

# Pharmacokinetics and pharmacodynamics of hydroxychloroquine in hospitalized patients with COVID-19

Noel Zahr, Saïk Urien, Benoit Llopis, Valerie Pourcher, Olivier Paccoud, Alexandre Bleibtreu, Julien Mayaux, Estelle Gandjbakhch, Guillaume Hekemian, Alain Combes, et al.

# ▶ To cite this version:

Noel Zahr, Saïk Urien, Benoit Llopis, Valerie Pourcher, Olivier Paccoud, et al.. Pharmacokinetics and pharmacodynamics of hydroxychloroquine in hospitalized patients with COVID-19. Alternative and Complementary Therapies, In press, 10.1016/j.therap.2021.01.056. hal-03127389

# HAL Id: hal-03127389 https://hal.sorbonne-universite.fr/hal-03127389v1

Submitted on 1 Feb 2021

**HAL** is a multi-disciplinary open access archive for the deposit and dissemination of scientific research documents, whether they are published or not. The documents may come from teaching and research institutions in France or abroad, or from public or private research centers. L'archive ouverte pluridisciplinaire **HAL**, est destinée au dépôt et à la diffusion de documents scientifiques de niveau recherche, publiés ou non, émanant des établissements d'enseignement et de recherche français ou étrangers, des laboratoires publics ou privés.

# Pharmacokinetics and Pharmacodynamics of Hydroxychloroquine in Hospitalized Patients with COVID-19

Noël Zahr<sup>1\*¥</sup>, Saik Urien<sup>2¥</sup>, Benoit Llopis<sup>1</sup>, Valérie Pourcher<sup>3</sup>, Olivier Paccoud<sup>3</sup>, Alexandre Bleibtreu 3, Julien Mayaux 4, Estelle Gandjbakhch 5, Guillaume Hekemian 6, Alain Combes 6, Olivier Benveniste<sup>7</sup>, David Saadoun<sup>7</sup>, Yves Allenbach<sup>7</sup>, Bruno Pinna<sup>1</sup>, Patrice Cacoub<sup>7</sup>, Christian Funck-Brentano<sup>1</sup> and Joe-Elie Salem<sup>1</sup> <sup>1</sup> AP-HP. Sorbonne Université, Pitié-Salpêtrière Hospital, Department of Pharmacology and Clinical Investigation Center; INSERM, CIC-1901; Sorbonne Université, Faculty of Medicine; F-75013 Paris, France. <sup>2</sup> AP-HP. Université de Paris, INSERM, Cochin Hospital, Department of Pediatric and Perinatal Pharmacology. Paris, France. <sup>3</sup> AP-HP. Sorbonne Université, INSERM 1136, Institut Pierre Louis d'Epidémiologie et de Santé Publique, Pitié-Salpêtrière Hospital, Service de Maladies Infectieuses et Tropicales, Paris, France. <sup>4</sup> AP-HP. Sorbonne Université, Service de Pneumologie, Médecine intensive - Réanimation (Département "R3S"), Groupe Hospitalier Universitaire Pitié-Salpêtrière Charles Foix, 75013, Paris, France <sup>5</sup> AP-HP. Sorbonne Université, Service de Cardiologie, Groupe Hospitalier Universitaire Pitié-Salpêtrière Charles Foix, 75013, Paris, France. <sup>6</sup> AP-HP. Sorbonne Université, Médecine intensive – Réanimation Médicale Groupe Hospitalier Universitaire Pitié-Salpêtrière Charles Foix, 75013, Paris, France. AP-HP. Sorbonne Université, Pitié-Salpêtrière Hospital, Department of Internal Medicine and Clinical Immunology; Centre de Référence des Maladies Auto-Immunes et Systémiques Rares. Paris, France \* Correspondence: Dr Noël Zahr Department of pharmacology & Therapeutic Drug Monitoring Pitié-Salpêtrière Hospital Sorbonne Université, APHP, Paris, France noel.zahr@aphp.fr ; Tel.: +33 1 42 16 20 22 <sup>¥</sup> Contributed equally 

Abstract: Background: Hydroxychloroquine (HCQ) dosage required to reach circulating levels that inhibit SARS-Cov-2 are extrapolated from pharmacokinetic data in non-COVID-19 patients. Methods: We performed a population-pharmacokinetic analysis from 104 consecutive COVID-19 hospitalized patients (31 in intensive care units, 73 in medical wards, n=149 samples). Plasma HCQ concentration were measured using high-performance liquid chromatography with fluorometric detection. Modelling used Monolix-2019R2. Results: HCQ doses ranged from 200 to 800 mg/day administered for 1 to 11 days and median HCQ plasma concentration was 151 ng/mL. Among the tested covariates, only bodyweight influenced elimination oral clearance (CL) and apparent volume of distribution (Vd). CL/F (F for unknown bioavailability) and Vd/F (relative standard-error, %) estimates were 45.9L/h (21.2) and 6690L (16.1). The derived elimination half-life (t1/2) was 102h. These parameters in COVID-19 differed from those reported in patients with lupus, where CL/F, Vd/F and t1/2 are reported to be 68L/h, 2440L and 19.5h, respectively. Within 72h of HCQ initiation, only 16/104 (15.4%) COVID-19 patients had HCQ plasma levels above the in-vitro half maximal effective concentration of HCQ against SARS-CoV-2 (240ng/mL).. HCQ did not influence inflammation status (assessed by C-reactive protein) or SARS-CoV-2 viral clearance (assessed by real-time reverse transcription-PCR nasopharyngeal swabs). Conclusion: The inter-individual variability of HCQ pharmacokinetic parameters in severe COVID-19 patients was important and differed from that previously reported in non-COVID-19 patients. Loading doses of 1600mg HCQ followed by 600mg daily doses are needed to reach concentrations relevant to SARS-CoV-2 inhibition within 72 hours in ≥60% (95% confidence-interval: 49.5-69.0%) of COVID-19 patients Keywords: Hydroxychloroquine; Covid-19; Pharmacokinetics; Pharmacodynamics

