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Maximum effect of the heterogeneity of tissue mineralization on the effective cortical bone elastic properties

Luc Brémaud^{1,2} · Xiran Cai^{1,3}
Renald Brenner² · Quentin Grimal¹

Abstract The mineralization level is heterogeneous in cortical bone extracellular matrix as a consequence of remodeling. Models of the effective elastic properties at the millimeter scale have been developed based on idealizations of the vascular pore network and matrix properties. Some popular models do not take into account the heterogeneity of the matrix. However, the errors on the predicted elasticity when the difference in elastic properties between osteonal and interstitial tissues is not modeled have not been quantified. This work provides an estimation of the maximum error. We compare the effective elasticity of a representative volume element (RVE) assuming (1) different elastic properties in osteonal and interstitial tissues vs. (2) average matrix properties. In order to account for the variability of bone microstructure, we use a collection of high resolution images of the pore network to build RVEs. In each RVE we assumed a constant osteonal wall thickness and we artificially varied this thickness between 35 and 140 μm to create RVEs with different amounts of osteonal tissue. The homogenization problem was solved with a fast Fourier transform (FFT) based numerical scheme. We found that the error depends on pore volume fraction and varies on average from 1% to 7% depending on the assumed diameter of the osteons. The results suggest that matrix heterogeneity may be disregarded in cortical bone models in most practical cases.

Keywords bone mineralization · porosity · elasticity · homogenization · mesoscale

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1 Introduction

About 10% of the adult skeleton is renewed every year by the remodeling process which, in cortical bone, produces secondary osteons. Bone cells deposit a collagen matrix which is progressively mineralized resulting in an increase of the average degree of mineralization of bone (DMB) (Boivin and Meunier 2002). As a consequence of this continuous remodeling process, cortical bone mineralized extracellular matrix (or shortly, matrix) is an assembly of portions of tissue of various ages characterized by different mineralization levels. The oldest part of this tissue is found in the interstitial tissue, which is composed of remnants of old osteons which have been partially remodeled. The difference of DMB between intact osteons and interstitial tissue is directly observed in microradiographs or indirectly as differences of elasticity with nanoindentation (Zysset et al. 1999), or acoustic impedance (a proxy for tissue elasticity) with scanning acoustic microscopy (Raum 2008) as illustrated in Fig. 1. Typically, osteons with the lowest mineral density contain 70% to 75% of the mineral content of the most highly calcified tissue parts (Boivin and Meunier 2002). For instance, Lefevre et al. found that DMB measured from quantitative microradiography in elderly adults was $1.260 (\pm 0.029) \text{ g cm}^{-3}$ in interstitial tissue vs. $1.079 (\pm 0.037) \text{ g cm}^{-3}$ in osteonal tissue (Lefèvre et al. 2019).

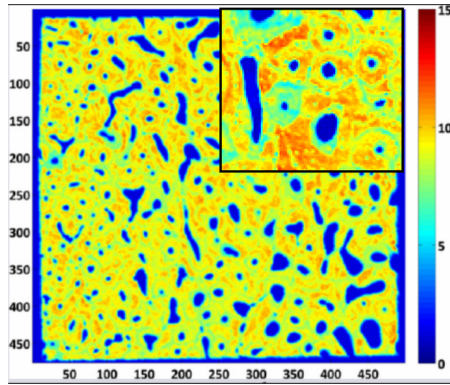


Fig. 1: Scanning acoustic microscopy (SAM) maps of acoustic impedance (MRayl) of human femoral cortical bone in a plane transverse to osteons. The relative differences in impedance reflect to a large extent the variations of mineralization levels in different portions of the tissue, a larger impedance (or elasticity) corresponding to more mineralized, and older, tissue (Raum et al. 2006). The main image is a region of $5 \times 5 \text{ mm}^2$ scanned at 50 MHz with a resolution of about $30 \mu\text{m}$. The inset is a SAM image at 200 MHz with a resolution of about $8 \mu\text{m}$ (Granke et al. 2011). The vascular porosity appears in dark blue.

The effective elastic properties of cortical bone at the mesoscale (Grimal et al. 2011a), or millimeter scale, are determined by the elastic properties of the mineralized tissue (matrix) and the properties of the pore network. Micromechanical models have been used for several decades to calculate the effective elastic properties for given representative volume elements (RVE) of bone. These models have been extensively

used to investigate the relationships between material composition, microarchitecture and elastic properties. The heterogeneity of the elastic properties of the matrix, i.e., the differences between osteonal and interstitial tissues, has been considered in some studies (Crolet et al. 1993; Dong and Guo 2006; Grimal et al. 2008; Hamed et al. 2010). However, many other authors have considered an homogeneous matrix (Hellmich and Ulm 2004; Sevostianov and Kachanov 2000; Parnell and Grimal 2009; Baumann et al. 2012; Gagliardi et al. 2017; Cai et al. 2019a). The errors on predicted mesoscale properties when the differences in elasticity between osteonal and interstitial tissues are disregarded have not been quantified. Noteworthy, these errors may strongly depend on the underlying microstructure (porosity and pore organization).

