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## Phenotypic Differences Between Polygenic and Monogenic Hypobetalipoproteinemia

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1 **GPR146 gene variants are associated with reduced plasma lipids and metabolic health:**  
2 **A novel role for GPR146 in hypolipidemia**

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46 **ABSTRACT**

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## BACKGROUND

In mice, G-protein coupled receptor 146 (GPR146) deficiency causes hypolipidemia and protects against atherosclerosis. We here aim to evaluate whether a loss of *GPR146* in humans results in a similarly beneficial phenotype.

## METHODS

We made use of common and rare genetic variants in the *GPR146* gene locus, as research instruments in population cohorts (UK-Biobank, >500 000 participants; Lifelines, ~165 000 participants and The Copenhagen-City Heart Study ~20 000) and in subjects with familial hypobetalipoproteinemia (FHBL).

## RESULTS

Carriers of the common variant rs2362529 present with gene dose-dependent decreased low-density lipoprotein cholesterol (LDL-C) and apolipoprotein B. Further delineating the role of GPR146 in lipid metabolism, we show that the exchange of a highly conserved proline for a leucine residue at position (p.Pro62Leu; predicted to be damaging), present in 2615 participants of the UK-Biobank cohort, is associated with markedly lower LDL-C, HDL-C, but also triglycerides, compared to non-carriers. Remarkably, carriers of this variant also show reduced plasma levels of liver enzymes and C-reactive protein. In a family with FHBL, we finally show that the p.Pro62Leu variant segregates with low LDL-C.

## CONCLUSIONS

This study shows that carriers of genetic variants in *GPR146* present with a beneficial lipidic and metabolic profile, low levels of LDL-C and reduced plasma liver enzymes which support the development of GPR146 antagonists against cardiometabolic diseases. The data furthermore support the hypothesis that loss of GPR146 is a novel cause of FHBL.

## 1 INTRODUCTION

2  
3 Carriers of mutations in *APOB*, *MTTP*, *PCSK9* and *ANGPTL3* are all characterized by  
4 reduced plasma cholesterol levels and are protected from atherosclerosis<sup>1-5</sup>. When studying these  
5 genes in individuals with extremely low levels of low-density lipoprotein cholesterol (LDL-C), we  
6 previously showed that in 30% of the cases, we have no explanation for this phenotype<sup>6,7</sup>, indicating  
7 that there is still room for new discoveries. The fact that genome-wide association studies (GWAS)  
8 (re)identified the above-mentioned key players in the metabolism of very low-density lipoprotein  
9 (VLDL), indicates that newly discovered genomic loci may play equally important roles.

10  
11 One of the GWAS hits in 2013 was *GPR146*<sup>8</sup>, encoding the G-protein coupled receptor 146.  
12 More specifically, a common DNA variation (rs1997243) in close proximity to the *GPR146* gene,  
13 present in 15% of individuals in the general population, was found to be associated with increased  
14 plasma levels of total cholesterol. Han *et al.* later showed that this increase in plasma cholesterol is  
15 caused by increased levels of *GPR146* mRNA<sup>9</sup>, indicating that the encoded protein is directly  
16 involved in the regulation of cholesterol metabolism. Data in the public domain show that *GPR146* is  
17 mostly expressed in white adipose tissue where it has been reported to play a role in adipocyte  
18 differentiation<sup>10</sup> but this is unlikely to explain its effect on plasma cholesterol. In mice, we have  
19 provided evidence that loss of *Gpr146* in the liver reduces plasma cholesterol levels<sup>11</sup>. Murine  
20 *Gpr146* deficiency was furthermore shown to decrease the hepatic secretion of VLDL, the precursor  
21 of LDL, and to protect against diet-induced atherosclerosis in LDL receptor knock-out mice.  
22 Considering that GPRs are druggable targets<sup>12</sup>, the latter finding nourished hope for the development  
23 of small molecules to inhibit GPR146 to improve the management of dyslipidemia and  
24 atherosclerosis, particularly in patients suffering from homozygous familial hypercholesterolemia  
25 (hoFH), lacking the LDL receptor<sup>13,14</sup>. The effects of GPR146 on plasma lipids in mice are  
26 associated with the fasting-refed state<sup>11</sup>, but it is not known what the natural ligand for cellular  
27 GPR146 activation is. C-peptide, a cleavage product from insulin has been reported to play a role<sup>15</sup>  
28 but this was later disputed<sup>16</sup>, and GPR146 remains an orphan receptor to date.

29  
30 The current study provides a positive answer to the key question whether loss of *GPR146* in  
31 humans is associated with reduced plasma cholesterol **as main factor for cardiovascular disease**. We  
32 show that genetic impairment of GPR146 is a novel genetic cause of low LDL-C, similar to reports  
33 for mutations in *APOB*, *MTTP*, *ANGPTL3* and *PCSK9*, all targets for pharmaceutical intervention to  
34 lower plasma cholesterol<sup>17</sup>. Our findings support the idea that GPR146 is a relevant target to reduce  
35 plasma cholesterol **with concomitant positive effects on liver function**.

## METHODS

The data that support the findings of this study are available from the corresponding author upon reasonable request.

### Association between genetic markers and *GPR146* gene expression

Expression quantitative trait loci (eQTL) analyses aim to connect genetic variants with the expression of genes, or specifically of nearby genes, in case of cis-eQTL. We used the eQTL dataset from the eQTLGen consortium<sup>18</sup>, the largest eQTL repository available so far (**supplementary materials**). We tested the association of rs2362529 and rs1997243 with the expression of nearby genes including *GPR146* in blood. Additionally, we studied the effects of rs2362529 and rs1997243 on *GPR146* expression in the human liver the eQTL dataset from GTEx (V8 release, <https://gtexportal.org/>)<sup>19</sup>.

### Genetic associations with plasma circulating biomarkers in the UK Biobank

The UK Biobank study is a population-based prospective cohort in the United Kingdom in which approximately 500,000 individuals aged between 40 and 69 years were recruited from 2006 through 2010. All participants have given informed consent for this study. The UK Biobank has ethical approval from North West - Haydock Research Ethics Committee (REC reference: 16/NW/0274). Details of the UK Biobank study have been described previously<sup>20</sup>. This research has been conducted using the UK Biobank resource under application number 15031. Clinical characteristics and biomarkers of individuals, variant genotyping, imputation and quality controls as well as genetic and statistical analyses are described in **supplementary materials, Tab.S1 and Fig.S2**.

### Genetic association with coronary artery disease

Genetic association with coronary artery disease was assessed by using summary data from the meta-analysis published by Nikpay et al.<sup>21</sup> (<http://www.cardiogramplusc4d.org/>). In order to test the relevance of these effects, we compared it with well-established LDL-C associated variants (as presented by Ference *et al.* in the consensus statement from the European Atherosclerosis Society Consensus Panel<sup>17</sup>) **using data from van der Harst et al.**<sup>22</sup>.

### Selection of individuals with extreme LDL-C and targeted sequencing

Individuals with extreme dyslipidemia are likely to carry rare genetic variants (loss or gain of function variants) in genes with large effects on lipid metabolism, and we set out to find such mutations in *GPR146* in two large population-based prospective cohort studies: the Lifelines (n=167,729 individuals from the Netherlands<sup>6,23,24</sup>) and the Copenhagen City Heart Study (CCHS n=20,000 individuals from Denmark<sup>25,26</sup>). We selected **unrelated** individuals with extremely low LDL-C (n=222 from Lifelines and n=195 from CCHS, LDL-C < 1<sup>st</sup> and 2<sup>nd</sup> percentiles respectively) and extremely high LDL-C (n=129 from Lifelines and n=194 from CCHS, LDL-C > 99<sup>th</sup> and 98<sup>th</sup> percentiles respectively) (**Tab.S2 and S3**). The next generation sequencing platform and analysis workflow that were used are detailed in the **supplementary materials**.

