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Cortical bone viscoelastic damping assessed with resonant ultrasound spectroscopy reflects porosity and mineral content

Fan Fan^{b,a,*}, Xiran Cai^{b,c}, Hélène Follet^d, Françoise Peyrin^e, Pascal Laugier^{b,a}, Haijun Niu^a, Quentin Grimal^b

^aBeijing Advanced Innovation Center for Biomedical Engineering, School of Biological Science and Medical Engineering, Beihang University, 100083, Beijing, China ^bSorbonne Université, INSERM UMR-S 1146, CNRS UMR 7371, Laboratoire d'Imagerie Biomédicale, F-75006, Paris, France

^cSchool of Information Science and Technology, ShanghaiTech University, 201210, Shanghai, China

^dUniv Lyon, Université Claude Bernard Lyon 1, INSERM, LYOS UMR 1033, F-69008, Lyon, France

^eUniv Lyon, INSA-Lyon, Université Claude Bernard Lyon 1, UJM-Saint Etienne, CNRS, Inserm, CREATIS UMR 5220, U1206, F-69621, Lyon, France

Abstract

Viscoelasticity is an essential property of bone related to fragility, which is altered in aging and bone disease. Bone viscoelastic behavior is attributed to several mechanisms involving collagen and mineral properties, porosities, and bone hierarchical tissue organization. We aimed to assess the relationships between cortical bone viscoelastic damping measured with Resonant Ultrasound Spectroscopy (RUS), microstructural and compositional characteristics. We measured 52 bone specimens from the femur of 26 elderly human donors. RUS provided a shear damping coefficient at a frequency of the order of 150 kHz. The characteristics of the structure of the vascular pore network and tissue mineral density were measured using synchrotron radiation high-resolution computed tomography (SR- μ CT). Fourier transformed infrared microspectroscopy (FTIRM) was used to quantify mineral-to-organic phase ratio, mineral maturity, crystallinity, and collagen maturity. Cross-links were quantified from biochemistry. Viscoelastic damping was found to increase with vascular porosity (r = 0.68), to decrease with the degree of mineralization of the extravascular matrix (r = -0.68), and was marginally affected by collagen. We built a multilinear model suggesting that when porosity is controlled, the variation of mineral content explains a small additional part of the variability of damping. The work supports the consideration of viscoelasticity measurement as a potential biomarker of fragility and provides a documentation of bone viscoelastic behavior and its determinants in a frequency range rarely investigated. Keywords: Cortical bone, Damping, Quality factor, Resonant ultrasound spectroscopy, Porosity, Mineral content

^{*}Corresponding author

Email address: fanfan@buaa.edu.cn (Fan Fan)

1. Introduction

Viscoelasticity is an essential property of biological tissues [1]. For example, hard biological tissues, such as bone, exhibit creep and stress relaxation, rate dependent response to a dynamic loading, phase lag between stress and strain during oscillatory loading, and damping of elastic waves [2]. Bone viscoelastic behavior is attributed to several mechanisms [3] involving all scales from the nanoscale to the mesoscale, i.e. the scale of a few millimeters in cortical bone [4]. The collagen of the extracellular matrix is itself viscoelastic. The movement of fluids within the pores, heat flow under mechanical loading between heterogeneous regions such as osteons and lamellae, are some of the other mechanisms associated to the viscoelastic behavior.

Viscoelasticity of bone has been investigated by many authors motivated by the relationship between damping mechanisms and mechanical behavior beyond the elastic limit and related to bone fragility. Firstly, 10 collagen was shown to have a profound effect on bone fragility, because changes in collagen content, or changes 11 to collagen cross-linking, reduce the energy required to cause failure [5]. Secondly, bone strength [6, 7] and 12 toughness [8] are rate dependent suggesting a role for viscous mechanisms. Thirdly, there is a close relationship 13 between microdamage and viscoelasticity as loading cortical bone past the yield stress changes its viscoelastic 14 properties [9]. Finally, the viscoelastic dissipation of energy at micron and sub-micron scales is related to crack 15 initiation and propagation [10, 11]. Bone material quality and fragility are multifaceted phenomena involving 16 several scales and they cannot be fully captured from a single mechanical measurement. This has led several 17 authors to suggest that the measurement of viscoelasticity could bring unique information related to fragility 18 associated with bone disease [12–14] and tissue alteration during aging [15]. Yet, viscoelastic data is scarce 19 in comparison to published data on elastic properties. In particular, the relationships between viscoelasticity, 20 extravascular matrix composition, and microstructure are poorly documented. 21

Viscoelasticity has been assessed with a variety of techniques such as microscale measurement of creep 22 with nanoindentation [15–17], mesoscale measurement of an oscillatory response in torsion (up to 50 kHz) 23 [18], or 3-point bending with dynamic mechanical analyzers (in the range 1-20 Hz) [19]. In material science, 24 another popular approach is to use resonant ultrasound spectroscopy (RUS), a technique to measure the 25 anisotropic stiffness and viscoelastic damping assessed from the width of a resonance of a freely vibrating 26 specimen [20-23]. In a viscoelastic material such as bone, the resonant peaks corresponding to the different 27 eigenmodes of the measured specimen tend to overlap. It follows that RUS is essentially practicable to measure 28 bone damping associated with the first eigenmode which is associated to a shear modulus [23, 24]. A typical 29 cortical bone specimen for RUS measurements is a cuboid with longest dimension around 5 mm, having its 30 first eigenfrequency around 150 kHz. 31

The purpose of the present work was to assess the relationships between cortical bone viscoelastic damping measured with RUS, tissue composition, porosity, and microstructure. We measured a collection of bone specimens from elderly human donors with RUS, providing a quality factor (equivalent to tan δ in torsion experiments), associated to a shear modulus, at a frequency of the order of 150 kHz. Vascular porosity and the degree of mineralization of bone were obtained from synchrotron radiation high-resolution computed tomography (SR- μ CT), cross-links were quantified from biochemistry, Fourier transformed infrared spectroscopy (FTIR) was used to quantify mineral-to-organic phase ratio, mineral maturity, crystallinity, and collagen maturity. Note that the relationship between elastic properties and these variables was previously reported in
 [25].