The objective of the present paper is to quantify the range of errors on predicted mesoscale effective elastic properties due to the assumption of an homogeneous mineralized matrix. We compare the effective elasticity of a RVE assuming (1) different elastic properties in osteonal and interstitial tissue and (2) average elastic properties. In order to account for the variability of bone microstructure, we use a collection of high resolution three-dimensionnal images of the pore network to build RVEs of cortical bone. For each RVE, we assume a constant osteonal wall thickness and we artificially vary this thickness between 35 and 140 μm to create RVEs with different amounts of osteonal tissue. With this parametric study, we estimate a maximum effect of the heterogeneity

This work provides data to quantitatively discuss the relevancy of important assumptions of cortical bone models. The results are helpful to guide the interpretation of effective elastic properties at the millimeter scale. These effective properties are involved in finite element models for the prediction of bone strength (van Rietbergen and Ito 2015; Engelke et al. 2016) or bone remodeling (Martínez-Reina and Pivonka 2019), and for the in vivo assessment of porosity by inverse approaches (Minonzi et al. 2019)

2 Model

2.1 Specimens and bone microstructure modelling

We have used a collection of synchrotron radiation micro-computed tomography (SR- μCT) 3D images of volumes of interest (VOI) from a previous study (Cai et al. 2019a) (Fig. 2). Briefly, cortical bone specimens were harvested from the mid-diaphysis of the left femur of 25 human cadavers. Among the donors, 13 were females and 12 were males (50 – 95 years old, 77 ± 11.5 , mean \pm SD). The nominal size of the specimens was $3 \times 4 \times 5 \text{ mm}^3$ in radial (x_1), circumferential (x_2) and axial (x_3) directions of the bone, respectively.

The femurs were provided by the Département Universitaire d'Anatomie Rockefeller (Lyon, France) through the French program on voluntary corpse donation to science. The tissue donors or their legal guardians provided informed written consent to give their tissue for investigations, in accord with legal clauses stated in the French Code of Public Health.

Images of the specimens were obtained with isotropic voxel size of 6.5 μm performed at the European Synchrotron Radiation Facility (ESRF, Grenoble, France), Figure 3. The 3D volume of each specimen was slightly rotated with the image processing software Fiji (Schindelin et al. 2012) using bilinear interpolation so that

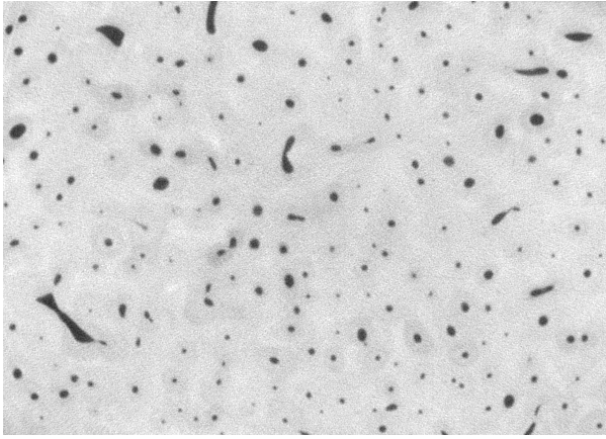


Fig. 2: A representative reconstructed cross-sectional slice from the SR- μ CT data (pixel size $6.5 \mu\text{m}$) before segmentation.

the geometric coordinates coincide with the material coordinates defined by the faces of the specimen. As perpendicularity and parallelism errors were about 1° (Cai et al. 2017), the orientation correction was small. Thereafter, axis x_3 was approximately along the direction of osteons and axes x_1 and x_2 were perpendicular to osteons. The images were then binarized by simple thresholding to obtain two phases : pores and mineralized matrix (Cai et al. 2019a). In each specimen, a VOI of approximately $2.8 \times 3.9 \times 4.8 \text{ mm}^3$ corresponding to one representative volume element (RVE) for the homogenization was selected manually. The VOI size was much larger than the minimum RVE size for cortical bone (Grimal et al. 2011a). After a convergence study (Cai et al. 2019a), the voxel size was then increased to $35 \mu\text{m}$ to reduce the computational cost, which lead to about 1.2 million voxels per specimen.

We then built RVEs made of three phases: the pores and two phases within the mineralized matrix to model its heterogeneity, that is, an assembly of osteonal (O) and interstitial (I) tissues. Images were processed to obtain segmentation masks for the osteonal and interstitial phases based on the observation that the Haversian canals run roughly along an osteon's center line (Maggiano et al. 2016). The osteonal tissue was defined to be within a ring of a given thickness e around pores. Each slice of the 3D image stack, which was roughly perpendicular to the osteon axis, was processed independently. The ring was obtained by erosion with a disk as structuring element (Figures 3 and 4).

