



**HAL**  
open science

## **Impact of Early Versus Late Antiretroviral Treatment Initiation on Naive T Lymphocytes in HIV-1-Infected Children and Adolescents – The-ANRS-EP59-CLEAC Study**

Pierre Frange, Thomas Montange, Jérôme Le Chenadec, Damien Batalie, Ingrid Fert, Catherine Dollfus, Albert Faye, Stéphane Blanche, Anne Chacé, Corine Fourcade, et al.

► **To cite this version:**

Pierre Frange, Thomas Montange, Jérôme Le Chenadec, Damien Batalie, Ingrid Fert, et al.. Impact of Early Versus Late Antiretroviral Treatment Initiation on Naive T Lymphocytes in HIV-1-Infected Children and Adolescents – The-ANRS-EP59-CLEAC Study. *Frontiers in Immunology*, 2021, 12, pp.662894. 10.3389/fimmu.2021.662894 . hal-03230769

**HAL Id: hal-03230769**

**<https://hal.sorbonne-universite.fr/hal-03230769>**

Submitted on 20 May 2021

**HAL** is a multi-disciplinary open access archive for the deposit and dissemination of scientific research documents, whether they are published or not. The documents may come from teaching and research institutions in France or abroad, or from public or private research centers.

L'archive ouverte pluridisciplinaire **HAL**, est destinée au dépôt et à la diffusion de documents scientifiques de niveau recherche, publiés ou non, émanant des établissements d'enseignement et de recherche français ou étrangers, des laboratoires publics ou privés.



# Impact of Early Versus Late Antiretroviral Treatment Initiation on Naive T Lymphocytes in HIV-1-Infected Children and Adolescents – The-ANRS-EP59-CLEAC Study

Pierre Frange<sup>1,2,3</sup>, Thomas Montange<sup>4,5</sup>, Jérôme Le Chenadec<sup>6</sup>, Damien Batalie<sup>4,5</sup>, Ingrid Fert<sup>4,5</sup>, Catherine Dollfus<sup>7</sup>, Albert Faye<sup>8</sup>, Stéphane Blanche<sup>1</sup>, Anne Chacé<sup>9</sup>, Corine Fourcade<sup>10</sup>, Isabelle Hau<sup>11</sup>, Martine Levine<sup>12</sup>, Nizar Mahlaoui<sup>1</sup>, Valérie Marcou<sup>13</sup>, Marie-Dominique Tabone<sup>7</sup>, Florence Veber<sup>1</sup>, Alexandre Hocin<sup>6</sup>, Thierry Wack<sup>6</sup>, Véronique Avettand-Fenoël<sup>2,14</sup>, Josiane Warszawski<sup>6,15</sup>, and Florence Buseyne<sup>4,5\*</sup>, on behalf of the ANRS-EP59-Study Group

## OPEN ACCESS

### Edited by:

Francesca Chiodi,  
Karolinska Institutet (KI), Sweden

### Reviewed by:

Daria Trabattoni,  
University of Milan, Italy  
Martyn Andrew French,  
University of Western Australia,  
Australia

### \*Correspondence:

Florence Buseyne  
florence.buseyne@pasteur.fr

### Specialty section:

This article was submitted to  
Viral Immunology,  
a section of the journal  
Frontiers in Immunology

Received: 01 February 2021

Accepted: 24 March 2021

Published: 22 April 2021

### Citation:

Frange P, Montange T,  
Le Chenadec J, Batalie D, Fert I,  
Dollfus C, Faye A, Blanche S, Chacé A,  
Fourcade C, Hau I, Levine M,  
Mahlaoui N, Marcou V, Tabone M-D,  
Veber F, Hocin A, Wack T,  
Avettand-Fenoël V, Warszawski J and  
Buseyne F (2021) Impact of Early  
Versus Late Antiretroviral  
Treatment Initiation on Naive  
T Lymphocytes in HIV-1-Infected  
Children and Adolescents –  
The-ANRS-EP59-CLEAC Study.  
Front. Immunol. 12:662894.  
doi: 10.3389/fimmu.2021.662894

<sup>1</sup> Immunologie, hématologie et rhumatologie pédiatrique, hôpital Necker-Enfants malades, AP-HP- Centre – Université de Paris, Paris, France, <sup>2</sup> Laboratoire de microbiologie clinique, hôpital Necker-Enfants malades, AP-HP-Centre – Université de Paris, Paris, France, <sup>3</sup> EHU 7328 PACT, Institut Imagine, Université de Paris, Paris, France, <sup>4</sup> Unité Epidémiologie et Physiopathologie des Virus Oncogènes, Institut Pasteur, Paris, France, <sup>5</sup> Département de Virologie, UMR CNRS 3569 Institut Pasteur, Paris, France, <sup>6</sup> Département d'épidémiologie, Centre de Recherche en Epidémiologie et Santé des Populations, INSERM U1018, Le Kremlin-Bicêtre, Villejuif, France, <sup>7</sup> Hémato-oncologie pédiatrique, Hôpital Trousseau, AP-HP, Paris, France, <sup>8</sup> Pédiatrie Générale, Hôpital Robert Debré, AP-HP, Paris, France, <sup>9</sup> Pédiatrie et néonatalogie, Centre hospitalier intercommunal de Villeeneuve-Saint-Georges, Villeeneuve-Saint-Georges, France, <sup>10</sup> Pédiatrie Générale, Hôpital Bicêtre, AP-HP, Paris, France, <sup>11</sup> Pédiatrie Générale, Centre hospitalier intercommunal de Créteil, Créteil, France, <sup>12</sup> Immuno-hématologie pédiatrique, Hôpital Robert Debré, AP-HP, Paris, France, <sup>13</sup> Médecine et réanimation néonatale, Hôpital Cochin, AP-HP-Centre – Université de Paris, Paris, France, <sup>14</sup> CNRS 8104/INSERM U1016, Institut Cochin, Université Paris Descartes, Paris, France, <sup>15</sup> INED, Université Paris Sud, Le Kremlin-Bicêtre, Orsay, France

**Background:** The early initiation of antiretroviral therapy (ART) in HIV-1-infected infants reduces mortality and prevents early CD4 T-cell loss. However, the impact of early ART on the immune system has not been thoroughly investigated in children over five years of age or adolescents. Here, we describe the levels of naive CD4 and CD8 T lymphocytes (CD4/CD8T<sub>N</sub>), reflecting the quality of immune reconstitution, as a function of the timing of ART initiation (early (<6 months) versus late (≥24 months of age)).

**Methods:** The ANRS-EP59-CLEAC study enrolled 27 children (5-12 years of age) and nine adolescents (13-17 years of age) in the early-treatment group, and 19 children (L-Ch) and 21 adolescents (L-Ado) in the late-treatment group. T lymphocytes were analyzed by flow cytometry and plasma markers were analyzed by ELISA. Linear regression analysis was performed with univariate and multivariate models.

**Results:** At the time of evaluation, all patients were on ART and had a good immunovirological status: 83% had HIV RNA loads below 50 copies/mL and the median CD4 T-cell count was 856 cells/μL (interquartile range: 685-1236 cells/μL). In children, early ART was associated with higher CD8T<sub>N</sub> percentages (medians: 48.7% vs. 31.0%, *P* = 0.001), and a marginally higher CD4T<sub>N</sub> (61.2% vs. 53.1%, *P* = 0.33). In adolescents, early ART was associated with low CD4T<sub>N</sub> percentages and less

differentiated memory CD8 T cells. CD4T<sub>N</sub> and CD8T<sub>N</sub> levels were inversely related to cellular activation and gut permeability.

**Conclusion:** In children and adolescents, the benefits of early ART for CD8T<sub>N</sub> were clear after long-term ART. The impact of early ART on CD4T<sub>N</sub> appears to be modest, because pediatric patients treated late respond to HIV-driven CD4 T-lymphocyte loss by the *de novo* production of T<sub>N</sub> cells in the thymus. Our data also suggest that current immune activation and/or gut permeability has a negative impact on T<sub>N</sub> levels.

**Clinical Trial Registration:** ClinicalTrials.gov, identifier NCT02674867.

**Keywords:** HIV-1, children, adolescents, early ART, T lymphocyte, naive T lymphocyte

## INTRODUCTION

HIV-1-infected children receiving antiretroviral treatment (ART) face a lifetime of chronic disease due to this virus (1–3). Early ART initiation has an immediate beneficial impact, by blocking clinical progression (4), decreasing the cell-associated viral reservoir in infants (5) and favoring faster CD4 T-lymphocyte recovery (1, 6, 7). However, the benefit of early ART for CD4 T-cell levels may decrease after longer periods on ART. Early ART initiation may preserve naive CD8 T lymphocytes (CD8T<sub>N</sub>) (8) and CD4/CD8 ratio (9). Early ART reduces the frequency of HIV-specific CD4 and CD8 T cells in children, but preserves the capacity of these cells to produce several cytokines (10–12). Overall, findings concerning the impact of early ART on CD4 and CD8 T cells in pediatric patients remain limited.

Several gaps in our knowledge of the long-term impact of early versus late ART remain. Few studies have investigated the immunological benefits of early ART initiation in children over the age of five years and those without sustained viral suppression. The benefit of early ART for CD4 T-cell levels may persist or decrease over longer periods of long-term effective ART. Finally, unplanned treatment interruptions frequently occur during early childhood and adolescence, and it remains unclear whether the deleterious impact of viral rebound on the benefits of early ART is transient or irreversible (13–15).