The proband of a familial case with hypocholesterolemia (LDL-C <5<sup>th</sup> percentile for age and **sex**) was recruited in the context of the HYPOCHOL trial (ClinicalTrials ID: NCT02354079). The genetic screening was performed using the DysliSEQ custom NGS panel as described previously<sup>27</sup>.

1 **Statistics and plots**

2 Statistical analyses and tools used for plotting are described in the **supplementary materials**.

3

## 1 RESULTS

### 2 3 COMMON VARIANTS IN THE *GPR146* GENE LOCUS

#### 4 5 *Rs1997243, rs2362529 and GPR146 mRNA expression*

6  
7 GPR146 is ubiquitously expressed but its expression in the liver has been shown to dictate changes  
8 in plasma lipids in *Gpr146*<sup>-/-</sup> mice. With this in mind we first consulted GPR146 expression in the  
9 human liver using the eQTL dataset from GTEx.

10  
11 We first studied rs1997243, the genetic variant that has led to the identification of *GPR146* as a  
12 candidate lipid gene<sup>8,9,11,28</sup> which is present in 14% of the general population. As can be appreciated  
13 from **Fig.S1A**, the alternative G-allele of rs1997243 tends to increase GPR146 expression as  
14 previously shown by Han *et al.*<sup>9</sup>. Following close examination of the *GPR146* gene locus, we noticed  
15 a second common *GPR146* variant, annotated as rs2362529. This variant has not been studied  
16 previously, and is present with a prevalence of 21% of the general population. Opposite to the  
17 findings of the alternative allele of rs1997243, the C-allele of rs2362529 tends instead to decrease  
18 *GPR146* expression (**Fig.S1B**). In both cases, however, these changes did not reach statistical  
19 significance which is likely related to the small number of samples (n=208) in this dataset.

20  
21 To further study GPR146 expression in carriers of both common variants, we made use of the fact that  
22 GPR146 is also expressed in circulating blood cells and made use of the eQTL dataset from the  
23 eQTLGen consortium which comprises data of 31,684 human subjects. In line with findings in  
24 human liver, carriers of the rs1997243-G allele exhibit higher *GPR146* expression levels in  
25 circulating blood than non carriers (p=3.3e<sup>-310</sup>). In contrast carriers of the rs2362529-C allele exhibit  
26 lower *GPR146* mRNA expression in blood than non carriers (p=1.8e<sup>-21</sup>) (**Fig.1A**).

27  
28 The two above described common *GPR146* variants are not in linkage disequilibrium ( $r^2=0.0172$  in  
29 the 1000 genomes population<sup>29</sup>). This indicates that rs1997243 and rs2362529 can be regarded as  
30 independent genetic research instruments with opposite effects on *GPR146* gene expression.

#### 31 32 *Rs1997243 and rs2362529, lipid traits, metabolic traits and CRP*

33  
34 To study the phenotypes of carriers of rs1997243 or rs2362529, we used the UK Biobank patients  
35 who passed our selection criteria (n=412,444 participants; **Tab.S1, Fig.S2**).

#### 36 37 *Plasma lipid traits*

38 **Fig.1B** shows that carriers of the rs1997243-G allele (heterozygotes and homozygotes combined)  
39 present with increased total cholesterol, LDL-C, HDL-C, with concomitant increases in  
40 apolipoproteins compared to non-carriers, without changes in triglycerides and lipoprotein (a)  
41 (Lp(a)) (**Tab.S4**). These effects were found to be gene-dose dependent (**Fig.S3A** and **Tab.S5**).

42  
43 In carriers of the rs2362529-C allele, we observed the exact opposite, *i.e.* reduced levels of total  
44 cholesterol, LDL-C and HDL-C with concomitant reductions in apolipoproteins and no changes in  
45 plasma triglycerides and Lp(a) compared to non-carriers (**Fig.1B** - **Tab.S4**). Just like for the other  
46 variant the effects were gene dose-dependent (**Fig.S3B** - **Tab.S5**).

#### 47 48 *Metabolic traits and C-reactive protein*

49 Although it has been reported that GPR146 plays a role in the differentiation of adipocytes<sup>10</sup>, carriers  
50 of rs1997243-G or rs2362529-C do not show differences in body mass index (BMI) compared to

1 non-carriers (data not shown). Compared to non-carriers, carriers of the rs1997243-G allele are, to  
2 our surprise, characterized by increased levels of circulating gamma-glutamyltransferase, alkaline  
3 phosphatase, aspartate aminotransferase while the opposite is true for carriers of the rs2362529-C  
4 allele (**Fig.1C - Tab.S4,5**).

5 In addition, carriers of rs1997243-G or rs2362529-C allele(s) have increased or decreased  
6 levels of C-reactive protein (CRP) plasma levels, compared to non-carriers, respectively (**Fig.1D -**  
7 **Tab.S4,5**). Finally, carriers of the rs2362529-C allele present with lower glycated haemoglobin  
8 (HbA<sub>1C</sub>) levels than non-carriers (**Fig.1E - Tab.S4,5**).

9  
10 These results indicate that carriers of the rs1997243-G or rs2362529-C allele(s) have unfavorable or  
11 favourable lipid and metabolic profiles, respectively. Combined, these findings prompt the question  
12 whether these differences translate into an altered risk of coronary artery disease (CAD).

### 13 14 ***Rs1997243, rs2362529 and risk of CAD***

15  
16 In a meta-analysis from the CARDIoGRAMplusC4D Consortium, carriers of the rs2362529-C allele  
17 show a **trend towards a** decreased risk of CAD compared to non-carriers (**Fig.2A**). The reduction in  
18 CAD risk in carriers of *rs2362529* appears marginal but this was anticipated when considering the  
19 magnitude of the effect size of this variant on LDL-C levels: homozygotes for *rs2362529* show an  
20 average decrease of 2.24 mg/dl compared to carriers of the wild-type allele. To study this further, we  
21 investigated the effects of both *GPR146* gene variants on LDL-C and CAD in the context of well  
22 characterized genetic variants in *APOB*, *PCSK9*, *HMGCR*, *ANGPTL3*<sup>17</sup>. As can be appreciated from  
23 **Fig.2B**, *GPR146 rs18997243* and *rs2362529* fit the regression line between the magnitude of LDL-C  
24 changes and the magnitude of effects on CAD **data from van der Harst et al.**<sup>22</sup>

### 25 26 27 **RARE VARIANTS IN THE *GPR146* GENE**

#### 28 29 ***Identification of a missense variant (GPR146-p.Pro62Leu)***

30  
31 With the finding that common *GPR146* variants are associated with opposite effects on *GPR146*  
32 expression and plasma lipid traits, we set out to identify *GPR146* mutations with large effects on  
33 plasma levels of LDL-C. Taken that *Gpr146* knockout mice have reduced LDL-C and are protected  
34 against atherosclerosis<sup>11</sup>, we hypothesized that loss or gain-of-function variants could be found in  
35 individuals with low and high LDL-C, respectively. In this effort, we sequenced genomic DNA of  
36 individuals with extreme LDL-C plasma levels in the Lifelines and CCHS cohorts (**Tab.S2 and S3**).

37  
38 Following targeted sequencing, we selected rare variants which affect the protein sequence (or  
39 splicing regions) of *GPR146* (**Tab.S6**). Most of these rare variants were singletons, however, one  
40 missense variant, annotated as rs151124717, was found eight times in individuals with low LDL-C  
41 and only once in an individual with high LDL-C (**Fig.3A**). The nucleotide substitution (c.185C>T)  
42 results in the exchange of Proline for a Leucine residue at position 62 of the mature GPR146 protein  
43 (p.Pro62Leu) (**Fig.3B**). The missense variant is located in the first intracellular loop of GPR146 and  
44 affects an highly evolutionary conserved protein domain (**Fig.3C - Fig.S4A**). It is predicted to be  
45 damaging for the protein function by seven *in silico* algorithms (**Fig.S4B**).