2.2 Notation for the elasticity tensor

We use the classical two-indices Voigt notation for the stiffness tensor components C_{ij} . For an orthotropic material with principal directions aligned with the frame axes $(\mathbf{x}_1, \mathbf{x}_2, \mathbf{x}_3)$, the stiffness tensor \mathbf{C} has nine independent moduli : C_{ii} ($i = 1, 2, 3$) denote the longitudinal stiffnesses, C_{ii} ($i = 4, 5, 6$) denote the shear moduli and only three non-diagonal terms are different from zero: C_{12} , C_{13} and C_{23} . For a transversely

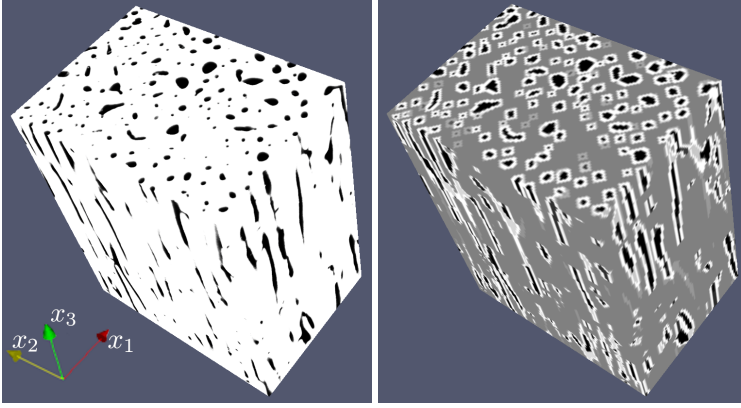


Fig. 3: Three-dimensional representative volume element (RVE) with vascular pores in black. Left: two-phase model where the matrix (white) is considered to be a mixture of interstitial and osteonal tissue. Right: three-phase model where the matrix is made of two phases (white: osteonal tissue; dark gray: interstitial tissue). The direction of the bone diaphysis axis (and approximate osteon axis) is given by \mathbf{x}_3 .

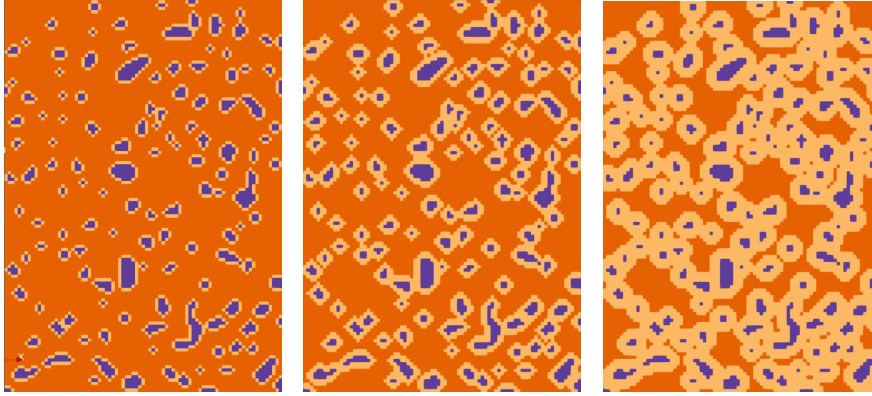


Fig. 4: A transverse cross-section of one cortical bone RVE showing the vascular pores (blue) and the two phases within the mineralized matrix (light orange: osteonal tissue; dark orange: interstitial tissue) for different values of osteon thickness: $e = 35\mu m$ (left), $e = 70\mu m$ (center), and $e = 140\mu m$ (right)

isotropic material with symmetry axis \mathbf{x}_3 , additional conditions hold: $C_{11} = C_{22}$, $C_{12} = C_{11} - 2C_{66}$, $C_{13} = C_{23}$, and $C_{44} = C_{55}$ (five independent moduli).

2.3 Mineralized matrix elasticity model

Following Grimal et al. (2008, 2011b), the mineralized matrix elastic properties are modeled in a transversely isotropic framework as functions of the mineral volume fraction f_{ha} . We use the “mineral foam matrix with collagen inclusions” micromechanical

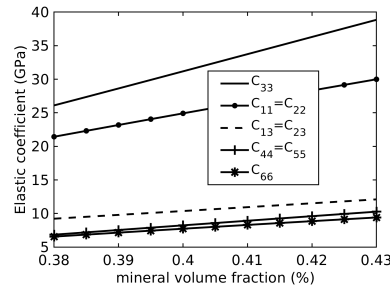


Fig. 5: Elastic coefficients of the mineralized matrix as a function of mineral volume fraction f_{ha} (Grimal et al. 2008, 2011b).

model introduced by Hellmich et al. (2004) which is based on two idealizations: (1) at a length scale of 100 nm, hydroxyapatite crystals and ultrastructural water with non-collagenous organic material constitutes a mineral foam; (2) at a length scale of 5 to 10 microns, collagen fibers are embedded into the mineral foam. Volume fractions of the collagen fibers and water-filled nanoscale pores are respectively denoted f_{col} and f_w . The model assumes fixed elastic properties of the constituents (mineral, collagen, and water) and uses self-consistent and Mori-Tanaka homogenization schemes to derive the effective properties. Since $f_{ha} + f_{col} + f_w = 1$ the model has in fact only two independent parameters. Furthermore, using an empirical relationship between f_{col}/f_w and f_{ha} (Raum et al. 2006; Broz et al. 1995), the mineralized matrix stiffness tensor \mathbf{C}^m is obtained as a function of a single parameter which we choose to be f_{ha} . The evolution of matrix elastic coefficients as a function of f_{ha} is plotted in Figure 5 for illustration. Different values of f_{ha} are assumed for the osteonal and interstitial tissues as described below.

