



HAL
open science

Portal hypertension and hepatocellular carcinoma: des liaisons dangereuses ...

Manon Allaire, Marika Rudler, Dominique Thabut

► **To cite this version:**

Manon Allaire, Marika Rudler, Dominique Thabut. Portal hypertension and hepatocellular carcinoma: des liaisons dangereuses Liver International, 2021, 10.1111/liv.14977 . hal-03247480

HAL Id: hal-03247480

<https://hal.sorbonne-universite.fr/hal-03247480v1>

Submitted on 3 Jun 2021

HAL is a multi-disciplinary open access archive for the deposit and dissemination of scientific research documents, whether they are published or not. The documents may come from teaching and research institutions in France or abroad, or from public or private research centers.

L'archive ouverte pluridisciplinaire **HAL**, est destinée au dépôt et à la diffusion de documents scientifiques de niveau recherche, publiés ou non, émanant des établissements d'enseignement et de recherche français ou étrangers, des laboratoires publics ou privés.

Portal hypertension and hepatocellular carcinoma: des liaisons dangereuses

...

Manon Allaire^{1,2}, Marika Rudler^{1,3}, Dominique Thabut^{1,3}

1. Sorbonne Université, Service d'Hépatolo-gastroentérologie, Hôpitaux Universitaires Pitié Salpêtrière - Charles Foix, AP-HP, Paris, France.
2. Inserm U1149, Centre de Recherche sur l'Inflammation, France Faculté de Médecine Xavier Bichat, Université Paris Diderot, Paris, France.
3. Sorbonne Université, INSERM, Centre de Recherche Saint-Antoine (CRSA), Institute of Cardiometabolism and Nutrition (ICAN), Paris, France

Running Title: Portal hypertension and Hepatocellular carcinoma

Correspondance and reprints to:

Pr Thabut
Service d'hépatogastroentérologie et oncologie
Groupe Hospitalier Pitié-Salpêtrière
47-83 Boulevard de l'Hôpital
75013 Paris, France
tél: 01.42.16.10.34 - fax: 01.42.16.11.60
mail: dominique.thabut@aphp.fr

Dr Manon ALLAIRE
Service d'hépatogastroentérologie et oncologie
Groupe Hospitalier Pitié-Salpêtrière
47-83 Boulevard de l'Hôpital
75013 Paris, France
tél: 01.42.16.10.34 - fax: 01.42.16.11.60
mail: manon.allaire@aphp.fr

Acknowledgement : none

Conflict of interest statement: nothing to declare regarding this work

Financial support: none

Number of tables: 2

Number of figures: 2

Electronic word count: 3548 excluding references

Guarantor of the article:

Dr Manon ALLAIRE
Service d'hépatogastroentérologie et oncologie
Groupe Hospitalier Pitié-Salpêtrière
47-83 Boulevard de l'Hôpital
75013 Paris, France
tél: 01.42.16.10.34 - fax: 01.42.16.11.60
mail: manon.allaire@aphp.fr

Author contributions:

Study concept and design: Allaire and Thabut
Drafting of the manuscript: Allaire, Rudler and Thabut
Final approval of the version to be submitted : Allaire, Rudler and Thabut

All authors approved the final version of the manuscript

Abstract:

Introduction: Portal hypertension (PHT) and Hepatocellular carcinoma (HCC) are major complications of cirrhosis which significantly contribute to morbidity and mortality. In this review, **we aim** to describe the consequences of both angiogenesis and inflammation in the pathogenesis of PHT and HCC, but also the difficulty to propose adapted treatment when PHT and HCC coexist in the same patients.

Methods: Studies for review in this article were retrieved from the PubMed database using literature published in English until March 2021.

Results: PHT occurs secondary to an increase of intrahepatic vascular resistances, the opening of portosystemic collateral vessels and the formation of neovessels, related to vascular endothelial growth factor (VEGF). Recently, bacterial translocation-mediated inflammation was also identified as a major contributor to PHT. Interestingly, VEGF and chronic inflammation also contribute to HCC occurrence. As PHT and HCC often coexist in the same patient, management of PHT and its related complications as well as HCC treatment appear more complex. Indeed, PHT-related complications such as significant ascites may hamper the access to HCC treatment and the presence of HCC is also independently associated with poor prognosis in patients with acute variceal bleeding related to PHT. Due to their respective mechanism of action, the combination of Atezolizumab and Bevacizumab for advanced HCC may impact the level of PHT and its related complications and to date, no real-life data are available.

Finally, appropriate evaluation and treatment of PHT remains a major issue in order to improve the outcome of HCC patients.

Key words: HCC, portal hypertension, VEGF, Atezolizumab/Bevacizumab, Inflammation

Abbreviations

AVB: acute variceal bleeding
CSPH: clinically significant portal hypertension
DAMP: damage-associated molecular pattern
HCC: hepatocellular carcinoma
HSC: hepatic stellate cell
HVPG: hepatic venous pressure gradient
IL: interleukine
LSEC: liver sinusoidal endothelial cell
NO: nitric oxid
PAMP: pathogen-associated molecular pattern
PDGF: platelet-derived growth factor
PHT: portal hypertension
PPR: pattern recognition receptors
RPS: reactive oxygen species
TKIs: tyrosine kinase inhibitors
TLR: toll like receptor
VEGF: vascular endothelial growth factor

Introduction

Portal hypertension (PHT) is a major complication of cirrhosis and is responsible for variceal bleeding, ascites and hepatorenal syndrome, which significantly contribute to morbidity and mortality. In the case of cirrhosis, PHT occurs secondary to an increase of intrahepatic vascular resistances due to sinusoidal alterations and the presence of fibrosis that cause distortion of the vascular architecture within the liver. The opening of portosystemic collateral vessels, and the formation of neovessels, related to vascular endothelial growth factor (VEGF) and platelet-derived growth factor (PDGF) production (so called angiogenic process) (1–3), as well as a subsequent increase of the splanchnic blood flow contribute and perpetuate PHT. Recently, bacterial translocation-mediated inflammation have also been identified as a major contributor to PHT occurrence (4,5). In clinical practice, PHT is defined by an increase in portal pressure above 12mmHg or by a pressure difference between portal vein pressure and hepatic venous pressure higher than 5 mmHg. Different methods to measure the pressure in the portal vein exist but the most used technique in patients is the measurement of the hepatic venous pressure gradient (HVPG) through a transjugular approach. A HVPG higher than 10mmHg is associated with ascites and variceal occurrence, so called clinically significant portal hypertension (CSPH) and a value higher than 12 mmHg exposes the patient to a risk of variceal bleeding.

Hepatocellular carcinoma (HCC) is also a major complication of cirrhosis. Similar to PHT, an increased level of VEGF and chronic inflammation also contribute to HCC occurrence (6). Moreover, patients with PHT present higher risk to develop HCC, and HCC through changes in the hepatic architecture and vascular invasion also contributes to PHT occurrence (7,8). As PHT and HCC often coexist in the same patient, management of PHT and its related complications as well as HCC treatment appear more complex. Indeed, CSPH may hamper the access to curative treatment for HCC such as liver resection/ablation but also to locoregional therapies or systemic treatment in case of significant ascites (6). Ascites and esophageal varices have been identified as predictive factors of death in patients with HCC independently of the severity of the underlying liver disease and the HCC stage (9,10). Conversely, the presence of HCC is also independently associated with poor prognosis in patients with acute variceal bleeding (AVB) related to PHT (8,9,11,12). Thus, understanding the interaction between HCC and PTH is a major issue in order to improve the management of such complications and the outcome of these patients.

