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β A3/A1-crystallin regulates apical polarity and EGFR endocytosis in retinal pigmented epithelial cells

Peng Shang¹, Nadezda Stepicheva¹, Kenneth Teel², Austin McCauley², Christopher Scott Fitting¹, Stacey Hose¹, Rhonda Grebe³, Meysam Yazdankhah¹, Sayan Ghosh¹, Haitao Liu¹, Anastasia Strizhakova ¹, Joseph Weiss¹, Imran A. Bhutto³, Gerard A. Luty³, Ashwath Jayagopal⁴, Jiang Qian ³, José-Alain Sahel ^{1,5}, J. Samuel Zigler Jr.³, James T. Handa³, Yuri Sergeev ⁶, Raju V. S. Rajala², Simon Watkins ⁷ & Debasish Sinha^{1,3,7}✉

The retinal pigmented epithelium (RPE) is a monolayer of multifunctional cells located at the back of the eye. High membrane turnover and polarization, including formation of actin-based apical microvilli, are essential for RPE function and retinal health. Herein, we demonstrate an important role for β A3/A1-crystallin in RPE. β A3/A1-crystallin deficiency leads to clathrin-mediated epidermal growth factor receptor (EGFR) endocytosis abnormalities and actin network disruption at the apical side that result in RPE polarity disruption and degeneration. We found that β A3/A1-crystallin binds to phosphatidylinositol transfer protein (PITP β) and that β A3/A1-crystallin deficiency diminishes phosphatidylinositol 4,5-bisphosphate (PI(4,5)P₂), thus probably decreasing ezrin phosphorylation, EGFR activation, internalization, and degradation. We propose that β A3/A1-crystallin acquired its RPE function before evolving as a structural element in the lens, and that in the RPE, it modulates the PI(4,5)P₂ pool through PITP β /PLC signaling axis, coordinates EGFR activation, regulates ezrin phosphorylation and ultimately the cell polarity.

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The retinal pigment epithelium (RPE) is a monolayer of highly polarized, differentiated, and mitotically quiescent cells, situated between the photoreceptors and the choriocapillaris in the retina, with diverse functions in retinal health, including roles in engulfment and digestion of shed photoreceptor outer segments, the visual cycle, and nutrient transport¹. The function of RPE cells critically relies on their intensive membrane recycling activities, including phagocytosis, autophagy, and endocytosis². In addition to their highly dynamic membrane activities, RPE cells have a highly polarized phenotype³. Like many other epithelial cell types, they have F-actin-based microvilli on their apical surface that serve as the platform for endocytosis, signal transduction, nutrient transport, etc⁴. With age and disease, the RPE microvilli become disorganized and atrophic⁵. The apical actin cytoskeletal architecture is organized by the membrane-cytoskeletal linker ezrin and its scaffolding protein-EBP50 (ezrin/radixin/moesin-binding phosphoprotein 50)⁶.

The RPE requires phosphoinositides (also known as phosphatidylinositol phosphates or PIPs), a group of phospholipids (PLs) that have fundamental roles in membrane dynamics and cytoskeleton organization, for cellular signaling⁷. In fact, dysregulation of PIP metabolism in photoreceptors has been reported to result in blindness^{8–12}. While a few studies have focused on phosphatidylinositol-3-phosphate in membrane recycling in RPE cells¹³, the specific roles of different PIPs in RPE function and physiology remain uncertain^{14,15}. For example, the specific role in the RPE of phosphatidylinositol-4,5-phosphate (PI(4,5)P₂), the most abundant PIP and regulates multiple intracellular activities, including membrane remodeling and trafficking as well as cytoskeleton assembly¹⁶, is unknown. Importantly, PI(4,5)P₂ has been shown to be critical for autophagy and phagocytosis, which are essential processes performed by the RPE for vision¹⁶.

Moonlighting proteins are multifunctional proteins that perform different functions in different cells or cellular locations¹⁷. The first described moonlighting proteins were crystallins, which provide transparency and refractivity of the lens¹⁸. In other tissues, α B-crystallin serves as a small heat shock protein¹⁹, while duck ϵ -crystallin is a lactate dehydrogenase²⁰. Recently, we found that β A3/A1-crystallin is a moonlighting protein because in addition to its structural function in the lens, it is also expressed in retinal astrocytes, ganglion cells, and RPE cells^{21,22}. In the RPE, β A3/A1-crystallin is required for normal lysosomal function, including maintenance of lysosomal pH, intracellular amino acid levels, and regulation of phagocytosis and autophagy^{23–25}. We have shown that *Cryba1* cKO mice (conditional knockout of *Cryba1* specifically in the RPE) exhibit an age-related macular degeneration (AMD)-like phenotype^{23,24,26,27}.

We have also found that β A3/A1-crystallin may play an important role in maintaining RPE polarity. This protein is enriched at the apical region of polarized RPE cells and is not expressed in non-polarized RPE cells, such as non-polarized cultured RPE cell lines²⁷. Furthermore, RPE cells lacking *Cryba1*, the gene encoding β A3/A1-crystallin, undergo an epithelial-mesenchymal transition (EMT)-like process²⁷. Importantly, β A3/A1-crystallin is unique among all crystallins in that two closely related polypeptides, β A3 and β A1, are produced from two different translation start sites on a single *Cryba1* mRNA by leaky ribosomal scanning²⁸. The production of two polypeptides may contribute to the complexity of the intriguingly diverse functions of this moonlighting protein. By utilizing our well-characterized *Cryba1* cKO mouse model and RPE/choroid/sclera flatmounts, we herein suggest another important function of β A3/A1-crystallin: modulating the PI(4,5)P₂ pool in RPE cells via the PIP2/PLC signaling axis, with subsequent effects on cell polarity and EGFR signaling.

Results

***Cryba1* cKO RPE show age-related microvilli defects.** We investigated the RPE microvilli on retinal sections and RPE flatmounts from *Cryba1* cKO and age-matched floxed control mice by immunostaining for F-actin and EBP50. At 1 month of age, the apical microvilli of RPE cells in floxed mice are robust and well interdigitated with photoreceptor outer segments, while in *Cryba1* cKO mice they are disorganized. This disorganization became more severe with age (Fig. 1a, b). By 4 months of age, microvilli in RPE cells from *Cryba1* cKO mice had collapsed, progressing to near complete loss of microvilli in some RPE cells by 9 months. These abnormalities can be visualized by the orthogonal projection of z-stack confocal images on RPE flatmounts where the XY panels are taken from the same apical plane (Fig. 1b). The lower magnification images of retina cryosections and RPE flatmounts showed a patchy pattern of microvilli abnormalities in cKO RPE cells (Supplementary Fig. 1a, b). Ultrastructurally, the RPE exhibited disorganized interdigitation of microvilli with photoreceptor outer segments in 2-month-old *Cryba1* cKO retinas, and complete loss of microvilli in some RPE cells by 20 months (Fig. 1c). It is worth noting that the control RPE cells also showed mild age-related microvilli disorganization (Fig. 1a–c). Our data are in agreement with another group that showed progressive microvilli atrophy (shortening) in aging rats⁵.

To investigate the mechanism of microvilli atrophy, we checked the levels of ezrin phosphorylation since its scaffold protein, EBP50, only binds to phosphorylated ezrin to bring microfilaments to the plasma membrane²⁹. We found that ezrin phosphorylation (Thr567) was substantially decreased in 3-week-old *Cryba1* cKO RPE cells (Fig. 2a). The small GTPase Rac1, which has been shown to be activated by ezrin that has been phosphorylated at Thr567^{30,31}, was also reduced in the RPE from cKO mice. Both changes persisted through at least 9 months of age (Fig. 2a). These results indicate that lack of β A3/A1-crystallin impairs microvilli organization through a decrease in ezrin phosphorylation. EBP50 protein was also decreased in the RPE from *Cryba1* cKO mice compared to controls (Fig. 2b). Furthermore, we detected a significant reduction in Na⁺/K⁺-ATPase, an apical marker for RPE cells³², at 4 and 9 months old, but not at 3 weeks of age in RPE cells from the mutant mice (Fig. 2c, d). The levels of monocarboxylate transporter 3 (MCT3), which is normally localized to the RPE basolateral membrane³³, were not significantly different in *Cryba1* cKO RPE cells compared to controls. However, in the 9-month-old cKO cells, MCT3 lost its basolateral localization and was present in the cytoplasm and at the apical domain, indicating that RPE polarity was disrupted in RPE cells lacking β A3/A1-crystallin (Fig. 2c, d).

Other important factors that are required to maintain RPE polarity are adherens and tight junctions^{34,35}. β -catenin, an adherens junction component that interacts with cadherins or actin in normal conditions, is anchored to cell membranes³⁶. Studies have shown that β -catenin plays a role in cell–cell contact and the EMT process in native mouse RPE^{27,37}. In RPE flatmounts, β -catenin immunostaining showed partial translocation of β -catenin to the cytoplasm and nucleus in RPE cells of both 4- and 9-month-old *Cryba1* cKO mice, while it remained restricted to the plasma membrane in RPE cells from age-matched control mice (Fig. 3a). We also observed that β -catenin phosphorylation was significantly decreased in the RPE cells of *Cryba1* cKO mice compared to age-matched control RPE cells, starting at 4 months of age (Fig. 3b). Phosphorylated β -catenin usually recruits ubiquitin E3 and is subsequently degraded by the proteasome. Non-phosphorylated β -catenin is able to translocate to the nucleus and initiate transcription of EMT-related genes³⁶.

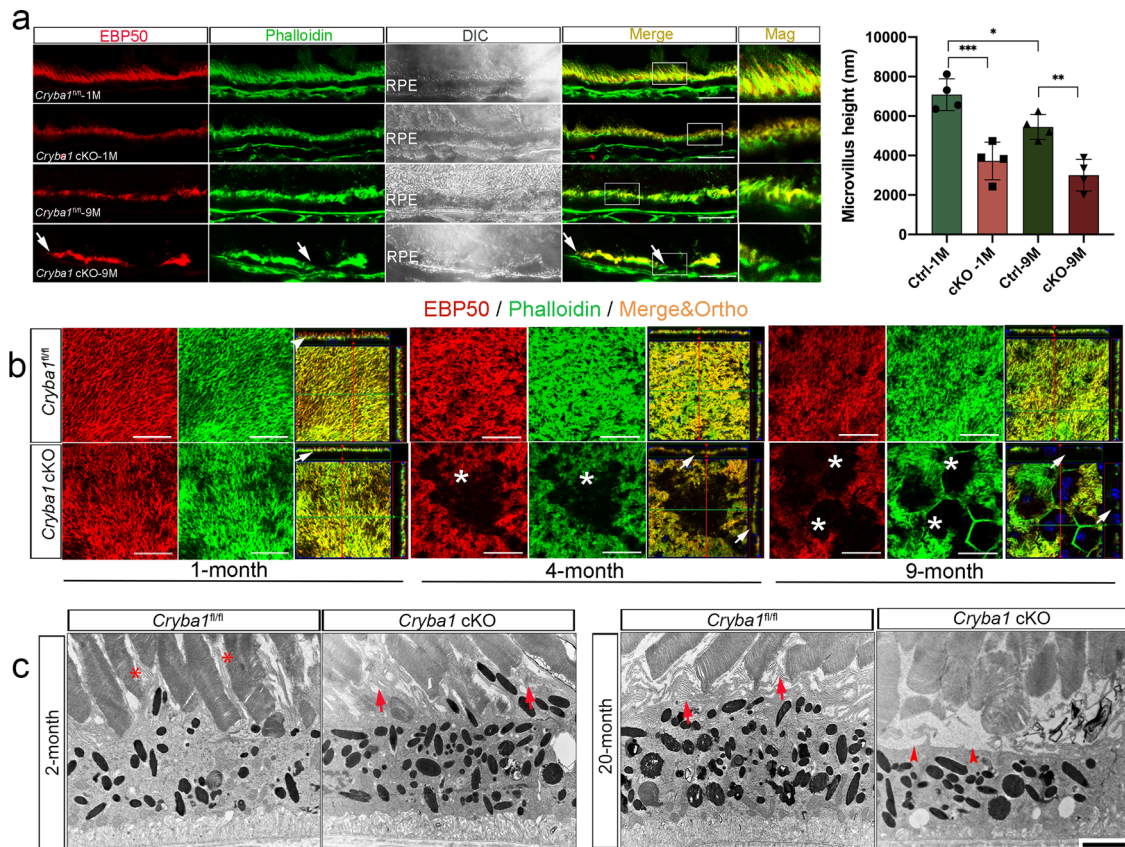


Fig. 1 *Cryba1* cKO RPE show age-related microvilli defects. **a** Immunostaining for EBP50 (red) and F-actin (phalloidin, green) on retina sections showed disorganized microvilli in 1-month-old cKO RPE cells, and microvilli loss (arrows) in RPE cells of 9-month-old cKO mice compared to age-matched control. Mag: magnified area outlined in merged image. DIC: differential interference contrast (DIC) microscopy showed the area of each sample for imaging. **b** Z-stack imaging for EBP50 (red) and F-actin (phalloidin, green) with an orthogonal projection showing upright and abundant microvilli in 1-month-old control (arrowhead), but not in cKO RPE cells (arrow). In 4-month-old mice, some cKO RPE cells presented collapsed microvilli indicated by sinking F-actin and EBP50 in the z-direction (arrows). Some RPE cells of 9-month-old cKO mice (asterisks) lacked microvilli completely. Scale bar: 20 μ m. **c** TEM images of cKO and control retina sections. Two-month-old control mice exhibited microvilli very well interdigitated with photoreceptor outer segments (asterisks). Arrows show disorganized microvilli of RPE cells in 2-month-old cKO and 20-month-old control mice. Arrowheads show microvilli loss in some RPE cells in 20-month-old cKO mice. Scale bar: 4 μ m. Statistical analysis was performed using one-way ANOVA. * $P < 0.05$, ** $P < 0.01$, *** $P < 0.001$, $n = 4$.