2.4 Choice of RVE parameters

Each RVE is defined by a specific microstructure θ (i.e, a 3D image of the vascular porosity), the transversely isotropic elastic properties of osteonal and interstitial tissues as well as osteon thickness e . As the elastic properties of the matrix phases are functions of f_{ha} , as explained above, the RVE effective elasticity tensor is a function $\tilde{\mathbf{C}} = f(\theta, e, \mathbf{C}^{m;O}, \mathbf{C}^{m;I})$ with $\mathbf{C}^{m;O}$ and $\mathbf{C}^{m;I}$ the elastic stiffness tensors of osteonal and interstitial tissues, respectively.

Interstitial tissue is assumed to be fully mineralized, that is, $\mathbf{C}^{m;I} = \mathbf{C}^m(f_{ha}^I = 0.43)$ and we consider a mineral volume fraction of osteonal tissue $f_{ha}^O = 0.40$, $\mathbf{C}^{m;O} = \mathbf{C}^m(f_{ha}^O = 0.4)$. These constitutive assumptions correspond to a longitudinal stiffness $C_{33}^{m;O}$ which is 19.75% smaller than $C_{33}^{m;I}$ (Figure 6). These values were selected based on the elasticity differences observed in nanoindentation measurements of osteonal and interstitial tissues. For instance, Rho et al. (1997) measured dehydrated tibia specimens embedded in epoxy resin and reported Young's moduli measured along the osteon axis of 22.5 ± 1.3 GPa and 25.8 ± 0.7 GPa in osteonal and interstitial tissues respectively, that is, a Young's modulus 12.8% smaller in osteonal tissue. Zysset et al. (1999) reported Young's moduli measured along the osteon axis in diaphyseal wet femoral bone specimens of 19.1 ± 5.4 GPa in osteonal and 21.2 ± 5.3 GPa in interstitial

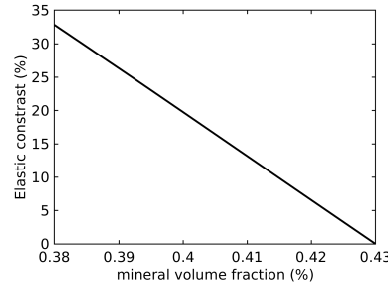


Fig. 6: Difference in elastic properties between the interstitial and osteonal tissues, calculated as $100 \times \frac{C_{33}^{m;I} - C_{33}^{m;O}}{C_{33}^{m;I}}$, as a function of the mineral content of the osteonal tissue f_{ha}^O . The mineral content of the interstitial tissue is kept constant : $f_{ha}^I = 0.43$

tissue, and in the femoral neck of 15.8 ± 5.3 GPa in osteonal and 17.5 ± 5.3 GPa in interstitial tissue, that is, a Young's modulus about 10% smaller in osteonal tissue.

For the purpose of comparison, we also calculated the stiffness tensor $\mathbf{C}^{m;hom}$ of an equivalent homogeneous matrix. The elasticity tensor is obtained from the mineralized matrix elasticity model by considering the average mineral content $\langle f_{ha} \rangle$, that is

$$\mathbf{C}^{m;hom} = \mathbf{C}^m(\langle f_{ha} \rangle), \quad \langle f_{ha} \rangle = \frac{\nu^I f_{ha}^I + \nu^O f_{ha}^O}{\nu^I + \nu^O} \quad (1)$$

where ν^I and ν^O are the volume fractions of interstitial and osteonal phase, respectively. Due to the quasi-linear variation of the elastic moduli in the considered range of mineral volume fraction (Figure 5), it almost coincides with the Voigt bound

$$\mathbf{C}^m(\langle f_{ha} \rangle) \approx \langle \mathbf{C}^m \rangle. \quad (2)$$

Osteon thickness e was chosen according to literature data reporting dimensions of the Haversian canals and of the osteons. Britz et al. (2009) conducted a two-dimensional histomorphometric analysis on microradiographs of femoral diaphyseal tissue from 88 donors (male and female 17 to 97 years old from the Melbourne Femur Research Collection (<https://dental.unimelb.edu.au/research/melbourne-femur-research-collection>)). They reported osteon diameter in the range $[155 - 325]$ μm with average of 220 ± 28 μm . Cooper et al. (2007) analyzed the properties of the vascular porous network in three-dimensions from Micro-CT images of 79 donors (males and females 18 to 92 years old, also from the Melbourne Femur Collection). They reported pore diameters in the range $[56.0 - 456.3]$ μm with average of 117.2 ± 89.6 μm . Gauthier et al. (2019) et al. conducted a SR- μCT study on samples from the radius of eight female donors (70.3 ± 13.7 y.o.). Osteonal canal diameter was (mean, SD) 64.7 ± 23.1 μm and osteon diameter was 184.0 ± 13.3 μm . Based on these mean values of osteon and pore diameters, the average thickness of the osteonal wall is between 50 and 60 μm with an expected large interval of variation which cannot be calculated from the experimental data of different sources.