Recently, the positive results of the Imbrave 150 study, a randomized study comparing Atezolizumab (anti PD-L1) in combination with Bevacizumab (anti VEGF) versus Sorafenib

(tyrosine kinase inhibitor) have prompted us to redefine our management strategy for advanced HCC by proposing the combination of Atezolizumab and Bevacizumab as a first line treatment (13). Patients were well selected and few complications related to PHT were noticed in this phase 3 study. However, due to their respective mechanism of action, the combination of Atezolizumab and Bevacizumab may potentially impact the level of PHT and its related complications and to date, no real-life data are available.

In this review, we will focus (i) on the consequences of both angiogenesis and inflammation in the pathogenesis of PHT and HCC, (ii) on the link between PHT and HCC and (iii) we will discuss the potential impact of the combination Atezolizumab and Bevacizumab on PHT.

Studies for review in this article were retrieved from the PubMed database using the search terms ‘hepatocellular carcinoma’, ‘liver cancer’, and ‘primary liver carcinoma’, both individually and in combination with the terms ‘portal hypertension’, ‘acute variceal bleeding’, ‘ascites’, and ‘TIPS’. The search included literature published in English until March 2021.

1. Angiogenesis and chronic inflammation contribute to PHT pathogenesis

In case of cirrhosis, PHT is initiated by an increased hepatic resistance caused by the distortion of liver vascular architecture due to fibrosis and regenerative nodules but also to an increase of vasoconstrictor stimuli. PHT is further perpetuated by opening of collateral vessels/formation of neovessels and changes in the systemic circulation that culminate in an increased portal-tributary inflow (1,14). The demonstration that angiogenesis is a main actor in the pathophysiology of PHT is relatively recent (3). During decades, the formation of portosystemic collateral vessels in PHT has been related to the opening of preexisting vascular channels secondary to the increased portal pressure. Recently, a neovascularization in the mesenteric vascular bed and in the portosystemic collateral vessels has been described and attributed to VEGF and PDGF production (1). VEGF and VEGF receptor type 2 expression were observed in the mesentery from portal vein-stenosed rats and the inhibition of VEGF receptor 2 signaling was associated with a 50% decrease in the formation of portal-systemic collateral blood vessels in rodent models with portal vein stenosis (15,16). VEGF increases vascular permeability and induces the migration and survival of endothelial cells favoring angiogenesis process (17,18). In addition to promoting neovessels formation in the mesentery and portosystemic system, VEGF also plays a role within the liver parenchyma. Increased hypoxia, inflammatory cytokines (IL-6, IL-1 α), growth factors (epidermal growth

factor, transforming growth factor- α and - β , fibroblast growth factor, PDGF), shear and oxidative stress within the liver lead to increase VEGF production by hepatocytes but also Hepatic Stellate Cells (HSCs) (19). HSCs are activated through its VEGF receptor leading to fibrosis occurrence and also VEGF production that interacts with adjacent liver sinusoidal endothelial cells (LSECs) (3). HSC-signaling pathways involving PDGF regulate microvascular structure and function in liver, and PHT (20). Portal myofibroblasts also appear to be critical in pathological angiogenesis through the production of collagen that stabilizes the newly formed vessels (21,22). Interestingly, sinusoidal angiogenesis depends on the liver stiffness and mechanotransduction appears as an important pathway in liver fibrosis progression (23–25). LSECs exhibit pro-angiogenic properties and promote fibrosis through HSCs activation on low stiffness substrates at early disease stage. On contrary, LSECs showed random migration and leaky sinusoids in high stiffness substrates (24). The newly formed intrahepatic neovessels present varying diameter and flow pattern leading to the impairment of oxygen and nutrient supply to hepatocytes and to the recruitment of inflammatory cells (3). Thus, angiogenesis mediated by VEGF promotes an extensive network of portosystemic collateral vessels, contribute to increased splanchnic and portal venous inflow that perpetuates and exacerbates PHT and favors liver fibrosis progression and inflammation (Figure 1). The use of anti VEGF such as Sorafenib was associated with PHT reduction and targeting angiogenesis at early stage of the liver disease may appear as a good strategy (26–29). However, VEGF also plays a role in hepatic tissue repair and fibrosis resolution (30).

Chronic inflammation and PHT are also linked since bacterial infection increases portal pressure and PHT favors bacterial translocation (31,32). PHT increases intestinal permeability, thus favoring bacterial translocation. Dysbiosis is associated with cirrhosis, and increases with the severity of liver disease (33,34). Bacterial translocation corresponds to the passage of microbial products through the mucosa into mesenteric lymph nodes and then to the liver. In the liver, the innate immune system recognizes damage-associated molecular pattern (DAMPs) and pathogen-associated molecular pattern (PAMPs) through pattern recognition receptors (PRRs), such as Toll-like receptor (TLRs) (35). Kupffer cells are the first cells to respond to PAMPs through TLR4 and consequently adopt a pro-inflammatory phenotype leading to the production of proinflammatory cytokines such as TNF, IL-6 and IL-12 and to the generation of reactive oxygen species (ROS) (36). ROS production will conduct to a decrease in nitric oxide (NO) secretion, which usually regulates intrahepatic vascular tone

by maintaining HSCs in a quiescent phenotype, and thus promote fibrosis process (37). ROS will also contribute to local tissue damage and propagating innate immune signaling through DAMPs and promote VEGF production (38). In addition, bacterial translocation will conduct to stimulation of TLR4-mediated signaling in HSCs and LSECs directly promoting angiogenesis and fibrosis progression (Figure 1). In animal models, TLR4 knockout mice are protected from fibrosis and PHT following bile duct ligation (39,40). Similar results were observed in mice treated with rifaximin after bile duct ligation (39,40). In humans, a trend towards HVPG reduction was observed in controlled study using rifaximin but these finding remain controversial (41,42). A similar reduction in portal pressure was observed in patients with Acute on Chronic liver Failure treated with anti-TNF, however this treatment was not adopted due to high rates of infection after anti-TNF treatment (43). Human data are scarce, further studies are needed to confirm the dialogue between VEGF, DAMPS and PAMPS in PHT progression.

