Strong ALK and PD-L1 positive IHC expression related *ALK* amplification in an advanced lung sarcomatoid carcinoma: a therapeutic trap?

Sébastien Gendarme a; Lise Matton a; Martine Antoine b; Khaledoun Kerrou c; Anne-Marie Ruppert a; Christelle Epaud a; Jacques Cadranel a; Vincent Fallet a

Affiliations:

a Department of Pneumology and Thoracic Oncology, Tenon Hospital, Assistance Publique-Hôpitaux de Paris and GRC 4, Theranoscan, Sorbonne Université, 75970 Paris, France.
b Pathology Department, AP-HP, Groupe Hospitalier HUEP, Hôpital Tenon, Paris, France.
c Department of Nuclear Medicine, Tenon Hospital. Sorbonne University, Paris, France.

Address for correspondence: Sébastien Gendarme, Department of Pneumology and Thoracic Oncology, Hôpital Tenon, 4 Rue de la Chine, 75 020 Paris, France; mail: s-ge@hotmail.fr

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Lung sarcomatoid carcinoma (LCS) is a rare aggressive Non-Small Lung Cancer (NSCLC), poorly differentiated and containing sarcoma or sarcoma-like elements. It is characterized by high mutation rates such as KRAS, EGFR, ALK or MET and PD-ligand 1 (PD-L1) overexpression [1,2]. Anaplastic Lymphoma Kinase (ALK) rearrangement is considered as one of the major oncogenic driver pathway in NSCLC, leading to target therapy [3]. Immunohistochemistry (IHC) is the primary therapy-determining test for ALK rearrangement. IHC is validated against Fluorescence in situ Hybridization (FISH) [4]. Occasionally, some discordance may occur in about 5% of cases [5]. In rare cases, this discordance reveal more complex biological mechanism, as 5' deleted pattern, split signals pattern or amplification pattern [6]. ALK amplification is defined by the presence of 6 copies or more of ALK per cell in more than 10% of analysed cell [7]. FISH for chromosome centromere 2 (CEP2) could help in order to assess the true ALK amplification, excluding the polysomy for chromosome 2. ALK amplification was mainly reported as a mechanism of resistance to ALK inhibitors [8,9]. However, ALK amplification may, as in our case, be a primary mutation in lung cancer. There is still few data about the management of the genetic pattern of ALK other than ALK rearrangements.

Here we report a case of NSCLC ALK highly positive in IHC, with initial life-threatening disease progression after beginning ALK tyrosine kinase inhibitors (ALK TKI). Further investigations reveal ALK amplification, which is not sensitive to ALK TKI. Dramatic response to immune checkpoint inhibitor (ICI) was then observed.

2- CASE REPORT

In November 2018, a 52-year-old woman, current smoker, with no history of disease or family history of cancer was diagnosed with an advanced non-small cell lung carcinoma of the right upper lobe with liver, adrenal and bone metastasis (Figure 1A). Liver biopsy was performed. Pathological and molecular analyses revealed a lung sarcomatoid carcinoma (LSC) highly expressing EML4-ALK (100% - clone 5A4) and PDL1 (60% of tumoral cells - clone E1L3N) on immunohistochemistry (IHC). Next-Generation DNA Sequencing revealed coexistence of a CTNNB1 (Catenin Beta 1); no ALK mutation or other molecular alteration were found. As recommended [4], ALK tyrosine kinase inhibitor (TKI) i.e. Alectinib (600mg daily) was started as first line treatment on the basis of the strong ALK-IHC. Two weeks later, motor deficit appeared in the lower part of the body due to tumor spinal cord compression in the 3rd thoracic vertebrae. Disease progression was confirmed with an increase in size of the primary tumor. Neurosurgical and radiotherapy treatment were performed in emergency with post-operative complications as abscess pneumonia and pulmonary embolism. However, motor deficit in the left leg did not improve.

Then, Vysis ALK break part fluorescence in situ hybridization (FISH) probe (Abbot Molecular, USA) revealed an ALK amplification (87.5% of malignant nuclei > 6 ALK copies in cell for orange fusioned spots) without ALK rearrangement (Figure 2). Alectinib was stopped and patient received pembrolizumab 200mg every 3 weeks as a second line, since January 2019.

Then, the motor deficit has decreased with a gradual improvement in her clinical condition. At the present date, the patient has received 21 cycles of immunotherapy with a persistant metabolic response on 18F-FDG PET (positron emission tomography) and a complete resolution of the neurologic deficit (Figure 1B).
Immunohistochemistry is the primary therapy-determining test for ALK rearrangement. An IHC result with strong marking (score 3+) is sufficient to indicate targeted treatment and does not require FISH control [4]. Occasionally, as in our case, clinical features such as current smoker or bronchopulmonary cancers other than adenocarcinoma, must raise suspicion of the ALK-IHC validity by performing further molecular tests [10].

Indeed, discrepancies between IHC and FISH are not uncommon because of the technical considerations and biological reasons such as ALK amplification, polysomy or type of ALK fusion gene [5]. Several clinical cases have, like us, reported cases of ALK amplification with positive results in immunohistochemistry and negative results in FISH [11]. This discrepancy is sometimes reversed with positive results in FISH and negative results in immunohistochemistry, due to the lack of correlation between mRNA/ALK protein level and to the lack of fusion protein expression [12]. Although therapeutic responses to ALK-TKI have been reported in cases of mismatch between IHC and FISH, treatment of ALK-amplified tumours remains controversial. Several clinical cases raise doubt about the therapeutic response of these cancers to ALK-TKI whether ALK amplification occurs as a secondary resistance mechanism or as a primary molecular alteration. (Table 1) In addition, further investigations into the use of clone 5A4 in our study, compared to clone D5F3 in other reported clinical cases, should be realized.