46  
47 Carriers of the *GPR146*-p.Pro62Leu variant in the Lifelines and CCHS cohorts were all  
48 heterozygotes and did not carry mutations in genes that are known to cause familial forms of  
49 hypocholesterolemia (*APOB*, *MTTP*, *PCSK9*, *ANGPTL3*) or hypercholesterolemia (*LDLR*, *APOB*,



1 *LDLRAP1, APOE, PCSK9*). In a next step we screened the UK Biobank for carriers of the *GPR146*-  
2 p.Pro62Leu to examine their cardiometabolic phenotype.

### 4 *Carriers of GPR146-p.Pro62Leu in the UK Biobank*

#### 6 *Plasma lipid traits*

7 In the UK Biobank, we identified 2,615 carriers of the *GPR146*-p.Pro62Leu variant (**Tab.S7**) which  
8 renders an allele frequency of 0.3% of the general population (in line with data from gnomAD,  
9 <https://gnomad.broadinstitute.org/>). In a direct comparison with carriers of the common rs2362529-  
10 C allele, it is clear that carriers of the *GPR146*-p.Pro62Leu present with larger reductions of total  
11 cholesterol, LDL-C and HDL-C, suggesting that this variant impairs the function of the protein as  
12 predicted (**Fig.3D, Tab.S4, Fig.S4B**).

13 Supporting, the relation between *GPR146* gene variants and atherosclerosis, the magnitude of  
14 the LDL-C lowering effect of *GPR146*-p.Pro62Leu (rs151124717) again correlates with the expected  
15 effect on CAD (**Fig.2B**). The effects of the *GPR146*-p.Pro62Leu variant on CAD did, however, not  
16 reach statistical significance, likely due to large standard errors.

#### 18 *Metabolic traits and CRP*

19 Carriers of the *GPR146*-p.Pro62Leu variant presented with significant reductions in gamma-  
20 glutamyltransferase and alkaline phosphatase when compared to non-carriers (**Fig.3E - Tab.S4**). In  
21 addition, carriers of the *GPR146*-p.Pro62Leu also presented with significantly reduced CRP levels  
22 when compared to non-carriers (**Fig.3F and Tab.S4**).

24 Combined, these data illustrate that carriers of the *GPR146*-p.Pro62Leu variant are characterized by  
25 and overall beneficial cardiometabolic phenotype. The remarkable lipid profile of individuals  
26 carrying this variant, i.e. reductions in all major plasma lipids (**Fig.3D**), resembles those suffering  
27 from familial hypocholesterolemia<sup>30</sup>, which prompted us to sequence the *GPR146* gene in familial  
28 cases with unexplained hypocholesterolemia.

### 30 *Segregation of GPR146 p.Pro62Leu with hypocholesterolemia*

32 In a familial case of hypobetalipoproteinemia from the HYPOCHOL clinical trial<sup>31</sup>, in which  
33 mutations in canonical lipid genes were excluded, one proband was found to be heterozygous for the  
34 *GPR146*-p.Pro62Leu variant (**Fig.4**). This 24 year-old woman presented with very low levels of total  
35 cholesterol (124 mg/dL), LDL-C (59 mg/dL), apoB (60 mg/dL) and triglycerides (50 mg/dL), but  
36 normal HDL-C (55 mg/dL). We recruited six family members and tested the co-segregation of the  
37 *GPR146*-p.Pro62Leu variant with plasma LDL-C levels in this family (**Fig.4 - Tab.S8**). The results  
38 show the cosegregation of the genotype with the hypobetalipoproteinemia phenotype: all three  
39 individuals carrying the *GPR146*-p.Pro62Leu variant present with LDL-C <5<sup>th</sup> percentile for age and  
40 sex<sup>32</sup>. A broader screening of *GPR146* in FHBL cases is needed in order to further support *GPR146*  
41 as a monogenic origin of FHBL.

## 1 DISCUSSION

2  
3 The key novel finding in this study is that carriers of new genetic variants affecting *GPR146* are  
4 characterized by low levels of plasma lipids. More specifically, we show that carriers of a common  
5 variant in the *GPR146* locus (rs2362529) have lower plasma levels of LDL-C, HDL-C, apoB, and  
6 apoAI with more pronounced effect in homozygotes compared to heterozygotes. The observed  
7 reduction in LDL-C in these carriers is shown to be concordant with the reduction in risk of CAD.  
8 These finding, combined with the notion that *Gpr146* deficient mice also have reduced LDL-C and  
9 HDL-C<sup>9,11</sup>, have led us to hypothesize that mutations in *GPR146* may constitute a novel cause of  
10 hypolipidemia. Support for this concept comes from carriers of a rare coding variant, *GPR146*-  
11 p.Pro62Leu (predicted to be damaging for GPR146 functions), who are indeed characterized by  
12 reduced LDL-C, HDL-C but also triglycerides compared to non-carriers. Segregation of this rare  
13 variant with low LDL-C in a family with hypocholesterolemia provides further support for a novel  
14 role for GPR146 in human hypolipidemia.

15  
16 Previous studies have already shown that GPR146 is associated with increased levels of plasma  
17 lipids<sup>8,28,33</sup>. Corroborating and extending these findings, we here show that carriers of rs1997243 or  
18 rs2362529, common variants in the *GPR146* locus, are characterized by opposite changes in plasma  
19 lipids. These findings bear similarity to opposite lipid profiles in carriers of rare variants in the  
20 *APOB* and *PCSK9* genes: loss-of-function mutations in *APOB* and *PCSK9* cause  
21 hypobetalipoproteinemia, while gain-of-function mutations cause hypercholesterolemia<sup>34</sup>.

22  
23 In a next step, we set out to identify *GPR146* mutations with larger effect sizes. In two white  
24 Caucasian general population samples, we found that *GPR146*-p.Pro62Leu was overrepresented in  
25 subjects with low LDL-C compared to individuals with high LDL-C. The proline residue at position  
26 62 is highly evolutionary conserved and accordingly, substitution to a leucine residue is predicted to  
27 be deleterious on the function of GPR146. Notably, carriers of the rare *GPR146*-p.Pro62Leu variant  
28 are characterized by stronger reductions in plasma lipids compared to carriers of the common  
29 rs2362529 variant which suggests that *GPR146*-p.Pro62Leu may be a functional mutation. This is  
30 further supported by the cosegregation of the *GPR146*-p.Pro62Leu heterozygous variant with a low  
31 LDL-C phenotype in a family with hypocholesterolemia. **However further functional investigations  
32 are needed to proof the functional impact of this variant in in-vitro/vivo models.**

33  
34 In contrast to products of established genes in lipid metabolism such as *APOB*, *MTTP*, *APOC3*,  
35 *ANGPTL3*, and *LDLR*, the GPR146 protein is not directly involved in the machinery to synthesize  
36 lipoproteins or to clear them from the circulation. It is, in fact, the first protein known to date to  
37 control plasma lipid traits through cellular signaling pathways: GPR146 has previously been shown  
38 to increase *de novo* cholesterol synthesis in the liver through activation of the extracellular signal-  
39 regulated kinase signaling pathway<sup>11</sup>. The downstream effect on hepatic VLDL secretion was shown  
40 to contribute to the plasma lipid phenotype of *Gpr146* knock out mice. Whether these findings  
41 translate to humans needs to be established.