2. Angiogenesis and chronic inflammation also contribute to hepatocarcinogenesis

Levels of VEGF are increased in HCC and correlate with tumor angiogenesis and progression. In fact, high levels of VEGF are associated with rapid disease progression and reduced survival, and VEGF gene amplifications have been described in 4 to 8% of HCCs (44). Interaction between VEGF and its receptor results in endothelial proliferation and migration, and formation of new tumor blood vessels generating rapid growth and dissemination. In addition, the neovessels have abnormally leaky vasculature resulting in areas of high interstitial pressure and severe hypoxia and acidosis, who further drive malignant potential, induced epithelial-mesenchymal transition and the recruitment of immune cells (45). VEGF induces tumor proliferation via the induction of hepatocyte growth factor by macrophages (46). VEGF also leads to disruption of hepatocellular tight junctions that may promote HCC spreading into normal liver parenchyma (47). Besides inducing tumor angiogenesis, VEGF also mediates immunosuppression within the tumor microenvironment (TME) by promoting immunosuppressive cells such as regulatory T cells, myeloid derived suppressive cells and tumor associated macrophages but also by suppressing antigen suppressive cells and cytotoxic T lymphocyte through TOX activation after binding VEGFR2 (48–51) (Figure 1). Thus, the use of anti-VEGF therapy results in vascular normalization and thereby reduces tumor hypoxia that improves the anticancer activity of infiltrating immune cells (52,53). However, results of phase 2 trial using anti-VEGF either alone or in combination with chemotherapy were disappointed in HCC patients (54). Recently,

Ramucirumab, an VEGFR-2 antagonist, was associated with improved overall survival in a phase III trial when compared to placebo in a subgroup of patients with baseline alpha-fetoprotein (AFP) ≥ 400 ng/mL (8.5 versus 7.3 months, $p=0.0199$); nevertheless Ramucirumab is poorly used in practice due to a limited increase in survival (55).

Chronic inflammation is a major trigger of HCC carcinogenesis and tumor progression. Infiltration of inflammatory cells within the premalignant environment produces various cytokines such as IL-6 that enhance the transformation of the liver progenitor cells into a more cancerous phenotype (56), but also synthesis of growth factors, chemokines and proangiogenic factor which create a TME that supports the transformation of hepatocytes, and also induces their survival through activation of anti-apoptotic pathway and inhibition of immune surveillance (Figure 1). Chronic inflammation also leads to tissue remodeling through the crosstalk between immune cells, HSCs and LSECs that promotes ECM degradation, the release of growth factors and consequently tumor growth. In addition, platelets also interact with immune cells and play a role in the hepatocarcinogenesis process. Once activated, platelets recruit inflammatory cells and induce growth factors production such as PDGF that mediates cellular proliferation and angiogenesis (57). High levels of platelets correlate with large HCC. In addition, platelets interact with LSECs that secrete specific chemokines regulating the recruitment and adhesion of leukocytes. Recently, the use of antiplatelet therapy allowed to prevent non-alcoholic steatohepatitis and subsequent HCC development in several dietary and genetic mouse models (58).

Immune cells also prevent tumor occurrence and growth: there is a complex interaction between pro-tumoral and anti-tumoral signals mediated by the immune system, the tumors cells and the TME, which can evolve over time resulting either in tumor elimination or tumor progression. TME of HCC is strongly immunosuppressive, as a result of VEGF interaction with immune cells (48–51), and also because HCC tumors cells may directly recruit TReg and tumor associated macrophages (59). TME, through HSCs, also promotes the differentiation of blood monocytes into myeloid derived suppressor cells through IL-6 signaling which enhances immune check point signaling, decreases anti tumoral immune response mediated by natural killer and T cells cytotoxicity and contributes to an immunosuppressive TME (60). Immune check point factors such as programmed death-1 (PD-1), PDL-1, cytotoxic t-lymphocyte antigen 4 (CTLA-4) down regulate the amplitude of immune response such as inhibition of T cell activity (CD8+ effectors and CD4+ helpers) that participates in suppressing tumor immunity and appears as a major mechanism of immune resistance (61).

Immune checkpoint inhibitors (ICIs) allow to maintain the physiological cell-mediated immune response against tumor cells and are the new promising approaches in oncology. However, even if phase III trials using nivolumab versus sorafenib in first line for advanced HCC or using pembrolizumab vs placebo in second line showed interesting results regarding overall survival and disease free survival, statistical analysis did not reach significance (62,63).

3. A link between HCC and PHT

CSPH is predictive of HCC occurrence independently of the severity of the underlying cirrhosis (7,8). HCC also increases HVPG through the presence of arteriovenous shunting within the tumor and modifications of liver architecture. Moreover, HCC can be associated with vascular invasion of the portal and/or its branches and contributes to PHT, thus exposing the patients to AVB risk. However, PHT evaluation remains difficult, especially in patients with advanced HCC. Baveno criteria (64) do not seem accurate to evaluate PHT as platelets level may be impacted by HCC presence and the presence of a large tumor, especially in the right lobe, may interfere with the elastometry results. Moreover, the presence of portal invasion makes HVPG measurement not always accurate. Thus, appropriate evaluation of PHT remains a major issue in patients with advanced HCC, and regular endoscopy should be preferred for the screening of esophageal varices, especially in patients with vascular invasion.

CSPH impacts the choice of HCC treatment. CSPH is a well identified predictive factor for liver decompensation and death after liver resection. Currently, optimal surgical candidacy for resection is based on a multiparametric evaluation including compensated Child-Pugh class A liver function with MELD score < 10, to be matched with grade of PHT, acceptable remaining parenchyma and the possibility of a laparoscopic/minimally invasive approach (6,65). CSPH is also associated with decrease overall survival after ablation and chemoembolization (66,67) Moreover, the presence of clinically significant ascites precludes any locoregional or systemic therapy. Interestingly, the occurrence of AVB and clinically significant ascites is associated with poor prognosis in HCC patients (10,11,68). Interestingly, secondary prophylaxis after AVB was associated with increased overall survival justifying the need to treat both PHT and HCC (68).

A treatment of CSPH by TIPS placement may be beneficial in some situations as it increases overall survival in case of AVB in high-risk patients (concept of preemptive TIPS), and transplant free survival in case of recurrent/refractory ascites (69–71). Moreover, in HCC

patients, TIPS can also allow the access to HCC treatments (either alone or in a downstaging perspective before liver transplantation) (69,72,73). However, HCC is generally considered as a contra-indication to TIPS placement, due to the fear of HCC spread, and the poor outcome of these patients (74–77). There is also a concern regarding the potential increased risk of liver failure after locoregional therapies and a decrease in efficacy of HCC treatments (74–77). In fact, few data are available regarding these points, and whether or not TIPS is a real risk factor for higher toxicity after locoregional therapy remains uncertain. At present, TIPS should be considered when needed in cirrhotic patients with Milan-In HCC to improve survival and as a bridge to liver transplant, in its classical indications (salvage, preemptive, failure of secondary prophylaxis, recurrent/refractory ascites). As for all cirrhotic patients, salvage TIPS should be performed when needed. However, there must be a careful selection of candidates in all other settings, and TIPS should be contraindicated in patients with heart failure, significant hepatic encephalopathy and advanced liver failure (Child-Pugh score >13 in case of preemptive for AVB, high bilirubin/low platelets count or a MELD score >19 in case of ascites) (Figure 2) (70). Moreover, as far as HCC treatment is concerned, one can envision pre-operative TIPS placement before liver resection in patients with CSPH, and in order to treat ascites and allow locoregional treatments, either as curative options or in a perspective of downstaging.

