



HAL
open science

Double-in vitro maturation increases the number of vitrified oocytes available for fertility preservation when ovarian stimulation is unfeasible

Nathalie Sermondade, Michaël Grynberg, Marjorie Comtet, Constance Valdelievre, Christophe Sifer, Charlotte Sonigo

► To cite this version:

Nathalie Sermondade, Michaël Grynberg, Marjorie Comtet, Constance Valdelievre, Christophe Sifer, et al.. Double-in vitro maturation increases the number of vitrified oocytes available for fertility preservation when ovarian stimulation is unfeasible. *Scientific Reports*, 2020, 10 (1), pp.18555. 10.1038/s41598-020-75699-x . hal-03160870

HAL Id: hal-03160870

<https://hal.sorbonne-universite.fr/hal-03160870v1>

Submitted on 5 Mar 2021


HAL is a multi-disciplinary open access archive for the deposit and dissemination of scientific research documents, whether they are published or not. The documents may come from teaching and research institutions in France or abroad, or from public or private research centers.

L'archive ouverte pluridisciplinaire **HAL**, est destinée au dépôt et à la diffusion de documents scientifiques de niveau recherche, publiés ou non, émanant des établissements d'enseignement et de recherche français ou étrangers, des laboratoires publics ou privés.



OPEN

Double-in vitro maturation increases the number of vitrified oocytes available for fertility preservation when ovarian stimulation is unfeasible

Nathalie Sermondade^{1,7}, Michaël Grynberg^{2,3,4,5}, Marjorie Comtet², Constance Valdelievre², Christophe Sifer¹ & Charlotte Sonigo^{2,3,6}

When ovarian stimulation is unfeasible, in vitro maturation (IVM) represents an alternative option for fertility preservation (FP). This retrospective study aims to evaluate the feasibility of performing within a short time frame two IVM cycles for FP. Seventeen women with breast cancer, 18–40 years of age, having undergone 2 cycles of IVM followed by oocyte vitrification were included. Non parametric analyses were used. No difference was observed between IVM1 and IVM2 outcomes. No complication was reported. The respective contributions of IVM1 and IVM2 for the number of cryopreserved oocytes were comparable irrespective of the delay between both procedures, even when performed during the same menstrual cycle. Those findings suggest that repeating IVM cycles may constitute a safe option for increasing the number of vitrified mature oocytes for FP. These two retrievals may be performed during the same cycle, providing additional argument for a physiologic continuous recruitment during follicular development.

Cancer treatments, such as chemotherapy, may induce early ovarian follicular depletion and impaired reproductive potential of survivors. Fertility preservation (FP) techniques have been developed to improve their possibilities of becoming genetic parents after healing^{1,2}. Among them, the most established is cryopreservation of mature oocytes after controlled ovarian stimulation (COS)^{1,3}. Although the actual success rate of oocyte vitrification in oncologic situations is unknown, it is commonly admitted that the more oocytes cryopreserved, the better the chances of live birth after thawing. As a consequence, two or more oocyte retrievals are often proposed in order to increase the total number of vitrified oocytes before the initiation of gonadotoxic treatments⁴. However, waiting for a new menstrual cycle before starting a second round of ovarian stimulation is often impossible for oncologic purposes. In this context, random start stimulation has been tried, based on the observation of small antral follicles, possibly responsive to exogenous FSH, anytime during the menstrual cycle^{5,6}. Further, the concept of the Duostim emerged, consisting in the repetition of 2 ovarian stimulations within the same menstrual cycle. Initially proposed in infertile poor responder patients^{7,8}, the Duostim has been extended to FP⁹. Even if the superiority of this strategy versus two stimulations during two consecutive cycles remains to be demonstrated in infertile patients, it constitutes a method to increase the number of cryopreserved oocytes during the shortest time frame.