42  
43 Using two common *GPR146* variants and one rare (*GPR146*-p.Pro62Leu) variant as genetic  
44 instruments, the current study shows that *GPR146* is also associated with changes in circulating  
45 levels of hepatic enzymes and HbA<sub>1C</sub>. When considering the previously suggested development of  
46 GPR146 antagonists<sup>9,11,13,14</sup>, the current genetic data show that no adverse effects on the onset of  
47 type 2 diabetes, as described for genetic variants in *HMGCR* (the target of statins) and *PCSK9*  
48 (targets of evolocumab and alirocumab), are anticipated<sup>35-37</sup>. In addition, the association of GPR146  
49 with plasma levels of CRP, a biomarker for systemic inflammation, is of particular interest<sup>38,39</sup> when  
50 considering the central role of inflammation in atherogenesis<sup>40</sup>. Finally, loss of GPR146 function is

1 not associated with elevated hepatic enzymes, commonly used as markers of hepatic steatosis or  
2 NAFLD, an adverse effect of Mipomersen (an *APOB* antisense oligonucleotide) and Lomitapide (a  
3 small molecule MTP inhibitor) used to treat hoFH<sup>41</sup>. In this regard we think it is of interest to note  
4 that, in mice, the protection against atherosclerosis is independent of the presence of the LDL  
5 receptor, which makes GPR146 an interesting therapeutic target for this group of patients<sup>11</sup>.

6  
7 The current study presents *GPR146* as a novel candidate gene for human hypolipidemia. The  
8 findings support the development of strategies to antagonize GPR146 to reduce plasma levels of  
9 cholesterol and atherosclerosis. This could be achieved through hepatic mRNA silencing approaches  
10 such as employed for PCSK9, ANGPTL3, APOC3 and LPA<sup>42</sup> (<https://clinicaltrials.gov/>) or through  
11 targeting the genomic DNA in hepatocytes with CRISPR<sup>43</sup>. As a G-coupled protein receptor,  
12 GPR146 also offers the option to find small molecule inhibitors but at this point deorphanization of  
13 GPR146 is warranted to unravel the molecular mechanisms that underlie the observations in this  
14 study.

### 15 **Limitations**

16 Despite great efforts by our groups, we have not been able to provide evidence that *GPR146*-  
17 p.Pro62Leu is a functional mutation. Our *in vitro* as well as *in vivo* systems have been inadequate to  
18 address the functionality of GPR146 variants. This is clearly complicated by the notion that the  
19 ligand for GPR146 as well as specific readouts of GPR146 activity remains to be identified.  
20 However, the circumstantial evidence that we provide in this study combined with the clearcut  
21 evidence that *Gpr146*<sup>-/-</sup> mice present with lower plasma lipids, makes, in our opinion, a strong case  
22 of the actual involvement of attenuated GPR146 function in low levels of plasma lipids and a  
23 favourable metabolic phenotype in humans. Hopefully the consequences of human genetic *GPR146*  
24 deficiency will soon give the required unequivocal evidence. It is in this regard good to note that  
25 whole body ablation of *Gpr146* in mice is compatible with life.  
26  
27

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32  
33 The authors have nothing to disclose

## 34 **LIST OF SUPPLEMENTAL MATERIALS**

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38 **Online References S1-S16**

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## 1 REFERENCES

- 2  
3 1. Peloso GM, Nomura A, Khera AV, Chaffin M, Won H-H, Ardissino D, Danesh J, Schunkert H,  
4 Wilson JG, Samani N, Erdmann J, McPherson R, Watkins H, Saleheen D, McCarthy S,  
5 Teslovich TM, Leader JB, Lester Kirchner H, Marrugat J, Nohara A, Kawashiri M-A, Tada H,  
6 Dewey FE, Carey DJ, Baras A, Kathiresan S. Rare Protein-Truncating Variants in APOB,  
7 Lower Low-Density Lipoprotein Cholesterol, and Protection Against Coronary Heart Disease.  
8 *Circ Genomic Precis Med.* 2019;12:e002376.
- 9 2. Musunuru K, Pirruccello JP, Do R, Peloso GM, Guiducci C, Sougnez C, Garimella KV, Fisher  
10 S, Abreu J, Barry AJ, Fennell T, Banks E, Ambrogio L, Cibulskis K, Kernysky A, Gonzalez E,  
11 Rudzicz N, Engert JC, DePristo MA, Daly MJ, Cohen JC, Hobbs HH, Altshuler D, Schonfeld  
12 G, Gabriel SB, Yue P, Kathiresan S. Exome sequencing, ANGPTL3 mutations, and familial  
13 combined hypolipidemia. *N Engl J Med.* 2010;363:2220–2227.
- 14 3. Stitzel NO, Peloso GM, Abifadel M, Cefalu AB, Fouchier S, Motazacker MM, Tada H, Larach  
15 DB, Awan Z, Haller JF, Pullinger CR, Varret M, Rabès J-P, Noto D, Tarugi P, Kawashiri M-A,  
16 Nohara A, Yamagishi M, Risman M, Deo R, Ruel I, Shendure J, Nickerson DA, Wilson JG,  
17 Rich SS, Gupta N, Farlow DN, Neale BM, Daly MJ, Kane JP, Freeman MW, Genest J, Rader  
18 DJ, Mabuchi H, Kastelein JJP, Hovingh GK, Averna MR, Gabriel S, Boileau C, Kathiresan S.  
19 Exome sequencing in suspected monogenic dyslipidemias. *Circ Cardiovasc Genet.*  
20 2015;8:343–350.
- 21 4. Wetterau JR, Aggerbeck LP, Bouma ME, Eisenberg C, Munck A, Hermier M, Schmitz J, Gay  
22 G, Rader DJ, Gregg RE. Absence of microsomal triglyceride transfer protein in individuals  
23 with abetalipoproteinemia. *Science.* 1992;258:999–1001.
- 24 5. Abifadel M, Varret M, Rabès J-P, Allard D, Ouguerram K, Devillers M, Cruaud C, Benjannet  
25 S, Wickham L, Erlich D, Derré A, Villéger L, Farnier M, Beucler I, Bruckert E, Chambaz J,  
26 Chanu B, Lecerf J-M, Luc G, Moulin P, Weissenbach J, Prat A, Krempf M, Junien C, Seidah  
27 NG, Boileau C. Mutations in PCSK9 cause autosomal dominant hypercholesterolemia. *Nat*  
28 *Genet.* 2003;34:154–156.
- 29 6. Balder J-W, Rimbart A, Zhang X, Viel M, Kanninga R, van Dijk F, Lansberg P, Sinke R,  
30 Kuivenhoven JA. Genetics, Lifestyle, and Low-Density Lipoprotein Cholesterol in Young and  
31 Apparently Healthy Women. *Circulation.* 2018;137:820–831.
- 32 7. Rimbart A, Vanhoye X, Coulibaly D, Marrec M, Pichelin M, Charrière S, Peretti N, Valéro R,  
33 Wargny M, Carrié A, Lindenbaum P, Deleuze J-F, Genin E, Redon R, Rollat-Farnier PA, Goxe  
34 D, Degraef G, Marmontel O, Divry E, Bigot-Corbel E, Moulin P, Cariou B, Di Filippo M.  
35 Phenotypic Differences Between Polygenic and Monogenic Hypobetalipoproteinemia.  
36 *Arterioscler Thromb Vasc Biol.* 2021;41:e63–e71.
- 37 8. Willer CJ, Schmidt EM, Sengupta S, Peloso GM, Gustafsson S, Kanoni S, Ganna A, Chen J,  
38 Buchkovich ML, Mora S, Beckmann JS, Bragg-Gresham JL, Chang H-Y, Demirkan A, Den  
39 Hertog HM, Do R, Donnelly LA, Ehret GB, Esko T, Feitosa MF, Ferreira T, Fischer K,  
40 Fontanillas P, Fraser RM, Freitag DF, Gurdasani D, Heikkilä K, Hyppönen E, Isaacs A,  
41 Jackson AU, Johansson Å, Johnson T, Kaakinen M, Kettunen J, Kleber ME, Li X, Luan J,  
42 Lyytikäinen L-P, Magnusson PKE, Mangino M, Mihailov E, Montasser ME, Müller-Nurasyid  
43 M, Nolte IM, O’Connell JR, Palmer CD, Perola M, Petersen A-K, Sanna S, Saxena R, Service  
44 SK, Shah S, Shungin D, Sidore C, Song C, Strawbridge RJ, Surakka I, Tanaka T, Teslovich

