Insights into the biology and therapeutic implications of TNF and regulatory T cells

Benoit L Salomon

► To cite this version:

Benoit L Salomon. Insights into the biology and therapeutic implications of TNF and regulatory T cells. Nature Reviews Rheumatology, 2021, 10.1038/s41584-021-00639-6 . hal-03287990

HAL Id: hal-03287990

<https://hal.sorbonne-universite.fr/hal-03287990>

Submitted on 16 Jul 2021

HAL is a multi-disciplinary open access archive for the deposit and dissemination of scientific research documents, whether they are published or not. The documents may come from teaching and research institutions in France or abroad, or from public or private research centers.

L'archive ouverte pluridisciplinaire **HAL**, est destinée au dépôt et à la diffusion de documents scientifiques de niveau recherche, publiés ou non, émanant des établissements d'enseignement et de recherche français ou étrangers, des laboratoires publics ou privés.

Insights into the biology and therapeutic implications of TNF and regulatory T cells

Benoit L. Salomon[†]

Sorbonne Université, INSERM, CNRS, Centre d'Immunologie et des Maladies Infectieuses (CIMI-Paris), F-75013, Paris, France.

[†]email: benoit.salomon@inserm.fr.

Abstract

Treatments that block tumour necrosis factor (TNF) have major beneficial effects in several autoimmune and rheumatic diseases, including rheumatoid arthritis. However, some patients do not respond to TNF inhibitor treatment and some rare occurrences of paradoxical disease exacerbation have been reported. These limitations on the clinical efficacy of TNF inhibitors can be explained by the differences between TNF receptor 1 (TNFR1) and TNFR2 signalling and by the diverse effects of TNF on multiple immune cells, including FOXP3⁺ regulatory T (T_{reg}) cells. This basic knowledge sheds light on the consequences of TNF inhibitor therapies on T_{reg} cells in treated patients and on the limitations of such treatment in the control of diseases with an autoimmune component. Accordingly, the next generation of drugs targeting TNF is likely to be based on agents that selectively block the binding of TNF to TNFR1 and on TNFR2 agonists. These approaches could improve the treatment of rheumatic diseases in the future.

[H1] Introduction

Tumour necrosis factor (TNF) is an inflammatory cytokine that is detected in the blood within minutes after an injury and has a major protective role in infectious diseases. In the late 1980s, TNF was detected in the joints of patients with rheumatoid arthritis (RA)^{1,2}. A few years later, overexpression of TNF in transgenic mice was shown to induce autoimmune arthritis³. Agents that block this cytokine, termed TNF inhibitors, include monoclonal antibodies (mAbs) and soluble TNF receptors. Anti-TNF therapy was first tested in patients with sepsis without clear success and then repurposed for the treatment of RA in the early 1990s^{1,2}. TNF inhibitors are now widely used and have greatly improved the medical care of patients with RA, juvenile idiopathic arthritis, psoriasis, psoriatic arthritis and ankylosing spondylitis. Five original TNF

inhibitors and numerous biosimilars have been approved, mostly for the treatment of arthritis, psoriasis or ankylosing spondylitis (**Table 1**). However, not all patients respond to TNF inhibitor treatment. One-third of patients with RA have to stop taking these drugs within the first year because of insufficient efficacy or adverse events⁴. About 20% of patients with psoriasis do not respond to treatment with a TNF inhibitor and around one-third of initial responders lose response over time⁵. Similar efficacy profiles are observed for patients with inflammatory bowel disease (IBD)⁶. Although this Review mainly focuses on the effects of TNF inhibitors in rheumatic diseases, particularly RA, I also discuss their effects and use in the treatment of other autoimmune and inflammatory diseases to illustrate the role and mechanisms of these agents in general.

Treatment with TNF inhibitors is also associated with adverse effects, such as infections, which are explained by the intrinsic anti-inflammatory effects of these agents. More intriguing (and counterintuitive) is the paradoxical exacerbation of pre-existing autoimmune disease or the development of new-onset autoimmune disease following TNF inhibitor therapy. Rarely, treated patients can develop lupus-like syndrome, vasculitis, antiphospholipid syndrome or sarcoidosis. For example, the reported prevalence of systemic lupus erythematosus (SLE) among recipients of TNF inhibitor therapy is 0.1–0.2%^{7–9}. A few patients develop organ-specific autoimmune conditions, such as interstitial lung disease, optical neuritis, demyelinating neuropathy, multiple sclerosis (MS), psoriasis or autoimmune hepatitis, with the highest prevalence (2.00–5.00%) reported for psoriasis and the lowest (0.05–0.20%) for demyelinating disease. Several reviews have discussed in depth the spectrum of autoimmune diseases occurring in TNF inhibitor-treated patients^{7–10}, among which MS is of particular interest. In the late 1990s, before the increased risk of demyelinating neuropathy associated with TNF inhibitor treatment was known, two clinical trials investigated the efficacy of TNF inhibitors in MS. However, these drugs induced unexpected disease exacerbations that led to the worldwide contraindication of these drugs in these patients^{11,12}. These observations sparked intense interest in elucidating why not all patients respond to TNF inhibitor therapy, developing biomarkers to predict response, and understanding why some treated patients develop paradoxical autoimmunity.

This Review focuses on the effects of TNF on inflammation and immunity. I describe the pro-inflammatory and regulatory roles of TNF, both of which are now well-established, and address the effects of this cytokine on diverse aspects of regulatory T (T_{reg}) cell biology, including their expansion, differentiation and suppressive function. Finally, I describe the effect of TNF inhibitors on T_{reg} cells and explore potential candidates for the next generation of drugs that target TNF or its receptors. Although TNF also has important roles in organogenesis and the

development of lymphoid organs, protection of tissues in the nervous system, heart and joints¹³⁻¹⁵ and inhibition of tumorigenesis¹⁶, these topics are outside the scope of the present Review and will not be considered.

[H1] The two Janus faces of TNF

TNF has complex regulatory and pro-inflammatory effects in diseases with an autoimmune component¹³⁻¹⁵, such as RA¹⁷. This cytokine is produced under various inflammatory conditions by multiple cell types and exists in two forms: a soluble form that acts as a ligand, and a membrane-bound form that can act as either a ligand or a receptor¹⁸⁻²⁰. Furthermore, TNF can induce multiple downstream signalling pathways⁹ as a result of binding to two different receptors, TNF receptor (TNFR) 1 and TNFR2, which are structurally related but have divergent biological properties. TNFR1 is broadly expressed whereas TNFR2 is expressed mostly by T cells, certain myeloid and endothelial cells and some cells of the central nervous system^{21,22}.

The next sections describe the distinct functions of TNFR1 and TNFR2 and discuss the proinflammatory and anti-inflammatory effects of TNF on innate immune cells and lymphocytes, and present information on the cellular source of TNF.

[H2] TNFR1 and TNFR2

TNF is one of the most potent pro-inflammatory cytokines²³, which explains the success of TNF inhibitor therapy in diseases with an inflammatory component. However, the paradoxical development or exacerbation of autoimmune disease in some patients treated with these drugs reveals the anti-inflammatory aspect of this cytokine, which is partly explained by effects downstream of TNFR2. Polymorphisms in *TNFRSF1B*, which encodes TNFR2, are frequently observed in patients with rheumatic diseases (RA, SLE, ankylosing spondylitis and systemic sclerosis) or IBD²⁴. These mutations seem to alter the binding kinetics between TNF and TNFR2 and lead to inhibition of downstream NF-κB signalling, which suggests that TNFR2 signalling has a protective role in these diseases²⁵. A single nucleotide polymorphism in *TNFRSF1A*, which encodes TNFR1, is specifically associated with an increased risk of MS. This allele results in the expression of a novel soluble form of TNFR1 that binds to and blocks TNF, and therefore mimics the MS-exacerbating effect of TNF inhibitor therapy²⁶. Other mutations in *TNFRSF1A* that cause TNFR1 misfolding and endoplasmic reticulum stress are found in patients with periodic fevers²⁷.

The differential functions of TNFR1 and TNFR2 in rheumatic and autoimmune diseases have been defined in mouse models. Generally, TNFR1-knockout mice have reduced disease

severity whereas TNFR2-knockout mice develop exacerbated disease (**Table 2**). In addition, treatment with either TNFR1 antagonists or TNFR2 agonists suppresses disease symptoms in mouse models of arthritis and in mice with experimental autoimmune encephalomyelitis (EAE), a model of MS, further supporting a pathogenic role of TNFR1 and a protective role of TNFR2 (**Table 2**). A pathogenic role of TNFR1 and a protective role of TNFR2 have also been observed in mouse models of IBD, at least during the chronic phase of the disease^{15,28}. Thus, TNFR1 and TNFR2 seem to be pathogenic and protective, respectively, in some autoimmune and chronic inflammatory diseases.

[H2] Effects of TNF on innate immunity

[H3] Pro-inflammatory effects.

The pro-inflammatory effects of TNF on innate immunity involve several distinct mechanisms (**Figure 1**). TNF is one of the main drivers of acute inflammation because it activates endothelial cells, induces chemokine release and promotes intense and early (within hours) recruitment of neutrophils and monocytes, via both TNFR1 and TNFR2^{29,30}. Acute inflammation is also attributed to the TNF-mediated activation of canonical NF- κ B signalling, which leads to the early induction of inflammatory cytokines, including TNF itself, IL-6, IL-8 and IL-1 β ¹⁴. TNF also sustains inflammation through the activation of receptor-interacting protein kinase (RIPK) 1 and RIPK3, which promote necroptosis and the release of inflammatory compounds termed damage-associated molecular patterns (DAMPs)³¹. In addition, via TNFR1 signalling, TNF promotes innate immunity by favouring the maturation of dendritic cells^{32,33}.

[H3] Regulatory effects.

The immunoregulatory functions of TNF are likely to involve multiple mechanisms (**Figure 1**). TNF might promote the extra-adrenal production of immunoregulatory glucocorticoids³⁴ and inhibit haematopoiesis³⁵. TNF also stimulates innate immunosuppressive cells (via TNFR2) and activates mesenchymal stem cells, which produce increased levels of immunosuppressive prostaglandin E2 (PGE₂), as has been shown in synovial fluid from patients with RA^{36,37}. TNF also promotes immunosuppression by favouring either the differentiation or the suppressive function of myeloid-derived suppressor cells via increasing their production of reactive oxygen species, arginase 1 and inducible nitric oxide synthase³⁸⁻⁴².

[H3] Effects on dendritic cells, monocytes and macrophages.

Although TNF seems to favour the production of T-bet and IL-12 by dendritic cells⁴³, other

studies suggest that the presence of TNF inhibits the production of p40 (the common chain of IL-12 and IL-23) by dendritic cells, macrophages and monocytes⁴⁴⁻⁴⁶. These divergent findings could be explained by differential actions of TNF depending on the maturation stage and type of both antigen presenting cells and dendritic cell subsets.

In addition, TNF can either promote or inhibit macrophage activation, effects that are both probably mediated by TNFR1 (reviewed elsewhere¹⁴). The early response of macrophages to incubation with TNF, observed after a few hours, is both NF- κ B-dependent and MAPK-dependent and involves the expression of genes encoding various inflammatory molecules and cytokines. This initial response is followed (after 24 hours) by a state of desensitization, also called cross-tolerance or endotoxin tolerance. Desensitized (also termed tolerized) macrophages are unable to produce inflammatory factors when stimulated by potent activators such as Toll-like receptor ligands. The mechanism of desensitization involves NF- κ B inhibition following activation of glycogen synthase kinase 3 (GSK3) and TNF-induced protein 3 (TNFAIP3)⁴⁷. Tolerized macrophages have a transiently reduced capacity to produce IL-12 and IL-23, which are pro-inflammatory^{46,48}. The physiological role of cross-tolerance is probably the prevention of life-threatening inflammation in the context of overwhelming macrophage activation by pathogens and Toll-like receptor ligands. Ultimately, after prolonged incubation with TNF, specifically in the presence of type 1 interferons, macrophages overcome this desensitized state and recover their inflammatory function by modifying their metabolism and epigenetic status^{49,50}.