# 82 Abbreviations

ALAT	Alanine aminotransferase
ASAT	Aspartate aminotransferase
β	Covariate effect parameter
BMI	Body Mass Index
BW	Bodyweight
CL/F	Apparent elimination Clearance
CNIL	National Commission on Informatics and Liberties
COVID-19	Novel coronavirus disease 2019
CRP	C-Reactive Protein
СҮР	Cytochrome P450
EC50	Half maximal effective concentration
F	Bioavailability
η	Between-subject variability
HCQ	Hydroxychloroquine
Ht	Hematocrit
Ka	Absorption rate constant
MDRD	Modification of Diet in Renal Disease equation
QTc	Corrected QT interval
RE	Rheumatoid Arthritis
RT-PCR	Reverse Transcription Polymerase Chain Reaction
SARS-CoV-2	Severe acute respiratory syndrome coronavirus
SLE	Systemic Lupus Erythematosus
T.i.d	Ter in die
U-HPLC	Ultra-High Performance Liquid Chromatography
V/F	Apparent Volume of Distribution
σ	Proportional residual variability

#### 93 1. Introduction

94 A new human respiratory-tropic coronavirus, SARS-CoV-2, has spread rapidly worldwide. 95 COVID-19, the disease caused by this virus, has a very variable clinical presentation, ranging from 96 pauci-symptomatic to acute respiratory distress syndrome. Several drugs are being evaluated for the 97 treatment of covid-19 including hydroxychloroquine (C18H26ClN3O) (HCQ). Some observational, 98 non-randomized studies have suggested the possible efficacy of HCQ associated or not with 99 azithromycin in COVID 19 patients contrasting with other studies [1-5]. Recent randomized 100 controlled trials showed that HCQ was not effective in hospitalized or non-hospitalized patients 101 with Covid-19 [6-9]. The results of over 120 randomized controlled trials for the treatment and 102 prevention of COVID-19 are pending. Doses of HCQ tested were highly variable, ranging from 103 400mg/day for few weeks up to 2.4g on day 1 as a loading dose followed by 400 mg/day for few 104 days, based on extrapolation from pharmacokinetics properties of HCQ derived from its approved 105 indications (malaria, auto-immune diseases) [10]. Yao et al. reported that HCQ possesses anti-viral 106 activity, against SARS-CoV2 in vitro [11] with an EC50 (half maximal effective concentration) of 107 0.72µM (240ng/mL) of HCQ on Vero-cells. The antiviral effect of HCQ, has been suggested to result 108 from increasing intracellular pH leading to decreased phago-lysosome fusion, and impaired viral 109 receptor glycosylation. Moreover, HCQ has immune-modulating effect by inhibiting toll-like 110 receptor signaling, decreasing production of cytokines, especially IL-1 and IL-6, potentially 111 mitigating the cytokine release syndrome induced by SARS-CoV-2 infection [12-14].

112 Steady-state pharmacokinetics of HCQ has previously been reported in healthy volunteers, 113 adult patients with malaria [15], systemic lupus erythematosus (SLE) [16, 17] and rheumatoid 114 arthritis (RA) [18-20] and are summarized in Table-1. Herein, we analyzed plasma and blood 115 concentration data in a cohort of consecutive patients hospitalized with COVID-19 who received 116 HCQ. The aim of this work was to characterize HCQ pharmacokinetics in the setting of COVID-19 117 and to identify its main influencing covariates. The pharmacokinetic model developed from 118 COVID-19 patients then allowed us to determine the best HCQ dosing regimen to rapidly reach 119 relevant theoretical antiviral concentrations, i.e. higher than HCQ EC50 on SARS-CoV-2. We finally 120 analyzed if there was any HCQ dose-efficacy relationship on SARS-CoV-2 clearance and 121 inflammation parameters.

## 122 2. Materials and Methods

123 We conducted a monocenter study in consecutive patients with confirmed COVID-19 (positive 124 for SARS-CoV-2 with reverse transcription polymerase chain reaction (RT-PCR), sampled for HCQ 125 therapeutic drug monitoring left at the discretion of the treating physicians. Patients were treated 126 with oral hydroxychloroquine sulfate (Plaquenil, Sanofi-Winthrop, Paris, France). Concentrations of 127 HCQ and its metabolites in whole blood and plasma were assayed by Ultra-High Performance 128 Liquid Chromatography (U-HPLC) with fluorometric detection [21]. This retrospective study was 129 based on data extracted from medical records, in strict compliance with the French reference 130 methodology MR-004, established by French National Commission on Informatics and Liberties 131 (CNIL) and was approved by Sorbonne University ethics Committee (CER-2020-14-JOCOVID).

#### 132 2.1. Pharmacokinetic-dynamic modelling

133 Hydroxychloroquine time-courses were analyzed using the nonlinear mixed effect modelling 134 software program Monolix 2019R2 (www.lixoft.eu). To ensure full convergence of the program, the 135 iteration number was fixed to 1000 with 50 Markov Chain Monte Carlo. The effect of the 136 demographic and clinical characteristics which were thought to influence pharmacokinetics were 137 evaluated for the following covariates: bodyweight (BW), height, age, sex, hepatic function using 138 ALAT (alanine aminotransferase), creatinine clearance using MDRD (modification of diet in renal 139 disease equation), CRP (c-reactive protein) level, serum albumin, co-prescription with 140 azithromycin or other macrolides, intensive care unit vs. medical wards patients, platelets/white cells counts and hematocrit. Parameter estimates were standardized for a mean standard covariateusing an allometric model:

143  $Pi = P_{STD} x (COVi/COVSTD)^{PWR}$ 

144 where PSTD is the standard value of parameter and Pi and COVi are the parameter and covariate 145 values of the i<sup>th</sup> individual. The superscript PWR denotes an exponent power.

