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Biochemical and mathematical lessons from the evolution of the SARS-CoV-2 virus: paths for novel antiviral warfare

Nicolas Cluzel¹, Amaury Lambert^{2,3}, Yvon Maday¹, Gabriel Turinici⁴, Antoine Danchin^{5,6,*}

1. Tremplin Carnot SMILES, 4 Place Jussieu, 75005 Paris, France

2. Laboratoire de Probabilités, Statistique & Modélisation (LPSM), Sorbonne Université, Université de Paris, CNRS UMR8001, 4 place Jussieu, 75005 Paris, France

3. Centre Interdisciplinaire de Recherche en Biologie (CIRB), Collège de France, CNRS UMR7241, INSERM U1050, PSL Research University, 11 place Marcelin Berthelot, 75005 Paris, France

4. Ceremade, Université Paris Dauphine – PSL

5. Kodikos Labs / Stellate Therapeutics, Institut Cochin, 24 rue du Faubourg Saint-Jacques, 75014 Paris, France

6. School of Biomedical Sciences, Li KaShing Faculty of Medicine, Hong Kong University, 21 Sassoon Road, Pokfulam, SAR Hong Kong, China

For correspondence :

E-mail : antoine.danchin@normalesup.org

Tel: +331 4441 2551; Fax: +331 4441 2559

Keywords

ddhCTP, D614G, F1757L, L37F, TN93, tRNA nucleotidyltransferase, non-homothetic growth

27 **Abstract**

28 In the fight against the spread of COVID-19 the emphasis is on vaccination or on reactivating
29 existing drugs used for other purposes. The tight links that necessarily exist between the virus as it
30 multiplies and the metabolism of its host are systematically ignored. Here we show that the
31 metabolism of all cells is coordinated by the availability of a core building block of the cell's
32 genome, cytidine triphosphate (CTP). This metabolite is also the key to the synthesis of the viral
33 envelope and to the translation of its genome into proteins. This unique role explains why evolution
34 has led to the early emergence in animals of an antiviral immunity enzyme, viperin, that
35 synthesizes a toxic analogue of CTP. The constraints arising from this dependency guide the
36 evolution of the virus. With this in mind, we explored the real-time experiment taking place before
37 our eyes using probabilistic modelling approaches to the molecular evolution of the virus. We have
38 thus followed, almost on a daily basis, the evolution of the composition of the viral genome to link it
39 to the progeny produced over time, particularly in the form of blooms that sparked a firework of
40 viral mutations. Some of those certainly increase the propagation of the virus. This led us to make
41 out the critical role in this evolution of several proteins of the virus, such as its nucleocapsid N, and
42 more generally to begin to understand how the virus ties up the host metabolism to its own benefit.
43 A way for the virus to escape CTP-dependent control in cells would be to infect cells that are not
44 expected to grow, such as neurons. This may account for unexpected body sites of viral
45 development in the present epidemic.

46

47 Introduction

48 The development of the COVID-19 pandemic is being explored in a myriad of articles. Despite this
 49 abundance, and because of our anthropocentrism, it is exceptional that these studies focus on the
 50 virus' standpoint. Of course, much work is looking into the details of the composition and structure
 51 of the SARS-CoV-2 virus genome, the proteins it codes for and its animal-infecting relatives.
 52 However, there are very few major studies on how the virus exploits the metabolism of its host's
 53 cells. The urgent necessity to contain the disease led investigators to emphasize vaccination or,
 54 more generally, the involvement of the host's immune system. It is well known, alas, that while it
 55 has sometimes been relatively easy to generate a vaccine that is both effective and harmless
 56 against a widespread disease, the opposite is also true. There are still very serious and very
 57 common diseases for which there is no vaccination. Vaccinating effectively assumes, in particular,
 58 that the progeny of a pathogen remains the same long enough to prevent escape of the immune
 59 response triggered by the vaccine. Coronaviruses are viruses made up of a long genome and an
 60 envelope. The length of the genome could have led to a very high mutation rate, but these viruses,
 61 thus avoiding the universal constraint of Muller's ratchet - see **Box** - have recruited a specific
 62 function that proofreads and corrects replication errors [1]. This means that, while coronaviruses do
 63 indeed tend to produce genetic variants over time, the number of these variants remains quite low.
 64 This mutation rate may appear very limited, but the sheer number of viral particles generated
 65 during an infection is enormous, while the human population currently recognized as infected
 66 exceeds twenty million people. It follows that the mutation rate per nucleotide - of course very
 67 heterogeneous due to the selection pressure on certain locations in the genome - is around 8×10^{-4}
 68 changes per site per year [2].

69 Here, this situation was placed in the perspective of the fundamental theorem of natural selection
 70 proposed by Fisher, which links the evolution of environmental fitness and genetic variance [3]. We
 71 wished to use the marks left by the evolution of the virus' fitness - observed in the form of genomic
 72 sequences - in the presence of the biochemical constraints that bias the choices available for
 73 evolution. We had to take into account, however, that the terms of the problem are not as explicit
 74 as one might have wished: fitness is not known, nor are the time markers (estimated from
 75 phylogenetic trees or simply taken as physical time) and the frequency of certain strains in the
 76 phylogenetic trees may be less due to natural selection than to heterogeneity in sampling and
 77 sequencing depth. This motivated our use of procedures that are robust enough to cope with these
 78 uncertainties. Nevertheless, the advantage of such an analysis is that it allowed us to propose
 79 anticipations for the evolution of the virus. It is therefore an explicit means of feeding
 80 epidemiological or clinical models with relevant observations.

81 In this context, it seemed to us of great interest to explore the details of how SARS-CoV-2 mutated
 82 over time, in the various places where COVID-19 has spread, highlighting relevant descents in

relation with the host metabolism. This should allow us to anticipate some of the future of the virus' progeny, with important consequences for control of the disease. The analysis of the constraints that govern access to the metabolism of the nucleotides that make up the virus genome has shown us that the content of cytosine (C) in its genome is subjected to strong negative pressure, leading to systematic depletion, over time, in cytosine monophosphate [4]. This bias has long been believed to result from a major causal effect of the "editing" of the C content of the genome by the family of APOBEC deaminating enzymes [5,6]. We now know that it is the organization of the metabolism of pyrimidines in animal cells, and more particularly of cytosine triphosphate [7], which drives the corresponding pressure on evolution (**Figure 1**).

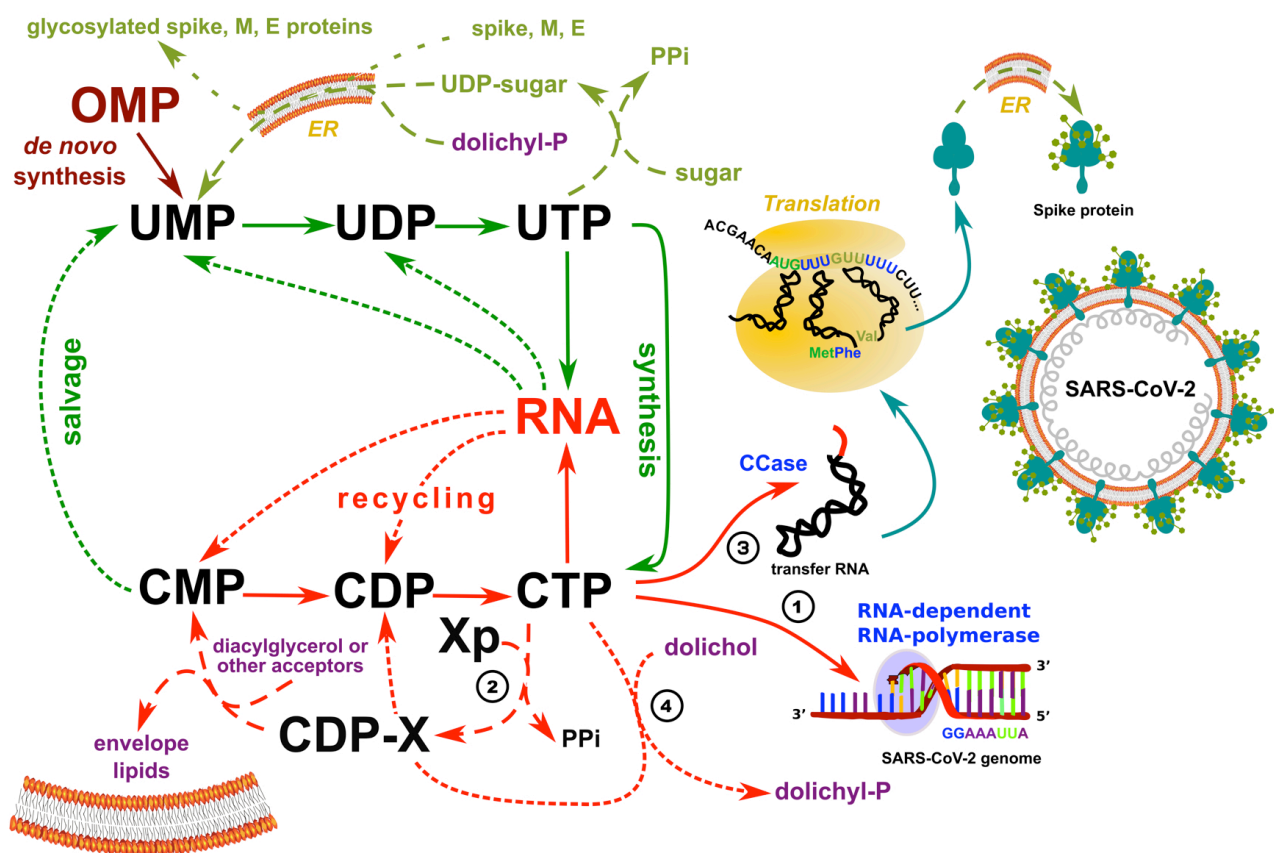


Figure 1. CTP controls all crucial metabolic steps required to build up a functional SARS-CoV-2 virus. 1/ CTP is a precursor of the virus genome; 2/ the lipids of its envelope derive from cytosine-based liponucleotide precursors; 3/ all transfer RNA molecules produced by the host must be matured to a form ending in a CCA triplet at their 3'OH end; and 4/ post-translational glycosylation of viral proteins, in particular its spike protein require a dolichyl-phosphate anchor in the endoplasmic reticulum (ER) and dolichol kinase is specifically dependent on CTP. See text and reference [7] for details.

Indeed, due to the extreme asymmetry of the replication of the virus - which replicates 50 to 100 times from its complementary template [8] - a genome editing effect of these highly context-dependent enzymes would only be significant when a C into U is modified on the negative RNA

103 template, which would lead to a major enrichment in A of the viral genome, or possibly from a U →
 104 C transition due to another class of deaminating enzymes acting on double stranded RNA, ADAR,
 105 that deaminates adenine into inosine [9]. Furthermore, both APOBEC and ADAR are highly specific
 106 enzymes and this hardly fits with the widespread C → U transitions that we keep observing as the
 107 virus evolves. Here, we have focused on the dynamics of the loss of C in the genome, and sought
 108 for the locations and the causes of changes in this driving force. In the first paragraph, we
 109 summarized the metabolic reasons accounting for this remarkable phenomenon. Subsequently, in
 110 the body of the article, we showed that the constraint on the C content of the genome leads to
 111 specific descents which can be used to reveal the existence of important functions of the virus as
 112 well as the role of the host's response.