[H2] Effects of TNF on lymphocytes

[H3] Pro-inflammatory effects.

TNF can either promote or suppress immunity through its differential effects on lymphocytes (**Figure 1**). The pro-inflammatory effects of TNF result from co-stimulation of T cells, mainly via TNFR2. TNF activates NF- κ B and AKT signalling pathways that lead to increased T cell proliferation and survival, which are associated with increased levels of BCL2, BCLXL, IL-2 and survivin⁵¹⁻⁵⁶. However, the co-stimulatory effect of TNF binding to TNFR2 on conventional T cells seems to be of marginal importance compared with its strong effect on T_{reg} cells⁵⁷, which is extensively discussed below.

[H3] Regulatory effects.

Although one report suggests that TNF promotes the expression of IL-10 by B regulatory (B_{reg}) cells⁵⁸, much more is known about the inhibitory effects of TNF on T cells. Prolonged exposure

to TNF attenuates T cell receptor signalling by impairing store-operated calcium influx^{59,60} and also favours T cell exhaustion; in one report, TNF blockade during chronic infection with lymphocytic choriomeningitis virus abrogated the inhibitory gene expression signature⁶¹. TNF is able to induce activation-induced cell death via TNFR1 engagement⁶². Interestingly, TNFR2 signalling also seems to increase T-cell apoptosis by interfering with signalling pathways downstream of TNFR1⁶³. However, TNFR2-dependent cell death might specifically occur in autoreactive T cells, which have altered TNFR2 signalling^{25,62,64-68}. Cross-talk between TNFR1 and TNFR2 signalling is discussed in more detail in subsequent sections.

TNF also inhibits the differentiation of T_H17 cells by increasing IL-2 production⁶⁹, and decreases IL-17 production by conventional T cells and effector T_{reg} cells via activation of TNFAIP3^{70,71}. This mechanism might explain the increase in numbers of T_H17 cells described in TNFR1-knockout mice or after treatment with TNF inhibitors in mouse models of RA and psoriasis^{44-46,72,73}. A similar increase in T_H17 cells has been reported specifically in non-responding patients with RA treated with TNF inhibitors^{44,74}. Interestingly, these non-responding patients showed a T_H1-mediated and T_H17-mediated immune response against the TNF inhibitor, which might have precipitated their lack of clinical response⁷⁵. Finally, in the late 2000s, regulatory properties of TNF were proposed to result from its effects on T_{reg} cells. These mechanisms are extensively discussed below.

[H2] Cellular sources of TNF

Multiple cell types are able to produce TNF, but the immune cells that produce this cytokine in the highest amounts are myeloid cells and activated T cells⁷⁶. The role of TNF produced by these two cell types in rheumatic and autoimmune diseases has been investigated using genetically modified mice with conditional knock-out of TNF only in myeloid cells or only in T cells. In mice with collagen-induced arthritis, conditional knockout of TNF in myeloid cells leads to reduced disease severity, showing that the TNF produced by these cells contributes to the pathology. By contrast, mice with TNF-deficient T cells have exacerbated arthritis, suggesting a protective role of the TNF produced by T cells⁷⁷. Similarly, mice with EAE and TNF-deficient myeloid cells have attenuated disease, which is either delayed in onset or reduced in severity depending on the model^{76,78}. Finally, the role of TNF produced by B cells has been analyzed in mice with TNF-deficient B cells. These mice have reduced arthritis and reduced levels of autoantibodies⁷⁷.

[H2] TNF structure and signalling

[H3] Soluble and transmembrane TNF.

Crystallographic studies show that trimers of TNF interact with trimers of either TNFR1 or TNFR2^{79,80}. This trimeric association of the cytokine with its receptor is characteristic of the TNF superfamily and is critical for downstream signalling^{63,81}.

TNF is initially produced as a transmembrane molecule that can be processed by disintegrin and metalloproteinase domain-containing protein 17 (also known as TNF converting enzyme (TACE)) encoded by the *ADAM17* gene⁸². Thus, activated myeloid and T cells produce transmembrane TNF and secreted soluble TNF, which are both biologically active^{77,83,84}. The role of soluble TNF in the pathophysiology of rheumatic and autoimmune diseases has been assessed in mice engineered to express a TNF protein that cannot be cleaved by TACE. Thus, these mice produce normal levels of transmembrane TNF but no soluble TNF⁸³. Importantly, such mice do not develop EAE or arthritis, showing that soluble TNF but not transmembrane TNF contributes to these diseases^{77,83,84}. By contrast, mice with global deletion of TNF (full knock-out) still develop EAE, which suggests that transmembrane TNF has protective effects in the disease⁸³.

Despite a similarly high binding affinity for its two receptors, trimeric soluble TNF triggers TNFR1 signalling much more efficiently than it does TNFR2 signalling⁸⁵. Although this observation requires further confirmation, TNFR1 signalling is usually considered to be triggered by both soluble and transmembrane TNF whereas TNFR2 signalling is preferentially triggered by transmembrane TNF⁸⁶. These observations suggest that soluble TNF (notably that produced by myeloid cells at the onset of a rheumatic or autoimmune disease) binds to TNFR1 to promote inflammation and precipitate the disease, whereas transmembrane TNF (probably that expressed by both myeloid and T cells) has regulatory effects mostly derived from triggering TNFR2. These observations might have implications for the use of TNF inhibitors. For example, etanercept (a TNFR2–Fc fusion protein) can efficiently block soluble TNF (as well as $\alpha 3$ and $\alpha 2\beta 1$ lymphotoxins) but not transmembrane TNF, whereas anti-TNF monoclonal antibodies block both soluble and transmembrane TNF⁸². This concept also has implications for the design of next-generation TNF inhibitors, as discussed below.

[H3] TNFR1 and TNFR2 signalling pathways.

TNFR1 and TNFR2 signalling pathways are complex and have been extensively reviewed elsewhere^{14,15,63,86}. Accordingly, only the pathways most relevant to this review are outlined here. Most of the available knowledge has been obtained in cell lines and non-immune cells and deserves further investigation to confirm its relevance in immune cells.

Upon binding of trimeric TNF to TNFR1, the cytoplasmic tail of the receptor recruits the adaptor protein TNFR1-associated death domain (TRADD) via its death domain. TRADD can then interact with other adaptor proteins, such as TNF receptor associated factor 2 (TRAF2), and kinases, such as receptor-interacting serine/threonine-protein kinase 1 (RIPK1) or cellular inhibitor of apoptosis (cIAP) 1 and cIAP2. The resulting molecular complex, named complex 1, is able to phosphorylate and ubiquitinate several other molecules, ultimately leading to potent activation of canonical NF- κ B and MAPK pathways. Members of these pathways, such as c-Jun N-terminal kinase (JNK) and p38, in turn, activate AP1 complex^{14,15,63,86}. This complex-1-dependent signalling pathway favours cell proliferation and survival. Alternatively, TNFR1 and TRADD interact with the Fas associated death domain (FADD) adaptors RIPK1 and RIPK3, forming the complex 2 interactome, which is able to induce cell death: either apoptosis (via caspase 8 activation) or necroptosis (via mixed lineage kinase domain-like (MLKL) protein activation)^{14,15,63,86}.

Complex 1 and complex 2 are downstream effectors of TNFR1 signalling. Complex 1 is probably involved in most of the effects of TNF on dendritic cells and macrophages, including activation of inflammatory target genes and production of inflammatory cytokines. Complex 2 is involved in TNF-dependent, activation-induced cell death and the formation of inflammation-dependent DAMPs.

The signal transduction pathway downstream of TNFR2 lacks a death domain and involves different adaptors. Binding of transmembrane TNF to TNFR2 recruits TRAF1 or TRAF2 adaptors to this receptor, leading to activation of cIAP1 or cIAP2 kinases and activation of canonical and non-canonical NF- κ B, JNK and AKT pathways that promote cell proliferation and survival^{14,15,63,86-88}. These pathways are likely to be involved in the TNF-dependent activation of mesenchymal stem cells and myeloid-derived stem cells as well as T cell co-stimulation. TRAF2 recruitment to TNFR2 also decreases the amount of cytoplasmic TRAF2, which interferes with TNFR1 signalling by favouring the formation of (cell death-promoting) complex 2 to the detriment of (survival-promoting) complex 1⁶³. This cross-talk between TNFR1 and TNFR2 signalling pathways seems to be responsible for TNFR2-dependent T cell death⁸⁹.

[H3] TNF reverse signalling.

Reverse (extracellular to intracellular) signalling induced by transmembrane TNF has been described but remains poorly documented. This phenomenon is only outlined here as it has been reviewed elsewhere¹⁸⁻²⁰. In this context, TNFR1 or TNFR2 can act as ligands for transmembrane TNF, which can function as a cell receptor transducing a signal in several different situations.

For example, TNFR2-expressing T cells promote the increased expression of TNF in monocytes and/or macrophages via transmembrane TNF, a phenomenon that has been observed in the joints of patients with RA. Also, TNFR1-expressing endothelial cells induce cross-tolerance in monocytes and/or macrophages via transmembrane TNF. Finally, TNF inhibitors are also able to bind to transmembrane TNF and thereby induce the apoptosis of transmembrane TNF-expressing cells; this phenomenon has been observed for instance in T cells and synovial macrophages from patients with RA^{90,91}. The mechanism of TNF reverse signalling involves increased intracellular levels of calcium and TGF- β and activation of the MAPK–ERK pathway. However, the in vivo relevance of reverse signalling is difficult to assess because this phenomenon has been poorly described. I consider that reverse signalling might contribute to the spectrum of effects of TNF and might have an important role in inducing cross-tolerance of macrophages and in the death of transmembrane TNF-expressing cells induced by administration of TNF inhibitors.

[H2] Integrative view of TNF functions

Here, I present a simplified and integrated view of what I believe is the major role of TNF in immunity (**Figure 1**). TNF is one of the most potent inflammatory cytokines owing to its capacity to activate endothelial cells, neutrophils, macrophages and dendritic cells, leading to leukocyte recruitment and massive release of inflammatory cytokines at sites of inflammation. Most of these pro-inflammatory phenomena are mediated by TNFR1 signalling. Besides its pro-inflammatory functions, TNF also has anti-inflammatory (regulatory) functions, although their role and mechanisms in immunity are yet to be clarified. The regulatory functions involve TNF-dependent activation of suppressive cells, such as mesenchymal stem cells, myeloid-derived stem cells and of course T_{reg} cells (which are extensively discussed below). TNF might also promote death or exhaustion of T cells and inhibit pathogenic T_H17 cells. Most of these regulatory phenomena are mediated by TNFR2 signalling.

The end result of TNF blockade depends on the type of autoimmune disease present and the timing of treatment. Blocking the interaction between TNF and TNFR1 led to increased numbers of pathogenic T_H1 and T_H17 cells in mouse models of arthritis and psoriasis^{44-46,72,73}. This increase was associated with exacerbation of psoriasis (as might logically be expected) but surprisingly with attenuation of arthritis because this treatment also blocked the migration of pathogenic T cells to the joints⁴⁶. Similarly, patients with RA treated with TNF inhibitors have increased levels of circulating T_H1 and T_H17 cells^{44,74,75}, which could explain some of the paradoxical inflammation observed in a subset of these patients. The effects of TNF blockade could also depend on the timing of treatment in relation to the course of disease. To investigate

the role of TNF signalling via TNFR2 in a model of collagen-induced arthritis, TNFR1-knockout mice were treated with TNF on either days 2–20 or days 22–40 after disease induction⁹². Interestingly, early TNF treatment led to disease exacerbation whereas late TNF treatment led to attenuation of arthritis. An opposite effect of TNF that is similarly dependent on the stage of disease progression has been documented in non-obese diabetic mice; TNF seems to exacerbate diabetes in young mice by activating dendritic cells and to attenuate it in adult mice by inhibiting conventional T cells and promoting T_{reg} cell activation^{59,93-95}.

