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Original Article

Notes on the biological development of the darkling beetle *Blaps nefrauensis nefrauensis* Seidlitz, 1893 (Coleoptera: Tenebrionidae)

Notas sobre o desenvolvimento biológico do besouro escuro *Blaps nefrauensis nefrauensis* Seidlitz, 1893 (Coleoptera: Tenebrionidae)

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Abstract

Several endemic species of *Blaps* occur in Tunisia, and the species *Blaps nefrauensis nefrauensis* has been reported in Moulares (urban zone in west-central Tunisia), where it lives and reproduces in home gardens and old buildings. The aim of this work is to study the life cycle of the darkling beetle, considering both field and laboratory rearing conditions. As a result, the beetle species has different developmental stages (egg, larva, prepupa, pupa, and adult) that last about 15 months. Each year during the same period, adults emerge (early summer) and expire (late autumn), larvae hatch (late summer) and pupate (early summer). There is only one generation per year. Females began laying eggs in late July. The eggs were ovoid, white, and about 2.7 mm in length and 1.5 mm in width. Embryogenesis took an average of nine days. The first instar larvae were at initially only 4.5 mm long and ivory white in color. A brief description of the newly egg hatched larva was provided; thus, the nerve fibers innervating the apical setae in the antennae and ligula were detected. Further light microscopic examination of the embryo before hatching from the egg pointed out that the antennal sensilla are protected during the embryogenesis stage.

Keywords: *Blaps nefrauensis nefrauensis*, life cycle stages, first instar larvae, larval antennae, larval ligula.

Resumo

Várias espécies endêmicas de *Blaps* ocorrem na Tunísia, e a espécie *Blaps nefrauensis nefrauensis* foi relatada em Moulares (zona urbana no centro-oeste da Tunísia), onde vive e se reproduz em jardins domésticos e prédios antigos. O objetivo deste trabalho é estudar o ciclo de vida do besouro escuro, considerando as condições de criação em campo e em laboratório. Como resultado, a espécie de besouro tem diferentes estágios de desenvolvimento (ovo, larva, prepupa, pupa e adulto) que duram cerca de 15 meses. Todos os anos, durante o mesmo período, os adultos emergem (início do verão) e expiram (final do outono), as larvas eclodem (final do verão) e se tornam pupas (início do verão). Existe apenas uma geração por ano. As fêmeas começaram a botar ovos no final de julho. Os ovos eram ovóides, brancos, com cerca de 2,7 mm de comprimento e 1,5 mm de largura. A embriogênese demorou em média nove dias. As larvas de primeiro instar tinham inicialmente apenas 4,5 mm de comprimento e uma cor branca marfim. Foi fornecida uma breve descrição da larva recém-eclodida; assim, as fibras nervosas que inervam as cerdas apicais nas antenas e ligulas foram detectadas. Um exame microscópico de luz posterior do embrião antes da eclosão do ovo mostrou que as sensilas antenais são protegidas durante o estágio de embriogênese.

Palavras-chave: *Blaps nefrauensis nefrauensis*, estágios do ciclo de vida, larvas de primeiro instar, antenas larvais, lígula larval.

1. Introduction

Tenebrionidae is a highly diverse family of Coleoptera is divided into ten subfamilies, 96 tribes, 61 subtribes, 2300 genera, and about 20000 species distributed worldwide (Löbl et al., 2008; Matthews et al., 2010; Ślipiński et al., 2011).

The subfamily Tenebrioninae Latreille, 1802 consists of 29 tribes in which the tribe Blaptini Leach, 1815 includes five subtribes (Blaptina Leach, 1815, Gnaptorina Medvedev, 2001, Gnaptorinina Medvedev, 2001, Prosodina Skopin,

