample since the majority of the microbial genome is sequenced. It also assesses the abundance (after comparison to published catalogs)
<b>Functional result</b>	Can be obtained but <b>partial</b>	Allows <b>the determination of functional capacity</b> using functional databases
<b>Cons</b>		
<b>Linked to the method</b>	Limits linked to the polymerase chain reaction (PCR) and examines <b>only selected</b> microorganism for which primers were present	Limits linked to the need of complex bioinformatic pipeline availability for quality control and analyses.

**Table 2: RCT using probiotic to improve NAFLD**

<b>Probiotic</b>	<b>Patient's number</b>	<b>Design RCT</b>	<b>Duration</b>	<b>Effects</b>
<b>Lepicol probiotic formula:</b> <i>Lactobacillus plantarum</i> , <i>Lactobacillus delbrueckii</i> , <i>Lactobacillus acidophilus</i> , <i>Lactobacillus rhamnosus</i> and <i>Bifidobacterium bifidum</i>	20 Adults with biopsy-proven NAFLD	Vs. placebo	6 M	↓ intra-hepatic triglyceride content (proton-magnetic resonance spectroscopy) <sup>87</sup>
<i>L. acidophilus</i> CBT LA1, <i>L. rhamnosus</i> CBT LR5, <i>L. paracasei</i> CBT LPC5, <i>P. pentosaceus</i> CBT SL4, <i>B. lactis</i> CBT BL3 and <i>B. breve</i> CBT BR3	75 Adults with biopsy-proven NAFLD	Vs. placebo	12W	↓ intra-hepatic triglyceride content MRI-PDF <sup>88</sup>
<i>Bifidobacterium</i> , <i>Lactobacillus</i> and <i>Enterococcus</i>	120 Adults with biopsy-proven NAFLD	Vs. placebo	3M	↓ steatosis (US) ↓ liver enzyme ↓ endotoxemia <sup>89</sup>
VSL#3 = <i>Streptococcus thermophilus</i> , <i>bifidobacteria</i> [ <i>B. breve</i> , <i>B. infantis</i> , <i>B. longum</i> ], <i>Lactobacillus acidophilus</i> , <i>L. plantarum</i> , <i>L. paracasei</i> , and <i>L. delbrueckii</i> subsp. <i>Bulgaricus</i>	44 Children with biopsy-proven NAFLD	Vs. placebo	4M	↓ steatosis (US) <sup>90</sup>

VSL#3 = <i>Streptococcus thermophilus</i> , <i>bifidobacteria</i> [ <i>B. breve</i> , <i>B. infantis</i> , <i>B. longum</i> ], <i>Lactobacillus acidophilus</i> , <i>L. plantarum</i> , <i>L. paracasei</i> , and <i>L. delbrueckii subsp. Bulgaricus</i>	22 NAFLD Adults with biopsy-proven NAFLD	Vs. VSL#3 given to 56 patients with other liver disease	<b>3M</b>	↓ liver enzyme <sup>91</sup>
<i>Lactobacillus bulgaricus</i> and <i>Streptococcus thermophilus</i>	<b>30 adults</b> with biopsy-proven NAFLD	Vs. starch	<b>3M</b>	↓ liver enzyme <sup>94</sup>
Multiprobiotic "Symbiter" (concentrated biomass of 14 probiotic bacteria genera <i>Bifidobacterium</i> , <i>Lactobacillus</i> , <i>Lactococcus</i> , <i>Propionibacterium</i> )	<b>58 adults with T2D</b> with US-proven NAFLD	Vs. placebo	<b>8W</b>	↓ fatty liver index (FLI) ↓ liver enzyme <sup>95</sup>
<u>On-going RCT trial:</u> ( <i>Lactobacillus acidophilus</i> , <i>Bifidobacterium lactis</i> , <i>Lactobacillus rhamnosus</i> , <i>Lactobacillus paracasei</i> )	<b>46 adults</b> with/without biopsy-proven NAFLD	Vs. placebo	<b>24W</b>	Liver fibrosis ? 100



**Table 3: Mediterranean diet composition** <sup>170</sup>

High intake	H i g h intake	H i g h intake	Moderate intake	Moderate intake	Moderate intake	L o w intake	L o w intake
Vegetables	Cereals	C o l d pressed olive oils	Meat	Red wine	D a i r y product	Sweets	eggs
Legumes		Nuts	Fish				
Fruits							

### Legend

#### Figure 1: NAFLD pathophysiology: focus on the liver axis and the effects of probiotics

HFD or western diet induces gut microbiota dysbiosis and is associated with increased intestinal permeability (due to reduced intestinal tight junctions), which translates into translocation of bacteria and LPS leading to increased endotoxemia. LPS triggers TLR4, activates inflammatory pathways in the liver and increased lipogenesis which translates into NAFLD development and its exacerbation into NASH. Probiotics have proven beneficial to improve gut barrier function, reduce endotoxemia and reduce TLR4 activation thus leading to improved NAFLD features.

#### Figure 2: Effects of other therapeutic options modulating the gut microbiota and the gut liver axis to improve NAFLD

Mostly originating from murine studies, polyphenols, exercise and prebiotics improve some features of NAFLD via their action on the gut-liver axis. Polyphenols, prebiotics and exercise improve NAFLD through improvement of gut microbiota dysbiosis, restore the gut barrier function, decrease endotoxemia. Prebiotics and exercise also reduce liver lipogenesis.