[H1] Effects of TNF on T_{reg} cells

T_{reg} cells are master regulators of autoimmune diseases. Mice and humans that are genetically deficient in T_{reg} cells die soon after birth from a massive and systemic autoimmune syndrome, which reveals the critical role of these cells in the suppression of autoimmunity⁹⁶. Functional or quantitative defects of T_{reg} cells have been reported in many human autoimmune diseases⁹⁷. Other indirect evidence supports the concept that T_{reg} cells contribute to human autoimmune diseases. For instance, some biomarkers of disease activity, such as C reactive protein levels, are inversely correlated with T_{reg} cell proportion in patients with RA⁹⁸ or IBD^{99,100}. Moreover, T_{reg} cell transfer seems to have beneficial effects in patients with various autoimmune diseases¹⁰¹.

Transcriptomic analyses that compared T_{reg} with conventional T cells in lymphoid tissues showed that several members of the TNFR superfamily, including TNFR2, TNFR superfamily member 4 (OX40), TNFR superfamily member 9 (41BB) and TNFR superfamily member 18 (GITR) are included in the T_{reg} cell signature¹⁰². These molecules are further upregulated upon stimulation of either the T cell receptor (TCR) or T cell-specific surface glycoprotein CD28 and are therefore preferentially expressed by effector T_{reg} cells rather than resting T_{reg} cells^{103,104}. At steady state, 30% of T_{reg} cells express TNFR2 and most of this subset are effector T_{reg} cells that have a stronger suppressive function in vitro than do TNFR2[−] resting T_{reg} cells^{105,106}. Thus, TNFR2 belongs to the T_{reg} cell signature and is a marker of highly suppressive T_{reg} cells.

[H2] Effects on T_{reg} cell expansion

Expansion is defined as an increase in cell numbers, and results from a combination of increased proliferation, prolonged survival and phenotypic stability. TNFR2 signalling seems to expand T_{reg} cells by increasing all three of these factors.

Initially, TNF and/or TNFR2 co-stimulation were shown to increase T_{reg} cell proliferation in mice^{22,107}. Our group and others showed that effector T cells, in particular T_H17 cells, are a major source of the TNF that induces this increase in the T_{reg} cell population in vivo¹⁰⁸⁻¹¹⁰.

Similar findings were obtained for human T_{reg} cells^{84,111,112}. TNF can also substantially prolong T_{reg} cell survival¹⁰³. Indirect evidence indicates that TNFR2 signalling also maintains *FOXP3* expression, which increases T_{reg} cell phenotypic stability and therefore their long-term expansion¹¹²⁻¹¹⁵.

In many of these in vitro studies, soluble TNF was capable of boosting T_{reg} cell expansion. Although transmembrane TNF has a stronger effect than soluble TNF on induction of TNFR2 signalling⁸⁵, strong evidence indicates that soluble TNF can indeed stimulate the expansion of T_{reg} cells by binding to TNFR2. Furthermore, TNFR1 expression has not been detected on T_{reg} cells (unlike TNFR2 expression)²². The expansion-promoting effect of soluble TNF on T_{reg} cells was lost in TNFR2-deficient T_{reg} cells and when TNFR2, but not TNFR1, was blocked¹¹³. Finally, treatment with TNF or TNFR2 agonists induced similar co-stimulation of T_{reg} cells¹¹¹. The capacity of soluble TNF to efficiently induce TNFR2 signalling could be explained by the use of high concentrations of this cytokine or the presence of TNF aggregates with crosslinking properties in the preparations. TNFR2 agonists, which are either multimers of mutated TNF or mAbs that bind only to TNFR2 (discussed in more detail below), strongly co-stimulate T_{reg} cells in both mice and humans^{57,103,111,116-118}. In a study of pre-activated T cells, TNFR2 co-stimulation strongly increased the proliferation of T_{reg} but had no effect on conventional T cells⁵⁷. The capacity of TNFR2 co-stimulation to promote T_{reg} cell expansion was confirmed in vivo in animals treated with TNFR2 agonists^{86,117,119-121}.

Although very little is known about TNFR2 signal transduction in T_{reg} cells, transcriptomic analyses showed that binding of TNF to TNFR2 on purified mouse or human T_{reg} cells induced a gene expression signature indicative of NF-κB pathway activation^{103,122}. More precisely, TNFR2 signalling induced nuclear translocation and binding of RelA to its DNA target sequence, which suggests that the canonical NF-κB pathway is activated by TNFR2 signalling in T_{reg} cells. Importantly, the increased proliferation and prolonged survival of T_{reg} cells induced by TNFR2 triggering was severely attenuated in RelA-deficient T_{reg} cells^{103,104}. Some evidence also suggests that the non-canonical NF-κB pathway is also activated by TNFR2 signalling in T_{reg} cells but this observation has to be treated with caution because these assays were conducted on a cell population with low T_{reg} cell purity¹²³. Other data suggest that TNFR2 signalling induces activation of the MAPK pathway, notably via p38^{124,125}. TNFR2-mediated co-stimulation of T_{reg} cells also induced a glycolytic switch associated with activation of mammalian target of rapamycin complex 1 (mTORC1) signalling via phosphoinositide-3 kinase (PI3K), although the signalling pathway connecting TNFR2 to PI3K was not identified⁵⁷.

Overall, strong evidence indicates that the boost in T_{reg} cell numbers induced by TNFR2 signalling involves activation of the canonical NF-κB pathway. The role of the other signalling pathways mentioned here requires further documentation.

[H2] Effects on T_{reg} suppressive function

The effects of TNF on the suppressive function of mouse and human T_{reg} cells have been assessed in vitro (**Table 3**). The first of these studies showed no effect of low-dose (≤5 ng/ml) TNF in human cells⁹⁸. Five subsequent reports showed that treatment with TNF, usually at a high dose (50 ng/ml), reduced the suppression of conventional T cell activation by human T_{reg} cells^{122,126-129}. By contrast, in vitro studies performed in mouse cells showed that the presence of high amounts of TNF either had no effect or even increased T_{reg} cell-mediated suppression of conventional T cell activation^{22,107}. Moreover, other evidence also suggests that TNF does not inhibit T_{reg} cell-mediated suppression of conventional T cells, and might even increase it. For instance, administration of a TNFR2 agonist to mice with graft versus host disease (GvHD) or collagen-induced arthritis promoted T_{reg} cell expansion and had a therapeutic effect^{117,119,121}. Also, treatment of cultured T_{reg} cells with TNF increased their capacity to suppress colitis or GvHD after transfer^{103,130}, whereas TNFR2-deficient T_{reg} cells had a reduced capacity to suppress colitis or GvHD^{114,131}. However, these observations provide only indirect evidence that TNF either had no effect on or increased T_{reg} cell suppressive function in mice. Indeed, this cytokine might influence other parameters of T_{reg} cell biology (such as proliferation, survival, functional stability or migration). Interestingly, EAE was exacerbated in genetically modified mice in which TNFR2 was ablated only in T_{reg} cells. Ablation of TNFR2 in T_{reg} cells seems to decrease their suppressive function specifically in the inflamed central nervous system¹³². In this context, the expression of TNFR2 by T_{reg} cells might be essential for their suppressive function and their capacity to control EAE.

Our group also performed an analysis of the suppressive capacity of T_{reg} cells from numerous different human donors under three different T cell activation conditions. We consistently found that TNF (added either before or during the suppression assay) either had no effect on or even slightly increased the suppressive activity of human T_{reg} cells¹³³. The preservation of T_{reg} cell suppressive activity after TNFR2 co-stimulation (achieved using a TNFR2 agonist) in humans has also been confirmed⁵⁷.

Several factors might account for the contrasting findings in mouse and human cells. First, as none of the available markers can exclusively characterize the population of human T_{reg} cells, the purified T_{reg} cell populations used in some of these studies might still have some level

of contamination by activated conventional T cells, especially when only CD4 and CD25 expression was used to sort the cells¹³⁴. Second, given the high inter-individual variability in T_{reg} cell phenotypes, responses to TNF and suppressive activity, it is important to collect data from a sufficiently large sample of individuals. Finally, a T_{reg} cell functional defect identified in a suppression assay could be due either to intrinsic T_{reg} cell dysfunction or to the presence of contaminating conventional T cells that are resistant to T_{reg} cell suppression. This last point is critical with regard to the effects of TNF. Indeed, in addition to its proliferation-promoting effect on T_{reg} cells, TNF not only increases the proliferation of conventional T cells^{51,53,133} but also increases their resistance to T_{reg} cell-mediated suppression¹³⁵. In several studies performed in human cells, TNF was present during the suppression assays and might act on any contaminating conventional T cells, which would impair the evaluation of T_{reg} cell suppressive function (**Table 3**). Accordingly, pre-incubation of the T_{reg} cells with TNF is appropriate before testing their capacity to suppress conventional T cells.

Another critical point is the choice of parameter used to assess the activation of conventional T cells. As TNF strongly increases T_{reg} cell proliferation (and possibly also cytokine production), measuring the activation of only the conventional T cells within the population is critical. This measurement can be done by analyzing fluorescent marker dilution or assessing intracellular cytokine production using flow cytometry techniques such as fluorescence-activated cell sorting (FACS). Researchers should not use thymidine incorporation or enzyme-linked immunosorbent assays (ELISA) to measure the proliferation or cytokine production of the whole cell population, which includes both conventional T cells and T_{reg} cells. For this reason, to accurately determine whether TNF alters the suppressive function of T_{reg} cells, we recommend that TNF is added only during the pre-incubation phase (that is, before the suppressive assay), and that activation of only the conventional T cells is measured by FACS. The absence of these two precautionary measures in some of the reports claiming that TNF inhibits T_{reg} cell suppressive activity in humans undermines their conclusions (**Table 3**).

To conclude, weak evidence indicates that TNF is able to either inhibit or increase the suppressive activity of T_{reg} cells. After careful analyses of the data from in vitro assays, I would say that TNF has no or only a minor effect on T_{reg} cell suppressive function in this context. However, this cytokine seems to have an essential role in stimulation of T_{reg} cell function in some conditions associated with inflammation.

The data derived from in vitro studies of mechanisms underlying the suppressive activity of T_{reg} cells reflect only the tip of the iceberg, as only two or three suppressive mechanisms have been analyzed in these studies to date. However, it is now well established that T_{reg} cells in vivo

are able to use a wide range of suppressive mechanisms depending on their tissue localization and the type of inflammation present^{136,137}. The suppressive activity of T_{reg} cells also involves many different effector molecules. Some have been thoroughly studied and shown to be essential for aspects of T_{reg} cell suppression, such as cytotoxic T-lymphocyte protein 4 (CTLA4) and IL-10¹³⁸. FOXP3 expression is also critical because its loss leads to loss of T_{reg} cell function¹³⁸, but no single marker has been shown to easily quantify the level of T_{reg} cell suppression.

Several mechanisms have been suggested to explain how TNF might increase the suppressive function of T_{reg} cells in mice. TNF promotes full differentiation of effector T_{reg} cells by stimulating NF-κB, which might increase some of these cells' suppressive functions^{103,104}. TNF also synergizes with IL-2 to increase the expression of CD25 (the IL-2 receptor α-chain) and FOXP3^{22,133}. Moreover, TNF increases the IL-2-induced phosphorylation of STAT5²² and limits the loss of FOXP3 expression in cultured cells by preventing re-methylation of the *Foxp3* promoter^{113,115}. Thus, TNF might increase T_{reg} cell suppression and stability by favouring both phosphorylation of STAT5 and FOXP3 expression, which are key determinants of these T_{reg} cell features^{139,140}. Finally, TNF limits IL-17 production by T_{reg} cells by activating TNFAIP3⁷¹.