- 1 TM, Thorleifsson G, Van den Herik EG, Voight BF, Volcik KA, Waite LL, Wong A, Wu Y,  
2 Zhang W, Absher D, Asiki G, Barroso I, Been LF, Bolton JL, Bonnycastle LL, Brambilla P,  
3 Burnett MS, Cesana G, Dimitriou M, Doney ASF, Döring A, Elliott P, Epstein SE, Ingi  
4 Eyjolfsson G, Gigante B, Goodarzi MO, Grallert H, Gravito ML, Groves CJ, Hallmans G,  
5 Hartikainen A-L, Hayward C, Hernandez D, Hicks AA, Holm H, Hung Y-J, Illig T, Jones MR,  
6 Kaleebu P, Kastelein JJP, et al. Discovery and refinement of loci associated with lipid levels.  
7 *Nat Genet.* 2013;45:1274–1283.
- 8 9. Han F, Liu X, Chen C, Liu Y, Du M, Zhou Y, Liu Y, Song B-L, He HH, Wang Y.  
9 Hypercholesterolemia risk-associated GPR146 is an orphan G-protein coupled receptor that  
10 regulates blood cholesterol levels in humans and mice. *Cell Res.* 2020;30:363–365.
- 11 10. [https://www.easd.org/virtualmeeting/home.html#!resources/crispr-cas9-induced-knockout-of-](https://www.easd.org/virtualmeeting/home.html#!resources/crispr-cas9-induced-knockout-of-gpr146-in-mouse-3t3-l1-fibroblasts-blunted-differentiation-into-adipocytes-28081dbf-21a1-4ae7-8e14-1f1c4209525d)  
12 [gpr146-in-mouse-3t3-l1-fibroblasts-blunted-differentiation-into-adipocytes-28081dbf-21a1-](https://www.easd.org/virtualmeeting/home.html#!resources/crispr-cas9-induced-knockout-of-gpr146-in-mouse-3t3-l1-fibroblasts-blunted-differentiation-into-adipocytes-28081dbf-21a1-4ae7-8e14-1f1c4209525d)  
13 [4ae7-8e14-1f1c4209525d.](https://www.easd.org/virtualmeeting/home.html#!resources/crispr-cas9-induced-knockout-of-gpr146-in-mouse-3t3-l1-fibroblasts-blunted-differentiation-into-adipocytes-28081dbf-21a1-4ae7-8e14-1f1c4209525d)
- 14 11. Yu H, Rimbert A, Palmer AE, Toyohara T, Xia Y, Xia F, Ferreira LMR, Chen Z, Chen T,  
15 Loaiza N, Horwitz NB, Kacergis MC, Zhao L, BIOS Consortium, Soukas AA, Kuivenhoven  
16 JA, Kathiresan S, Cowan CA. GPR146 Deficiency Protects against Hypercholesterolemia and  
17 Atherosclerosis. *Cell.* 2019;179:1276-1288.e14.
- 18 12. Wacker D, Stevens RC, Roth BL. How Ligands Illuminate GPCR Molecular Pharmacology.  
19 *Cell* [Internet]. 2017 [cited 2021 May 28];170:414–427. Available from:  
20 <https://linkinghub.elsevier.com/retrieve/pii/S0092867417308164>
- 21 13. Crunkhorn S. GPR146 inhibition protects against atherosclerosis. *Nat Rev Drug Discov.*  
22 2020;19:22.
- 23 14. Fernández-Ruiz I. GPR146 is a potential new therapeutic target for lipid lowering. *Nat Rev*  
24 *Cardiol.* 2019;
- 25 15. Yosten GLC, Kolar GR, Redlinger LJ, Samson WK. Evidence for an interaction between  
26 proinsulin C-peptide and GPR146. *J Endocrinol.* 2013;218:B1-8.
- 27 16. Lindfors L, Sundström L, Fröderberg Roth L, Meuller J, Andersson S, Kihlberg J. Is GPR146  
28 really the receptor for proinsulin C-peptide? *Bioorg Med Chem Lett.* 2020;127208.
- 29 17. Ference BA, Ginsberg HN, Graham I, Ray KK, Packard CJ, Bruckert E, Hegele RA, Krauss  
30 RM, Raal FJ, Schunkert H, Watts GF, Borén J, Fazio S, Horton JD, Masana L, Nicholls SJ,  
31 Nordestgaard BG, van de Sluis B, Taskinen M-R, Tokgözoğlu L, Landmesser U, Laufs U,  
32 Wiklund O, Stock JK, Chapman MJ, Catapano AL. Low-density lipoproteins cause  
33 atherosclerotic cardiovascular disease. 1. Evidence from genetic, epidemiologic, and clinical  
34 studies. A consensus statement from the European Atherosclerosis Society Consensus Panel.  
35 *Eur Heart J.* 2017;38:2459–2472.
- 36 18. Võsa U, Claringbould A, Westra H-J, Bonder MJ, Deelen P, Zeng B, Kirsten H, Saha A,  
37 Kreuzhuber R, Kasela S, Pervjakova N, Alvaes I, Fave M-J, Agbessi M, Christiansen M, Jansen  
38 R, Seppälä I, Tong L, Teumer A, Schramm K, Hemani G, Verlouw J, Yaghootkar H, Sönmez  
39 R, Brown A, Kukushkina V, Kalnapekis A, Rieger S, Porcu E, Kronberg-Guzman J, Kettunen  
40 J, Powell J, Lee B, Zhang F, Arindrarto W, Beutner F, Brugge H, Dmitreva J, Elansary M,  
41 Fairfax BP, Georges M, Heijmans BT, Kähönen M, Kim Y, Knight JC, Kovacs P, Krohn K, Li