The ANRS-EP59-CLEAC study investigated the immunological and virological characteristics of HIV-1-infected children over five years of age and adolescents, as a function of age at ART initiation (< 6 months vs. ≥ 24 months of age). We included participants with an initial period of viral suppression but without any criteria on the persistence of viral control (16). Here, we focused on the proportions of CD4T<sub>N</sub> and CD8T<sub>N</sub>, the major indicators of qualitative immune reconstitution (17), and their relationships with current and past HIV disease parameters and current immune activation (3).

## PATIENTS AND METHODS

### Patients

The ANRS-EP59-CLEAC study was conducted in accordance with the Helsinki Declaration and the protocol was approved by

the “Comité de protection des personnes île-de-France V”. Agreement to participate was obtained from the participants, if they were old enough to give an opinion, and written informed consent was obtained from at least one parent. The main inclusion criteria were (1) HIV-1 infection after vertical transmission, (2) participant aged 5 to 17 years at the time of the study, (3) ART initiation for therapeutic purposes before six months of age or after two years of age, (4) with initial virological success (HIV-1 RNA < 400 copies/mL achieved within 24 months of treatment initiation), regardless of the subsequent course of viremia. Between 2016 and 2019, two 15 mL blood samples were collected for biological evaluations, at two consecutive routine clinical follow-up visits.

### Biological Evaluations

Total blood cell-associated HIV-1 DNA was quantified by ultrasensitive real-time PCR (18). The CD4 and CD8 T-lymphocyte subsets were quantified in fresh blood by flow cytometry with combinations of antibodies targeting the CD3, CD4, CD8 $\alpha$ , CD45RA, CCR7, CD27, CD28, CD31, CD95, HLA-DR, and CD38 molecules (**Supplementary Material**). Data were collected on a Gallios cytometer (Beckman Coulter) and analyzed with Kaluza software (Beckman Coulter). The gating strategy involved the sequential definition of lymphocytes, CD4 and CD8 T lymphocytes, and their subsets according to CD45RA and CCR7 expression. The subsets were further characterized by using CD27 and CD28 expression to define naive (T<sub>N</sub>), central memory (T<sub>CM</sub>), transitional memory (T<sub>TM</sub>), effector memory (T<sub>EM</sub>) and effector (T<sub>E</sub>) cells; CD31 and CD95 expression was used to define recent thymic emigrants (T<sub>RTE</sub>) and stem cell memory cells (T<sub>SCM</sub>); and HLA-DR and CD38 expression was used to define activated T cells (**Supplementary Material**).

C-reactive protein (CRP), interleukin-6 (IL-6), CXCL10, soluble CD14 (sCD14), soluble CD163 (sCD163), and intestinal fatty acid-binding protein (iFABP) were quantified by ELISA (**Supplementary Table 1**). The Liaison XL CMV IgG (Diasorin) chemiluminescence immunoassay and the CMV R-Gene (Biomérieux) PCR assay were used to quantify cytomegalovirus (CMV) antibodies and DNA in plasma samples. The frequency of interferon (IFN)- $\gamma$ -producing cells was quantified in an Elispot assay with anti-IFN- $\gamma$  capture/detection antibodies (Diaclone 869.060.010), streptavidin

alkaline phosphatase conjugate (GE Healthcare RPN4402) and BCIP/NBT color development substrate (S3771, Promega), according to the kit manufacturers' instructions. Fresh peripheral blood mononuclear cells (PBMCs) were used to seed High Protein-Binding Immobilon-P Membrane P96 plates (MSIPS4510, Merk Millipore) at a density of  $2 \times 10^5$  and  $5 \times 10^4$  cells/well in RPMI supplemented with 10% fetal calf serum, with the addition of pp65 CMV peptides (#11549, NIH AIDS Reagent Program, 1  $\mu\text{g}/\text{mL}$  of each peptide) or peptide diluent (DMSO). The cells were then cultured for 24 h. Spot-forming cells (SFCs) were counted with an ImmunoSpot S6 UV Image Analyzer (CTL, Bonn, Germany).

## HIV History Variables

The patients were either included at birth in The Agence Nationale de Recherche sur le SIDA et les Hépatites Virales (ANRS) EPF/ANRS CO10 national prospective multicenter cohort or were managed at the centers of the ANRS/CO10 cohort and diagnosed before the age of 13 years (1). Clinical and biological data were therefore collected for HIV-infected children *via* the completion of standardized questionnaires at six-month intervals, or were collected retrospectively after inclusion in the CLEAC study. All ART regimens initiated consisted of highly active antiretroviral treatment (HAART; i.e., any combination of at least three different antiretroviral drugs or any combination including one protease inhibitor, or one non-nucleoside reverse transcriptase inhibitor or one integrase inhibitor). During the first few weeks of life, 30 children received prophylactic ART consisting of zidovudine ( $n=21$ ), zidovudine+lamivudine ( $n=5$ ), zidovudine+lamivudine+lopinavir ( $n=2$ ) or zidovudine+lamivudine+nevirapine ( $n=2$ ). For the last four of these children, the date of first HAART (ART1) initiation was the date of prophylactic ART initiation, as treatment was continued without interruption after diagnosis. At the time of the study, all but one of the children were on HAART, the remaining child being on two nucleoside reverse transcriptase inhibitors and having an undetectable viral load.

CD4 and CD8 T cells and plasma HIV RNA levels were quantified at each clinical site. The threshold values used in HIV RNA assays depended on study site, and a value of 50 copies/mL was selected as the common cutoff for current HIV RNA detection. A cutoff of 400 copies/mL was used for the assessment of virological history, because of changes in HIV RNA quantification assays and cutoffs over time. Cumulative viremia was defined as the area under the HIV RNA curve over time (19). Past immunological parameters were defined using CD4 T-cell percentages, because the age-related variations of this parameter are smaller than those for CD4 T-cell counts. The duration of viral suppression and cumulative viremia were assessed from ART1 initiation. These parameters were normalized by dividing their values by the time since ART1 initiation.

## Statistical Analysis

Analyses were stratified according to age at evaluation (children: 5-12 years of age; adolescents: 13-17 years of age). Wilcoxon-Mann-Whitney, Kruskal-Wallis and the Fisher's exact tests were used to compare the characteristics of children and adolescents

between the early and late treatment initiation groups. The *corrplot* package of R was used to display Spearman's rank correlation coefficients. Univariate and multivariate analyses were performed by linear regression, with CD4/CD8T<sub>N</sub> percentages as the continuous dependent variables. Multivariate models included early ART as the main independent variable, together with noncollinear variables with  $P$  values  $<.2$  in univariate analysis. Most models were built with a single biological variable, to prevent over-adjustment. Observations with high leverage values or studentized residuals were excluded to test the robustness of the models. Analyses were conducted with SAS statistical software.  $P$  values  $<.05$  were considered statistically significant.

## RESULTS

### Patient Characteristics

We prospectively enrolled 27 children (E-Ch) and nine adolescents (E-Ado) in the early ART (E) group, and 19 children (L-Ch) and 21 adolescents (L-Ado) in the late ART (L) group (**Table 1**). Just over half (54%) of the patients were girls. Most patients (74%) were born to mothers originating from Sub-Saharan Africa. However, 60% of patients were born in mainland France, and ART was initiated early more frequently in these children than in those born abroad. At the time of the study, children were more frequently aviremic (89% vs. 73%,  $P = .07$ ), and had higher CD4 T-cell counts than adolescents (median ([IQR]: 954 [745;1320] vs. 766 [622;1092] cells/ $\mu\text{L}$ ,  $P = .04$ ). Immunovirological status was good in most patients at the time of the study.

### Early ART Was Not Significantly Associated With Higher CD4T<sub>N</sub> Percentages in Children, and Was Associated With Lower CD4T<sub>N</sub> Percentages In Adolescents

Blood CD4T<sub>N</sub> percentages were not significantly higher in E-Ch than in L-Ch (medians: 61.2% vs. 53.1%,  $P = .33$ ). E-Ado had significantly lower CD4T<sub>N</sub> than L-Ado (36.0% vs. 51.5%,  $P = 0.02$ ). The interaction between age and treatment group was significant ( $P = .008$ ) and we performed analyses separately for children and adolescents.

High interindividual variation in the distribution of CD4 T-cell subsets was observed in all groups (**Figures 1A, B**). No significant differences were observed between the E-Ch and L-Ch groups (**Figure 1A**). E-Ado had significantly lower levels of recent thymic emigrants (T<sub>RE</sub>, 27.2% vs. 41.4%,  $P = .03$ ) and CD31<sup>neg</sup>T<sub>N</sub> (4.0% vs. 6.5%,  $P = .01$ ), and higher levels of transitional memory (T<sub>TM</sub>) cells (26.6% vs. 15.7%,  $P = .04$ ) than L-Ado. CD4 T-cell counts were positively correlated with CD4T<sub>N</sub> and CD4T<sub>RE</sub>, and negatively correlated with the levels of most memory subsets (**Figure 1C**).

We then searched for factors related to CD4T<sub>N</sub> levels, including demographic factors, immunovirological status at the time of the study and during the time since ART1 initiation.