We used $e = 35, 70, 105$ and 140 μm . The smallest value corresponds to the typical (average) size of an osteon. The other values were chosen in order to model a maximum effect of the matrix heterogeneity on effective elastic properties at the mesoscale.

Pores were assumed to be filled with a fluid whose elastic properties are similar to those of water, that is a bulk modulus equal to 2.2 GPa and a null shear modulus. The parameters of the RVE are summarized in Table 1.

3 Overall elasticity inferred from numerical homogenization

To investigate the influence of the bone tissue heterogeneity on the effective elastic properties, we have performed unit-cell computations on the RVE samples with the microstructural assumptions previously described. The size of the constitutive heterogeneities (i.e pores and osteons) being much smaller than the sample volume, the Hill-Mandel macro-homogeneity condition is fulfilled. The overall properties can thus advantageously be determined by assuming periodic-boundary conditions on the microstructural unit-cell Ω . The local problem to be solved thus reads, $\forall \mathbf{x} \in \Omega$,

$$\begin{cases} \text{curl}(\text{curl}^T \boldsymbol{\varepsilon}(\mathbf{x})) = \mathbf{0}, & \text{div } \boldsymbol{\sigma}(\mathbf{x}) = \mathbf{0}, \\ \boldsymbol{\sigma}(\mathbf{x}) = \mathbf{C}(\mathbf{x}) : \boldsymbol{\varepsilon}(\mathbf{x}), \end{cases} \quad (3)$$

with periodicity conditions on the boundary $\partial\Omega$ (Suquet 1987). $\boldsymbol{\varepsilon}$ and $\boldsymbol{\sigma}$ are respectively the linearized strain and Cauchy stress. \mathbf{C} is the tensor field of the local elastic moduli which values correspond to the elastic properties of the pores, the osteonal or the interstitial tissue (Table 1) depending on the coordinate. To solve the problem (3), we have used a fast Fourier transform (FFT) based numerical scheme (Moulinec and Suquet 1998; Moulinec and Silva 2014). The principle of this method is to solve iteratively the implicit integral equation for the strain field $\boldsymbol{\varepsilon}(\mathbf{x})$

$$\begin{cases} \boldsymbol{\varepsilon}(\mathbf{x}) = \mathbf{E} + \int_{\Omega} \boldsymbol{\Gamma}^{(0)}(\mathbf{x} - \mathbf{x}') : \boldsymbol{\tau}(\mathbf{x}') d\mathbf{x}', \\ \boldsymbol{\tau}(\mathbf{x}) = (\mathbf{C}(\mathbf{x}) - \mathbf{C}^{(0)}) : \boldsymbol{\varepsilon}(\mathbf{x}) \end{cases} \quad (4)$$

with \mathbf{E} the macroscopic strain and $\boldsymbol{\Gamma}^{(0)}$ denoting the strain Green operator corresponding to a reference homogeneous medium with elasticity $\mathbf{C}^{(0)}$. This numerical method, which is widely used in engineering material mechanics, has been recently used for cortical and trabecular bones (Colabella et al. 2017; Gagliardi et al. 2018; Cai et al. 2019a, 2020). It can be mentioned that it allows to perform calculations directly on the microstructure digital image. The effective elastic moduli tensor $\tilde{\mathbf{C}}$ is classically defined by

$$\langle \boldsymbol{\sigma} \rangle_{\Omega} = \tilde{\mathbf{C}} : \langle \boldsymbol{\varepsilon} \rangle_{\Omega} \quad (5)$$

where $\langle \cdot \rangle_{\Omega}$ denotes a volume average over the unit-cell. No symmetry assumptions have been made on $\tilde{\mathbf{C}}$ and six independent loadings were performed, for each sample, to determine the whole set of overall elastic coefficients (21). The reader is referred to Cai et al. (2019a) for a detailed convergence analysis.

4 Results and discussion

The mineral content of the interstitial and osteonal tissues being chosen, the effective elasticity tensor $\tilde{\mathbf{C}}$ is solely a function of the specific pore network microstructure

θ and the considered osteon thickness e . Unit-cell FFT computations have been performed for each parameter pairs (θ, e) by considering either an heterogeneous matrix (pores coated with osteons) or an equivalent homogeneous matrix. Numerical results are reported in Figure 7 for the 25 different pores network microstructures with porosities ranging from about 3 to 20%. The relative errors of elastic moduli (i.e., the relative difference between effective elastic moduli of heterogenous matrix mineralization and homogeneous matrix mineralization) as a function of porosity are shown in Figure 8 for the largest osteon thickness. The average error values are given in Table 2 for each osteon thickness.