4. Impact of systemic therapy for HCC on PHT

During the last decades, tyrosine kinase inhibitors (TKIs) were the standard of treatment for advanced HCC. Sorafenib and Lenvatinib were used in first line treatment while Regorafenib and Cabozantinib in second line (78–81). All of these TKIs target the VEGFR and may impact PHT pathophysiology (Table 1). Inhibition of VEGF-mediated angiogenesis by drugs such as sorafenib allowed to reduce portal pressure, portosystemic collateralization and fibrosis in animal models (15,26–29,82). Similar results were observed in rodents models treated with regorafenib and exposed to fibrotic models such as bile duct ligation, chronic carbon tetrachloride injections and partial portal vein ligation (83), and with Lenvatinib (84). In humans, portal venous area but not portal venous flow velocity was also significantly decreased, as well as the congestion index (portal venous area divided by portal venous flow velocity) in 25 Child-Pugh A patients after two weeks administration of Sorafenib, suggesting a potential beneficial effect on PHT (85). On the contrary, the congestion index was significantly worsened in 28 Child-Pugh A patients who received two weeks of Lenvatinib for advanced HCC (86). Contrary to Sorafenib, Lenvatinib also targets Fibroblast Growth Factor

(FGF) receptors, and FGF19 and FGF 21 are known to promote liver regeneration and the maintenance of hepatic metabolism which could impact the underlying liver disease and favor PHT, however further studies are needed to address this point. As discussed before, AVB is a major issue in patients with HCC and similar PHT bleeding complications rates were observed in the phase 3 studies for advanced HCC (less than 3%) (Table 1) (13,55,62,78–81). However, in these series, patients were well selected as recent history of bleeding was an exclusion criterion, and real-life data are needed (Table 2). Interestingly, neoplastic portal vein thrombosis was independently associated with AVB in patients treated with Sorafenib (87). Such patients should benefit from regular screening and adapted bleeding prevention therapy (88).

5. PHT and the new combination Bevacizumab and Atezolizumab

Bevacizumab is an antibody directed against VEGF who plays a major role in PHT pathophysiology as described before. Atezolizumab is an immune checkpoint inhibitor (anti-PDL1) which aims to restore anti-tumor immunity and may contribute to PAMPs and DAMPs elimination. Therefore, these two molecules could theoretically limit the onset of PHT. However, in the Imbrave 150 study, bleeding events were more frequently observed with the combination Bevacizumab and Atezolizumab than with Sorafenib (25.2% vs 17.3%), including 7% and 4.5% of gastrointestinal bleeding respectively, and 2.4% and 0.6% of AVB linked to PHT (13). In this study, the patients were well selected, with a majority of patients with controlled viral hepatitis who benefitted from optimal PHT prophylaxis. In series where prophylactic treatment to prevent AVB was less standardized, 10% of patients included in Phase II trials of HCC using bevacizumab presented bleeding complications related to PHT (54). Therefore, the impact of the combination of Bevacizumab and Atezolizumab on PHT remains to be determined. Indeed, even if VEGF contributes to PHT through angiogenesis process, this growth factor is also required for sinusoidal homeostasis. Inhibition of VEGF may impact preserved liver sinusoids in the non-tumoral liver leading to sinusoidal alterations, impairment of oxygen and nutrient supply to hepatocytes causing cell death and the recruitment of inflammatory cells, consequently worsening the underlying liver disease and PHT. In addition, the use of Atezolizumab, which promotes cytotoxic lymphocytes, may also contribute to increase production of proinflammatory cytokines that might activate innate immune cells, as well as HSCs and LSECs, and consequently drive liver fibrosis, chronic inflammation and PHT (89), especially in NASH and alcoholic liver disease where chronic inflammation plays a major role in the progression of the liver disease. Immunotherapy agents

have also been linked to the occurrence of colitis (90). This immuno-mediated side effect might promote bacterial translocation in the setting of cirrhosis and favors chronic inflammation. Thus, promoting inflammation in a setting of diseases where chronic inflammation plays a major role could be also detrimental. Last, but not least, hepatic sinusoidal obstruction syndrome leading to PHT was also noticed after nivolumab treatment, and one can hypothesize that this is also the case with other immunotherapies (91). Hence, further studies are needed to evaluate the impact of the new combination Atezolizumab and Bevacizumab in real life, especially in patients with more severe PHT status. Until then, one can only recommend strict screening and monitoring of PHT, by regular endoscopies, and optimal prophylaxis of AVB.

In conclusion, PHT and HCC are linked in a pathophysiological point of view through angiogenesis process and chronic inflammation, but also in the clinical management of the patients. Combination of PHT and HCC is associated with poorer prognosis in patients with cirrhosis. A better understanding of the crosstalk between these two complications of cirrhosis is mandatory to improve the outcome of these patients, especially in the era of Atezolizumab and Bevacizumab as a first line for advanced HCC.

References

1. Fernandez M. Molecular pathophysiology of portal hypertension. *Hepatology*. 2015;61(4):1406- 15.
2. Lemoine S, Thabut D, Housset C, Moreau R, Valla D, Boulanger CM, et al. The emerging roles of microvesicles in liver diseases. *Nat Rev Gastroenterol Hepatol*. 2014;11(6):350- 61.
3. Thabut D, Shah V. Intrahepatic angiogenesis and sinusoidal remodeling in chronic liver disease: New targets for the treatment of portal hypertension? *J Hepatol*. 2010;53(5):976- 80.
4. Mehta G, Gustot T, Mookerjee RP, Garcia-Pagan JC, Fallon MB, Shah VH, et al. Inflammation and portal hypertension – The undiscovered country. *J Hepatol*. 2014;61(1):155- 63.
5. Thabut D, Massard J, Gangloff A, Carbonell N, Francoz C, Nguyen- Khac E, et al. Model for end-stage liver disease score and systemic inflammatory response are major prognostic factors in patients with cirrhosis and acute functional renal failure. *Hepatology*. 2007;46(6):1872- 82.
6. European Association for the Study of the Liver. Electronic address: easloffice@easloffice.eu, European Association for the Study of the Liver. *EASL Clinical Practice Guidelines: Management of hepatocellular carcinoma*. *J Hepatol*. 2018;69(1):182- 236.
7. Nakayama H, Masuda H, Miyake H, Takayama T, Yokoyama E. Endoscopic Prediction of Hepatocellular Carcinoma by Evaluation of Bleeding Esophageal Varices. *DIG*. 2004;70(4):233- 9.
8. Ripoll C, Groszmann RJ, Garcia-Tsao G, Bosch J, Grace N, Burroughs A, et al. Hepatic venous pressure gradient predicts development of hepatocellular carcinoma independently of severity of cirrhosis. *J Hepatol*. 2009;50(5):923- 8.
9. Giannini EG, Risso D, Testa R, Trevisani F, Di Nolfo MA, Del Poggio P, et al. Prevalence and prognostic significance of the presence of esophageal varices in patients with hepatocellular carcinoma. *Clin Gastroenterol Hepatol*. 2006;4(11):1378- 84.
10. Hsu CY, Lee YH, Huang YH, Hsia CY, Su CW, Lin HC, et al. Ascites in patients with hepatocellular carcinoma: prevalence, associated factors, prognostic impact, and staging strategy. *Hepatol Int*. 2013;7(1):188- 98.
11. Lee YR, Park SY, Tak WY. Treatment Outcomes and Prognostic Factors of Acute Variceal Bleeding in Patients with Hepatocellular Carcinoma. *Gut Liver*. 2020;14(4):500- 8.
12. Lang BH, Poon RT, Fan ST, Wong J. Outcomes of patients with hepatocellular carcinoma presenting with variceal bleeding. *Am J Gastroenterol*. 2004;99(11):2158- 65.
13. Finn RS, Qin S, Ikeda M, Galle PR, Ducreux M, Kim T-Y, et al. Atezolizumab plus Bevacizumab in Unresectable Hepatocellular Carcinoma. *N Engl J Med*. 2020;382(20):1894- 905.
14. Fernández M, Semela D, Bruix J, Colle I, Pinzani M, Bosch J. Angiogenesis in liver disease. *J Hepatol*. 2009;50(3):604- 20.
15. Fernandez M, Vizzutti F, Garcia-Pagan JC, Rodes J, Bosch J. Anti-VEGF receptor-2 monoclonal antibody prevents portal-systemic collateral vessel formation in portal hypertensive mice. *Gastroenterology*. 2004;126(3):886- 94.
16. Fernandez M, Mejias M, Angermayr B, Garcia-Pagan JC, Rodés J, Bosch J. Inhibition of VEGF receptor-2 decreases the development of hyperdynamic splanchnic circulation and portal-systemic collateral vessels in portal hypertensive rats. *J Hepatol*. juill 2005;43(1):98- 103.
17. Carmeliet P. Angiogenesis in life, disease and medicine. *Nature*. 2005;438(7070):932- 6.