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## References

1. Estes C, Razavi H, Loomba R, et al. Modeling the epidemic of nonalcoholic fatty liver disease demonstrates an exponential increase in burden of disease. *Hepatology* 2018;67:123-133.
2. European Association for the Study of the Liver (EASL), European Association for the Study of Diabetes (EASD), European Association for the Study of Obesity (EASO). EASL-EASD-EASO Clinical Practice Guidelines for the management of non-alcoholic fatty liver disease. *J Hepatol* 2016;64:1388-1402.
3. Barrera F, George J. The role of diet and nutritional intervention for the management of patients with NAFLD. *Clin Liver Dis* 2014;18:91-112.
4. Day CP, James OF. Steatohepatitis: a tale of two "hits"? *Gastroenterology* 1998;114:842-845.
5. Neuschwander-Tetri BA. Hepatic lipotoxicity and the pathogenesis of nonalcoholic steatohepatitis: the central role of nontriglyceride fatty acid metabolites. *Hepatology* 2010;52:774-788.
6. Bäckhed F, Ding H, Wang T, et al. The gut microbiota as an environmental factor that regulates fat storage. *Proc Natl Acad Sci USA* 2004;101:15718-15723.
7. Ridaura VK, Faith JJ, Rey FE, et al. Gut microbiota from twins discordant for obesity modulate metabolism in mice. *Science* 2013;341:1241-1244.
8. De Filippo C, Cavalieri D, Di Paola M, et al. Impact of diet in shaping gut microbiota revealed by a comparative study in children from Europe and rural Africa. *Proc Natl Acad Sci USA* 2010;107:14691-14696.
9. Wu GD, Chen J, Hoffmann C, et al. Linking long-term dietary patterns with gut microbial enterotypes. *Science* 2011;334:105-108.
10. Falony G, Joossens M, Vieira-Silva S, et al. Population-level analysis of gut microbiome variation. *Science* 2016;352:560-564.
11. Cotillard A, Kennedy SP, Kong LC, et al. Dietary intervention impact on gut microbial gene richness. *Nature* 2013;500:585-588.
12. Thomas V, Clark J, Doré J. Fecal microbiota analysis: an overview of sample collection methods and sequencing strategies. *Future Microbiol* 2015;10:1485-1504.
13. Le Chatelier E, Nielsen T, Qin J, et al. Richness of human gut microbiome correlates with metabolic markers. *Nature* 2013;500:541-546.
14. Aron-Wisnewsky J, Prifti E, Belda E, et al. Major microbiota dysbiosis in severe obesity: fate after bariatric surgery. *Gut* 2018.

15. Vandeputte D, Kathagen G, D'hoë K, et al. Quantitative microbiome profiling links gut community variation to microbial load. *Nature* 2017;551:507-511.
16. Marjot T, Moolla A, Cobbold JF, et al. Non-alcoholic fatty liver disease in adults: Current concepts in etiology, outcomes and management. *Endocr Rev* 2019.
17. Zhang H, DiBaise JK, Zuccolo A, et al. Human gut microbiota in obesity and after gastric bypass. *Proc Natl Acad Sci USA* 2009;106:2365-2370.
18. Dao MC, Everard A, Aron-Wisnewsky J, et al. *Akkermansia muciniphila* and improved metabolic health during a dietary intervention in obesity: relationship with gut microbiome richness and ecology. *Gut* 2015.
19. Allin KH, Tremaroli V, Caesar R, et al. Aberrant intestinal microbiota in individuals with prediabetes. *Diabetologia* 2018;61:810-820.
20. Kayser BD, Prifti E, Lhomme M, et al. Elevated serum ceramides are linked with obesity-associated gut dysbiosis and impaired glucose metabolism. *Metabolomics* 2019;15:140.
21. Karlsson FH, Tremaroli V, Nookaew I, et al. Gut metagenome in European women with normal, impaired and diabetic glucose control. *Nature* 2013;498:99-103.
22. Qin J, Li Y, Cai Z, et al. A metagenome-wide association study of gut microbiota in type 2 diabetes. *Nature* 2012;490:55-60.
23. Pedersen HK, Gudmundsdottir V, Nielsen HB, et al. Human gut microbes impact host serum metabolome and insulin sensitivity. *Nature* 2016;535:376-381.
24. Koh A, Molinaro A, Ståhlman M, et al. Microbially Produced Imidazole Propionate Impairs Insulin Signaling through mTORC1. *Cell* 2018;175:947-961.e17.
25. Yki-Järvinen H. Non-alcoholic fatty liver disease as a cause and a consequence of metabolic syndrome. *Lancet Diabetes Endocrinol* 2014;2:901-910.
26. Brandl K, Schnabl B. Intestinal microbiota and nonalcoholic steatohepatitis. *Curr Opin Gastroenterol* 2017;33:128-133.
27. Tilg H, Zmora N, Adolph TE, et al. The intestinal microbiota fuelling metabolic inflammation. *Nat Rev Immunol* 2019.
28. Hoyles L, Fernández-Real J-M, Federici M, et al. Molecular phenomics and metagenomics of hepatic steatosis in non-diabetic obese women. *Nat Med* 2018;24:1070-1080.
29. Shen F, Zheng R-D, Sun X-Q, et al. Gut microbiota dysbiosis in patients with non-alcoholic fatty liver disease. *HBPD INT* 2017;16:375-381.
30. Raman M, Ahmed I, Gillevet PM, et al. Fecal microbiome and volatile organic compound metabolome in obese humans with nonalcoholic fatty liver disease. *Clin Gastroenterol Hepatol* 2013;11:868-875.e1-3.
31. Loomba R, Seguritan V, Li W, et al. Gut Microbiome-Based Metagenomic Signature for Non-invasive Detection of Advanced Fibrosis in Human Nonalcoholic Fatty Liver Disease. *Cell Metab* 2017;25:1054-1062.e5.