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1960, Remipedellina Semenov, 1907). The largest in the subtribe Blaptina is the genus *Blaps* Fabricius, 1775 (type genus) which encompasses more than 250 species. It is an emblematic group of large flightless beetles adapted to arid and semi-arid terrestrial ecosystems. With about 230 species, the subgenus *Blaps* is the most diverse of the four extant subgenera (*Arenoblaps* Medvedev, *Blaps* Fabricius, *Dineria* Motschnhulsky and *Prosoblapsia* Skopin and Kaszab) (Löbl et al., 2008). Within the subgenus *Blaps* about 140 species are Eurasian, whereas about 90 species mostly occur in the Mediterranean pool (Soldati et al., 2017). In Tunisia (North Africa), 17 taxa (species and subspecies belonging to the subgenus *Blaps*) at least were listed (*B. appendiculata* Motschulsky, 1851, *B. approximans* Seidlitz, 1893 ; *B. binominata* Escalera, 1914, *B. divergens* Fairmaire, 1875, *B. emondi* Solier, 1848, *B. bifurcata strauchii* Reiche, 1861b: 88, *B. gigas* Linnaeus, 1767, *B. nefrauensis nefrauensis* Seidlitz, 1893, *B. nitens nitens* Laporte, 1840, *B. nitens praedeserta* Koch, 1944, *B. nitens requieni* Solier, 1848, *B. nitiduloides* L. Soldati, 2017, *B. plana* Solier, 1848, *B. propheta propheta* Reiche, 1861, *B. pubescens* Allard, 1880, *B. superstitiosa superstitiosa* Erichson, 1841) (Bouchard et al., 2005; Löbl et al., 2008; Condamine et al., 2011, 2013; Soldati et al., 2017).

Among the coleopteran species reported in Tunisia, *Blaps nefrauensis nefrauensis* Seidlitz, 1893, occurred in Gafsa (Soldati et al., 2017). This desert darkling beetle was widespread in summer in Moulares (Gafsa), invading local gardens and ruins, where it mates, feeds and lays its eggs. This makes it useful as a model organism to study the life cycle of the *Blaps* species. Knowing that the life histories of the tenebrionid species of North Africa have received little attention to date. (Halstead, 1967; Menon and Putnam, 1988; De Los Santos et al., 1988, 2006; Zanuncio et al., 2001). Therefore, the studies conducted in this zoogeography focused mainly on the biogeography and taxonomy of beetles. On the other hand, first instar larvae in *Blaps* species were small and white transparent, so that more details could be seen and investigated under the light microscope.

The present work was a contribution to know more about the life cycle of *B. nefrauensis* and to provide notes on the first instar larvae, especially on the innervations of the ligular and antennal setae. This may be evaluated for use with other species of the *Blaps* genus or Blabtni tribe.

2. Material and Methods

2.1. Identification

The identification of the specimens was based on diagnostic keys and characteristics from the literature (original descriptions): (Seidlitz, 1893; Normand, 1937), and in comparison with the *Blaps* species illustrated in the following work by Laurent Soldati and Fabian Condamine (Western Palearctic tenebrionid specialists): (Soldati et al., 2009, 2017; Condamine et al., 2011, 2013).

2.2. Field and laboratory investigation

The different life cycle stages of *B. nefrauensis* were followed in a domestic garden in Moulares center

(34°29'26.30"N, 8°16'19.95"E) (Figure 1) during three years (from June 2017 to June 2020), where the beetles mate, oviposit and feed while exploring its environment (decomposing plant matter, dead insect, human food waste, wet soil, stones, boards, concealed shelters...), they were simultaneously observed in the rearing laboratory. Field searches for adult *Blaps* and sampling were conducted fortnightly at night from early May to late December each year for three years. Beetles were counted at each sampling and most were returned to the field after having measured their size (♂ and ♀). About Five pairs of beetles were taken for breeding at each sampling. Field searches for eggs, larvae, prepupae and pupae were carried out every week throughout the year. Mating was monitored both in the field and during laboratory rearing. Larval molting was monitored weekly in laboratory (daily for the first weeks of larval age). Considering that it was difficult to detect egg hatch in the field, some copulating pairs of beetles were placed in buckets (40 cm diameter 50 cm depth) in the breeding laboratory (at room temperature away from wind, rain, and other climatic factors). About one-quarter of the buckets were filled with garden soil. A part of the soil was covered with cement planks to maintain humidity, and soil was sprinkled with water every ten days. Decomposing plant matter, cereals, and occasionally fruits were used as a source of food. From late summer to late fall, laid eggs were collected from soil and counted every three days. Eggs were placed in Petri dishes lined with damp filter papers at a temperature fluctuating between 25 to 30 °C where hatching time and hatched eggs were daily recorded. To monitor the first mating of beetles and their beginning of oviposition, some of tens of newly emerged adults (male and female) carried out in rearing laboratory were daily observed especially after sunset. Prior to that, the size (length and width) of beetles was measured. As they were small and ivory transparent in color, some newly egg hatched larvae were observed under light microscopy; it is there that the antennal and ligula setae innervation have been investigated.