113 **A universal metabolic requisite, the biosynthesis of cytidine triphosphate (CTP), guides the** 114 **evolution of the virus**

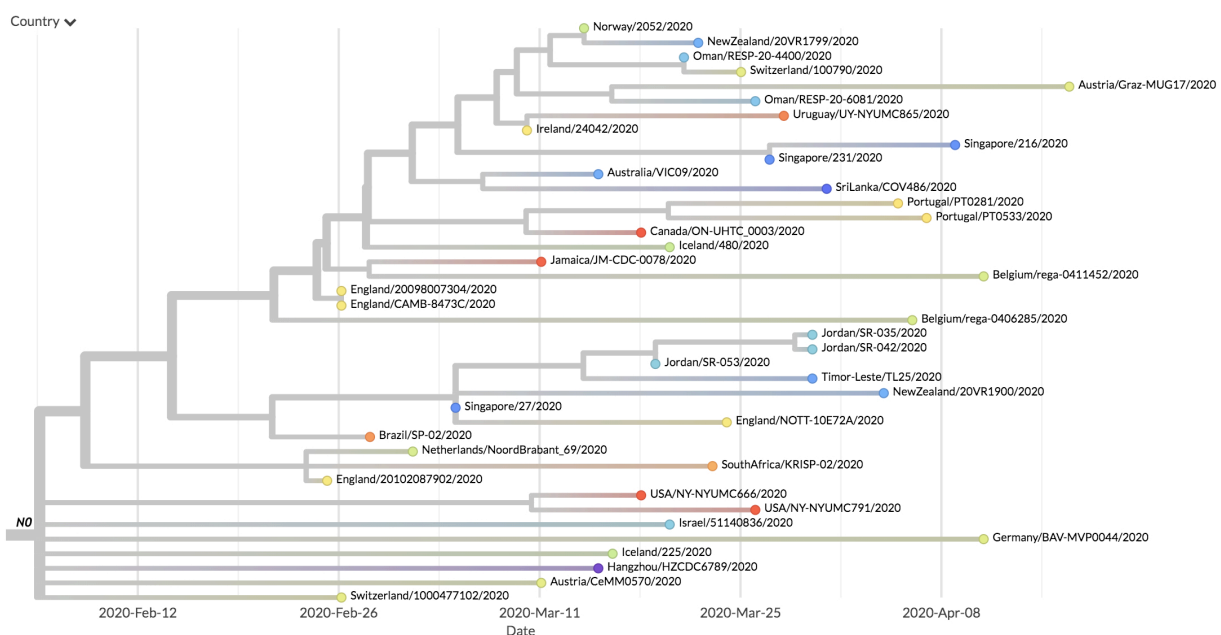
115 What do we know about the synthesis of the building blocks that allow the generation of a viral
 116 particle (a virion)? During a viral infection cells usually stop multiplying. All their resources are
 117 quickly diverted in favour of the multiplication of the virus. Yet, growth is a universal property of life.
 118 This means that, almost always - differentiated neurons are an exception - the cell's metabolism
 119 that the virus faces is organized to allow cell growth as soon as the opportunity to multiply arises.
 120 The moment it infects a cell - again, with the exception of those that do not multiply - any virus will
 121 therefore have to manage the metabolic pressure that organizes the availability of the building
 122 blocks necessary for its construction. In our usual physical space (three-dimensional), growing
 123 introduces an inevitable constraint. The cell must put together the growth of its cytoplasm (three-
 124 dimensional, therefore), that of the membrane that encloses it (two-dimensional) and that of its
 125 genome (one-dimensional, because nucleic acids are linear polymers). However, it is a common
 126 metabolism, developed mainly in the cytoplasm, which produces the building materials needed to
 127 build up these three major compartments. So, here we have a question similar to the one asked by
 128 economists when they raise the question of "non-homothetic" growth [10]. Unfortunately, because
 129 life developed from a primitive metabolism in several stages over 3.5 billion years [10], we might
 130 fear that many organisms had found an idiosyncratic solution to this constraint, as often witnessed
 131 in the huge diversity of life forms. Unexpectedly, it appears that the solution to this quandary is
 132 universal: a single metabolite, the nucleotide cytidine triphosphate (CTP), has been recruited to
 133 this purpose [4,7].

134 The key role of CTP appears in four essential places in cellular metabolism, and these places are
 135 essential for the formation of new virions. 1/ It is the immediate precursor of one of the four
 136 nucleotides forming the genome of the virus; 2/ CTP is required for the synthesis of liponucleotide
 137 precursors of the viral envelope; 3/ human transfer RNAs are synthesized from 415 genes which
 138 do not encode their 3'OH-CCA terminal end - this sequence is synthesized from CTP by a specific

nucleotidyltransferase [12]; and finally 4/ the "decoration" of proteins by complex glycosylations is performed in parallel with their translation in the endoplasmic reticulum (ER) *via* the anchoring of substrates by dolichyl-phosphate, produced by a kinase which uses CTP, not ATP, as its phosphate donor [13]. In addition, intermediate metabolism is based on an original organization of the metabolism of pyrimidines, which systematically recycles and salvages them *via* uridine triphosphate (UTP) which makes CTP a pivot metabolite and limits considerably its availability (**Figure 1**). As a result, accidental replication errors will tend to replace cytosine with uracil in the genome.

General evolution of the SARS-CoV-2 virus

Using the available sequence data gathered in the SARS-CoV-2 GISAID database (<https://www.gisaid.org>) we have, like others [14,15], reconstituted a phylogenetic tree of the evolution of the virus. As the sequences of each viral genome, as well as the date of identification of these sequences are known with fairly great precision, this tree makes it possible to explore the orderly lineage of the mutations which appear over time. In particular, unless we can suspect a recombination event due to the infection of the same patient by two or more viruses, when two identical mutations appear in separate branches of the tree, we can assume that this is the result of evolutionary convergence [16]. The reasons for this convergence are discussed on a case by case basis when analysing each relevant mutation. A second observation, which needs to be put in perspective (see below), is that the shape of the tree is not at all homogeneous. We noticed indeed the presence of "blooms" where, at a particular node of the tree, a large number of branches appear, demonstrating an "explosive" appearance of new mutations (**Figure 2**). We have therefore devised a statistical approach that allowed us to characterize them explicitly.



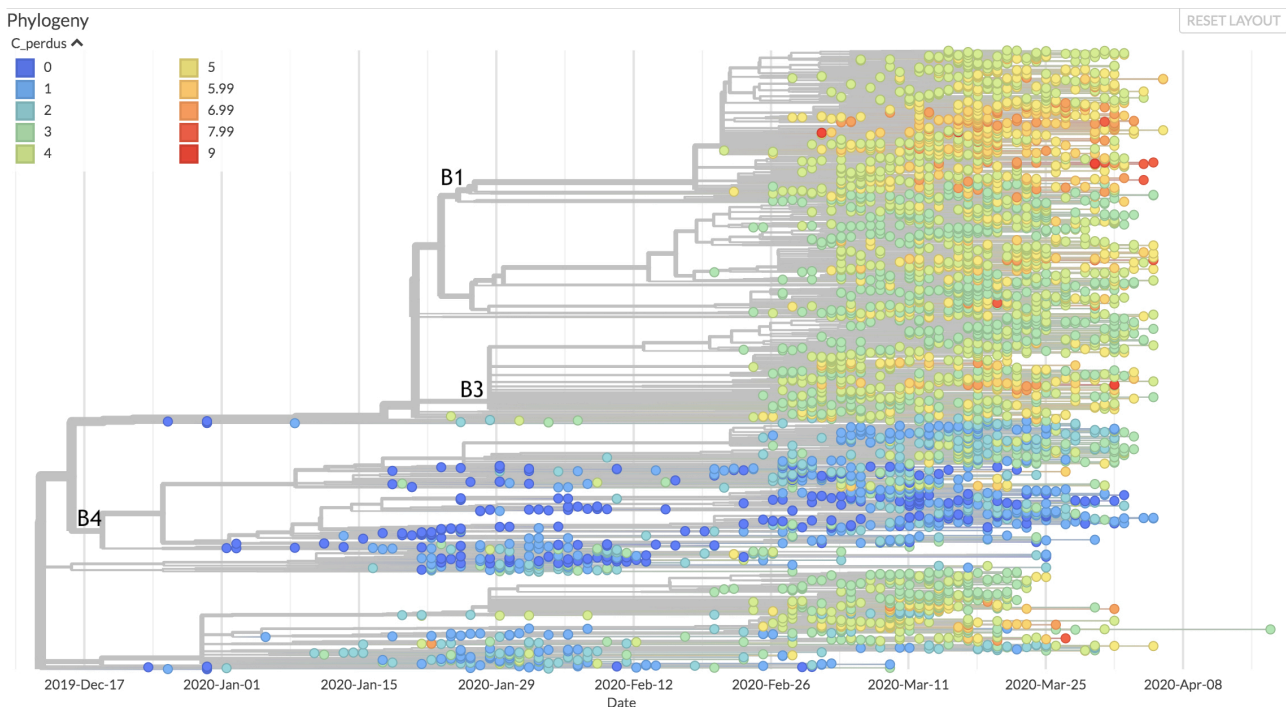
162 **Figure 2. An example of bloom detected by our statistical approach.** At node N0, there are 25
 163 different states in the 40 samples of the subtree and a high number of branches. This behaviour
 164 differs significantly from that of the other sub-trees.

165 The causes of these blooms are multiple, but the adaptation of important viral functions can be at
 166 their origin, and we retained a few cases of this kind for further discussion (see **Materials and**
 167 **Methods** for the statistical definition of blooms).

168 Description and analysis of the evolution of the C content of the genome

169 Generally speaking, the coronavirus genome tends to evolve by adapting its C content to the
 170 metabolism of its host. More specifically SARS-CoV-2 evolves towards forms less rich in C as the
 171 epidemic develops [7]. However, this development is not homogeneous.

172 In the two data sets of interest, 77% of the transitions between pyrimidines are represented by
 173 transitions from cytosine to uracil. These transitions represent 48% of all substitutions identified in
 174 the first set (respectively, 49% in the second). An important imbalance can also be noted at the
 175 level of the transversions, knowing that more than 73% of those pertain to a substitution from
 176 purine to pyrimidine in the first set (respectively, 74%). However, only 20% of these 73% lead to
 177 the occurrence of cytosine (respectively, 17%), indicating once again a tendency to favour the
 178 generation of uracil, thus demonstrating that the major constraint of the mutagenic process is the
 179 availability of each one of the nucleoside triphosphates in the cell. This inhomogeneity is also
 180 salient at the tree level. At the level of branch B4 (20% of the samples), the tendency is strongly
 181 marked to lose less C as compared to the rest of the tree (**Figure 3**).



183 **Figure 3: Heat map of C losses from the original sequence.** Branches 1 and 4 can be readily
 184 discriminated by their extreme values.