For bodyweight, allometric scaling theory dictates that PWR are typically 1 and 0.75 for volumes and clearance terms, respectively [22]. The goodness-of-fit of each model was evaluated by the observed-predicted (population and individual) concentration scatter plots, by the visual inspection of the individual concentration-time courses, and the prediction-corrected visual predictive checks.

A one-compartment open model best described HCQ pharmacokinetics, whatever the sampling reference, blood or plasma. The parameters of the model were the elimination oral clearance (CL/F), the apparent volume of distribution (V/F) and the absorption rate constant, Ka (with F, as the unknown bioavailability). Given the lack of data on the absorption phase, Ka was fixed to 0.75 and 1.15 h<sup>-1</sup> in blood and plasma respectively as previously reported [22]. Between-subject variabilities were estimated for CL/F and V/F parameters and the residual variability was described by a proportional model. F stands for unknown bioavailability.

158 2.2. HCQ and Viral clearance

Different covariates, including HCQ concentration, thought to influence the time-to-PCR negativation were tested using the R-program [23] and the survival package [24]. The Kaplan-Meier method and log-Rank test were used for this purpose. Patients were split according to their individual model-predicted HCQ plasma concentration at 48h using the 1<sup>st</sup>, 50<sup>th</sup> or 75<sup>th</sup> quartile. Thereafter, two Kaplan-Meyer curves were generated for each splitting factor. The time to negativation was the first occurrence when two successive RT-PCR were negative.

## 165 2.3. HCQ effect on CRP

166 The CRP time-courses were modelled as a function of time and plasma HCQ Concentration 167 (Cp) as:

168  $CRP = CRP_0 \{1 - f_{HCQ} Cp / (Cp_{50} + Cp) - (1 - f_{HCQ}) t / (t + t_{50}) \}$ 

169 where CRP<sub>0</sub>, f<sub>HCQ</sub>, Cp<sub>50</sub> and t<sub>50</sub> denote the initial CRP concentration, fractional effect of HCQ, HCQ

- 170 concentration or time that produce a 50% decrease in the CRP<sub>0</sub> level. The model stands for the effect 171 of HCQ ( $f_{HCQ}$  and Cp<sub>50</sub>) plus an independent time-related effect ([1 -  $f_{HCQ}$ ] and  $t_{50}$ ) which
- 172 simultaneously decrease the initial CRP<sub>0</sub> level.
- 173 **3. Results**

## 174 3.1. Demographic and biological characteristics

175 A total of 149 plasma samples were obtained from 104 COVID-19 patients, (n=31 in intensive 176 care units and n=73 in medical wards). Time point of drug sampling was performed at various times 177 after HCQ dosing, i.e., mean 16.2 h (SD 30 h). Characteristics of included COVID-19 patients are 178 detailed in Table-2. At the time of HCQ blood sampling, 10/104 patients (9.6%) had severe renal 179 failure with a glomerular filtration rate <30 mL/min/1.73m2 and 34/104 (32.7%) had ALAT levels 3 180 times higher than the upper normal limit. In all patients, SARS-CoV-2 was confirmed by a positive 181 (RT-PCR) assay on a nasopharyngeal sample. All patients were treated with HCQ and 75/104 (72%) 182 had a post-treatment follow-up with RT-PCR on nasopharyngeal samples. HCQ was combined with 183 a macrolide antibiotic in 29 patients (n=6 with azithromycin). The mean time between the 184 introduction of HCQ and the onset of symptoms was  $8.6 \pm 5$  days. The usual HCQ dosage was

200mg t.i.d (78/104 patients) for 1 to 11 days (3 patients received an 800mg loading dose). Figure-1

shows plasma and blood HCQ concentrations available in our cohort.

Table 1. Median plasma and blood pharmacokinetics parameters of HCQ in several pathologies.

Plasma	COVID-19***	Lupus*	Malaria¤	Healthy+
V/F, L	6696	2440	2363	2851
CL/F, L/h	45.5	68.2	15.5	12.0
t1/2, h	102	19.5	106	172.3
Blood	COVID-19**	LUPUS*	RA¥	RA§
V/F, L	1990	903	605	2283
CL/F, L/h	14.7	18.6	9.9	15
t1/2, h	93.8	25.9	43.3	124.3

\*HCQ sulfate 400 mg/day [16], \$HCQ sulfate 400, 800, or 1,200 mg/day [19], \*HCQ sulfate 200 or

 400mg/day [20], #HCQ sulfate dose 800mg then 400 mg at 6, 24, and 48 h afterward [15], \*\*96 patients (135 samples), \*\*\*104 patients (149 samples) in our study were assessed.

	Mean	Minimum	Maximum
	± standard deviation		
Age, years	$63.0 \pm 14.4$	25	99
Weight, kg	$79 \pm 16$	40	150
Height, cm	$169 \pm 11$	146	192
BMI, kg/m <sup>2</sup>	$27.3 \pm 5.0$	17.8	51.9
Sex, (Female) %	32	NA	NA
Patient in intensive care, %	23	NA	NA
MDRD, mL/min/1.73m2.	$86.0 \pm 33.6$	5	194
Creatinine, µmol/L	98 ± 82	34	808
Albumin, g/L.	$29.0 \pm 6.9$	12	63
HT, %.	$35.0 \pm 6.5$	18	49
Platelet, 10º L	$313 \pm 134$	52	753
White blood cells, 109 L	$7.5 \pm 4.7$	2	32.4
C-reactive protein (CRP), mg/L	$86 \pm 98$	2	469
ALAT, U/L	$69 \pm 74$	11	486
ASAT, U/L	$63 \pm 49$	13	252
Dose HCQ, mg/day	563 ± 99	200	800
Observation duration, Days	$5.3 \pm 2.3$	1	12
Blood concentration			
HCQ, ng/mL	$586 \pm 457$	50	2792
Plasma concentration			
HCQ, ng/mL	$193 \pm 152$	12	795
HCQBlood/HCQPlasma	$4.0 \pm 2.3$	1	15

# Table 2. Demographic and biological characteristics of 104 patients.

BMI: Body Mass Index, NA: non-applicable, MDRD: (modification of diet in renal disease equation),
HT: hematocrit, ALAT: alanine aminotransferase, ASAT: aspartate aminotransferase, HCQ:
hydroxychloroquine. All data were collected at the time of HCQ sampling.