3. Results

Blaps nefrauensis is one of about 17 desert darkling beetles (*Blaps* genus) occurring in Tunisia. Living in urban zones (Moulares, Tunisia), it is more accessible to study its biological development and mating behavior as it inhabits major domestic gardens where it mates, oviposits and feeds. A field survey combined with the laboratory rearing can probably provide a more clear idea about the life history (egg, larva, prepupa, pupae, and imago ♂ and ♀) of this coleopteran species (Figures 2, 3a-f).

3.1. Life cycle

3.1.1. Emergence and imago

In Moulares (urban zone in the west south of Tunisia), *B. nefrauensis* populations occurred generally in the domestic gardens. Adults appeared in early June. The majority imago emerges from soil burrowing and



Figure 1. Map of Tunisia (North Africa) showing the localization of the sampling area.

leaving tunnels about 2 cm in diameter and 30 cm in depth. Males emerged smaller in size than females (mean values: 4 cm in length and 1.5 cm in width vs. 4.5 cm in length and 1.8 cm in width for males and females, respectively). In general, males and females were sexually dimorphic: Females are larger in size than males. Males have abdominal intersternite hair tuft and longer caudal extensions (mucron) (Figure 2a-b). Both males and females began to mate about one week after their emergence (June) and continue mating throughout their lifetime. The mating period starts from June to November peaking in August when beetles are most evident. At night (from dusk till dawn), the majority of pairs adopted almost the same sexual behavior. The male approaches the female, mounts, and remains mounted for up to thirty minutes. During this time female extrudes her ovipositor out of her abdomen (Figure 2c) to smooth the insertion and facilitate the fixation of male genitalia on the female vagina (Figure 2d) during more than five minutes. Consequently, male completely extrudes spermatophores inside female abdomen during copulation.

3.1.2. Oviposition and egg hatching

In the field, females began the oviposition from late July to late November, peaking in September. They lay their eggs in soil covered by decomposition matter, under stones, and under cement and wooden planks. Because of the high temperature and low humidity, embryogenesis can really be accomplished only from mid-August. Laboratory conditions show that females dig in the soil (2-40 mm) and lay eggs scattered (soil moisture and softness may help females dig deep into the soil and lay their eggs). Females lay between 1 to 6 eggs per day during summer and autumn seasons. In the breeding laboratory females stay alive, but give up laying eggs. Eggs were ovoid in form, white in color and approximately 2.7 mm in length and 1.5 mm in width. The duration of eggs development till hatching was around 9 days for the majority. New hatching larvae (1st instar) were transparent white in color and 4.5 mm in length (Figure 3a).

3.1.3. Larvae and pupation

As well-known, species belonging to order Coleoptera present a complete metamorphosis involving several



Figure 2. Morphological dimorphism and mating in *B. nefrauensis*. (a) Male ventral view, (b) Female ventral view, (c) Female ovipositor, (d) Male genitalia.

larval instars. These instars are followed by a non-feeding pupal stage, which gives rise directly to the adult. An active survey in the laboratory combined with a field examination showed that, in *B. nefrauensis* populations, the number of instars intervening between the egg and the pupa ranged from 14 to 16. Figure 3g-j can give an idea about the moulting phenomena in the *Blaps* larva. As shown in Figure 3a-f, the larval stage began with the newly hatched larva (hereafter, first instar). First larval instar was white in color and very small, a little bit longer than an egg (Figure 3a). Within three days approximately, it changes gradually to yellowish giving the second larval instar (Figure 3b) which is clearly different in size (7 mm vs. 4.5 mm long in mean) and color (shiny yellow). In the field, larvae in different instars were hidden under stones and planks or inside their borrowed tunnels. Larvae began to appear in the garden at the end of August. The majority of *B. nefrauensis* larvae moulted between 14 and 16 times. In every inter-moulting period, larvae become white in color, leaving its old yellow cuticle (they turn yellow in color within two days on average. Figure 3g-j showed

some of moulting larvae in different instar stages. In mid-May, most of the larvae were in the last instar (Figure 3c) reaching a constant size (~6 cm). They enlarge the external part of their tunnels and enter in to the prepupal stage (Figure 3d). This was a semi-dormant and non-feeding stage in which the prepupae shortened in size and lost weight. After about four days, they pupate on the enlarged base of the tunnels. At the population level, pupation occurred from middle May to mid-June. Pupa appears to be white in color (Figure 2e) lasting about 12 days in that non-feeding stage. In June, adults (Figure 3f) emerge from the soil, fulfill their biological functions during summer and autumn and die before winter leaving their larvae in the field as a new generation.