Other mechanisms have been proposed to explain how TNF might decrease T_{reg} cell function. TNF decreases FOXP3 expression by increasing the expression of deleted in breast cancer 1 (DBC1) and miR-34a, which respectively promote FOXP3 degradation and reduce *FOXP3* transcription and translation^{128,141,142}. Alternatively, TNF might increase the expression of serine/threonine-protein phosphatase PP1, which dephosphorylates FOXP3, thereby decreasing its effect on T_{reg} cell suppressive function¹²⁶.

[H2] Effects on T_{reg} cell differentiation

The population of FOXP3⁺ T_{reg} cells is composed of thymic T_{reg} cells, which acquire their T_{reg} cell state during their development in the thymus, and peripheral T_{reg} cells, which acquire their T_{reg} cell state during peripheral differentiation of mature naive conventional T cells. Finally, induced T_{reg} cells can be differentiated in vitro from naive conventional T cells by TCR stimulation in the presence of IL-2 and TGF-β. Thus, induced T_{reg} cells are the in vitro counterpart of peripheral T_{reg} cells.

However, whereas TNF alone has no effect on thymic T_{reg} cell differentiation, experiments in mice show that TNF inhibits the differentiation of induced T_{reg} cells, whereas treatment with TNF inhibitors increased the differentiation of induced T_{reg} cells^{143,144}. This inhibitory effect of TNF was also observed on peripheral T_{reg} cells in vivo. In mice with EAE, injection of anti-TNF or anti-TNFR2 mAbs at the time of disease induction led to reduced

disease severity, which was associated with an increased proportion of T_{reg} cells and evidence of increased peripheral T_{reg} cell differentiation¹⁴⁴. Two other papers do not support this observation and even suggest that the TNF–TNFR2 axis promotes the differentiation of induced T_{reg} and peripheral T_{reg} cells^{28,73}. However, the design of these two studies meant that contaminating natural T_{reg} cells were present in the starting inoculum, and thus treatment with TNF might boost the expansion of these contaminating cells rather than increase the differentiation of induced T_{reg} cells^{28,73}. TNF does not seem to affect thymic T_{reg} cell differentiation at steady state, because mice lacking TNFR2 have normal thymic T_{reg} cell numbers. However, ablation or neutralization of TNFR2 combined with ablation or neutralization of two other members of the TNFR superfamily, namely OX40 and GITR, led to reduced differentiation of thymic T_{reg} cells¹⁴⁵. Overall, whereas the effect of TNF on T_{reg} cell differentiation is still open to discussion, an excess of TNF seems to impair the differentiation of induced T_{reg} cells and peripheral T_{reg} cells in mice.

In humans, the inhibition of T_{reg} cell differentiation by TNF was first observed in patients with RA. TNF inhibitor treatment increased the in vitro differentiation of induced T_{reg} cells derived from patients with RA but not those from healthy controls¹⁴⁶. This observation explained why blood samples from patients with RA treated with infliximab had an increased proportion of T_{reg} cells, which might result from increased differentiation of peripheral T_{reg} cells^{98,146}. Other members of the TNF family, such as 41BB, OX40 or TNFR superfamily member 25 (also known as death receptor 3 (DR3)), can also inhibit the differentiation of induced T_{reg} cells^{147–149}. These observations suggest that a shared mechanism is involved, perhaps implicating NF-κB, PI3K or MAPK pathways. IFNγ produced by T cells following TNFR co-stimulation has also been proposed to inhibit the differentiation of induced T_{reg} cells. Alternatively, the increased activation of the PI3K–AKT pathway resulting from TNFR signalling could lead to reduced activation of phosphorylated SMAD3, which transactivates *Foxp3* expression in mouse induced T_{reg} cells¹⁴⁴.

[H2] Overall effects of TNF on T_{reg} cells

In summary, TNF has multiple negative and positive effects on T_{reg} cell biology, most probably resulting from TNFR2 rather than TNFR1 signalling (**Figure 2**). The best-characterized of the positive effects of TNF are increased T_{reg} cell proliferation and expansion. TNF also seems to promote T_{reg} cell survival in vitro, although the relevance of this effect in vivo is difficult to evaluate. The TNF-dependent increases in T_{reg} cell proliferation and survival are at least partially dependent on RelA and activation of the canonical NF-κB pathway. Involvement of p38 and

PI3K–AKT pathway activation has also been suggested but requires further investigation. Finally, weak evidence indicates that TNF increases the stability and suppressive function of T_{reg} cells, a phenomenon that might be partially due to TNF signalling synergizing with IL-2 signalling and with phosphorylation of STAT5. Other reports suggest a negative effect of TNF on T_{reg} cell biology in vitro. Whether this cytokine truly has a negative effect on T_{reg} cell function is questionable. By contrast, TNF seems to increase T_{reg} cell suppressive function in vivo, at least in some inflammatory contexts. However, the evidence of an inhibitory effect of TNF on differentiation of induced T_{reg} cells is fairly solid and might involve the PI3K–AKT pathway (**Figure 2**).

[H1] T_{reg} cells in RA

As T_{reg} cells have an important role in the suppression of autoimmunity, numerous studies have attempted to identify whether these cells have a quantitative or functional defect in patients with autoimmune diseases. Major drawbacks of these studies include the use of sample sizes too small to account for interindividual variability and the absence of a specific marker for human T_{reg} cells, which has led to the utilization of different combinations of markers. As a result, the literature is full of conflicting data.

I present here the main findings on T_{reg} cell proportion and function in patients with RA. This disease is particularly interesting as T_{reg} cells can be obtained from both the blood and joints (the target tissue of the disease), which are easily accessible for analysis. Data obtained in other autoimmune diseases are also included where relevant.

[H2] T_{reg} cell proportion

Contrasting findings have been reported in studies of the proportion of T_{reg} cells in the blood of patients with RA receiving conventional immunosuppressive treatment (the effects of TNF inhibitors are discussed below). Among studies that compared patients with RA with healthy control individuals, four described a decreased T_{reg} cell proportion^{75,150-152}, five found no difference^{98,126,146,153,154} and one found an increased T_{reg} cell proportion in the patients with RA¹⁵⁵.

Most studies that have analyzed both blood and synovial fluid of patients with RA concluded that the proportion of T_{reg} cells was higher in synovial fluid than in blood, and remained stable over time in individual patients^{152,155-157}. The T_{reg} cells isolated from synovial fluid seem to be bona fide T_{reg} cells because they exhibit *FOXP3* promoter demethylation. Also, the phenotype of these T_{reg} cells indicates that they have an activated status¹⁵⁷. The synovial fluid

of patients with RA contains high amounts of IL-6, TNF and IFN γ , low levels of IL-17A, IL-10 or IL-13 and does not contain IL-1^{126,157}. Which of these factors is responsible for the increased proportion and activation of synovial T_{reg} cells remains unclear. However, IL-6 is not likely to be involved, because this cytokine (which is produced by joint fibroblasts) induces transdifferentiation of T_{reg} cells into highly pathogenic T_H17 cells in a mouse model of autoimmune arthritis, a phenomenon that might also take place in patients with RA¹⁵⁸. IL-6 also induced proteasomal degradation of FOXP3 and loss of the suppressive activity of T_{reg} cells^{159,160}. We do not know much about the effect of IFN γ on T_{reg} cells. Therefore, the activation and/or expansion of T_{reg} cells in the synovial fluid of patients with RA is likely to be caused by high local levels of TNF.

[H2] T_{reg} cell function

Compared with T_{reg} cells obtained from the blood of healthy control individuals, T_{reg} cells isolated from the blood of patients with RA were shown to have similar suppressive activity in one study¹⁵⁵ and decreased suppressive activity in another¹²⁸. In a third study, the capacity of these cells to suppress conventional T cell proliferation was maintained but their cytokine production was reduced⁹⁸. Contrasting findings have also been reported for the suppressive activity of T_{reg} cells isolated from the synovial fluid of patients with RA. Several studies showed that synovial fluid T_{reg} cells from patients with RA were as active, or were more active, than blood T_{reg} cells from either patients with RA or healthy control individuals in terms of suppression of proliferation or IFN γ production^{152,155-157}. Another publication reported that synovial fluid T_{reg} cells from patients with RA had decreased suppressive activity¹²⁶. Importantly, these studies noted considerable variation between patients, with T_{reg} cells from some individuals but not others showing a high level of suppression¹⁵⁷. This observation might explain the contrasting results and further emphasizes the importance of generating data from at least 7–10 different patients, which was not the case for most of these studies.

Firm conclusions are difficult to draw because the available evidence does not provide a clear picture of whether T_{reg} cells in the blood of patients with RA have similar proportions and functions to those of healthy control individuals. The situation is a little bit clearer for synovial fluid T_{reg} cells, which seem to be present at an increased proportion in patients with RA.

[H2] Effects of TNF inhibitors

[H3] T_{reg} cell proportion

The proportion of T_{reg} cells in the blood has been analyzed in many studies of patients with RA 3–6 months (typically 3 months) after initiation of TNF inhibitor treatment. In studies of infliximab-treated patients with RA, the T_{reg} cell proportion increased^{75,98,115,146,151} after treatment (**Table 4**). In studies of patients with RA treated with either adalimumab or etanercept, the T_{reg} cell proportion was either increased^{150,154,161} or unchanged^{153,154,162} (**Table 4**). This T_{reg} cell increase was more often observed in responding than in non-responding patients.

Moreover, in studies of infliximab-treated patients with Crohn disease or IBD (**Supplementary Table 1**), the T_{reg} cell proportion was also either unchanged^{99,163} or increased^{100,115,163-168}. Some of the studies in patients with IBD or Crohn disease also analyzed the kinetics of this treatment-related increase in T_{reg} cell proportion. In a study of patients with Crohn disease, the increase was transient and only occurred after the first injection¹⁶⁵. In two studies of patients with IBD, the increase occurred 2 weeks after the first injection and was maintained for ≥22 weeks^{100,166}, whereas in another study in patients with Crohn disease no increase was detected after 1 week but an increase was detected at week 24 in patients who had low T_{reg} cell proportions before treatment⁹⁹ (**Supplementary Table 1**).

Two studies in patients with uveitis^{169,170} and one in patients with ankylosing spondylitis¹⁷¹ showed an increase in the T_{reg} cell proportion after TNF inhibitor therapy. However, one study in patients with juvenile idiopathic arthritis observed no difference¹⁷² and one in patients with sarcoidosis observed a decrease in the T_{reg} cell proportion¹⁷³ following TNF inhibitor therapy (**Table 4**).

Some general conclusions can be drawn from these data. Most publications described an increase in the proportion of T_{reg} cells in blood after TNF inhibitor therapy. Discrepancies between some studies could be due to the following factors: first, infliximab seems to induce an increase in the T_{reg} cell proportion more consistently than either adalimumab or etanercept. Second, a T_{reg} cell increase seems to be more consistent among patients who responded to TNF inhibitor treatment. The type of concomitant medications might also matter. For instance, although methotrexate monotherapy induces an increase in T_{reg} cell proportion¹⁵⁰, combination therapy with methotrexate and a TNF inhibitor provided an optimal increase in T_{reg} cells in vitro¹⁷⁴. Also, steroid treatment might increase T_{reg} cell proportion and function^{175,176}. Finally, as discussed above, technical factors related to the way that T_{reg} cells were purified might influence the conclusions of these studies. Some activated conventional T cells (which also express CD25) are likely to contaminate the population identified as T_{reg} cells. Thus, the findings of these studies have to be considered carefully because the level of conventional T cell contamination could differ between healthy control individuals and patients with rheumatic or autoimmune

disease, or before and after TNF inhibitor treatment. Use of the CD45RA (naive T cell) or CD45RO (memory T cell) markers, in addition to CD25 or CD127, would help to limit the risk of such contamination¹³⁴.