185 Interestingly, this branch is also the one that comprises on average the strains with the least
 186 divergence from the original strain of the virus. By contrast, in branch B1, the loss of C looks larger.
 187 The rate of virus mutation also seems to be accelerating in this branch, with a rate of transversions
 188 20% higher than the rest of the tree (and also higher transition rates, but in more anecdotal
 189 proportions). Finally, for branch B3, the main site of blooms, a 29% decrease in the transition rate
 190 of pyrimidines and a 30% decrease in the rate of purines compared to the rest of the tree is
 191 noteworthy.

192 This inhomogeneity can be the consequence of many constraints:

193 1/ The very structure of the genome, which must fold into a compact capsid envelope requires
 194 certain regions to maintain the presence of specific C residues. This is the case of the regions
 195 which control the origin of replication [8] or transcription, AACGAAC, for example [17]. In the case
 196 of the translated regions, the pressure on the presence of C varies depending on its position in the
 197 codon trinucleotides. When C is located at the first position of a codon, it is used to input arginine,
 198 glutamine, histidine, leucine or proline into proteins. Histidine and glutamine are coded in two
 199 codon families, discussed below. For arginine, the selection pressure is lower because the CGN
 200 codons can be replaced by AGR codons - we used here the IUPAC convention for labelling
 201 nucleotides or aminoacids, e.g. N is for aNy, R for puRine, etc.
 202 (<https://www.bioinformatics.org/sms/iupac.html>). The selection pressure on the leucine content is
 203 also lower, since in addition to the CUN codons, this amino acid can be input using the UUR
 204 codons. In the second codon position, C is again used to code for proline, but also threonine
 205 (ACN), alanine (GCN) and serine (UCN). Again, the latter amino acid escapes a large part of the
 206 constraint imposed by the availability of C because it can also use the AGY codons. Finally, the
 207 third position of the codons is much less constrained because it can be replaced by U but also by A
 208 or G in the families with four codons (alanine, proline, threonine, valine). The two codon families
 209 UGY, AGY and NAY are discriminated along a pyrimidine / purine axis. A pyrimidine is used to
 210 maintain the same nature of the coded residue, as the codon uses a U or C as the 3 'end
 211 (aspartate, asparagine, cysteine, histidine and tyrosine). Finally, isoleucine is coded by three
 212 codons (AUH), and ending in U or C is taken into account by relevant tRNAs [18];

213 2/ The function of the virus proteins can impose the presence of certain amino acids in their
 214 sequence. For example, the proline residue encoded by the CCN codons is not strictly an amino
 215 acid, but is essential for the folding of key domains of viral proteins [19];

216 3 / Further stressing the importance of CTP, during evolution, innate antiviral immunity recruited the
 217 activity of an enzyme, viperin, which modifies CTP into a form toxic to the development of the virus,

218 3'-deoxy-3',4'-didehydro-CTP (ddhCTP) [20]. An interesting consequence of this pathway is that
 219 decreasing the C content of the genome will allow the virus replication process to be less sensitive
 220 to the presence of this nucleobase. It follows that, during the transfer of a virus relatively rich in C
 221 from an animal host to human beings, the evolution towards the loss of C may be transiently
 222 concomitant with an increase in its pathogenicity. In the long term, however, the loss of C severely
 223 restricts the evolutionary landscape of the virus and most likely will tend to its attenuation [21].

224 **Examples of correlations allowing us to propose a function for viral proteins**

225 Thousands of mutations have been identified at this date. It is possible to follow their emergence
 226 along the tree of its phylogenetic evolution of the virus and then highlight some interesting features
 227 that may allow us to anticipate some of its future.

228 *Mutations leading to an early translation termination*

229 Mutations leading to premature termination of the virus protein synthesis are expected to appear
 230 with high frequency. In the present context, this is all the more likely because the translation
 231 termination codons UAA, UAG and UGA do not contain C, and are therefore favoured by the
 232 disappearance of this nucleotide. Since most of these mutations lead to non-functional
 233 polypeptides, it is generally probable that the affected viruses do not give rise to a significant
 234 progeny. It follows that when these mutations are observed - and that they do not result from
 235 sequencing errors - they indicate that the role of the truncated protein corresponds to a function
 236 which is not critical, or that the protein has remained functional at a sufficient level to allow virus
 237 reproduction. However, a few observations allowed us to offer an explanation for the fact that the
 238 viruses in question may have survived. Here are three examples which reveal interesting features
 239 of the virus.

240 Example 1: In a strain from Iceland, the succession of mutations G1440A (Gly392Asp, protein
 241 Nsp2) and G2891A (Ala876Thr, ubiquitin-like domain of protein Nsp3) is now present in multiple
 242 world locations [22]. This sequence ends up with C27661U (which modifies amino acid Gln90 into
 243 a premature translation end, near the carboxy-terminal end of protein Orf7a). This viral protein is
 244 found in the endoplasmic reticulum, the Golgi apparatus and the perinuclear space [23]. Several
 245 variants have been identified in the course of the epidemic [24]. Remarkably, several deletions
 246 have been isolated in the gene, which suggests that the function of this region is not essential [25].
 247 However, we noticed that many of these mutations, as the one discussed here, keep the small
 248 hydrophobic protein Orf7b gene intact, downstream of Orf7a. This very small protein is present in
 249 the Golgi apparatus and is also found in the purified virus [26]. It must be noticed that it is
 250 synthesized *in vivo* via a frameshift that spans the termination codon of the Orf7a frame (...GAA
 251 TGA TT... becomes ...GAATG ATT...). This can be interpreted as a conflict in this region between
 252 translation of Orf7a and Orf7b, creating a cost / benefit dilemma for the expression of either one of

these proteins. Hence it will be important to monitor the future descent of the virus in this region as it may result in interesting attenuated forms.

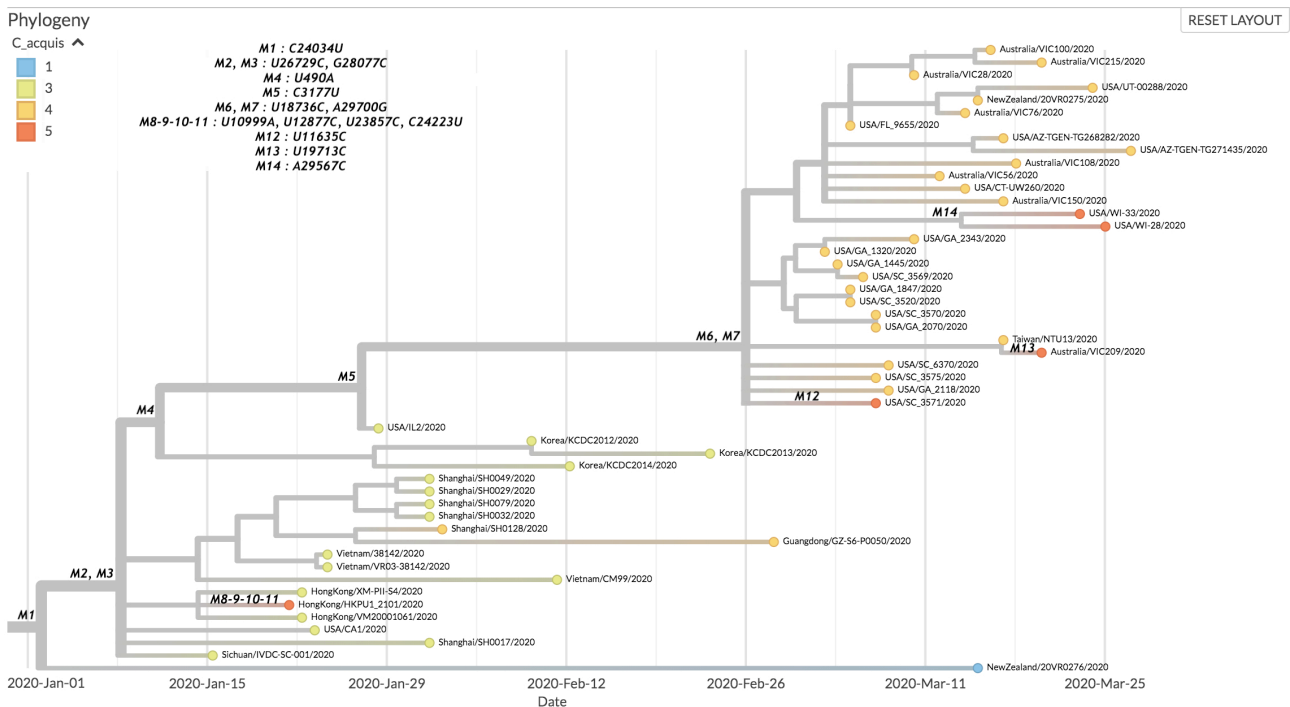
Example 2: Another succession of mutations that leads to premature translation termination of a viral protein begins with G11083U (protein Nsp6, Leu37Phe). This mutation is now widely distributed worldwide. It is likely to induce a more stable binding of the protein to the ER, possibly favouring coronavirus infection by compromising delivery of viral components to lysosomes for degradation [27]; then we have G1397A (Nsp2, Val378Ile), also likely to favour virus propagation [28]; followed by G29742U (3'UTR of the virus), and U28688C (synonymous); subsequently, we have the couple of mutations C884U (Nsp2 again, Arg207Cys [28]) and G8653U (Nsp4, essential for envelope assembly [29]. The corresponding change (Met2796Ile) is located at the border of the ER lumenal domain of the protein. It is known that, in order to function properly, the ER requires the presence of oxygen [30], and reactive oxygen species (ROS) are associated to misfolding of proteins in this compartment. Nsp4 has a number of cysteine residues, prone to be oxidized. The role of methionine in the parent might be to act as a buffer against ROS, so that the mutant would be slightly attenuated). These mutations are followed by A19073G (in the methylase domain of protein Nsp14, Asp1869Gly, a position that already evolved from SARS-CoV-1 [31], hence likely to be more or less neutral), then the couple with the mutation resulting in end of translation: G27915U, Gly8 to end of translation at the N-terminus of Orf8 and C29077U (synonymous); the succession ends with the couple of mutations leading to synonymous changes C19186U and G23608U. This region of SARS-related coronaviruses is hypervariable. It changes during the course of epidemics, showing that it is subject to ongoing selection pressure, sometimes producing two peptides Orf8a and Orf8b [32]. It corresponds to proteins expressed at the end of the infection cycle. It will be important to monitor the way they function in the course of the evolution of virulence of the virus. This displays a branching that appeared in four different countries and in seven samples, spanning six weeks between the first and the last mutation.

Example 3: Here we have a succession of mutations that begin within the 5'end of the virus genome, C241U, followed by mutation C14408U (Pro314Leu) at the end of a zinc finger in replicase Nsp12, which appears in many branches of the evolution tree of the virus. It is discussed in details below (origin of blooms). This mutation is followed by A23403G (Asp614Gly) a widely spread mutation of the spike protein (also discussed below), C3037U (synonymous), mutation G25563U (Gln57His) in Orf3a forming potassium channels is supposed to negatively interfere with the function of the protein [33], C1059U (Thr265Ile) in protein Nsp2, discussed previously, and the triplet G4181A (Ala1305Thr) in the SUD-N domain of protease Nsp3, then mutations G4285U (Glu1340Asp), and G28209U which results in an end of translation at glutamate 106 of protein Orf8. As discussed previously, many mutations, including deletions in Orf8, were frequently observed. This is again an indication that evolution of this regions should be carefully monitored to

look for attenuated forms of the virus. This particular mutation to an end of translation is significant as it was found in a sample from Croatia, another one from Thailand, on two significantly separated branches and with one month difference. The sequence of mutations here corresponds to the Thailand sample.