The work provides a documentation of bone viscoelastic behavior and its determinants in a frequency range rarely investigated, complementing existing data. Comparison with the viscoelastic behavior at other frequencies may provide insight into the mechanics of bone viscoelasticity. While our results do not reflect the viscoelastic behavior of bone at the frequencies of physiological loading, they are of practical interest for some ultrasonic applications around 10⁵ Hz such as bone ultrasonic drilling, ultrasonic stimulation of bone healing and bone health assessment with guided waves.

47 2. Materials and methods

48 2.1. Specimens

We have used a collection of specimens from a previous study [25]. The preparation of specimens is briefly 49 recalled here. Cortical bone specimens were harvested from the left femur of 26 human cadavers. The femurs 50 were provided by the Départment Universitaire d'Anatomie Rockefeller (Lyon, France) through the French 51 program on voluntary corpse donation to science. Among the donors, 14 were females and 12 were males 52 $(50-95 \text{ years old}, 77.3 \pm 11.5, \text{mean} \pm \text{SD})$. As shown in Figure 1, in each of the lateral and medial anatomical 53 quadrants, adjacent specimens (# 1, # 2 and # 3) were prepared along the axial direction, to be measured 54 by several techniques described below. All specimens were frozen and stored at -20° C between tests. They 55 were then slowly thaved and immersed in 0.9% NaCl saline before testing to ensure full hydration [26]. The 56 nominal size of the specimens #1 used for RUS was $3 \times 4 \times 5mm^3$ in radial (x_1) , circumferential (x_2) and axial 57 (x_3) directions, respectively. These specific dimensions were chosen so as to maximize the size of the specimens 58 while complying with the technical requirements of RUS [22]. Figure 2 is the three-dimensional rendering of 59 the SR- μ CT image of a bone specimen. Specimens #1 used in RUS were kept hydrated prior to experiment 60 and all RUS measurements were made on a fully hydrated specimen. After a first set of RUS measurements, 61 specimens #1 were defatted following a protocol which prevents the risk of infections and allows the specimen 62 conservation at room temperature [26]. 63

64 2.2. RUS experiment

Setup and signal processing methods dedicated to the RUS measurement of attenuative materials, exten-65 sively described elsewhere [22, 27], were used in this study. Briefly, the specimen #1 was mounted on opposite 66 corners between two shear wave transducers (V154RM, Panametrics, Waltham, MA, USA). The frequency 67 response (vibration spectrum) was recorded using a vectorial network analyzer (Bode 100, Omicron Electron-68 ics GmbH, Klaus, Austria) and a broadband charge amplifier (HQA-15M-10T, Femto Messtechnik GmbH, 69 Berlin, Germany). The frequency band of analysis was 100-700 kHz, including the first resonant frequency 70 of the specimen. Six successive spectra were recorded for each specimen, with intermediate rotation (without 71 unmounting) of the specimen by a small angle between each measurement to vary the relative amplitudes of 72



Figure 1: Specimen preparation procedure.(a) A cross-section of femoral bone at the mid-diaphysis was extracted. (b) The vertical and upper-front view of the cross section, which was then cut into 4 pieces (lateral, medial, anterior and posterior). Two of these pieces (lateral and medial) were then used. (c) 3 rectangular parallelepiped shaped specimens (set #1, #2 and #3) were prepared along the axial direction at both the lateral and medial quadrants. (d) RUS measurements for bone viscoelastic damping. (e) SR- μ CT scanning for Bone microstructural parameters and DMB. (f) Bone residues close-by #2 after cutting for #2 went to biochemistry experiments for the collagen and cross-links. The data experiments carried on #2 were not shown in this work. (g) FTIRM tests for bone compositional information.



Figure 2: Three-dimensional rendering of the SR- μ CT image of a bone specimen of approximate dimensions $3 \times 4 \times 5mm^3$.

- ⁷³ the excited resonant modes in order to maximize the number of detectable resonant frequencies (Fig. 3). A
- ⁷⁴ selected portion of each complex spectrum can be fitted by a sum of M Lorentzian line-shapes (each describing

⁷⁵ the behavior of a one degree-of-freedom mechanical resonator) :

$$L(f) = \sum_{k=1}^{M} \frac{a_k}{(f_k^2 - f^2) + i(f_k f/Q_k)},$$
(1)

with f the frequency, a_k the complex amplitudes, f_k the resonant frequencies and Q_k the quality factors. The quality factor is related to the width of the resonant peak as $Q_k \sim f_k/\Delta f$, where Δf is the -3db bandwidth (half-power bandwidth).