Therapeutic alternatives can then be considered. Although data suggests poor efficacy of immune checkpoint inhibitors (ICI) in ALK-rearranged tumours [13], its efficacy remains uncertain in ALK-amplified tumours. In our clinical case, the rapid progression after starting Alectinib and the prolonged response to ICI may lead us to consider the PDL1 marker as a real predictor of ICI efficacy in this situation i.e. smoking exposure and sarcomatoid histology.

Indeed, LSC may present mutations such as KRAS, EGFR, ALK and MET in more than 70% of cases, and PD-L1 expression in 70%-90% of cases [14]. Little is known on the best therapeutic strategy to apply in case of both molecular alterations and PD-L1 expression. Several clinical cases have reported the efficacy of ICI in LSC with molecular alterations such as c-MET amplifications [15]. In the Immuntarget registry, the objective tumour response to ICI in non-small cell lung cancers by driver alteration was KRAS = 26%, BRAF = 24%, ROS1 = 17%, MET = 16%, EGFR = 12%, HER2 = 7%, RET = 6%, ALK = 0%. [13] ALK-rearranged tumours were refractory to ICI despite PD-L1 expression. Tumours with ALK rearrangement thus appeared as poor candidates for ICI, even after failure of ALK TKI or chemotherapy. PD-L1 marking is often consider as a trap in this situation with a low predictive value of response to ICI. Our clinical case thus demonstrates the usual response of a tumor with ALK molecular alteration. ALK rearrangements, mutations and amplifications probably need to be apprehended separately on the predictive value of PD-L1 and on the response to ICI. To our knowledge, our clinical case is the first one to report a prolonged response to ICI with ALK amplification.

Apart from the expression of PD-L1, other factors may explain the response to ICI in our case. Smoking status is often associated with a higher tumor mutation burden. The presence of the CTNNB1 mutation has also been associated with a better response to immunotherapy in other cancers [16,17]. This could be explained by the role of CTNNB1 in modulating the activity of NK cells by affecting CD96. Further investigations should be performed to better understand the role of this mutation in non-small cell lung cancer.
CONCLUSION

In case of an unusual phenotype, an ALK-IHC positive test should be confirmed by FISH or ARN/DNA molecular testing before starting an ALK-TKI. The concomitant presence of $ALK$ amplification and PD-L1 expression in LSC should not be an impediment to the initiation of ICI.
**Figure 1.** 18F-FDG PET 3D Maximum intensity Projection (3D MIP) anterior views at baseline (A) and after 14 months of treatment with pembrolizumab (B) with baseline (A1, A2) and post therapy (B1, B2) transaxial slices on 18F-FDG PET, CT and fusion PET/CT respectively from left to right passing through the right lung apical mass and invaded T3 vertebra on first row (A1) and same slices after therapy on second row (B1). Transaxial slices passing through the liver and right adrenal gland (2 sites) metastasis on third row (A2) and same slices after therapy on fourth row (B2).
Figure 2. Histological analyses of the biopsied sample. Eosin hematoxylin staining is presented at magnification of (A) x 40 and IHC is presented at magnification of (B) x 10. FISH analysis shows no ALK translocation (C) but amplification (D).
Table 1. Clinical cases or series of cases reporting clinical and pathological data from subjects with lung cancers with ALK amplification (M: male; F: female; ADC: adenocarcinoma; SCC: squamous cell carcinoma; BAC: bronchioalveolar carcinoma; LSC: lung sarcomatoid carcinoma; PD: progressive disease, DOD: Died of disease, BA: Break Apart)

<table>
<thead>
<tr>
<th>Study</th>
<th>Cases</th>
<th>Sex</th>
<th>Histological type of tumor</th>
<th>Smoking</th>
<th>ALK IHC</th>
<th>ALK FISH</th>
<th>ALK FISH Pattern</th>
<th>Treatment</th>
<th>Overall Patient Response</th>
</tr>
</thead>
<tbody>
<tr>
<td>Scattone et al, [6]</td>
<td>1</td>
<td>F</td>
<td>ADC</td>
<td>Yes</td>
<td>Negative (clone D5F3)</td>
<td>Positive</td>
<td>Del 5' 32%, Split 16% Gains 64%</td>
<td>Crizotinib</td>
<td>PD</td>
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<tr>
<td></td>
<td>1</td>
<td>M</td>
<td>ADC</td>
<td>N/A</td>
<td>Negative (clone D5F3)</td>
<td>Positive</td>
<td>Del 5' 80%, Split 6% Gains 20% Polysomy 65%</td>
<td>Crizotinib</td>
<td>PD</td>
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<tr>
<td>Pelosi et al, [18]</td>
<td>5</td>
<td>2F / 3M</td>
<td>LSC (4 pleomorphic carcinoma and 1 pulmonary blastoma)</td>
<td>4 current 1 never</td>
<td>Negative</td>
<td>Positive</td>
<td>N/A</td>
<td>Surgery</td>
<td></td>
</tr>
<tr>
<td></td>
<td>11</td>
<td>2F / 9M</td>
<td>ADC, 5 SCC, 1 BAC</td>
<td>7 current 3 former 1 never</td>
<td>Negative (clone D5F3)</td>
<td>Positive</td>
<td>N/A</td>
<td>Surgery N = 9 N/A N = 2</td>
<td></td>
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<tr>
<td>Ilie et al, [11]</td>
<td>2</td>
<td>1F / 1H</td>
<td>ADC</td>
<td>N/A</td>
<td>Positive (clone D5F3)</td>
<td>Negative</td>
<td>BA + high level of ALK polysomy</td>
<td>Chemotherapy</td>
<td>PD</td>
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REFERENCES