**TABLE 1 |** Patient's characteristics.

Characteristics <sup>a</sup>	Children (5-12 years)		Adolescents (13-17 years)		All n = 76	P value <sup>a</sup>
	Early treatment n = 27	Late treatment n = 19	Early treatment n = 9	Late treatment n = 21		
Age, years	9 [6;11]	8 [7;10]	15 [14;16]	14 [13;15]	11 [8;14]	.0001
Sex						
Male	33.3 (9)	42.1 (8)	44.4 (4)	66.7 (14)	46.0 (35)	.14
Female	66.7 (18)	57.9 (11)	55.6 (5)	33.3 (7)	54.0 (41)	
Sub-Saharan African origin						
No	25.9 (7)	26.3 (5)	22.2 (2)	23.8 (5)	25.0 (19)	1.00
Yes	74.1 (20)	73.7 (14)	77.8 (7)	76.2 (16)	75.0 (57)	
Born in mainland France						
No	7.4 (2)	73.7 (14)	0.0 (0)	71.4 (15)	40.8 (31)	<.0001
Yes	92.6 (25)	26.3 (5)	100.0 (9)	28.6 (6)	59.2 (45)	
Age at ART1 initiation (months)	2.1 [0.5;3.4]	54.3 [49.9;81.4]	2.3 [1.7;2.7]	93.0 [55.4;137.3]	25.2 [2.3;79.0]	.0001
Time since ART1 initiation (months)	119 [75;137]	52 [23;70]	188 [178;198]	85 [40;124]	93 [57;137]	.0001
Current HIV RNA						
<50 copies/ml	88.9 (24)	89.5 (17)	66.7 (6)	76.2 (16)	82.9 (63)	.30
≥50 copies/ml	11.1 (3)	10.5 (2)	33.3 (3)	23.8 (5)	17.1 (13)	
CD4 count, cells/μL	951 [725;1320]	1009 [745;1527]	840 [713;1078]	740 [622;1092]	856 [685;1236]	.20
CD4 percentage	39 [37;44]	37 [33;41]	39 [34;44]	36 [29;41]	38 [32.5;42]	.18
CD8 count, cells/μL	574 [478;876]	939 [724;1035]	675 [539;767]	672 [510;1016]	714 [519.5;985]	.02
CD8 percentage	26 [22;31]	31 [27;36]	30 [25;43]	31 [26;42]	29 [25;36]	.07
CD4/CD8 ratio	1.57 [1.10;1.90]	1.26 [0.89;1.48]	1.34 [0.92;1.80]	1.06 [0.76;1.67]	1.34 [0.92;1.76]	.10
Lymphocyte count, 10 <sup>3</sup> cells/μL	2.4 [2.0;2.9]	2.8 [2.5;3.6]	1.8 [1.4;2.3]	2.0 [1.7;2.6]	2.4 [1.8;2.8]	.003
CD4 <sub>N</sub> %	61.2 [49.9;68.9]	53.1 [50.0;58.5]	36.0 [33.2;48.9]	51.5 [43.1;59.1]	54.0 [46.4;62.5]	.01
CD8 <sub>N</sub> %	48.7 [35.9;55.8]	31.0 [21.9;36.7]	29.0 [15.9;32.0]	29.0 [20.2;37.6]	35.1 [24.1;45.9]	.0002

<sup>a</sup>For categorical variables, frequencies, counts and Fisher's exact test P values are indicated; for quantitative variables, median, interquartile range and Kruskal-Wallis' test P values are indicated.

ART1, first highly active antiretroviral therapy; CD4<sub>N</sub> %, naive CD4 T lymphocyte percentage; CD8<sub>N</sub> %, naive CD8 T lymphocyte percentage.

Biomarkers of immune activation (activated CD4/CD8<sub>T<sub>M</sub></sub>, sCD14, sCD163, and CXCL10), inflammation (CRP and IL-6) and gut permeability (iFABP) (**Figure 2** and **Supplementary Table 2**) were assessed in the samples used for T<sub>N</sub> quantification.

In children, higher CD4<sub>T<sub>N</sub></sub> percentages were associated with higher CD4 T-cell counts and lower activated CD4<sub>T<sub>M</sub></sub> percentages (**Figure 2**). Sex and geographic origin were not significantly associated with CD4<sub>T<sub>N</sub></sub> percentages, but children of sub-Saharan origin tended to have lower CD4<sub>T<sub>N</sub></sub> percentages than other children (56.5% vs 62.5%,  $P = 0.08$ , **Figure 2** and **Supplementary Table 2**). After adjustment for treatment group, geographic origin and age, CD4<sub>T<sub>N</sub></sub> percentage was not associated with CD4 T-cell counts (model 1), but was inversely correlated with activated CD4<sub>T<sub>M</sub></sub> percentage (model 2).

In adolescents, higher CD4<sub>T<sub>N</sub></sub> percentages were associated with late treatment, younger age, higher CD4 T-cell counts, and lower plasma CRP and iFABP levels, but not with either sex or geographic origin (**Figure 2** and **Supplementary Table 2**). In multivariate analysis, higher CD4<sub>T<sub>N</sub></sub> percentages were associated with late treatment and higher CD4 T-cell counts (model 1). Multivariate models including other variables were not robust to outliers (data not shown).

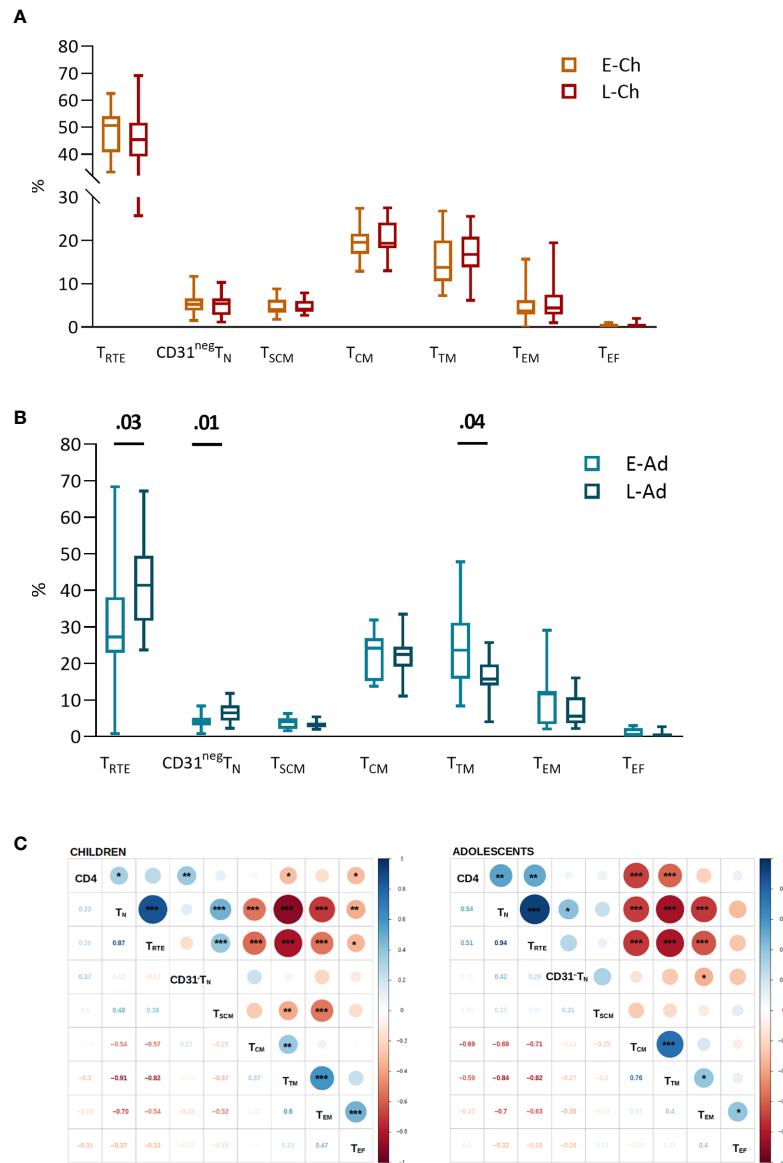
In conclusion, early ART was not significantly associated with higher CD4<sub>T<sub>N</sub></sub> percentages in children, and was associated with lower CD4<sub>T<sub>N</sub></sub> percentages in adolescents. In both groups, CD4<sub>T<sub>N</sub></sub> percentages were associated with CD4 T-cell count and levels of immune activation and/or intestinal permeability.

## Early ART Is Associated With Higher Naive CD8<sub>T<sub>N</sub></sub> Percentages in Children but Not in Adolescents

E-Ch had significantly higher CD8<sub>T<sub>N</sub></sub> percentages than L-Ch (48.7% vs. 31.0%,  $P = .001$ ), and lower percentages of the most differentiated effector memory cells (CD27<sup>-</sup>CD28<sup>-</sup>CD8<sub>T<sub>EM</sub></sub>, 6.1 vs. 14.6%,  $P = .009$ ) (**Figure 3A**). E-Ado had significantly higher percentages of the least differentiated CD27<sup>+</sup>CD28<sup>+</sup>CD8<sub>T<sub>EM</sub></sub> than L-ado (24.6% vs. 17.9%,  $P = .02$ ), and similar levels of CD8<sub>T<sub>N</sub></sub> (29.0% vs. 29.0%,  $P = .50$ ) (**Figure 3B**). CD4 T-cell counts were correlated with CD8 T-cell subset levels (**Figure 3C**). Overall, early ART was associated with the least differentiated CD8 T lymphocytes.

In children, higher CD8<sub>T<sub>N</sub></sub> percentages were associated with early ART, a longer normalized duration of viral suppression, lower blood total HIV DNA levels, higher CD4/CD8 ratio, and lower sCD163 levels, but not with either sex or geographic origin (**Figure 4** and **Supplementary Table 3**). Higher CD8<sub>T<sub>N</sub></sub> percentages were independently associated with early ART after adjustment for the duration of viral suppression (model 1), and with higher CD4/CD8 ratio and lower HIV DNA levels (model 2). The associations with CMV serostatus and sCD163 were not significant in multivariate models (data not shown).