⁷⁹ Upon combining the six spectra and fitting with Eq. (1), between 20 to 30 resonant frequencies f_k were ⁸⁰ extracted for each specimen [22]. Frequencies f_k , which are nearly equal to the eigenfrequencies of the freely ⁸¹ vibrating specimen, were then used to determine the coefficients C_{ij} of the stiffness tensor [20, 27]. These ⁸² were obtained in a previous work presented in [25]. We used Voigt notation for the stiffness tensor and we ⁸³ assumed that bone is a transversely isotropic material (plane 1–2 is the plane of isotropy). As a consequence, ⁸⁴ $C_{11} = C_{22}, C_{13} = C_{23}, C_{44} = C_{55}.$

For the purpose of the present study, we specifically processed the first resonant peak (around 150 kHz in Fig. 3) to assess shear mode damping. As explained in section 2.3, the shear damping coefficient Q_{44}^{-1} (associated to elastic coefficient C_{44}) can be obtained from the quality factor of the first resonant mode Q_1 (defined in Eq. (1)). The quality factor Q_1 in each spectrum was obtained as follows :

• A portion of the spectrum (bandwidth) was manually selected, containing only the first peak f_1 (Case 1, Fig. 4 left) or the two first peaks (Case 2, Fig. 4 right). Precisely, Case 1 corresponds to specimens with a well-isolated first resonant peak. For other specimens with first two relatively close resonant peaks (Case 2), we selected a portion of the spectrum with the first two peaks to account for the potential influence of the second resonance on the Lorentzian lineshape of the first peak. The distance between the two first peaks depends on the exact dimensions of the specimen and its elastic properties. Among the 52 specimens, 10 were in Case 1 and 42 were in Case 2. The effect on frequency and Quality factor determination of the bandwidth selection method is discussed further in Appendix A.

• To determine Q_1 , Eq. (1) was fitted to the spectrum assuming M = 1 (Case 1) or M = 2 (Case 2) using a time domain estimation method based on a linear predictive filter (black dash line in Fig 5), followed by a frequency domain nonlinear optimization (black solid line) [22, 28].

Specimens were measured with RUS before defatting, after defatting, and finally after X-ray irradiation during $SR-\mu CT$.

¹⁰² 2.3. Calculation of material damping

In this section, we present how the measured quality factor Q_1 was used to calculate shear damping coefficient Q_{44}^{-1} following [23, 24]. Bone viscoelasticity was modeled by introducing the complex modulus:

$$C_{ij}^{*} = C_{ij} + iC_{ij}^{'} = C_{ij}(1 + iQ_{ij}^{-1}), \qquad (2)$$



Figure 3: Typical set of spectra measured for a cortical bone specimen. The relative amplitudes of the resonant peaks vary as the specimen is rotated. The width of the first peak provided a measurement of damping. The inset figure is a zoom of in the frequency range of 100 to 200 kHz.



Figure 4: To calculate Q_1 , a portion of the spectrum was selected between the two dash-dot lines. (left) Case 1: for specimens with a well-isolated first resonant peak, only the peak was selected. (right) Case 2: for other specimens, the first two peaks were selected together.



Figure 5: Illustration of the fit of the first peak with Eq. (1). Red dash lines show experimental frequency response of Case 1 (left) and Case 2 (right), frequency response reconstructed with a linear predictive filter (black dash line), and then followed by a frequency domain nonlinear optimization (black solid line).

where C_{ij} and C'_{ij} are the stiffness coefficient and loss modulus, respectively, and $Q_{ij}^{-1} = \frac{C'_{ij}}{C_{ij}}$. This definition of the complex modulus holds for a dynamic loading at a given frequency, and C_{ij} and C'_{ij} are in general functions of frequency. Note that $Q_{ij}^{-1} \sim \tan \delta$, where $\tan \delta$, another popular way of reporting damping, is the phase shift between a harmonic loading and the mechanical response in a vibrational mechanical test [2]. In the present work, the complex modulus is evaluated using the first resonant peak centered at f_1 of each specimen.

In a RUS experiment, a series of resonant frequencies f_k and Quality factors Q_k can in principle be obtained from the resonant spectrum. These are related to the complex modulus through [29–31] :

$$Q_k^{-1} = \sum_{i,j} \frac{2C_{ij}}{f_k} \frac{\partial f_k}{\partial C_{ij}} Q_{ij}^{-1}, \tag{3}$$

113 where

$$\sum_{i,j} \frac{2C_{ij}}{f_k} \frac{\partial f_k}{\partial C_{ij}} = 1.$$
(4)

The linear system of equations Eq. (3) between Q_{ij}^{-1} and Q_k^{-1} is derived with the assumption that $Q_{ij}^{-1} \ll 1$, which is in practice the case for cortical bone [23, 24]. The coefficients of the linear system are the relative sensitivities of the eigenmode k to the loss moduli Q_{ij} .

¹¹⁷ When the stiffness coefficients C_{ij} are available, the system of equations (3) can be inverted to derive ¹¹⁸ Q_{ij}^{-1} from the measured Q_k^{-1} . In practice, quality factors Q_k for $k = 2, 3, \cdots$ can hardly be estimated with ¹¹⁹ a sufficient precision in high damping materials such as bone because of strong resonant peak overlapping. ¹²⁰ Furthermore, several relative sensitivity terms are small, so that the inversion is ill-conditioned and sensitive ¹²¹ to errors in the measured Q_k . Here, we take advantage of the fact that:

1. The first resonant peak is well-separated from the subsequent peaks (Fig. 4) for all specimens. This is a result of the specific aspect ratio of the specimens which was selected so as to optimize the separation of low frequency resonances. As a consequence, Q_1 is estimated with a satisfactory precision ;

2. The terms

$$\frac{2C_{ij}}{f_1}\frac{\partial f_1}{\partial C_{ij}}$$

for $ij \neq 44$ are relatively small.

Hence, the relationship between Q_1^{-1} and Q_{44}^{-1} simplifies to,

$$Q_1^{-1} \approx \frac{2C_{44}}{f_1} \frac{\partial f_1}{\partial C_{44}} Q_{44}^{-1}.$$
 (5)

¹²⁷ The quality of this approximation was checked by calculating the relative sensitivity $\frac{2C_{44}}{f_1} \frac{\partial f_1}{\partial C_{44}}$ of the first ¹²⁸ eigenmode to the shear coefficient C_{44} for the collection of specimens. We found that close to 90% of the value of ¹²⁹ Q_1^{-1} is determined by Q_{44}^{-1} , i.e., $\frac{2C_{44}}{f_1} \frac{\partial f_1}{\partial C_{44}} = 89.7\% \pm 2.0\%$ (mean±SD). From a physical perspective, the first ¹³⁰ eigenmode of the specimens is a pure shear mode involving mostly the shear modulus C_{44} and corresponding ¹³¹ damping Q_{44}^{-1} [20].