32. Zhu L, Baker SS, Gill C, et al. Characterization of gut microbiomes in nonalcoholic steatohepatitis (NASH) patients: a connection between endogenous alcohol and NASH. *Hepatology* 2013;57:601-609.
33. Wang B, Jiang X, Cao M, et al. Altered Fecal Microbiota Correlates with Liver Biochemistry in Nonobese Patients with Non-alcoholic Fatty Liver Disease. *Sci Rep* 2016;6:32002.
34. Del Chierico F, Nobili V, Vernocchi P, et al. Gut microbiota profiling of pediatric nonalcoholic fatty liver disease and obese patients unveiled by an integrated meta-omics-based approach. *Hepatology* 2017;65:451-464.
35. Wong VW-S, Tse C-H, Lam TT-Y, et al. Molecular characterization of the fecal microbiota in patients with nonalcoholic steatohepatitis--a longitudinal study. *PLoS ONE* 2013;8:e62885.
36. Boursier J, Mueller O, Barret M, et al. The severity of nonalcoholic fatty liver disease is associated with gut dysbiosis and shift in the metabolic function of the gut microbiota. *Hepatology* 2016;63:764-775.
37. Dewulf EM, Cani PD, Claus SP, et al. Insight into the prebiotic concept: lessons from an exploratory, double blind intervention study with inulin-type fructans in obese women. *Gut* 2013;62:1112-1121.
38. Caussy C, Tripathi A, Humphrey G, et al. A gut microbiome signature for cirrhosis due to nonalcoholic fatty liver disease. *Nat Commun* 2019;10:1406.
39. Qin N, Yang F, Li A, et al. Alterations of the human gut microbiome in liver cirrhosis. *Nature* 2014;513:59-64.
40. Mouzaki M, Comelli EM, Arendt BM, et al. Intestinal microbiota in patients with nonalcoholic fatty liver disease. *Hepatology* 2013;58:120-127.
41. Michail S, Lin M, Frey MR, et al. Altered gut microbial energy and metabolism in children with non-alcoholic fatty liver disease. *FEMS Microbiol Ecol* 2015;91:1-9.
42. Da Silva HE, Teterina A, Comelli EM, et al. Nonalcoholic fatty liver disease is associated with dysbiosis independent of body mass index and insulin resistance. *Sci Rep* 2018;8:1466.
43. Caussy C, Soni M, Cui J, et al. Nonalcoholic fatty liver disease with cirrhosis increases familial risk for advanced fibrosis. *J Clin Invest* 2017;127:2697-2704.
44. Rothschild D, Weissbrod O, Barkan E, et al. Environment dominates over host genetics in shaping human gut microbiota. *Nature* 2018;555:210-215.
45. Henaoui-Mejia J, Elinav E, Jin C, et al. Inflammasome-mediated dysbiosis regulates progression of NAFLD and obesity. *Nature* 2012;482:179-185.
46. Le Roy T, Llopis M, Lepage P, et al. Intestinal microbiota determines development of non-alcoholic fatty liver disease in mice. *Gut* 2013;62:1787-1794.
47. Chiu C-C, Ching Y-H, Li Y-P, et al. Nonalcoholic Fatty Liver Disease Is Exacerbated in High-Fat Diet-Fed Gnotobiotic Mice by Colonization with the Gut Microbiota from Patients with Nonalcoholic Steatohepatitis. *Nutrients* 2017;9.
48. Arab JP, Arrese M, Trauner M. Recent Insights into the Pathogenesis of Nonalcoholic Fatty Liver Disease. *Annu Rev Pathol* 2018;13:321-350.

49. Mouzaki M, Loomba R. Insights into the evolving role of the gut microbiome in nonalcoholic fatty liver disease: rationale and prospects for therapeutic intervention. *Therap Adv Gastroenterol* 2019;12:1756284819858470.
50. Tripathi A, Debelius J, Brenner DA, et al. The gut-liver axis and the intersection with the microbiome. *Nat Rev Gastroenterol Hepatol* 2018;15:397-411.
51. Aron-Wisnewsky J, Gaborit B, Dutour A, et al. Gut microbiota and non-alcoholic fatty liver disease: new insights. *Clin Microbiol Infect* 2013;19:338-348.
52. Kolodziejczyk AA, Zheng D, Shibolet O, et al. The role of the microbiome in NAFLD and NASH. *EMBO Mol Med* 2019;11.
53. Leung C, Rivera L, Furness JB, et al. The role of the gut microbiota in NAFLD. *Nat Rev Gastroenterol Hepatol* 2016;13:412-425.
54. Anjani K, Lhomme M, Sokolovska N, et al. Circulating phospholipid profiling identifies portal contribution to NASH signature in obesity. *J Hepatol* 2015;62:905-912.
55. Chávez-Talavera O, Tailleux A, Lefebvre P, et al. Bile Acid Control of Metabolism and Inflammation in Obesity, Type 2 Diabetes, Dyslipidemia, and Nonalcoholic Fatty Liver Disease. *Gastroenterology* 2017;152:1679-1694.e3.
56. Ji Y, Yin Y, Li Z, et al. Gut Microbiota-Derived Components and Metabolites in the Progression of Non-Alcoholic Fatty Liver Disease (NAFLD). *Nutrients* 2019;11.
57. Canfora EE, Meex RCR, Venema K, et al. Gut microbial metabolites in obesity, NAFLD and T2DM. *Nat Rev Endocrinol* 2019;15:261-273.
58. Dumas M-E, Barton RH, Toye A, et al. Metabolic profiling reveals a contribution of gut microbiota to fatty liver phenotype in insulin-resistant mice. *Proc Natl Acad Sci USA* 2006;103:12511-12516.
59. Yuan J, Chen C, Cui J, et al. Fatty Liver Disease Caused by High-Alcohol-Producing *Klebsiella pneumoniae*. *Cell Metab* 2019;30:675-688.e7.
60. Volynets V, Küper MA, Strahl S, et al. Nutrition, intestinal permeability, and blood ethanol levels are altered in patients with nonalcoholic fatty liver disease (NAFLD). *Dig Dis Sci* 2012;57:1932-1941.
61. Caussy C, Hsu C, Lo M-T, et al. Link between gut-microbiome derived metabolite and shared gene-effects with hepatic steatosis and fibrosis in NAFLD. *Hepatology* 2018.
62. Luther J, Garber JJ, Khalili H, et al. Hepatic Injury in Nonalcoholic Steatohepatitis Contributes to Altered Intestinal Permeability. *Cell Mol Gastroenterol Hepatol* 2015;1:222-232.
63. Giorgio V, Miele L, Principessa L, et al. Intestinal permeability is increased in children with non-alcoholic fatty liver disease, and correlates with liver disease severity. *Dig Liver Dis* 2014;46:556-560.
64. Miele L, Valenza V, La Torre G, et al. Increased intestinal permeability and tight junction alterations in nonalcoholic fatty liver disease. *Hepatology* 2009;49:1877-1887.
65. Cani PD, Amar J, Iglesias MA, et al. Metabolic endotoxemia initiates obesity and insulin resistance. *Diabetes* 2007;56:1761-1772.