In adolescents, higher CD8<sub>T<sub>N</sub></sub> percentages were not associated with early ART, sex or geographic origin, but were significantly associated with younger age, higher CD4/CD8 ratio, lower normalized cumulative viremia, and lower levels of activated CD4



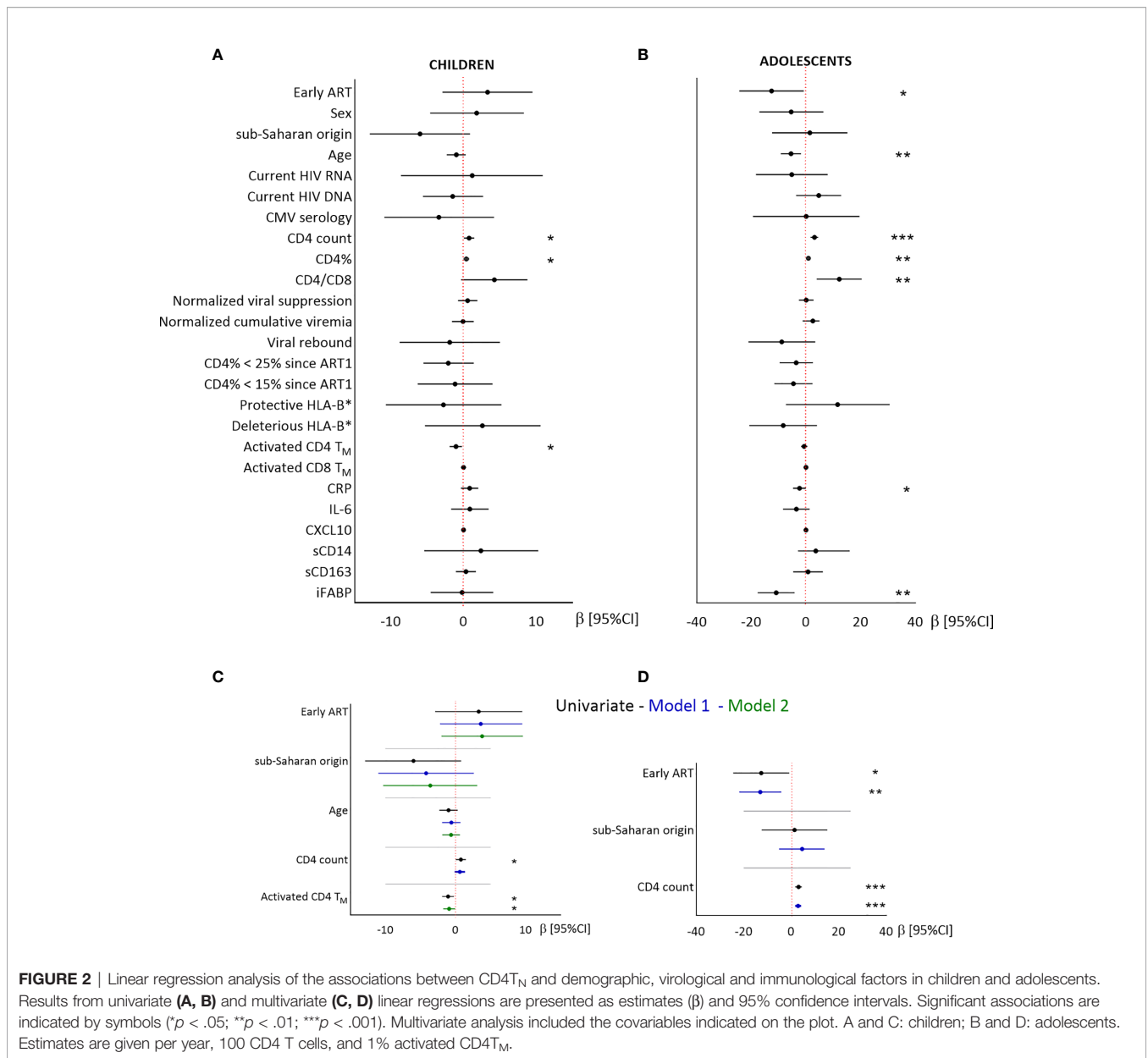
**FIGURE 1 |** CD4 T-cell subsets in patients treated early and late. T-cell phenotypes were assessed by flow cytometry on fresh whole blood (Supplementary Material). The percentage of each subset among total CD4 T lymphocytes is presented for children (A) and adolescents (B), on box and whiskers plots showing the minimum and maximum values. The early and late treatment groups were compared in Mann-Whitney tests; when significant, P values are shown on the graph. Subsets are presented from the least to the most differentiated, and were defined as naive T<sub>N</sub> (CD45RA<sup>+</sup>CCR7<sup>+</sup>); recent thymic emigrants, T<sub>RTE</sub> (CD45RA<sup>+</sup>CCR7<sup>+</sup>CD31<sup>+</sup>); CD31<sup>neg</sup>T<sub>N</sub> (CD45RA<sup>+</sup>CCR7<sup>+</sup>CD31<sup>-</sup>); central memory, T<sub>CM</sub> (CD45RA<sup>-</sup>CCR7<sup>+</sup>); transitional memory T<sub>TM</sub> (CD45RA<sup>-</sup>CCR7<sup>-</sup>CD27<sup>+</sup>); effector memory T<sub>EM</sub> (CD45RA<sup>-</sup>CCR7<sup>-</sup>CD27<sup>-</sup>) and effector T<sub>EF</sub> (CD45RA<sup>+</sup>CCR7<sup>-</sup>CD27<sup>-</sup>CD28<sup>-</sup>) cells. (C) The correlograms present Spearman's rank correlation coefficients as symbols (upper quadrants) and values. Significant associations are indicated by symbols (\*p < .05; \*\*p < .01; \*\*\*p < .001). CD4 T-cell counts were used for correlation analyses.

and CD8T<sub>M</sub> cells, IL-6 and iFABP (Figure 4 and Supplementary Table 3). In multivariate models, higher CD8T<sub>N</sub> percentages were associated with younger age and higher CD4/CD8 ratio (model 1), or with younger age and lower activated CD4T<sub>M</sub> cell levels (model 2).

In conclusion, early ART was significantly associated with higher CD8T<sub>N</sub> percentages in children, but not in adolescents. In both groups, higher CD8T<sub>N</sub> percentages were associated with higher CD4/CD8 ratio and stronger immune activation.

### Low CD4T<sub>N</sub> Levels in Adolescents Treated Early may Reflect Limited HIV-Driven T<sub>N</sub> Production by the Thymus

The lower CD4T<sub>N</sub> level in E-Ado than in other groups was unexpected. One patient had <5% CD4T<sub>N</sub> and this very low value was confirmed on the second blood sample. Relative to the reference values for Spain, a second patient had a CD4T<sub>N</sub> percentage below the 10<sup>th</sup> percentile, two had values above the

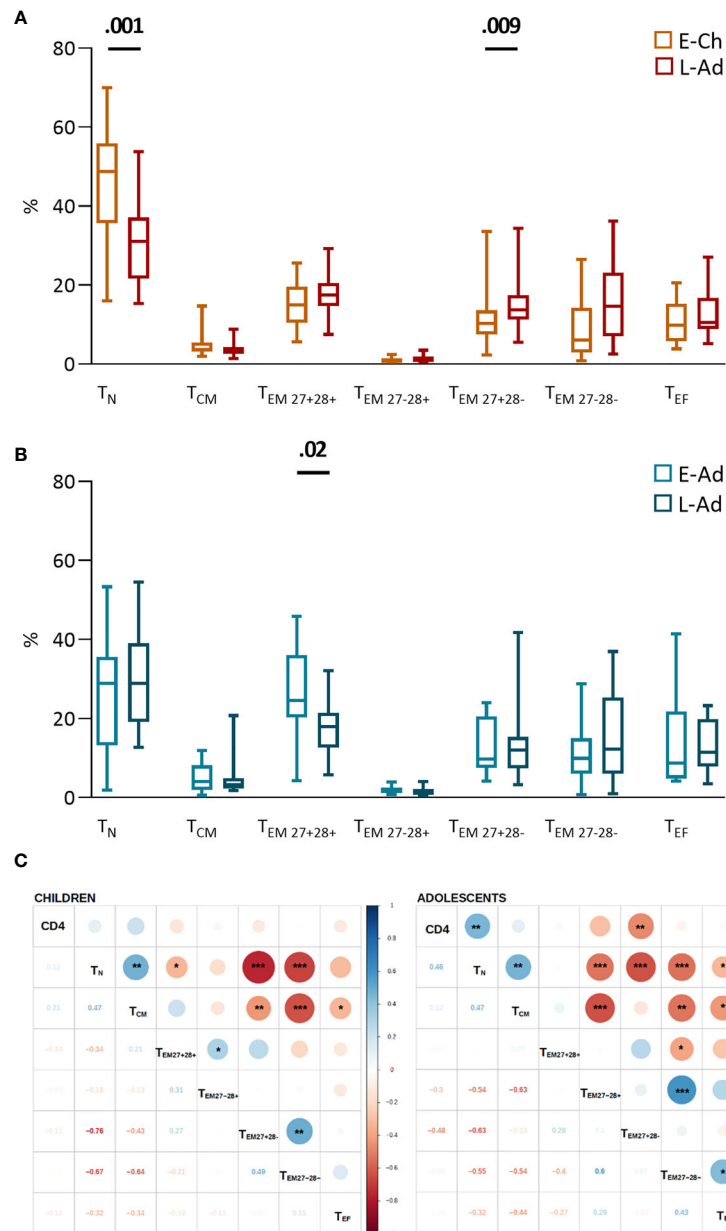


90<sup>th</sup> percentile, and five had CD4<sub>T<sub>N</sub></sub> percentages between the 10<sup>th</sup> and 90<sup>th</sup> percentiles (20).