Several mechanisms by which T_{reg} cells might increase after TNF inhibitor treatment are supported by experimental evidence. First, T_{reg} cells might increase because treatment with TNF blockers such as infliximab favour the differentiation of peripheral T_{reg} cells¹⁴⁶. Second, T_{reg} cells might increase because treatment with anti-TNF mAbs such as adalimumab augments the expression of transmembrane TNF on monocytes, which then triggers T_{reg} cell expansion via TNFR2 signalling. Thus, anti-TNF mAbs that are intended to inhibit TNF might paradoxically increase its activity⁸⁴. The preferential expansion of activated T_{reg} cells rather than resting T_{reg} cells in patients receiving anti-TNF mAbs supports this hypothesis¹⁰⁰. In patients with RA¹⁵¹ or IBD¹⁶⁸, T_{reg} cells that are more sensitive to spontaneous apoptosis than are those of healthy control individuals, might be present at an increased proportion in patients treated with TNF inhibitors because they are protected from cell death by this therapy. In patients with IBD, TNF inhibitor therapy blocks T_{reg} cell migration to inflamed tissues, which results in increased T_{reg} cell levels in blood and decreased levels in the intestinal mucosa¹⁶⁶. Finally, TNF inhibitor therapy leads to a decrease in inflammatory cytokine levels and pathogenic T cells while sparing T_{reg} cells in patients with Crohn disease¹⁶¹ or ankylosing spondylitis^{165,171}. Therefore, this treatment might target conventional T cells in preference to T_{reg} cells, thereby explaining the relative increase in the T_{reg} cell proportion within the population of CD4⁺ T cells. As the increased proportion of blood T_{reg} cells following TNF inhibitor treatment is an in vivo phenomenon that occurs over a long time period, determining which of the above-described mechanisms is most relevant remains a challenge.

[H3] Suppressive function.

Treatment with mAb TNF inhibitors affects not only the T_{reg} cell proportion but also their suppressive function. Early work showed that T_{reg} cells from patients with RA obtained before the initiation of TNF inhibitor therapy had a poor capacity to suppress cytokine production by conventional T cells, and that the suppressive activity of these T_{reg} cells was restored following anti-TNF treatment⁹⁸. These functional T_{reg} cells resulted from either the generation of new peripheral T_{reg} cells following infliximab treatment¹⁴⁶ or from the expansion of differentiated T_{reg} cells following adalimumab treatment⁸⁴. These restored T_{reg} cells were even able to suppress pathogenic T_H17 cells, unlike the T_{reg} cells of healthy control individuals¹⁵⁴. Dysfunction of T_{reg} cells obtained from the blood or synovial fluid of patients with RA and restoration of their

suppressive function after TNF inhibitor treatment (infliximab) were also confirmed in two other studies^{126,128}. Restoration of functional blood T_{reg} cells after TNF inhibitor treatment has also been described in patients with IBD¹⁶⁴.

TNF inhibitors based on mAbs seem to act, at least in part, by restoring the functional T_{reg} cell compartment. By contrast, etanercept is likely to act by suppressing conventional T cells and/or rendering them sensitive to the suppressive effects of T_{reg} cells^{172,177}. In another study, T_{reg} cells obtained from patients with Crohn disease were shown to be functional even before initiation of infliximab treatment¹⁷⁸. However, the T_{reg} cell purification strategy used in this paper meant that activated conventional T cells might have contaminated the population of T_{reg} cells, thereby resulting in an inaccurate measurement of the suppressive activity of genuine T_{reg} cells. Therefore, the conclusions of this report have to be interpreted with caution.

In summary, the beneficial effects of TNF inhibitor therapies could be due to either restoration of fully functional T_{reg} cells or to an increased susceptibility of conventional T cells to the suppressive effects of T_{reg} cells.

[H3] T_{reg} cell biomarkers of response.

The development of biomarkers to identify the 20–30% of patients with RA or IBD who will not respond to TNF inhibitor therapy is highly desirable, and some T_{reg} cell-related biomarkers are potential candidates. In some studies, an increase in the T_{reg} cell proportion after TNF inhibitor treatment was observed only in patients who responded to this therapy (**Table 4**). Thus, the T_{reg} cell proportion before TNF inhibitor treatment has been proposed as a predictive biomarker to predict treatment response. However, patients with IBD who respond to TNF inhibitors could have T_{reg} cell proportions before therapy that are either higher^{99,100} or lower¹⁷⁸ than those of non-responding patients. Moreover, in patients with ankylosing spondylitis^{171,174} or RA^{171,174}, the T_{reg} cell proportion before TNF inhibitor therapy was not predictive of treatment efficacy.

As discussed above, a possible mechanism for the observed increase of T_{reg} cells upon mAb TNF inhibitor treatment is binding of the mAb to transmembrane TNF on myeloid cells, leading first to its increased expression and then to a boost in T_{reg} cell numbers mediated by TNFR2 signalling⁸⁴. As the expression of transmembrane TNF on monocytes can be readily assessed by flow cytometry, the capacity of adalimumab to provoke an increase in T_{reg} cell numbers in a 3-day culture has been used to identify which patients with RA would respond to this treatment¹⁷⁴.

In summary, pretreatment T_{reg} cell proportion does not seem to be a reliable biomarker of response to anti-TNF therapies. The expression of transmembrane TNF on myeloid cells as a biomarker of treatment response deserves to be confirmed in other studies.

[H1] Next-generation drugs targeting TNF

The putative mechanisms underlying non-response and paradoxical autoimmunity to TNF inhibitor treatment could be explained by the regulatory aspect of TNF. Blocking TNF is associated with an increased risk of impairing the activity of some suppressor cells, including T_{reg} cells, or increasing the activation of autoreactive T cells. The overall effect of these treatments is likely to depend on the specific autoimmune disease present, its stage and severity, and on genetic and environmental factors unique to each patient. At the time of treatment, if TNF has a dominant inflammatory and pathogenic role, TNF inhibitors will be beneficial. By contrast, if TNF has a dominant regulatory and protective role, TNF inhibitors will be detrimental.

Given that most of the pro-inflammatory properties of TNF are due to TNFR1 signalling induced by soluble TNF and most of the regulatory properties of are due to TNFR2 signalling induced by transmembrane TNF, the next generation of TNF inhibitors might preferentially target TNFR1 or TNFR2^{14,25,82,86,88,179,180}. Two types of TNFR-specific agents have been proposed: mAbs and so-called TNF muteins, which are forms of this cytokine harbouring mutations in the receptor-interacting domains¹⁸¹⁻¹⁸³.

[H2] Selective TNFR1 antagonists

The capacity of TNFR1 antagonists to block the pro-inflammatory interaction of TNF with TNFR1 has been investigated in mouse models of autoimmune diseases.

Both mAbs and TNF muteins have been developed that have potent TNFR1 antagonist activity and a strong therapeutic effect in mouse models of autoimmune diseases (**Table 5**). For example, the mAb DMS5540 was as effective as etanercept in the treatment of collagen-induced arthritis. In addition, the effects of DMS5540 on immune cells were superior to those of etanercept, as DMS5540 induced T_{reg} cell activation and reduced the activation of conventional T cells, a phenomenon not observed with etanercept¹⁸⁴. Several anti-TNFR1 mAbs (namely atosab, trivalent nanobody TNFR1 silencer (TROS) and HM1097) were able to suppress EAE¹⁸⁵⁻¹⁸⁷. Finally, the muteins XPro1595 and R1antTNF had therapeutic effects in arthritis or EAE; when these agents were compared with etanercept, they sometimes demonstrated improved efficacy¹⁸⁸⁻¹⁹².

[H2] TNFR2 agonists

As TNFR2 signalling stimulates the expansion of T_{reg} cells, TNFR2 agonists such as the mAbs MR2-1 and another unnamed version are interesting candidates for improving T_{reg} cell therapy in autoimmune diseases (**Table 5**). When added to human T_{reg} cell cultures, these mAbs promote the expansion and improve the stability and purity of T_{reg} cells over time^{111,118}.

Two additional TNF muteins with human TNFR2 agonist activity (TNF07 and TNC-scTNFR₂) have been generated^{116,193} and TNF07 has been shown to promote T_{reg} cell activation in vitro. In the future, mAbs or TNF muteins with TNFR2 agonist activity might be used to improve cell culture methods used to generate T_{reg} cell preparations for use in cell therapy. This notion is supported by mouse studies showing that adding TNF or a TNFR2 agonist to T_{reg} cell cultures increased the capacity of these cells to suppress colitis^{103,130} or GvHD^{103,130} after their reintroduction in vivo^{103,130}.

The capacity of TNFR2 agonists to stimulate T_{reg} cells in vivo has been tested in mouse models. Treatment with either of two TNF muteins with TNFR2 agonist activity (STAR2 and EHD2-sc-mTNFR₂)^{194,195} induced in vivo T_{reg} cell activation and expansion^{117,120} that was associated with prevention or amelioration of arthritis^{119,121}, EAE¹³² or GvHD¹¹⁷. These agents also protected the central nervous system of treated animals from inflammation and neuronal injury induced by chronic nerve constriction or drug treatment, respectively^{195,196}.

Whereas classical immunosuppressive drugs aim to suppress autoimmunity by neutralizing pathogenic cells, an alternative approach is based on increasing the expansion or suppressive capacity of T_{reg} cells. A prototype of this new class of drugs is IL-2¹⁹⁷. Our group showed that administration of low-dose IL-2 boosts the proliferation of T_{reg} cells and induces remission of type 1 diabetes mellitus in non-obese diabetic mice^{198,199}. Low-dose IL-2 is now being investigated as a treatment for other autoimmune diseases in multiple clinical trials. One study has investigated this treatment in 14 different autoimmune diseases, including RA and ankylosing spondylitis (NCT01988506). TNFR2 agonists are another type of drug that are able to boost the number or function of T_{reg} cells. No clinical trial has so far investigated the use of TNFR2 agonists to treat an autoimmune disease. However, bacillus Calmette–Guérin (BCG) vaccine can induce TNF release without secondary effects, thereby providing an indirect way to trigger TNFR2 signalling. BCG vaccine has been tested for efficacy in type 1 diabetes (NCT00607230 and NCT02081326).

[H1] Conclusions

TNF has a long and fascinating yet chaotic history. This cytokine was discovered in the mid-

1970s and named for its effect as a tumour cell killer. Major milestones in its history include its cloning in the mid-1980s, the discovery that TNF binds to two receptors, that its signalling transduction is highly complex (and still remains to be fully explored), and that it has multiple effects at steady state.

TNF is now known to be one of the most important inflammatory cytokines. Although TNF is critical for beneficial immune responses, the realization that TNF is also harmful in many autoimmune diseases led to the great success of TNF inhibitors and ultimately to the flowering of research into other biological therapies. The regulatory role of this cytokine is also important to consider. Here again, the mechanisms underlying the immunosuppressive activity of TNF are complex. However, one of its main features seems to involve the expression of transmembrane TNF on myeloid or T cells, which interacts with TNFR2 on T_{reg} cells to boost their proliferation and maybe also their stability and suppressive function.

The inflammatory and regulatory roles of TNF are both essential to take into account in the design of future generations of TNF inhibitors. Preclinical studies have shown that selective antagonists of TNFR1 inhibit the inflammatory action of TNF whereas selective agonists of TNFR2 boost T_{reg} cell numbers and potentially also improve their function. Therefore, TNFR1 antagonists and TNFR2 agonists could be beneficial in future treatments of several diseases with an autoimmune component. In the future, biotechnology and pharmaceutical companies are expected to work hand in hand with academic laboratories towards the successful translation of these fascinating observations into the clinic.

Acknowledgements

The author thanks current and past members of his laboratory for their hard work and fruitful and passionate exchanges on the effects of TNF on T_{reg} cells. His research work is supported by Agence Nationale de la Recherche (grants ANR-15-CE15-0015-03 and ANR-17-CE15-0030-01) and Fondation pour la Recherche Médicale (équipe FRM).