In some oncologic situations, COS cannot be considered, as a result of contraindication to exogenous gonadotropins administration or when cancer treatment is urgent. Therefore, alternative options have been

¹Department of Cytogenetic and Reproductive Biology, Hôpital Jean Verdier, Hôpitaux Universitaires Paris-Seine-Saint-Denis, Assistance Publique—Hôpitaux de Paris, 93143 Bondy, France. ²Department of Reproductive Medicine and Fertility Preservation, Hôpital Jean Verdier, Hôpitaux Universitaires Paris-Seine-Saint-Denis, Assistance Publique—Hôpitaux de Paris, 93143 Bondy, France. ³Department of Reproductive Medicine and Fertility Preservation, Hôpital Antoine Béclère, Hôpitaux Universitaires Paris-Sud, Assistance Publique—Hôpitaux de Paris, 92140 Clamart, France. ⁴Université Paris-Sud, Université Paris Saclay, 94276 Le Kremlin Bicêtre, France. ⁵Inserm U1133, Université Paris Diderot, 75013 Paris, France. ⁶Inserm U1185 Université Paris-Sud, Université Paris Saclay, 94276 Le Kremlin Bicêtre, France. ⁷Present address: Department of Reproductive Biology, Hôpital Tenon, Hôpitaux Universitaires Est Parisien, Assistance Publique—Hôpitaux de Paris, 75020 Paris, France.  email: nathalie.sermondade@aphp.fr

Age (years)	32.6 ± 5.0 (23.4–40.0)
BMI (kg/m ²)	22.2 ± 3.0 (19.6–29.4)
Antral follicle count	22.8 ± 8.7 (10–38)
Serum anti-Müllerian hormone levels (ng/mL)	3.0 ± 1.7 (0.8–6.3)

Table 1. Clinical characteristics of patients that underwent dual IVM for fertility preservation. Data are expressed as mean ± SD (min–max).

IVM outcomes	IVM1	IVM2	P
Number of cumulus oocyte complexes recovered	7.7 ± 4.5	7.8 ± 5.5	0.9
Oocyte retrieval rate (%)	36.8 ± 18.3	32.4 ± 16.6	0.6
Total number of matured oocytes (MII)	4.8 ± 3.2	4.4 ± 4.2	0.7
Total oocyte maturation rate (%)	62.7 ± 23.9	51.6 ± 32.4	0.4
Percentage of mature vitrified oocyte per procedure/total number of mature vitrified oocyte	57%	43%	/

Table 2. Biological outcomes of IVM procedures. Data are expressed as mean ± SD. Oocyte retrieval rate = number of retrieved oocytes / AFC × 100. Total oocyte maturation rate = number of mature oocytes / number of retrieved oocytes × 100. Wilcoxon matched-paired signed rank tests were used.

proposed, such as oocyte vitrification after unstimulated in vitro maturation (IVM) and/or ovarian tissue cryopreservation^{10,11}. Although data about IVM for FP remain scarce, it is likely that the number of cryopreserved oocytes may also be a crucial issue and approaches resulting in an increase in this number could represent a significant achievement.

The present investigation aims to evaluate the feasibility, safety and effectiveness of performing, within a short time frame, two successive unstimulated IVM cycles before chemotherapy, in young women diagnosed with breast cancer.

Results

Patients' characteristics. Overall, mean age of the population was 32.6 ± 5.0 years (23–40 years) (Table 1). Mean serum anti-Müllerian hormone (AMH) levels and antral follicle counts (AFC) performed on the day of oncofertility counselling irrespective of the menstrual phase were 3.0 ± 1.7 ng/mL and 22.8 ± 8.7 follicles, respectively.

IVM results. IVM results are reported in Table 2. A similar number of follicles were punctured during both cycles, allowing the retrieval of a similar mean numbers of COCs following IVM1 and IVM2 (7.7 ± 4.5 and 7.8 ± 5.5, respectively ($p = 0.90$)). After 24 and 48 h of IVM, the mean total numbers of metaphase II oocytes available for vitrification were comparable (4.8 ± 3.2 and 4.4 ± 4.2, $p = 0.75$). As expected, the total number of vitrified oocytes per patient was significantly higher following two IVM cycles (9.2 ± 6.1 vs. 4.8 ± 3.2, $p < 0.001$). Among the total number of vitrified oocytes, a mean of 57.0% (0–100) and 43.0% (0–100) were obtained following IVM1 and IVM2, respectively.