Reversal of the tendency of the viral genome to lose its cytosine residues

We have here retained two examples of a situation where, from an upstream branching point in the evolution tree, it appears that the descendants of the virus stop losing their cytosines, and may even tend to regain them. These examples are as follows (**Figure 4**).



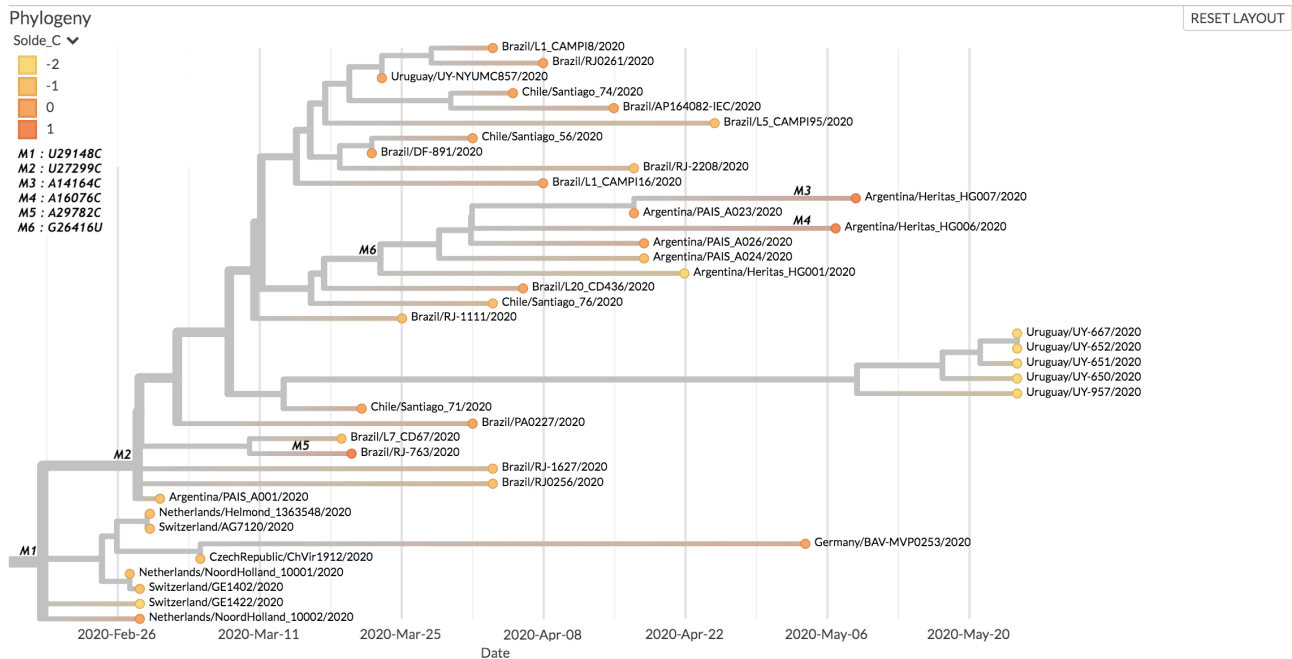


Figure 4. Two sub-trees in the first dataset where the tendency of the genome to lose its cytosine residues is reversed. Upper panel. First sub-tree. The sub-samples displayed are those that have acquired the most C, apart from a few isolated samples on other branches. The node with the M1 mutation directly follows those respectively associated with the C8782U and U28144C mutations. **Lower panel. Second sub-tree.** This tree contains a majority of strains with a neutral C balance (both gained and lost), as well as 3 strains with more C gained than lost.

In dataset 1, there are two sub-trees, the first of which is more of an Asian sub-tree with the root of the node associated with the M2 and M3 mutations. The second contains samples from North America and Oceania, and its root node is related to the M6 and M7 mutations. The first tree arises from the succession of C8782U (synonymous), U28144C (Leu84Ser) mutations in the Orf8 protein, whose function was discussed above. It defines a major clade of variants of the virus [24], C24034U (synonymous), and finally the doublet U26729C (synonymous), G28077C (Val62Leu), in the Orf8 protein again. As this is the origin of the observed phenomenon, we are led to believe that it is the alteration of the role of Orf8 (8a or 8b) that is responsible. The Orf8 region is particularly variable and has been clearly implicated in interspecies transmission [34]. A common hypothesis is that the alteration of this gene corresponds to a loss of active function in chiropteran ancestors [35]. Since these are generally richer in cytosine than the human forms [21], one might ask whether one of the functions of this protein is to modulate the activity of CTP synthase.

In fact, the second branch comes from the same descent, to which is added the U490A (Asp75Glu) mutation in the Nsp1 protein, which controls the specific translation of viral RNA [36], systematically associated with the mutation C3177U (Pro971Leu) in the acidic domain, without any clearly identified function, of the multifunctional protease Nsp3 [37], and finally the U18736C (Phe1757Leu) doublet of the exonuclease, N7-methyltransferase Nsp14, and A29700G in the 3'UTR region of the virus. The Phe1757Leu modification is located in the middle of a zinc binding

site at the interface between the two domains of the Nsp3 protein. It can therefore be surmised that this mutation could subtly change the proofreading process correcting replication errors in a way that would be less amenable to the entry of UTP opposite an A in the negative viral template. We noted that 3 out of the 5 samples that acquired the most C did so through a transition from U to C. The first one, HongKong/HKPU1_2101, shows two simultaneous transitions at positions 12877 and 23857. These mutations being synonymous, they are unlikely to change the replication-correction mechanism. The second one, USA_SC_3571, and the third one, Australia/VIC209, show transitions of the same type, also synonymous, at positions 11635 and 19713 respectively. Finally, the last two samples, USA/WI-33 and USA/WI-28, were derived from the transversion from A to C at position 29567, a mutation at the end of ORF9b.

For dataset number 2, this reversal of the trend concerns mostly Latino-American strains. The succession of mutations C241U, C14408U, then A23403G discussed in relation to the generation of end of translation codons in the virus genes, is followed by C3037U (synonymous), and the triplet G28881A, G28882A, G28883C, overlapping the codons at position 203-204 of the N nucleocapsid N gene. They mutate an arginine-glycine dipeptide into a lysine-arginine dipeptide. This alters the positive charge of the protein and may help improve its role in the assembly of the virus genome in the capsid, as discussed below in relation to the appearance of blooms (36). After this triple modification, we see several reversals of the tendency to lose C in the genome. U29148C (Ile292Thr) is found again in the nucleocapsid N gene, then U27299C (Ile33Thr) in the Orf6 gene, resulting in a set of samples that have at worst gained as much C as they have lost. There are also 3 samples among the 39 in the subtree that gained one more C than they lost (Brazil/RJ-763, Argentina/Heritas_HG007, Argentina/Heritas_HG006). Each time, the last C acquisition comes from a transversion from an adenine (in positions 14164 (Met233Leu), 16076 (Asp870Ala), and 29782, in the late 3'UTR of the viral genome. Overall, it is the change in the nucleocapsid that appears to be most conducive to reversing the tendency to lose C. Indeed, this protein, expressed at a high level during the infection, regulates the process of replication / transcription of the virus and this may account for this remarkable observation [39].

Emergence of blooms

The succession C3037U, (C241U, A23403G), C14408U is present upstream of 10 sub-trees, which we considered to be significant (see **Figure 5** and **Materials and Methods**).

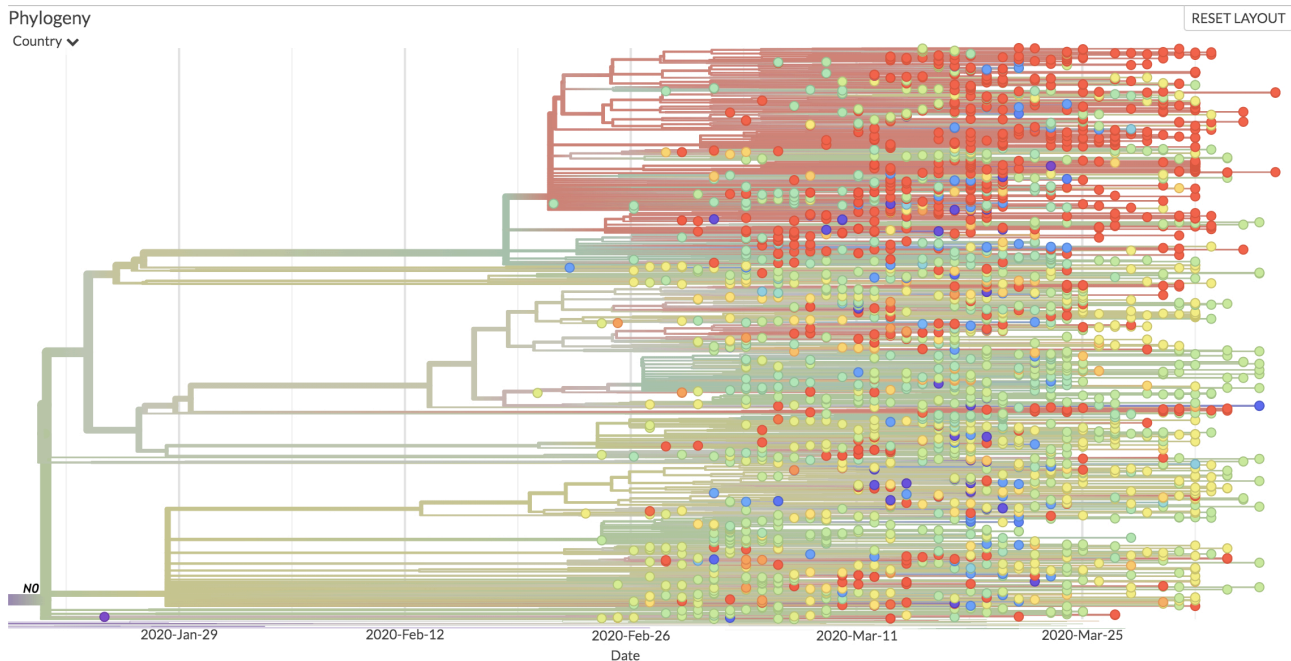


Figure 5: Example of blooms The subtree shown here contains 10 of the 20 most significant blooms in the sense of the method we used. Node N0 is the place where mutation C14408U emerges.