Figure 1. Observed blood (red) and plasma (green) hydroxychloroquine concentrations. Numbers
 stand for the patient identity and lines for the corresponding spline describing the overall trend for
 each matrix.

#### 217 3.2. Pharmacokinetic modelling

218 The population plasma and blood HCQ pharmacokinetic parameter estimates and their 219 influencing covariates are summarized in Table-3, and supplementary Table-1, respectively. These 220 parameters estimates were different from those reported in other diseases (lupus, malaria, 221 rheumatoid arthritis) or in healthy volunteers (Table-1). Figure-2A shows the visual predictive 222 checks for the HCQ plasma final model in COVID-19 (for blood final model, see Figure-2B). The 223 observed concentrations percentiles are well included in the corresponding model-predicted 90% 224 confidence interval bands. Among the tested covariates (age, bodyweight, gender, hepatic and renal 225 function, CRP, intensive care vs. medical wards care, macrolide/azithromycin co-prescription, 226 platelet count), bodyweight (based on allometry principles) was the sole variable having an effect on 227 plasma or blood HCQ CL/F and V/F prediction that improved the model. Platelet count had an 228 additional significant effect on V/F estimation for blood HCQ (supplementary Table-1).

Parameter	Estimate	% res
ka, h-1	1.15	fixed
V/F, L	6690	16.1
β, V/F*(BW/70) <sup>β</sup>	1	fixed
CL/F	45.9	21.2
β, CL/F*(BW/70) <sup>β</sup>	0.75	fixed
ηV/F	0.61	18.9
ηCL/F	0.69	25.1
σ, ng/mL	64.1	9.76

Table 3. Median plasma hydroxychloroquine population pharmacokinetic parameters in 104
 COVID-19 adult patients.

232CL/F, apparent elimination clearance; V/F, apparent volume of distribution; Ka, absorption rate233constant; F, unknown bioavailability; β, covariate effect parameter; η, between-subject variability; σ,234proportional residual variability; BW, bodyweight (CL/F and V/F estimates are normalized to a 70 kg235BW, i.e., for the i<sup>th</sup> patient CL/F<sub>i</sub> = CL/F\*(BW<sub>i</sub>/70)<sup>0.75.</sup>





Figure 2. Prediction-corrected visual predictive check for plasma (A) and blood (B) hydroxychloroquine
population pharmacokinetics. Plain (•) and green lines stand for prediction-corrected observed
concentrations and their 5<sup>th</sup>, 50<sup>th</sup> and 95<sup>th</sup> percentiles. Light blue and red bands stand for the corresponding
model predicted 90% confidence intervals.

251 Relying on our final pharmacokinetics parameters modelling, we generated representative 252 plasma HCQ concentrations-time courses using various dosing regimens of major COVID-19 253 prospective trials testing HCQ (Figure-3A). Concentration vs. time profiles were also drawn 254 according to documented plasma HCQ pharmacokinetics parameters estimates (Table-1) derived 255 from healthy volunteers, lupus and malaria patients (Figure-3B, 3C, 3D, respectively). Depending on 256 the diseases-specific estimates used, results were dramatically different. Figure-4 shows 4 dosing 257 regimens based on our COVID-19 plasma HCQ pharmacokinetics estimates leading to HCQ plasma 258 concentration above the HCQ EC50 against SARS-CoV-2 value 48 to 72h after treatment initiation. 259 Day 1 loading doses of HCQ ≥1600mg followed by daily dose ≥600mg reached theoretical 260 concentrations in  $\geq$ 40% (95% confidence interval 30-50%) and  $\geq$ 60% (95% confidence-interval: 261 49.5-69.0%) of COVID-19 patients within 48 and 72hours, respectively, assuming a distribution of 262 body weights generally similar to that of our population. For a selected dosing scheme, effect of 1st 263 and 3rd body weight quartiles on CL and Vd population parameters are shown in Figure-5A and 264 HCQ plasma concentration-times courses for patients weighing 79kg (median bodyweight) using 265 their individualized pharmacokinetic parameters are depicted in Figure-5B. An important between 266 patient's variability, leading to low or unexpectedly high (potentially toxic) HCQ plasma 267 concentrations, ensues despite administering a standardized HCQ dosing (Figure-5B).



269



Dosing schedules are 2.4 g loading dose then 400 mg/12h (RECOVERY), 1.2 g loading dose then 200 mg/8h (SANOFI), 200mg/8h (IHU.Marseille) and 800 mg loading dose then 400 mg/24h (DISCOVERY). Curves shown are using Covid-19 patients (A), lupus patients (B), malaria patients (C), and healthy subjects (D) parameters.

278



Figure 4. Possible dosing regimen in COVID-19 patients (weighing 79 kg) according to our final model.
Dosing schedules represented are 800mg/12h (total 1600 mg) the 1<sup>st</sup> day, then 400mg/12h (RED);
800mg/12h (total 1600 mg) the 1<sup>st</sup> day, then 200mg/8h (ORANGE); 400mg/8h (total 1200mg) loading dose
the 1<sup>st</sup> day, then 400mg/12h (BLUE); 600mg/8h (total 1800mg) loading dose the 1<sup>st</sup> day, then 200mg/8h (GREEN).



