Strong ALK and PD-L1 positive IHC expression related *ALK* amplification in an advanced lung sarcomatoid carcinoma: a therapeutic trap?

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1- INTRODUCTION

Lung sarcomatoid carcinoma (LSC) is a rare aggressive Non-Small Lung Cancer (NSCLC), poorly differentiated and containing sarcoma or sarcoma-like elements. It is characterized by high mutations rates such as KRAS, EGFR, ALK or MET and PD-ligand 1 (PD-L1) overexpression [1,2]. *Anaplastic Lymphoma Kinase (ALK)* rearrangement is considered as one of the major oncogenic driver pathway in NSCLC, leading to target therapy [3]. Immunohistochemistry (IHC) is the primary therapy-determining test for *ALK* rearrangement. IHC is validated against Fluorescence in situ Hybridization (FISH) [4]. Occasionally, some discordance may occur in about 5% of cases [5]. In rare cases, this discordance reveal more complex biological mechanism, as 5' deleted pattern, split signals pattern or amplification pattern [6]. *ALK* amplification is defined by the presence of 6 copies or more of ALK per cell in more than 10% of analysed cell [7]. FISH for chromosome centromere 2 (CEP2) could help in order to assess the true *ALK* amplification, excluding the polysomy for chromosome 2. *ALK* amplification was mainly reported as a mechanism of resistance to ALK inhibitors [8,9]. However, *ALK* amplification may, as in our case, be a primary mutation in lung cancer. There is still few data about the management of the genetic pattern of *ALK* other than *ALK* rearrangements.

Here we report a case of NSCLC ALK highly positive in IHC, with initial life-threatening disease progression after beginning ALK tyrosine kinase inhibitors (ALK TKI). Further investigations reveal *ALK* amplification, which is not sensitive to ALK TKI. Dramatic response to immune checkpoint inhibitor (ICI) was then observed.

2- CASE REPORT

In November 2018, a 52-year-old woman, current smoker, with no history of disease or family history of cancer was diagnosed with an advanced non-small cell lung carcinoma of the right upper lobe with liver, adrenal and bone metastasis (Figure 1A). Liver biopsy was performed. Pathological and molecular analyses revealed a lung sarcomatoid carcinoma (LSC) highly expressing EML4-ALK (100% - clone 5A4) and PDL1 (60% of tumoral cells - clone E1L3N) on immunohistochemistry (IHC). [Next-Generation DNA Sequencing revealed coexistence of a CTNNB1 \(Catenin Beta 1\); no ALK mutation or other molecular alteration were found.](#)

As recommended [4], ALK tyrosine kinase inhibitor (TKI) i.e. Alectinib (600mg daily) was started as first line treatment on the basis of the strong ALK-IHC. Two weeks later, motor deficit appeared in the lower part of the body due to tumor spinal cord compression in the 3rd thoracic vertebrae. Disease progression was confirmed with an increase in size of the primary tumor. Neurosurgical and radiotherapy treatment were performed in emergency with post-operative complications as abscess pneumonia and pulmonary embolism. However, motor deficit in the left leg did not improve.

Then, Vysis *ALK* break part fluorescence *in situ* hybridization (FISH) probe (Abbot Molecular, USA) revealed an *ALK* amplification (87.5% of malignant nuclei > 6 ALK copies in cell for orange fusioned spots) without *ALK* rearrangement (Figure 2). Alectinib was stopped and patient received pembrolizumab 200mg every 3 weeks as a second line, since January 2019. Then, the motor deficit has decreased with a gradual improvement in her clinical condition. At the present date, the patient has received 21 cycles of immunotherapy with a persistent metabolic response on 18F-FDG PET (positron emission tomography) and a complete resolution of the neurologic deficit (Figure 1B).

3- DISCUSSION

Immunohistochemistry is the primary therapy-determining test for *ALK* rearrangement. An IHC result with strong marking (score 3+) is sufficient to indicate targeted treatment and does not require FISH control [4]. Occasionally, as in our case, clinical features such as current smoker or bronchopulmonary cancers other than adenocarcinoma, must raise suspicion of the ALK-IHC validity by performing further molecular tests [10].

Indeed, discrepancies between IHC and FISH are not uncommon because of the technical considerations and biological reasons such as *ALK* amplification, polysomy or type of *ALK* fusion gene [5]. Several clinical cases have, like us, reported cases of *ALK* amplification with positive results in immunohistochemistry and negative results in FISH [11]. This discrepancy is sometimes reversed, with positive results in FISH and negative results in immunohistochemistry, due to the lack of correlation between mRNA/*ALK* protein level and to the lack of fusion protein expression [12]. Although therapeutic responses to ALK-TKI have been reported in cases of mismatch between IHC and FISH, treatment of *ALK*-amplified tumours remains controversial. Several clinical cases raise doubt about the therapeutic response of these cancers to ALK-TKI whether *ALK* amplification occurs as a secondary resistance mechanism or as a primary molecular alteration. (Table 1) In addition, further investigations into the use of clone 5A4 in our study, compared to clone D5F3 in other reported clinical cases, should be realized.