132 2.4. Microstructural, mineral and collagen variables

In the present work, we use the same data set as [25] where the measurement protocols of the microstructural, collagen and mineral variables were extensively described. Briefly, (i) Fourier transform infrared mi-

crospectroscopy (FTIRM) was used to measure collagen maturity (CollMat), mineral-to-organic ratio (Mi-135 nOrga), mineral maturity (Minmat), carbonation, and crystallinity index (CryInd); (ii) biochemical measure-136 ments on hydrolyzates prepared from powdered demineralized bone residues provided the amount of enzymatic 137 cross-links (DHLNL, HLNL, PYD and DPD), non-enzymatic cross-links (PEN), and the amount of collagen 138 (Coll); (iii) SR- μ CT (pixel size 6.5 μ m) was used to determine vascular porosity (Ct.Po) and other microstruc-139 tural variables (PoS/PoV, PoN, PoDm, PoSp, PoPf, ConnD and SMI), and the degree of mineralization of 140 bone (DMB). The definitions of these variables are collected in Table 1 grouped as collagen, mineral, and 141 microstructural variables. 142

143 2.5. Data analysis

Normality of the variables was evaluated using the Shapiro-Wilk test. One-way analysis of variance (ANOVA) and Wilcoxon test (for the variables failing the normality test) were performed to evaluate the differences of the data sets from lateral and medial anatomical quadrants. As some variables were not normally distributed, Spearman rank correlation coefficients between Q_{44}^{-1} and each of the mineral, collagen, and microstructural variables were calculated.

The variables significantly correlated with Q_{44}^{-1} were retained for stepwise multiple linear regression analyses. To highlight the relative importance of the explanatory variables in the model, all the variables except Q_{44}^{-1} were normalized between -1 and 1 using the equation,

$$\bar{x} = 2\frac{x - \min x}{\max x - \min x} - 1,\tag{6}$$

where x is the variable to be normalized. The multiple linear regression analyses were firstly carried out in each group of variables (microstructure, mineral and collagen). Then, the most significant variable of each group was retained to find the optimal multiple linear regression model. Linear models were evaluated using the adjusted-r² (Adj- r^2) and root-mean-square-error (*RMSE*).

Note that for some variables which showed significant differences between lateral and medial specimens, the multiple linear regression analyses were carried out both on the data sets of lateral and medial specimens separately and the corresponding model are reported. If these variables were not retained as explanatory variables in the regression model, analyses were run again pooling the data sets from lateral and medial specimens.

Data were considered statistically significant for p < 0.05. Statistical analyses were made using the Matlab 2017a Statistics Toolbox (Mathworks Inc., Natick, MA, USA).

We present below the statistical results for Q_{44}^{-1} measured by RUS before defatting. We found that defatting or irradiation by X-ray for tomography imaging did not significantly modify Q_{44}^{-1} (see Appendix B).

Table 1: Microstructure, mineral and collagen variables and their definition
--

Variable	Unit	Definition	Modality			
Microstructure group						
Ct.Po	%	pore volume fraction	$SR-\mu CT$			
PoS/PoV	mm^{-1}	pore surface to pore volume ratio	$SR-\mu CT$			
PoN	mm^{-1}	pore number per millimeter	$SR-\mu CT$			
PoDm	$\mu { m m}$	average diameter of the pores	$SR-\mu CT$			
\mathbf{PoSp}	μ m	average separation between pores	$SR-\mu CT$			
PoPf	mm^{-1}	pore pattern factor, lower PoPf indicates	$SR-\mu CT$			
		higher concavity, i.e., better-connected pore network				
ConnD	mm^{-3}	connectivity density, a measure of the degree	$SR-\mu CT$			
		to which a pore is multiply connected				
\mathbf{SMI}	a.u.	structure model index, the relative prevalence	$SR-\mu CT$			
		of rods and plates in a 3D pore network				
Mineral	group					
DMB	$ m g/cm^3$	Degree of mineralization of bone	$SR-\mu CT$			
MinOrga	no unit	Mineral to organic ratio, the ratio of the	FTIRM			
		$\nu_1 \nu_3 PO_4$ area (910 - 1184 cm ⁻¹) over the				
		Amide I area $(1592 - 1730 \text{ cm}^{-1})$				
MinMat	no unit	Mineral maturity, the ratio of the ap-	FTIRM			
		atitic ($\sim 1030 \text{ cm}^{-1}$ peak) over non apatitic				
		$(\sim 1110 \text{ cm}^{-1} \text{ peak})$				
Carbon	no unit	Carbonation, the ratio of the $\nu_2 CO_3$ area	FTIRM			
		$(862 - 894 \text{ cm}^{-1})$ over the $\nu_1 \nu_3 \text{PO}_4$ area				
CryInd	cm	Crystallinity index, the inverse of the full	FTIRM			
		width FTIRM at half maximum of the				
		$\sim 604 \text{ cm}^{-1} \text{ peak}$				
Collagen	group					
CollMat	no unit	Collagen maturity, $\sim 1660 \text{ cm}^{-1}$ peak over $\sim 1690 \text{ cm}^{-1}$ peak	FTIRM			
DHLNL	$\rm mmol/mol\ colla-$	Didhydroxylysinonorleucine, immature en-	Biochemistry			
	gen	zymatic cross-links				
HLNL	mmol/mol colla-	Hydroxylysinonorleucine, immature enzy-	Biochemistry			
	gen	matic cross-links				
PYD	mmol/mol colla-	Pyridinoline, mature enzymatic cross-links	Biochemistry			
	gen					
DPD	mmol/mol colla-	Deoxypyridinoline, mature enzymatic	Biochemistry			
DEM	gen	cross-links				
PEN	mmol/mol colla-	Pentosidine, non-enzymatic cross-links	Biochemistry			
C 11	gen					
Coll	%	Collagen percentage by weight	Biochemistry			