Figure 5. Mean plasma hydroxychloroquine concentration-time courses for a patient with bodyweight (WT) <58Kg or >103Kg and half-life >70h, red and blue curves, respectively (A) and for typical patients with 79kg WT and clearance (CL) ranging between 30-68 L/h and volume of distribution (Vd) between 4765-13470 L, black curves drawn from variable CL and Vd Bayesian estimates (B). Dosing regimen is 200 mg HCQ/8h, with no loading dose.

291

# 292 3.3. Pharmacodynamic effects of HCQ in COVID-19

293 A total of 75 patients were available for a SARS-CoV-2 viral status analysis using 294 nasopharyngeal swab. PCR follow-up was negative in 40 (53%). To assess the effect of plasma HCQ 295 concentration on time-to-PCR negativation, patients were grouped as follows: individual predicted 296 plasma HCQ concentration at 48h below versus above 25th (72 ng/mL), 50th (95.5 ng/mL), 75th 297 quantile (129 ng/mL). There were no significant differences in time-to-PCR negativation for all tested 298 comparisons (Figure-6). In our cohort, only 4 and 16 patients among 104 had observed or imputed 299 (in patients with data available after 72 hours) HCQ plasma levels >240ng/mL, the in-vitro half 300 maximal effective concentration of HCQ against SARS-CoV-2, at 48 and 72 hours, respectively. 301 There was also no significant effect of HCQ plasma concentration on the CRP time-course. All 302 attempts gave non-significant values for fHCQ, or Cp50 parameters that stand for the effect of HCQ 303 on CRP time-course, meaning that the effect of HCQ on the inflammation status could not be 304 demonstrated.







Figure 6. Time-to-Sars-Cov-2 PCR negativation curves as a function of HCQ plasma levels within 48
hours of HCQ start. Blue and red curves represent patients with an HCQ plasma concentration at 48
h below or above the 1<sup>st</sup> HCQ plasma concentration quartile observed in our cohort, respectively (72
ng/mL, A), median (95 ng/mL, B) and 3<sup>rd</sup> quartile (129 ng/mL, C).

#### 313 4. Discussion

314 In this study, we developed a plasma and blood population pharmacokinetics models of HCQ 315 based on data obtained in hospitalized COVID-19 patients in intensive care units and in medical 316 wards. The blood and plasma pharmacokinetics were described by a one-compartment model with 317 first-order absorption. Body weight had a significant effect on CL and Vd in both matrices. HCQ 318 pharmacokinetic parameters in COVID-19 patients are different from those of other pathologies 319 (lupus, malaria, rheumatoid arthritis) and healthy volunteers [15, 16, 20]. The theoretical ideal lowest 320 dose to achieve a target plasma concentration >EC50 (240ng/ml) within 48/72 hours in most patients 321 was 1600mg as a loading dose, followed by 200mg/8h thereafter. Nevertheless, plasma 322 concentrations of HCQ showed a high interindividual variability (see Figure-1) mainly influenced 323 by body weight. In COVID 19, either HCQ dosage adjusted on body weight or HCQ plasma 324 therapeutic drug monitoring may be useful options if HCQ is clinically effective on COVID-19. 325 However, in our cohort study, there was no significant influence of HCQ plasma concentrations on 326 inflammation (CRP) or on viral clearance (RT-PCR).

327 Interestingly, recent studies used HCQ pharmacokinetics parameters derived from 328 autoimmune diseases, to propose dosing regimen of HCQ to be used in COVID-19 patients [25-27]. 329 Thus, our data suggest that relevance of these type of modelling might be toned down given the 330 importance of difference observed between HCQ pharmacokinetic parameters in COVID-19 versus 331 other settings (Table-1). Supporting our findings, preliminary pharmacokinetics data from a small 332 cohort of 7 hospitalized COVID-19 patients treated with HCQ as part of the RECOVERY trial (2.4g as 333 loading dose then 400 mg/12h) have recently been pre-published [28]. The results indicate that HCQ 334 concentrations are lower than those expected based on previous modelling, even though a high dose 335 regimen was used.

Of note, our PK blood parameters estimates were concordant with those estimated by Thémans et al [29] and other groups [27, 29, 30] providing evidence that a high HCQ loading dose is needed to reach circulating levels in COVID-19 patients theoretically relevant as compared to in-vitro SARS-CoV-2 inhibitory concentrations.

In our cohort including over 100 COVID-19 patients, subjects had different profiles ranging from hospitalization in medicine to intensive care unit, with variable renal and hepatic functions, as well as co-prescription with macrolides, most of which are cytochrome P-450 inhibitors [31]. None influenced HCQ plasma and blood pharmacokinetics in COVID-19 except weight, or weight and platelet count, respectively. This finding is concordant with other HCQ pharmacokinetic studies in lupus and malaria settings, in which body mass index and platelet count were also significant contributing covariates in the model [16, 32].

347 Of note, the relationship between circulating concentrations of HCQ and clinical efficacy has 348 been demonstrated in rheumatoid arthritis and systemic lupus erythematosus [17-19, 33]. However, 349 our study did not show any association between plasma HCQ concentration and time to 350 negativation of SARS-CoV-2 viral load in hospitalized patients, or resolution of inflammation 351 (assessed by CRP). We are currently studying the association between the blood and plasma 352 concentration of HCQ and QTc (i.e the duration of ventricular repolarization corrected for heart rate, 353 a predictor of ventricular arrhythmias) [26] in patients with COVID-19 to further assess 354 cardiovascular safety of HCQ in COVID-19 setting [34]. Indeed, the risks of cardiotoxicity associated 355 with HCQ during the COVID-19 pandemic might increase for several reasons. Patients with 356 COVID-19 have multiple risk factors for drug-induced QT prolongation and proarrhythmia: 357 hypokalemia; fever amplifying drug-induced IKr blockade; and an increase in interleukin-6, as seen 358 in COVID-19 infection which has been suggested as a mechanism of the QT prolongation associated 359 with inflammation [35]. The French Pharmacovigilance Network has reported 103 notifications of cardiac adverse drug reactions associated with "off-label" use of hydroxychloroquine since March
2020 up to April 2020 [36]. These observations, on top of its lack of efficacy, justified limiting the
prescription of HCQ in COVID-19 patients [37].