The E-Ado were born between 1999 and 2003, before early ART was recommended for all newborns. We hypothesized that they might have received early ART because of early diagnosis, reflecting *in utero* infection, or clinical symptoms observed from birth or developing during the first six months of life. We compared E-Ado to E-Ch, who were born between 2004 and 2012, when early ART was the standard of care (Table 2). HIV DNA and/or RNA tests were performed during the first week of life for 29 of the participants receiving early treatment: five of six E-Ado and 13 of 23 E-Ch tested positive, indicating prenatal HIV contamination (Fisher’s exact test,  $P = .36$ ). Two patients in each group were diagnosed on the basis of suggestive symptoms ( $P =$

.25). Two of nine E-Ado and five of 27 E-Ch had at least one CDC stage C event ( $P = .99$ ); these events occurred before the age of six months in both groups. Graphical analyses of CD4<sub>T<sub>N</sub></sub> values as a function of early clinical events and early HIV diagnosis revealed no obvious relationship (Supplementary Figure 1). HIV RNA and CD4 levels before, at, and after ART1 initiation were not significantly different between E-Ch and E-Ado (Table 2). In conclusion, we found no statistically significant prescription bias (earlier ART related to early and/or severe infection) in E-Ado relative to E-Ch.

HIV replication leads to a depletion of CD4 T cells and drives the *de novo* generation of T<sub>N</sub> by the thymus in young viremic patients (21–23). We compared E-Ado to L-Ado, whose median percentages of lifetime with HIV RNA levels > 400 copies/mL



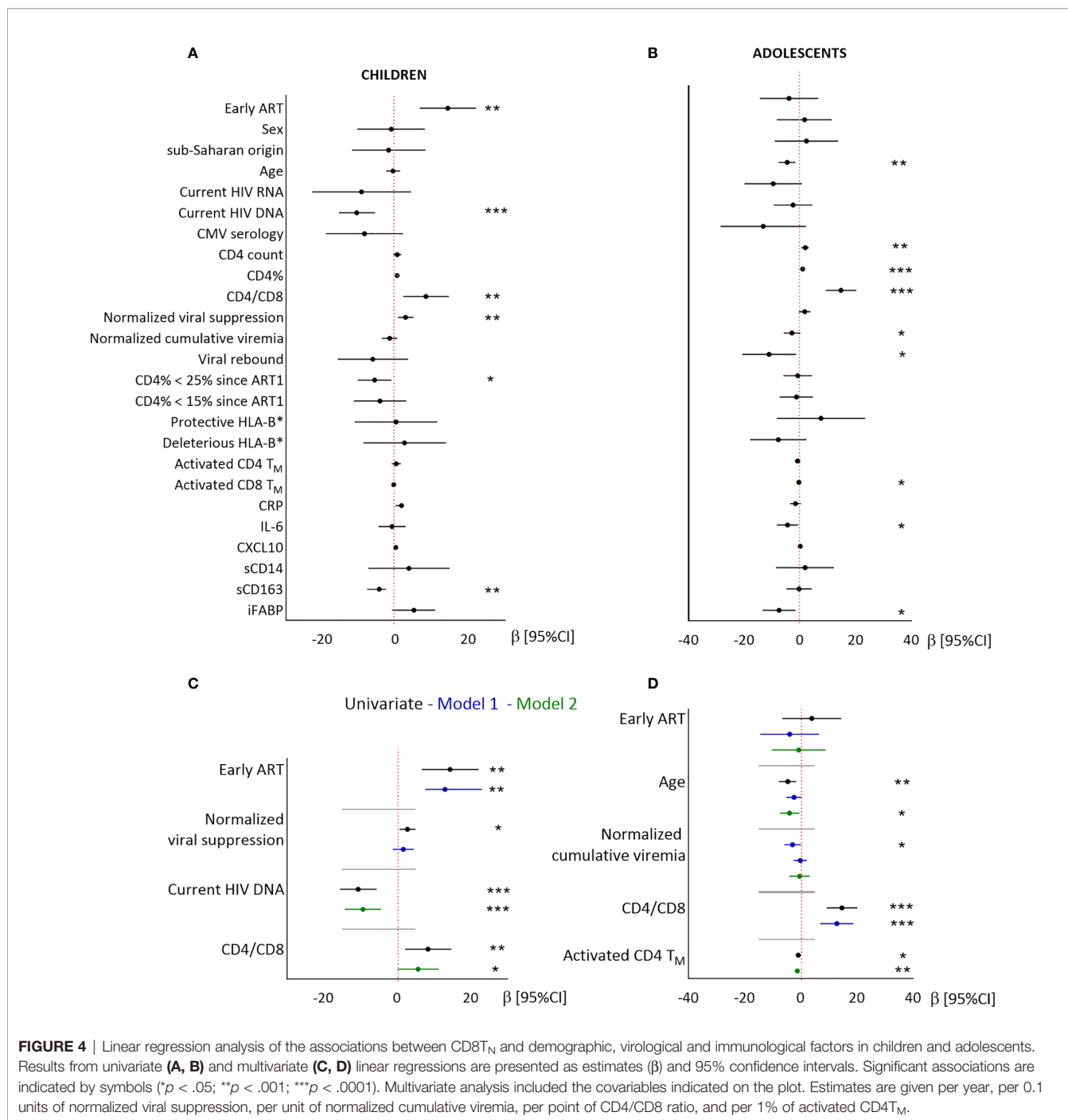
**FIGURE 3** | CD8 T-cell differentiation subsets in patients with early and late treatment initiation. T-cell phenotypes were assessed by flow cytometry on fresh whole blood (**Supplementary Material**). The percentage of each subset among total CD8 T lymphocytes is presented for children (**A**) and adolescents (**B**) on box and whiskers plots showing the minimum and maximum values. Early and late treatment groups were compared in Mann-Whitney tests; when significant, *P* values are shown on the graph. Subsets are presented from the least to the most differentiated, and were defined as naive  $T_N$  (CD45RA<sup>+</sup>CCR7<sup>+</sup>); central memory  $T_{CM}$  (CD45RA<sup>+</sup>CCR7<sup>+</sup>); effector memory  $T_{EM27+28+}$  (CD45RA<sup>+</sup>CCR7<sup>+</sup>CD27<sup>+</sup>CD28<sup>+</sup>);  $T_{EM27-28+}$  (CD45RA<sup>+</sup>CCR7<sup>+</sup>CD27<sup>+</sup>CD28<sup>+</sup>);  $T_{EM27+28-}$  (CD45RA<sup>+</sup>CCR7<sup>+</sup>CD27<sup>+</sup>CD28<sup>-</sup>);  $T_{EM27-28-}$  (CD45RA<sup>+</sup>CCR7<sup>+</sup>CD27<sup>-</sup>CD28<sup>-</sup>) and effector  $T_{EF}$  (CD45RA<sup>+</sup>CCR7<sup>-</sup>CD27<sup>-</sup>CD28<sup>-</sup>) cells. (**C**) The correlograms present Spearman's rank correlation coefficients as symbols (upper quadrants) and values. Significant associations are indicated by symbols (\**p* < .05; \*\**p* < .01; \*\*\**p* < .001). CD4 T-cell counts were used to calculate correlations.

were 9% and 60%, respectively. The percentage  $T_{RTE}$  among CD4 $T_N$  — an indicator of thymic output — was lower in E-Ado than in L-Ado, but this difference was not significant (**Table 3**). The absolute duration of HIV RNA levels < 400 copies/mL since ART1 was inversely correlated with both CD4 $T_N$  and CD4 $T_{RTE}$

levels in adolescents (Spearman's rho = -0.402, *P* = .02 and rho = -0.452, *P* = .01). Thus, the low CD4 $T_N$  levels in E-Ado may reflect limited HIV-driven  $T_N$  production by the thymus.

The immune response to pathogens, inflammation and immune activation drive the recruitment of naive





**FIGURE 4 |** Linear regression analysis of the associations between CD8T<sub>N</sub> and demographic, virological and immunological factors in children and adolescents. Results from univariate (**A, B**) and multivariate (**C, D**) linear regressions are presented as estimates (β) and 95% confidence intervals. Significant associations are indicated by symbols (\**p* < .05; \*\**p* < .001; \*\*\**p* < .0001). Multivariate analysis included the covariables indicated on the plot. Estimates are given per year, per 0.1 units of normalized viral suppression, per unit of normalized cumulative viremia, per point of CD4/CD8 ratio, and per 1% of activated CD4T<sub>M</sub>.

lymphocytes to the memory compartment and reduce thymic function (24). CMV prevalence, and the magnitude of CMV-specific serologic and T-cell responses were similar in L-Ado and E-Ado (Table 3), and blood samples from all patients tested negative for CMV DNA. These data argue against the low CD4T<sub>N</sub> levels being driven by an expansion of CMV-specific lymphocytes. E-Ado and L-Ado had similar levels of cellular and plasma markers of immune activation, inflammation, and gut permeability (Table 3). Overall, we did not observe higher

levels of immune factors specific to E-Ado that could explain their low CD4T<sub>N</sub> levels.