18. Ferrara N, Gerber HP, LeCouter J. The biology of VEGF and its receptors. *Nat Med.* 2003;9(6):669- 76.
19. Corpechot C, Barbu V, Wendum D, Kinnman N, Rey C, Poupon R, et al. Hypoxia-induced VEGF and collagen I expressions are associated with angiogenesis and fibrogenesis in experimental cirrhosis. *Hepatology.* 2002;35(5):1010- 21.
20. Semela D, Das A, Langer D, Kang N, Leof E, Shah V. Platelet-Derived Growth Factor Signaling Through Ephrin-B2 Regulates Hepatic Vascular Structure and Function. *Gastroenterology.* 2008;135(2):671-679.e2.
21. Lemoine S, Thabut D, Housset C. Portal myofibroblasts connect angiogenesis and fibrosis in liver. *Cell Tissue Res.* 2016;365(3):583- 9.
22. Lemoine S, Cadoret A, Rautou PE, El Mourabit H, Ratziu V, Corpechot C, et al. Portal myofibroblasts promote vascular remodeling underlying cirrhosis formation through the release of microparticles. *Hepatology.* 2015;61(3):1041- 55.
23. Duscher D, Maan ZN, Wong VW, Rennert RC, Januszyk M, Rodrigues M, et al. Mechanotransduction and fibrosis. *J Biomech.* 2014;47(9):1997- 2005.
24. Liu L, You Z, Yu H, Zhou L, Zhao H, Yan X, et al. Mechanotransduction-modulated fibrotic microniches reveal the contribution of angiogenesis in liver fibrosis. *Nature Materials.* 2017;16(12):1252- 61.
25. Greuter T, Shah VH. Too Stiff, Too Late . . . Timing Is Everything in Antiangiogenic Treatment of Liver Fibrosis. *Hepatology.* 2019;69(1):449- 51.
26. Reiberger T, Angermayr B, Schwabl P, Rohr-Udilova N, Mitterhauser M, Gangl A, et al. Sorafenib attenuates the portal hypertensive syndrome in partial portal vein ligated rats. *J Hepatol.* 2009;51(5):865- 73.
27. Thabut D, Routray C, Lomberk G, Shergill U, Glaser K, Huebert R, et al. Complementary vascular and matrix regulatory pathways underlie the beneficial mechanism of action of sorafenib in liver fibrosis. *Hepatology.* 2011;54(2):573- 85.
28. Pinter M, Sieghart W, Reiberger T, Rohr-Udilova N, Ferlitsch A, Peck-Radosavljevic M. The effects of sorafenib on the portal hypertensive syndrome in patients with liver cirrhosis and hepatocellular carcinoma--a pilot study. *Aliment Pharmacol Ther.* 2012;35(1):83- 91.
29. Ma R, Chen J, Liang Y, Lin S, Zhu L, Liang X, et al. Sorafenib: A potential therapeutic drug for hepatic fibrosis and its outcomes. *Biomed Pharmacother.* 2017;88:459- 68.
30. Yang L, Kwon J, Popov Y, Gajdos GB, Ordog T, Brekken RA, et al. Vascular endothelial growth factor promotes fibrosis resolution and repair in mice. *Gastroenterology.* 2014;146(5):1339-1350.e1.
31. Neugebauer H, Hartmann P, Krenn S, Glück T, Schölmerich J, Straub R, et al. Bacterial translocation increases phagocytic activity of polymorphonuclear leucocytes in portal hypertension: priming independent of liver cirrhosis. *Liv Int.* 2008;28(8):1149- 57.
32. Sersté T, Bourgeois N, Lebecq D, Evrard S, Devière J, Le Moine O. Relationship between the degree of portal hypertension and the onset of spontaneous bacterial peritonitis in patients with cirrhosis. *Acta Gastroenterol Belg.* 2006;69(4):355- 60.
33. Chen Y, Ji F, Guo J, Shi D, Fang D, Li L. Dysbiosis of small intestinal microbiota in liver cirrhosis and its association with etiology. *Scientific Reports.* 2016;6(1):34055.
34. Wiest R, Lawson M, Geuking M. Pathological bacterial translocation in liver cirrhosis. *J Hepatol.* 2014;60(1):197- 209.
35. Akira S, Takeda K. Toll-like receptor signalling. *Nature Reviews Immunology.* 2004;4(7):499- 511.
36. Schwabe RF, Seki E, Brenner DA. Toll-like receptor signaling in the liver. *Gastroenterology.* 2006;130(6):1886- 900.