The retrospective, observational design of our work is the main limitation. The blood and nasopharyngeal samples were not systematically assessed for all patients during the treatment period. This may have biased our results by precluding to demonstrate that there was an association between plasma HCQ levels and negative viral loads. Unfortunately, the detailed time course of viral load was unknot available, precluding further analysis. However, multiple lines of evidence are emerging against HCQ efficacy in hospitalized COVID-19, even with theoretically effective high dosing regimen such as in the RECOVERY randomized controlled trial [38-40]. In that study, patients received a loading dose of 2.4g then 400mg every 12 hours. HCQ was not associated with reduced mortality but was associated with an increased length of hospital stay and a trend towards increased risk of progression to invasive mechanical ventilation or death. [35, 41] Indeed, the dosing regimen used in the RECOVERY trial was even higher than the adapted dosing regimen that we can recommend based on in vitro HCQ EC50 on SARS-CoV-2 and HCQ human pharmacokinetic parameters in COVID-19, identified in this work.

## 376 5. Conclusions

Interindividual variability of HCQ pharmacokinetics parameters in hospitalized COVID-19
 patients was important and parameters differed from those identified in non-COVID-19 patients. No
 effect of HCQ was found on SARS-CoV-2 (nasopharyngeal) viral clearance nor on inflammation
 resolution. Loading doses of 1600mg HCQ followed by 600mg daily doses reached within 72 hours,
 concentrations relevant to SARS-CoV-2 inhibition in ≥60% (95% confidence-interval: 49.5-69.0%) of
 COVID-19 patients.

#### **Funding:** This research received no external funding

- 386 Acknowledgments: NA
- **Conflicts of Interest:** The authors declare no conflict of interest.

410 **Table S1**. Median blood Hydroxychloroquine population pharmacokinetic parameters in 98 COVID-19 adult

- 411 patients
- 412

Parameter	Estimate	%res
ka, h-1	0.75	fixed
V/F, L	1,990	15.9
β, V/F*(BW/70) <sup>β</sup>	1	fixed
β, V/F*(PLAT/300,000) <sup>β</sup>	-0.726	37
CL/F	14.7	13.5
β, CL/F*(BW/70) <sup>β</sup>	0.75	fixed
ηV/F		
ηCL/F		
σ, proportional	0.272	12.2

414 CL/F, apparent elimination clearance; V/F, apparent volume of distribution; Ka, absorption rate constant; F,

415 unknown bioavailability;  $\beta$ , covariate effect parameter;  $\eta$ , between-subject variability;  $\sigma$ , proportional residual

416 variability; BW, bodyweight (CL/F and V/F estimates are normalized to a 70 kg BW plus V/F to a 300,000 417 platelets count, i.e., for the i<sup>th</sup> patient CL/F<sub>i</sub> = CL/F\*(BW<sub>i</sub>/70)<sup>0.75</sup> and V/F<sub>i</sub> = V/F\*(BW<sub>i</sub>/70)\*(PLAT<sub>i</sub>/300,000)-<sup>0.726</sup>

418

419

420

#### 422 References

- 423 1. Arshad S, Kilgore P, Chaudhry ZS, Jacobsen G, Wang DD, Huitsing K et al. Treatment with 424 hydroxychloroquine, azithromycin, and combination in patients hospitalized with COVID-19. Int J Infect Dis.
- 425 2020;97:396-403. doi:10.1016/j.ijid.2020.06.099.
- 426 2. Gautret P, Lagier JC, Parola P, Hoang VT, Meddeb L, Mailhe M et al. Hydroxychloroquine and azithromycin
- 427 as a treatment of COVID-19: results of an open-label non-randomized clinical trial. Int J Antimicrob Agents.
- 428 2020;56(1):105949. doi:10.1016/j.ijantimicag.2020.105949.
- 429 3. Gautret P, Lagier JC, Parola P, Hoang VT, Meddeb L, Sevestre J et al. Clinical and microbiological effect of a
- 430 combination of hydroxychloroquine and azithromycin in 80 COVID-19 patients with at least a six-day follow
- 431 up: A pilot observational study. Travel Med Infect Dis. 2020;34:101663. doi:10.1016/j.tmaid.2020.101663.
- 432 4. Mikami T, Miyashita H, Yamada T, Harrington M, Steinberg D, Dunn A et al. Risk Factors for Mortality in
- 433 Patients with COVID-19 in New York City. J Gen Intern Med. 2020. doi:10.1007/s11606-020-05983-z.
- 434 5. Million M, Lagier JC, Gautret P, Colson P, Fournier PE, Amrane S et al. Early treatment of COVID-19 patients

435 with hydroxychloroquine and azithromycin: A retrospective analysis of 1061 cases in Marseille, France. Travel