In conclusion, we considered three explanations for the low CD4T<sub>N</sub> levels in E-Ado and analyzed the characteristics of the patients and correlations between variables. We found no indication for either a more severe disease profile or a more activated immune profile in E-Ado than in the other groups. Our data suggest that E-Ado have lower CD4T<sub>N</sub> levels than L-Ado because thymic activity is enhanced to maintain T-cell homeostasis in L-ado.

**TABLE 2 |** HIV history variables in E-Ch and E-Ado.

Variable	E-Ch		E-Ado		P value <sup>a</sup>
	%	(n) Median [IQR]	%	(n) Median [IQR]	
Positive HIV RNA and/or HIV DNA assay before 7 days of age					
No	45.0	(10)	0.0	(0)	.36
Yes	55.0	(13)	100.0	(5)	
Diagnosis based on suggestive symptoms					
No	92.6	(22)	78.8	(7)	.26
Yes	7.4	(2)	22.2	(2)	
Occurrence of a CDC stage C event over lifetime					
No	81.5	(22)	77.8	(7)	1.00
Yes	18.5	(5)	22.2	(2)	
Age at the time of the CDC stage C event (months)	2	[1;3]	4	[3;4]	.29
Age at first HIV RNA < 50 copies/mL (days)	302	[150;536]	226	[173;682]	.72
Normalized lifetime spent with HIV RNA ≥ 400 copies/mL	0.16	[0.08;0.24]	0.09	[0.07;0.12]	.11
Zenith HIV RNA before ART1 initiation, log <sub>10</sub> copies/mL	5.6	[4.1;6.5]	5.5	[5.1;6.0]	.82
HIV RNA at ART1 initiation, log <sub>10</sub> copies/mL	4.8	[3.4;5.9]	5.5	[4.3;6.0]	.46
Zenith HIV RNA since ART1 initiation, log <sub>10</sub> copies/mL	3.9	[2.9;5.7]	3.7	[3.0;4.2]	.59
Viral rebound ≥ 400 copies/ml					
Never	66.7	(18)	66.7	(6)	1.00
Ever	33.3	(9)	33.3	(3)	
Nadir CD4 T-cell % before ART1 initiation	52	[29;60]	50	[36;56]	.81
CD4 T-cell % at ART1 initiation	52	[29;60]	50	[36;62]	.99
Nadir CD4 T-cell % since ART1 initiation	27	[20;34]	22	[14;26]	.06

<sup>a</sup>For categorical variables, frequencies, counts and Fisher's exact test P values are indicated; for quantitative variables, median, interquartile range and Mann-Whitney test P values are indicated. ART1, first highly active antiretroviral therapy.

**TABLE 3 |** Immune parameters in E-Ado and L-Ado.

Variable	E-Ado		L-Ado		P value <sup>a</sup>
	%	(n) Median [IQR]	%	(n) Median [IQR]	
% CD4 <sub>TRE</sub> (among CD4 <sub>TN</sub> )	75.0	[74.5;78.8]	78.7	[75.5;83.8]	.27
CMV serology					
Negative	22.2	(2)	4.8	(1)	.21
Positive	77.8	(7)	95.2	(20)	
CMV serology, titer	102.0	[73.3;109.0]	104.0	[74.6;124.0]	.28
CMV IFN-γ EliSpot					
Negative	0.0	(0)	12.5	(1)	1.00
Positive	100.0	(5)	87.5	(7)	
CMV IFN-γ EliSpot (cells/10 <sup>6</sup> PBMCs)	1 752	[592;3 754]	1 933	[724;3 344]	.94
HLA-DR <sup>+</sup> CD38 <sup>+</sup> CD4 <sub>TM</sub> (% among CD4 <sub>TN</sub> )	9.2	[8.3;10.0]	9.9	[9.0;13.3]	.29
HLA-DR <sup>+</sup> CD38 <sup>+</sup> CD8 <sub>TM</sub> (% among CD8 <sub>TN</sub> )	11.2	[5.9;14.1]	12.4	[10.2;19.2]	.42
Soluble CD14 (sCD14), μg/mL	1.52	[1.35;1.96]	1.45	[1.25;1.57]	.35
Soluble CD163 (sCD163), ng/mL	287	[279;311]	281	[217;355]	.90
CXCL10, pg/mL	27.1	[19.4;35.1]	40.5	[14.2;124.4]	.39
Interleukin-6 (IL-6), pg/mL	0.53	[0.40;1.33]	0.51	[0.33;1.04]	.72
C-reactive protein (CRP), μg/mL	1.04	[0.73;3.05]	0.57	[0.42;1.31]	.20
Intestinal fatty acid-binding protein (IFABP), ng/mL	0.55	[0.47;1.05]	0.56	[0.46;0.82]	.87

<sup>a</sup>For categorical variables, frequencies, counts and Fisher's exact test P values are indicated; for quantitative variables, median, interquartile range and Mann-Whitney test P values are indicated. IQR, interquartile range; CD4<sub>TRE</sub>, CD4 recent thymic emigrant; CD4<sub>TN</sub>%, naive CD4 T lymphocyte percentage; CMV, cytomegalovirus; IFN-γ, gamma interferon; CD4<sub>TM</sub>, memory CD4 T lymphocyte; CD8<sub>TM</sub>, memory CD8 T lymphocyte.

## DISCUSSION

The ANRS-EP59-CLEAC study included children and adolescents on ART with a good immunovirological status at the time of evaluation. In children, early ART had a beneficial effect for the maintenance of higher proportions of CD8<sub>TN</sub> lymphocytes and a marginal impact on CD4<sub>TN</sub> lymphocyte levels. By contrast, early ART had no beneficial effect in

adolescents. CD4<sub>TN</sub> and CD8<sub>TN</sub> levels were negatively related to cellular activation or gut permeability.

The reported percentages of CD4<sub>TN</sub> were in the range of those published for age-matched healthy subjects, and only two E-Ado had values below the 10<sup>th</sup> percentile (20). CD4<sub>TN</sub> percentages were strongly associated with CD4 T-cell counts, as previously reported (21, 25–27). In our study population, CD4 T-cell counts were in the same range in the early and late treatment groups for both age strata

(**Table 1**), reflecting a lessening of the benefits of early ART after several years of treatment (28–31). Most CD4T<sub>N</sub> are CD4T<sub>RTE</sub>, consistent with the thymus being active in pediatric patients (22, 23, 25, 32).

By contrast to these findings for CD4 T lymphocytes, early ART had a beneficial effect on CD8 T lymphocytes during childhood. E-Ch had median CD8T<sub>N</sub> values in the same range as uninfected controls, whereas the median values for L-Ch, E-Ado and L-Ado were lower than those of the reference groups (20); eight children and seven adolescents had values below the 10<sup>th</sup> percentile. E-Ch had a higher CD8T<sub>N</sub> and CD4/CD8 ratio, and lower total and CD8 T-cell counts than L-Ch (**Table 1**). The dichotomy between robust thymus-driven CD4T<sub>N</sub> recovery and the persistence of HIV-driven CD8T<sub>N</sub> loss is a key feature of pediatric HIV infection, reported in ART-treated young adults infected during the perinatal period, until their third decade of life (33). We show here that early ART keeps CD8T<sub>N</sub> levels high in children, with a possible impact on the future health of the child, because T<sub>N</sub> lymphocytes have a broad TCR repertoire and a high capacity to respond to new antigens (33).

T<sub>N</sub> percentages displayed a high level of interindividual variability in all groups. Among the expected factors (17, 34, 35), we found a trend towards lower CD8T<sub>N</sub> percentages in CMV-infected patients. CD4T<sub>N</sub> percentages were lower in patients born to mothers of Sub-Saharan origin than in those born to mothers of other geographic origins, although this difference was not significant. We found no association between T<sub>N</sub> levels and sex.

We document associations between higher immune activation and gut permeability biomarkers and lower T<sub>N</sub> percentages in at least one of the two age strata. In children, activated CD4T<sub>M</sub> were more strongly associated with CD4T<sub>N</sub> than CD4 T-cell counts. iFABP is a marker of intestinal permeability, and impaired mucosal immunity and decreases in thymic output influence each other (3, 36, 37). Higher CD4T<sub>N</sub> and CD8T<sub>N</sub> levels were found to be associated with lower iFABP levels, but only in adolescents. No difference in iFABP levels was observed between HIV-1-infected and uninfected infants (38–40), by contrast to what has been observed in adults (41). Thus, the data reported here and in other pediatric studies indicate that gut permeability and its impact on the immune system vary with age.

We found no association between CD4T<sub>N</sub> and virological history in either children or adolescents. Indeed, E-Ado had the lowest CD4T<sub>N</sub>, despite having the lowest normalized exposure to HIV RNA > 400 copies/ml (median 9% of their lifetime) and low HIV-DNA levels (16). In HIV-infected adolescents who had no access to early ART, CD4T<sub>N</sub> and T<sub>RTE</sub> levels were, paradoxically, higher in those with uncontrolled viremia, and HIV replication was correlated with CD4T<sub>N</sub> levels (21). Our observations are consistent with lower *de novo* thymic T<sub>N</sub> production in E-Ado than in L-Ado, whose CD4 T-cell compartment was less depleted by HIV replication. Reduced thymic activity may also account for the lack of association between early ART and higher CD8T<sub>N</sub> in adolescents. Such an association was expected because HIV replication is the main driver of CD8T<sub>N</sub> loss due to recruitment to the memory pool. However, this association was observed in children but not in adolescents. Thus, at adolescence, thymus-

driven T-cell reconstitution may have a stronger influence on naive T lymphocytes than HIV replication.