66. Cani PD, Possemiers S, Van de Wiele T, et al. Changes in gut microbiota control inflammation in obese mice through a mechanism involving GLP-2-driven improvement of gut permeability. *Gut* 2009;58:1091-1103.
67. Sharifnia T, Antoun J, Verriere TGC, et al. Hepatic TLR4 signaling in obese NAFLD. *Am J Physiol Gastrointest Liver Physiol* 2015;309:G270-278.
68. Harte AL, Silva NF da, Creely SJ, et al. Elevated endotoxin levels in non-alcoholic fatty liver disease. *J Inflamm (Lond)* 2010;7:15.
69. Ruiz AG, Casafont F, Crespo J, et al. Lipopolysaccharide-binding protein plasma levels and liver TNF-alpha gene expression in obese patients: evidence for the potential role of endotoxin in the pathogenesis of non-alcoholic steatohepatitis. *Obes Surg* 2007;17:1374-1380.
70. Mouries J, Brescia P, Silvestri A, et al. Microbiota-driven gut vascular barrier disruption is a prerequisite for non-alcoholic steatohepatitis development. *J Hepatol* 2019.
71. Rahman K, Desai C, Iyer SS, et al. Loss of Junctional Adhesion Molecule A Promotes Severe Steatohepatitis in Mice on a Diet High in Saturated Fat, Fructose, and Cholesterol. *Gastroenterology* 2016;151:733-746.e12.
72. Nighot M, Al-Sadi R, Guo S, et al. Lipopolysaccharide-Induced Increase in Intestinal Epithelial Tight Permeability Is Mediated by Toll-Like Receptor 4/Myeloid Differentiation Primary Response 88 (MyD88) Activation of Myosin Light Chain Kinase Expression. *Am J Pathol* 2017;187:2698-2710.
73. Guo S, Nighot M, Al-Sadi R, et al. Lipopolysaccharide Regulation of Intestinal Tight Junction Permeability Is Mediated by TLR4 Signal Transduction Pathway Activation of FAK and MyD88. *J Immunol* 2015;195:4999-5010.
74. Guo S, Al-Sadi R, Said HM, et al. Lipopolysaccharide causes an increase in intestinal tight junction permeability in vitro and in vivo by inducing enterocyte membrane expression and localization of TLR-4 and CD14. *Am J Pathol* 2013;182:375-387.
75. Elinav E, Henao-Mejia J, Flavell RA. Integrative inflammasome activity in the regulation of intestinal mucosal immune responses. *Mucosal Immunol* 2013;6:4-13.
76. Tsutsui H, Imamura M, Fujimoto J, et al. The TLR4/TRIF-Mediated Activation of NLRP3 Inflammasome Underlies Endotoxin-Induced Liver Injury in Mice. *Gastroenterol Res Pract* 2010;2010:641865.
77. Mehal WZ. The Gordian Knot of dysbiosis, obesity and NAFLD. *Nat Rev Gastroenterol Hepatol* 2013;10:637-644.
78. Pierantonelli I, Rychlicki C, Agostinelli L, et al. Lack of NLRP3-inflammasome leads to gut-liver axis derangement, gut dysbiosis and a worsened phenotype in a mouse model of NAFLD. *Sci Rep* 2017;7:12200.
79. El-Agroudy NN, Kurzbach A, Rodionov RN, et al. Are Lifestyle Therapies Effective for NAFLD Treatment? *Trends Endocrinol Metab* 2019;30:701-709.
80. European Association for the Study of the Liver (EASL), European Association for the Study of Diabetes (EASD), European Association for the Study of Obesity (EASO). EASL-EASD-EASO Clinical Practice Guidelines for the management of non-alcoholic fatty liver disease. *Diabetologia* 2016;59:1121-1140.

81. Promrat K, Kleiner DE, Niemeier HM, et al. Randomized controlled trial testing the effects of weight loss on nonalcoholic steatohepatitis. *Hepatology* 2010;51:121-129.
82. Younossi ZM, Koenig AB, Abdelatif D, et al. Global Epidemiology of Non-Alcoholic Fatty Liver Disease-Meta-Analytic Assessment of Prevalence, Incidence and Outcomes. *Hepatology* 2015.
83. Seganfredo FB, Blume CA, Moehlecke M, et al. Weight-loss interventions and gut microbiota changes in overweight and obese patients: a systematic review. *Obes Rev* 2017;18:832-851.
84. Hill C, Guarner F, Reid G, et al. Expert consensus document. The International Scientific Association for Probiotics and Prebiotics consensus statement on the scope and appropriate use of the term probiotic. *Nat Rev Gastroenterol Hepatol* 2014;11:506-514.
85. Porras D, Nistal E, Martínez-Flórez S, et al. Intestinal Microbiota Modulation in Obesity-Related Non-alcoholic Fatty Liver Disease. *Front Physiol* 2018;9:1813.
86. Caussy C, Reeder SB, Sirlin CB, et al. Noninvasive, Quantitative Assessment of Liver Fat by MRI-PDFF as an Endpoint in NASH Trials. *Hepatology* 2018;68:763-772.
87. Wong VW-S, Won GL-H, Chim AM-L, et al. Treatment of nonalcoholic steatohepatitis with probiotics. A proof-of-concept study. *Ann Hepatol* 2013;12:256-262.
88. Ahn SB, Jun DW, Kang B-K, et al. Randomized, Double-blind, Placebo-controlled Study of a Multispecies Probiotic Mixture in Nonalcoholic Fatty Liver Disease. *Sci Rep* 2019;9:5688.
89. Anon. Clinical study of probiotics in treatment of non-alcoholic fatty liver disease--《Chinese Journal of Gastroenterology and Hepatology》2013年03期. Available at: [http://en.cnki.com.cn/Article\\_en/CJFDTotat-WCBX201303009.htm](http://en.cnki.com.cn/Article_en/CJFDTotat-WCBX201303009.htm) [Accessed October 16, 2019].
90. Alisi A, Bedogni G, Baviera G, et al. Randomised clinical trial: The beneficial effects of VSL#3 in obese children with non-alcoholic steatohepatitis. *Aliment Pharmacol Ther* 2014;39:1276-1285.
91. Loguercio C, Federico A, Tuccillo C, et al. Beneficial effects of a probiotic VSL#3 on parameters of liver dysfunction in chronic liver diseases. *J Clin Gastroenterol* 2005;39:540-543.
92. Mencarelli A, Cipriani S, Renga B, et al. VSL#3 resets insulin signaling and protects against NASH and atherosclerosis in a model of genetic dyslipidemia and intestinal inflammation. *PLoS ONE* 2012;7:e45425.
93. Velayudham A, Dolganiuc A, Ellis M, et al. VSL#3 probiotic treatment attenuates fibrosis without changes in steatohepatitis in a diet-induced nonalcoholic steatohepatitis model in mice. *Hepatology* 2009;49:989-997.
94. Aller R, De Luis DA, Izaola O, et al. Effect of a probiotic on liver aminotransferases in nonalcoholic fatty liver disease patients: a double blind randomized clinical trial. *Eur Rev Med Pharmacol Sci* 2011;15:1090-1095.
95. Kobyliak N, Abenavoli L, Mykhalchyshyn G, et al. A Multi-strain Probiotic Reduces the Fatty Liver Index, Cytokines and Aminotransferase levels in NAFLD Patients: Evidence from a Randomized Clinical Trial. *J Gastrointestin Liver Dis* 2018;27:41-49.