3.1.4. Summary

Blaps nefrauensis is usually found at night around human waste food, and near plant and stones. In an urban area it has been associated with domestic gardens. Because of its North Africa origin (Soldati et al., 2017), adults were well suited with summer climate (high temperature and

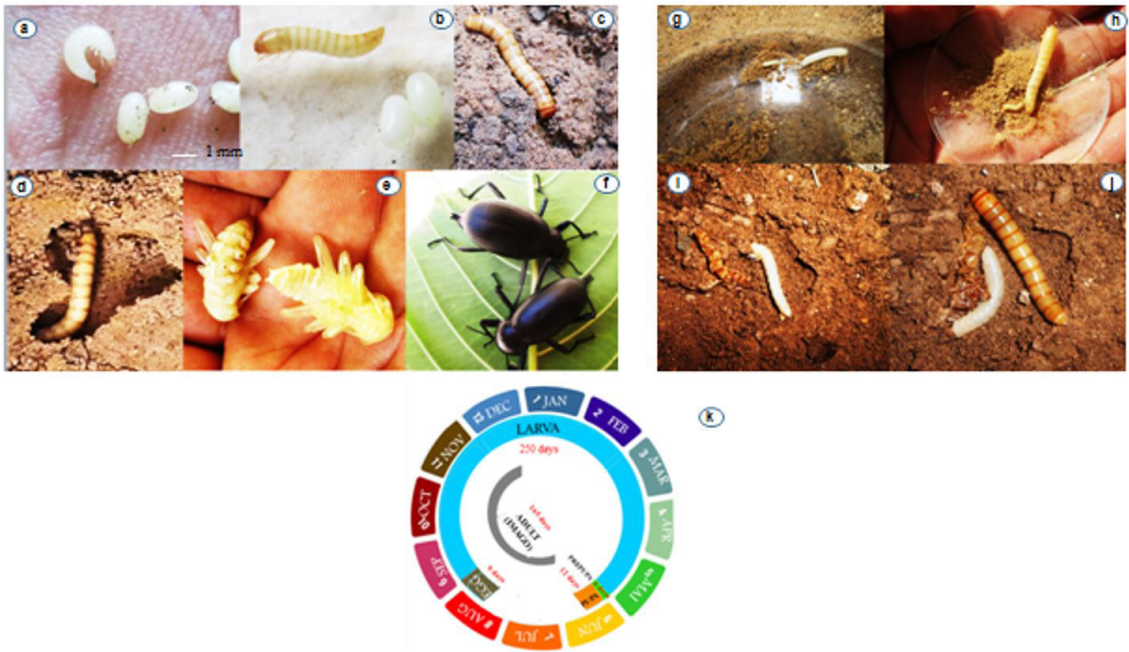


Figure 3. *B. nefrauensis* life cycle: (a) Eggs and newly hatched larva (1st instar). (b) 2nd instar larva beside eggs. (c) Larvae in the last instar. (d) Prepupa. (e) Pupa. (f) Adults. (g) Newly moulted 2nd and 4th instar larvae. (h) Moulded 2nd and 6th instar larva began to yellowish. (i) Newly moulted larva beside its old yellow cuticle. (j) Newly moulted larva compared with larva in the 8th instar. Diagram representation of the life cycle of *B. nefrauensis* (k).

low humidity). They were active during summer and autumn before their expiration in winter. Beetle activity decreases when it's windy and intensifies mostly after the rain if there is no wind. Adults can live six months continuing to feed, mate and produce eggs most of their lives. They feed on decomposing plant matter, fruits, cereals, bread, cooked meat and vegetable, pasta, rest of soup, chickpea etc.) They began to mate about seven days after their emergence. Mating increases normally after rain peaking in August. Female laid their eggs from late July in moist soil, and under stones and planks. Larvae normally hatch in nine days and develop into adults in about 250 days (autumn, winter and spring). Since summer days are characterized by high temperature (more than 35 °C reaching 45 °C) and low humidity, larvae survived only from the beginning of autumn. There are approximately fifteen larval instars. Larvae were usually hidden in soil digging tunnels or under stones. Sometime when the weather is moderate (no wind, no rain, no sirocco), larvae may leave their tunnel and shelters for a night walking on the ground crossing decomposing organic matter. As all beetles belonging to order Coleoptera, *B. nefrauensis* is a holometabolous insect with complete metamorphosis. Figure 3k depicts its life cycle. The field observation revealed that during summer months, eggs and larvae died under high temperature (in the field, second instar larvae can be observed from September). Then, the life cycle began with eggs laid at the beginning of autumn with embryogenesis duration of about 9 days. Newly hatched larva continues its development (fifteen instars) to the