Timing between two successive oocyte retrievals. Median time between IVM1 and IVM2 was 28.0 days (6.0–56.0). There was no association between the time between both procedures and the proportion of vitrified oocytes resulting from IVM2 ($r = 0.39$, $p = 0.11$) (Fig. 1). IVM1 and IVM2 were performed during the same or different menstrual cycles for 6 and 11 patients, respectively. There was no difference between the respective contributions of IVM1 and IVM2 in either situation. The second IVM provided 45.9 ± 29.9% vs. 37.7 ± 24.1% of the total number of cryopreserved oocytes, when performed during the same or different menstrual cycles, respectively, $p = 0.67$.

Discussion

Although it was commonly thought that antral follicles recruitment occurs in a cyclic manner following the luteo-follicular transition, increasing data challenges this theory. In 2003, Baerwald et al. ultrasonographically documented follicular waves throughout the ovarian cycle¹², paving the way for other theories. This finding was followed by clinical case reports of successful luteal phase oocyte retrieval for IVM or after COS^{13,14}. Based on these data, novel ovarian stimulation regimens have been developed, such as random-start protocols^{5,6} and double stimulation in the same menstrual cycle (Duostim)^{7,8}. Used in order to maximize the number of oocytes for poor prognosis patients and/or to facilitate urgent FP procedures, the feasibility of these protocols relies on two further theories of follicle recruitment: (i) the waves theory, according to which many cohorts of antral follicles are recruited during the menstrual cycle or (ii) the continuous recruitment theory, hypothesizing that follicles are recruited continuously throughout the menstrual cycle¹⁵. The present investigation confirms that

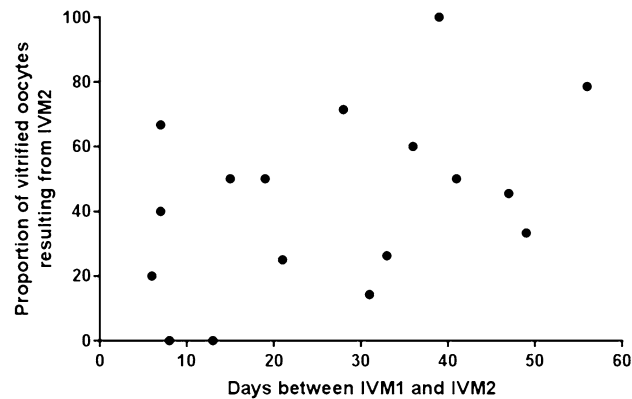


Figure 1. Association between the time between IVM1 and IVM2, and the proportion of vitrified oocytes resulting from IVM2.

recruitable antral follicles are present within the ovaries throughout the menstrual cycle, endowed with oocytes capable of IVM. The delay between both immature retrievals was not associated with the success of the second procedure, even when less than 10 days separated the two procedures, providing additional arguments for a continuous follicular recruitment.

Data from in vitro fertilization (IVF) indicates that the number of oocytes is a key point for success. Cumulative live birth rates following IVF or egg donation continue to increase with the number of oocytes harvested^{16–18}, suggesting that the higher the oocyte yield, the better the likelihood of achieving a live birth. However, the generalization of those results obtained with fresh oocytes to frozen eggs may be questioned. Although, data on the outcomes of oocyte vitrification cycles is scarce, evidence indicate that the number of cryopreserved oocytes represents a crucial issue for the success of FP strategies¹⁹. In this context, it is likely that approaches resulting in an increase in this number could represent a significant achievement. In particular, some authors applied the double stimulation protocol for FP patients, aiming at maximizing the number of vitrified oocytes without delaying cancer treatments⁹. However, few data are currently available regarding safety of COS in neoadjuvant situations, still leading to reluctance for some oncologists to agree with this strategy, especially if a double stimulation is planned within a limited time frame. Other options can then be proposed, including unstimulated IVM or ovarian tissue cryopreservation or a combination of both techniques²⁰. Although preliminary, our findings suggest that double IVM might constitute a feasible and safe option for increasing the total number of cryopreserved oocytes, especially in cases where patients are reluctant to undergo a surgical procedure for ovarian tissue cryopreservation.