We identified a small number of adolescents with low T<sub>N</sub> levels despite long-term viral suppression. Thymic failure may occur in pediatric patients, albeit less frequently than in adults (32). Rare cases of perinatally infected adults on suppressive ART with low CD4 T-cell counts were reported in an Italian study (33). By contrast, low CD4T<sub>N</sub> counts were mostly associated with poor adherence and active viral replication in an older population of perinatally infected patients living in France (42).

One limitation of this study is a probable survivor bias, particularly for adolescents, potentially resulting in an underestimation of the deleterious impact of late ART initiation. It was not possible to attribute differences (or the lack of difference) between E and L groups definitively to the timing of first ART, because data from an observational study necessarily reflect differences in care according to chronological time and place of birth. Nevertheless, pathophysiological description according to the timing of ART initiation is worthwhile, because it reflects the diversity of patients managed in France and other high-income countries.

In conclusion, we report that, in children over five years of age and adolescents, early ART clearly has a beneficial effect on CD8T<sub>N</sub> cells. Children in whom treatment was initiated late had high CD4T<sub>N</sub> levels, probably because of their robust thymic activity. They achieved CD4 T-cell counts similar to those of their peers receiving early treatment, whilst on suppressive therapy. Thus, CD8T<sub>N</sub> levels, and their principal clinical correlate, CD4/CD8 ratio, are valuable indicators for use in long-term immune reconstitution studies in pediatric patients.

## DATA AVAILABILITY STATEMENT

The raw data supporting the conclusions of this article will be made available by the authors, without undue reservation.

## ETHICS STATEMENT

The studies involving human participants were reviewed and approved by Comité de protection des personnes Île-de-France V. Written informed consent to participate in this study was provided by the participants' legal guardian/next of kin.

## AUTHOR CONTRIBUTIONS

PF, JLC, CD, VA-F, JW and FB conceived the study and analyzed the data. PF supervised the clinical study. VA-F supervised the virological study. JW supervised the study methodology and statistical analyses. FB supervised the immunological study. TM, DB, and IF performed the immunological tests and analyzed immunological data. JLC performed the statistical analyses. PF, CD, AF, SB, AC, CF, IH, ML, NM, VM, M-DT and FV followed

the patients and contributed to clinical data. AH participated to data collection, TW managed the database. FB wrote the manuscript. All authors contributed to the article and approved the submitted version.

## FUNDING

ANRS (France Recherche Nord et Sud SIDA-HIV Hépatites) funded this study.

## ACKNOWLEDGMENTS

We thank the children and adolescents who participated in the study. We thank M. Hasan, S. Novault, P-H. Commere and T. Stephen from the *Unité de Technologie et Service Cytométrie et Biomarqueurs* (UTechS CB) at Institut Pasteur. The CMV pp65 peptide pool #11549 was obtained through the AIDS Reagent Program, Division of AIDS, NIAID, NIH. This text has been verified by a native English speaker.

## THE ANRS CLEAC STUDY GROUP

Hôpital Armand Trousseau, Paris: Mary-France Courcoux, Catherine Dollfus, Marie-Dominique Tabone, Geneviève

Vaudre; Hôpital Bicêtre, Le Kremlin-Bicêtre: Corinne Fourcade, Josiane Warszawski, Jérôme Lechenadec, Olivia Dialla, Laura Nailler, Lamy Ait-Si-Selmi, Isabelle Leymarie, Thierry Wack, Alexandre Hocin, Razika Feraon-Nanache; Centre hospitalier intercommunal de Créteil: Isabelle Hau; Hôpital Delafontaine, Saint-Denis: Cécile Gakobwa; Hôpital Necker-Enfants Malades, Paris: Véronique Avettand-Fenoël, Stéphane Blanche, Marine Fillion, Pierre Frange, Nizar Mahlaoui, Adeline Mélard, Florence Veber, Marie-Christine Mourey; Maternité Port-Royal, Paris: Valérie Marcou; Hôpital Robert Debré, Paris: Albert Faye, Martine Lévine, Sandrine Richard; Pitié-Salpêtrière, Paris: Brigitte Autran, Assia Samri, Mariama Diallo; Hôpital Saint-Louis: Sophie Caillat-Zucman, Kahina Amokrane, Rayna Ivanova-Derin, Centre hospitalier intercommunal de Villeneuve-Saint-Georges: Anne Chacé; Institut Pasteur Paris: Florence Buseyne, Thomas Montange, Damien Batalie, Ingrid Fert, Asier Saez-Cirion, Valérie Monceaux, Daniel Scott-Algara; ANRS: Lucie Marchand, Delphine Lebrasseur, Axel Levier. ANRS (*France Recherche Nord et Sud SIDA-HIV Hépatites*) was the sponsor of the study.

## SUPPLEMENTARY MATERIAL

The Supplementary Material for this article can be found online at: <https://www.frontiersin.org/articles/10.3389/fimmu.2021.662894/full#supplementary-material>

## REFERENCES

- Dollfus C, Le Chenadec J, Faye A, Blanche S, Briand N, Rouzioux C, et al. Long-term outcomes in adolescents perinatally infected with HIV-1 and followed up since birth in the French Perinatal Cohort (EPF/ANRS CO10). *Clin Infect Dis* (2010) 51:214–24. doi: 10.1086/653674
- Chiappini E, Bianconi M, Dalzini A, Petrara MR, Galli L, Giaquinto C, et al. Accelerated aging in perinatally HIV-infected children: clinical manifestations and pathogenetic mechanisms. *Aging* (2018) 10:3610–25. doi: 10.18632/aging.101622
- Zicari S, Sessa L, Cotugno N, Ruggiero A, Morrocchi E, Concato C, et al. Immune activation, inflammation, and non-AIDS co-morbidities in HIV-infected patients under long-term ART. *Viruses* (2019) 11:200. doi: 10.3390/v11030200
- Cotton MF, Violari A, Otwombe K, Panchia R, Dobbels E, Rabie H, et al. Early time-limited antiretroviral therapy versus deferred therapy in South African infants infected with HIV: results from the children with HIV early antiretroviral (CHER) randomised trial. *Lancet* (2013) 382:1555–63. doi: 10.1016/S0140-6736(13)61409-9
- Shiau S, Abrams EJ, Arpadi SM, Kuhn L. Early antiretroviral therapy in HIV-infected infants: can it lead to HIV remission? *Lancet HIV* (2018) 5:e250–e58. doi: 10.1016/S2352-3018(18)30012-2
- Sabin CA, Smith CJ, Monforte AD, Battegay M, Gabiano C, Galli L, et al. Response to combination antiretroviral therapy: variation by age - The Collaboration of Observational HIV Epidemiological Research Europe (COHERE) study group. *AIDS* (2008) 22:1463–73. doi: 10.1016/S0140-6736(11)60208-0
- Picat MQ, Lewis J, Musiime V, Prendergast A, Nathoo K, Kekitiinwa A, et al. Predicting patterns of long-term CD4 reconstitution in HIV-infected children starting antiretroviral therapy in sub-Saharan Africa: a cohort-based modelling study. *PLoS Med* (2013) 10:e1001542. doi: 10.1371/journal.pmed.1001542
- Azzoni L, Barbour R, Papasavvas E, Glencross DK, Stevens WS, Cotton MF, et al. Early ART results in greater immune reconstitution benefits in HIV-infected infants: Working with data missingness in a longitudinal dataset. *PLoS One* (2015) 10:e0145320. doi: 10.1371/journal.pone.0145320
- Guillen S, Prieto L, Jimenez de Ory S, Gonzalez-Tome MI, Rojo P, Navarro ML, et al. Prognostic factors of a lower CD4/CD8 ratio in long term viral suppression HIV infected children. *PLoS One* (2019) 14:e0220552. doi: 10.1371/journal.pone.0220552
- Buseyne F, Scott-Algara D, Bellal N, Burgard M, Rouzioux C, Blanche S, et al. The frequency of HIV-specific interferon-gamma-producing CD8 T cells is associated with both age and level of antigenic stimulation in HIV-1-infected children. *J Infect Dis* (2005) 192:1781–86. doi: 10.1086/497152
- Scott ZA, Chadwick EG, Gibson LL, Catalina MD, McManus MM, Yogev R, et al. Infrequent detection of HIV-1-specific, but not cytomegalovirus-specific, CD8+ T cell responses in young HIV-1-infected infants. *J Immunol* (2001) 167:7134–40. doi: 10.4049/jimmunol.167.12.7134
- Rinaldi S, Pallikkuth S, Cameron M, de Armas LR, Cotugno N, Dinh V, et al. Impact of early antiretroviral therapy initiation on HIV-specific CD4 and CD8 T-cell function in perinatally infected children. *J Immunol* (2020) 204:540–49. doi: 10.4049/jimmunol.1900856
- Aupiais C, Faye A, Le Chenadec J, Rouzioux C, Bouallag N, Laurent C, et al. Interruption of cART in clinical practice is associated with an increase in the long-term risk of subsequent immunosuppression in HIV-1-infected children. *Pediatr Infect Dis J* (2014) 33:1237–45. doi: 10.1097/inf.0000000000000450
- Palladino C, Briz V, Bellon JM, Climent FJ, de Ory SJ, Mellado MJ, et al. Determinants of highly active antiretroviral therapy duration in HIV-1-infected children and adolescents in Madrid, Spain, from 1996 to 2012. *PLoS One* (2014) 9:e96307. doi: 10.1371/journal.pone.0096307
- Kakkar F, Lee T, Hawkes MT, Brophy J, Lindy S, Singer J, et al. Challenges to achieving and maintaining viral suppression among children living with HIV. *AIDS* (2020) 34:687–97. doi: 10.1097/qad.0000000000002454