The synonymous mutation C3037U is located at the end of the ubiquitin-like domain 1 of protein Nsp3. This leads to a sequence (UUUUUU) that promotes changes in the reading frame and could decrease the translation efficiency of the proteins of the ORF1a region. The C241U mutation is observed very often [40]. It is found in the region that initiates replication of the virus. We can therefore assume that this may alter the frequency of replication. A23403G is a widely spread non-synonymous mutation which leads to the replacement of an aspartate by a glycine at position 614 of the spike protein, which is used by the virus to bind its host cell's receptor. For this reason, several previous analyses have suggested that this mutation has an important role in the spread of the virus [41,42]. Here, the fact that it is part of a major bloom can be considered as an additional argument favouring this interpretation. The C14408U changes an amino acid from proline to leucine (Pro314Leu) just after the end of the NiRAN domain (nido virus RdRp-associated nucleotidyl transferase) of the protein Nsp12 ending in a "zinc finger". The NiRAN domain, essential for the replication of the virus, acts as a nucleotidyltransferase, preferring UTP as a substrate for a function which has not yet been clarified [43]. The proline modified in the mutant is part of a dipeptide diproline which plays the role of hinge of separation between the NiRAN domain and the following domain.

A second bloom, which shares several elements with the preceding one begins with the same sequence C3037U, (C241U, A23403G) and C14408U. However it continues with a series of contiguous mutations resulting in a change (G28881A, G28882A, G28883C) in nucleocapsid N, as we saw previously. It is worth noticing that this change might have a role in assembling the virus

377 genome in the capsid by phase separation [38]. This might increase the efficiency of virus
 378 transmission and thus contribute to the formation of blooms. The fact that it is a cluster of
 379 mutations involving G is intriguing. It may result from the fact that it spans a GGGG sequence.

380 We have previously seen that mutation G11083U (protein Nsp6, Leu37Phe) has initiated another
 381 succession of mutations that led to premature translation termination of a viral protein. Here, this
 382 widely distributed mutation is at the root of blooms. As discussed, it is possibly favouring
 383 coronavirus infection by compromising delivery of viral components to lysosomes for degradation.
 384 This would certainly favour blooms. The mutation is followed, in a first bloom-generating
 385 succession, by G26144U (Gly251Val) in protein Orf3a, that forms potassium channels important for
 386 innate immunity response - but the exact function of the protein still remains open to question [44].
 387 Subsequent mutations are C14805U (synonymous) and U17247C (synonymous). This succession
 388 suggests that the first mutation in protein Nsp6 and perhaps the second one are the primary
 389 causes of the bloom [27]. The role of the first mutation is further substantiated by the second bloom-
 390 generating succession where it is followed by a quadruplet: C6312A (Thr2016Lys) in the inter-
 391 domain region that precedes domain G2M of multi-domain protease Nsp3, then associated with
 392 three C → U mutations, hence expected to be more frequent: C13730U (Ala88Val) in the NiRAN
 393 domain of protein Nsp12, C23929U (synonymous), and finally C28311U (in a sequence of four C,
 394 Pro13Leu) at the beginning of the nucleocapsid protein, N.

395 A second succession of mutations that ends up in blooms is C8782U (synonymous), U28144C
 396 (Leu84Ser) in protein Orf8, the function of which has been discussed previously and defines a
 397 significant clade of the virus variants [24], ending up with C26088U (synonymous). The Leu84Ser
 398 mutation co-evolves significantly with the Asp614Gly mutations of the spike protein discussed
 399 above [37], which makes it another likely candidate for positive selection leading to increased
 400 spreading of the virus, hence blooms.

401 *Change in frequency of transitions / transversions*

402 Among the mutations upstream of branches showing significant changes in the transition /
 403 transversion flow is the mutation C17747U, which modifies a proline residue into a leucine residue
 404 in the protein Nsp13 (**Figure 6** and **Materials and Methods**).

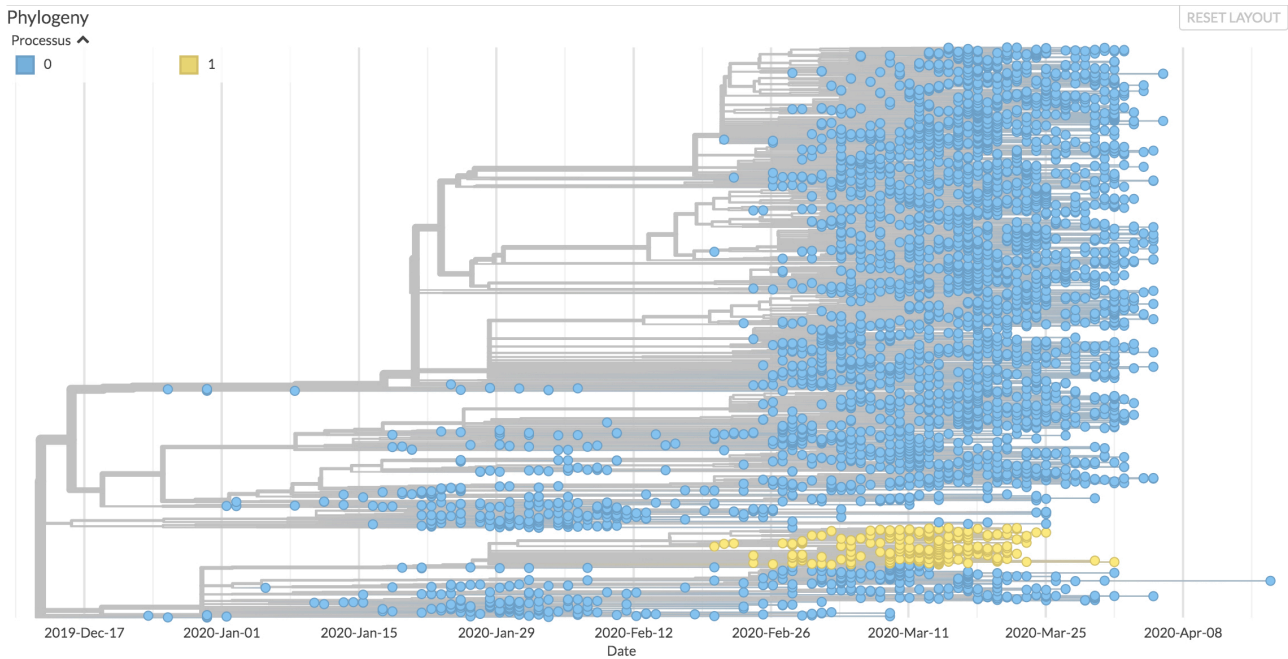


Figure 6. One of the descents considered to be significant for the change in the process of molecular evolution. The progeny resulting from the C17747U mutation is shown in yellow, and its evolutionary process is modelled by a 6-parameter TN93 model (process 1). The rest of the tree (blue leaves) is modelled by a 3-parameter TN93 model.

This mutation affects the protein domain which has nucleoside triphosphatase activity, the exact role of which is unknown but consistent with a proofreading activity [45]. We might propose that it is involved in the quality control of the product of the replication of the virus for example *via* stabilizing the “anti” form of nucleotides, thus avoiding the mismatching leading to transversions. In fact, this protein has been identified among those which lead to a significant alteration in the diversity of the viral genome [16]. The existence of a notable change in the type of mutations located downstream of the tree is therefore a strong argument for the discriminating role of the corresponding region of the protein. Furthermore, to the extent that this mutation increases the frequency of mutations in a biased manner, we can expect the ensuing descent to lead to an attenuation of the virus. However, as this changes the evolutionary landscape, this evolution could lead to “innovative” mutations modifying the pathogenicity of the virus, and this especially under conditions where recombination due to co-infections would be favoured. This is yet another argument for choosing a strong public health policy which tends to avoid the formation of clusters of infection.

Conclusions and perspectives

The COVID-19 epidemic is a life-size experiment in virus evolution. Remarkably, we neither know the real origin of the virus [46], nor where it will lead us. This explains why the vast majority of studies of the SARS-CoV-2 virus and its evolution are essentially descriptive. Here, we tried to make use of the ongoing evolution of the virus to investigate some of its related constraints using a hypothesis-driven probabilistic modelling approach to the molecular evolution of the virus. Based

on the assumption that the virus' metabolism is ruled by its host. Based on the metabolic set up of the host cells, acting as a compulsory material framework for the multiplication of viral particles, we pointed out specific changes in the evolution pattern of the virus descent, witnessed by changes in the virus genome composition as time passes. Using the widely spread C to U change in this genome's composition as a base line, we identified nodes where the change is shifted from this direction to another one, favouring transversions rather than transitions, reversing the C to U trend towards U to C enrichment or generating blooms with sudden appearance of multiple branches in the evolution tree. This allowed us to point out a series of functions that are evolving towards a more efficient spread of the virus (e.g. the previously identified Asp214Gly mutation of the spike protein, but also the Gln57His mutation of the Orf3a potassium channel). We also noticed that Orf8 is the likely site of an ongoing competition for expression of two frameshift-dependent overlapping proteins Orf8a and Orf8b. Similarly, the unstable region of Orf7 could promote the synthesis of the very small membrane protein Orf7b, whose function remains unknown to date. Finally, the reversion of the tendency to favour U over C indicates that nucleocapsid protein N may be involved in the control of CTP synthesis in the host, suggesting an interesting target for future control of the virus development. We hope that this combination of mathematical and biochemical knowledge will help us devise further enterprises against the dire consequences of COVID-19. We noticed that among the possible way for the virus to escape CTP-dependent control in cells would be to infect cells that are not expected to grow, such as neurons. This may account for unexpected body sites of viral development observed in the present epidemic.

Acknowledgements

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Materials and Methods

Data processing

A total of 4,792 sequences of the SARS-CoV-2 virus were recovered from the GISAID databank [47] on April 21, 2020 for the first dataset. Only the genomes of viruses from the human hosts of SARS-CoV-2 of a length greater than 25,000 bp were retained. Sequences for which the sampling date was insufficiently informed (absence of the harvest day, sometimes of the month) were also excluded. For sequences present multiple times, only the first isolate was retained. We also reused the work of the Nextstrain teams and discarded the too divergent or unstable samples that they themselves had left out (github.com/nextstrain/ncov/blob/master/defaults/exclude.txt). The sequence of 26 coding regions (Nsp1, Nsp2, Nsp3, Nsp4, Nsp5, Nsp6, Nsp7, Nsp8, Nsp9, Nsp10,

464 Nsp11, Nsp12, Nsp13, Nsp14, Nsp15, Nsp16, S, ORF3a, E, M, ORF6, ORF7a , ORF7b, ORF8, N
 465 and ORF10) was characterized using NC_045512 as a reference. The total number of sequences
 466 retained at the end of the treatment is 4,088 sequences. A second dataset of 3,246 sequences,
 467 510 of which are common with the first dataset, was retrieved on July 6, 2020 using directly the
 468 Nextstrain API [48].

469 We note here that, over time, data availability kept being altered, with some sequences deleted
 470 from the samples, while other ones entered the database. Furthermore, it was generally difficult to
 471 extract large samples of sequences so that it was extremely difficult to build up a consistent data
 472 repository where correct statistical approaches could be implemented. It seems very awkward that
 473 the bulk of the sequences of a virus of worldwide importance has not been made available at the
 474 International Nucleotide Sequence Database despite recommendations of the major research
 475 institutions [49,50].