The relative error between effective elasticity coefficients using a homogeneous versus a heterogeneous matrix depends on the porosity and the osteonal thickness. The error weakly depends on the elastic modulus and varies from $\sim 1\%$ for $e = 35 \mu\text{m}$ to $\sim 7\%$ for $e = 140 \mu\text{m}$. For $e = 140 \mu\text{m}$, it can be observed that the difference is almost constant for a porosity between 5 and 10%. Nevertheless, fluctuations of the error of about 2% are observed between samples of similar porosity but different microstructure, evidencing an influence of the distribution of pores on the error. For lower or higher porosity values, the difference decreases. More bone samples with a high porosity would be necessary for a more quantitative analysis.

The equivalent homogeneous matrix elasticity (Eq. 1) is almost defined by the Voigt upper bound because the 'homogeneous' mineral content is calculated as the arithmetic mean. Accordingly, accounting for the matrix morphological heterogeneity in the homogenization is expected to lead to lower effective moduli, which is observed in our results. Furthermore, for a given microstructure, this softening effect consistently increases with the osteon thickness, except for high porosity values. At a larger scale, by modeling the global stiffness of a mouse femur from high-resolution CT, Blanchard et al. (2013) also observed that calculating apparent properties from the average mineral content overestimates the stiffness compared to a model incorporating the heterogeneity of the mineral distribution in the femur.

In our work, the calculated error should be considered as an upper bound because we have designed the study to model a maximum effect of the heterogeneity. We selected values of f_{ha} for the osteonal and interstitial phases leading to differences in elastic properties in the upper range of the differences observed experimentally. We have also considered that all pores are surrounded by softer tissue (young osteon) and that the thickness of the osteons' wall around canals is constant inside each RVE. In reality many pores are surrounded by older tissue, reducing the difference in mechanical properties with interstitial tissue.

We used an idealized model of cortical bone that incorporates the main features of the heterogeneity and that was parametrized in order to observe a range of effects. More realistic and sample-specific models could be designed using calibrated SR- μCT data providing DMB for each voxel. The specific elastic heterogeneity could be reconstructed after converting DMB to elasticity for each voxel. A major drawback of this approach is that the law to convert DMB to a stiffness tensor at this scale is not known and likely dependent on the location in the tissue. Another sample-specific approach would be to perform a segmentation of osteons based on the differences of gray level between osteons and interstitial tissue (Gauthier et al. 2019) and then allocate a different stiffness per tissue type. However, even with high quality acquisitions this would have required a prohibitive amount of work for the samples of the present study.

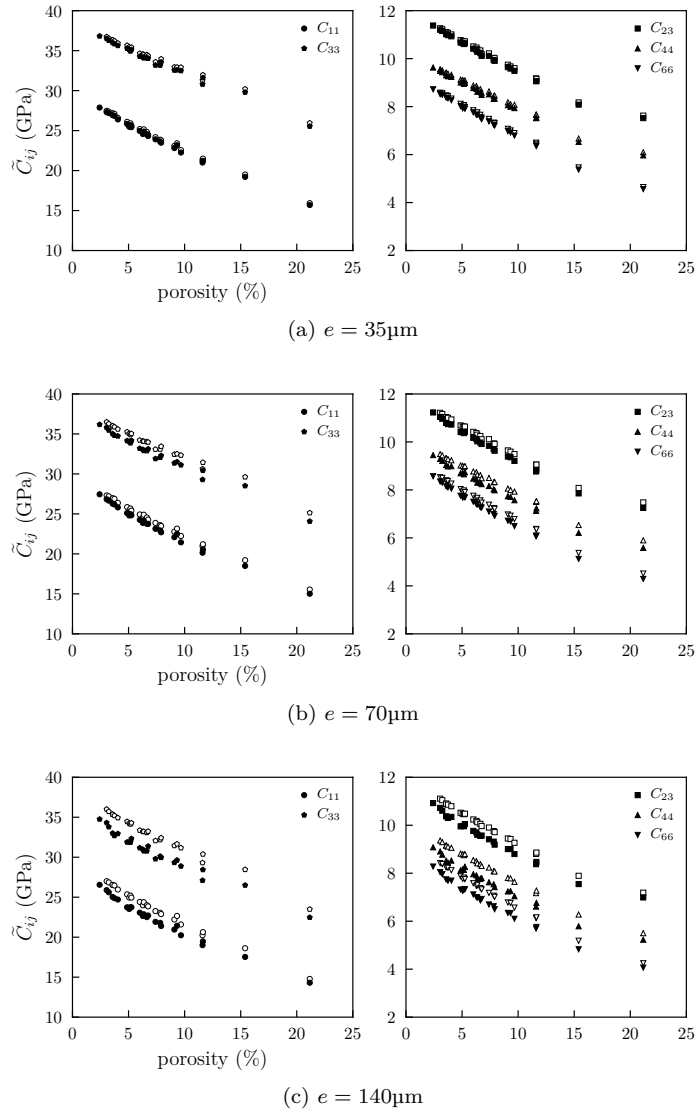


Fig. 7: Evolution of the effective elastic coefficients \tilde{C}_{ij} with porosity for different osteon thicknesses: (a) $e = 35\mu\text{m}$, (b) $e = 70\mu\text{m}$ and (c) $e = 140\mu\text{m}$ (solid symbols: two-phase bone matrix composed of interstitial tissue and osteons; open symbols: equivalent homogeneous matrix)