165 3. Results

166 3.1. Descriptive statistics

¹⁶⁷ Microstructural, mineral and collagen variables and Q_{44}^{-1} values have been obtained for 52 specimens. For ¹⁶⁸ each specimen measured by RUS, in average 5 values of Q_1 were successfully retrieved from the 6 available ¹⁶⁹ spectra. The average of these values, which was used for the analyses, ranged from 23.93 to 35.31 (30.07 ± ¹⁷⁰ 2.02). The frequency f_1 of the first peak ranged from 115.4 to 160.3 kHz (146.1 ± 8.4 kHz). Descriptive statistics of all variables are given in Table 2. Except for Q_{44}^{-1} , the data is the same as published

¹⁷² in [25] but is recalled here for the convenience of the reader.

Table 2: Descriptive statistics (Mean \pm SD) of Q_{44}^{-1} , microstructure and compositional variables. * Variables in which significant difference were found between the data from lateral and medial specimens.

Q	44							
	0.0373 ± 0.0031							
N	Microstructure variables							
	Ct.Po (%)	$PoS/PoV (mm^{-1})$	$PoN (mm^{-1})$	PoDm (μm)	PoSp (μ m)			
	7.47 ± 4.03	60.53 ± 18.29	0.80 ± 0.22	89.39 ± 31.98	320.22 ± 31.51			
	$PoPf (mm^{-1})$	$ConnD^*$ ((mm^{-3})	SMI^*	(a.u.)			
		L	Μ	L	Μ			
	30.85 ± 8.75	10.32 ± 4.99	24.75 ± 5.84	3.23 ± 0.25	3.05 ± 0.22			
Ν	fineral variables							
	DMB (g/cm^3)	MinOrga	* (n.u.)	* (n.u.) MinMat* (n.u.)				
		L	Μ	L	Μ			
	1.02 ± 0.02	5.26 ± 0.30	5.55 ± 0.26	1.84 ± 0.10	1.72 ± 0.07			
	Carbon [*] (n.u.)		CryInd [*] (cm)					
	Carbo	n* (n.u.)	CryInc	l* (cm)				
	Carbo L	n* (n.u.) M	CryInc L	l* (cm) M				
	$\begin{array}{c} \text{Carbox}\\ \text{L}\\ 0.0071 \pm 0.0003 \end{array}$	$\frac{\text{n* (n.u.)}}{\text{M}}$ 0.0066 ± 0.0002	$\begin{array}{c} \text{CryInc}\\ \text{L}\\ 0.0384 \pm 0.0011 \end{array}$	$1^{*} (cm)$ M 0.0396 ± 0.0006				
C	$\begin{array}{c} \text{Carbox}\\ \text{L}\\ \hline 0.0071 \pm 0.0003\\ \hline \textbf{collagen variables} \end{array}$	$\frac{m^{*} (n.u.)}{M}$ 0.0066 ± 0.0002 s	$\begin{array}{c} {\rm CryInol}\\ {\rm L}\\ 0.0384\pm 0.0011\end{array}$	1^{*} (cm) M 0.0396 \pm 0.0006				
C	$\begin{array}{c} \text{Carbox}\\ \text{L}\\ \hline 0.0071 \pm 0.0003\\ \hline \textbf{collagen variables}\\ \hline \text{Col}\\ \end{array}$	$n^{*} (n.u.) M$ 0.0066 ± 0.0002 s $1Mat^{*}$	CryInc L 0.0384 ± 0.0011 DHLNL	1^{*} (cm) M 0.0396 ± 0.0006 HLNL	PYD			
	Carbo L 0.0071 ± 0.0003 Collagen variables Col (n)	$\frac{n^{*} (n.u.)}{M}$ 0.0066 ± 0.0002 s IMat [*] u.)	$\begin{array}{c} {\rm CryInc}\\ {\rm L}\\ \hline\\ 0.0384 \pm 0.0011\\\\ \hline\\ {\rm DHLNL}\\ {\rm (mmol/mol}\\ \end{array}$	${ m M}^{ m M}$ 0.0396 \pm 0.0006 HLNL (mmol/mol	PYD (mmol/mol			
	Carbo L 0.0071 ± 0.0003 Collagen variables Col (r	$\frac{n^{*} (n.u.)}{M}$ 0.0066 ± 0.0002 s IMat [*] n.u.)	CryInd L 0.0384 ± 0.0011 DHLNL (mmol/mol collagen)	\mathbb{R}^* (cm) <u>M</u> 0.0396 \pm 0.0006 HLNL (mmol/mol collagen)	PYD (mmol/mol collagen)			
C	Carbo L 0.0071 ± 0.0003 Collagen variables Col (r L	$\frac{M}{M} = \frac{M}{0.0066 \pm 0.0002}$ s = M Mat* n.u.) M	CryInd L 0.0384 ± 0.0011 DHLNL (mmol/mol collagen)	$\frac{M}{M}$ 0.0396 ± 0.0006 HLNL (mmol/mol collagen)	PYD (mmol/mol collagen)			
C	Carbo L 0.0071 ± 0.0003 Collagen variables Col (r L 4.54 ± 0.37	$\frac{M}{M}$ 0.0066 ± 0.0002 s 1Mat* h.u.) M 4.33 ± 0.29	CryInd L 0.0384 ± 0.0011 DHLNL (mmol/mol collagen) 567.8 ± 195.6	M^{*} (cm) M 0.0396 ± 0.0006 HLNL (mmol/mol collagen) 260.4± 73.4	$\begin{array}{c} {\rm PYD}\\ {\rm (mmol/mol}\\ {\rm collagen})\\ {\rm 353.1 \pm 44.5} \end{array}$			
C	Carbon L 0.0071 ± 0.0003 Collagen variables Col (r L 4.54 ± 0.37 DPD	$\frac{M}{M} = \frac{M}{0.0066 \pm 0.0002}$ s M M M 4.33 \pm 0.29 PEN	CryInd L 0.0384 ± 0.0011 DHLNL (mmol/mol collagen) 567.8 ± 195.6 Coll	M^* (cm) M 0.0396 ± 0.0006 HLNL (mmol/mol collagen) 260.4± 73.4	PYD (mmol/mol collagen) 353.1 ± 44.5			
C	Carbo L 0.0071 ± 0.0003 Collagen variables Col (r L 4.54 ± 0.37 DPD (mmol/mol	$\frac{M}{M}$ 0.0066 ± 0.0002 s IMat* h.u.) M 4.33 ± 0.29 PEN (mmol/mol	CryInd L 0.0384 ± 0.0011 DHLNL (mmol/mol collagen) 567.8 ± 195.6 Coll (mmol/mol	$\frac{M}{M} = 0.0396 \pm 0.0006$ HLNL (mmol/mol collagen) 260.4± 73.4	PYD (mmol/mol collagen) 353.1 ± 44.5			
	Carbo L 0.0071 ± 0.0003 Collagen variables Col (r L 4.54 ± 0.37 DPD (mmol/mol collagen)	$\frac{M}{M} = \frac{M}{0.0066 \pm 0.0002}$ s M M M 4.33 \pm 0.29 PEN (mmol/mol collagen)	CryInd L 0.0384 ± 0.0011 DHLNL (mmol/mol collagen) 567.8 ± 195.6 Coll (mmol/mol collagen)	M^{*} (cm) M 0.0396 ± 0.0006 HLNL (mmol/mol collagen) 260.4± 73.4	PYD (mmol/mol collagen) 353.1 ± 44.5			