It can be speculated that in *Cryba1* cKO RPE cells, the decreased β -catenin phosphorylation allows more β -catenin translocation to the cytoplasm and nucleus, thereby initiating the EMT-like processes. We previously showed that RPE cells from *Cryba1* cKO mice exhibit elevated mRNA for the EMT transcription factors *Snai1* and *Snai2*, elevated vimentin protein, and loss of E-cadherin with dynamic reorganization of the actin cytoskeleton by 5 months of age²⁷. Interestingly, the phosphorylation level of β -catenin was not different in the cKO RPE at 3 weeks of age (Fig. 3b), indicating that activated β -catenin signaling in cKO RPE cells is likely to be a secondary effect of the cytoskeletal disorganization.

EGFR activation and internalization is compromised in *Cryba1* cKO RPE. Since phosphorylated ezrin also transduces extracellular signals by binding to receptor tyrosine kinases (RTKs) at the cell membrane^{38,39}, and RTKs are important signaling transducers that initiate signaling cascades for various intracellular activities⁴⁰, we investigated if loss of β A3/A1-crystallin affects RTK activation in the RPE. A phospho-RTK array was performed on RPE lysates from 2-month-old floxed control

and *Cryba1* cKO mice (Supplementary Fig. 2a). In order to stimulate RTKs and reduce individual variation, all mice were fasted for 36 h with or without subsequent refeeding for 3 h prior to the assay. We found that the phosphorylation of epidermal growth factor receptor (EGFR or ErbB1) and ErbB2, two members of the ErbB family of RTKs, was lower in *Cryba1* cKO compared to floxed control RPE cells from mice that were refed for 3 h following fasting (Fig. 4a). However, other phosphorylated RTKs, such as platelet-derived growth factor subunit A (PDGFR α) (Fig. 4a), either showed no significant difference between the groups, or had low abundance in RPE cells (Supplementary Fig. 2a). ErbB2, which has no identified activating soluble ligand and usually forms a heterodimer with other ErbB members⁴¹, may form a heterodimer with EGFR in RPE cells. Thus, *Cryba1* deficiency in the RPE did not alter the phosphorylation of all RTKs, but rather, specifically regulated the phosphorylation of EGFR and its interactive RTK (ErbB2). These data are supported by our previous observation that β A3/A1-crystallin interacted with EGFR in a proteome high-throughput array (CDI NextGen Proteomics Laboratories, Inc.)²³. While EGFR is a well-studied RTK in cancer cell biology⁴², little is known about its function in RPE cells. The specific effect of β A3/A1-crystallin on EGFR

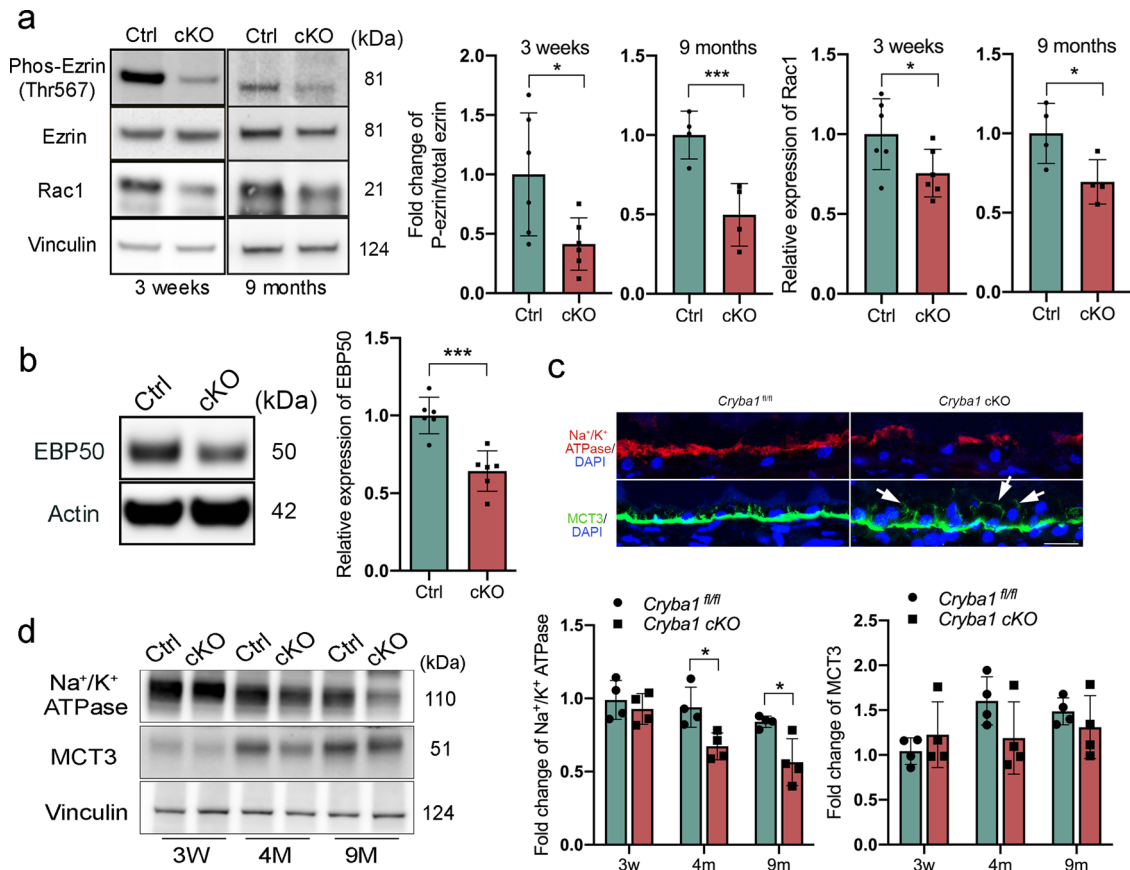


Fig. 2 Disrupted apical-basal polarity in RPE cells lacking *Cryba1*. **a** Western blot analysis of 3-week and 9-month-old RPE extracts showing that ezrin phosphorylation and Rac1 expression were reduced in cKO RPE cells at both ages. **b** Western blot analysis of 4-month-old RPE extracts confirmed downregulation of EBP50 protein in cKO RPE cells. **c** Immunostaining for Na⁺/K⁺ ATPase and MCT3 on sections of 9-month-old retina showed reduced Na⁺/K⁺ ATPase and mis-located MCT3 in *Cryba1* cKO RPE cells (indicated by arrows). Scale bar: 20 μm. **d** Western blot analysis of RPE extracts from control and cKO mice at different ages showed reduced Na⁺/K⁺ ATPase; however, there was no significant change in MCT3 abundance in cKO RPE. Statistical analysis was performed using either a two-tailed unpaired Student's *t*-test (**a**, **b**) or two-way ANOVA (**d**). **P* < 0.05, ****P* < 0.001, *n* = 4–6.

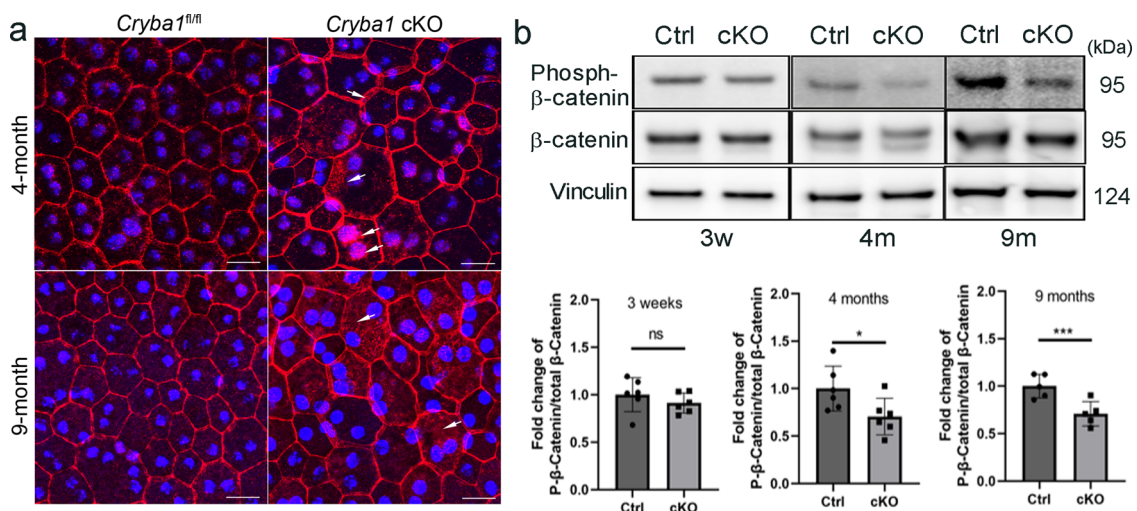


Fig. 3 Activated β-catenin in *Cryba1* cKO RPE. **a** RPE flatmounts prepared from 4-month and 9-month-old *Cryba1* cKO and *Cryba1*^{fl/fl} control mice were stained with anti-β-catenin antibody (red) and DAPI (blue). Arrows show cytoplasmic and nuclear β-catenin in cKO RPE cells. Scale bar: 20 μm. **b** RPE lysates of *Cryba1* cKO and *Cryba1*^{fl/fl} control mice at different ages were collected and analyzed by western blotting for phosphorylated β-catenin and total β-catenin. Vinculin was used as an internal control. Graphs show decreased phosphorylated β-catenin at 4 and 9 months of age in the cKO cells. Statistical analysis was performed using a two-tailed unpaired Student's *t*-test. **P* < 0.05, ****P* < 0.001, *n* = 5–6.

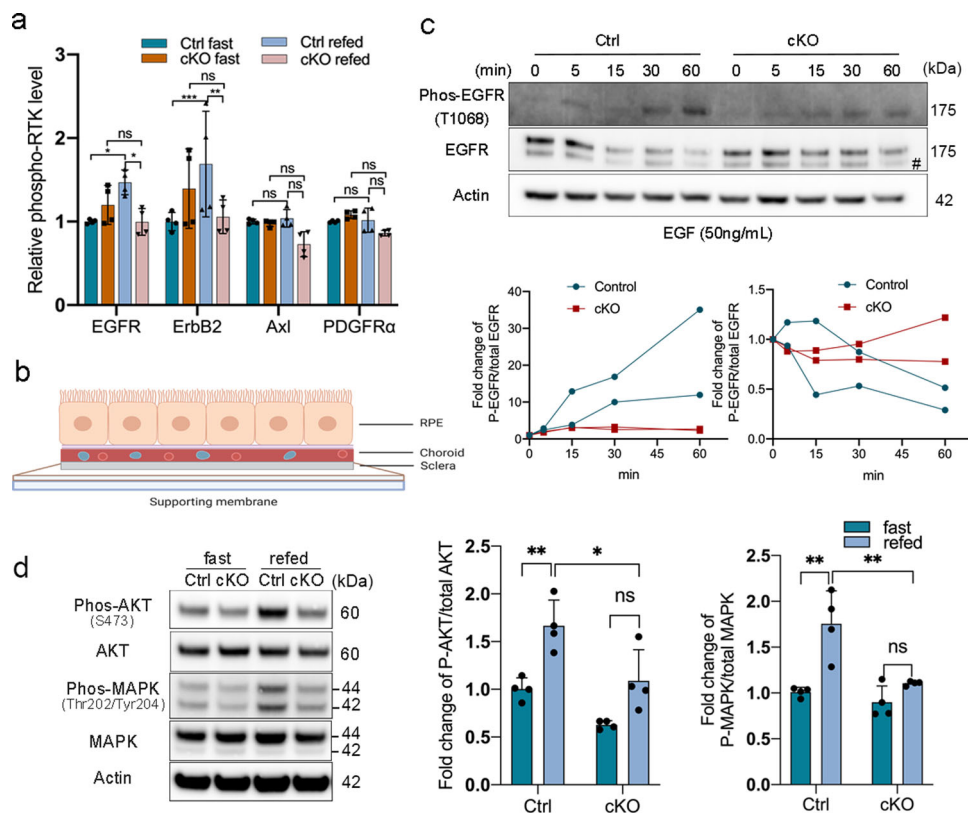


Fig. 4 Compromised EGFR activation in *Cryba1* cKO RPE. **a** RPE lysates from 2-month-old control and cKO mice fasted or fasted and then refed were isolated for a phospho-RTK array assay. Phosphorylated EGFR and ErbB2 were decreased in refed cKO RPE cells compared to refed floxed controls. $n = 4$. **b** A diagram showing RPE-choroid-sclera flatmount preparation. **c** RPE flatmounts of 2-month-old control and cKO mice stimulated with EGF for times indicated were analyzed by western blot. cKO RPE cells presented less phosphorylated EGFR in response to EGF stimulation at each time point. While total EGFR gradually decreased after ligand stimulation in control RPE cells, EGFR was not reduced in cKO RPE cells. Graphs show the EGFR phosphorylation level and total EGFR level in both cell types. Immunoblots are representative of two independent experiments. #: non-specific band. **d** Phosphorylated AKT and MAPK levels in RPE from fasted or refed mice were measured by western blot. Both phosphorylated AKT and MAPK were significantly upregulated in control RPE after refeeding, but not in cKO RPE. $n = 3$. Statistical analysis was performed using two-way ANOVA (**a**, **d**). * $P < 0.05$, ** $P < 0.01$, *** $P < 0.001$, ns: not significant.

activation implies an important physiological role of β A3/A1-crystallin/EGFR signaling in RPE.