Competing interests

B.S. declares that he received consultancy fees from HiFiBio Therapeutics regarding the applications of TNFR2 agonists and antagonists in cancer and autoimmunity.

Peer review information

Nature Reviews Rheumatology thanks [Referee#1 name], [Referee#2 name] and the other, anonymous, reviewer(s) for their contribution to the peer review of this work.

778

779 **Supplementary information**

780 Supplementary information is available for this paper at [https://doi.org/10.1038/s415XX-XXX-](https://doi.org/10.1038/s415XX-XXX-XXXX-X)
781 XXXX-X

782

783 **Key points**

- 784 • Tumour necrosis factor (TNF) is a major inflammatory cytokine that has deleterious
785 effects in several rheumatic and autoimmune diseases, as attested by the success of TNF
786 inhibitor therapy.
- 787 • Some patients do not respond to TNF inhibitors and others develop paradoxical
788 autoimmune exacerbations that can be explained by the immunoregulatory role of TNF.
- 789 • The pro-inflammatory and anti-inflammatory properties of TNF are largely segregated by
790 the capacity of this cytokine to bind to TNF receptor 1 (TNFR1) and TNF receptor 2
791 (TNFR2), respectively.
- 792 • The anti-inflammatory effects of TNF are explained by its capacity to increase the
793 proliferation, stability and suppressive function of FOXP3⁺ regulatory T cells via TNFR2
794 signalling.
- 795 • Antagonists of TNFR1 and agonists of TNFR2 constitute a new generation of drugs that
796 might be more effective and have fewer adverse effects than classical TNF inhibitors.

797

798

799

Table 1. Clinically approved TNF inhibitors in the USA and Europe

Drug	Molecule	Biosimilars	Approved rheumatic disease indications ^a
Etanercept	Human TNFR2—IgG1–Fc fusion protein	Benpali, Erelzi, Nepexto	RA, JIA, psoriatic arthritis, plaque psoriasis, AS
Infliximab	Humanized chimeric anti-TNF IgG1/κ mAb	Remsima, Inflectra, Flixabi, Ixifi, Zessly, Avsola	RA, psoriatic arthritis, plaque psoriasis, AS
Adalimumab	Fully human anti-TNF IgG1/κ mAb	Exemptia, Adfrar, Amjevita, Cyltezo, Amgevita, Solymbic, Imraldi, Cyltezo, Halimatoz, Hefiya, Hyrimoz, Hulio, Idacio, Kromea, Hadlima, Abrilada, Amsparity	RA, JIA, psoriatic arthritis, plaque psoriasis, AS, hidradenitis suppurativa, non-infectious uveitis
Certolizumab pegol	PEGylated human Fab fragment of anti-TNF mAb	NA	RA (Europe only), psoriatic arthritis
Golimumab	Fully human anti-TNF IgG1/κ mAb	NA	RA, psoriatic arthritis, AS

800

801

802

803

804

^aDisease indications for biosimilars can differ from those of the original drug and depend on the countries where they are registered. AS, ankylosing spondylitis; JIA, juvenile idiopathic arthritis; mAb, monoclonal antibody; NA, not applicable; RA, rheumatoid arthritis.

Commenté [JM1]: Caroline: Check against table 1 in the published paper below for overlap:

<https://www.mdpi.com/2073-4468/4/1/48/html?>

This table might need to be adjusted to avoid overlap (or else might need permissions).

(see handover sheet for one idea of alt layout)

805 Table 2. Pathogenic and protective roles of TNFR1 and TNFR2 in models of rheumatic and
806 autoimmune diseases

Mouse model	<i>TNFR1</i> knockout	<i>TNFR2</i> knockout	TNFR1 antagonist	TNFR2 agonist	Refs
Collagen-induced arthritis	Attenuated	Exacerbated	Attenuated	Attenuated	200,201,184,119,121
Antigen-induced arthritis	ND	Exacerbated	ND	ND	201
DTHA	ND	Exacerbated	ND	ND	115
Arthritis in TNF-transgenic mice	Attenuated	Exacerbated	ND	ND	202
EAE	Attenuated	Exacerbated	Attenuated	Attenuated	28,187,203-205,28,187,191,203,204, 206,207,208,132

807 DTHA, delayed-type hypersensitivity arthritis; EAE, experimental autoimmune
808 encephalomyelitis; ND, not determined.

809

810

811 Table 3. Effect of TNF on T_{reg} cell function in vitro.

T _{reg} cell population	Culture condition(s)	<i>n</i>	TNF added	Assay type	Effect of TNF on T _{reg} cell suppression	Refs
Human CD4 ⁺ CD25 ^{hi}	Soluble anti-CD3 and anti-CD28 mAbs	6	Before or during	Cytokine FACS	No change	98
Human CD4 ⁺ CD25 ^{hi}	Coated anti-CD3 mAbs	6	Before	Proliferation ³ H	Decreased	128
Human CD4 ⁺ CD25 ⁺	HBV e-antigen	7	None	Proliferation ³ H	Decreased	127
Human CD4 ⁺ CD25 ⁺	Coated anti-CD3 mAbs	NR	During	Proliferation FACS, cytokine ELISA	Decreased	129
Human CD4 ⁺ CD25 ^{hi}	Coated anti-CD3 mAbs	3	Before or during	Proliferation FACS	Decreased	122
Human CD4 ⁺ CD25 ^{hi} CD127 ^{low}	Coated anti-CD3 and anti-CD28 mAbs	5?	Before	Proliferation FACS	Decreased	126
Human CD4 ⁺ CD25 ^{hi} CD127 ^{low} CD45RA ⁺	Coated anti-CD3 and anti-CD28 mAbs; APC and soluble anti-CD3 mAbs; APC and coated anti-CD3 mAbs	28	Before or during	Proliferation FACS	No change or increased	133
Mouse CD4 ⁺ CD25 ⁺	APC and soluble anti-CD3 mAbs	6	Before or during	Proliferation FACS	No change or increased	22
Mouse CD4 ⁺ CD25 ⁺	APC and soluble anti-CD3 mAbs	3	Before	Proliferation FACS	Increased	107

812 APC, antigen presenting cells; ELISA, enzyme-linked immunosorbent assay; FACS,
813 fluorescence-activated cell sorting; *n*, number of healthy individuals; NR, not reported; TNF,
814 tumour necrosis factor.

815

816

817 Table 4. T_{reg} cell proportions in blood before and after TNF inhibitor therapy

Study population	TNF inhibitor (concomitant medications)	Sampling time points ^a	T _{reg} cells			Refs
			Cell population	Pre-treatment (proportion) ^b	Post-treatment (proportion)	
27 patients with RA; 8 healthy controls	Infliximab (NSAIDs, methotrexate)	Baseline, 1.5 and 3.0 months	CD4 ⁺ CD25 ^{hi}	Same	Increased from baseline; increased in responders vs nonresponders	98
17 patients with RA; 15 healthy controls	Infliximab (NSAIDs, methotrexate)	Baseline and 3.0 months	CD4 ⁺ CD25 ^{hi}	Decreased ^c	Increased ^c from baseline	151
31 patients with RA; 20 healthy controls	Infliximab (NSAIDs, methotrexate)	Baseline and 4.0–6.0 months	CD4 ⁺ FOXP3 ⁺	Same	Increased from baseline	146
40 patients with RA; 10 healthy controls	Infliximab (methotrexate, salazopyrin, hydroxychloroquine, steroids)	NR	CD4 ⁺ CD25 ⁺ FOX P3 ⁺	Decreased	Increased from baseline and in responders vs nonresponders	75
10 patients with RA; 10 healthy controls	Adalimumab (NSAIDs, methotrexate, steroids)	Baseline and 3.0 months	CD4 ⁺ CD25 ^{hi}	Same	No change from baseline	153
50 patients with RA; 15 healthy controls	Adalimumab or etanercept (NR)	NR	CD4 ⁺ FOXP3 ⁺	Same	Increased from baseline; increased in responders vs nonresponders to adalimumab; no change from baseline with etanercept	154
48 patients with RA	Adalimumab or etanercept (methotrexate, leflunomide)	Baseline, 1.5 and 3.0 months	CD4 ⁺ FOXP3 ⁺ , CD25 ^{hi} 127 ^{low}	ND	No change from baseline, no difference between responders and nonresponders	162
20 patients with RA; 10 healthy controls	Etanercept (methotrexate)	Baseline and 3.0 months	CD4 ⁺ CD25 ^{hi} FOX P3 ⁺	Decreased	Increased from baseline	150
33 patients with RA	Etanercept (methotrexate)	Baseline, 3.0 and 6.0 months	CD4 ⁺ CD25 ⁺ FOX P3 ⁺	ND	Increased from baseline	161
16 patients with RA	Infliximab or etanercept ^d (NR)	Baseline and 3.0 months	CD4 ⁺ CD25 ⁺ 127 ^{lo} w FOX P3 ⁺	ND	Increased from baseline	115
7 patients with JIA	Etanercept (NSAIDs, methotrexate)	Baseline and 1.0–5.0 months	CD4 ⁺ FOXP3 ⁺	ND	No change from baseline	172
222 patients with AS; 68 healthy controls	Infliximab or etanercept (NSAIDs)	Baseline and 6.0 months	CD4 ⁺ CD25 ^{hi} FOX P3 ⁺	Decreased	Increased from baseline; increased in responders versus nonresponders	171
46 patients with sarcoidosis; 26 healthy controls	Infliximab (NR)	Baseline, 3.5 and 6.0 months	CD4 ⁺ CD25 ^{hi}	Increased	Decreased from baseline	173
12 patients with uveitis	Adalimumab (NR)	Baseline, 1.0 and 6.0 months	CD4 ⁺ CD25 ^{hi} 127 ^{lo} w FOX P3 ⁺	ND	Increased from baseline	169
16 patients with uveitis; 15 healthy controls	Infliximab (NR)	Baseline, 4.0–27.0 months	CD4 ⁺ FOXP3 ⁺	Same	Increased from baseline ^e	170

818 ^aBaseline (before initiation of TNF inhibitor treatment). ^bIn patients versus controls. ^cAbsolute

819 number. ^dThree patients also received golimumab, adalimumab or certolizumab. ^eVersus patients

820 treated only with ciclosporine or colchicine. AS, ankylosing spondylitis; JIA, juvenile idiopathic
821 arthritis; ND, not determined; NR, not reported; NSAID, nonsteroidal anti-inflammatory drug.

822

823 **Table 5. Therapeutic effects of drugs targeting TNFRs in autoimmune disease models**

Agent	Structure	Therapeutic efficacy	Refs
<i>Antagonists of TNFR1</i>			
DMS5540	Bispecific anti-TNFR1 and anti-albumin mAb	Arthritis (CIA)	184
ASTROSAB	Humanized anti-TNFR1 IgG1 mAb, mutated in the Fc fragment to abrogate complement and immune complex activation	EAE	185
TROS	Trivalent nanobody comprising two mAb domains binding to TNFR1 and one mAb domain binding to albumin	EAE	186
HM1097	Hamster IgG	EAE	187
XPro1595	Dominant-negative PEGylated TNF muteins that interact with soluble TNF to form inactive heterotrimers, which have low binding and signalling activity	Arthritis (CIA) and EAE	188,190,192
R1antTNF	PEGylated TNF mutein that binds specifically to TNFR1 without signalling activity	Arthritis (CIA) and EAE	189,191
<i>Agonists of TNFR2</i>			
MR2-1	Mouse mAb against human TNFR2	Increased expansion and stability of T _{reg} cells; not tested in vivo	57, 118
Unnamed	Mouse mAb against human TNFR2	Increased expansion and stability of T _{reg} cells; not tested in vivo	111
TNF07	Human TNF mutein trimer	Increased expansion of T _{reg} cells; not tested in vivo	116
TNC-scTNFR ₂	Human TNF mutein trimer	Not tested in vitro or vivo	193
STAR2	Mouse TNF mutein nanomer	Increased expansion, survival and function of T _{reg} cells; effective in CIA, EAE and GvHD	121,103,117,120,132
EHD2-sc-mTNFR ₂	Mouse TNF mutein hexamer	Increased expansion of T _{reg} cells; effective in EAE and CIA	119,208

824 CIA, collagen-induced arthritis; EAE, experimental autoimmune encephalomyelitis; GvHD,
825 graft versus host disease; mAb, monoclonal antibody; TNF, tumour necrosis factor; TNFR, TNF
826 receptor.