Considering more realistic models of tissue heterogeneity would likely lead to errors smaller than the errors we calculated. Interestingly, the error is found to be the largest in the most common porosity interval for non-pathological bone ($\sim 5\text{-}10\%$). For high porosities we have observed a reduction of the error that could be due to the fact that in these cases, the osteonal tissue covers a very large fraction of the matrix volume resulting in a quasi-homogeneous matrix. However, in the case of high

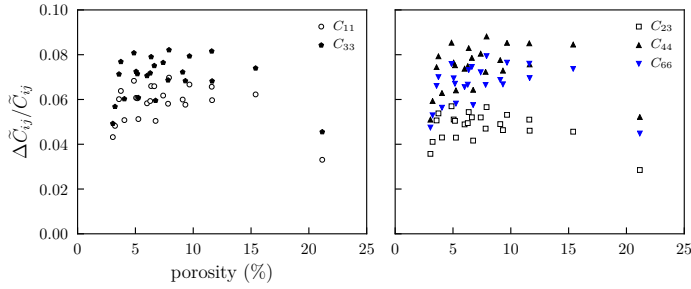


Fig. 8: Relative difference for thickness $e = 140\mu\text{m}$. Note that the denominator represents the effective elastic moduli considering homogeneous matrix mineralization $\tilde{C} = \tilde{C}_{hom. matrix}$ and $\Delta\tilde{C} = |\tilde{C}_{hom. matrix} - \tilde{C}_{het. matrix}|$.

porosities, this error might be somewhat underestimated : some of the osteons are in reality more mineralized leading to an increased heterogeneity.

Modeling cortical bone elastic properties, as any modeling approach, is a matter of idealizations consisting in neglecting certain details of the organization of the tissue. The results of the present study complete a detailed analysis of some idealizations relevant for cortical bone mesoscale elasticity modeling conducted during the last decade. The accuracy of more or less refined models has been evaluated by comparing model predictions to experimental values of anisotropic elastic coefficients obtained from samples of human donors (Granke et al. 2011; Baumann et al. 2012; Cai et al. 2019b). The samples were typically imaged with high-resolution μCT (as for the present work) to obtain a representation of the microstructure and the matrix mineral content. Then, the question arises as to whether an accurate estimation of elastic properties can be obtained from this information, that is, whether the variability of mesoscale elastic properties can be explained from the mineral content of the matrix and an image of the microstructure.

Porosity (volume fraction of pores) alone is known to explain the largest part of elasticity variations (Granke et al. 2011; Cai et al. 2019b). Indeed, a popular simple model for cortical bone is a homogeneous transversely isotropic matrix with fixed properties pervaded by elongated pores with a single orientation (e.g., (Hellmich and Ulm 2004; Parnell et al. 2012; Baumann et al. 2012; Gagliardi et al. 2017)). This simple model, which disregards the details of the pore structure and the inter-individual variations of matrix elasticity, accounts well for the variations of elastic properties with porosity (Hellmich and Ulm 2004; Granke et al. 2015; Cai et al. 2019b), or with the average orientation of mineral crystals (Baumann et al. 2012), with errors compared to experimental data typically below 10% (average error for a collection of samples). Nevertheless this model has several potential biases and its accuracy for predicting the elasticity of a given specimen may not be as good as 10%. As shown by comparing this simple model to an FFT model accounting for the details of the pore network structure (similar to the present work), disregarding the complexity of the pore network and model it as a collection of cylindrical pores leads to an overestimation of effective elasticity of 1% to 15% depending on the elastic coefficient C_{ij} and porosity level (Cai et al. 2019a).

Apart from the porosity, the average matrix mineral content explains a part of the experimental variation of effective mesoscale elasticity: taken together, these

two variables were found to explain 76 to 91% of the variability of the different elastic coefficients (Cai et al. 2019b). In the elderly population without specific bone pathology, the coefficient of variation of the average mineral content was found to be 2% (Cai et al. 2019b). Using inverse homogenization based on mesoscale anisotropic stiffness data for the same samples, Cai et al. (2020) calculated a coefficient of variation of matrix elastic coefficients between 3 and 7% depending on the elastic constant, which was correlated to the mineral content. (Note that above reported percentages are for coefficients of variations, and that the actual ranges of variation are somewhat larger). Accordingly, an accurate model of cortical bone at the mesoscale should also consider an accurate modeling of the anisotropic elastic properties of the matrix, which would reflect the mineral content of the bone matrix. Then, the question arises as to whether such a model of matrix elasticity should account for the heterogeneous distribution of the mineral at the scale of 10-100 μm , e.g., lower mineral content in osteons compared to interstitial tissue. This is precisely the contribution of the present work where we have shown that disregarding the heterogeneity of the matrix and assuming that its elasticity can be derived from the average matrix mineral content leads to an overestimation of effective elasticity of a maximum of about 7% in extreme cases.