In order to confirm that EGFR phosphorylation in cKO RPE cells is defective, we established an in vitro model to culture RPE explants from mice (see Methods for details). This method maintains RPE cells in their original milieu, while allowing in vitro manipulations. The RPE-choroid-sclera (RCS) complexes were carefully flattened on a supporting membrane and put into culture (Fig. 4b). RPE cell viability was determined by the level of the cell death marker, receptor interacting protein kinase 1 (RIP1) (Supplementary Fig. 2b). *Cryba1* cKO RPE cells in situ had reduced levels of phosphorylated EGFR following EGF stimulation. While total EGFR levels were decreased after ligand stimulation in floxed control RPE cells due to degradation⁴³, their levels were unchanged in *Cryba1* cKO RPE cells (Fig. 4c and Supplementary Fig. 2c) because of compromised EGFR activation. Interestingly, we observed that EGFR is activated at later time points (30–60 min post EGF stimulation) in cultured RCS complexes compared to the mouse RPE cell line (HPV E6/E7). The RCS model data are in sharp contrast to several previous reports using multiple cell lines^{44,45} including HPV E6/E7 RPE cells, that maximum EGFR activation occurs within 5–10 min after ligand stimulation (Supplementary Fig. 2d and Fig. 4c).

To better delineate if an EGFR activation defect would influence downstream signaling pathways, we measured the phosphorylation of AKT1 and MAPK, both well-known EGFR

downstream effectors⁴⁶, and found that their phosphorylation was upregulated in native RPE cells of control mice 3 h after refeeding, but not in *Cryba1* cKO RPE cells (Fig. 4d).

Since EGFR activation is perturbed in *Cryba1* cKO RPE cells, we asked if clathrin-mediated endocytosis of EGFR is also compromised. The high melanin content and autofluorescence of the RPE impairs visualization of the RPE apical surface⁴⁷. Therefore, we developed a live-cell technique to image clathrin-mediated endocytosis from the apical side of RPE cells using total internal reflection fluorescence (TIRF) microscopy. As shown in Fig. 5a, RPE flatmounts were cultured in glass-bottom dishes and infected with two recombinant human serotype 5 adenovirus vectors with deleted E1/E3. One of the vectors contained a clathrin light chain A construct with a CMV promoter and a fluorogen-activated peptide (FAP) reporter (Ad-CMV-FAP-mClita). This reporter only fluoresces after being activated by a fluorogen such as MG-ester, a far-red, cell-permeable FAP fluorogenic dye. The other vector contained a LifeAct-GFP fusion protein to fluorescently label the F-actin cytoskeleton. Infection efficiency was similar for control and *Cryba1* cKO RPE flatmounts (Supplementary Fig. 3a). After starving in fetal bovine serum (FBS)-free growth medium, RPE flatmounts were stimulated with EGF. To exclude the possibility that the clathrin signals were from non-specific autofluorescent particles, RPE flatmounts were incubated with or without MG-ester dye. Those without MG-ester dye showed no signal, which confirmed specificity (Supplementary Fig. 3b). The TIRF

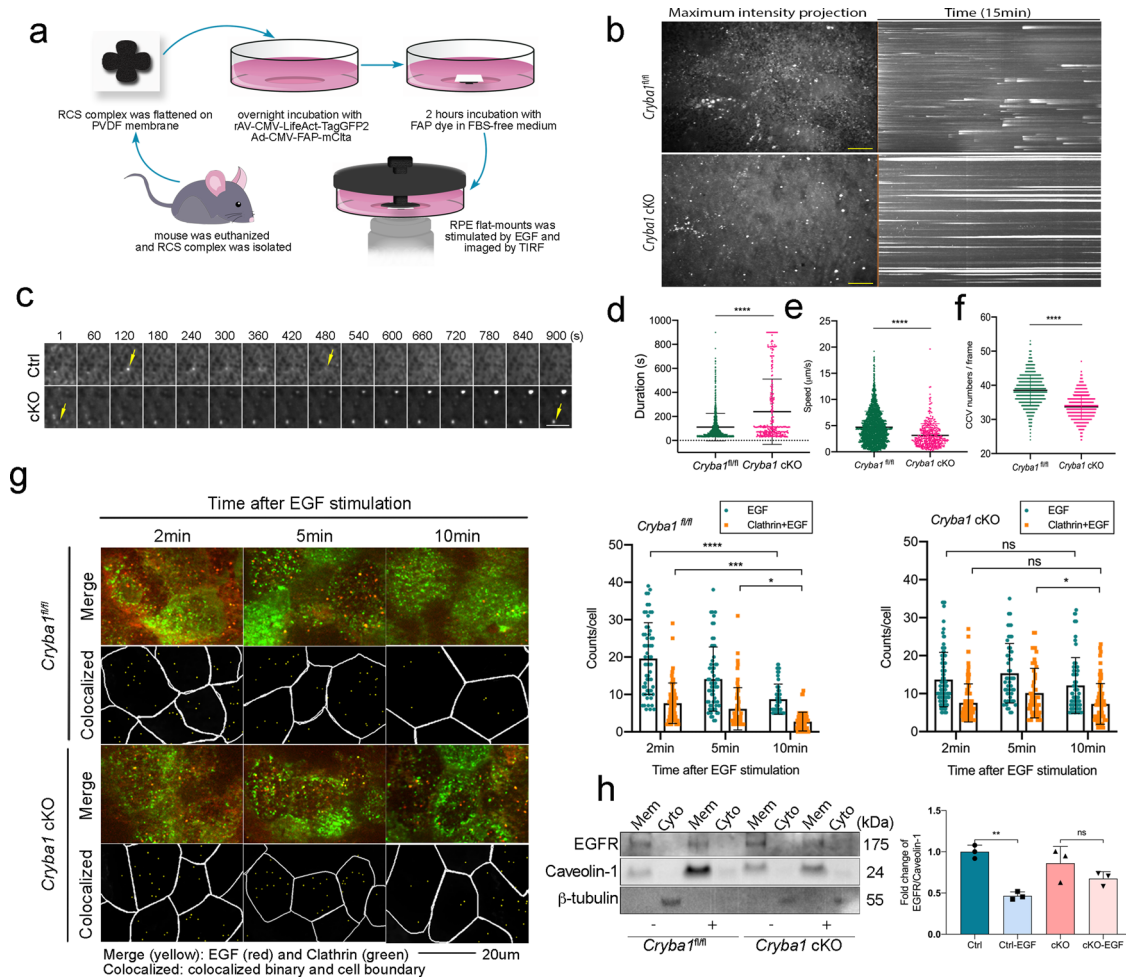


Fig. 5 Compromised clathrin-mediated EGFR internalization in *Cryba1* cKO RPE. **a** A diagram showing sample preparation for TIRF and HiLo imaging of endocytosis on the apical side of RPE cells in situ. **b** RPE flatmounts from 2-month-old control and cKO mice were infected with Ad-CMV-*mClna*-FAP and rAV-CMV-LifeAct-TagGFP2 and stimulated with EGF. Representative time-lapse TIRF imaging of clathrin from four biological repeats is shown in maximum intensity projection (x - y axis). The corresponding kymographs (x - t axis projection) show that a significant number of CCPs in *Cryba1* cKO RPE were stagnant at the apical side of the membrane and did not pinch off from the plasma membrane, indicated by long lines. Scale bar: 10 μ m. **c** A representative CCP from each cell type was tracked over the acquisition time. As time proceeded, the CCP in the control RPE cell pinched off from the membrane after showing up in the imaging frame as the arrows show. However, the CCP in the cKO RPE cell was stuck at the cell membrane during the acquisition time window. Scale bar: 5 μ m. **d** Duration of CCPs in cKO (CCP number = 310) and control RPE (CCP number = 1217) captured by TIRF. CCPs with a duration time less than 30 s were filtered out for this analysis. **e** Speed of each CCP in control (CCV number = 1639) and cKO (CCV number = 590) captured by HiLo microscopy were plotted. **f** CCVs were counted in each frame of live-cell imaging of *Cryba1* cKO RPE and control RPE samples. n (frames) = 1000. **g** RPE flatmounts from 2-month-old control and cKO mice infected with Ad-CMV-*mClna*-FAP and rAV-CMV-LifeAct-TagGFP2 were collected at times indicated after Alexa-Fluo-555 EGF stimulation and imaged by TIRF microscopy. EGF and EGF-loaded CCPs represented by colocalization of clathrin and EGF were counted, respectively. n (cell number) = 54–90. Data suggest a failure of EGF-loaded CCPs to internalize into the cell in the cKO RPE. **h** Membrane and cytoplasmic protein fraction were separated from RPE flatmounts stimulated with 50 ng/mL EGF for 30 min. Western blot analysis were performed to detect EGFR level in membrane fraction from different cell types. Caveolin-1 and β -tubulin were used as membrane and cytoplasm marker, respectively. The plot shows the EGFR level in membrane fraction significantly decreased in control RPE cells due to internalization, but not in *Cryba1* cKO RPE cells. $n = 3$. Statistical analysis was performed using a two-tailed unpaired Student's t -test (**d-f**), two-way ANOVA (**g**), and one-way ANOVA (**h**). * $P < 0.05$, ** $P < 0.01$, *** $P < 0.001$, **** $P < 0.0001$, ns: not significant.

time-lapse imaging data showed that within a 15 min timeframe, the clathrin-coated pits (CCPs) of *Cryba1* cKO RPE cells remained at the apical membrane without pinching off from the plasma membrane, while the CCPs in the floxed control cells remained at the apical membrane only briefly before internalization (Supplementary Movies 1 and 2 and Fig. 5b–d). To better depict this difference, we generated a kymograph of representative images from floxed control and *Cryba1* cKO samples, where solid, horizontal lines depict CCPs staying on the cell membrane (Fig. 5b). The data suggest that a significant number of CCPs in cKO RPE cells have a relatively long duration time at the apical membrane.

We also employed highly inclined and laminated optical sheet (HiLo) microscopy to image the area beneath the apical surface. HiLo imaging also showed that movement of clathrin-coated vesicles was dynamic in floxed control RPE cells, but slower in *Cryba1* cKO RPE cells (Supplementary Movies 3 and 4 and Fig. 5e). In addition, fewer and larger clathrin-coated vesicles were observed in *Cryba1* cKO RPE cells compared to control RPE cells (Supplementary Movies 3 and 4 and Fig. 5f). The above observations suggest a critical role of β A3/A1-crystallin not only in clathrin-mediated EGFR endocytosis, but also in the membrane trafficking dynamics of polarized RPE cells.

To determine if the CCPs that failed to pinch off from the plasma membrane were properly loaded with the cargo (EGF) and if EGFR internalization was hindered in the RPE due to loss of β A3/A1-crystallin, RPE flatmounts from *Cryba1* cKO mice were infected with Ad-CMV-m*Ctla*-FAP and rAV-CMV-LifeAct-TagGFP2, stimulated with Alexa-Fluo 555 EGF for 2 min after serum starvation, and fixed at 2, 5, and 10 min after EGF stimulation. Using TIRF microscopy, the number of EGF-loaded CCPs at the apical side gradually decreased over the tracking time in control RPE cells (Fig. 5g). However, a significant number of EGF-loaded CCPs remained at the apical side of *Cryba1* cKO RPE cells at 10 min post EGF stimulation, suggesting a failure of EGF-loaded CCPs to pinch off from the cell membrane of *Cryba1* cKO RPE cells (Fig. 5g). We also separated membrane fractions from lysates of the RPE flatmounts from control and cKO mice with or without EGF stimulation and detected EGFR in these fractions (Fig. 5h). We observed decreased membrane EGFR after 30 min of EGF stimulation in control RPE flatmounts. However, we did not observe significant reduction of membrane EGFR in *Cryba1* cKO RPE flatmounts. These data indicate that less EGFR was degraded in cKO RPE cells compared to the control, and further imply that the defective EGFR internalization might lead to less degradation in the RPE lacking β A3/A1-crystallin.

Interestingly, we found that a phosphorylation proteomics experiment we had performed previously showed some abnormally phosphorylated endocytosis-related proteins, such as formin-binding protein 17 (FBNP1) and amphiphysin II (BIN1), which are key mediators in CCP fission⁴⁸ in *Cryba1* cKO RPE cells (Supplementary Table 1). We wondered if insufficient CCP fission was the rate-limiting factor preventing CCP pinching off from the plasma membrane in *Cryba1* cKO RPE cells. Therefore, we measured the FBNP1 activity during clathrin-mediated endocytosis in control and cKO RPE flatmounts. RPE flatmounts from *Cryba1* cKO and floxed control mice were infected with rAV-CMV-LifeAct-TagGFP2, Ad-CMV-FAP-m*Ctla*, and Ad-CMV-m*Fnbp1*, and stimulated with EGF. With time-lapse imaging by TIRF microscopy, CCPs that were trapped at the apical membrane surface co-localized with FBNP1 in *Cryba1* cKO RPE cells, indicating defective fission in RPE cells of *Cryba1* cKO mice (Supplementary Movies 5 and 6 and Supplementary Fig. 3c, d). We speculate that the defective EGFR internalization may be due to the insufficient CCP fission. How β A3/A1-crystallin regulates EGFR activation and internalization remains to be determined.