- 1 S, Loeffler M, Marigorta UM, Mei H, Momozawa Y, Müller-Nurasyid M, Nauck M, Nivard M,  
2 Penninx B, Pritchard J, Raitakari O, Rotzchke O, Slagboom EP, Stehouwer CDA, Stumvoll M,  
3 Sullivan P, Hoen PAC 't, Thiery J, Tönjes A, van Dongen J, van Iterson M, Veldink J, Völker  
4 U, Wijmenga C, Swertz M, Andiappan A, Montgomery GW, Ripatti S, Perola M, Kutalik Z,  
5 Dermitzakis E, Bergmann S, Frayling T, van Meurs J, Prokisch H, Ahsan H, Pierce B,  
6 Lehtimäki T, Boomsma D, Psaty BM, Gharib SA, Awadalla P, Milani L, Ouwehand W,  
7 Downes K, Stegle O, Battle A, Yang J, Visscher PM, Scholz M, Gibson G, et al. Unraveling the  
8 polygenic architecture of complex traits using blood eQTL metaanalysis. *bioRxiv* [Internet].  
9 2018 [cited 2019 Dec 3]; Available from: <http://biorxiv.org/lookup/doi/10.1101/447367>
- 10 19. GTEx Consortium, Laboratory, Data Analysis & Coordinating Center (LDACC)—Analysis  
11 Working Group, Statistical Methods groups—Analysis Working Group, Enhancing GTEx  
12 (eGTEx) groups, NIH Common Fund, NIH/NCI, NIH/NHGRI, NIH/NIMH, NIH/NIDA,  
13 Biospecimen Collection Source Site—NDRI, Biospecimen Collection Source Site—RPCI,  
14 Biospecimen Core Resource—VARI, Brain Bank Repository—University of Miami Brain  
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16 Data Integration & Visualization—EBI, Genome Browser Data Integration & Visualization—  
17 UCSC Genomics Institute, University of California Santa Cruz, Lead analysts:, Laboratory,  
18 Data Analysis & Coordinating Center (LDACC):, NIH program management:, Biospecimen  
19 collection:, Pathology:, eQTL manuscript working group:, Battle A, Brown CD, Engelhardt BE,  
20 Montgomery SB. Genetic effects on gene expression across human tissues. *Nature*.  
21 2017;550:204–213.
- 22 20. Sudlow C, Gallacher J, Allen N, Beral V, Burton P, Danesh J, Downey P, Elliott P, Green J,  
23 Landray M, Liu B, Matthews P, Ong G, Pell J, Silman A, Young A, Sprosen T, Peakman T,  
24 Collins R. UK Biobank: An Open Access Resource for Identifying the Causes of a Wide Range  
25 of Complex Diseases of Middle and Old Age. *PLOS Med* [Internet]. 2015 [cited 2021 Jun  
26 10];12:e1001779. Available from: <https://dx.plos.org/10.1371/journal.pmed.1001779>
- 27 21. Nikpay M, Goel A, Won H-H, Hall LM, Willenborg C, Kanoni S, Saleheen D, Kyriakou T,  
28 Nelson CP, Hopewell JC, Webb TR, Zeng L, Dehghan A, Alver M, Armasu SM, Auro K,  
29 Bjornes A, Chasman DI, Chen S, Ford I, Franceschini N, Gieger C, Grace C, Gustafsson S,  
30 Huang J, Hwang S-J, Kim YK, Kleber ME, Lau KW, Lu X, Lu Y, Lyytikäinen L-P, Mihailov  
31 E, Morrison AC, Pervjakova N, Qu L, Rose LM, Salfati E, Saxena R, Scholz M, Smith AV,  
32 Tikkanen E, Uitterlinden A, Yang X, Zhang W, Zhao W, de Andrade M, de Vries PS, van  
33 Zuydam NR, Anand SS, Bertram L, Beutner F, Dedoussis G, Frossard P, Gauguier D, Goodall  
34 AH, Gottesman O, Haber M, Han B-G, Huang J, Jalilzadeh S, Kessler T, König IR, Lannfelt L,  
35 Lieb W, Lind L, Lindgren CM, Lokki M-L, Magnusson PK, Mallick NH, Mehra N, Meitinger  
36 T, Memon F-U-R, Morris AP, Nieminen MS, Pedersen NL, Peters A, Rallidis LS, Rasheed A,  
37 Samuel M, Shah SH, Sinisalo J, Stirrups KE, Trompet S, Wang L, Zaman KS, Ardissino D,  
38 Boerwinkle E, Borecki IB, Bottinger EP, Buring JE, Chambers JC, Collins R, Cupples LA,  
39 Danesh J, Demuth I, Elosua R, Epstein SE, et al. A comprehensive 1,000 Genomes-based  
40 genome-wide association meta-analysis of coronary artery disease. *Nat Genet*. 2015;47:1121–  
41 1130.
- 42 22. van der Harst P, Verweij N. Identification of 64 Novel Genetic Loci Provides an Expanded  
43 View on the Genetic Architecture of Coronary Artery Disease. *Circ Res*. 2018;122:433–443.
- 44 23. Klijs B, Scholtens S, Mandemakers JJ, Snieder H, Stolk RP, Smidt N. Representativeness of the  
45 LifeLines Cohort Study. *PloS One*. 2015;10:e0137203.

- 1 24. Scholtens S, Smidt N, Swertz MA, Bakker SJL, Dotinga A, Vonk JM, van Dijk F, van Zon  
2 SKR, Wijmenga C, Wolffenbittel BHR, Stolk RP. Cohort Profile: LifeLines, a three-generation  
3 cohort study and biobank. *Int J Epidemiol*. 2015;44:1172–1180.
- 4 25. Aguib Y, Al Suwaidi J. The Copenhagen City Heart Study (Østerbrounder søgelsen). *Glob  
5 Cardiol Sci Pract*. 2015;2015:33.
- 6 26. Rimbart A, Dalila N, Wolters JC, Huijkman N, Smit M, Kloosterhuis N, Riemsma M, van der  
7 Veen Y, Singla A, van Dijk F, Biobank-Based Integrative Omics Studies Consortium, Frikke-  
8 Schmidt R, Burstein E, Tybjærg-Hansen A, van de Sluis B, Kuivenhoven JA. A common  
9 variant in CCDC93 protects against myocardial infarction and cardiovascular mortality by  
10 regulating endosomal trafficking of low-density lipoprotein receptor. *Eur Heart J*. 2019;
- 11 27. Marmontel O, Rollat-Farnier PA, Wozny A-S, Charrière S, Vanhoye X, Simonet T, Chatron N,  
12 Collin-Chavagnac D, Nony S, Dumont S, Mahl M, Jacobs C, Janin A, Caussy C, Poinso P,  
13 Tauveron I, Bardel C, Millat G, Peretti N, Moulin P, Marçais C, Di Filippo M. Development of  
14 a new expanded next-generation sequencing panel for genetic diseases involved in  
15 dyslipidemia. *Clin Genet*. 2020;98:589–594.
- 16 28. Klarin D, Damrauer SM, Cho K, Sun YV, Teslovich TM, Honerlaw J, Gagnon DR, DuVall SL,  
17 Li J, Peloso GM, Chaffin M, Small AM, Huang J, Tang H, Lynch JA, Ho Y-L, Liu DJ, Emdin  
18 CA, Li AH, Huffman JE, Lee JS, Natarajan P, Chowdhury R, Saleheen D, Vujkovic M, Baras  
19 A, Pyarajan S, Di Angelantonio E, Neale BM, Naheed A, Khera AV, Danesh J, Chang K-M,  
20 Abecasis G, Willer C, Dewey FE, Carey DJ, Global Lipids Genetics Consortium, Myocardial  
21 Infarction Genetics (MIGen) Consortium, Geisinger-Regeneron DiscovEHR Collaboration, VA  
22 Million Veteran Program, Concato J, Gaziano JM, O'Donnell CJ, Tsao PS, Kathiresan S, Rader  
23 DJ, Wilson PWF, Assimes TL. Genetics of blood lipids among ~300,000 multi-ethnic  
24 participants of the Million Veteran Program. *Nat Genet*. 2018;50:1514–1523.
- 25 29. Machiela MJ, Chanock SJ. LDlink: a web-based application for exploring population-specific  
26 haplotype structure and linking correlated alleles of possible functional variants. *Bioinforma  
27 Oxf Engl*. 2015;31:3555–3557.
- 28 30. Kuivenhoven JA, Hegele RA. Mining the genome for lipid genes. *Biochim Biophys Acta*.  
29 2014;1842:1993–2009.
- 30 31. Rimbart A. Phenotypic Differences Between Polygenic and Monogenic  
31 Hypobetalipoproteinemia. *Arterioscler Thromb Vasc Biol* [Internet]. 2020; Available from:  
32 <http://doi.org/10.1161/atvbaha.120.315491>
- 33 32. Nurmohamed NS, Collard D, Balder JW, Kuivenhoven JA, Stroes ESG, Reeskamp LF. From  
34 evidence to practice: development of web-based Dutch lipid reference values. *Neth Heart J  
35 Mon J Neth Soc Cardiol Neth Heart Found*. 2021;
- 36 33. Liu DJ, Peloso GM, Yu H, Butterworth AS, Wang X, Mahajan A, Saleheen D, Emdin C, Alam  
37 D, Alves AC, Amouyel P, Di Angelantonio E, Arveiler D, Assimes TL, Auer PL, Baber U,  
38 Ballantyne CM, Bang LE, Benn M, Bis JC, Boehnke M, Boerwinkle E, Bork-Jensen J,  
39 Bottinger EP, Brandslund I, Brown M, Busonero F, Caulfield MJ, Chambers JC, Chasman DI,  
40 Chen YE, Chen Y-DI, Chowdhury R, Christensen C, Chu AY, Connell JM, Cucca F, Cupples  
41 LA, Damrauer SM, Davies G, Deary IJ, Dedoussis G, Denny JC, Dominiczak A, Dubé M-P,  
42 Ebeling T, Eiriksdottir G, Esko T, Farmaki A-E, Feitosa MF, Ferrario M, Ferrieres J, Ford I,



