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Electronic Supporting Information

Intercalation-exfoliation processes during ionic exchange reaction from the sodium lepidocrocite-type titanate toward the proton-based trititanate structure

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Fig. S1. Structural representation of a) lepidocrocite and b) trititanate structure.

Fig. S2. ATR FT-IR spectrum of Na-titanate.

Fig. S3. Thermogravimetric analysis curve of Na-titanate.

Fig. S4. PDF refinements of Na-titanate using different structural models.

Fig. S5. Nyquist plots of conductivity at RT.

- a)** Low-frequency contribution (double layer capacitance) C1 fitted with a circular arc.
b) Grain bulk contribution fitted with a straight line obtained after the subtraction of C1.
 $\sigma_a = 6.4 \cdot 10^{-4}$ S/m and $\sigma_b = 1.5 \cdot 10^{-3}$ S/m are sample and grain conductivities, respectively.

Fig. S6. TEM images of Na-titanate (a, c) and H-titanate (b, d).

Fig. S7. SEM images of Na-titanate (a) and H-titanate (b)

Fig. S8. Atomic force micrograph with a section of an aggregate of elongated particles of H-titanate. Horizontal and vertical distances are between black, green, and red arrows. The longest sizes of two elongated particles are given in the picture.

Fig. S9. Real-space refinements of the PDFs of the samples collected at different pH values.

Fig. S10. Real-space refinements of the PDF of the sample collected at pH 4 using lepidocrocite (a), trititanate (b), and both models (c).

Fig. S11. Experimental (blue) and fitted using the Gaussian Isotropic Model¹ (red dashed line) ²³Na MAS (44 kHz) NMR spectra of three samples collected at different pH values. This model involves a Gaussian distribution of chemical shifts and a distribution of quadrupolar interactions. The parameters used for these fits are reported in **Table S1**.

Table S1. Average isotropic chemical shift ($\langle\delta_{iso}\rangle$ /ppm), full width at half maximum (FWHM) of the isotropic chemical shift Gaussian distribution (ΔCS /ppm), average quadrupolar frequency ($\langle\nu_Q\rangle$ /kHz) and quadrupolar constant ($\langle C_Q\rangle$ /MHz) used to fit the ²³Na MAS NMR spectra of three samples collected at different pH values (see **Fig. S8**).

pH	$\langle\delta_{iso}\rangle$	ΔCS	$\langle\nu_Q\rangle$	$\langle C_Q\rangle$
9	-12.6	9.1	515	1.03
7	-11.3	7.0	668	1.34
4	-11.7	5.8	631	1.26

¹ D. Massiot, F. Fayon, M. Capron, I. King, S. Le Calvé, B. Alonso, J.-O. Durand, B. Bujoli, Z. Gan and G. Hoatson, Modelling one- and two-dimensional solid-state NMR spectra: Modelling 1D and 2D solid-state NMR spectra, *Magn. Reson. Chem.*, 2002, **40**, 70–76.

Fig. S12. Na/Ti ratio quantified using energy dispersive X-ray spectroscopy for the samples collected at different pH.

Fig. S13. Intensity / Intensity (pH = 9) ratio of the ^{23}Na non-normalized NMR spectra for the samples collected at different pH. Intensity estimated as the area under the curve of the fit of each non-normalized spectrum. The ^{23}Na non-normalized spectra correspond, for each sample, to the same number of scans and the same sample mass in the rotor.

Fig. S14. Experimental (blue) and fitted (red dashed line) ^1H MAS (60 kHz) NMR spectra of the samples collected at different pH. The individual resonances used for these fits are shown below (see **Table S2**).

Table S2. Isotropic chemical shifts (δ_{iso} /ppm), line widths (LW/ppm), relative intensities (I/%), of the NMR resonances used for the fit of the ^1H MAS (64 kHz) NMR spectra of the samples collected at different pH (see **Fig. S11**).

pH	δ_{iso}	LW	I
1	0.85	0.13	0.4
	1.26	0.31	1.3
	6.3	2.3	84.9
	8.1	5.7	13.4
2	0.92	0.14	0.6
	1.35	0.36	1.3
	5.9	1.6	60.3
	7.2	7.3	37.5
	11.4	1.5	0.3
4	0.85	0.14	0.5
	1.25	0.34	1.0
	5.2	1.5	74.6
	6.5	9.1	23.9
7	0.84	0.12	0.7
	1.24	0.28	1.4
	5.0	1.5	73.5
	5.3	5.3	24.4
9	0.86	0.13	0.6
	1.28	0.20	1.0
	1.8	0.09	0.3
	5.1	1.6	74.5
	6.0	5.4	23.6

Fig. S15. Results of to fit the decay of the ionic conductivity during the $\text{Na}^+ \text{-H}^+$ ionic exchange.