β A3/A1-crystallin interacts with PITP β and may regulate PIP levels in RPE cells. Having demonstrated that β A3/A1-crystallin has a role in the regulation of the apical cytoskeleton and in EGFR endocytosis, we investigated in our human proteome high-throughput array for upstream regulators that could interact with β A3/A1-crystallin to initiate these effects, and identified a very significant binding signal between β A3/A1-crystallin and PITP β (phosphatidylinositol transfer protein beta isoform)²³. PITP β catalyzes the transfer of phosphatidylinositol and phosphatidylcholine between membranes and participates in PL metabolism during signal transduction and vesicular trafficking⁴⁹. As mentioned above, a single *Cryba1* mRNA uses alternative translation start sites to produce β A3 and β A1-crystallin isoforms by leaky ribosomal scanning²⁸. Since we identified multiple functions of this protein in RPE cells, and we have also previously found that β A3- and β A1-crystallins have different roles in astrocytes⁵⁰, we wondered if the binding to PITP β was isoform-specific. Using a co-immunoprecipitation assay on mouse RPE flatmounts overexpressing PITP β and either the β A1- or β A3-crystallin isoform by adenovirus constructs, we confirmed that

PITP β binds to both isoforms (Fig. 6a). The binding was increased after EGF stimulation. A significant colocalization of PITP β and β A3/A1-crystallin was also observed in ARPE19 cells overexpressing PITP β and β A3/A1-crystallin either with or without EGF stimulation (Fig. 6c). Moreover, molecular modeling predicted an interaction of PITP β with either β A3- and β A1-crystallin (Fig. 6b), with β A1 likely the better docking isoform.

One of the main functions of PITP β in cellular signaling is to present phosphatidylinositol to lipid kinases for localized production of PI(4,5)P₂⁵¹. Therefore, we measured PI(4,5)P₂ in floxed control and cKO RPE cells. PI(4,5)P₂ is the dominant phosphoinositide at the cell plasma membrane and the most important phosphoinositide for endocytosis⁵². Interestingly, we found a significantly decreased level of PI(4,5)P₂ in *Cryba1* cKO RPE compared to control RPE (Fig. 6d). It has been reported that PITP β and other PITPs enhance phospholipase C (PLC) activity, which mediates PI(4,5)P₂ hydrolysis^{53–55}. Notably, a recent study indicates that Sec14I3, a PITP family member, regulates the PI(4,5)P₂ level by modulating PLC activation by a mechanism not involving PI transfer activity⁵⁴. Therefore, we postulated that β A3/A1-crystallin may affect PI(4,5)P₂ levels by influencing PLC activity. As expected, phosphorylated PLC γ 1, which catalyzes the reaction of PI(4,5)P₂ into second messengers inositol triphosphate (IP3) and diacylglycerol (DAG), was upregulated in *Cryba1* cKO RPE (Fig. 6e), indicating enhanced PI(4,5)P₂ hydrolysis in cKO RPE cells. IP3 and DAG, the hydrolysis products of PI(4,5)P₂, activate protein kinase C (PKC)⁵⁶. Using the phospho-PKC (pan) (β II Ser660) antibody that detects PKC α , β I, β II, δ , ϵ , η , and θ isoforms only when phosphorylated at a carboxyl-terminal residue homologous to serine 660 of PKC β II, we found that PKC was significantly phosphorylated in *Cryba1* cKO RPE (Fig. 6f), consistent with the decreased levels of PI(4,5)P₂ observed. We also found significantly reduced levels of phosphatidylinositol (3,4,5)-trisphosphate (PI(3,4,5)P₃) in *Cryba1* cKO RPE compared to control cells. This may be due to reduced levels of PI(4,5)P₂, since PI(4,5)P₂ serves as a substrate for phosphoinositide 3-kinase (PI3K) when generating PI(3,4,5)P₃⁷. The decrease in PI(3,4,5)P₃ in *Cryba1* cKO RPE could also be due to reduced EGFR activation in *Cryba1* cKO RPE (Fig. 2b) because PI3K can be activated by phosphorylated EGFR⁵⁷. It has been well-documented that PLC interacts with EGFR and relays signals from activated EGFR⁵⁸. However, it has also been reported that PKC, the downstream effector of PLC, counteracts EGFR dimerization, acting as a negative feedback regulator of EGFR signaling⁵⁹. Here, we found that in the absence of β A3/A1-crystallin, PLC γ 1 signaling is activated while EGFR activation is compromised, which indicates that the β A3/A1-crystallin/PLC γ /PKC axis may play a pivotal role in EGFR activation at the apical surface of RPE cells.

Moreover, we observed that myo-inositol, an important component of PI and PIPs⁶⁰, was also lower in *Cryba1* cKO RPE-choroid samples compared to controls, indicating an essential role of β A3/A1-crystallin in maintaining the PI(4,5)P₂ pool (Fig. 6g). It is known that PI(4,5)P₂ participates in CCP assembly and fission, activating EGFR, and potentiating its subsequent internalization^{48,61}. We propose that the compromised EGFR activation and the defects in clathrin-coated structures in *Cryba1* cKO RPE cells are due to decreased PI(4,5)P₂. Since PI(4,5)P₂ is required for activating ezrin to link microfilaments to the plasma membrane⁴, the loss of PI(4,5)P₂ could also induce microvilli disorganization observed in *Cryba1* cKO RPE cells.

β A3/A1-crystallin attenuates PLC γ signaling, thereby activating ezrin phosphorylation and the EGFR signaling network. In order to confirm our findings that β A3/A1-crystallin regulates PI

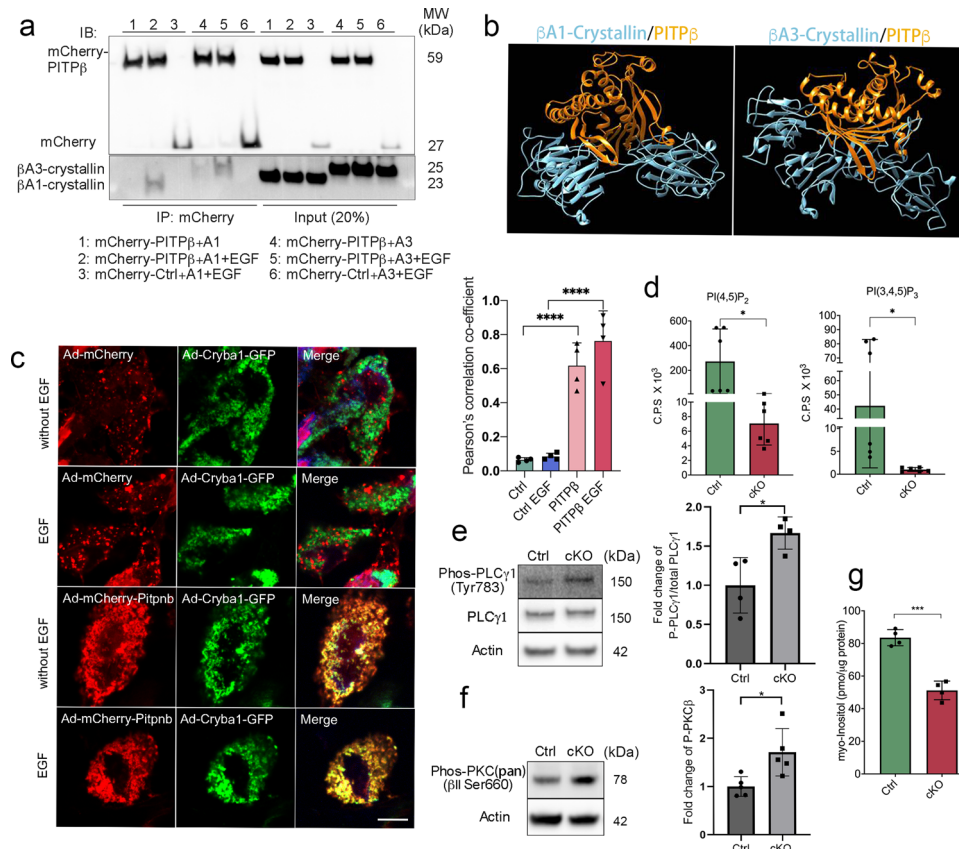


Fig. 6 β A3/A1-crystallin interacts with PITP β and may regulate PIP levels in RPE cells. **a** RPE flatmounts isolated from wildtype mice were cultured and infected either with Ad-mCherry-*Pitpnb* or Ad-mCherry-control and Ad-*Cryba1*-A3 or Ad-*Cryba1*-A1, respectively. RPE flatmounts were starved in FBS-free medium for 2 h with or without 30 min of EGF stimulation. RPE cell lysates were subjected to co-immunoprecipitation using RFP magnetic beads and the immunoprecipitants were analyzed by western blotting. Data show interaction of PITP β with β A3/A1-crystallin, especially during EGF stimulation. **b** Protein complexes of β A3-crystallin/PITP β and β A1-crystallin/PITP β as predicted by molecular modeling. β A1-crystallin (left) and β A3-crystallin (right) are shown in blue. PITP β is shown in orange. Both β A3- and β A1-crystallins are represented as the dimers formed by two “open” conformers. **c** ARPE19 cells were infected with Ad-mCherry-*Pitpnb* or Ad-mCherry-control and Ad-*Cryba1*-GFP. Ad-*Cryba1*-GFP vector contains the wildtype *Cryba1* gene, and produces both β A3-crystallin and β A1-crystallin. Cells were treated with or without 10 ng/mL EGF for 10 min and fixed for imaging and colocalization analysis. Data show the colocalization of PITP β and β A3/A1-crystallin, especially during EGF stimulation. Scale bar: 10 μ m. **d** PI(4,5)P₂ and PI(3,4,5)P₃ were analyzed on RPE-choroid complexes of 2-month-old control and cKO mice. Data indicate a markedly reduced amount of PI(4,5)P₂ and PI(3,4,5)P₃ in cKO RPE-choroid complexes compared to that of control. $n = 6$. **e** Phosphorylated PLC γ 1 levels were determined in RPE lysates of 3-week-old control and cKO mice by western blot. Data suggest increased PLC γ 1 activity in cKO RPE cells. $n = 4$. **f** Phosphorylated PKC (pan) (β II Ser660) levels were determined in RPE lysates of 3-week-old control and cKO mice. Data suggest the activation of PKC in cKO RPE cells. $n = 4$. **g** Myo-inositol was analyzed on RPE-choroid complexes of 2-month-old control and cKO mice. Data indicate reduced myo-inositol level in cKO RPE-choroid complexes compared to that of control. $n = 4$. Statistical analysis was performed using two-tailed unpaired Student’s *t*-test (**d-g**) and one-way ANOVA (**c**). * $P < 0.05$, *** $P < 0.001$, **** $P < 0.0001$.

(4,5)P₂ and PI(3,4,5)P₃ levels and is involved in the EGFR signaling network, we overexpressed β A3/A1-crystallin in *Cryba1* cKO RPE flatmounts using a *Cryba1* adenovirus construct. We found that phosphorylated PLC γ 1 and PKC were decreased in *Cryba1* cKO RPE cells with *Cryba1* overexpression (Fig. 7a). Importantly, when stimulated by EGF, EGFR phosphorylation was significantly upregulated in RPE cells with *Cryba1* overexpression compared to the cells infected with control constructs. The EGFR downstream effectors AKT and MAPK were also activated after *Cryba1* overexpression (Fig. 7b). These data confirmed that β A3/A1-crystallin promotes EGFR activation and its downstream signaling pathways. In RPE flatmounts with either *Cryba1* or control expression vectors, we found that the PI(3,4,5)P₃ level was significantly increased when *Cryba1* was overexpressed (Fig. 7d). However, unexpectedly, the PI(4,5)P₂ level was decreased in cKO RPE cells overexpressing β A3/A1-crystallin compared to that in cKO RPE cells with control vectors (Fig. 7c). It is possible that the PI(4,5)P₂ pool is highly dynamic and the EGFR/PI3K/AKT signaling pathway is activated after *Cryba1*

overexpression, and that more PI(4,5)P₂ is converted to PI(3,4,5)P₃, accounting for the decrease in PI(4,5)P₂. In addition, both control and cKO RPE flatmounts have elevated ezrin phosphorylation levels following *Cryba1* overexpression (Fig. 7e). Overall, our data demonstrate that β A3/A1-crystallin regulates PIP levels by modulating PLC γ activity, thus activating EGFR signaling and ezrin phosphorylation in RPE cells.

Discussion

In this study we made the unique observation that in RPE cells, β A3/A1-crystallin regulates PIP metabolism, most likely by binding to PITP β , a protein that transfers phosphatidylinositol and phosphatidylcholine between membranes, and presents phosphatidylinositol to lipid kinases for localized production of PI(4,5)P₂. Mutations in PITP β or other PITP proteins such as rdgB (retinal degeneration B) have been shown to result in retinal degeneration in both invertebrate and vertebrate models^{62–66}.