- 1 Fornage M, Franks PW, Frayling TM, Frikke-Schmidt R, Fritsche LG, Frossard P, Fuster V,  
2 Ganesh SK, Gao W, Garcia ME, Gieger C, Giulianini F, Goodarzi MO, Grallert H, Grarup N,  
3 Groop L, Grove ML, Gudnason V, Hansen T, Harris TB, Hayward C, Hirschhorn JN, Holmen  
4 OL, Huffman J, Huo Y, Hveem K, Jabeen S, Jackson AU, Jakobsdottir J, Jarvelin M-R, Jensen  
5 GB, Jørgensen ME, Jukema JW, Justesen JM, Kamstrup PR, Kanoni S, Karpe F, Kee F, Khera  
6 AV, Klarin D, Koistinen HA, Kooner JS, Kooperberg C, Kuulasmaa K, Kuusisto J, et al.  
7 Exome-wide association study of plasma lipids in >300,000 individuals. *Nat Genet.*  
8 2017;49:1758–1766.
- 9 34. Berberich AJ, Hegele RA. The complex molecular genetics of familial hypercholesterolaemia.  
10 *Nat Rev Cardiol.* 2018;
- 11 35. Lotta LA, Sharp SJ, Burgess S, Perry JRB, Stewart ID, Willems SM, Luan J, Ardanaz E,  
12 Arriola L, Balkau B, Boeing H, Deloukas P, Forouhi NG, Franks PW, Grioni S, Kaaks R, Key  
13 TJ, Navarro C, Nilsson PM, Overvad K, Palli D, Panico S, Quirós J-R, Riboli E, Rolandsson O,  
14 Sacerdote C, Salamanca EC, Slimani N, Spijkerman AM, Tjonneland A, Tumino R, van der A  
15 DL, van der Schouw YT, McCarthy MI, Barroso I, O’Rahilly S, Savage DB, Sattar N,  
16 Langenberg C, Scott RA, Wareham NJ. Association Between Low-Density Lipoprotein  
17 Cholesterol-Lowering Genetic Variants and Risk of Type 2 Diabetes: A Meta-analysis. *JAMA.*  
18 2016;316:1383–1391.
- 19 36. Schmidt AF, Swerdlow DI, Holmes MV, Patel RS, Fairhurst-Hunter Z, Lyall DM, Hartwig FP,  
20 Horta BL, Hyppönen E, Power C, Moldovan M, van Iperen E, Hovingh GK, Demuth I, Norman  
21 K, Steinhagen-Thiessen E, Demuth J, Bertram L, Liu T, Coassin S, Willeit J, Kiechl S, Willeit  
22 K, Mason D, Wright J, Morris R, Wanamethee G, Whincup P, Ben-Shlomo Y, McLachlan S,  
23 Price JF, Kivimaki M, Welch C, Sanchez-Galvez A, Marques-Vidal P, Nicolaides A,  
24 Panayiotou AG, Onland-Moret NC, van der Schouw YT, Matullo G, Fiorito G, Guarrera S,  
25 Sacerdote C, Wareham NJ, Langenberg C, Scott R, Luan J, Bobak M, Malyutina S, Pajak A,  
26 Kubinova R, Tamosiunas A, Pikhart H, Husemoen LLN, Grarup N, Pedersen O, Hansen T,  
27 Linneberg A, Simonsen KS, Cooper J, Humphries SE, Brilliant M, Kitchner T, Hakonarson H,  
28 Carrell DS, McCarty CA, Kirchner HL, Larson EB, Crosslin DR, de Andrade M, Roden DM,  
29 Denny JC, Carty C, Hancock S, Attia J, Holliday E, O’Donnell M, Yusuf S, Chong M, Pare G,  
30 van der Harst P, Said MA, Eppinga RN, Verweij N, Snieder H, LifeLines Cohort study group,  
31 Christen T, Mook-Kanamori DO, Gustafsson S, Lind L, Ingelsson E, Pazoki R, Franco O,  
32 Hofman A, Uitterlinden A, Dehghan A, Teumer A, Baumeister S, et al. PCSK9 genetic variants  
33 and risk of type 2 diabetes: a mendelian randomisation study. *Lancet Diabetes Endocrinol.*  
34 2017;5:97–105.
- 35 37. Sattar N, Preiss D, Murray HM, Welsh P, Buckley BM, de Craen AJM, Seshasai SRK,  
36 McMurray JJ, Freeman DJ, Jukema JW, Macfarlane PW, Packard CJ, Stott DJ, Westendorp  
37 RG, Shepherd J, Davis BR, Pressel SL, Marchioli R, Marfisi RM, Maggioni AP, Tavazzi L,  
38 Tognoni G, Kjekshus J, Pedersen TR, Cook TJ, Gotto AM, Clearfield MB, Downs JR,  
39 Nakamura H, Ohashi Y, Mizuno K, Ray KK, Ford I. Statins and risk of incident diabetes: a  
40 collaborative meta-analysis of randomised statin trials. *Lancet Lond Engl.* 2010;375:735–742.
- 41 38. Nissen SE, Tuzcu EM, Schoenhagen P, Crowe T, Sasiela WJ, Tsai J, Orazem J, Magorien RD,  
42 O’Shaughnessy C, Ganz P, Reversal of Atherosclerosis with Aggressive Lipid Lowering  
43 (REVERSAL) Investigators. Statin therapy, LDL cholesterol, C-reactive protein, and coronary  
44 artery disease. *N Engl J Med.* 2005;352:29–38.
- 45 39. Ligthart S, Vaez A, Vösa U, Stathopoulou MG, de Vries PS, Prins BP, Van der Most PJ,

- 1 Tanaka T, Naderi E, Rose LM, Wu Y, Karlsson R, Barbalic M, Lin H, Pool R, Zhu G, Macé A,  
2 Sidore C, Trompet S, Mangino M, Sabater-Lleal M, Kemp JP, Abbasi A, Kacprowski T,  
3 Verweij N, Smith AV, Huang T, Marzi C, Feitosa MF, Lohman KK, Kleber ME, Milaneschi Y,  
4 Mueller C, Huq M, Vlachopoulou E, Lyytikäinen L-P, Oldmeadow C, Deelen J, Perola M,  
5 Zhao JH, Feenstra B, LifeLines Cohort Study, Amini M, CHARGE Inflammation Working  
6 Group, Lahti J, Schraut KE, Fornage M, Suktitipat B, Chen W-M, Li X, Nutile T, Malerba G,  
7 Luan J, Bak T, Schork N, Del Greco M F, Thiering E, Mahajan A, Marioni RE, Mihailov E,  
8 Eriksson J, Ozel AB, Zhang W, Nethander M, Cheng Y-C, Aslibekyan S, Ang W, Gandin I,  
9 Yengo L, Portas L, Kooperberg C, Hofer E, Rajan KB, Schurmann C, den Hollander W,  
10 Ahluwalia TS, Zhao J, Draisma HHM, Ford I, Timpson N, Teumer A, Huang H, Wahl S, Liu  
11 Y, Huang J, Uh H-W, Geller F, Joshi PK, Yanek LR, Trabetti E, Lehne B, Vozzi D, Verbanck  
12 M, Biino G, Saba Y, Meulenberg I, O'Connell JR, Laakso M, et al. Genome Analyses of  
13 >200,000 Individuals Identify 58 Loci for Chronic Inflammation and Highlight Pathways that  
14 Link Inflammation and Complex Disorders. *Am J Hum Genet.* 2018;103:691–706.
- 15 40. Libby P. The changing landscape of atherosclerosis. *Nature.* 2021;592:524–533.
- 16 41. Blom DJ, Raal FJ, Santos RD, Marais AD. Lomitapide and Mipomersen-Inhibiting Microsomal  
17 Triglyceride Transfer Protein (MTP) and apoB100 Synthesis. *Curr Atheroscler Rep.*  
18 2019;21:48.
- 19 42. Nordestgaard BG, Nicholls SJ, Langsted A, Ray KK, Tybjaerg-Hansen A. Advances in lipid-  
20 lowering therapy through gene-silencing technologies. *Nat Rev Cardiol.* 2018;15:261–272.
- 21 43. Musunuru K, Chadwick AC, Mizoguchi T, Garcia SP, DeNizio JE, Reiss CW, Wang K, Iyer S,  
22 Dutta C, Clendaniel V, Amaonye M, Beach A, Berth K, Biswas S, Braun MC, Chen H-M,  
23 Colace TV, Ganey JD, Gangopadhyay SA, Garrity R, Kasiewicz LN, Lavoie J, Madsen JA,  
24 Matsumoto Y, Mazzola AM, Nasrullah YS, Nneji J, Ren H, Sanjeev A, Shay M, Stahley MR,  
25 Fan SHY, Tam YK, Gaudelli NM, Ciaramella G, Stolz LE, Malyala P, Cheng CJ, Rajeev KG,  
26 Rohde E, Bellinger AM, Kathiresan S. In vivo CRISPR base editing of PCSK9 durably lowers  
27 cholesterol in primates. *Nature.* 2021;593:429–434.