173 3.2. Univariate correlation analysis

Spearman rank correlation coefficients (r) between Q_{44}^{-1} and the other variables are summarized in Table 3. For the variables displaying a significant difference between the lateral and medial quadrants, i.e., ConnD, SMI, MinOrga, MinMat, Carbon, CryInd and CollMat, r was calculated for lateral and medial group separately.

Among the microstructure variables, Ct.Po, PoN, and PoDm were positively correlated with Q_{44}^{-1} , r was

 $_{178}$ 0.68, 0.51 and 0.68, respectively. Negative correlations were found between PoS/PoV, PoSp, PoPf and Q_{44}^{-1} , r

was -0.68, -0.44 and -0.61, respectively. Among the mineral variables, DMB was significantly correlated with

 Q_{44}^{-1} (r = -0.68). Carbon from medial quadrant was significantly correlated with Q_{44}^{-1} (r = 0.40). Among the

collagen variables, Q_{44}^{-1} , was weakly correlated with DHLNL (r = 0.32) and HLNL (r = 0.28).

The variables that are not significantly correlated with Q_{44}^{-1} , as shown in Table 3, were not included in the subsequent regression analyses.

Μ	Microstructure variables							
	Ct.Po	PoS/PoV	PoN	PoDm	\mathbf{PoSp}	PoPf	$\operatorname{Conn} D$	\mathbf{SMI}
r	0.68^{3}	-0.68^3	0.51^{3}	0.68^{3}	-0.44^2	-0.61^3	n.s.	n.s.
\mathbf{M}	Mineral variables							
	DMB	MinOrga	MinMat	Carl	bon	CryInd		
				L	Μ			
r	-0.68^3	n.s.	n.s.	n.s.	0.40^{1}	n.s.		
Co	Collagen variables							
	CollMat	DHLNL	HLNL	PYD	DPD	PEN	Coll	
r	n.s.	0.32^{1}	0.28^{1}	n.s.	n.s.	n.s.	n.s.	

Table 3: Spearman rank correlation coefficient r between Q_{44}^{-1} and microstructural properties. ${}^{1}p < 0.05$, ${}^{2}p < 0.01$, ${}^{3}p < 0.001$, n.s. not significant.

184 3.3. Multivariate regression model

In the multivariate regression models, Q_{44}^{-1} is the dependent variable and the microstructure, mineral and collagen variables are the independent variables (Table 4). Overall, Ct.Po and DMB are the most significant factors contributing to the variability of Q_{44}^{-1} . As for the collagen variables, the only significant variable is DHLNL which only accounts for a minor part of the variability of Q_{44}^{-1} (Adj- r^2 is 5.7%). In the multiple regression models using microstructure variables, Ct.Po explains most of the variations of Q_{44}^{-1} (Adj- r^2 is 53.2%). Among the mineral variables, DMB is the most significant factor (Adj- r^2 is 43.2%).

The most significant variable of each group, i.e. Ct.Po, DMB and DHLNL, was then retained to derive a multiple linear regression model. The result is a model with only two variables, Ct.Po and DMB, explaining 59.1% of the variability of Q_{44}^{-1} (Table 4). The contribution of Ct.Po and DMB to Q_{44}^{-1} are illustrated in Figure 6.

Table 4: Multiple linear regression models of Q_{44}^{-1} . In the two-variable models, only Ct.Po and DMB are included. Note that the explanatory variables have been normalized . ${}^{1}p < 0.05$, ${}^{3}p < 0.0001$.