96. Yadav H, Lee J-H, Lloyd J, et al. Beneficial metabolic effects of a probiotic via butyrate-induced GLP-1 hormone secretion. *J Biol Chem* 2013;288:25088-25097.
97. Koutnikova H, Genser B, Monteiro-Sepulveda M, et al. Impact of bacterial probiotics on obesity, diabetes and non-alcoholic fatty liver disease related variables: a systematic review and meta-analysis of randomised controlled trials. *BMJ Open* 2019;9:e017995.
98. Pais R, Charlotte F, Fedchuk L, et al. A systematic review of follow-up biopsies reveals disease progression in patients with non-alcoholic fatty liver. *J Hepatol* 2013;59:550-556.
99. Hagström H, Nasr P, Ekstedt M, et al. SAF score and mortality in NAFLD after up to 41 years of follow-up. *Scand J Gastroenterol* 2017;52:87-91.
100. Silva-Sperb AS, Moraes HA, Moura BC de, et al. Effect of probiotic supplementation in nonalcoholic steatohepatitis patients: PROBILIVER TRIAL protocol. *Trials* 2019;20:580.
101. Martín R, Langella P. Emerging Health Concepts in the Probiotics Field: Streamlining the Definitions. *Front Microbiol* 2019;10:1047.
102. Chang C-J, Lin T-L, Tsai Y-L, et al. Next generation probiotics in disease amelioration. *J Food Drug Anal* 2019;27:615-622.
103. Cani PD, Vos WM de. Next-Generation Beneficial Microbes: The Case of *Akkermansia muciniphila*. *Front Microbiol* 2017;8:1765.
104. Everard A, Lazarevic V, Derrien M, et al. Responses of gut microbiota and glucose and lipid metabolism to prebiotics in genetic obese and diet-induced leptin-resistant mice. *Diabetes* 2011;60:2775-2786.
105. Everard A, Lazarevic V, Gaïa N, et al. Microbiome of prebiotic-treated mice reveals novel targets involved in host response during obesity. *ISME J* 2014;8:2116-2130.
106. Everard A, Geurts L, Caesar R, et al. Intestinal epithelial MyD88 is a sensor switching host metabolism towards obesity according to nutritional status. *Nat Commun* 2014;5:5648.
107. Goodrich JK, Waters JL, Poole AC, et al. Human genetics shape the gut microbiome. *Cell* 2014;159:789-799.
108. Chang C-J, Lin C-S, Lu C-C, et al. *Ganoderma lucidum* reduces obesity in mice by modulating the composition of the gut microbiota. *Nat Commun* 2015;6:7489.
109. Chang C-J, Lin C-S, Lu C-C, et al. Corrigendum: *Ganoderma lucidum* reduces obesity in mice by modulating the composition of the gut microbiota. *Nat Commun* 2017;8:16130.
110. Wu T-R, Lin C-S, Chang C-J, et al. Gut commensal *Parabacteroides goldsteinii* plays a predominant role in the anti-obesity effects of polysaccharides isolated from *Hirsutella sinensis*. *Gut* 2019;68:248-262.
111. Derrien M, Vaughan EE, Plugge CM, et al. *Akkermansia muciniphila* gen. nov., sp. nov., a human intestinal mucin-degrading bacterium. *Int J Syst Evol Microbiol* 2004;54:1469-1476.