28

## HIGHLIGHTS

- lower *GPR146* gene expression, is associated with beneficial effects on plasma: -lipids, -liver enzymes and hsCRP, with a concordant reduction in CAD risk.
- The *GPR146*-p.Pro62Leu coding rare variant induces larger reductions of plasma lipids, liver enzymes and CRP compared to the common variant.
- *GPR146* deficiency may be a new genetic cause of familial hypobetalipoproteinemia and a potential target for intervention to reduce plasma cholesterol levels and atherosclerosis with a particularly good metabolic safety profile.

## FIGURES LEGENDS

### FIGURE 1

**Panel A:** Effects of rs2362529 and rs1997243 on the expression of *GPR146* in whole blood (Source: <https://www.eqtlgen.org> ). The upper part of the figure shows the effects of rs2362529 (green arrows) while the bottom part shows the effects of rs1997243 (red arrows) on nearby genes. + and - signs show the direction of the effects with corresponding p values.

**Panel B:** Lipids, lipoproteins, and apolipoproteins in plasma as a function of the C-allele of rs2362529 (green symbols) and the G-allele of rs1997243 (red symbols) in individuals from UK Biobank cohort. The effects are shown as beta-coefficients (per z-score unit) per C-allele or G-allele compared with non-carriers respectively. Horizontal bars depict standard errors. N; number of individuals included in the analysis, LDL; low density lipoprotein, ApoB; apolipoprotein B, ApoAI; apolipoprotein AI, Lp(a); lipoprotein a.

**Panel C:** Plasma liver enzymes as a function of the C-allele of rs2362529 (green symbols) and the G-allele of rs1997243 (red symbols) in individuals from UK Biobank cohort. The effects are shown as beta-coefficients (per z-score unit) per C-allele or G-allele compared with non-carriers respectively. Horizontal bars depict standard errors. N; number of individuals included in the analysis, ALT; alanine transaminase, AST; aspartate transaminase, GGT; ammaglutamyl-transferases, ALP, alkaline phosphatase.

**Panel D:** Plasma C-reactive protein as a function of the C-allele of rs2362529 (green symbols) and the G-allele of rs1997243 (red symbols) in individuals from UK Biobank cohort. The effects are shown as beta-coefficients per z-score unit) per C-allele or G-allele compared with non-carriers respectively. Horizontal bars depict standard errors, N; number of individuals included in the analysis, CRP; C-reactive protein.

**Panel E:** Plasma glucose or HbA<sub>1C</sub> levels as a function of the C-allele of rs2362529 (green symbols) and the G-allele of rs1997243 (red symbols) in individuals from UK Biobank cohort. The effects are shown as beta-coefficients (per z-score unit) per C-allele or G-allele compared with non-carriers. Horizontal bars depict standard errors. N; number of individuals included in the analysis, HbA<sub>1C</sub>; glycated haemoglobin.

### FIGURE 2

**Panel A:** Odds ratio for coronary artery disease as a function of the C-allele of rs2362529 and the G-allele of rs1997243 compared with non-carriers. The effects are shown as odds-ratio per C-allele or

1 G-allele respectively compared with non-carriers. Horizontal bars depict standard errors. N; number  
2 of individuals included in the analysis, CAD; coronary artery disease.

3  
4 **Panel B:** Proportional risk of CAD associated with different genetic variants that affect LDL-c  
5 levels. All values are presented as magnitude of change (positive values). The regression line shows  
6 the proportional risk of CAD as a function of genetically determined change in LDL cholesterol (in  
7 mmol/L). The vertical and horizontal error bars are standard errors for the risk of CAD and LDL-c  
8 levels respectively.

### 9 10 **FIGURE 3**

11  
12 **Panel A:** Selection of individuals with extreme LDL-C plasma levels (<1<sup>st</sup> or 2<sup>nd</sup> percentile for age  
13 and sex from Lifelines cohort and CCHS, respectively and >99<sup>th</sup> or 98<sup>th</sup> percentile for age and sex  
14 from Lifelines cohort and CCHS respectively). Green crosses depict carriers of the *GPR146*-  
15 p.Pro62Leu.

16  
17 **Panel B:** Schematic view of the *GPR146* gene located on the 7p22.3 locus and the variant *GPR146*-  
18 p.Pro62Leu (chr7:1057700 (GRCh38.p13); NM\_138445.4:c.185C>T; NP\_612454.1:p.Pro62Leu).

19  
20 **Panel C:** Schematic view of the predicted amino-acids sequence and protein structure of GPR146.  
21 The red arrow indicates the Proline residue at position 62 of the protein located in the first  
22 intracellular loop. The total protein consists in 333 amino-acids.

23  
24 **Panel D:** Lipids, lipoproteins, and apolipoproteins in plasma as a function of the T-allele of  
25 rs151124717 (green symbols) and the C-allele of rs2362529 (gray symbols) in individuals from the  
26 UK Biobank cohort. The effects are shown as beta-coefficients (per z-score unit) per T-allele or C-  
27 allele compared with non-carriers respectively. Horizontal bars depict standard errors. N; number of  
28 individuals included in the analysis, LDL; low density lipoprotein, ApoB; apolipoprotein B, ApoAI;  
29 apolipoprotein AI, Lp(a); apolipoprotein a.

30  
31 **Panel E:** Plasma liver enzymes as a function of the minor T-allele of rs151124717 (green symbols)  
32 and the minor C-allele of rs2362529 (gray symbols) in individuals from the UK Biobank cohort. The  
33 effects are shown as beta-coefficients (per z-score unit) per T-allele or C-allele compared with non-  
34 carriers respectively. Horizontal bars depict standard errors. N; number of individuals included in the  
35 analysis, ALT; alanine transaminase, AST; aspartate transaminase, GGT; gammaglutamyl-  
36 transferases, ALP, alkaline phosphatase.

37  
38 **Panel F:** Plasma C-reactive protein levels as a function of the minor T-allele of rs151124717 (green  
39 symbols) and the minor C-allele of rs2362529 (gray symbols) in individuals from the UK Biobank  
40 cohort. The effects are shown as beta-coefficients (per z-score unit) per T-allele or C-allele compared  
41 with non-carriers. Horizontal bars depict standard errors. N; number of individuals included in the  
42 analysis, CRP; C-reactive protein.

### 43 44 **FIGURE 4**

1 Pedigree of a family with familial hypobetalipoproteinemia. Carriers of the GPR146-p.Pro62Leu  
2 variant are depicted with green symbols (II:6, III:3, and III:5/proband) and all have LDL-C values  
3 below the 5<sup>th</sup> percentile. Abbreviations: TC (total cholesterol), LDL-C (low-density lipoprotein  
4 cholesterol), HDL-C (high-density lipoprotein cholesterol) and triglycerides plasma levels are  
5 presented in mg/dL. Circles = women and squares = men; \*, Values between brackets show  
6 percentile for corresponding values adjusted for age and sex.

7

8