pupation stage lasting about 9 months (from September to late May). Then the larva stopped feeding and entered into the prepupa stage for about four days. During the first half of June it submits a metamorphosis transforming into the pupal stage which lasts twenty days on average. In mid-June the adult emerges and remains alive until late autumn fulfilling its biological function. In winter adults expire leaving their larvae in the soil as a new generation. Approximately, the duration of every stage: eggs, larvae, prapupae, pupa and adults were 9, 250, 4, 12, and 165 days, respectively (about 14.5 months total). In the breeding laboratory, adults males and females in captivity reached their second year and still alive and active. Females laid eggs for the second season.

3.2. Notes on the first instar larvae

Newly hatched eggs larvae were small in size and white in color, yet when seen under light microscope, they appear to be rather transparent. This has allowed visualizing the external morphology with more details and occasionally observing nerve fibers through the cuticle in some areas. Herein, the external morphology will be described briefly. The innervation of the antennal and the ligular setae are the main object of this contribution.

Body of the first instar larvae (Figure 4a) elongate and subcylindrical, length 4.5-5 mm, color ivory-white with dark red mandibles, integument soft, thoracic and abdominal segments subcylindrical ninth abdominal segment triangular with two small urogomphi and two

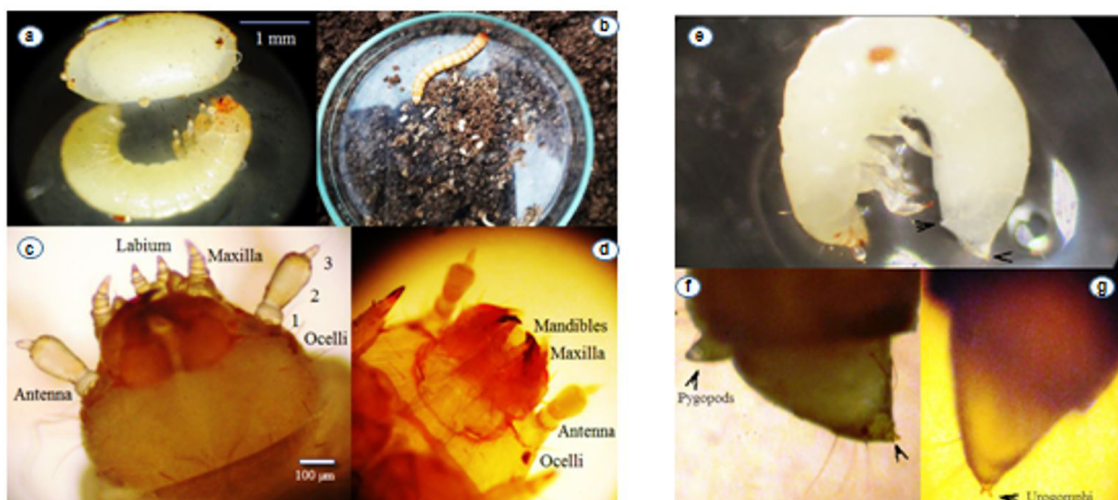


Figure 4. Newly egg hatched larva: (a) First instar larva compared with egg, scale = 1 mm. (b) First instar larva compared with 8th instar larva. (c) Head (dorsal view). (d) Head (ventral view). (e-g) Pygopods and urogomphi in first instar larva

pygopods subconical well visible (Figure 4e-g) Compared with older instar, newly egg hatched larvae are very small (Figure 4b).

Head (Figure 4c, d) Prognathous, wider than long, convex above, lateral margins rounded lateral side behind antenna bears brown ocelli. Ocelli arranged in an anterior group of three and a posterior group of two on each side. Antenna three-segmented, antennomere 1 cylindrical as wide as long, antennomere 2 widest distally (pear or spinning top-shaped), three time longer than wide and about 3.5 times than the first (longer than the other two segments together), bearing apically small sensilla surrounding the base of segment 3, antennomere 3 short and cylindrical, about 1/5 diameter and long of second, apically bearing a long erect seta surrounded with a diversity of small sensilla. Maxillary palpi (Figure 5c) distinctly three-segmented with sensory area on apex and single seta in each segment, lacinia (Figure 5b) with some of ten teeth, galea with a tuft of setae at apex surrounded by small sensilla. Labial palpi (Figure 5c), two-segmented (appear to be three-segmented in dorsal view having the same shape of maxillary palpi). Ligula with two long erect setae on tip, visible in ventral view and occasionally in dorsal view (Figure 5a,c). Mandibles well developed with two apical teeth. The dorsal view showed that paired mandibles composed of two lobes superimposed and inserted well in the clypeus (Figure 4c).