112. Plovier H, Everard A, Druart C, et al. A purified membrane protein from *Akkermansia muciniphila* or the pasteurized bacterium improves metabolism in obese and diabetic mice. *Nat Med* 2017;23:107-113.
113. Everard A, Belzer C, Geurts L, et al. Cross-talk between *Akkermansia muciniphila* and intestinal epithelium controls diet-induced obesity. *Proc Natl Acad Sci USA* 2013;110:9066-9071.
114. Schneeberger M, Everard A, Gómez-Valadés AG, et al. *Akkermansia muciniphila* inversely correlates with the onset of inflammation, altered adipose tissue metabolism and metabolic disorders during obesity in mice. *Sci Rep* 2015;5:16643.
115. Depommier C, Everard A, Druart C, et al. Supplementation with *Akkermansia muciniphila* in overweight and obese human volunteers: a proof-of-concept exploratory study. *Nat Med* 2019;25:1096-1103.
116. Moreira GV, Azevedo FF, Ribeiro LM, et al. Liraglutide modulates gut microbiota and reduces NAFLD in obese mice. *J Nutr Biochem* 2018;62:143-154.
117. Grander C, Adolph TE, Wieser V, et al. Recovery of ethanol-induced *Akkermansia muciniphila* depletion ameliorates alcoholic liver disease. *Gut* 2018;67:891-901.
118. Fraga CG, Croft KD, Kennedy DO, et al. The effects of polyphenols and other bioactives on human health. *Food Funct* 2019;10:514-528.
119. Braune A, Blaut M. Bacterial species involved in the conversion of dietary flavonoids in the human gut. *Gut Microbes* 2016;7:216-234.
120. Selma MV, Espín JC, Tomás-Barberán FA. Interaction between phenolics and gut microbiota: role in human health. *J Agric Food Chem* 2009;57:6485-6501.
121. Liu B, Zhang J, Sun P, et al. Raw Bowl Tea (Tuocha) Polyphenol Prevention of Nonalcoholic Fatty Liver Disease by Regulating Intestinal Function in Mice. *Biomolecules* 2019;9.
122. Tan Y, Kim J, Cheng J, et al. Green tea polyphenols ameliorate non-alcoholic fatty liver disease through upregulating AMPK activation in high fat fed Zucker fatty rats. *World J Gastroenterol* 2017;23:3805-3814.
123. Ushiroda C, Naito Y, Takagi T, et al. Green tea polyphenol (epigallocatechin-3-gallate) improves gut dysbiosis and serum bile acids dysregulation in high-fat diet-fed mice. *J Clin Biochem Nutr* 2019;65:34-46.
124. Sheng L, Jena PK, Liu H-X, et al. Obesity treatment by epigallocatechin-3-gallate-regulated bile acid signaling and its enriched *Akkermansia muciniphila*. *FASEB J* 2018:fj201800370R.
125. Porrás D, Nistal E, Martínez-Flórez S, et al. Protective effect of quercetin on high-fat diet-induced non-alcoholic fatty liver disease in mice is mediated by modulating intestinal microbiota imbalance and related gut-liver axis activation. *Free Radic Biol Med* 2017;102:188-202.
126. Porrás D, Nistal E, Martínez-Flórez S, et al. Functional Interactions between Gut Microbiota Transplantation, Quercetin, and High-Fat Diet Determine Non-Alcoholic Fatty Liver Disease Development in Germ-Free Mice. *Mol Nutr Food Res* 2019;63:e1800930.

127. Feng W, Wang H, Zhang P, et al. Modulation of gut microbiota contributes to curcumin-mediated attenuation of hepatic steatosis in rats. *Biochim Biophys Acta Gen Subj* 2017;1861:1801-1812.
128. Li W, Yang H, Zhao Q, et al. Polyphenol-Rich Loquat Fruit Extract Prevents Fructose-Induced Nonalcoholic Fatty Liver Disease by Modulating Glycometabolism, Lipometabolism, Oxidative Stress, Inflammation, Intestinal Barrier, and Gut Microbiota in Mice. *J Agric Food Chem* 2019;67:7726-7737.
129. Song H, Chu Q, Yan F, et al. Red pitaya betacyanins protects from diet-induced obesity, liver steatosis and insulin resistance in association with modulation of gut microbiota in mice. *J Gastroenterol Hepatol* 2016;31:1462-1469.
130. Zhang C, Yuan W, Fang J, et al. Efficacy of Resveratrol Supplementation against Non-Alcoholic Fatty Liver Disease: A Meta-Analysis of Placebo-Controlled Clinical Trials. *PLoS ONE* 2016;11:e0161792.
131. Elgebaly A, Radwan IAI, AboElnas MM, et al. Resveratrol Supplementation in Patients with Non-Alcoholic Fatty Liver Disease: Systematic Review and Meta-analysis. *J Gastrointestin Liver Dis* 2017;26:59-67.
132. Gibson GR, Hutkins R, Sanders ME, et al. Expert consensus document: The International Scientific Association for Probiotics and Prebiotics (ISAPP) consensus statement on the definition and scope of prebiotics. *Nat Rev Gastroenterol Hepatol* 2017;14:491-502.
133. Canfora EE, Beek CM van der, Hermes GDA, et al. Supplementation of Diet With Galacto-oligosaccharides Increases Bifidobacteria, but Not Insulin Sensitivity, in Obese Prediabetic Individuals. *Gastroenterology* 2017;153:87-97.e3.
134. Delzenne NM, Neyrinck AM, Cani PD. Modulation of the gut microbiota by nutrients with prebiotic properties: consequences for host health in the context of obesity and metabolic syndrome. *Microb Cell Fact* 2011;10 Suppl 1:S10.
135. Kok N, Roberfroid M, Robert A, et al. Involvement of lipogenesis in the lower VLDL secretion induced by oligofructose in rats. *Br J Nutr* 1996;76:881-890.
136. Delzenne NM, Kok NN. Biochemical basis of oligofructose-induced hypolipidemia in animal models. *J Nutr* 1999;129:1467S-70S.
137. Sugatani J, Osabe M, Wada T, et al. Comparison of enzymatically synthesized inulin, resistant maltodextrin and clofibrate effects on biomarkers of metabolic disease in rats fed a high-fat and high-sucrose (cafeteria) diet. *Eur J Nutr* 2008;47:192-200.
138. Daubioul C, Rousseau N, Demeure R, et al. Dietary fructans, but not cellulose, decrease triglyceride accumulation in the liver of obese Zucker fa/fa rats. *J Nutr* 2002;132:967-973.
139. Agheli N, Kabir M, Berni-Canani S, et al. Plasma lipids and fatty acid synthase activity are regulated by short-chain fructo-oligosaccharides in sucrose-fed insulin-resistant rats. *J Nutr* 1998;128:1283-1288.
140. Busserolles J, Gueux E, Rock E, et al. Oligofructose protects against the hypertriglyceridemic and pro-oxidative effects of a high fructose diet in rats. *J Nutr* 2003;133:1903-1908.