37. Gracia-Sancho J, Lavina B, Rodriguez-Vilarrupla A, Garcia-Caldero H, Fernandez M, Bosch, et al. Increased oxidative stress in cirrhotic rat livers: A potential mechanism contributing to reduced nitric oxide bioavailability. *Hepatology*. 2008;47(4):1248-56.
38. Wheeler MD, Kono H, Yin M, Nakagami M, Uesugi T, Arteel GE, et al. The role of Kupffer cell oxidant production in early ethanol-induced liver disease. *Free Radic Biol Med*. 2001;31(12):1544- 9.
39. Seki E, De Minicis S, Osterreicher CH, Kluwe J, Osawa Y, Brenner DA, et al. TLR4 enhances TGF-beta signaling and hepatic fibrosis. *Nat Med*. 2007;13(11):1324- 32.
40. Zhu Q, Zou L, Jagavelu K, Simonetto DA, Huebert RC, Jiang ZD, et al. Intestinal decontamination inhibits TLR4 dependent fibronectin-mediated cross-talk between stellate cells and endothelial cells in liver fibrosis in mice. *J Hepatol*. 2012;56(4):893- 9.
41. Kemp W, Colman J, Thompson K, Madan A, Vincent M, Chin-Dusting J, et al. Norfloxacin treatment for clinically significant portal hypertension: results of a randomised double-blind placebo-controlled crossover trial. *Liver Int*. 2009;29(3):427- 33.
42. Mendoza YP, Rodrigues SG, Bosch J, Berzigotti A. Effect of poorly absorbable antibiotics on hepatic venous pressure gradient in cirrhosis: A systematic review and meta-analysis. *Dig Liver Dis*. 2020;52(9):958- 65.
43. Mookerjee RP, Sen S, Davies NA, Hodges SJ, Williams R, Jalan R. Tumour necrosis factor α is an important mediator of portal and systemic haemodynamic derangements in alcoholic hepatitis. *Gut*. 2003;52(8):1182- 7.
44. Cao G, Li X, Qin C, Li J. Prognostic Value of VEGF in Hepatocellular Carcinoma Patients Treated with Sorafenib: A Meta-Analysis. *Med Sci Monit*. 2015;21:3144- 51.
45. Xiong XX, Qiu XY, Hu DX, Chen XQ. Advances in Hypoxia-Mediated Mechanisms in Hepatocellular Carcinoma. *Mol Pharmacol*. 2017;92(3):246- 55.
46. Horwitz E, Stein I, Andreozzi M, Nemeth J, Shoham A, Pappo O, et al. Human and mouse VEGFA-amplified hepatocellular carcinomas are highly sensitive to sorafenib treatment. *Cancer Discov*. 2014;4(6):730- 43.
47. Schmitt M, Horbach A, Kubitz R, Frilling A, Häussinger D. Disruption of hepatocellular tight junctions by vascular endothelial growth factor (VEGF): a novel mechanism for tumor invasion. *J Hepatol*. 2004;41(2):274- 83.
48. Palazon A, Tyrakis PA, Macias D, Veliça P, Rundqvist H, Fitzpatrick S, et al. An HIF-1 α /VEGF-A Axis in Cytotoxic T Cells Regulates Tumor Progression. *Cancer Cell*. 2017;32(5):669-683.e5.
49. Fukumura D, Kloepper J, Amoozgar Z, Duda DG, Jain RK. Enhancing cancer immunotherapy using antiangiogenics: opportunities and challenges. *Nat Rev Clin Oncol*. 2018;15(5):325- 40.
50. Gabrilovich D, Ishida T, Oyama T, Ran S, Kravtsov V, Nadaf S, et al. Vascular endothelial growth factor inhibits the development of dendritic cells and dramatically affects the differentiation of multiple hematopoietic lineages in vivo. *Blood*. 1998;92(11):4150- 66.
51. Kim CG, Jang M, Kim Y, Leem G, Kim KH, Lee H, et al. VEGF-A drives TOX-dependent T cell exhaustion in anti-PD-1-resistant microsatellite stable colorectal cancers. *Sci Immunol*. 2019;4(41).
52. Huang Y, Yuan J, Righi E, Kamoun WS, Ancukiewicz M, Nezivar J, et al. Vascular normalizing doses of antiangiogenic treatment reprogram the immunosuppressive tumor microenvironment and enhance immunotherapy. *Proc Natl Acad Sci U S A*. 2012;109(43):17561- 6.
53. Huang Y, Chen X, Dikov MM, Novitskiy SV, Mosse CA, Yang L, et al. Distinct roles of VEGFR-1 and VEGFR-2 in the aberrant hematopoiesis associated with elevated levels of VEGF. *Blood*. 2007;110(2):624- 31.
54. Fang P, Hu J, Cheng Z, Liu Z, Wang J, Jiao S. Efficacy and Safety of Bevacizumab

for the Treatment of Advanced Hepatocellular Carcinoma: A Systematic Review of Phase II Trials. *PLoS One*. 2012;7(12):e49717.

55. Zhu AX, Kang Y-K, Yen C-J, Finn RS, Galle PR, Llovet JM, et al. Ramucirumab after sorafenib in patients with advanced hepatocellular carcinoma and increased α -fetoprotein concentrations (REACH-2): a randomised, double-blind, placebo-controlled, phase 3 trial. *The Lancet Oncology*. 2019;20(2):282- 96.

56. He G, Dhar D, Nakagawa H, Font-Burgada J, Ogata H, Jiang Y, et al. Identification of liver cancer progenitors whose malignant progression depends on autocrine IL-6 signaling. *Cell*. 2013;155(2):384- 96.

57. Pavlovic N, Rani B, Gerwins P, Heindryckx F. Platelets as Key Factors in Hepatocellular Carcinoma. *Cancers*. 2019;11(7):1022.

58. Malehmir M, Pfister D, Gallage S, Szydlowska M, Inverso D, Kotsiliti E, et al. Platelet GPIIb/IIIa is a mediator and potential interventional target for NASH and subsequent liver cancer. *Nature Medicine*. 2019;25(4):641- 55.

59. Nan J, Xing YF, Hu B, Tang J-X, Dong H-M, He Y-M, et al. Endoplasmic reticulum stress induced LOX-1+ CD15+ polymorphonuclear myeloid-derived suppressor cells in hepatocellular carcinoma. *Immunology*. 2018;154(1):144- 55.

60. Deng Y, Cheng J, Fu B, Liu W, Chen G, Zhang Q, et al. Hepatic carcinoma-associated fibroblasts enhance immune suppression by facilitating the generation of myeloid-derived suppressor cells. *Oncogene*. 2017;36(8):1090- 101.

61. Chen DS, Mellman I. Oncology meets immunology: the cancer-immunity cycle. *Immunity*. 2013;39(1):1- 10.

62. Finn RS, Ryoo BY, Merle P, Kudo M, Bouattour M, Lim HY, et al. Pembrolizumab As Second-Line Therapy in Patients With Advanced Hepatocellular Carcinoma in KEYNOTE-240: A Randomized, Double-Blind, Phase III Trial. *J Clin Oncol*. 2020;38(3):193- 202.

63. El-Khoueiry AB, Sangro B, Yau T, Crocenzi TS, Kudo M, Hsu C, et al. Nivolumab in patients with advanced hepatocellular carcinoma (CheckMate 040): an open-label, non-comparative, phase 1/2 dose escalation and expansion trial. *Lancet*. 2017;389(10088):2492- 502.

64. de Franchis R, Baveno VI Faculty. Expanding consensus in portal hypertension: Report of the Baveno VI Consensus Workshop: Stratifying risk and individualizing care for portal hypertension. *J Hepatol*. 2015;63(3):743- 52.

65. Allaire M, Goumard C, Lim C, Le Cleach A, Wagner M, Scatton O. New frontiers in liver resection for hepatocellular carcinoma. *JHEP Reports*. 2020;2(4):100134.

66. Allaire M, Rekik S, Layese R, Mumana A, Guyot E, Nkontchou G, et al. Virologic control and severity of liver disease determine survival after radiofrequency ablation of hepatocellular carcinoma on cirrhosis. *Dig Liver Dis*. 2019;51(1):86- 94.

67. Choi JW, Chung JW, Lee DH, Kim H-C, Hur S, Lee M, et al. Portal hypertension is associated with poor outcome of transarterial chemoembolization in patients with hepatocellular carcinoma. *Eur Radiol*. 2018;28(5):2184- 93.