The main limitation of the present study is that we have used a relatively simple micromechanical model to obtain the elasticity tensor with a single parameter f_{na} associated to the mineral content. Due to the coarse assumptions of the micromechanical model, we argue that this parameter should not be seen as the actual volume fraction of mineral content but rather a lumped parameter driving the matrix stiffness. The specific values of this parameter for osteonal and interstitial tissues in the present study were defined based on the calculated elastic parameters and corresponding experimental data. An alternative could be to use empirical relationships between each matrix elastic coefficient and mineral content (Cai et al. 2020) or more sophisticated models (Hellmich et al. 2004; Hamed et al. 2010). The rationale for modeling the elasticity tensor with a single parameter is that experimentally, in a population of individuals without documented bone pathology, the collagen properties or the properties of the mineral crystals (such as cristallinity) are not correlated with elastic properties contrary to the mineral content (Cai et al. 2019b).

Another potential modeling issue is the bias introduced by the scheme to calculate effective properties. This, however, can be ruled out as we have shown that for a sufficiently large RVE (typically larger than $(1.5)^3 \text{ mm}^3$) the upper (kinematic uniform boundary conditions) and lower (static uniform boundary conditions) bounds of apparent properties are very close (Grimal et al. 2011a).

5 Conclusion

For typical experimental values of osteon thickness and realistic porous microstructures, the error on effective elasticity due to neglecting the mineralized matrix heterogeneity (e.g., not distinguishing osteonal and interstitial tissue) is about 7% at the maximum. This suggests that matrix heterogeneity can likely be disregarded in cortical bone models in most practical cases. In particular, it seems less important to model this heterogeneity compared to (1) the individual-specific matrix elastic properties scaled to the average matrix mineral content and (2) the effect of the complex pore architecture which is not fully captured when the porosity is modeled

as a collection of ellipsoids or cylinders. These conclusions should hold for bone tissue without a specific pathology. The effect of matrix heterogeneity may however turn out to be more important in some pathologies (Roschger et al. 2008).

This work provides data to quantitatively discuss the handling of mineralized matrix heterogeneity in cortical bone elasticity models. The results should be helpful to guide the interpretation of effective elastic properties at the millimeter scale. These are involved in finite element models for the prediction of bone response to loads for the assessment of strength (van Rietbergen and Ito 2015; Engelke et al. 2016) or remodeling (Martelli et al. 2014). Experimentally-derived effective elastic coefficients can be used to determine mineralized matrix properties by inverse approaches (Sanz-Herrera et al. 2019; Cai et al. 2020). Finally, effective elasticity can also be probed in vivo with ultrasound, from which porosity can be estimated by an inverse approach (Minonzio et al. 2019).

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Conflict of interest

The authors declare that they have no conflict of interest.

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Microstructure (θ)	Osteon thickness (e , μm)	Elastic material inside pores	Elasticity of osteonal tissue	Elasticity of interstitial tissue
According to a high-resolution image (25 samples)	35 ; 70 ; 105 ; 140	Bulk modulus 2.2 GPa, null shear modulus.	$\mathbf{C}^{m;O} = \mathbf{C}^m(f_{ha}^O = 0.4)$.	$\mathbf{C}^{m;I} = \mathbf{C}^m(f_{ha}^I = 0.43)$

Table 1: Summary of parameters of the representative volume element. Elasticity tensors of osteons $\mathbf{C}^{m;O}$ and interstitial tissue $\mathbf{C}^{m;I}$ are calculated with a micromechanical model for a fixed volume fraction of mineral f_{ha} .

e (μm)	35	70	105	140
$\Delta\tilde{C}_{11}/\tilde{C}_{11}$ (%)	1.05	2.94	4.75	5.84
$\Delta\tilde{C}_{33}/\tilde{C}_{33}$ (%)	1.1	3.22	5.42	6.97
$\Delta\tilde{C}_{23}/\tilde{C}_{23}$ (%)	0.94	2.51	3.87	4.78
$\Delta\tilde{C}_{44}/\tilde{C}_{44}$ (%)	1.34	3.77	5.92	7.4
$\Delta\tilde{C}_{66}/\tilde{C}_{66}$ (%)	1.24	3.64	5.16	6.64

Table 2: Relative difference averaged over all microstructural samples with porosities ranging from 3 to 20% for different osteon thicknesses e (See the definition of $\Delta\tilde{C}/\tilde{C}$ in the legend of Fig. 8)