The present results should be confirmed in a larger population referred for FP. In addition, even if all procedures were only performed by experienced practitioners in our center, the unpredictable nature of immature oocyte retrieval might have influenced the recovery rate between IVM1 and IVM2 and further results of each procedure. Few data are currently available regarding the outcome of cryopreserved oocytes. In particular, the competence of oocytes retrieved during the luteal phase was initially questioned. Recent data indicates that luteal phase ovarian stimulation results in similar euploid blastocyst formation^{8,21} and similar miRNomic signatures within follicular fluids²². Although it still merits our attention, reassuring obstetrical and neonatal data have also been preliminary outlined^{23,24}. However, the competence of IVM oocytes remains poorly known, even if live births were recently reported²⁵.

In conclusion, unstimulated IVM represents an alternative for preserving fertility of breast cancer patients when ovarian stimulation is unfeasible. Our findings suggest that repeating IVM cycles may constitute a feasible and safe option for increasing the total number of cryopreserved mature oocytes, a key point to success. In addition, the second IVM can be performed during the same menstrual cycle, even within a short time frame (less than 10 days), providing additional argument for a continuous follicular recruitment throughout the menstrual cycle. Further analyses should be performed to evaluate the relevance and efficiency of this strategy in terms of live births.

Methods

Patients. From July 2013 and July 2017, we studied 17 women, 23 to 40 years of age, having undergone 2 cycles of IVM followed by oocyte vitrification before a first chemotherapy for breast cancer. All of them were characterized by early referral for oncofertility counseling. A first IVM cycle (IVM1) was scheduled rapidly, irrespective of the menstrual cycle phase²⁶, while waiting for therapeutic decision. At this time, oncologists did not give agreement for ovarian stimulation. Finally, when COS could not be considered due to urgent chemotherapy and/or oncologic reasons, patients were offered a second IVM cycle (IVM2) combined with ovarian tissue cryopreservation. The present retrospective study was approved by our Local Ethic Committee (CLEA-2017-46), all methods were performed in accordance with the relevant guidelines and regulations and informed consent was obtained from all subjects.

Ovarian reserve assessment. Measurement of serum AMH and transvaginal ovarian ultrasound scan for AFC were systematically performed, irrespective of the menstrual phase, on the day of oncofertility counseling (before IVM1) and just before IVM2. Serum AMH levels were determined using a fully automated AMH assay (Elecys AMH; Roche Diagnostics International). The limit of detection, limit of quantification, and maximum imprecision were 0.01 ng/mL, 0.03 ng/mL, and 3.5%, respectively. Ultrasound scans were performed using a 5–9 MHz multi-frequency transvaginal probe (RIC 5–9-D; Voluson E8 Expert, General Electric Medical Systems, Paris, France) by one single operator, who was blinded to the results of hormone assays. The objective of ultrasound examination was to carefully and exhaustively determine the number and sizes of small antral follicles in both ovaries.

Oocytes in vitro maturation and vitrification procedure. Oocyte retrieval was performed as previously described for all IVM procedures²⁷, irrespective of their rank. Briefly, cumulus-oocyte complexes (COCs) were recovered transvaginally under ultrasound guidance. All visible follicles were fully cleared out. After 24 h of incubation in IVM culture medium (Origio, Denmark) enriched with 20% inactivated patient's serum and FSH and LH (Menopur, Ferring, Germany)²⁸, COCs were denuded. Mature metaphase II oocytes were frozen using Kitazato Vitrification Kit (Kitazato BioPharma Co., Ltd., Japan) according to manufacturers' instructions, and loaded in a minimal volume on the Cryotop device. Vitrification was induced by immediate plunging into liquid nitrogen²⁹. Oocytes having failed to mature after 24 h were kept for 24 additional hours of culture. After 48 h, oocytes that have reached metaphase II stage were frozen.

Statistical analysis. Continuous data are presented as mean \pm standard deviation (SD) and categorical data are presented as percentages. Wilcoxon matched-paired signed rank tests were used to compare IVM1 and IVM2 outcomes, as well as the total number of mature vitrified oocytes after one or two IVM cycles. The respective proportion of vitrified oocytes resulting from IVM1 or IVM2 was calculated as the number of metaphase II oocytes following IVM1 or IVM2, divided by the total number of vitrified oocytes after both procedures. The correlation between the delay separating both oocyte retrievals and the gain of a second IVM procedure was assessed by Spearman test. Mann Whitney test was used to compare the gain of IVM2 when performed during the same or the subsequent menstrual cycle. A p-value < 0.05 was considered significant.