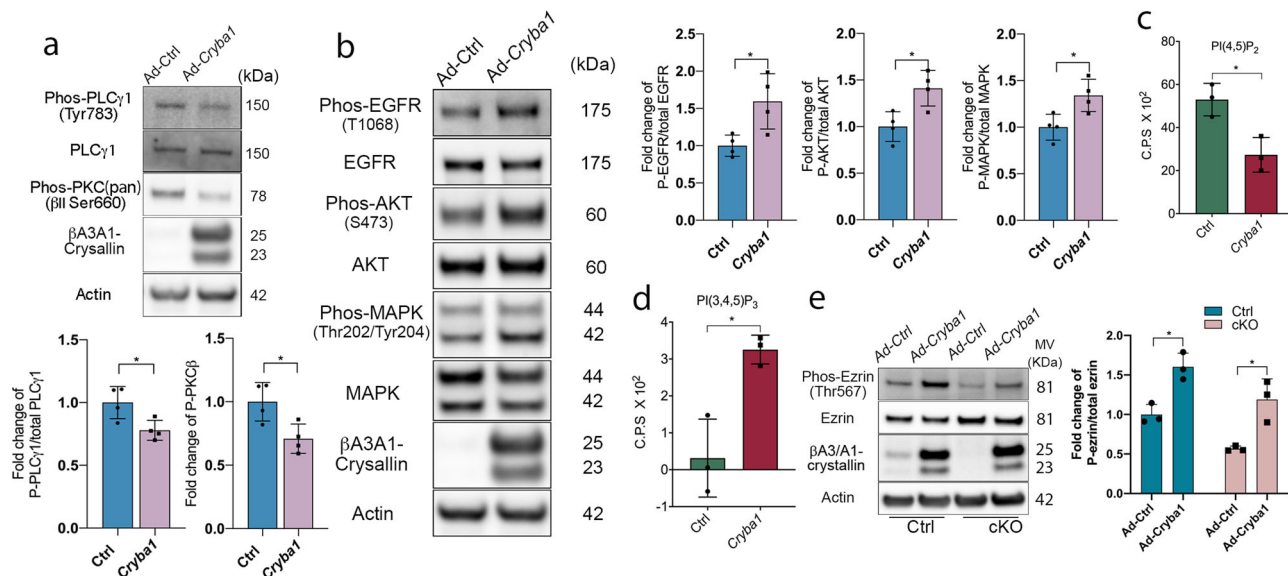


Fig. 7 β A3/A1-crystallin attenuates PLC γ signaling and activates ezrin phosphorylation and the EGFR signaling network. **a** RPE flatmounts from 2-month-old *Cryba1* cKO mice were cultured and infected with either Ad-RFP-*mCryba1* or Ad-CMV-RFP control viral vector. Western blot analysis was performed on lysates to detect PLC γ 1 and PKC activity. Graphs show reduced PLC γ 1 and PKC phosphorylation in *Cryba1* cKO RPE cells overexpressing *Cryba1*. **b** RPE flatmounts from 5-month-old *Cryba1* cKO mice were cultured and infected with either Ad-RFP-*mCryba1* or Ad-CMV-RFP control viral vectors. Flatmounts were stimulated with 50 ng/mL EGF for 60 min after serum starvation. Cells were then lysed and western blot analysis was performed on the lysates to detect EGFR phosphorylation and its downstream signaling molecules. Upregulated phosphorylation of EGFR, AKT1, and MAPK was observed in *Cryba1* cKO RPE cells overexpressing *Cryba1*. **c, d** RPE flatmounts from 2-month-old *Cryba1* cKO mice were cultured and infected with either Ad-RFP-*mCryba1* or Ad-CMV-RFP control viral vector. PI(4,5)P₂ (**c**) and PI(3,4,5)P₃ (**d**) levels were analyzed. **e** RPE flatmounts from 2-month-old *Cryba1* cKO mice and control mice were cultured and infected with either Ad-RFP-*mCryba1* or Ad-CMV-RFP control viral vector. Western blot analysis was performed on lysates to detect ezrin phosphorylation. Graph shows increased phosphorylation of ezrin in both control and *Cryba1* cKO RPE cells overexpressing *Cryba1*. *n* = 3. The error bars represent the SD of the mean. The statistical analysis was performed using two-tailed unpaired Student's *t*-test (**a–d**) and two-way ANOVA (**e**). **P* < 0.05.

In RPE lacking β A3/A1-crystallin, we found decreased PI(4,5)P₂ levels and increased activity of phospholipase C, the enzyme that hydrolyzes PI(4,5)P₂⁵⁸. In addition, upon overexpression of β A3/A1-crystallin in RPE flatmounts, a reduction in PLC/PKC activity was observed. We speculate that β A3/A1-crystallin regulates PLC activity by influencing PIP β activity. Previously, PIPs were shown to enhance PLC activity^{53–55}. We also postulate that PI(4,5)P₂ levels are responsive to β A3/A1-crystallin through the PIP β /PLC axis. While dysregulation of PIP metabolism in photoreceptors has been reported to cause blindness^{8–12}, the roles of PIPs in RPE function and physiology remain uncertain.

PI(4,5)P₂ is the most abundant phosphoinositide at the plasma membrane, and it binds to proteins that are required for actin filament assembly (e.g., ezrin/radixin/moesin, profilin, cofilin, etc.) and endocytosis (e.g., picalm, dynamin, epsin, etc.)⁶⁷. We propose that PI(4,5)P₂ is essential for maintaining the RPE apical cytoskeletal network and normal endocytosis. PI(4,5)P₂ is the most important phosphoinositide for endocytosis;⁵² it is required for EGFR activation and clathrin-mediated endocytosis⁶¹. Indeed, we found compromised EGFR activation and internalization following EGF stimulation in RPE cells that lack β A3/A1-crystallin. With TIRF imaging of RPE flatmounts, we observed impaired CCP fission and EGFR internalization. Importantly, the defective EGFR activation in *Cryba1* cKO RPE cells can be rescued by overexpressing β A3/A1-crystallin. Interestingly, while a β A3/A1-crystallin deficiency altered the phosphorylation of EGFR (and the related ErbB2), it did not affect other RTKs. We speculate that during evolution, β A3/A1-crystallin obtained the specific ability to regulate EGFR signaling in RPE cells, and that the molecular coordination of this signaling pathway is unique to

RPE cells. This may be analogous to the recent postulation that the RPE has a specialized and unique clearance machinery that differs from traditional phagocytic cells⁶⁸. The possible role of β A3/A1-crystallin in the EGFR signaling network is demonstrated in Fig. 8a. Our data showed that a β A3/A1-crystallin deficiency in RPE cells led to increased PLC γ /PKC activity, thus decreased PI(4,5)P₂ and EGFR activation, whereas β A3/A1-crystallin overexpression inhibited phosphorylation of PLC γ 1, probably resulting in a rapid increase in PI(4,5)P₂ levels, which led to EGFR activation. Meanwhile, the decreased PKC activity in the presence of β A3/A1-crystallin might disrupt the negative feedback effect on EGFR dimerization. Due to activated EGFR, the conversion of PI(4,5)P₂ to PI(3,4,5)P₃ was enhanced because of AKT activation by PI3K (Fig. 8a). Interestingly, it was previously reported that PIP is required for EGFR signaling and that it can bind to EGFR and PLC γ in response to EGF stimulation. Therefore, we propose here that β A3/A1-crystallin modulates EGFR activation by influencing PIP β /PLC γ activity (Fig. 8a). The EGFR activation defects might also contribute to RPE polarity loss and RPE degeneration; however, this possibility needs to be investigated in the future (Fig. 8b).

Furthermore, PI(4,5)P₂ is known to bind to ezrin, a member of the ezrin/radixin/moesin (ERM) family of proteins. ERM proteins provide a critical link between the plasma membrane and the underlying actin cytoskeleton. Ezrin is highly expressed in epithelial cells that have microvilli, such as RPE. Its phosphorylation is regulated by PI(4,5)P₂, and it is essential for establishing microvilli and maintaining polarity⁴. Here, we show that β A3/A1-crystallin is required by RPE cells to maintain normal ezrin phosphorylation and polarity. When β A3/A1-crystallin is absent, PI(4,5)P₂ and ezrin phosphorylation are also substantially

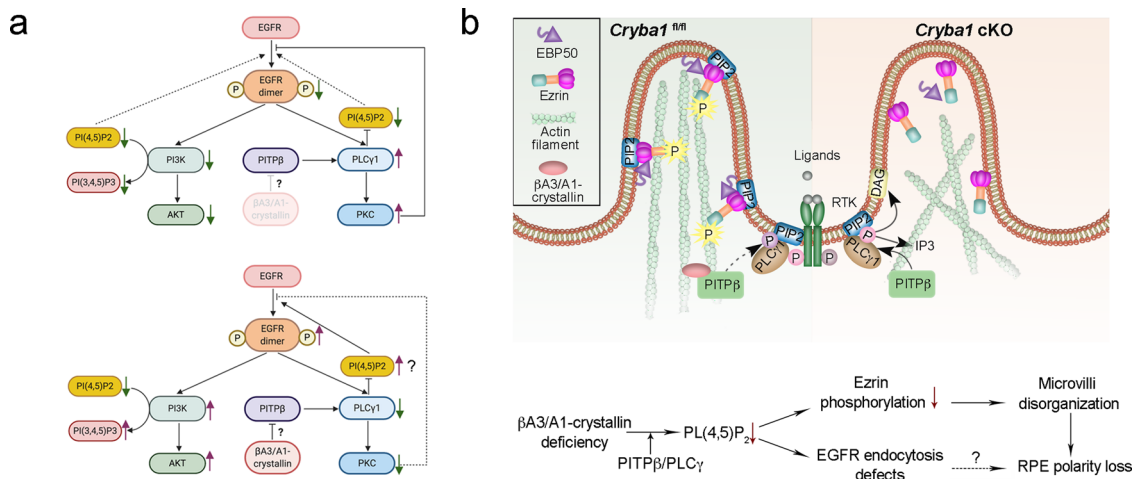


Fig. 8 The possible role of β A3/A1-crystallin in the EGFR signaling network and RPE polarity. **a** A model showing the possible role of β A3/A1-crystallin in the PITP β /PLC γ signaling axis and PI(4,5)P $_2$ and PI(3,4,5)P $_3$ turnover, and consequently in EGFR activation and internalization. Top: RPE cells lacking β A3/A1-crystallin have increased PLC γ 1 and PKC activity, which leads to decreased PI(4,5)P $_2$. EGFR activation is inhibited by the PI(4,5)P $_2$ deficiency and activated PKC through a negative feedback loop. Bottom: overexpression of β A3/A1-crystallin attenuated phosphorylated PLC γ 1, possibly by influencing PITP β activity. The PI(4,5)P $_2$ level may be temporarily increased. Having sufficient PI(4,5)P $_2$ promotes EGFR activation. On the other hand, decreased PKC activity may not affect EGFR dimerization. Due to the activation of EGFR, more PI(4,5)P $_2$ was converted to PI(3,4,5)P $_3$ because of the activation of AKT by PI3K, which may subsequently lead to PI(3,4,5)P $_3$ upregulation and PI(4,5)P $_2$ downregulation. **b** Cartoon depicting the pathway initiated by the β A3/A1-crystallin deficiency leading to RPE apical surface disruption and impaired EGFR endocytosis. Loss of functional β A3/A1-crystallin prevents its interaction with PITP β causing a modification of PIP metabolism that eventually impairs both EGFR endocytosis and ezrin phosphorylation, thereby altering the actin cytoskeleton and affecting RPE cell polarity.

reduced, and interaction between ezrin and the cytoskeleton is disrupted. As a result, morphological and functional changes at the apical surface are altered, and RPE cells lose polarity (Fig. 8b). In this study, we provide direct evidence that β A3/A1-crystallin affects microvilli organization by modulating PI(4,5)P $_2$ and ezrin phosphorylation.

These changes are highly relevant to AMD, the most common cause of blindness among the elderly in Western societies⁶⁹. Rod photoreceptor death secondary to RPE degeneration in the perifoveal macula is one of the earliest changes in AMD⁷⁰, and loss of apical microvilli is an early sign of this process⁵. With a similar RPE and rod/cone density as the human perifovea, mice are a reasonable model for studying early AMD changes⁷¹. Thus, the apical microvilli changes in the RPE of *Cryba1* cKO mice, an established model of early perifoveal AMD-like pathobiology, provide us with new avenues for investigating both how the RPE loses its polarity, especially in the apex, and the impact that this loss has on the health of both the RPE and the overlying photoreceptors during disease onset and progression. Interestingly, loss of apical processes is considered integral to the loss of RPE polarity in atrophic AMD. Immunohistochemistry studies have shown decreased expression of ezrin in the macula of geographic atrophy eyes⁷². By utilizing this unique animal model, we identified the role of β A3/A1-crystallin in regulating two processes most likely through modulating PI(4,5)P $_2$ pool at the apical side of RPE: ezrin's interaction with the actin cytoskeleton and membrane trafficking during EGFR endocytosis, which are absolutely essential for maintaining RPE cellular polarity. We previously reported the dysfunction of the retina in *Cryba1* cKO mice; assessment included fundus imaging and electroretinogram measurements²⁴. We can only speculate that the retina dysfunction is partially caused by the RPE polarity loss, since the lysosomal function is also impaired in the RPE cells in the *Cryba1* cKO model. In this study, we focus on the function of β A3/A1-crystallin in RPE polarity and the molecular mechanism underlying this regulation. It is worth investigating the detailed

consequences of RPE polarity disruption on retinal function in *Cryba1* cKO, but it is beyond the scope of the current study. Taken together, our data suggest a pathway initiated by a β A3/A1-crystallin deficiency that explains how apical microvilli become disorganized and how EGFR endocytosis is impaired during RPE degeneration in a model that is highly relevant to early AMD.