476 *Phylogenetic reconstruction*

477 The reconstruction process begins with aligning all of the sequences to the reference sequence.
 478 Insertions and deletions of genome regions were not taken into account. It was out of the question
 479 here to take into account nucleotide insertions and deletions. We retained only the potential one to
 480 one substitutions. We used program MAFFT [51] to generate these alignments. Some ambiguous
 481 positions were highlighted during the alignment process. For example, some regions of the
 482 genome may display high instability and wide variability depending on the parameters of the
 483 algorithm used to perform the alignment. To overcome this problem, we used the same masks as
 484 those used by the Nextstrain team. Sites 18529, 29849, 29851, 29853, as well as the first 130 and
 485 last 50 sites of the genome were therefore omitted from the substitution analysis. We used a
 486 General Time Reversible (GTR) model to infer the substitution process at work using the IQTREE
 487 software [52]. This first tree is a fairly raw version which does not take into account the temporal
 488 aspect of evolution. The Treetime software [53] allows you to refine this tree by also taking into
 489 account the sampling dates of the sequences. It then reconstructs the tree with maximum
 490 likelihood compared to the sampled sequences. Using maximum likelihood approaches it also
 491 infers the compositions of the ancestral sequences of the samples, as well as a 90% confidence
 492 interval around the most likely date of these common ancestors. Once the tree has been created,
 493 we could then reconstruct the order in which the mutations in each sample appeared, in the sense
 494 of maximum likelihood. For the visualization of the tree and the production of Figures 2 to 6, we
 495 used the Auspice program developed by Nextstrain, to which we made some modifications to
 496 display the parameters we were interested in. To this purpose, we developed a Python script to
 497 modify the JSON file used as input by the Auspice program. This allowed us to enrich the
 498 visualization capabilities of the software by adding quantities such as the number of C acquired or
 499 lost by a sample compared to the reference and to generate original tree presentations.

500 *Identification of blooms*

501 The main pitfall we had to face when identifying blooms was the bias introduced when selecting
 502 samples from the phylogenetic tree. In particular, some hospitals were likely to provide more
 503 samples than others, due to the different health policies and means implemented depending on the
 504 country. In order to avoid selecting nodes likely to generate a bloom due to oversampling, we
 505 chose to develop a custom-made statistical method meant to cope with this difficulty.

506 A subtree is any set of nodes and leaves rooted in one of the nodes of the main tree. The idea is to
 507 use the information provided by the identity of the countries represented in each sub-tree: the
 508 easier a strain is spread, the higher the number of countries in which it is expected to be observed.
 509 To implement this heuristic, it is necessary to control two factors: the size of the tree (two trees of
 510 unequal depth, that is to say rooted on different dates, naturally show diversity as different
 511 countries) and the heterogeneity of sampling (countries where sampling and sequencing are
 512 carried out with different intensities have different probabilities of appearing in a given sub-tree).

513 These two factors interact, because the size of a tree (the number of its leaves for example)
 514 obviously varies with the sampling intensity. One way to control this interaction is to measure the
 515 size of a tree by its total length, or sum of branch lengths, in time units. Indeed, this observable is
 516 not very sensitive to the effects of oversampling because the presence of many sequences
 517 sampled in the same place at about the same time generates a sub-tree whose length is close to
 518 zero.

519 To control the effect of the length factor L on the number of countries represented, N , we sought to
 520 learn the relation $N = f(L)$ in a typical tree in order to be subsequently able to identify the sub-trees
 521 whose number of countries represented, for a known length L , exceeds the expected $f(L)$. A simple
 522 statistical model consists in supposing that the number of occurrences of country i in a tree of
 523 length L is a Poisson distribution of parameter $\theta_i L$ and that these numbers are independent. If K is
 524 the total number of countries referenced by Nextstrain, the number of countries N represented in a
 525 tree of length L is therefore the sum of K Bernoulli variables independent of parameters $1 - \exp(-\theta_i L)$.
 526 For example, if countries are divided into two groups, the k_1 'frequent' of intensity θ_1 , and the
 527 k_2 'rare' of intensity $\theta_2 \ll \theta_1$, N has the mean $K - k_1 \exp(-\theta_1 L) - k_2 \exp(-\theta_2 L)$,

528 which behaves when L is large like $K - k_2 \exp(-\theta_2 L)$.

529 In addition, when L is large, assuming that $\theta_2 L = O(1)$, the distribution of N is approximately equal
 530 to $k_1 + N_2$, where N_2 follows a Poisson law of parameter $k_2(1 - \exp(-\theta_2 L))$.

531 So, we used the parameterization:

532 $N = a - b \exp(-cL)$, interpreting the parameters as follows: a is the maximum number of countries, b
 533 is the number of countries with low sampling/sequencing intensities and c is a density of presence
 534 of these countries per unit length of tree. Under the null hypothesis, N is distributed as $a - b + N_1$,
 535 where N_1 follows the Poisson's law of parameter $b(1 - \exp(-cL))$. Finally, we selected the 20 most
 536 significant blooms, i.e., those whose behaviour deviated the most from that expected by our
 537 estimator. This then allowed us to reconstruct the lineages and the mutations that appeared
 538 successively upstream of each node at which a bloom had occurred. This allowed us to identify the
 539 succession of mutations common to some of these nodes and thus those giving rise to the majority
 540 of statistically significant blooms. Furthermore, we restricted the automatic selection of nodes so
 541 that no selected node was present in the lineage of another one. The selected blooms are
 542 therefore mutually independent, even though they may obviously have common ancestors. To
 543 arbitrate the choice between two nodes present in the same lineage, we have systematically kept
 544 the oldest node, and thus the most dense tree.

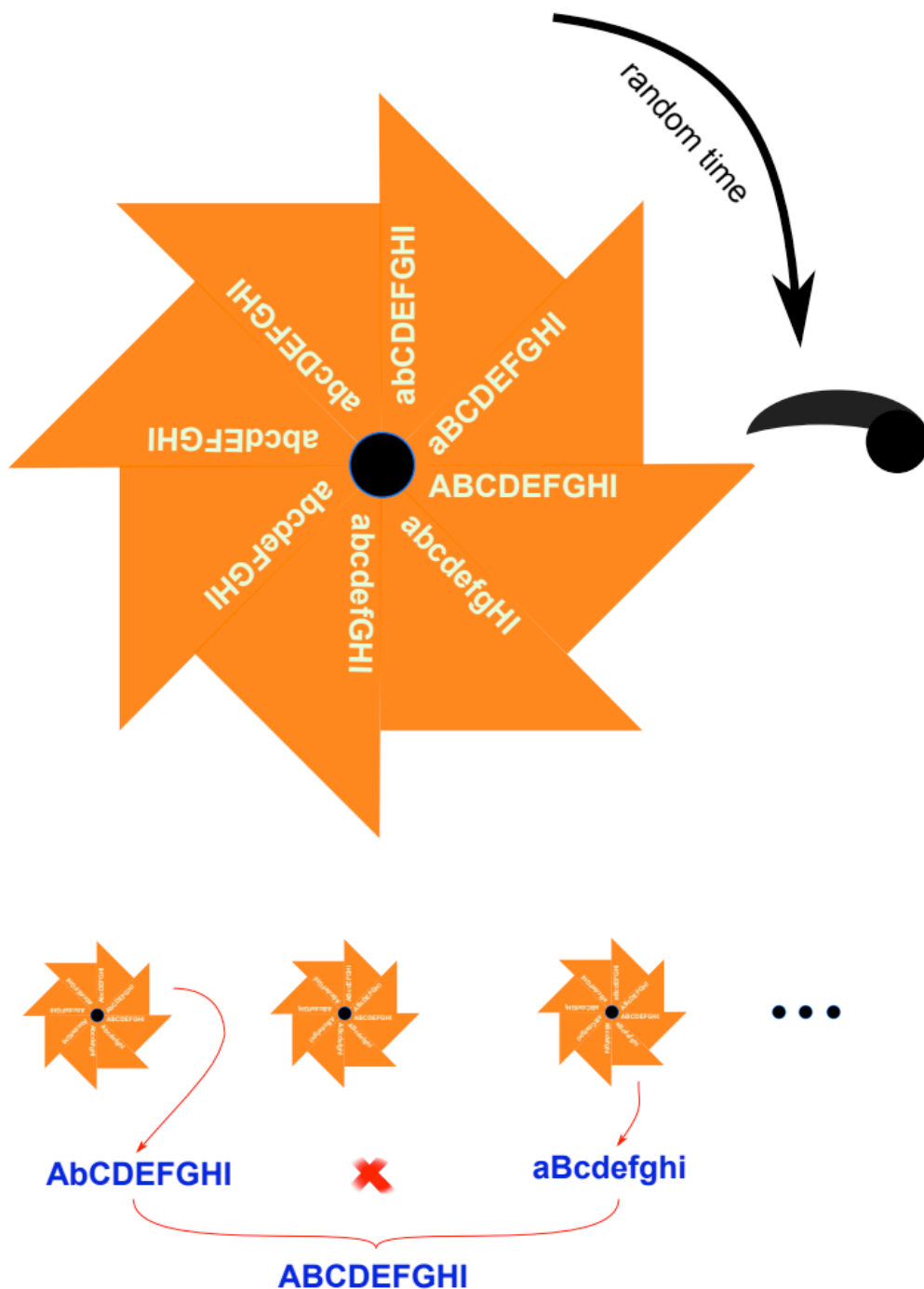
545 *Detection of changes in the molecular evolutionary process*

546 We investigated whether the substitution process in some sub-trees behaved differently from what
 547 was observed in the rest of the tree, statistically speaking. To this aim, we used the classical TN93
 548 model from Tamura and Nei [54] with 3 parameters (purine transition rate, pyrimidine transition rate
 549 and transversion rate) and allowed these three rates to take, downstream of a candidate node N_i ,
 550 values that differed from those they take in the rest of the tree. We then used a second (6-
 551 parameter) model. Since this model is nested in the first (3-parameter) model, we used as test
 552 statistic the likelihood ratio $2\Delta l = 2(l_1 - l_0)$, where l_0 is the log-likelihood under assumption H_0 (3-
 553 parameter TN93 model estimating all the elements of the tree) and l_1 is the log-likelihood under
 554 assumption H_1 (6-parameter TN93 model with local differentiation of the parameters downstream of
 555 a node of interest). We then compared the likelihood ratio to a distribution of the χ^2 with 3
 556 degrees of freedom, whose significance threshold at 5% is 7.81. We were then able to identify the
 557 nodes from which the evolution process varied significantly and to quantify the variations of the
 558 different substitution rates, i.e. the nodes for which we can reject the H_0 hypothesis that the 3-
 559 parameter model TN93 produces better estimates of tree substitution rates than the 6-parameter
 560 model TN93. We have chosen to implement these models ourselves in Python, in order to keep
 561 this parametrization flexibility. The program allows us to determine the set of nodes and leaves
 562 present downstream of a node of interest and to perform hypothesis testing by calculating the
 563 likelihood ratio and the different substitution rates.