Predicted variable	Explanatory variables	Linear model	$\begin{array}{c} \text{Adj-}r^2 \\ (\%) \end{array}$	RMSE
$Q_{44}^{-1} \ Q_{44}^{-1} \ Q_{-1}^{-1}$	microstructure mineral	$\begin{array}{l} 0.0396 + 0.0055 \times \overline{\text{Ct.Po}} \\ 0.0379 - 0.0051 \times \overline{\text{DMB}} \\ 0.0377 + 0.0016 \times \overline{\text{DHLNL}} \end{array}$	53.2^3 43.2^3	0.0021 0.0023
$Q_{44} Q_{44}^{-1}$	Ct.Po + DMB	$0.0377 + 0.0016 \times \overline{\text{DHLNL}}$ $0.0392 + 0.0039 \times \overline{\text{Ct.Po}} - 0.0025 \times \overline{\text{DMB}}$	5.7^{-5} 59.1^{3}	0.0030 0.0020



Figure 6: Q_{44}^{-1} as a function of Ct.Po (left) and DMB (right)

¹⁹⁵ 4. Discussion

¹⁹⁶ We measured with RUS the shear damping coefficient Q_{44}^{-1} , equivalent to a torsion loss tangent (usually ¹⁹⁷ denoted tan δ), in 52 specimens of human cortical bone from 26 elderly donors. We then investigated the ¹⁹⁸ relationships between Q_{44}^{-1} and some compositional and microstructural characteristics measured with FTIRM, ¹⁹⁹ biochemical analysis, and SR- μ CT.

The shear damping values in the present study (0.0371 ± 0.0031) fall in the range of values usually reported 200 [18]. The variations of Q_{44}^{-1} were essentially determined by the variations of vascular porosity and mineral 201 content: a multiple linear regression model with these two variables explained 59.1% of the variability of Q_{44}^{-1} . 202 Damping increased with specimen's porosity and decreased with mineral content. Adding collagen variables 203 did not improve this model. These relationships between damping, vascular porosity and mineral content 204 have not been reported before as far as we know. These results are consistent with the finding previously 205 reported [23] that Q_{44} (also measured with RUS) is correlated to mass density ($r^2 = 0.72$); indeed, density 206 increases as porosity decreases and mineral content increases. Interestingly, this behavior for Q_{44}^{-1} is similar 207 to that observed for stiffness coefficients [25] which decrease with porosity and increase with mineral but are 208 also weakly dependent on collagen variables. 209

Viscoelasticity in bone may arise from a variety of mechanisms, including fluid motion inside pores, ther-210 moelastic coupling, motions at interfaces such as the cement line and between lamellae of mineralized collagen, 211 and molecular deformation of collagen [3]. The relative importance of these mechanisms depend on the time 212 scale of the experiment (or excitation frequency in a dynamic experiment). Garner et al. [18] have investigated 213 the shear damping with an excitation frequency between 10^{-2} and 10^{5} Hz, showing a minimum of damping 214 around 10 Hz and a range of $\tan \delta$ of approximately 0.01-0.08. With RUS, the measurement frequency cor-215 responds to the natural resonance of the specimen; in this study, this frequency varied in a narrow range 216 centered at 146.1 (\pm 8.4 kHz). Measurement of damping around 150 kHz have seldom been reported as most 217 of the viscoelastic data was obtained with dynamic mechanical analyzers (DMA) below 20 Hz or creep tests. 218 Accordingly, our results can only be compared with that of others with caution. 219

The relative importance of damping mechanisms also depends on the length scale of the measurement. 220 Shepherd et al. [32] reported concurrent measurement of shear damping with a creep test on dogbone-shape 221 specimens (several millimeters) and nanoindentation measurement of creep and found millimeter scale damping 222 (relaxation time) about an order of magnitude larger than microscopic damping; furthermore the damping 223 values at the two scales did not correlate. Accordingly, it is hypothesized that damping mechanisms, not 224 captured by nanoindentation, dominate at the scale of a few millimeters: e.g., viscous damping related to 225 fluid flow in pores, motion at mesoscale interfaces in the Haversian microstructure, or thermoelastic coupling 226 at the mesoscale in the heterogeneous mineralized matrix. The correlation we found in the present study 227 between porosity and damping is consistent with this hypothesis: with increased porosity the heterogeneity of 228 the microstructure increases (leading to inhomogeneous thermoelastic damping) as well as the contact surface 229 between fluid in pores and bone matrix (viscous damping due to fluid flow). 230

We built a multilinear model suggesting that when porosity is controlled, the variation of mineral content 231 explains a small additional part of the variability of damping (Adj- r^2 of 59.1 vs. 53.2). Inter-specimen variation 232 of mineral content may reflect different degree of homogeneity of the bone matrix (e.g., proportions of osteonal 233 vs. interstitial tissue, age of osteons) which could affect damping through inhomogeneous thermoelastic effect 234 and affect the viscous loss in nanoscale motion within the mineralized collagen fibrils [3]. At the scale of the 235 mineralized collagen molecule, it was evidenced with molecular dynamics simulations that the mineral content 236 contributes to the attenuation of stress waves [33]. The mineral characteristics other than DMB were weakly or 237 non-significantly correlated to damping. Using nanoindentation and assessing mineral properties with FTIRM, 238 Ojanen et al. [17] also found that mineral variables, except crystallinity, were not correlated to creep viscosity. 239

The role of matrix proteins in damping is well established [34, 35]. However, authors who have investigated the relationships between collagen and viscoelastic properties of native specimens (not chemically altered) reported weak or non significant correlations [16, 17]. In line with these works, we found that collagen variables, including cross-links properties were weakly or non significantly correlated to Q_{44}^{-1} . One possible explanation is that the variations of collagen properties in the population of donors considered are too small and that at the macroscale, the variations of damping due to changes in microstructure are dominant.