Finally, crystallins are intriguing for their multifunctionality in non-lens tissues as moonlighting proteins¹⁷. We have previously shown that β A3/A1-crystallin is localized to the lysosomal lumen in RPE cells, which is required for autophagy and phagocytosis^{21,24}. In this study, to the best of our knowledge, we, for the first time, identified the functions of β A3/A1-crystallin in the apical region of polarized RPE cells, including its regulation of PIP metabolism through interaction with PITP β , its role in EGFR signaling, and its role in ezrin activation and microvilli organization. We postulate that β A3/A1-crystallin may regulate diverse subcellular processes as a moonlighting protein. The binding of β A3/A1-crystallin and PITP β indicates its location in the Golgi and endoplasmic reticulum (ER). It is known that newly synthesized lysosomal proteins are first transferred into the ER lumen, followed by the Golgi apparatus and are then transported from the trans Golgi network to late endosomes. This is consistent with the β A3/A1-crystallin protein being localized to the ER and Golgi at some point of its existence. In addition, Ma et al. demonstrated that β A3/A1-crystallin is required for trafficking the anoikis regulator, Bit1, to the Golgi in the astrocytes, thus putting this protein's function in the context of processes that take place in the Golgi apparatus.

It is worth mentioning that *Cryba1* is unusual among eukaryotic genes in producing two proteins by leaky ribosomal scanning. In eukaryotes, genes with leaky ribosomal scanning produce functionally important protein isoforms with distinct roles⁷³. When translation is initiated using alternative start sites (β A3-crystallin has 17 additional N-terminal amino acid residues), it is predicted that the isoforms are delivered to different subcellular

compartments^{74,75}. While our studies to date do not address whether β A3- and β A1-crystallins are functionally different in RPE cells, we have shown β A3/A1-crystallin in different compartments of RPE cells and astrocytes^{23,24,50,76}. In future studies, we intend to determine whether the β A3- and β A1-crystallin isoforms possess different properties and functions at different subcellular locations.

Methods

Animals. β A3/A1-crystallin conditional knockout (*Cryba1* cKO) mice and control *Cryba1*^{fl/fl} mice were generated as previously described²⁴. All animal studies were conducted in accordance with the Guide for the Care and Use of Animals (National Academy Press) and were approved by the Animal Care and Use Committee of the University of Pittsburgh.

Immunostaining of retina cryosections and RPE flatmounts. Fresh eyes were enucleated and fixed in 2% paraformaldehyde (PFA) for 10 min followed by the removal of the anterior parts including cornea, lens, and the attached iris pigmented epithelium. The resulting posterior eyecups were fixed in 2% PFA for 1 h at room temperature either for cryosections or RPE flatmounts. For the retina cryosections, the eyecups were dehydrated using gradient sucrose solutions and embedded in OCT compound for sectioning. For the RPE flatmounts, the eyecups were quartered like a petaloid structure and the retinas were removed. The resulting RCS complexes or retina cryosections were permeabilized with 0.25% Triton X-100 in PBS for 15 and 10 min, respectively. RCS complexes or retina sections were blocked in blocking buffer (2% goat serum, 1% BSA, and 0.1% Triton X-100 in PBS) for 30 min at room temperature followed by incubation in EBP50 antibody (1:200, PA1090, Thermo Fisher Scientific) in blocking buffer in a humidified chamber overnight at 4°C. RCS complexes or retina sections were then washed with 1x PBS and incubated with the secondary antibody (1:200, Goat anti-Rabbit, Alexa Fluor 568, A-11011, Invitrogen), Alexa Fluor 488 Phalloidin (1 unit/mL, A12379, Invitrogen), and 1 μ g/mL DAPI (D1306, Thermo Fisher) in blocking buffer for 1 h in the dark at room temperature. After washing, RCS complexes were flattened on a microscope slide with the RPE layer up. Retina sections or RCS complexes were then mounted with DAKO mounting medium (S3023, Agilent) and sealed with a coverslip. Images were acquired on a Zeiss LSM 710 confocal workstation. Microvilli height was measured by the length measurement tool available in ZEN software (Zeiss, Germany) based on the staining results. Other antibodies used for immunostaining: MCT3 (1:200, AMT013, Alomone Labs), Sodium Potassium ATPase, (1:200, ab76020, Abcam), and β -catenin (1:200, ab16051, Abcam).

Transmission electron microscopy. Samples were prepared and transmission electron microscopy was performed as previously described²¹. Briefly, mouse eyes were enucleated immediately after CO₂ euthanasia; a small slit was made at the limbus and then fixative (2.5% glutaraldehyde with 2% PFA in 0.1 mol/L cacodylate buffer, pH 7.4) was injected into the globe. Each eye was then immersed in the same fixative solution. Anterior segments were removed and the resulting eyecups were washed in 0.05 mol/L cacodylate buffer and post-fixed in 1% OsO₄ in 0.05 mol/L cacodylate buffer for 90 min. The eyecups were then washed with 0.05% cacodylate buffer and dehydrated in a graded series of ethyl alcohol and stained en bloc with 1% uranyl acetate in 100% ethanol followed by two 15 min incubations in propylene oxide. The tissue was then kept overnight in a 1:1 propylene oxide and resin mixture. The tissue was then infiltrated in 100% LX112 resin (Ladd Research Industry, Burlington, VT) for 4–6 h under vacuum, embedded in a final change of 100% LX112 resin and then polymerized at 60°C for 36–48 h. Ultrathin sections were cut with a Leica Ultramicrotome UCT (Leica Microsystems, Wetzlar, Germany), stained with uranyl acetate and lead citrate and visualized with an H7600 transmission electron microscope (Hitachi, Tokyo, Japan).

Tissue collection and RPE protein preparation. Eyeballs were enucleated from euthanized mice, and anterior cornea, lens, and neural retina were removed. The resulting RCS complexes were snap-frozen on dry ice. RPE cells were mechanically isolated from RCS complexes by tapping for 30 s in RIPA lysis buffer (20–188, Millipore) with a mixed protease and phosphatase inhibitor cocktail as described⁷⁷. Choroid-sclera complexes were then removed from the lysate buffer. The resulting RPE cells were sonicated for 15 s on ice and incubated in lysis buffer for 15 min. RPE lysates were separated from the cell debris by centrifugation at 15,000 rpm for 20 min. The protein concentrations were determined by BCA assay.

Phospho-RTK array. Mice were fasted for 36 h by withholding food without restriction on water availability, with or without a subsequent 3 h refeeding. All mice were sacrificed between 10 am and 12 pm. The RPE lysates were extracted as described above. Each RPE sample was pooled from four mice that had received the same treatment. An equal amount of each sample was used to quantify phospho-RTK proteins with a proteome profiler mouse phospho-RTK array kit (ARY014,

R&D Systems). Quantification of the spot intensity in the arrays was conducted with Quantity One software (Bio-Rad).

RPE flatmount culture and EGF stimulation. Eyeballs were enucleated from 2-month-old mice immediately after euthanasia, and the anterior segments were removed thereafter. The resulting posterior eyecups were cut into four petals and the neural retina was carefully removed. The resulting RCS complexes were flattened onto PVDF membranes with the RPE cells facing up and cultured in a 37°C incubator in complete Dulbecco Modified Eagle Medium (DMEM), high glucose, supplemented with 2% FBS, 1% penicillin/streptomycin, 2.5 mM L-glutamine, and 1x MEM nonessential amino acids. The mouse HPV E6/E7 RPE cells (T0574, ABM) were cultured in Prigrow III Medium (TM003, ABM) with 10% FBS and 1% penicillin/streptomycin. Before EGF stimulation, RPE flatmounts/mouse HPV E6/E7 RPE cells were starved in FBS-free medium for 2 h. RPE flatmounts/mouse HPV E6/E7 RPE cells were then cultured in DMEM medium containing 50 ng/mL EGF (AF-100-15, Peprotech) for 5, 15, 30, and 60 min, respectively. RPE flatmounts were then immediately washed with PBS and snap-frozen on dry ice. RPE lysates were isolated from flatmounts as described above. Mouse HPV E6/E7 RPE cells were washed in PBS and collected for western blot analysis. For the positive control of cell death, RPE flatmounts were cultured in complete medium containing 10 and 100 μ M H₂O₂ for 16 h. RPE cell lysates were collected for western blot analysis.

EGFR endocytosis analysis by total internal reflection fluorescence (TIRF) microscopy and highly inclined and laminated optical sheet (HiLo) microscopy. For endocytosis analysis in live RPE cells, RPE flatmounts were cultured in complete medium in a glass-bottom dish and infected with recombinant human serotype 5 adenoviral constructs with deleted E1/E3. Ad-CMV-FAP-m*Clta* is a CMV promoter-driven vector that contains mouse *Clta* cDNA and a FAP reporter. rAV-CMV-LifeAct-TagGFP2 (60121, ibidi) was used for labeling the F-actin network. Ad-CMV-m*Fnbp1* was made by inserting the cloned mouse *Fnbp1* cDNA into the recombinant human serotype 5 adenoviral constructs. The flatmounts were infected at 5 \times 10⁶ PFU/mL of each adenovirus for 12–16 h. The RPE flatmounts were then starved in DMEM medium containing a cell-permeable FAP fluorogenic dye (MG-ester, 50 nM) for 2 h. The starvation medium was replaced with 50 ng/mL EGF containing medium immediately and the flatmounts were placed apical surface down against the coverslip of the glass-bottom dish and is gently compressed such that the apical surface enters the TIRF plane. The device consists of a 3D printed black lid that clips onto the imaging dish and a plunger that is axially positioned using a fine pitch thread. As the plunger was screwed down, the RPE flatmount (mounted on a non-moving, bearing mounted plate at the end of the plunger) came into contact with the coverslip and can be imaged. This was done on the microscope to ensure that there was no compressive damage to the cells as they were imaged. RPE flatmounts were immediately taken for TIRF time-lapse imaging. TIRF images of RPE flatmounts were taken on an inverted Nikon Ti with a Plan Apo TIRF 1.49 NA 100x oil objective and Prime 95b camera (Photometrics). Images were acquired with a 100 ms exposure time. All samples were imaged in a chamber at 37°C with 5% CO₂. Subsequent image analysis included a modified Richardson-Lucy deconvolution step and no other processing was performed. Kymographs of the images in maximum intensity projection were generated using NIS-Elements (Nikon). Duration analyses of CCPs were performed using Imaris software. CCPs with a duration time of less than 30 s were filtered out for the analysis. HiLo microscopy was conducted on the same microscope. Illumination used a 640 nm laser at an incident angle that allowed the lower ~0.5 μ m of the cell to be imaged. Images were acquired with a 100 ms exposure time. All samples were imaged in a chamber 37°C with 5% CO₂. Clathrin structures were determined with the spot detection function to create binary masks of clathrin by NIS-Elements. Speed and count analysis of CCVs were generated using NIS-Elements software. For EGFR endocytosis tracking analysis, clathrin and F-actin labeled mouse RCS complexes were transferred to DMEM medium containing a cell-permeable FAP fluorogenic dye (MG-ester, 50 nM) for 2 h. RCS complexes from *Cryba1* cKO and control mice were then stimulated with 50 ng/mL Alexa Fluor 555 EGF complex (E35350, Thermo Fisher Scientific) for 2 min in an incubator and then immediately moved either to PBS for washing and for subsequent fixation in 2% PFA or to FBS/EGF-free DMEM medium to extend the culture for extra 3 or 8 min for tracking purpose. Imaging quantification was performed in NIS-Elements. Clathrin and EGF localization and quantification were determined with the spot detection function to create binary masks of clathrin and EGF signals.

RPE membrane fraction assay. RPE flatmounts from 2-month-old *Cryba1* cKO and age-matched control mice were cultured and stimulated with 50 ng/mL EGF (as described above) for 30 min. RPE flatmounts were washed in PBS three times and immediately snap-frozen on dry ice. The RPE cells were collected in permeabilization buffer (89842, Thermo Fisher) as described above. Membrane and cytoplasmic protein fractions of RPE cells were obtained with the Mem-PER plus membrane protein extraction kit (89842, Thermo Fisher).

Phospho-proteomics. RPE samples were collected from 8-month-old *Cryba1* cKO and age-matched control mice. Each sample was pooled from four mice from the same group. RPE samples were digested by trypsin and the tryptic peptides were labeled using the TMT-10 plex labeling kit and fractionated by UHPLC (Poochon Scientific). The phosphorylated peptides were enriched from the fractions of TMT-10 plex labeled peptide using a TiO₂ column from Pierce and analyzed by LC/MS/MS. The LC/MS/MS analysis of samples was carried out using a Thermo Scientific Q-Exactive Hybrid Quadrupole-Orbitrap Mass Spectrometer and a Thermo Dionex UltiMate 3000 RSLCnano System. For protein identification, MS files were analyzed using the Thermo Proteome Discoverer 1.4.1 platform (Thermo Scientific, Bremen, Germany) for peptide identification and protein assembly. A database search against public mouse proteins obtained from the NCBI website was performed based on the SEQUEST and percolator algorithms through the Proteome Discoverer 1.4.1 platform. The maximum false peptide discovery rate was specified as 0.01.

RFP pulldown experiment. To produce adenovirus constructs for the β A1 isoform, the first codon of *Cryba1* was mutated and a silent mutation was also introduced to prevent the binding and re-cutting of the sequence by gRNA after homology-directed repair. To produce the longer (β A3) isoform, CCACC was inserted immediately before the first start site to strengthen the Kozak consensus sequence, thereby making the first start codon stronger. The second mutation was introduced to disrupt the possible Kozak sequence that was located upstream of the A1 start codon. Wildtype mouse RPE flatmounts were infected with Ad-CMV-mCherry-*Pitpnb* (or Ad-CMV-mCherry-control) and either Ad-CMV-A3 or Ad-CMV-A1 for 16 h. RPE flatmounts were then stimulated with 50 ng/mL EGF for 30 min. RPE lysates were collected. Cell extracts were incubated with RFP magnetic beads (rtmak-20, Chromotek) for 2 h at 4 °C. Collected beads were washed with washing buffer (10 mM Tris/Cl pH 7.5, 150 mM NaCl, 0.5 mM EDTA, and 0.018% sodium azide) and eluted with 2x SDS-sample buffer. The eluted samples were analyzed in a SDS-PAGE gel.