827

Figure 1. The proinflammatory and anti-inflammatory activities of TNF are driven by effects on innate and adaptive immunity. Tumour necrosis factor (TNF) is a major proinflammatory cytokine (top panel) that activates both innate (left side) and adaptive (right side) immunity. TNF promotes recruitment of leukocytes, favours the production of other proinflammatory cytokines, activates neutrophils and participates in co-stimulation of conventional T cells. TNF also has regulatory activities (bottom panel) such as inhibition of haematopoiesis, increased glucocorticoid production, activation of suppressive cells (such as mesenchymal stem cells (MSC) and myeloid-derived suppressor cells (MDSC)) or altering the function of dendritic cells (DCs) and macrophages. TNF also regulates immunity by promoting IL-10-producing B cells, inducing T cell apoptosis, altering T cell receptor (TCR) signalling, inhibiting T_H17 cell differentiation and boosting numbers and function of regulatory T (T_{reg}) cells. APC, antigen-presenting cell; FLS, fibroblast-like synoviocyte; HSC, haematopoietic stem cell.

Figure 2. The overall effects of TNF on regulatory T cells. Most of the effects of tumour necrosis factor (TNF) on regulatory T (T_{reg}) cells are due to induction of TNF receptor 1 (TNFR2) signalling, which is probably preferentially mediated by transmembrane TNF rather than soluble TNF. Signal transduction downstream of TNFR2 that does not involve kinase activity involves TNF receptor-associated factor (TRAF) adaptor proteins. Multiple downstream signalling pathways lead to positive (left) and negative (right) effects on T_{reg} cell biology. TNFR2 signalling strongly induces T_{reg} cell proliferation and has a moderate survival-promoting effect on T_{reg} cells; both of these effects depend on RelA and probably also on the activation of p38, AKT and mTORC1 by phosphorylation (p). Weak evidence indicates that TNF also promotes the stability and suppressive function of T_{reg} cells, perhaps via TNF-induced protein 3 (TNFAIP3) and signal transducer and activator of transcription 5 (STAT5) signalling pathways. In addition to these positive effects of TNF, the negative effects of this cytokine are clear in relation to the inhibition of induced T_{reg} cell differentiation (which involves phosphoinositide 3-kinase (PI3K) and/or phosphorylated RAC α serine/threonine-protein kinase (AKT) pathway activation). Weak evidence suggests that TNF induces T_{reg} cell dysfunction, perhaps via a mechanism involving deleted in breast cancer 1 (DBC1), microRNA 34a (miR-34a) and serine/threonine-protein phosphatase PP1. Arrow thickness and box shading intensity is proportional to the importance of the effect or the likelihood that a given molecule is involved in the pathway.

References

1. Feldmann, M. Translating molecular insights in autoimmunity into effective therapy. *Annu Rev Immunol* **27**, 1-27 (2009).
2. Monaco, C., Nanchahal, J., Taylor, P. & Feldmann, M. Anti-TNF therapy: past, present and future. *Int Immunol* **27**, 55-62 (2015).
3. Keffer, J. et al. Transgenic mice expressing human tumour necrosis factor: a predictive genetic model of arthritis. *EMBO J* **10**, 4025-31 (1991).
4. Silva-Fernandez, L. & Hyrich, K. Rheumatoid arthritis: When TNF inhibitors fail in RA-weighing up the options. *Nat Rev Rheumatol* **10**, 262-4 (2014).
5. Roda, G., Jharap, B., Neeraj, N. & Colombel, J.F. Loss of Response to Anti-TNFs: Definition, Epidemiology, and Management. *Clin Transl Gastroenterol* **7**, e135 (2016).
6. Esposito, M. et al. Survival rate of antitumour necrosis factor-alpha treatments for psoriasis in routine dermatological practice: a multicentre observational study. *Br J Dermatol* **169**, 666-72 (2013).
7. Ramos-Casals, M., Brito-Zeron, P., Soto, M.J., Cuadrado, M.J. & Khamashta, M.A. Autoimmune diseases induced by TNF-targeted therapies. *Best Pract Res Clin Rheumatol* **22**, 847-61 (2008).
8. Ramos-Casals, M. et al. Autoimmune diseases induced by biological agents: a double-edged sword? *Autoimmun Rev* **9**, 188-93 (2010).
9. Brenner, D., Blaser, H. & Mak, T.W. Regulation of tumour necrosis factor signalling: live or let die. *Nat Rev Immunol* **15**, 362-74 (2015).
10. Conrad, C. et al. TNF blockade induces a dysregulated type 1 interferon response without autoimmunity in paradoxical psoriasis. *Nat Commun* **9**, 25 (2018).
11. van Oosten, B.W. et al. Increased MRI activity and immune activation in two multiple sclerosis patients treated with the monoclonal anti-tumor necrosis factor antibody cA2. *Neurology* **47**, 1531-4 (1996).
12. Study-group. TNF neutralization in MS: results of a randomized, placebo-controlled multicenter study. The Lenercept Multiple Sclerosis Study Group and The University of British Columbia MS/MRI Analysis Group. *Neurology* **53**, 457-65 (1999).
13. Aggarwal, B.B., Gupta, S.C. & Kim, J.H. Historical perspectives on tumor necrosis factor and its superfamily: 25 years later, a golden journey. *Blood* **119**, 651-65 (2012).
14. Kalliolias, G.D. & Ivashkiv, L.B. TNF biology, pathogenic mechanisms and emerging therapeutic strategies. *Nat Rev Rheumatol* **12**, 49-62 (2016).
15. Atretkhany, K.N., Gogoleva, V.S., Drutskaya, M.S. & Nedospasov, S.A. Distinct modes of TNF signaling through its two receptors in health and disease. *J Leukoc Biol* (2020).
16. Salomon, B.L. et al. Tumor Necrosis Factor alpha and Regulatory T Cells in Oncoimmunology. *Front Immunol* **9**, 444 (2018).
17. Davignon, J.L. et al. Modulation of T-cell responses by anti-tumor necrosis factor treatments in rheumatoid arthritis: a review. *Arthritis Res Ther* **20**, 229 (2018).
18. Horiuchi, T., Mitoma, H., Harashima, S., Tsukamoto, H. & Shimoda, T. Transmembrane TNF-a: structure, function and interaction with anti-TNF agents. *Rheumatology (Oxford)* **49**, 1215-1228 (2010).
19. Lee, W.H., Seo, D., Lim, S.G. & Suk, K. Reverse Signaling of Tumor Necrosis Factor Superfamily Proteins in Macrophages and Microglia: Superfamily Portrait in the Neuroimmune Interface. *Front Immunol* **10**, 262 (2019).
20. Qu, Y., Zhao, G. & H., L. Forward and Reverse Signaling Mediated by Transmembrane Tumor Necrosis Factor-Alpha and TNF Receptor 2: Potential Roles in an immunosuppressive Tumor Microenvironment. *Front Immunol* **8**, 1675 (2017).
21. Aggarwal, B.B. Signalling pathways of the TNF superfamily: a double-edged sword. *Nat Rev Immunol* **3**, 745-56 (2003).

- 912 22. Chen, X., Baumel, M., Mannel, D.N., Howard, O.M. & Oppenheim, J.J. Interaction of
913 TNF with TNF receptor type 2 promotes expansion and function of mouse CD4+CD25+
914 T regulatory cells. *J Immunol* **179**, 154-61 (2007).
- 915 23. Ware, C.F. Network communications: lymphotoxins, LIGHT, and TNF. *Annu Rev*
916 *Immunol* **23**, 787-819 (2005).
- 917 24. Yang, S., Wang, J., Brand, D.D. & Zheng, S.G. Role of TNF-TNF Receptor 2 Signal in
918 Regulatory T Cells and Its Therapeutic Implications. *Front Immunol* **9**, 784 (2018).
- 919 25. Faustman, D. & Davis, M. TNF receptor 2 pathway: drug target for autoimmune
920 diseases. *Nat Rev Drug Discov* **9**, 482-93 (2010).
- 921 26. Gregory, A.P. et al. TNF receptor 1 genetic risk mirrors outcome of anti-TNF therapy in
922 multiple sclerosis. *Nature* **488**, 508-511 (2012).
- 923 27. Park, H., Bourla, A.B., Kastner, D.L., Colbert, R.A. & Siegel, R.M. Lighting the fires
924 within: the cell biology of autoinflammatory diseases. *Nat Rev Immunol* **12**, 570-80
925 (2012).
- 926 28. Yang, S. et al. Differential roles of TNFalpha-TNFR1 and TNFalpha-TNFR2 in the
927 differentiation and function of CD4(+)Foxp3(+) induced Treg cells in vitro and in vivo
928 periphery in autoimmune diseases. *Cell Death Dis* **10**, 27 (2019).
- 929 29. Rampart, M., De Smet, W., Fiers, W. & Herman, A.G. Inflammatory properties of
930 recombinant tumor necrosis factor in rabbit skin in vivo. *J Exp Med* **169**, 2227-32 (1989).
- 931 30. Venkatesh, D. et al. Endothelial TNF receptor 2 induces IRF1 transcription factor-
932 dependent interferon-beta autocrine signaling to promote monocyte recruitment.
933 *Immunity* **38**, 1025-37 (2013).
- 934 31. Duprez, L. et al. RIP kinase-dependent necrosis drives lethal systemic inflammatory
935 response syndrome. *Immunity* **35**, 908-18 (2011).
- 936 32. Ding, X. et al. TNF receptor 1 mediates dendritic cell maturation and CD8 T cell
937 response through two distinct mechanisms. *J Immunol* **187**, 1184-91 (2011).
- 938 33. Menges, M. et al. Repetitive injections of dendritic cells matured with tumor necrosis
939 factor alpha induce antigen-specific protection of mice from autoimmunity. *J Exp Med*
940 **195**, 15-21 (2002).
- 941 34. Noti, M., Corazza, N., Mueller, C., Berger, B. & Brunner, T. TNF suppresses acute
942 intestinal inflammation by inducing local glucocorticoid synthesis. *J Exp Med* **207**, 1057-
943 66 (2010).
- 944 35. Wang, W. et al. Enhanced human hematopoietic stem and progenitor cell engraftment by
945 blocking donor T cell-mediated TNFalpha signaling. *Sci Transl Med* **9** (2017).
- 946 36. Ghannam, S., Pene, J., Moquet-Torcy, G., Jorgensen, C. & Yssel, H. Mesenchymal stem
947 cells inhibit human Th17 cell differentiation and function and induce a T regulatory cell
948 phenotype. *J Immunol* **185**, 302-12 (2010).
- 949 37. Sayegh, S. et al. Rheumatoid Synovial Fluids Regulate the Immunomodulatory Potential
950 of Adipose-Derived Mesenchymal Stem Cells Through a TNF/NF-kappaB-Dependent
951 Mechanism. *Front Immunol* **10**, 1482 (2019).
- 952 38. Raveney, B.J., Copland, D.A., Dick, A.D. & Nicholson, L.B. TNFR1-dependent
953 regulation of myeloid cell function in experimental autoimmune uveoretinitis. *J Immunol*
954 **183**, 2321-9 (2009).
- 955 39. Zhao, X. et al. TNF signaling drives myeloid-derived suppressor cell accumulation. *J*
956 *Clin Invest* **122**, 4094-104 (2012).
- 957 40. Chavez-Galan, L. et al. Transmembrane Tumor Necrosis Factor Controls Myeloid-
958 Derived Suppressor Cell Activity via TNF Receptor 2 and Protects from Excessive
959 Inflammation during BCG-Induced Pleurisy. *Front Immunol* **8**, 999 (2017).
- 960 41. Hu, X. et al. Transmembrane TNF-alpha promotes suppressive activities of myeloid-
961 derived suppressor cells via TNFR2. *J Immunol* **192**, 1320-31 (2014).