- 436 Med Infect Dis. 2020;35:101738. doi:10.1016/j.tmaid.2020.101738.
- 437 6. Geleris J, Sun Y, Platt J, Zucker J, Baldwin M, Hripcsak G et al. Observational Study of Hydroxychloroquine
- 438 in Hospitalized Patients with Covid-19. N Engl J Med. 2020;382(25):2411-8. doi:10.1056/NEJMoa2012410.
- 439 7. Rosenberg ES, Dufort EM, Udo T, Wilberschied LA, Kumar J, Tesoriero J et al. Association of Treatment With
- 440 Hydroxychloroquine or Azithromycin With In-Hospital Mortality in Patients With COVID-19 in New York
- 441 State. JAMA. 2020. doi:10.1001/jama.2020.8630.
- 442 8. Boulware DR, Pullen MF, Bangdiwala AS, Pastick KA, Lofgren SM, Okafor EC et al. A Randomized Trial of
- 443 Hydroxychloroquine as Postexposure Prophylaxis for Covid-19. N Engl J Med. 2020;383(6):517-25.
  444 doi:10.1056/NEIMoa2016638.
- 9. Tang W, Cao Z, Han M, Wang Z, Chen J, Sun W et al. Hydroxychloroquine in patients with mainly mild to
  moderate coronavirus disease 2019: open label, randomised controlled trial. BMJ. 2020;369:m1849.
  doi:10.1136/bmj.m1849.
- 448 10. Le MP, Peiffer-Smadja N, Guedj J, Neant N, Mentre F, Ader F et al. Rationale of a loading dose initiation for
- 449 hydroxychloroquine treatment in COVID-19 infection in the DisCoVeRy trial. J Antimicrob Chemother. 2020.450 doi:10.1093/jac/dkaa191.
- 451 11. Yao X, Ye F, Zhang M, Cui C, Huang B, Niu P et al. In Vitro Antiviral Activity and Projection of Optimized
- 452 Dosing Design of Hydroxychloroquine for the Treatment of Severe Acute Respiratory Syndrome Coronavirus 2
- 453 (SARS-CoV-2). Clin Infect Dis. 2020;71(15):732-9. doi:10.1093/cid/ciaa237.
- 454 12. Akpovwa H. Chloroquine could be used for the treatment of filoviral infections and other viral infections
- that emerge or emerged from viruses requiring an acidic pH for infectivity. Cell Biochem Funct.
  2016;34(4):191-6. doi:10.1002/cbf.3182.
- 457 13. Savarino A, Boelaert JR, Cassone A, Majori G, Cauda R. Effects of chloroquine on viral infections: an old
  458 drug against today's diseases? Lancet Infect Dis. 2003;3(11):722-7. doi:10.1016/s1473-3099(03)00806-5.
- 459 14. Rossi B, Nguyen LS, Zimmermann P, Boucenna F, Dubret L, Baucher L et al. Effect of Tocilizumab in
- 460 Hospitalized Patients with Severe COVID-19 Pneumonia: A Case-Control Cohort Study. Pharmaceuticals
- 461 (Basel). 2020;13(10). doi:10.3390/ph13100317.
- 462 15. Lim HS, Im JS, Cho JY, Bae KS, Klein TA, Yeom JS et al. Pharmacokinetics of hydroxychloroquine and its
- 463 clinical implications in chemoprophylaxis against malaria caused by Plasmodium vivax. Antimicrob Agents
- 464 Chemother. 2009;53(4):1468-75. doi:10.1128/AAC.00339-08.

- 465 16. Morita S, Takahashi T, Yoshida Y, Yokota N. Population Pharmacokinetics of Hydroxychloroquine in
- Japanese Patients With Cutaneous or Systemic Lupus Erythematosus. Ther Drug Monit. 2016;38(2):259-67.
   doi:10.1097/FTD.00000000000261.
- 468 17. Costedoat-Chalumeau N, Amoura Z, Hulot JS, Hammoud HA, Aymard G, Cacoub P et al. Low blood
- 469 concentration of hydroxychloroquine is a marker for and predictor of disease exacerbations in patients with
   470 systemic lupus erythematosus. Arthritis Rheum. 2006;54(10):3284-90. doi:10.1002/art.22156.
- 471 18. Tett SE, Cutler DJ, Beck C, Day RO. Concentration-effect relationship of hydroxychloroquine in patients
- 472 with rheumatoid arthritis--a prospective, dose ranging study. J Rheumatol. 2000;27(7):1656-60.
- 473 19. Munster T, Gibbs JP, Shen D, Baethge BA, Botstein GR, Caldwell J et al. Hydroxychloroquine
  474 concentration-response relationships in patients with rheumatoid arthritis. Arthritis Rheum. 2002;46(6):1460-9.
  475 doi:10.1002/art.10307.
- 476 20. Carmichael SJ, Charles B, Tett SE. Population pharmacokinetics of hydroxychloroquine in patients with
- 477 rheumatoid arthritis. Ther Drug Monit. 2003;25(6):671-81. doi:10.1097/00007691-200312000-00005.
- 478 21. Noe G, Amoura Z, Combarel D, Lori L, Tissot N, Seycha A et al. Development and Validation of a Fast
- 479 Ultra-High Performance Liquid Chromatography-Fluorescent Method for the Quantification of
- 480 Hydroxychloroquine and Its Metabolites in Patients With Lupus. Ther Drug Monit. 2019;41(4):476-82.
  481 doi:10.1097/FTD.00000000000614.
- 482 22. Anderson BJ, Holford NH. Mechanism-based concepts of size and maturity in pharmacokinetics. Annu Rev
  483 Pharmacol Toxicol. 2008;48:303-32. doi:10.1146/annurev.pharmtox.48.113006.094708.
- 484 23. R. Core Team. R: A language and environment for statistical computing. R Foundation for Statistical485 Computing. Vienna, Austria. 2018.
- 486 24. Therneau T. A Package for Survival Analysis in S. version 2.38,
   487 <u>https://CRAN.R-project.org/package=survival</u>. 2015.
- 488 25. Martin-Blondel G, Ruiz S, Murris M, Faguer S, Duhalde V, Eyvrard F et al. Hydroxychloroquine in
  489 COVID-19 patients: what still needs to be known about the kinetics. Clin Infect Dis. 2020.
  490 doi:10.1093/cid/ciaa558.
- 491 26. Funck-Brentano C, Salem JE, Nguyen LS, Drici MD, Roden DM. Response to the editorial "COVID-19 in
- 492 patients with cardiovascular diseases": Covid-19 treatment with hydroxychloroquine or chloroquine and
- 493 azithromycin: A potential risk of Torsades de Pointes. Arch Cardiovasc Dis. 2020;113(5):367-8.
  494 doi:10.1016/j.acvd.2020.04.001.
- 495 27. Garcia-Cremades M, Solans BP, Hughes E, Ernest JP, Wallender E, Aweeka F et al. Optimizing
  496 Hydroxychloroquine Dosing for Patients With COVID-19: An Integrative Modeling Approach for Effective
  407 Description of the Descri
- 497 Drug Repurposing. Clin Pharmacol Ther. 2020;108(2):253-63. doi:10.1002/cpt.1856.
- 498 28. MacGowan AP, Hamilton F, Bayliss M, Read L, Attwood M, Noel A et al. Hydroxychloroquine serum
- 499 concentrations in non-critical care patients infected with SARS CoV 2. medRxiv. 2020;2020.06.23.20137992.
  500 doi:10.1101/2020.06.23.20137992.
- 501 29. Themans P, Belkhir L, Dauby N, Yombi JC, De Greef J, Delongie KA et al. Population Pharmacokinetics of
- 502 Hydroxychloroquine in COVID-19 Patients: Implications for Dose Optimization. Eur J Drug Metab
- 503 Pharmacokinet. 2020. doi:10.1007/s13318-020-00648-y.
- 504 30. Karatza E, Ismailos G, Marangos M, Karalis V. Optimization of hydroxychloroquine dosing scheme based
- 505 on COVID-19 patients' characteristics: a review of the literature and simulations. Xenobiotica. 2020:1-12.
- 506 doi:10.1080/00498254.2020.1824301.