16. Avettand-Fenoel V, Le Chenadec J, Diallo M, Fillion M, Mélard A, Dollfus C, et al. Initiating antiretroviral treatment early in infancy has long-term benefits on the HIV reservoir in late childhood and adolescence. *Clin Infect Dis* (2021). doi: 10.1093/cid/ciaa1931. Advanced online publication.
17. Gaballa A, Clave E, Uhlin M, Toubert A, Arruda LCM. Evaluating thymic function after human hematopoietic stem cell transplantation in the personalized medicine era. *Front Immunol* (2020) 11:1341. doi: 10.3389/fimmu.2020.01341
18. Avettand-Fenoel V, Chaix ML, Blanche S, Burgard A, Floch C, Toure K, et al. LTR real-time PCR for HIV-1 DNA quantitation in blood cells for early diagnosis in infants born to seropositive mothers treated in HAART Area (ANRS CO 01). *J Med Virol* (2009) 81:217–23. doi: 10.1002/jmv.21390
19. Avettand-Fenoel V, Blanche S, Le Chenadec J, Scott-Algara D, Dollfus C, Viard JP, et al. Relationships between HIV disease history and blood HIV-1 DNA load in perinatally infected adolescents and young adults: the ANRS-EP38-IMMIP Study. *J Infect Dis* (2012) 205:1520–28. doi: 10.1093/infdis/jis233
20. Garcia-Prat M, Álvarez-Sierra D, Aguiló-Cucurull A, Salgado-Perandrés S, Briongos-Sebastian S, Franco-Jarava C, et al. Extended immunophenotyping reference values in a healthy pediatric population. *Cytom B Clin Cytom* (2019) 96:223–33. doi: 10.1002/cyto.b.21728
21. Blanche S, Scott-Algara D, Le Chenadec J, Didier C, Montange T, Avettand-Fenoel V, et al. Naive T lymphocytes and recent thymic emigrants are associated with HIV-1 disease history in French adolescents and young adults infected in the perinatal period: The ANRS-EP38-IMMIP study. *Clin Infect Dis* (2014) 58:573–87. doi: 10.1093/cid/cit729[doi]
22. Nobile M, Correa R, Borghans JAM, D'Agostino C, Schneider P, de Boer RJ, et al. De novo T-cell generation in patients at different ages and stages of HIV-1 disease. *Blood* (2004) 104:470–77. doi: 10.1182/blood-2003-12-4265
23. Aguilera-Sandoval CR, Yang OO, Jovic N, Lovato P, Chen DY, Boechar MI, et al. Supranormal thymic output up to 2 decades after HIV-1 infection. *AIDS* (2016) 30:701–11. doi: 10.1097/QAD.0000000000001010
24. Appay V, Fastenackels S, Katlama C, Ait-Mohand H, Schneider L, Guihot A, et al. Old age and anti-cytomegalovirus immunity are associated with altered T-cell reconstitution in HIV-1-infected patients. *AIDS* (2011) 25:1813–22. doi: 10.1097/QAD.0b013e32834640e6
25. Lewis J, Walker AS, Castro H, De Rossi A, Gibb DM, Giaquinto C, et al. Age and CD4 count at initiation of antiretroviral therapy in HIV-infected children: effects on long-term T-cell reconstitution. *J Infect Dis* (2012) 205:548–56. doi: 10.1093/infdis/jir787
26. Zakhour R, Tran DQ, Degaffe G, Bell CS, Donnachie E, Zhang W, et al. Recent thymus emigrant CD4<sup>+</sup> T cells predict HIV disease progression in patients with perinatally acquired HIV. *Clin Infect Dis* (2016) 62:1029–35. doi: 10.1093/cid/ciw030
27. Lewis J, Payne H, Walker AS, Otway K, Gibb DM, Babiker AG, et al. Thymic output and CD4 T-cell reconstitution in HIV-infected children on early and interrupted antiretroviral treatment: Evidence from the children with HIV early antiretroviral therapy trial. *Front Immunol* (2017) 8:1162. doi: 10.3389/fimmu.2017.01162
28. Goetghebuer T, Le Chenadec J, Haelterman E, Galli L, Dollfus C, Thorne C, et al. Short- and long-term immunological and virological outcome in HIV-infected infants according to the age at antiretroviral treatment initiation. *Clin Infect Dis* (2012) 54:878–81. doi: 10.1093/cid/cir950
29. Cohen S, Smit C, van Rossum AM, Fraaij PL, Wolfs TF, Geelen SP, et al. Long-term response to combination antiretroviral therapy in HIV-infected children in the Netherlands registered from 1996 to 2012. *AIDS* (2013) 27:2567–75. doi: 10.1097/01.aids.0000432451.75980.1b
30. Desmonde S, Dicko F, Koueta F, Eboua T, Balestre E, Amani-Bosse C, et al. Association between age at antiretroviral therapy initiation and 24-month immune response in West-African HIV-infected children. *AIDS* (2014) 28:1645–55. doi: 10.1097/QAD.0000000000000272
31. Novelli S, Lecroux C, Avettand-Fenoel V, Seng R, Essat A, Morlat P, et al. Long-term therapeutic impact of the timing of antiretroviral therapy in patients diagnosed with primary human immunodeficiency virus type 1 infection. *Clin Infect Dis* (2018) 66:1519–27. doi: 10.1093/cid/cix1068
32. Ferrando-Martinez S, De Pablo-Bernal RS, De Luna-Romero M, De Ory SJ, Genebat M, Pacheco YM, et al. Thymic function failure is associated with human immunodeficiency virus disease progression. *Clin Infect Dis* (2017) 64:1191–97. doi: 10.1093/cid/cix095
33. Paghara S, Quiros-Roldan E, Sottini A, Properzi M, Castelli F, Imberti L. Lymphocyte homeostasis is maintained in perinatally HIV-infected patients after three decades of life. *Immun Ageing* (2019) 16:26. doi: 10.1186/s12979-019-0166-7
34. Turner JE, Campbell JP, Edwards KM, Howarth LJ, Pawelec G, Aldred S, et al. Rudimentary signs of immunosenescence in Cytomegalovirus-seropositive healthy young adults. *Age* (2014) 36:287–97. doi: 10.1007/s11357-013-9557-4
35. van den Heuvel D, Jansen MAE, Nasserinejad K, Dik WA, van Lochem EG, Bakker-Jonges LE, et al. Effects of nongenetic factors on immune cell dynamics in early childhood: The Generation R Study. *J Allergy Clin Immunol* (2017) 139:1923–34.e17. doi: 10.1016/j.jaci.2016.10.023
36. Bourgeois C, Hao Z, Rajewsky K, Potocnik AJ, Stockinger B. Ablation of thymic export causes accelerated decay of naive CD4 T cells in the periphery because of activation by environmental antigen. *PNAS* (2008) 105:8691–96. doi: 10.1073/pnas.0803732105
37. De Voeght A, Martens H, Renard C, Vaira D, Debruche M, Simonet J, et al. Exploring the link between innate immune activation and thymic function by measuring sCD14 and TRECs in HIV patients living in Belgium. *PLoS One* (2017) 12:e0185761. doi: 10.1371/journal.pone.0185761
38. Koay WLA, Lindsey JC, Uprety P, Bwakura-Dangarembizi M, Weinberg A, Levin MJ, et al. Intestinal integrity biomarkers in early antiretroviral-treated perinatally HIV-1-infected infants. *J Infect Dis* (2018) 218:1085–89. doi: 10.1093/infdis/jiy271
39. Prendergast AJ, Chasekwa B, Rukobo S, Govha M, Mutasa K, Ntozini R, et al. Intestinal damage and inflammatory biomarkers in human immunodeficiency virus (HIV)-exposed and HIV-infected Zimbabwean infants. *J Inf Dis* (2017) 216:651–61. doi: 10.1093/infdis/jix367
40. Dirajlal-Fargo S, El-Kamari V, Weiner L, Shan L, Sattar A, Kulkarni M, et al. Altered intestinal permeability and fungal translocation in Ugandan children with human immunodeficiency virus. *Clin Infect Dis* (2020) 70:2413–22. doi: 10.1093/cid/ciz561
41. Sandler NG, Wand H, Roque A, Law M, Nason MC, Nixon DE, et al. Plasma levels of soluble CD14 independently predict mortality in HIV infection. *J Infect Dis* (2011) 203:780–90. doi: 10.1093/infdis/jiq118
42. Fastenackels S, Sauce D, Vigouroux C, Avettand-Fenoel V, Bastard J-P, Fellahi S, et al. HIV-mediated immune aging in young adults infected perinatally or during childhood. *AIDS* (2019) 33:1705–10. doi: 10.1097/QAD.0000000000002275

**Conflict of Interest:** The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Copyright © 2021 Frange, Montange, Le Chenadec, Batalie, Fert, Dollfus, Faye, Blanche, Chacé, Fourcade, Hau, Levine, Mahlaoui, Marcou, Tabone, Veber, Hocin, Wack, Avettand-Fenoel, Warszawski and Buseyne. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.