Received: 8 June 2020; Accepted: 19 October 2020

Published online: 29 October 2020

References

1. Practice Committee of American Society for Reproductive Medicine. Fertility preservation in patients undergoing gonadotoxic therapy or gonadectomy: A committee opinion. *Fertil. Steril.* **100**, 1214–1223 (2013).
2. ISFP Practice Committee *et al.* Recommendations for fertility preservation in patients with lymphoma, leukemia, and breast cancer. *J. Assist. Reprod. Genet.* **29**, 465–468 (2012).
3. Loren, A. W. *et al.* Fertility preservation for patients with cancer: American Society of Clinical Oncology clinical practice guideline update. *J. Clin. Oncol.* **31**, 2500–2510 (2013).
4. Turan, V., Bedoschi, G., Moy, F. & Oktay, K. Safety and feasibility of performing two consecutive ovarian stimulation cycles with the use of letrozole-gonadotropin protocol for fertility preservation in breast cancer patients. *Fertil. Steril.* **100**, 1681–1685 (2013).
5. von Wolff, M. *et al.* Ovarian stimulation to cryopreserve fertilized oocytes in cancer patients can be started in the luteal phase. *Fertil. Steril.* **92**, 1360–1365 (2009).
6. Cakmak, H., Katz, A., Cedars, M. I. & Rosen, M. P. Effective method for emergency fertility preservation: Random-start controlled ovarian stimulation. *Fertil. Steril.* **100**, 1673–1680 (2013).
7. Kuang, Y. *et al.* Double stimulations during the follicular and luteal phases of poor responders in IVF/ICSI programmes (Shanghai protocol). *Reprod. Biomed. Online* **29**, 684–691 (2014).
8. Ubaldi, F. M. *et al.* Follicular versus luteal phase ovarian stimulation during the same menstrual cycle (DuoStim) in a reduced ovarian reserve population results in a similar euploid blastocyst formation rate: New insight in ovarian reserve exploitation. *Fertil. Steril.* **105**, 1488–1495 (2016).
9. Tsampras, N., Gould, D. & Fitzgerald, C. T. Double ovarian stimulation (DuoStim) protocol for fertility preservation in female oncology patients. *Hum. Fertil. (Camb.)* **20**, 248–253 (2017).
10. Huang, J. Y. J., Tulandi, T., Holzer, H., Tan, S. L. & Chian, R. C. Combining ovarian tissue cryobanking with retrieval of immature oocytes followed by in vitro maturation and vitrification: An additional strategy of fertility preservation. *Fertil. Steril.* **89**, 567–572 (2008).
11. Berwanger, A. L. *et al.* New trends in female fertility preservation: In vitro maturation of oocytes. *Fut. Oncol.* **8**, 1567–1573 (2012).
12. Baerwald, A. R., Adams, G. P. & Pierson, R. A. Characterization of ovarian follicular wave dynamics in women. *Biol. Reprod.* **69**, 1023–1031 (2003).
13. Demirtas, E. *et al.* Immature oocyte retrieval in the luteal phase to preserve fertility in cancer patients. *Reprod. Biomed. Online* **17**, 520–523 (2008).
14. Bedoschi, G. M., de Albuquerque, F. O., Ferriani, R. A. & Navarro, P. A. Ovarian stimulation during the luteal phase for fertility preservation of cancer patients: Case reports and review of the literature. *J. Assist. Reprod. Genet.* **27**, 491–494 (2010).
15. Baerwald, A. R., Adams, G. P. & Pierson, R. A. Ovarian antral folliculogenesis during the human menstrual cycle: A review. *Hum. Reprod. Update* **18**, 73–91 (2012).
16. Briggs, R., Kovacs, G., MacLachlan, V., Motteram, C. & Baker, H. W. G. Can you ever collect too many oocytes?. *Hum. Reprod.* **30**, 81–87 (2015).
17. Law, Y. J., Zhang, N., Venetis, C. A., Chambers, G. M. & Harris, K. The number of oocytes associated with maximum cumulative live birth rates per aspiration depends on female age: A population study of 221 221 treatment cycles. *Hum. Reprod.* **34**, 1778–1787 (2019).
18. Cobo, A., Garrido, N., Pellicer, A. & Remohi, J. Six years' experience in ovum donation using vitrified oocytes: Report of cumulative outcomes, impact of storage time, and development of a predictive model for oocyte survival rate. *Fertil. Steril.* **104**, 1426–1434 (2015).
19. Cobo, A., García-Velasco, J., Domingo, J., Pellicer, A. & Remohi, J. Elective and onco-fertility preservation: Factors related to IVF outcomes. *Hum. Reprod.* **33**, 2222–2231 (2018).