Further investigation using light microscopy and zoom of the camera (Canon IXUS) showed that the two ligular setae appear to be innervated with two nerve fibers across the ligula (Figure 5e). Similarly, the apical antennal seta seems to be innervated with single nerve fiber across the third antennal segment (Figure 5f).

Note, an examination on the larvae (embryo) just before egg hatching showed furthermore that the antennae are covered by a fine cylindrical film protecting apically the antennal sensilla (Figure 5g, h).

4. Discussion

4.1. Life cycle and biology

Tunisia is located in the central part of North Africa. The Country belongs to the Mediterranean region, which is well known for its high biodiversity. It is also a part of the Maghreb area which is an important Pleistocene glacial refuge both in North Africa and in the Western Palearctic (Myers et al., 2000; Husemann et al., 2013). The first data on Tunisian beetles come from the 19th and starting of the 20th centuries (Vigors 1825; Fairmaire 1858; Seidlitz 1893; Bedel 1895, 1900; Bodemeyer, 1927; Normand, 1933; Mandl, 1935). Among about 250 species belonging to the *Blaps* genus distributed worldwide, about 17 *Blaps* species was recorded in Tunisia (including *B. nitiduloides* Soldati, 2017 and *B. teocchii* Soldati, 2017 were recently identified). Of these, *B. nefrauensis nefrauensis* Seidlitz, 1893, was recorded in the Tunisian desert between Gabes and Gafsa (Soldati et al., 2017). Moulares is a town and commune in the Gafsa Governorate, Tunisia. It is part of the west-central arid region of Tunisia and has a dry climate marked by Mediterranean and continental influences. The city is located in a transition zone between the steppe and desert (Karaouli et al., 2009; Mohamed et al., 2013; Raja et al., 2014). It seems that our study beetle preferred Moulares, inhabiting the domestic gardens where they have easy access to food and shelter (humidity, decomposing matters, concealed shelters...). Its complete life cycle with egg, larva, pupa, and adult was in summary about 15 months. Every year in the same month (June) the adults emerged. Furthermore, in the same period (mid-May) larvae began to pupate. Here again, every year before winter adults expired (the same phenomena is repeated every year). Therefore, we assumed that the duration of every generation is one year. Beetles belonging to the *Blaps* genus vary significantly in their biological development duration.