141. Daubioul CA, Horsmans Y, Lambert P, et al. Effects of oligofructose on glucose and lipid metabolism in patients with nonalcoholic steatohepatitis: results of a pilot study. *Eur J Clin Nutr* 2005;59:723-726.
142. Bomhof MR, Parnell JA, Ramay HR, et al. Histological improvement of non-alcoholic steatohepatitis with a prebiotic: a pilot clinical trial. *Eur J Nutr* 2019;58:1735-1745.
143. Lambert JE, Parnell JA, Eksteen B, et al. Gut microbiota manipulation with prebiotics in patients with non-alcoholic fatty liver disease: a randomized controlled trial protocol. *BMC Gastroenterol* 2015;15:169.
144. Liu L, Li P, Liu Y, et al. Efficacy of Probiotics and Synbiotics in Patients with Nonalcoholic Fatty Liver Disease: A Meta-Analysis. *Dig Dis Sci* 2019.
145. Bakhshimoghaddam F, Shateri K, Sina M, et al. Daily Consumption of Synbiotic Yogurt Decreases Liver Steatosis in Patients with Nonalcoholic Fatty Liver Disease: A Randomized Controlled Clinical Trial. *J Nutr* 2018;148:1276-1284.
146. Eslamparast T, Poustchi H, Zamani F, et al. Synbiotic supplementation in nonalcoholic fatty liver disease: a randomized, double-blind, placebo-controlled pilot study. *Am J Clin Nutr* 2014;99:535-542.
147. Scorletti E, Afolabi PR, Miles EA, et al. Design and rationale of the INSYTE study: A randomised, placebo controlled study to test the efficacy of a synbiotic on liver fat, disease biomarkers and intestinal microbiota in non-alcoholic fatty liver disease. *Contemp Clin Trials* 2018;71:113-123.
148. Li SS, Zhu A, Benes V, et al. Durable coexistence of donor and recipient strains after fecal microbiota transplantation. *Science* 2016;352:586-589.
149. Nood E van, Vrieze A, Nieuwdorp M, et al. Duodenal infusion of donor feces for recurrent *Clostridium difficile*. *N Engl J Med* 2013;368:407-415.
150. Bakker GJ, Nieuwdorp M. Fecal Microbiota Transplantation: Therapeutic Potential for a Multitude of Diseases beyond *Clostridium difficile*. *Microbiol Spectr* 2017;5.
151. Aron-Wisnewsky J, Clément K, Nieuwdorp M. Fecal Microbiota Transplantation: a Future Therapeutic Option for Obesity/Diabetes? *Curr Diab Rep* 2019;19:51.
152. Youngster I, Russell GH, Pindar C, et al. Oral, capsulized, frozen fecal microbiota transplantation for relapsing *Clostridium difficile* infection. *JAMA* 2014;312:1772-1778.
153. Cheminet G, Kapel N, Bleibtreu A, et al. Faecal microbiota transplantation with frozen capsules for relapsing *Clostridium difficile* infections: the first experience from 15 consecutive patients in France. *J Hosp Infect* 2018.
154. Kootte RS, Levin E, Salojärvi J, et al. Improvement of Insulin Sensitivity after Lean Donor Feces in Metabolic Syndrome Is Driven by Baseline Intestinal Microbiota Composition. *Cell Metab* 2017;26:611-619.e6.
155. Vrieze A, Van Nood E, Holleman F, et al. Transfer of intestinal microbiota from lean donors increases insulin sensitivity in individuals with metabolic syndrome. *Gastroenterology* 2012;143:913-916.e7.
156. Allegretti JR, Kassam Z, Mullish BH, et al. Effects of Fecal Microbiota Transplantation With Oral Capsules in Obese Patients. *Clin Gastroenterol Hepatol* 2019.

157. Groot P de, Scheithauer T, Bakker GJ, et al. Donor metabolic characteristics drive effects of faecal microbiota transplantation on recipient insulin sensitivity, energy expenditure and intestinal transit time. *Gut* 2019.
158. Zhou D, Pan Q, Shen F, et al. Total fecal microbiota transplantation alleviates high-fat diet-induced steatohepatitis in mice via beneficial regulation of gut microbiota. *Sci Rep* 2017;7. Available at: <https://www.ncbi.nlm.nih.gov/pmc/articles/PMC5431549/> [Accessed April 1, 2019].
159. Bajaj JS, Kassam Z, Fagan A, et al. Fecal microbiota transplant from a rational stool donor improves hepatic encephalopathy: A randomized clinical trial. *Hepatology* 2017;66:1727-1738.
160. Bajaj JS, Salzman NH, Acharya C, et al. Fecal Microbial Transplant Capsules Are Safe in Hepatic Encephalopathy: A Phase 1, Randomized, Placebo-Controlled Trial. *Hepatology* 2019.
161. Colberg SR, Sigal RJ, Yardley JE, et al. Physical Activity/Exercise and Diabetes: A Position Statement of the American Diabetes Association. *Diabetes Care* 2016;39:2065-2079.
162. Schuler G, Adams V, Goto Y. Role of exercise in the prevention of cardiovascular disease: results, mechanisms, and new perspectives. *Eur Heart J* 2013;34:1790-1799.
163. Asano RY, Sales MM, Browne RAV, et al. Acute effects of physical exercise in type 2 diabetes: A review. *World J Diabetes* 2014;5:659-665.
164. Huber Y, Pfirrmann D, Gebhardt I, et al. Improvement of non-invasive markers of NAFLD from an individualised, web-based exercise program. *Aliment Pharmacol Ther* 2019;50:930-939.
165. Zhao X, Zhang Z, Hu B, et al. Response of Gut Microbiota to Metabolite Changes Induced by Endurance Exercise. *Front Microbiol* 2018;9:765.
166. Carbajo-Pescador S, Porras D, García-Mediavilla MV, et al. Beneficial effects of exercise on gut microbiota functionality and barrier integrity, and gut-liver crosstalk in an in vivo model of early obesity and non-alcoholic fatty liver disease. *Dis Model Mech* 2019;12.
167. Kang SS, Jeraldo PR, Kurti A, et al. Diet and exercise orthogonally alter the gut microbiome and reveal independent associations with anxiety and cognition. *Mol Neurodegener* 2014;9:36.
168. Denou E, Marcinko K, Surette MG, et al. High-intensity exercise training increases the diversity and metabolic capacity of the mouse distal gut microbiota during diet-induced obesity. *Am J Physiol Endocrinol Metab* 2016;310:E982-993.
169. Luo B, Xiang D, Nieman DC, et al. The effects of moderate exercise on chronic stress-induced intestinal barrier dysfunction and antimicrobial defense. *Brain Behav Immun* 2014;39:99-106.
170. Davis C, Bryan J, Hodgson J, et al. Definition of the Mediterranean Diet; a Literature Review. *Nutrients* 2015;7:9139-9153.
171. Velasco N, Contreras A, Grassi B. The Mediterranean diet, hepatic steatosis and nonalcoholic fatty liver disease. *Curr Opin Clin Nutr Metab Care* 2014;17:453-457.