ARPE19 cell culture and colocalization analysis. ARPE19 cells were obtained from American Type Culture Collection (Manassas, VA), and cultured in DMEM/F12 medium supplemented with 10% FBS and 1% penicillin/streptomycin. Cells were infected with Ad-CMV-m*Cryba1*-GFP and either Ad-CMV-mCherry-*Pitpnb* or Ad-CMV-mCherry control constructs for 24 h. Cells were then starved in FBS-free medium for 2 h and stimulated with or without 10 ng/mL EGF for 10 min. The colocalization analysis for dual-color observations was performed by using Fiji ImageJ software.

Molecular modeling. The structures of β A3-crystallin, β A1-crystallin, and PITP β were built by homology modeling using Yasara (<http://www.yasara.org/>). The molecules were docked and the complex structure was energy minimized in ucsc chimera (<https://www.cgl.ucsf.edu/chimera>).

Phosphoinositide extraction and measurement. RPE-choroid complexes were isolated from 2-month-old *Cryba1* cKO and age-matched control mice. Each sample was pooled from four mice from the same group. All mice were sacrificed between 10 am and 12 pm. Mouse RPE was homogenized in PBS, and 2:1 chloroform:methanol was added to extract the phosphatidylinositol (PI). The extraction was repeated with chloroform to remove bulk PLs (PE, phosphatidylethanolamine; PC, phosphatidylcholine; PS, phosphatidylserine). The chloroform layers were pooled into one tube, which represents the PLs (PE, PC, and PS). This fraction does not contain phosphoinositides. To the remaining solution, chloroform:methanol:HCl (2:4:0.1) was added to extract the PI. The procedure was repeated with chloroform:methanol:HCl. Then, the chloroform layers were pooled into a clean tube, which represents the PI. The PI pooled fractions were dried under nitrogen gas and the dried lipids were dissolved in 150 μ L of 1:9 chloroform:methanol, and sonicated. We measured lipid phosphorus from the PI fractions according to the method described⁷⁸ and converted the determined lipid phosphorus concentration into PL concentration. In the conversion factor, PL = lipid phosphorus \times 24.0, an “average phospholipid” of molecular weight 744 (i.e., an equimolar mixture of oleopalmitoyl lecithin and cephalin) is assumed. The PI sample was used for the measurement of PI (4,5)P₂ and PI(3,4,5)P₃. The phosphoinositide assay is based on the interaction of PI with phosphoinositide-binding proteins⁷⁹. Specifically, we measured lipid phosphorus from the PI fraction and converted the quantified lipid phosphorus into PL. We have expressed the PLC δ -PH domain that specifically binds to PI(4,5)P₂ and Grp1-PH domain that binds specifically with PI(3,4,5)P₃ in the bacteria as a maltose-binding protein (MBP) fused with a rhodopsin-1D4 epitope. The expressed proteins were purified on an amylose affinity column chromatography. An equivalent amount of PL from different samples was coated onto 96-well ELISA plate, and the dried lipids were blocked with 3%BSA in PBS at room temperature for 2 h. The ELISA plate was incubated with MBP-PLC δ -PH-1D4 and MBP-Grp1-1D4 (0.25–0.5 μ g/mL) fusion proteins overnight. The plate was washed, then incubated with the rhodopsin-1D4 antibody for 2 h at RT. After washing the plate, a mouse secondary antibody coupled to the HRP enzyme was added and incubated for 60 min at RT. The plate was washed and the luminescence was measured using an ELISA plate reader.

Relative luminescence units (or counts per second) of each PI was normalized to PL.

Myo-inositol measurement. RPE-choroid complexes were isolated from 2-month-old *Cryba1* cKO and age-matched control mice. Each sample was pooled from four mice from the same group. All mice were sacrificed between 10 am and 12 pm. The myo-inositol concentration in the RPE samples was determined as described in the reference⁸⁰. Specifically, the method is based on the oxidation of myo-inositol by NAD⁺-dependent myo-inositol dehydrogenase, coupled to reoxidation of NADH by iodinitrotetrazolium chloride and diaphorase. RPE-choroid complexes were homogenized in (16%, w/v) perchloric acid and centrifuged. All supernatants were neutralized with 2.0 M K₂CO₃ and centrifuged. The supernatants were then added to hexokinase reagent (200 mM Tris-HCl buffer, 400 mM adenosine triphosphate disodium (pH = 8.6), and 115 U/mL hexokinase). The mixtures were incubated at 37 °C for 90 min, and heated for 3 min to stop the reaction. Endogenous NADH and NADPH were then eliminated by adding 4.5 M HCl. Then, 3.0 M K₂CO₃ was added to the mixtures for neutralization. Then, samples were mixed with 210 mM triethanolamine hydrochloride–32 mM K₂HPO₄–KOH buffer (pH 8.6), 1.2% (v/v) Triton X-100, 10 mM β -NAD, 1.0 U/mL diaphorase, 0.1% (w/v) bovine serum albumin, and 60 μ g/mL INT in a 96-well microplate. The absorbance of the solutions was measured at 492 nm before and after the reaction by adding 2.1 U/mL MIDH dissolved in 20 mM potassium phosphate buffer (pH 7.0) to each well for 30 min at room temperature. The myo-inositol content was calculated based on the increase in absorbance during the reaction.

***Cryba1* overexpression in *Cryba1* cKO RPE flatmounts.** An Ad-CMV-RFP-m*Cryba1* vector was made by inserting mouse *Cryba1* cDNA into a human type 5 (de1/E3) adenoviral backbone, with a RFP under its own CMV promoter. *Cryba1* cKO or control RPE flatmounts were prepared and infected with either Ad-RFP-m*Cryba1* or Ad-CMV-RFP at a concentration of 1×10^7 PFU/mL for 16 h. RPE flatmounts were then processed for subsequent analysis.

Western blotting analysis and antibodies. The native protein samples were mixed with 4x protein LDS sample buffer (NP0007, Invitrogen) containing 10% 2-mercaptoethanol and heated at 100 °C for 10 min. Then, 8–12 μ g of protein from each RPE sample was loaded onto a 4–12% Bis-Tris Nu-PAGE gel (NP0323BOX, Invitrogen) and transferred to a nitrocellulose membrane. Membranes were blocked in either 5% non-fat milk (170-6404, Bio-Rad) or 5% BSA (A3912-50G, Sigma Aldrich), and then incubated with primary antibodies overnight at 4 °C followed by three washes (30 min total) and incubation with the appropriate HRP-conjugated secondary antibody for 1 h at room temperature. Blots were visualized using the ECL western blotting detection reagent (RPN2209, GE Healthcare Life Sciences). The following antibodies were used: phospho-Ezrin (Thr567) (1:1000, PA537763, Thermo Fisher Scientific), Ezrin (1:1000, 3145, Cell Signaling Technology), EBP50 (1:2000, PA1090, Thermo Fisher Scientific), Rac1 (1:1000, 610651, BD Biosciences), phospho-EGFR (Tyr1068) (1:1000, 2234, Cell Signaling Technology), EGFR (1:1000, 2646, Cell Signaling Technology), mCherry (1:1000, 43590, Cell Signaling Technology), phospho-PLC γ 1 (Tyr783) (1:1000, 2821, Cell Signaling Technology), PLC γ 1 (1:1000, 5690, Cell Signaling Technology), phospho-PKC (pan) (β II Ser660) (1:1000, 9371, Cell Signaling Technology), phospho-AKT1 (S473) (1:1000, 9018, Cell Signaling Technology), AKT1 (1:1000, 2938, Cell Signaling Technology), phospho-p44/42 MAPK (Erk1/2) (Thr202/Tyr204) (1:1000, 9106, Cell Signaling Technology), p44/42 MAPK (Erk1/2) (1:1000, 9102, Cell Signaling Technology), and β A3A1-crystallin (1:1000, ab151722, Abcam), MCT3 (1:1000, AMT013, Alomone Labs), Sodium Potassium ATPase. (1:2000, ab7671, Abcam), Vinculin (1:1000, ab73412, Abcam), β -catenin (1:1000, ab16051, Abcam), phospho- β -catenin (1:1000, 2009, Cell Signaling Technology), β -tubulin (1:1000, 2128S, Cell Signaling Technology), Caveolin-1 (1:1000, 3267, Cell Signaling Technology), Actin (1:1000, A2066, Sigma Aldrich), RRP1 (1:1000, MAB3585, R&D Systems), CLTA (1:1000, 10852-1-AP, Proteintech). Quantification of the band intensity was conducted with Quantity One software (Bio-Rad).

Statistics and reproducibility. Statistical analysis was performed using GraphPad Prism v.8 software. All graphs show mean \pm SD. *P* values were obtained by either two-way ANOVA (multiple comparisons) or a two-tailed unpaired Student's *t*-test (comparison between two groups). The numbers of biological repeats (*n*) of each experiment are provided in the figure legends. All figures show the data from independent samples or otherwise are indicated in the figure legend as representative data from independent experiments.

Reporting summary. Further information on research design is available in the Nature Research Reporting Summary linked to this article.

Data availability

All data generated or analyzed during this study are included in the article (and its Supplementary Data files). Source data for figures can be found in Supplementary Data 1. Original blots/gel images are shown in Supplementary Fig. 4. All relevant data relating to

this manuscript are available upon request. The mass spectrometry proteomics data have been deposited to the ProteomeXchange Consortium via the PRIDE partner repository with the dataset identifier PXD026777.