68. Ripoll C, Genescà J, Araujo IK, Graupera I, Augustin S, Tejedor M, et al. Rebleeding prophylaxis improves outcomes in patients with hepatocellular carcinoma. A multicenter case-control study. *Hepatology*. 2013;58(6):2079- 88.

69. Bureau C, Garcia-Pagan JC, Ota P, Pomier-Layrargues G, Chabbert V, Cortez C, et al. Improved clinical outcome using polytetrafluoroethylene-coated stents for TIPS: results of a randomized study. *Gastroenterology*. 2004;126(2):469- 75.

70. Rudler M, Mallet M, Sultanik P, Bouzbib C, Thabut D. Optimal management of ascites. *Liver International*. 2020;40(S1):128- 35.

71. Nicoară-Farcău O, Han G, Rudler M, Angrisani D, Monescillo A, Torres F, et al.

Effects of Early Placement of Transjugular Portosystemic Shunts in Patients With High-Risk Acute Variceal Bleeding: a Meta-analysis of Individual Patient Data. *Gastroenterology*. 2021;160(1):193-205.e10.

72. Allaire M, Walter A, Sutter O, Nahon P, Ganne-Carrié N, Amathieu R, et al. TIPS for management of portal-hypertension-related complications in patients with cirrhosis. *Clin Res Hepatol Gastroenterol*. 2020;44(3):249-63.

73. García-Pagán JC, Caca K, Bureau C, Laleman W, Appenrodt B, Luca A, et al. Early use of TIPS in patients with cirrhosis and variceal bleeding. *N Engl J Med*. 2010;362(25):2370- 9.

74. Kuo YC, Kohi MP, Naeger DM, Tong RT, Kolli KP, Taylor AG, et al. Efficacy of TACE in TIPS Patients: Comparison of Treatment Response to Chemoembolization for Hepatocellular Carcinoma in Patients With and Without a Transjugular Intrahepatic Portosystemic Shunt. *Cardiovasc Intervent Radiol*. 2013;36(5):1336-43.

75. Padia SA, Chewing RH, Kogut MJ, Ingraham CR, Johnson GE, Bhattacharya R, et al. Outcomes of Locoregional Tumor Therapy for Patients with Hepatocellular Carcinoma and Transjugular Intrahepatic Portosystemic Shunts. *Cardiovasc Intervent Radiol*. 2015;38(4):913- 21.

76. Kohi MP, Fidelman N, Naeger DM, LaBerge JM, Gordon RL, Kerlan RK. Hepatotoxicity after transarterial chemoembolization and transjugular intrahepatic portosystemic shunt: do two rights make a wrong? *J Vasc Interv Radiol*. 2013;24(1):68- 73.

77. Wang Z, Zhang H, Zhao H, Wang X, Tsao J, Luo X, et al. Repeated transcatheter arterial chemoembolization is safe for hepatocellular carcinoma in cirrhotic patients with transjugular intrahepatic portosystemic shunt. *Diagn Interv Radiol*. 2014;20(6):487- 91.

78. Llovet JM, Ricci S, Mazzaferro V, Hilgard P, Gane E, Blanc JF, et al. Sorafenib in Advanced Hepatocellular Carcinoma. *N Engl J Med*. 2008;359(4):378- 90.

79. Bruix J, Qin S, Merle P, Granito A, Huang YH, Bodoky G, et al. Regorafenib for patients with hepatocellular carcinoma who progressed on sorafenib treatment (RESORCE): a randomised, double-blind, placebo-controlled, phase 3 trial. *Lancet*. 2017;389(10064):56- 66.

80. Kudo M, Finn RS, Qin S, Han KH, Ikeda K, Piscaglia F, et al. Lenvatinib versus sorafenib in first-line treatment of patients with unresectable hepatocellular carcinoma: a randomised phase 3 non-inferiority trial. *Lancet*. 2018;391(10126):1163- 73.

81. Abou-Alfa GK, Meyer T, Cheng A-L, El-Khoueiry AB, Rimassa L, Ryoo B-Y, et al. Cabozantinib in Patients with Advanced and Progressing Hepatocellular Carcinoma. *New England Journal of Medicine*. 2018;379(1):54- 63.

82. Tugues S, Fernandez-Varo G, Muñoz-Luque J, Ros J, Arroyo V, Rodés J, et al. Antiangiogenic treatment with sunitinib ameliorates inflammatory infiltrate, fibrosis, and portal pressure in cirrhotic rats. *Hepatology*. 2007;46(6):1919- 26.

83. Uschner FE, Schueller F, Nikolova I, Klein S, Schierwagen R, Magdaleno F, et al. The multikinase inhibitor regorafenib decreases angiogenesis and improves portal hypertension. *Oncotarget*. 2018;9(90):36220- 37.

84. O Ogawa H, Kaji K, Nishimura N, Takagi H, Ishida K, Takaya H, et al. Lenvatinib prevents liver fibrosis by inhibiting hepatic stellate cell activation and sinusoidal capillarization in experimental liver fibrosis. *J Cell Mol Med*. 2021; doi: 10.1111/jcmm.16363.

85. Hidaka H, Nakazawa T, Kaneko T, Minamino T, Takada J, Tanaka Y, et al. Portal hemodynamic effects of sorafenib in patients with advanced hepatocellular carcinoma: a prospective cohort study. *J Gastroenterol*. 2012;47(9):1030- 5.

86. Hidaka H, Uojima H, Nakazawa T, Shao X, Hara Y, Iwasaki S, et al. Portal hemodynamic effects of lenvatinib in patients with advanced hepatocellular carcinoma: A prospective cohort study. *Hepatol Res*. 2020;50(9):1083- 90.

87. Iavarone M, Primignani M, Vavassori S, Sangiovanni A, La Mura V, Romeo R, et al. Determinants of esophageal varices bleeding in patients with advanced hepatocellular carcinoma treated with sorafenib. *United European Gastroenterol J.* 2016;4(3):363- 70.
88. Colecchia A, Ravaioli F, Marasco G, Colli A, Dajti E, Di Biase AR, et al. A combined model based on spleen stiffness measurement and Baveno VI criteria to rule out high-risk varices in advanced chronic liver disease. *J Hepatol.* 2018;69(2):308- 17.
89. Honma Y, Shibata M, Gohda T, Matsumiya H, Kumamoto K, Miyama A, et al. Rapid Progression of Liver Fibrosis Induced by Acute Liver Injury Due to Immune-related Adverse Events of Atezolizumab. *Intern Med.* 2021; doi: 10.2169/internalmedicine.6535-20.
90. Miyashita H, Mikami T, Satoi S, Cruz C, Galsky MD. Incidence and Risk of Colitis With Programmed Death 1 Versus Programmed Death Ligand 1 Inhibitors for the Treatment of Cancer. *J Immunother.* 2020;43(9):291- 8.
91. Riveiro-Barciela M, González-Sans D, Marmolejo D, Salcedo M-T, Muñoz-Couselo E. Hepatic sinusoidal obstruction syndrome associated with nivolumab: an uncommon adverse event related to immune checkpoint inhibitors. *J Gastrointest Liver Dis.* 2021;30(1):171- 2.
92. Yau T, Park JW, Finn RS, Cheng A-L, Mathurin P, Edeline J, et al. LBA38_PR - CheckMate 459: A randomized, multi-center phase III study of nivolumab (NIVO) vs sorafenib (SOR) as first-line (1L) treatment in patients (pts) with advanced hepatocellular carcinoma (aHCC). *Annals of Oncology.* 2019;30:v874- 5.