- 507 31. von Rosensteil NA, Adam D. Macrolide antibacterials. Drug interactions of clinical significance. Drug Saf.
  508 1995;13(2):105-22. doi:10.2165/00002018-199513020-00005.
- 509 32. Jallouli M, Galicier L, Zahr N, Aumaitre O, Frances C, Le Guern V et al. Determinants of
- 510 hydroxychloroquine blood concentration variations in systemic lupus erythematosus. Arthritis Rheumatol.
- 511 2015;67(8):2176-84. doi:10.1002/art.39194.
- 512 33. Chasset F, Arnaud L, Costedoat-Chalumeau N, Zahr N, Bessis D, Frances C. The effect of increasing the dose
- 513 of hydroxychloroquine (HCQ) in patients with refractory cutaneous lupus erythematosus (CLE): An open-label
- 514 prospective pilot study. J Am Acad Dermatol. 2016;74(4):693-9 e3. doi:10.1016/j.jaad.2015.09.064.
- 515 34. Nguyen LS, Dolladille C, Drici MD, Fenioux C, Alexandre J, Mira JP et al. Cardiovascular Toxicities
- 516 Associated With Hydroxychloroquine and Azithromycin: An Analysis of the World Health Organization
- 517 Pharmacovigilance Database. Circulation. 2020;142(3):303-5. doi:10.1161/CIRCULATIONAHA.120.048238.
- 518 35. Funck-Brentano C, Nguyen LS, Salem JE. Retraction and republication: cardiac toxicity of 519 hydroxychloroquine in COVID-19. Lancet. 2020;396(10245):e2-e3. doi:10.1016/S0140-6736(20)31528-2.
- 520 36. Gerard A, Romani S, Fresse A, Viard D, Parassol N, Granvuillemin A et al. "Off-label" use of
- 521 hydroxychloroquine, azithromycin, lopinavir-ritonavir and chloroquine in COVID-19: A survey of cardiac
- 4 adverse drug reactions by the French Network of Pharmacovigilance Centers. Therapie. 2020;75(4):371-9.
- 523 doi:10.1016/j.therap.2020.05.002.
- 37. Roustit M, Guilhaumou R, Molimard M, Drici MD, Laporte S, Montastruc JL et al. Chloroquine and
  hydroxychloroquine in the management of COVID-19: Much kerfuffle but little evidence. Therapie.
  2020;75(4):363-70. doi:10.1016/j.therap.2020.05.010.
- 527 38. Cavalcanti AB, Zampieri FG, Rosa RG, Azevedo LCP, Veiga VC, Avezum A et al. Hydroxychloroquine with
- 528 or without Azithromycin in Mild-to-Moderate Covid-19. N Engl J Med. 2020. doi:10.1056/NEJMoa2019014.
- 529 39. Skipper CP, Pastick KA, Engen NW, Bangdiwala AS, Abassi M, Lofgren SM et al. Hydroxychloroquine in
- 530 Nonhospitalized Adults With Early COVID-19: A Randomized Trial. Ann Intern Med. 2020. 531 doi:10.7326/M20-4207.
- 532 40. Mitja O, Corbacho-Monne M, Ubals M, Tebe C, Penafiel J, Tobias A et al. Hydroxychloroquine for Early
- 533 Treatment of Adults with Mild Covid-19: A Randomized-Controlled Trial. Clin Infect Dis. 2020. 534 doi:10.1093/cid/ciaa1009.
- 535 41. Horby P, Mafham M, Linsell L, Bell JL, Staplin N, Emberson JR et al. Effect of Hydroxychloroquine in
- 536 Hospitalized Patients with COVID-19: Preliminary results from a multi-centre, randomized, controlled trial.
- 537 medRxiv. 2020:2020.07.15.20151852. doi:10.1101/2020.07.15.20151852.
- 538