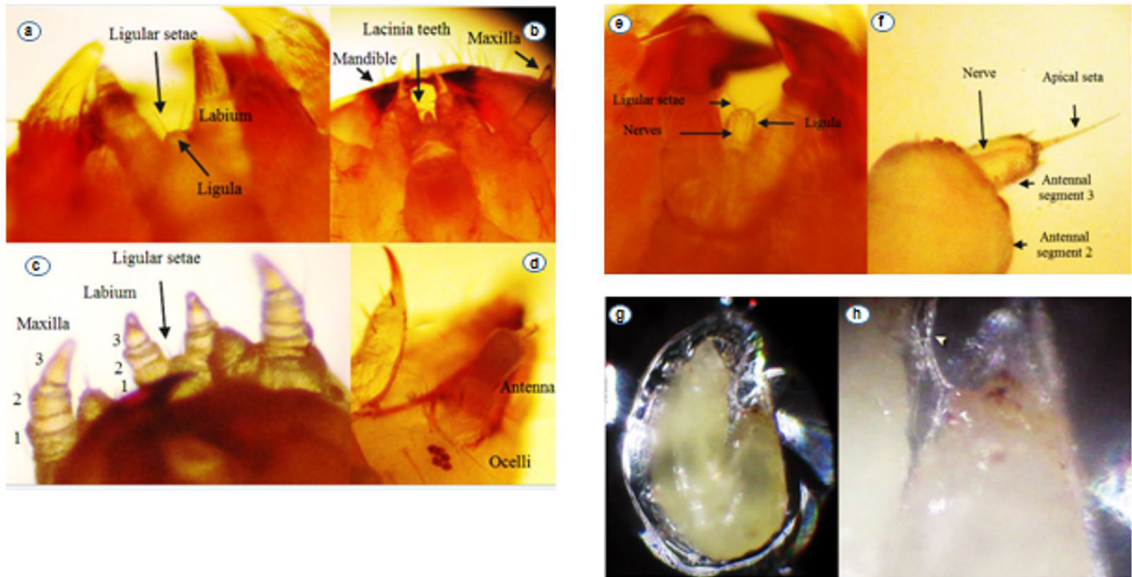


Figure 5. Light microscopy of some structure of the newly hatched egg larva of *B. nefrauensis*: (a) Ligula (ventral view). (b) Lacinia teeth (ventral view). (c) Ligula, and Labial and Maxillary palpi (dorsal view). (d) Ocelli (lateral view). (e) Nerves across ligula. (f) Nerve across third antennomere. Light microscopy on the antennal protection in the embryo of *B. nefrauensis* (g) Larva just before egg hatching. (h) Cylindrical membrane covering apically the antennae.

According to Knor (1975), *B. lusitanica* populations have a three-year life cycle. However, the development period of *B. kollari* doesn't reach one year (Menon and Putnam, 1988). In laboratory conditions, larval development of *B. Kollari* was reported as being very rapid, lasting only 80 days or less. Authors attributed the rapid development to an adaptation in Blaptinii living in arid conditions to make the best use ephemeral moisture. De Los Santos et al. (2006) examined the correlation between thermal habitats and the life cycle of two species belonging to the *Pimelia* genus (Tenebrionidae) on two extreme sites along an elevation ranges on Tenerife (Canary Islands). They concluded that *P. ascendens* presented a two years life cycle with larvae and adults overwintering. However *P. canariensis* had one year life cycle. This difference was explained that in addition to the two species inhabiting different ecological systems, they also need to synchronize their life history with climatic fluctuations of different intensities. Multiyear life cycles are common in order Coleoptera. Larval development of some species of the Scarabaeidae family takes up to two years reaching 5 years in different climatic zones. In Lucanidae, Carambycidae, Alleculidae and many other Coleopteran families, the biological development of various species can reach 5 years or more (Saulich, 2010). Short life cycles are also somewhat prevalent in beetles (Danks, 2006). Concerning our study beetle *B. nefrauensis*, the annual variation in temperature plays the principal role in its biological development. Thus, adults expire under the low temperature in winter while larva stage can't exceed late spring due to high temperature. In the beginning of summer larvae pupate or expire under the high temperature and low humidity.

However, larvae of certain darkling beetles as tribe Helopini (Tenebrionidae) in the arid zone of European Russia can migrate deep into the soil and have a summer diapause (Byzova and Ghilarov, 1956).

In the gardens, the soil was frequently watered and the humidity appears to be kept up during all seasons in soil. Consequently, the majority of laid eggs hatched and the majority of larvae in the first instar attain pupation. Adults found in gardens were able to oviposit, feed and find shelter. Therefore, we assume that the availability of moisture and food in the urban area increase the frequency of our study insect. Hence, *B. nefrauensis* populations inhabited urban zones and occupied gardens as an adaptive strategy to survive and to increase their abundance keeping their life cycle duration around one year under the arid and desert climate. In rearing conditions, males and females collected two years ago are still alive and active. This is maybe not strange because other species of the genus *Blaps* from Tunisia like *B. nitens requini* lived in the laboratory for about six years (Cloudsley-Thompson, 1956). Our finding suggests that imago expire in the early years in the fields under various factors (cold, wind, snow, and rain) which tend to intensify in wintertime. All of these factors together, in addition to starvation can begin to disrupt the species' refuges and habitats to the point that they reach levels that would be lethal to adult *Blaps* during the cold season. It is believed that if adults spend only the winter in rearing and the remaining seasons in the field, they can remain alive and active for many years.

4.2. Innervation in ligula and antennae

In insects, the chemical senses are important to detect various types of environmental chemical information.