172. De Lorenzo A, Noce A, Bigioni M, et al. The effects of Italian Mediterranean organic diet (IMOD) on health status. *Curr Pharm Des* 2010;16:814-824.
173. Browning JD, Baker JA, Rogers T, et al. Short-term weight loss and hepatic triglyceride reduction: evidence of a metabolic advantage with dietary carbohydrate restriction. *Am J Clin Nutr* 2011;93:1048-1052.
174. Shai I, Schwarzfuchs D, Henkin Y, et al. Weight loss with a low-carbohydrate, Mediterranean, or low-fat diet. *N Engl J Med* 2008;359:229-241.
175. Gepner Y, Shelef I, Komy O, et al. The beneficial effects of Mediterranean diet over low-fat diet may be mediated by decreasing hepatic fat content. *J Hepatol* 2019;71:379-388.
176. Katsagoni CN, Papatheodoridis GV, Ioannidou P, et al. Improvements in clinical characteristics of patients with non-alcoholic fatty liver disease, after an intervention based on the Mediterranean lifestyle: a randomised controlled clinical trial. *Br J Nutr* 2018;120:164-175.
177. De Filippis F, Pellegrini N, Vannini L, et al. High-level adherence to a Mediterranean diet beneficially impacts the gut microbiota and associated metabolome. *Gut* 2016;65:1812-1821.
178. Gutiérrez-Díaz I, Fernández-Navarro T, Sánchez B, et al. Mediterranean diet and faecal microbiota: a transversal study. *Food Funct* 2016;7:2347-2356.
179. Mitsou EK, Kakali A, Antonopoulou S, et al. Adherence to the Mediterranean diet is associated with the gut microbiota pattern and gastrointestinal characteristics in an adult population. *Br J Nutr* 2017;117:1645-1655.
180. Garcia-Mantrana I, Selma-Royo M, Alcantara C, et al. Shifts on Gut Microbiota Associated to Mediterranean Diet Adherence and Specific Dietary Intakes on General Adult Population. *Front Microbiol* 2018;9:890.
181. Zelber-Sagi S, Salomone F, Mlynarsky L. The Mediterranean dietary pattern as the diet of choice for non-alcoholic fatty liver disease: Evidence and plausible mechanisms. *Liver Int* 2017;37:936-949.
182. Ross AB, Bruce SJ, Blondel-Lubrano A, et al. A whole-grain cereal-rich diet increases plasma betaine, and tends to decrease total and LDL-cholesterol compared with a refined-grain diet in healthy subjects. *Br J Nutr* 2011;105:1492-1502.
183. Langkamp-Henken B, Nieves C, Culpepper T, et al. Fecal lactic acid bacteria increased in adolescents randomized to whole-grain but not refined-grain foods, whereas inflammatory cytokine production decreased equally with both interventions. *J Nutr* 2012;142:2025-2032.
184. Carvalho-Wells AL, Helmolz K, Nodet C, et al. Determination of the in vivo prebiotic potential of a maize-based whole grain breakfast cereal: a human feeding study. *Br J Nutr* 2010;104:1353-1356.
185. Costabile A, Klinder A, Fava F, et al. Whole-grain wheat breakfast cereal has a prebiotic effect on the human gut microbiota: a double-blind, placebo-controlled, crossover study. *Br J Nutr* 2008;99:110-120.
186. Shen J, Zhang B, Wei G, et al. Molecular profiling of the *Clostridium leptum* subgroup in human fecal microflora by PCR-denaturing gradient gel electrophoresis and clone library analysis. *Appl Environ Microbiol* 2006;72:5232-5238.

187. Guilloteau P, Martin L, Eeckhaut V, et al. From the gut to the peripheral tissues: the multiple effects of butyrate. *Nutr Res Rev* 2010;23:366-384.
188. Endo H, Niioka M, Kobayashi N, et al. Butyrate-producing probiotics reduce nonalcoholic fatty liver disease progression in rats: new insight into the probiotics for the gut-liver axis. *PLoS ONE* 2013;8:e63388.
189. Perumpail BJ, Li AA, John N, et al. The Therapeutic Implications of the Gut Microbiome and Probiotics in Patients with NAFLD. *Diseases* 2019;7.
190. Cani PD, Neyrinck AM, Fava F, et al. Selective increases of bifidobacteria in gut microflora improve high-fat-diet-induced diabetes in mice through a mechanism associated with endotoxaemia. *Diabetologia* 2007;50:2374-2383.
191. Bozzetto L, Prinster A, Annuzzi G, et al. Liver fat is reduced by an isoenergetic MUFA diet in a controlled randomized study in type 2 diabetic patients. *Diabetes Care* 2012;35:1429-1435.
192. Bozzetto L, Costabile G, Luongo D, et al. Reduction in liver fat by dietary MUFA in type 2 diabetes is helped by enhanced hepatic fat oxidation. *Diabetologia* 2016;59:2697-2701.
193. Mujico JR, Baccan GC, Gheorghe A, et al. Changes in gut microbiota due to supplemented fatty acids in diet-induced obese mice. *Br J Nutr* 2013;110:711-720.
194. Wolters M, Ahrens J, Romani-Pérez M, et al. Dietary fat, the gut microbiota, and metabolic health - A systematic review conducted within the MyNewGut project. *Clin Nutr* 2018.