564 TEXT BOX**565 Molecular evolution and Muller's Ratchet**

566 Biology rests on the laws of physics. Because it develops at approximately 300K, it is subject to the
567 universal stress of thermal noise, involving an energy that does not differ considerably from that
568 involved in the chemical bonds of biological chemistry. It follows that the reactions that come
569 through and organize living things cannot develop with strict reproducibility. Inevitable errors cause
570 the product of a reaction to differ from what it is supposed to be. Genome replication cannot
571 escape this constraint. The consequence is that, in the progeny of a virus, there is always a
572 number of variants, named mutants when they carry over alterations of the genome. In most
573 cases, these mutants correspond to the change from one of the four nucleotides to a different one.
574 This process, as a rough approximation, is random — the mutant position can be anywhere in the
575 genome, and the replacement of one nucleotide is by any of the other three. As time goes by, all
576 the nucleotides of the genome are likely to change into others. This will affect the functions
577 necessary for the multiplication of the virus, and some changes will continue to be propagated (be
578 fixated), while others will end up without a progeny. A mutation followed by a fixation is called a
579 substitution. The substitution of a purine for a pyrimidine (or vice versa) is called a transversion;
580 other substitutions are called transitions. The likelihood of a particular mutation returning to the
581 ancestral state is very low. The probability that a particular mutation will return to the ancestral
582 state is very low. This forces evolution to always go forward, without the possibility of going back.
583 This process was noticed in 1932 by Hermann Muller in the special case of the effects of irradiation
584 on mutagenesis. His reflection has since been simplified and popularized. It is now known as the
585 "Muller's ratchet" [55]. It is obviously highly probable that the majority of mutations leads to the
586 partial or total loss of the functions coded by the altered regions of the genome. It follows that this
587 generally leads, in the long term - but not in the short term - to the attenuation of the functions
588 allowing multiplication and virulence of pathogenic species. This is why Louis Pasteur and his
589 successors could have the luck to isolate attenuated organisms which, in some - rare - cases,
590 could then be used for the vaccination of infected persons [56]. However, this process becomes
591 unproductive as soon as co-infection with different mutants occurs under circumstances where
592 recombination is possible. Two different mutants can recombine into the ancestral form of the
593 pathogen and erase the entire benefit of attenuation. This is all the more harmful since the old
594 forms are also, very often, those which spread most easily.

595



596 **Boxed text Figure. Muller's ratchet and recombination** The figure is reprinted from reference
 597 [57]. Genes (capitals) are mutated at random in a different form (low case). Mutations accumulate
 598 ratchet-like because the probability of reversion to the parent form is negligible. This happens
 599 independently for viruses of different descents. However, if viruses from different descent happen
 600 to be in the same cell, they can recombine. This allows them to recreate the ancestral form of the
 601 virus.

602 **References**

- [1] M. Romano, A. Ruggiero, F. Squeglia, G. Maga, R. Berisio, A structural view of SARS-CoV-2 RNA replication machinery: RNA synthesis, proofreading and final capping, *Cells*. 9 (2020) 1267. <https://doi.org/10.3390/cells9051267>.
- [2] A. Lai, A. Bergna, C. Acciarri, M. Galli, G. Zehender, Early phylogenetic estimate of the effective reproduction number of SARS-CoV-2, *J. Med. Virol.* 92 (2020) 675–679. <https://doi.org/10.1002/jmv.25723>.
- [3] R.A. Fisher, *The genetical theory of natural selection*, Clarendon Press, Oxford, 1930. <https://doi.org/10.5962/bhl.title.27468>.
- [4] A. Danchin, P. Marlière, Cytosine drives evolution of SARS-CoV-2, *Environ. Microbiol.* 22 (2020) 1977–1985. <https://doi.org/10.1111/1462-2920.15025>.
- [5] P. Simmonds, Rampant C→U hypermutation in the genomes of SARS-CoV-2 and other coronaviruses: causes and consequences for their short- and long-term evolutionary trajectories, *MSphere*. 5 (2020) e00408-20. <https://doi.org/10.1128/mSphere.00408-20>.
- [6] P.C.Y. Woo, B.H.L. Wong, Y. Huang, S.K.P. Lau, K.-Y. Yuen, Cytosine deamination and selection of CpG suppressed clones are the two major independent biological forces that shape codon usage bias in coronaviruses, *Virology*. 369 (2007) 431–442. <https://doi.org/10.1016/j.virol.2007.08.010>.
- [7] Z. Ou, C. Ouzounis, D. Wang, W. Sun, J. Li, W. Chen, P. Marlière, A. Danchin, A path towards SARS-CoV-2 attenuation: metabolic pressure on CTP synthesis rules the virus evolution, 2020. <https://doi.org/10.1101/2020.06.20.162933>.
- [8] I. Sola, F. Almazán, S. Zúñiga, L. Enjuanes, Continuous and discontinuous RNA synthesis in coronaviruses, *Annu Rev Virol.* 2 (2015) 265–288. <https://doi.org/10.1146/annurev-virology-100114-055218>.
- [9] S. Di Giorgio, F. Martignano, M.G. Torcia, G. Mattiuz, S.G. Conticello, Evidence for host-dependent RNA editing in the transcriptome of SARS-CoV-2, *Sci Adv.* 6 (2020) eabb5813. <https://doi.org/10.1126/sciadv.abb5813>.
- [10] J. Alonso-Carrera, C. de Miguel, B. Manzano, Economic growth and environmental degradation when preferences are non-homothetic, *Environ Resource Econ.* 74 (2019) 1011–1036. <https://doi.org/10.1007/s10640-019-00357-4>.
- [11] Freeman J Dyson, *Origins of life*, Cambridge University Press, Cambridge, UK, 1985.
- [12] K. Wellner, H. Betat, M. Mörl, A tRNA's fate is decided at its 3' end: Collaborative actions of CCA-adding enzyme and RNases involved in tRNA processing and degradation, *Biochim Biophys Acta Gene Regul Mech.* 1861 (2018) 433–441. <https://doi.org/10.1016/j.bbagrm.2018.01.012>.
- [13] P. Shridas, C.J. Waechter, Human dolichol kinase, a polytopic endoplasmic reticulum membrane protein with a cytoplasmically oriented CTP-binding site, *J. Biol. Chem.* 281 (2006) 31696–31704. <https://doi.org/10.1074/jbc.M604087200>.
- [14] C. Wang, Z. Liu, Z. Chen, X. Huang, M. Xu, T. He, Z. Zhang, The establishment of reference sequence for SARS-CoV-2 and variation analysis, *J. Med. Virol.* 92 (2020) 667–674. <https://doi.org/10.1002/jmv.25762>.
- [15] X. Yang, N. Dong, E.W.-C. Chan, S. Chen, Genetic cluster analysis of SARS-CoV-2 and the identification of those responsible for the major outbreaks in various countries, *Emerg Microbes Infect.* 9 (2020) 1287–1299. <https://doi.org/10.1080/22221751.2020.1773745>.
- [16] L. van Dorp, M. Acman, D. Richard, L.P. Shaw, C.E. Ford, L. Ormond, C.J. Owen, J. Pang, C.C.S. Tan, F.A.T. Boshier, A.T. Ortiz, F. Balloux, Emergence of genomic

- diversity and recurrent mutations in SARS-CoV-2, *Infect. Genet. Evol.* 83 (2020) 104351. <https://doi.org/10.1016/j.meegid.2020.104351>.
- [17] Y. Yang, W. Yan, B. Hall, X. Jiang, Characterizing transcriptional regulatory sequences in coronaviruses and their role in recombination, *BioRxiv.* (2020). <https://doi.org/10.1101/2020.06.21.163410>.
- [18] H. Grosjean, V. de Crécy-Lagard, C. Marck, Deciphering synonymous codons in the three domains of life: co-evolution with specific tRNA modification enzymes, *FEBS Lett.* 584 (2010) 252–264. <https://doi.org/10.1016/j.febslet.2009.11.052>.
- [19] S.S. Rout, M. Singh, K.S. Shindler, J. Das Sarma, One proline deletion in the fusion peptide of neurotropic mouse hepatitis virus (MHV) restricts retrograde axonal transport and neurodegeneration, *J. Biol. Chem.* 295 (2020) 6926–6935. <https://doi.org/10.1074/jbc.RA119.011918>.
- [20] E.E. Rivera-Serrano, A.S. Gizzi, J.J. Arnold, T.L. Grove, S.C. Almo, C.E. Cameron, Viperin reveals its true function, *Annu. Rev. Virol.* 7 (2020) annurev-virology-011720-095930. <https://doi.org/10.1146/annurev-virology-011720-095930>.
- [21] J. Armengaud, A. Delaunay-Moisan, J.-Y. Thuret, E. van Anken, D. Acosta-Alvear, T. Aragón, C. Arias, M. Blondel, I. Braakman, J.-F. Collet, R. Courcol, A. Danchin, J.-F. Deleuze, J.-P. Lavigne, S. Lucas, T. Michiels, E.R.B. Moore, J. Nixon-Abell, R. Rossello-Mora, Z.-L. Shi, A.G. Siccardi, R. Sitia, D. Tillett, K.N. Timmis, M.B. Toledano, P. van der Sluijs, E. Vicenzi, The importance of naturally attenuated SARS-CoV-2 in the fight against COVID-19, *Environ. Microbiol.* 22 (2020) 1997–2000. <https://doi.org/10.1111/1462-2920.15039>.
- [22] S. Liu, J. Shen, L. Yang, C.-D. Hu, J. Wan, Distinct genetic spectrums and evolution patterns of SARS-CoV-2, *Health Informatics*, 2020. <https://doi.org/10.1101/2020.06.16.20132902>.
- [23] C.A. Nelson, A. Pekosz, C.A. Lee, M.S. Diamond, D.H. Fremont, Structure and intracellular targeting of the SARS-Coronavirus Orf7a accessory protein, *Structure.* 13 (2005) 75–85. <https://doi.org/10.1016/j.str.2004.10.010>.
- [24] J.-S. Kim, J.-H. Jang, J.-M. Kim, Y.-S. Chung, C.-K. Yoo, M.-G. Han, Genome-wide identification and characterization of point mutations in the SARS-CoV-2 genome, *Osong Public Health Res Perspect.* 11 (2020) 101–111. <https://doi.org/10.24171/j.phrp.2020.11.3.05>.
- [25] A. Addetia, H. Xie, P. Roychoudhury, L. Shrestha, M. Loprieno, M.-L. Huang, K.R. Jerome, A.L. Greninger, Identification of multiple large deletions in ORF7a resulting in in-frame gene fusions in clinical SARS-CoV-2 isolates, *J. Clin. Virol.* 129 (2020) 104523. <https://doi.org/10.1016/j.jcv.2020.104523>.
- [26] S.R. Schaecher, J.M. Mackenzie, A. Pekosz, The ORF7b protein of severe acute respiratory syndrome coronavirus (SARS-CoV) is expressed in virus-infected cells and incorporated into SARS-CoV particles, *J. Virol.* 81 (2007) 718–731. <https://doi.org/10.1128/JVI.01691-06>.
- [27] D. Benvenuto, S. Angeletti, M. Giovanetti, M. Bianchi, S. Pascarella, R. Cauda, M. Ciccozzi, A. Cassone, Evolutionary analysis of SARS-CoV-2: how mutation of Non-Structural Protein 6 (NSP6) could affect viral autophagy, *J. Infect.* 81 (2020) e24–e27. <https://doi.org/10.1016/j.jinf.2020.03.058>.
- [28] S. Angeletti, D. Benvenuto, M. Bianchi, M. Giovanetti, S. Pascarella, M. Ciccozzi, COVID-2019: The role of the nsp2 and nsp3 in its pathogenesis, *J. Med. Virol.* 92 (2020) 584–588. <https://doi.org/10.1002/jmv.25719>.
- [29] M.C. Hagemeijer, I. Monastyrska, J. Griffith, P. van der Sluijs, J. Voortman, P.M. van Bergen en Henegouwen, A.M. Vonk, P.J.M. Rottier, F. Reggiori, C.A.M. de Haan,