Figure 1. Impact of increased vascular endothelial growth factor (VEGF) level on portal hypertension and hepatocellular occurrence

(A) In physiological condition, VEGF plays an important role in cellular homeostasis, especially to maintain sinusoidal capillary network homeostasis; (B) Inflammation, shear and oxidative stress within the liver induced by liver injury increase VEGF production by hepatocytes and HSCs. The augmentation of VEGF levels will favor fibrosis through activation of HSCs and angiogenesis through activation of LSECs that will increase intra hepatic resistance contributing to portal hypertension occurrence. Chronic inflammation favors by bacterial translocation also contribute to the phenomena. An increase of VEGF within the mesenteric vascular bed favors angiogenesis and lead to the creation of portal systemic collateral vessel, contributing to the increase splanchnic blood flow and portal hypertension; (C) With time, VEGF will promote hepatocytes proliferation and angiogenesis participating to the pathophysiology of hepatocellular carcinoma genesis. Chronic inflammation will also promote cell proliferation and creates a tumor microenvironment that supports the transformation of hepatocytes, and also induces their survival through activation of anti-apoptotic pathway and inhibition of immune surveillance. VEGF also mediates immunosuppression within the tumor microenvironment by promoting immunosuppressive cells such as Treg, MDSCs and TAMs and suppressing antigen suppressive cells and cytotoxic T lymphocyte.

LSECs: liver sinusoidal endothelial cells, HCC: hepatocellular carcinoma; HSCs: hepatic stellate cells, MDSCs: myeloid derived suppressive cells, TAM: tumor associated macrophages, TReg: regulatory T cells, VEGF: vascular endothelial growth factor

Figure 2. TIPS placement in patients with HCC.

TIPS placement in well selected patients with HCC may be beneficial as TIPS increases survival (overall survival and transplant free survival in the setting of variceal bleeding and recurrent/refractory ascites). In addition, TIPS may allow to perform locoregional HCC treatment by modeling hypertension portal level and improving liver function in order to access to liver transplant. However, the decision of TIPS placement should consider a potential risk of liver dysfunction that may prevent patients to access to HCC treatments, but also a potential risk of HCC spread especially in case of central localization and vascular invasion and the risk of less effective transarterial HCC treatment.

Table 1. Targets of HCC systemic therapy and impact on portal hypertension

Systemic treatment	Target of Systemic treatment	Effect on portal hypertension	
Tyrosine kinase inhibitors		Mice models	Humans' data
Sorafenib	VEGFR-2, VEGFR-3 PDGFR-β CRAF, BRAF, V600E BRAF c-KIT, FLT-3	↓ Angiogenesis (26,27)	↓ PHT (28,85)
Lenvatinib	VEGFR-1, VEGFR-2, VEGFR-3 PDGFR, c-KIT, RET FGFR-1, FGFR-2, FGFR-3, FGFR-4,	↓ Angiogenesis (84)	↑ PHT? (86)
Regorafenib	VEGFR-1, VEGFR-2, VEGFR-3 PDGFR, TIE2, FGFR c-KIT, RET, RAF-1, BRAF, V600E BRAF	↓ Angiogenesis (83)	No data
Cabozantinib	VEGFR-2, c-MET TIE2, AXL, RET, ROS1, TYR03, MER, KIT, TRKB, FLT3	No data	No data
Monoclonal antibody		Mice models	Human data
Bevacizumab	VEGF	No data	No data
Ramucirumab	VEGFR-2	No data	No data
Immunotherapy		Mice models	Human data
Atezolizumab	PD-L1	No data	No data
Nivolumab	PD1	No data	No data
Pembrolizumab	PD1	No data	No data

PDGFR: VEGF: vascular endothelial growth factor; VEGFR: vascular endothelial growth factor receptor; PHT: portal hypertension;

Table 2. Bleeding events related to portal hypertension in phase 3 systemic treatment studies for advanced hepatocellular carcinoma

Study	Design of the study	Exclusion criteria regarding bleeding events	Bleeding events related to PHT
First line treatments			
Llovet et al. NEJM 2008 (78)	SHARP Study Phase 3 RCT Sorafenib (n=299) vs Placebo (n=303)	Gastrointestinal bleeding within 30 days prior to randomization	Sorafenib (2.69%) Placebo (4.3%)
Kudo et al. Lancet 2018 (80)	REFLECT study Phase 3 RCT Sorafenib (n=476) vs Lenvatinib (n=478)	Gastrointestinal bleeding and interventional treatment for varices within 28 days prior to randomization Antiplatelet agents were prohibited Thrombotic disorders or use of anticoagulants requiring therapeutic INR monitoring	Sorafenib (1.05%) Lenvatinib (1.47%)
Finn et al. NEJM 2020 (13)	IMBRAVE 150 study Phase 3 RCT Atezolizumab/Bevacizumab (n=336) vs Sorafenib (n=165)	A prior bleeding event due to PHT within 6 months prior to randomization Untreated or incompletely treated esophageal and/or gastric varices with bleeding or high-risk for bleeding	Atezolizumab/Bevacizumab (2.6%) Sorafenib (0.4%)
Yau et al. Ann of Oncol 2020 (92)	CHECKMATE 459 Phase 3 RCT Nivolumab (n=371) vs Sorafenib (n=372)	No specific exclusion criteria regarding bleeding history	Nivolumab (1.09%) Sorafenib (1.93%)
Second line treatments			
Bruix et al. Lancet 2017 (79)	RESORCE study Phase 3 RCT Regorafenib (n=379) vs Placebo (n=194)	Gastrointestinal bleeding within 30 days prior to randomization	Regorafenib (1.60%) Placebo (0.52%)
Abou-alfa et al. NEJM 2018 (81)	CELESTIAL Study Phase 3 RCT Cabozantinib (n=470) vs Placebo (n=237)	Subjects with untreated or incompletely treated varices with bleeding or high risk for bleeding Concomitant anticoagulation, at therapeutic doses, with anticoagulants	Cabozantinib (1.71%) Placebo (2.52%)
Zhu et al. Lancet 2019 (55)	REACH-2 Study Phase 3 RCT Ramucirumab (n=470) vs Placebo (n=237)	Any bleeding episode considered life-threatening, or any Grade 3 or 4 gastrointestinal bleeding episode in the 3 months prior to randomization Esophageal or gastric varices that require intervention or represent high bleeding risk.	Ramucirumab (1.02%) Placebo (1.05%)
Finn et al. J Clin Oncol 2020 (62)	KEYNOTE-240 Study Phase 3 RCT Pembrolizumab (n=279) vs Placebo (n=134)	Esophageal or gastric variceal bleeding within the last 6 months	Pembrolizumab (1.08%) Placebo (0.75%)

*Data obtained on clinical trial.gov

INR: international normalized ratio; PHT: portal hypertension; RCT: randomized-controlled trial