20. Sermondade, N. *et al.* Serum antimüllerian hormone is associated with the number of oocytes matured in vitro and with primordial follicle density in candidates for fertility preservation. *Fertil. Steril.* **111**, 357–362 (2019).
21. Cimadomo, D. *et al.* Luteal phase anovulatory follicles result in the production of competent oocytes: Intra-patient paired case-control study comparing follicular versus luteal phase stimulations in the same ovarian cycle. *Hum. Reprod.* **33**, 1442–1448 (2018).
22. Cimadomo, D. *et al.* Similar miRNomic signatures characterize the follicular fluids collected after follicular and luteal phase stimulations in the same ovarian cycle. *J. Assist. Reprod. Genet.* **37**, 149–158 (2020).
23. Vaiarelli, A. *et al.* Double stimulation in the same ovarian cycle (DuoStim) to maximize the number of oocytes retrieved from poor prognosis patients: A multicenter experience and SWOT analysis. *Front. Endocrinol.* **9**, 317 (2018).
24. Chen, H. *et al.* Comparison of live-birth defects after luteal-phase ovarian stimulation vs. conventional ovarian stimulation for in vitro fertilization and vitrified embryo transfer cycles. *Fertil. Steril.* **103**, 1194–1201 (2015).
25. Grynberg, M., Le Mayeur, B. A., Hesters, L., Gallot, V. & Frydman, N. First birth achieved after fertility preservation using vitrification of in vitro matured oocytes in a woman with breast cancer. *Ann. Oncol.* **31**, 541–542 (2020).
26. Grynberg, M. *et al.* Similar in vitro maturation rates of oocytes retrieved during the follicular or luteal phase offer flexible options for urgent fertility preservation in breast cancer patients. *Hum. Reprod.* **31**, 623–629 (2016).
27. Sonigo, C. *et al.* What threshold values of antral follicle count and serum AMH levels should be considered for oocyte cryopreservation after in vitro maturation?. *Hum. Reprod.* **31**, 1493–1500 (2016).
28. Chian, R. C. & Tan, S. L. Maturational and developmental competence of cumulus-free immature human oocytes derived from stimulated and intracytoplasmic sperm injection cycles. *Reprod. Biomed. Online* **5**, 125–132 (2002).
29. Cobo, A. *et al.* Oocyte vitrification as an efficient option for elective fertility preservation. *Fertil. Steril.* **105**, 755–764 (2016).

Author contributions

N.S.: collection of data; analysis and interpretation of data; writing of the manuscript. M.G.: study design; analysis and interpretation of data; critical revisions of the manuscript. M.C.: collection of data; analysis and interpretation of data. C.V.: collection of data; analysis and interpretation of data. C.Si.: collection of data; critical revision of the manuscript. F.E.: collection of data; critical revision of the manuscript. C.So.: study design; collection of data; analysis and interpretation of data; statistical analysis; critical revisions of the manuscript.

Competing interests

The authors declare no competing interests.

Additional information

Correspondence and requests for materials should be addressed to N.S.

Reprints and permissions information is available at www.nature.com/reprints.

Publisher's note Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.



Open Access This article is licensed under a Creative Commons Attribution 4.0 International License, which permits use, sharing, adaptation, distribution and reproduction in any medium or format, as long as you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons licence, and indicate if changes were made. The images or other third party material in this article are included in the article's Creative Commons licence, unless indicated otherwise in a credit line to the material. If material is not included in the article's Creative Commons licence and your intended use is not permitted by statutory regulation or exceeds the permitted use, you will need to obtain permission directly from the copyright holder. To view a copy of this licence, visit <http://creativecommons.org/licenses/by/4.0/>.

© The Author(s) 2020