Our data fills a gap of knowledge as we have measured cortical bone shear damping at frequencies around 246 150 kHz which have seldom been considered [18]. This frequency regime is not accessible with most widespread 247 measurement techniques: it is intermediate between the lower frequencies typically accessible with commercial 248 DMA devices to measure the phase shift δ between a forced excitation and the oscillatory response, and 249 higher frequencies (MHz range) of conventional ultrasonic transducers to measure wave attenuation. The 250 measurement frequency region considered is a priori not relevant for the study of the physiological behavior of 251 bone as physiological loading hardly contains frequencies above 100 Hz. However, some engineering applications 252 could benefit from a better quantification of damping around 150 kHz. Low intensity pulsed ultrasound 253 (LIPUS) devices used to stimulate bone healing work in a frequency range between 45 kHz and 3 MHz [36]. 254 Ultrasonic bone drilling in orthopedic surgery uses frequencies in the range 20-50kHz [37, 38]. Finally, some 255 devices for the assessment of cortical bone properties for the monitoring of bone health with guided waves use 256

²⁵⁷ frequencies around 100 kHz [39, 40].

It has been suggested that viscoelasticity could serve as a biomarker of skeletal fragility and bone disease 258 [13, 14, 16]. Most often, this idea is related to the assumption that interindividual variations of viscoelasticity 259 reflect the variability in the quality of the bone matrix which should prevent crack propagation. Our results 260 suggest that collagen variations only have a minor effect at the mesoscale (i.e., the scale of a few millimeters), 261 if any, on damping variability. In contrast, we found that damping is correlated to porosity, which is a well 262 documented risk factor for fragility fracture [41] and which is related to bone strength ex vivo [42]. This means 263 that damping, as it reflects porosity, is related to bone fragility, supporting the consideration of viscoelasticity 264 measurement as a potential biomarker of fragility. 265

This study has some limitations. The accuracy of shear damping measurement is limited by the fact that 266 it was evaluated from the measurement of the quality factor of a resonance peak by using an approximate 267 formulae (Eq. 5). We evaluated that only about 90% of the damping value was correctly captured. The 268 resonance peak of the specimens ranged from 115.4 to 160.3 kHz depending on the specimens dimensions, 269 stiffness, and mass. We have disregarded a possible effect of frequency on shear damping independent of the 270 effect of the microstructural and compositional variables. We believe this is reasonable as we did not find 271 any correlation between the frequency of the peak and damping. Besides, bone specimens were collected at a 272 single skeletal site (lateral and medial quadrants of the femoral diaphysis) of bones from elderly donors without 273 documentation on the existence of bone pathologies. Therefore, the findings in this work are limited to the 274 femoral mid-diaphysis of an aged group of donors. Further studies are warranted to investigate whether these 275 conclusions can be extended to other skeletal sites of bone. Finally, studies with bone material representative 276 of that of patients (e.g., osteopenic and osteoporotic patients) should be conducted in order to assess the extent 277 to which viscoelasticity could reflect fragility for specific bone diseases. 278

279 Declaration of interests

The authors declare that they have no known competing financial interests or personal relationships that could have appeared to influence the work reported in this paper.

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Appendix A. Effect of the selected bandwidth for signal analysis on the peak frequency and quality factor.

The quality factor of the first peak Q_1 in the RUS spectrum (Fig. 4) was determined by fitting the Lorentzian 294 model, as explained in Methods, to a portion (bandwidth) of the spectrum containing one (Case 1) or two 295 (Case 2) peaks. To test the effect of the choice of bandwidth we also fitted the spectrum with a bandwidth 296 limited to the first peak for the 40 specimens initially in Case 2 (first two peaks relative close). Results are 297 given in Table A.5. There was no significant difference for Q_1 and f_1 tested by Wilcoxon test. There was a 298 slightly larger standard deviation of Q_1 calculated from the several repetitions of the measurements for each 299 specimen due to the uptrend of the low frequency part of the second peak that was also taken into account. 300 Overall, these results indicate that the bandwidth selection method has a negligible effect on the values of f_1 301 and Q_1 in this study. 302

Table A.5: Comparison of the results (mean \pm SD) of f_1 and Q_1 obtained by fitting the first peak or alternatively the first two peaks. The last line summarizes the standard deviation (mean (SD)) of Q_1 calculated from the several repetitions of the measurements for each specimen.

f_1 — Two peaks	\mathbf{f}_1 — One peak
145.9 ± 6.5	145.9 ± 6.5
Q_1 — Two peaks	Q_1 — One peak
29.98 ± 1.81	30.04 ± 1.73
SD of Q_1 — Two peaks	SD of Q_1 — One peak
1.04 ± 0.70	1.18 ± 1.01

³⁰³ Appendix B. Effect of defatting and X-ray radiation on viscoelastic damping

In order to clarify whether Q_{44}^{-1} would be affected by defatting and irradiation, RUS measurements were conducted three times on each specimen from a subset of 24 specimens: i) on the native specimen right after preparation, ii) after being defatted for 18 h in a chemical bath of diethyl ether and methanol (1:1), iii) after SR- μ CT imaging which delivers a moderate radiation dose of 2.5 kGy. More details concerning defatting and imaging protocols were given in [25]. Table B.6 summarizes the Q_{44}^{-1} values determined from the three measurements.

Table B.6: A summary of the results (Mean \pm SD) of Q_{44}^{-1} in native, defatted and irradiated specimens.

Q_{44}^{-1} — Native	\mathbf{Q}_{44}^{-1} — Defatted	\mathbf{Q}_{44}^{-1} — Irradiated
0.0375 ± 0.0034	0.0385 ± 0.0031	0.0378 ± 0.0028

Multiple comparison of means did not show significant difference between the different states. There was no bias and the mean and standard deviation of differences appeared to be constant throughout the range of Q_{44}^{-1} values. This analysis complements the analysis, conducted on the same specimens, of the effect of defatting and irradiation on elastic properties which was reported in [26].

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