Therapeutic alternatives can then be considered. Although data suggests poor efficacy of immune checkpoint inhibitors (ICI) in *ALK*-rearranged tumours [13], its efficacy remains uncertain in *ALK*-amplified tumours. In our clinical case, the rapid progression after starting Alectinib and the prolonged response to ICI may lead us to consider the PDL1 marker as a real predictor of ICI efficacy in this situation i.e. smoking exposure and sarcomatoid histology.

Indeed, LSC may present mutations such as *KRAS*, *EGFR*, *ALK* and *MET* in more than 70% of cases, and PD-L1 expression in 70%-90% of cases [14]. Little is known on the best therapeutic strategy to apply in case of both molecular alterations and PD-L1 expression. Several clinical cases have reported the efficacy of ICI in LSC with molecular alterations such as *c-MET* amplifications [15]. In the Immunotarget registry, the objective tumour response to ICI in non-small cell lung cancers by driver alteration was KRAS = 26%, BRAF = 24%, ROS1 = 17%, MET = 16%, EGFR = 12%, HER2 = 7%, RET = 6%, ALK = 0%. [13] ALK-rearranged tumours were refractory to ICI despite PDL1 expression. Tumours with ALK rearrangement thus appeared as poor candidates for ICI, even after failure of ALK TKI or chemotherapy. PDL1 marking is often consider as a trap in this situation with a low predictive value of response to ICI. Our clinical case thus demonstrates the unusual response of a tumor with ALK molecular alteration. ALK rearrangements, mutations and amplifications probably need to be apprehended separately on the predictive value of PDL1 and on the response to ICI. To our knowledge, our clinical case is the first one to report a prolonged response to ICI with *ALK* amplification.

Apart from the expression of PDL1, other factors may explain the response to ICI in our case. Smoking status is often associated with a higher tumor mutation burden. The presence of the CTNNB1 mutation has also been associated with a better response to immunotherapy in other cancers. [16,17] This could be explained by the role of CTNNB1 in modulating the activity of NK cells by affecting CD96. Further investigations should be performed to better understand the role of this mutation in non-small cell lung cancer.

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CONCLUSION

In case of an unusual phenotype, an ALK-IHC positive test should be confirmed by FISH or ARN/DNA molecular testing before starting an ALK-TKI. The concomitant presence of *ALK* amplification and PD-L1 expression in LSC should not be an impediment to the initiation of ICI.

Figure 1. 18F-FDG PET 3D Maximum intensity Projection (3D MIP) anterior views at baseline (A) and after 14 months of treatment with pembrolizumab (B) with baseline (A1, A2) and post therapy (B1, B2) transaxial slices on 18F-FDG PET, CT and fusion PET/CT respectively from left to right passing through the right lung apical mass and invaded T3 vertebra on first row (A1) and same slices after therapy on second row (B1). Transaxial slices passing through the liver and right adrenal gland (2 sites) metastasis on third row (A2) and same slices after therapy on fourth row (B2).

Figure 2. Histological analyses of the biopsied sample. Eosin hematoxylin staining is presented at magnification of (A) x 40 and IHC is presented at magnification of (B) x 10. FISH analysis shows no *ALK* translocation (C) but amplification (D).

Table 1. Clinical cases or series of cases reporting clinical and pathological data from subjects with lung cancers with *ALK* amplification
(M: male; F: female; ADC: adenocarcinoma; SCC: squamous cell carcinoma; BAC: bronchioalveolar carcinoma; LSC: lung sarcomatoid carcinoma; PD: progressive disease, DOD: Died of disease, BA: Break Apart)

Study	Cases	Sex	Histological type of tumor	Smoking	ALK IHC	<i>ALK</i> FISH	<i>ALK</i> FISH Pattern	Treatment	Overall Patient Response
Scattone et al, [6]	1	F	ADC	Yes	Negative (clone D5F3)	Positive	Del 5' 32% Split 16% Gains 64%	Crizotinib	PD
	1	M	ADC	N/A	Negative (clone D5F3)	Positive	Del 5' 80% Split 6% Gains 20% Polysomy 65%	Crizotinib	PD
Pelosi et al, [18]	5	2F / 3M	LSC (4 pleomorphic carcinoma and 1 pulmonary blastoma)	4 current 1 never	Negative	Positive	N/A	Surgery	Early recurrence after radical surgery and died of disease shortly thereafter (N=2)
Salido et al, (8)	11	2F / 9M	5 ADC, 5 SCC, 1 BAC	7 current 3 former 1 never	Negative (clone D5F3)	Positive	N/A	Surgery N = 9 N/A N = 2	N/A
Ilie et al, [11]	2	1F / 1M	ADC	N/A	Positive (clone D5F3)	Negative	BA + high level of <i>ALK</i> polysomy	Chemotherapy	PD

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