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References

- Lakkaraju, A. et al. The cell biology of the retinal pigment epithelium. *Prog. Retin. Eye Res.* 100846 (2020).
- Storm, T., Burgoyne, T. & Futter, C. E. Membrane trafficking in the retinal pigment epithelium at a glance. *J. Cell Sci.* 133, jcs238279 (2020).
- Lehmann, G. L., Benedicto, I., Philp, N. J. & Rodriguez-Boulán, E. Plasma membrane protein polarity and trafficking in RPE cells: past, present and future. *Exp. Eye Res.* 126, 5–15 (2014).
- Pelaseyed, T. & Bretscher, A. Regulation of actin-based apical structures on epithelial cells. *J. Cell Sci.* 131, jcs221853 (2018).
- Bonilha, V. L. Age and disease-related structural changes in the retinal pigment epithelium. *Clin. Ophthalmol.* 2, 413–424 (2008).
- Viswanatha, R., Bretscher, A. & Garbett, D. Dynamics of ezrin and EBP50 in regulating microvilli on the apical aspect of epithelial cells. *Biochem Soc. Trans.* 42, 189–194 (2014).
- Falkenburger, B. H., Jensen, J. B., Dickson, E. J., Suh, B. C. & Hille, B. Phosphoinositides: lipid regulators of membrane proteins. *J. Physiol.* 588, 3179–3185 (2010).
- Jacoby, M. et al. INPP5E mutations cause primary cilium signaling defects, ciliary instability and ciliopathies in human and mouse. *Nat. Genet.* 41, 1027–1031 (2009).
- George, A. A. et al. Synaptojanin 1 is required for endolysosomal trafficking of synaptic proteins in cone photoreceptor inner segments. *PLoS One* 9, e84394 (2014).
- Van Epps, H. A. et al. The zebrafish nrc mutant reveals a role for the polyphosphoinositide phosphatase synaptojanin 1 in cone photoreceptor ribbon anchoring. *J. Neurosci.* 24, 8641–8650 (2004).
- Hagstrom, S. A., North, M. A., Nishina, P. L., Berson, E. L. & Dryja, T. P. Recessive mutations in the gene encoding the tubby-like protein TULP1 in patients with retinitis pigmentosa. *Nat. Genet.* 18, 174–176 (1998).
- Jacobson, S. G. et al. TULP1 mutations causing early-onset retinal degeneration: preserved but insensitive macular cones. *Invest. Ophthalmol. Vis. Sci.* 55, 5354–5364 (2014).
- He, F., Agosto, M. A., Nichols, R. M. & Wensel, T. G. Multiple phosphatidylinositol(3)phosphate roles in retinal pigment epithelium membrane recycling. *bioRxiv*. <https://doi.org/10.1101/2020.01.09.899815> (2020).
- Wensel, T. G. Phosphoinositides in retinal function and disease. *Cells* 9, 866 (2020).
- Rajala, R. V. S. Signaling roles of phosphoinositides in the retina. *J. Lipid Res.* 62, 100041 (2021).
- Sun, Y., Thapa, N., Hedman, A. C. & Anderson, R. A. Phosphatidylinositol 4,5-bisphosphate: targeted production and signaling. *Bioessays* 35, 513–522 (2013).
- Huberts, D. H. & van der Klei, I. J. Moonlighting proteins: an intriguing mode of multitasking. *Biochim Biophys. Acta* 1803, 520–525 (2010).
- Roskamp, K. W., Paulson, C. N., Brubaker, W. D. & Martin, R. W. Function and aggregation in structural eye lens crystallins. *Acc. Chem. Res.* 53, 863–874 (2020).
- Klemenz, R., Frohli, E., Steiger, R. H., Schafer, R. & Aoyama, A. Alpha B-crystallin is a small heat shock protein. *Proc. Natl Acad. Sci. USA* 88, 3652–3656 (1991).
- Hendriks, W. et al. Duck lens epsilon-crystallin and lactate dehydrogenase B4 are identical: a single-copy gene product with two distinct functions. *Proc. Natl Acad. Sci. USA* 85, 7114–7118 (1988).
- Zigler, J. S. Jr. et al. Mutation in the betaA3/A1-crystallin gene impairs phagosome degradation in the retinal pigmented epithelium of the rat. *J. Cell Sci.* 124, 523–531 (2011).
- Sinha, D. et al. betaA3/A1-crystallin in astroglial cells regulates retinal vascular remodeling during development. *Mol. Cell Neurosci.* 37, 85–95 (2008).
- Shang, P. et al. The amino acid transporter SLC36A4 regulates the amino acid pool in retinal pigmented epithelial cells and mediates the mechanistic target of rapamycin, complex 1 signaling. *Aging Cell* 16, 349–359 (2017).
- Valapala, M. et al. Lysosomal-mediated waste clearance in retinal pigment epithelial cells is regulated by CRYBA1/betaA3/A1-crystallin via V-ATPase-MTORC1 signaling. *Autophagy* 10, 480–496 (2014).
- Sinha, D. et al. Lysosomes: regulators of autophagy in the retinal pigmented epithelium. *Exp. Eye Res.* 144, 46–53 (2016).
- Valapala, M. et al. Increased Lipocalin-2 in the retinal pigment epithelium of Cryba1 cKO mice is associated with a chronic inflammatory response. *Aging Cell* 13, 1091–1094 (2014).
- Ghosh, S. et al. A role for betaA3/A1-crystallin in type 2 EMT of RPE cells occurring in dry age-related macular degeneration. *Invest. Ophthalmol. Vis. Sci.* 59, AMD104–AMD113 (2018).
- Werten, P. J., Stege, G. J. & de Jong, W. W. The short 5' untranslated region of the betaA3/A1-crystallin mRNA is responsible for leaky ribosomal scanning. *Mol. Biol. Rep.* 26, 201–205 (1999).
- Viswanatha, R., Wayt, J., Ohouo, P. Y., Smolka, M. B. & Bretscher, A. Interactome analysis reveals ezrin can adopt multiple conformational states. *J. Biol. Chem.* 288, 35437–35451 (2013).
- Pujuguet, P., Del Maestro, L., Gautreau, A., Louvard, D. & Arpin, M. Ezrin regulates E-cadherin-dependent adherens junction assembly through Rac1 activation. *Mol. Biol. Cell* 14, 2181–2191 (2003).
- Hatano, R. et al. Loss of ezrin expression reduced the susceptibility to the glomerular injury in mice. *Sci. Rep.* 8, 4512 (2018).
- Lobato-Alvarez, J. A. et al. The apical localization of Na(+), K(+)-ATPase in cultured human retinal pigment epithelial cells depends on expression of the beta2 subunit. *Front Physiol.* 7, 450 (2016).
- Philp, N. J., Wang, D., Yoon, H. & Hjelmeland, L. M. Polarized expression of monocarboxylate transporters in human retinal pigment epithelium and ARPE-19 cells. *Invest. Ophthalmol. Vis. Sci.* 44, 1716–1721 (2003).
- Tamiya, S. & Kaplan, H. J. Role of epithelial-mesenchymal transition in proliferative vitreoretinopathy. *Exp. Eye Res.* 142, 26–31 (2016).
- Pei, D., Shu, X., Gassama-Diagne, A. & Thiery, J. P. Mesenchymal-epithelial transition in development and reprogramming. *Nat. Cell Biol.* 21, 44–53 (2019).
- MacDonald, B. T., Tamai, K. & He, X. Wnt/beta-catenin signaling: components, mechanisms, and diseases. *Dev. Cell* 17, 9–26 (2009).
- Han, J. W., Lyu, J., Park, Y. J., Jang, S. Y. & Park, T. K. Wnt/beta-catenin signaling mediates regeneration of retinal pigment epithelium after laser photocoagulation in mouse eye. *Invest. Ophthalmol. Vis. Sci.* 56, 8314–8324 (2015).
- Crepaldi, T., Gautreau, A., Comoglio, P. M., Louvard, D. & Arpin, M. Ezrin is an effector of hepatocyte growth factor-mediated migration and morphogenesis in epithelial cells. *J. Cell Biol.* 138, 423–434 (1997).
- Krieg, J. & Hunter, T. Identification of the two major epidermal growth factor-induced tyrosine phosphorylation sites in the microvillar core protein ezrin. *J. Biol. Chem.* 267, 19258–19265 (1992).
- Lemmon, M. A. & Schlessinger, J. Cell signaling by receptor tyrosine kinases. *Cell* 141, 1117–1134 (2010).
- Lemmon, M. A., Schlessinger, J. & Ferguson, K. M. The EGFR family: not so prototypical receptor tyrosine kinases. *Cold Spring Harb. Perspect. Biol.* 6, a020768 (2014).
- Normanno, N. et al. Epidermal growth factor receptor (EGFR) signaling in cancer. *Gene* 366, 2–16 (2006).
- Goh, L. K. & Sorkin, A. Endocytosis of receptor tyrosine kinases. *Cold Spring Harb. Perspect. Biol.* 5, a017459 (2013).
- Fukuda, S. et al. Reversible interconversion and maintenance of mammary epithelial cell characteristics by the ligand-regulated EGFR system. *Sci. Rep.* 6, 20209 (2016).
- Harris, D. L. & Joyce, N. C. Protein tyrosine phosphatase, PTP1B, expression and activity in rat corneal endothelial cells. *Mol. Vis.* 13, 785–796 (2007).
- Scartozzi, M. et al. Epidermal growth factor receptor (EGFR) downstream signalling pathway in primary colorectal tumours and related metastatic sites: optimising EGFR-targeted treatment options. *Br. J. Cancer* 97, 92–97 (2007).
- Sparrow, J. R. & Duncker, T. Fundus autofluorescence and RPE lipofuscin in age-related macular degeneration. *J. Clin. Med.* 3, 1302–1321 (2014).
- Kaksonen, M. & Roux, A. Mechanisms of clathrin-mediated endocytosis. *Nat. Rev. Mol. Cell Biol.* 19, 313–326 (2018).
- Nile, A. H., Bankaitis, V. A. & Grabon, A. Mammalian diseases of phosphatidylinositol transfer proteins and their homologs. *Clin. Lipido.* 5, 867–897 (2010).
- Ghosh, S. et al. betaA1-crystallin regulates glucose metabolism and mitochondrial function in mouse retinal astrocytes by modulating PTP1B activity. *Commun. Biol.* 4, 248 (2021).
- Wiedemann, C. & Cockcroft, S. The role of phosphatidylinositol transfer proteins (PITPs) in intracellular signalling. *Trends Endocrinol. Metab.* 9, 324–328 (1998).
- Di Paolo, G. & De Camilli, P. Phosphoinositides in cell regulation and membrane dynamics. *Nature* 443, 651–657 (2006).
- Currie, R. A., MacLeod, B. M. & Downes, C. P. The lipid transfer activity of phosphatidylinositol transfer protein is sufficient to account for enhanced phospholipase C activity in turkey erythrocyte ghosts. *Curr. Biol.* 7, 184–190 (1997).

54. Gong, B. et al. The Sec14-like phosphatidylinositol transfer proteins Sec14I3/SEC14L2 act as GTPase proteins to mediate Wnt/Ca(2+) signaling. *Elife* **6**, e26362 (2017).
55. Cunningham, E. et al. The yeast and mammalian isoforms of phosphatidylinositol transfer protein can all restore phospholipase C-mediated inositol lipid signaling in cytosol-depleted RBL-2H3 and HL-60 cells. *Proc. Natl Acad. Sci. USA* **93**, 6589–6593 (1996).
56. Fukami, K., Inanobe, S., Kanemaru, K. & Nakamura, Y. Phospholipase C is a key enzyme regulating intracellular calcium and modulating the phosphoinositide balance. *Prog. Lipid Res.* **49**, 429–437 (2010).
57. Liu, P., Cheng, H., Roberts, T. M. & Zhao, J. J. Targeting the phosphoinositide 3-kinase pathway in cancer. *Nat. Rev. Drug Disco.* **8**, 627–644 (2009).
58. Kadamur, G. & Ross, E. M. Mammalian phospholipase C. *Annu Rev. Physiol.* **75**, 127–154 (2013).
59. Kluba, M., Engelborghs, Y., Hofkens, J. & Mizuno, H. Inhibition of receptor dimerization as a novel negative feedback mechanism of EGFR signaling. *PLoS One* **10**, e0139971 (2015).
60. Zhu, X. & Eichberg, J. A myo-inositol pool utilized for phosphatidylinositol synthesis is depleted in sciatic nerve from rats with streptozotocin-induced diabetes. *Proc. Natl Acad. Sci. USA* **87**, 9818–9822 (1990).
61. Michailidis, I. E. et al. Phosphatidylinositol-4,5-bisphosphate regulates epidermal growth factor receptor activation. *Pflug. Arch.* **461**, 387–397 (2011).
62. Ile, K. E. et al. Zebrafish class 1 phosphatidylinositol transfer proteins: PITPbeta and double cone cell outer segment integrity in retina. *Traffic* **11**, 1151–1167 (2010).
63. Hotta, Y. & Benzer, S. Abnormal electroretinograms in visual mutants of *Drosophila*. *Nature* **222**, 354–356 (1969).
64. Kohn, L. et al. Mutation in the PYK2-binding domain of PITPNM3 causes autosomal dominant cone dystrophy (CORD5) in two Swedish families. *Eur. J. Hum. Genet* **15**, 664–671 (2007).
65. Reinis, A., Golovleva, I., Kohn, L. & Sandgren, O. Ocular phenotype of CORD5, an autosomal dominant retinal dystrophy associated with PITPNM3 p.Q626H mutation. *Acta Ophthalmol.* **91**, 259–266 (2013).
66. Guo, J. & Yu, F. X. Cloning and characterization of human homologue of *Drosophila* retinal degeneration B: a candidate gene for degenerative retinal diseases. *Dev. Genet* **20**, 235–245 (1997).
67. Katan, M. & Cockcroft, S. Phosphatidylinositol(4,5)bisphosphate: diverse functions at the plasma membrane. *Essays Biochem* **64**, 513–531 (2020).
68. Penberthy, K. K., Lysiak, J. J. & Ravichandran, K. S. Rethinking phagocytes: clues from the retina and testes. *Trends Cell Biol.* **28**, 317–327 (2018).
69. Bourne, R. R. A. et al. Prevalence and causes of vision loss in high-income countries and in Eastern and Central Europe in 2015: magnitude, temporal trends and projections. *Br. J. Ophthalmol.* **102**, 575–585 (2018).
70. Wang, J. et al. ATAC-Seq analysis reveals a widespread decrease of chromatin accessibility in age-related macular degeneration. *Nat. Commun.* **9**, 1364 (2018).
71. Volland, S., Esteve-Rudd, J., Hoo, J., Yee, C. & Williams, D. S. A comparison of some organizational characteristics of the mouse central retina and the human macula. *PLoS One* **10**, e0125631 (2015).
72. Vogt, S. D. et al. Retinal pigment epithelial expression of complement regulator CD46 is altered early in the course of geographic atrophy. *Exp. Eye Res.* **93**, 413–423 (2011).
73. Kochetov, A. V. Alternative translation start sites and hidden coding potential of eukaryotic mRNAs. *Bioessays* **30**, 683–691 (2008).
74. Levine, C. G., Mitra, D., Sharma, A., Smith, C. L. & Hegde, R. S. The efficiency of protein compartmentalization into the secretory pathway. *Mol. Biol. Cell* **16**, 279–291 (2005).
75. Cai, J., Huang, Y., Li, F. & Li, Y. Alteration of protein subcellular location and domain formation by alternative translational initiation. *Proteins* **62**, 793–799 (2006).
76. Ma, B. et al. betaA3/A1-Crystallin controls anoikis-mediated cell death in astrocytes by modulating PI3K/AKT/mTOR and ERK survival pathways through the PKD/Btk1-signaling axis. *Cell Death Dis.* **2**, e217 (2011).
77. Wei, H., Xun, Z., Granado, H., Wu, A. & Handa, J. T. An easy, rapid method to isolate RPE cell protein from the mouse eye. *Exp. Eye Res.* **145**, 450–455 (2016).
78. Rouser, G., Siakotos, A. N. & Fleischer, S. Quantitative analysis of phospholipids by thin-layer chromatography and phosphorus analysis of spots. *Lipids* **1**, 85–86 (1966).
79. Rajala, A., McCauley, A., Brush, R. S., Nguyen, K. & Rajala, R. V. S. Phosphoinositide lipids in ocular tissues. *Biology (Basel)* **9**, 125 (2020).
80. Ashizawa, N., Yoshida, M. & Aotsuka, T. An enzymatic assay for myo-inositol in tissue samples. *J. Biochem. Biophys. Methods* **44**, 89–94 (2000).

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Author contributions

P. S. and D. S. designed the study and D. S. supervised the work. P. S., N. S., A. M., K. T., C. S. F., R. G., M. Y., S. H., S. G., H. L., A. S., J. W., and I. A. B. conducted the experiments and/or analyzed the data. R. V. S. R. analyzed the PIP data. J. Q. and Y. S. developed the molecular models and bioinformatics studies. S. W. developed the TIRF based live-cell technique to image clathrin-mediated endocytosis from the apical side of RPE cells. P. S., N. S., S. H., J. S. Z., J. T. H., R. V. S. R., and D. S. wrote the paper. G. A. L., A. J., J.-A. S., and S. W. reviewed the data and revised the manuscript.

Competing interests

The authors declare no competing interests.

Additional information


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