The gustatory and olfactory receptors play an important role in feeding and reproduction, where sensory neurons for the perception are found (Hallem et al., 2006; Shields, 1994). The gustatory receptor neurons are present mostly in the mouthparts; whereas the olfactory receptors neurons are located on the antennae (Van der Goes van Naters and Carlson, 2006). Crouau (1997) mentioned that mechanosensitive neurons associated with insect setae are bipolar cells with an axon and a dendrite, and linked further to the physical environment. Coleopteran larvae are provided in a family of sensory structures known as sensilla, present on the antennae, mouthparts, wing, legs and the ovipositors (Crouau, 1997). The larval antenna is an important sensory organ thanks to a variety of associated sensilla that perform various functions: mechanoreception, vibroreception, proprioception, and thermo-hygroreception. In Elateridae, Faucheux (2014) listed sex groups of sensilla (aporous sensilla chaetica, uniporous sensilla chaetica, long aporous sensillum chaeticum, sensillum campaniformium, sensillum styloconicum, and sensilla basiconica) distributed over the antennal three-segments in *Drilus mauritanicus* larvae. He mentioned that the number of long setae (chaetica) in the larvae of *D. mauritanicus* (Elateridae) is 16 as against only one in the antennal larvae of the tenebrionid *Tribolium* ssp. This difference in the number of long sensilla can be explained the different lifestyle in the two larvae. Thus, the variety and the high number of sensilla chaetica can be useful as gustative and mechanoreceptors for the predator larvae of *Drillus*.

The larval palps are covered by different categories of sensilla which have various receptions. Corbière-Tichané (1969) has classified the sensory tuft crowning the extremity of palps in *Speophyes lucidulus* larvae (Coleoptera, Catopidae) into eight different groups. The main types of sensory organs were present: mechanoreceptors, olfactory, and gustatory receptors. He has also described a sensillum with a particular structure. The author especially described eight pairs of sensilla located on the ventral surface of the ligula, number of them were campaniformia and basiconica sensilla. He affirmed that all the receptors of the ligula have a simple innervation (dendritic) and may present various types of mechanoreceptors.

In tenebrionidae, the last instar larvae of *Tribolium* ssp and *Tenebrio molitor* have three-segmented antennae. The reduced terminal segment bears apically a single long seta, the trichoid terminal sensillum surrounded by several types of small sensilla. Behan and Ryan (1978) cited five categories of sensilla (trichoid, styloconic, placoid, campaniform, and coeloconic sensilla) as antennal receptors in *Tribolium* larvae. The trichoid sensillum is a long, thin and aporous seta innervated by one or two bipolar neurons (Bloom et al., 1982; Behan and Ryan 1978). It can be considered as a sensillum filiformium sensitive to air currents (vibroreceptor or anemoreceptor), possess also probably gustative or chemoreceptive functions (Faucheux, 2014).

Our results supported by findings of Behan and Ryan (1978) for the genus *Tribolium* and Bloom et al. (1982) for *Tenebrio molitor*, were shown that the third antennal segment in *B. nefrauensis* larvae bears different types of

sensilla with single long seta crowning apically the tip. Under light microscopy, the apical seta appears to be linked with single nerve fiber cross the terminal segment. Hence, the structure agrees with larvae of these cited species placed on Tenebrionidae in number, form and associated innervation of the antennal sensory organ.

An examination of the ligula in the newly egg hatched larvae revealed the presence of two nerve fibers innervating two apical setae; it is there that each seta was associated with one fiber crossing the ligula. It is important here to notice that this structure was shown for the first time under light microscopy. As these two elongate setae can be filiform sensilla, and if we consider that they located centrally in the middle of the larva mouthparts and innervated by paired nerves extend from the tip of the ligula to the nervous system (brain), they give the impression of having gustatory and taste functions sensitive mainly to the organic matter. The structure suggests that ligula is notably involved in the chemo-detection of food sources.

Results in our study found also that the antennae in the embryo just before egg hatching were protected apically by a fine cylindrical membrane. This can be explained by the need for the protection of the antennal sensilla (especially the apical seta) during the embryonic stage and the fact that the embryo must stay further from all antennal detections to keep its physiological hemostasis.

5. Conclusion

Blaps nefrauensis nefrauensis is a desert beetle found in Tunisia. Populations of this species colonise urban areas in search of a better environment to live and reproduce. The results of our study showed that the lifespans of larval, prepupal, and pupal stages are almost identical to those found in fields or rearing. Conversely, the lifespan of an adult beetle is particularly dependent on the biogeographical conditions and environment of the area and may be extended if living conditions change. Home gardens in good condition provide numerous services to maintain the insect's life cycle. However, Moulares like other cities shows continued and hurried building expansion and management by limiting the garden areas and old structures. So, the removal of soil and construction of new buildings could be fatal to both the imago and its larvae. Moreover, Moulares is a town that belongs to the Gafsa mining area and is known for its phosphate production. Therefore, the beetles inhabiting this industrial area could be affected by the harmful effects of toxic emissions (gas and dust) related to industrialization, but this requires further studies.

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