- Membrane rearrangements mediated by coronavirus nonstructural proteins 3 and 4, *Virology*. 458–459 (2014) 125–135. <https://doi.org/10.1016/j.virol.2014.04.027>.
- [30] B. Short, A call for oxygen in the ER, *The Journal of Cell Biology*. 203 (2013) 552–552. <https://doi.org/10.1083/jcb.2034iti3>.
- [31] C. Selvaraj, D.C. Dinesh, U. Panwar, R. Abhirami, E. Boura, S.K. Singh, Structure-based virtual screening and molecular dynamics simulation of SARS-CoV-2 Guanine-N7 methyltransferase (nsp14) for identifying antiviral inhibitors against COVID-19, *J. Biomol. Struct. Dyn.* (2020) 1–12. <https://doi.org/10.1080/07391102.2020.1778535>.
- [32] S. Chen, X. Zheng, J. Zhu, R. Ding, Y. Jin, W. Zhang, H. Yang, Y. Zheng, X. Li, G. Duan, Extended ORF8 Gene Region Is Valuable in the Epidemiological Investigation of Severe Acute Respiratory Syndrome-Similar Coronavirus, *J. Infect. Dis.* 222 (2020) 223–233. <https://doi.org/10.1093/infdis/jiaa278>.
- [33] E. Issa, G. Merhi, B. Panossian, T. Salloum, S. Tokajian, SARS-CoV-2 and ORF3a: nonsynonymous mutations, functional domains, and viral pathogenesis, *MSystems*. 5 (2020) e00266-20. <https://doi.org/10.1128/mSystems.00266-20>.
- [34] M. Bolles, E. Donaldson, R. Baric, SARS-CoV and emergent coronaviruses: viral determinants of interspecies transmission, *Current Opinion in Virology*. 1 (2011) 624–634. <https://doi.org/10.1016/j.coviro.2011.10.012>.
- [35] V.M. Corman, H.J. Baldwin, A.F. Tateno, R.M. Zerbinati, A. Annan, M. Owusu, E.E. Nkrumah, G.D. Maganga, S. Oppong, Y. Adu-Sarkodie, P. Vallo, L.V.R.F. da Silva Filho, E.M. Leroy, V. Thiel, L. van der Hoek, L.L.M. Poon, M. Tschapka, C. Drosten, J.F. Drexler, Evidence for an ancestral association of human coronavirus 229E with bats, *J. Virol.* 89 (2015) 11858–11870. <https://doi.org/10.1128/JVI.01755-15>.
- [36] M. Thoms, R. Buschauer, M. Ameismeier, L. Koepke, T. Denk, M. Hirschenberger, H. Kratzat, M. Hayn, T. Mackens-Kiani, J. Cheng, C.M. Stürzel, T. Fröhlich, O. Berninghausen, T. Becker, F. Kirchhoff, K.M.J. Sparrer, R. Beckmann, Structural basis for translational shutdown and immune evasion by the Nsp1 protein of SARS-CoV-2, *Molecular Biology*, 2020. <https://doi.org/10.1101/2020.05.18.102467>.
- [37] S. Laha, J. Chakraborty, S. Das, S.K. Manna, S. Biswas, R. Chatterjee, Characterizations of SARS-CoV-2 mutational profile, spike protein stability and viral transmission, *Infect. Genet. Evol.* 85 (2020) 104445. <https://doi.org/10.1016/j.meegid.2020.104445>.
- [38] C. Iserman, C. Roden, M. Boerneke, R. Sealfon, G. McLaughlin, I. Jungreis, C. Park, A. Boppana, E. Fritch, Y.J. Hou, C. Theesfeld, O.G. Troyanskaya, R.S. Baric, T.P. Sheahan, K. Weeks, A.S. Gladfelter, Specific viral RNA drives the SARS CoV-2 nucleocapsid to phase separate, *Biochemistry*, 2020. <https://doi.org/10.1101/2020.06.11.147199>.
- [39] Y. Cong, M. Ulasli, H. Schepers, M. Mauthe, P. V'kovski, F. Kriegenburg, V. Thiel, C.A.M. de Haan, F. Reggiori, Nucleocapsid protein recruitment to replication-transcription complexes plays a crucial role in coronaviral life cycle, *J Virol.* 94 (2019) e01925-19, [/jvi/94/4/JVI.01925-19.atom](https://doi.org/10.1128/JVI.01925-19). <https://doi.org/10.1128/JVI.01925-19>.
- [40] O.M. Ugurel, O. Ata, D. Turgut-Balik, An updated analysis of variations in SARS-CoV-2 genome, *Turk. J. Biol.* 44 (2020) 157–167. <https://doi.org/10.3906/biy-2005-111>.
- [41] Z. Daniloski, X. Guo, N.E. Sanjana, The D614G mutation in SARS-CoV-2 Spike increases transduction of multiple human cell types, 2020. <https://doi.org/10.1101/2020.06.14.151357>.
- [42] R. Lorenzo-Redondo, H.H. Nam, S.C. Roberts, L.M. Simons, L.J. Jennings, C. Qi, C.J. Achenbach, A.R. Hauser, M.G. Ison, J.F. Hultquist, E.A. Ozer, A unique clade of SARS-CoV-2 viruses is associated with lower viral loads in patient upper airways, 2020. <https://doi.org/10.1101/2020.05.19.20107144>.

- [43] C.C. Posthuma, A.J.W. Te Velthuis, E.J. Snijder, Nidovirus RNA polymerases: Complex enzymes handling exceptional RNA genomes, *Virus Res.* 234 (2017) 58–73. <https://doi.org/10.1016/j.virusres.2017.01.023>.
- [44] S.-Y. Fung, K.-S. Yuen, Z.-W. Ye, C.-P. Chan, D.-Y. Jin, A tug-of-war between severe acute respiratory syndrome coronavirus 2 and host antiviral defence: lessons from other pathogenic viruses, *Emerg Microbes Infect.* 9 (2020) 558–570. <https://doi.org/10.1080/22221751.2020.1736644>.
- [45] K.A. Ivanov, J. Ziebuhr, Human coronavirus 229E nonstructural protein 13: characterization of duplex-unwinding, nucleoside triphosphatase, and RNA 5'-triphosphatase activities, *J. Virol.* 78 (2004) 7833–7838. <https://doi.org/10.1128/JVI.78.14.7833-7838.2004>.
- [46] M. Letko, S.N. Seifert, K.J. Olival, R.K. Plowright, V.J. Munster, Bat-borne virus diversity, spillover and emergence, *Nat. Rev. Microbiol.* 18 (2020) 461–471. <https://doi.org/10.1038/s41579-020-0394-z>.
- [47] S. Elbe, G. Buckland-Merrett, Data, disease and diplomacy: GISAID's innovative contribution to global health: Data, Disease and Diplomacy, *Global Challenges.* 1 (2017) 33–46. <https://doi.org/10.1002/gch2.1018>.
- [48] J. Hadfield, C. Megill, S.M. Bell, J. Huddleston, B. Potter, C. Callender, P. Sagulenko, T. Bedford, R.A. Neher, Nextstrain: real-time tracking of pathogen evolution, *Bioinformatics.* 34 (2018) 4121–4123. <https://doi.org/10.1093/bioinformatics/bty407>.
- [49] R.I. Amann, S. Baichoo, B.J. Blencowe, P. Bork, M. Borodovsky, C. Brooksbank, P.S.G. Chain, R.R. Colwell, D.G. Daffonchio, A. Danchin, V. de Lorenzo, P.C. Dorrestein, R.D. Finn, C.M. Fraser, J.A. Gilbert, S.J. Hallam, P. Hugenholtz, J.P.A. Ioannidis, J.K. Jansson, J.F. Kim, H.-P. Klenk, M.G. Klotz, R. Knight, K.T. Konstantinidis, N.C. Kyrpides, C.E. Mason, A.C. McHardy, F. Meyer, C.A. Ouzounis, A.A.N. Patrinos, M. Podar, K.S. Pollard, J. Ravel, A.R. Muñoz, R.J. Roberts, R. Rosselló-Móra, S.-A. Sansone, P.D. Schloss, L.M. Schriml, J.C. Setubal, R. Sorek, R.L. Stevens, J.M. Tiedje, A. Turjanski, G.W. Tyson, D.W. Ussery, G.M. Weinstock, O. White, W.B. Whitman, I. Xenarios, Toward unrestricted use of public genomic data, *Science.* 363 (2019) 350–352. <https://doi.org/10.1126/science.aaw1280>.
- [50] I. Karsch-Mizrachi, T. Takagi, G. Cochrane, International Nucleotide Sequence Database Collaboration, The international nucleotide sequence database collaboration, *Nucleic Acids Res.* 46 (2018) D48–D51. <https://doi.org/10.1093/nar/gkx1097>.
- [51] K.D. Yamada, K. Tomii, K. Katoh, Application of the MAFFT sequence alignment program to large data-reexamination of the usefulness of chained guide trees, *Bioinformatics.* 32 (2016) 3246–3251. <https://doi.org/10.1093/bioinformatics/btw412>.
- [52] B.Q. Minh, H.A. Schmidt, O. Chernomor, D. Schrempf, M.D. Woodhams, A. von Haeseler, R. Lanfear, IQ-TREE 2: New models and efficient methods for phylogenetic inference in the genomic era, *Mol. Biol. Evol.* 37 (2020) 1530–1534. <https://doi.org/10.1093/molbev/msaa015>.
- [53] P. Sagulenko, V. Puller, R.A. Neher, TreeTime: Maximum-likelihood phylodynamic analysis, *Virus Evol.* 4 (2018) vex042. <https://doi.org/10.1093/ve/vex042>.
- [54] Estimation of the number of nucleotide substitutions in the control region of mitochondrial DNA in humans and chimpanzees., *Molecular Biology and Evolution.* (1993). <https://doi.org/10.1093/oxfordjournals.molbev.a040023>.
- [55] H.J. Muller, Some genetic aspects of sex, *The American Naturalist.* 66 (1932) 118–138. <https://doi.org/10.1086/280418>.
- [56] K.A. Smith, Louis Pasteur, the father of immunology?, *Front Immunol.* 3 (2012) 68. <https://doi.org/10.3389/fimmu.2012.00068>.

- [57] A. Danchin, K. Timmis, SARS-CoV-2 variants: Relevance for symptom granularity, epidemiology, immunity (herd, vaccines), virus origin and containment?, *Environ. Microbiol.* 22 (2020) 2001–2006. <https://doi.org/10.1111/1462-2920.15053>.

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