- 962 42. Sade-Feldman, M. et al. Tumor necrosis factor- α blocks differentiation and enhances
963 suppressive activity of immature myeloid cells during chronic inflammation. *Immunity*
964 **38**, 541-54 (2013).
- 965 43. Bachus, H. et al. Impaired Tumor-Necrosis-Factor- α -driven Dendritic Cell Activation
966 Limits Lipopolysaccharide-Induced Protection from Allergic Inflammation in Infants.
967 *Immunity* **50**, 225-240 e4 (2019).
- 968 44. Alzabin, S. et al. Incomplete response of inflammatory arthritis to TNF α blockade is
969 associated with the Th17 pathway. *Ann Rheum Dis* **71**, 1741-1748 (2012).
- 970 45. Mayordomo, A.C. et al. IL-12/23p40 overproduction by dendritic cells leads to an
971 increased Th1 and Th17 polarization in a model of Yersinia enterocolitica-induced
972 reactive arthritis in TNFRp55 $^{-/-}$ mice. *PLoS One* **13**, e0193573 (2018).
- 973 46. Notley, C.A. et al. Blockade of tumor necrosis factor in collagen-induced arthritis reveals
974 a novel immunoregulatory pathway for Th1 and Th17 cells. *J Exp Med* **205**, 2491-7
975 (2008).
- 976 47. Park, S.H., Park-Min, K.H., Chen, J., Hu, X. & Ivashkiv, L.B. Tumor necrosis factor
977 induces GSK3 kinase-mediated cross-tolerance to endotoxin in macrophages. *Nat*
978 *Immunol* **12**, 607-15 (2011).
- 979 48. Zakharova, M. & Ziegler, H.K. Paradoxical anti-inflammatory actions of TNF- α :
980 inhibition of IL-12 and IL-23 via TNF receptor 1 in macrophages and dendritic cells. *J*
981 *Immunol* **175**, 5024-33 (2005).
- 982 49. Kusnadi, A. et al. The Cytokine TNF Promotes Transcription Factor SREBP Activity and
983 Binding to Inflammatory Genes to Activate Macrophages and Limit Tissue Repair.
984 *Immunity* **51**, 241-257 e9 (2019).
- 985 50. Park, S.H. et al. Type I interferons and the cytokine TNF cooperatively reprogram the
986 macrophage epigenome to promote inflammatory activation. *Nat Immunol* **18**, 1104-1116
987 (2017).
- 988 51. Tartaglia, L.A. et al. Stimulation of human T-cell proliferation by specific activation of
989 the 75-kDa tumor necrosis factor receptor. *J Immunol* **151**, 4637-41 (1993).
- 990 52. Kim, E.Y. & Teh, H.S. Critical role of TNF receptor type-2 (p75) as a costimulator for
991 IL-2 induction and T cell survival: a functional link to CD28. *J Immunol* **173**, 4500-9
992 (2004).
- 993 53. Kim, E.Y., Priatel, J.J., Teh, S.J. & Teh, H.S. TNF receptor type 2 (p75) functions as a
994 costimulator for antigen-driven T cell responses in vivo. *J Immunol* **176**, 1026-35 (2006).
- 995 54. Calzascia, T. et al. TNF- α is critical for antitumor but not antiviral T cell immunity in
996 mice. *J Clin Invest* **117**, 3833-45 (2007).
- 997 55. Chen, X. et al. TNFR2 expression by CD4 effector T cells is required to induce full-
998 fledged experimental colitis. *Sci Rep* **6**, 32834 (2016).
- 999 56. Solovieva, K., Puliaiev, M., Haas, M. & Via, C.S. In vivo maturation of allo-specific
1000 CD8 CTL and prevention of lupus-like graft-versus-host disease is critically dependent
1001 on T cell signaling through the TNF p75 receptor but not the TNF p55 receptor. *J*
1002 *Immunol* **190**, 4562-72 (2013).
- 1003 57. de Kivit, S. et al. Stable human regulatory T cells switch to glycolysis following TNF
1004 receptor 2 costimulation. *Nat Metab* **2**, 1046-1061 (2020).
- 1005 58. Schioppa, T. et al. B regulatory cells and the tumor-promoting actions of TNF- α
1006 during squamous carcinogenesis. *Proc Natl Acad Sci U S A* **108**, 10662-7 (2011).
- 1007 59. Cope, A.P. et al. Chronic tumor necrosis factor alters T cell responses by attenuating T
1008 cell receptor signaling. *J Exp Med* **185**, 1573-84 (1997).
- 1009 60. Aspalter, R.M., Wolf, H.M. & Eibl, M.M. Chronic TNF- α exposure impairs TCR-
1010 signaling via TNF-RII but not TNF-RI. *Cell Immunol* **237**, 55-67 (2005).

61. Beyer, M. et al. Tumor-necrosis factor impairs CD4(+) T cell-mediated immunological control in chronic viral infection. *Nat Immunol* **17**, 593-603 (2016).
62. Qin, H.Y., Chaturvedi, P. & Singh, B. In vivo apoptosis of diabetogenic T cells in NOD mice by IFN-gamma/TNF-alpha. *Int Immunol* **16**, 1723-32 (2004).
63. Naude, P.J., den Boer, J.A., Luiten, P.G. & Eisel, U.L. Tumor necrosis factor receptor cross-talk. *FEBS J* **278**, 888-98 (2011).
64. Lin, R.H., Hwang, Y.W., Yang, B.C. & Lin, C.S. TNF receptor-2-triggered apoptosis is associated with the down-regulation of Bcl-xL on activated T cells and can be prevented by CD28 costimulation. *J Immunol* **158**, 598-603 (1997).
65. Ban, L. et al. Selective death of autoreactive T cells in human diabetes by TNF or TNF receptor 2 agonism. *Proc Natl Acad Sci U S A* **105**, 13644-9 (2008).
66. Bhattacharyya, S. et al. Tumor-induced oxidative stress perturbs nuclear factor-kappaB activity-augmenting tumor necrosis factor-alpha-mediated T-cell death: protection by curcumin. *Cancer Res* **67**, 362-70 (2007).
67. Kim, E.Y., Teh, S.J., Yang, J., Chow, M.T. & Teh, H.S. TNFR2-deficient memory CD8 T cells provide superior protection against tumor cell growth. *J Immunol* **183**, 6051-7 (2009).
68. Luckey, U. et al. T cell killing by tolerogenic dendritic cells protects mice from allergy. *J Clin Invest* **121**, 3860-71 (2011).
69. Miller, P.G., Bonn, M.B. & McKarns, S.C. Transmembrane TNF-TNFR2 Impairs Th17 Differentiation by Promoting IL2 Expression. *J Immunol* **195**, 2633-47 (2015).
70. Urbano, P.C.M. et al. TNF-alpha-induced protein 3 (TNFAIP3)/A20 acts as a master switch in TNF-alpha blockade-driven IL-17A expression. *J Allergy Clin Immunol* **142**, 517-529 (2018).
71. Urbano, P.C.M. et al. TNFalpha-Signaling Modulates the Kinase Activity of Human Effector Treg and Regulates IL-17A Expression. *Front Immunol* **10**, 3047 (2019).
72. Elicabe, R.J. et al. Lack of TNFR p55 results in heightened expression of IFN-gamma and IL-17 during the development of reactive arthritis. *J Immunol* **185**, 4485-95 (2010).
73. Ma, H.L. et al. Tumor necrosis factor alpha blockade exacerbates murine psoriasis-like disease by enhancing Th17 function and decreasing expansion of Treg cells. *Arthritis Rheum* **62**, 430-40 (2010).
74. Hull, D.N. et al. Anti-tumour necrosis factor treatment increases circulating T helper type 17 cells similarly in different types of inflammatory arthritis. *Clin Exp Immunol* **181**, 401-6 (2015).
75. Talotta, R. et al. Paradoxical Expansion of Th1 and Th17 Lymphocytes in Rheumatoid Arthritis Following Infliximab Treatment: a Possible Explanation for a Lack of Clinical Response. *J Clin Immunol* **35**, 550-7 (2015).
76. Kruglov, A.A., Lampropoulou, V., Fillatreau, S. & Nedospasov, S.A. Pathogenic and protective functions of TNF in neuroinflammation are defined by its expression in T lymphocytes and myeloid cells. *J Immunol* **187**, 5660-5670 (2011).
77. Kruglov, A. et al. Contrasting contributions of TNF from distinct cellular sources in arthritis. *Ann Rheum Dis* **79**, 1453-1459 (2020).
78. Wolf, Y. et al. Autonomous TNF is critical for in vivo monocyte survival in steady state and inflammation. *J Exp Med* **214**, 905-917 (2017).
79. Mukai, Y. et al. Solution of the structure of the TNF-TNFR2 complex. *Sci Signal* **3**, ra83 (2010).
80. Chan, F.K. The pre-ligand binding assembly domain: a potential target of inhibition of tumour necrosis factor receptor function *Ann Rheum Dis* **59 Suppl 1**, i50-53 (2000).
81. Croft, M. & Siegel, R.M. Beyond TNF: TNF superfamily cytokines as targets for the treatment of rheumatic diseases. *Nat Rev Rheumatol* **13**, 217-233 (2017).

- 1061 82. Van Hauwermeiren, F., Vandenbroucke, R.E. & Libert, C. Treatment of TNF mediated
1062 diseases by selective inhibition of soluble TNF or TNFR1. *Cytokine Growth Factor Rev*
1063 **22**, 311-9 (2011).
- 1064 83. Alexopoulou, L. et al. Transmembrane TNF protects mutant mice against intracellular
1065 bacterial infections, chronic inflammation and autoimmunity. *Eur J Immunol* **36**, 2768-
1066 2780 (2006).
- 1067 84. Nguyen, D.X. & Ehrenstein, M.R. Anti-TNF drives regulatory T cell expansion by
1068 paradoxically promoting membrane TNF-TNF-RII binding in rheumatoid arthritis. *J Exp*
1069 *Med* **213**, 1241-53 (2016).
- 1070 85. Grell, M. et al. The transmembrane form of tumor necrosis factor is the prime activating
1071 ligand of the 80 kDa tumor necrosis factor receptor *Cell* **83**, 793-802 (1995).
- 1072 86. Medler, J. & Wajant, H. Tumor necrosis factor receptor-2 (TNFR2): an overview of an
1073 emerging drug target. *Expert Opin Ther Targets* **23**, 295-307 (2019).
- 1074 87. So, T. & Croft, M. Regulation of PI-3-Kinase and Akt Signaling in T Lymphocytes and
1075 Other Cells by TNFR Family Molecules. *Front Immunol* **4**, 139 (2013).
- 1076 88. Wajant, H. & Beilhack, A. Targeting Regulatory T Cells by Addressing Tumor Necrosis
1077 Factor and Its Receptors in Allogeneic Hematopoietic Cell Transplantation and Cancer.
1078 *Front Immunol* **10**, 2040 (2019).
- 1079 89. Twu, Y.C., Gold, M.R. & Teh, H.S. TNFR1 delivers pro-survival signals that are
1080 required for limiting TNFR2-dependent activation-induced cell death (AICD) in CD8+ T
1081 cells. *Eur J Immunol* **41**, 335-44 (2011).
- 1082 90. Catrina, A.C. et al. Evidence that anti-tumor necrosis factor therapy with both etanercept
1083 and infliximab induces apoptosis in macrophages, but not lymphocytes, in rheumatoid
1084 arthritis joints: extended report. *Arthritis Rheum* **52**, 61-72 (2005).
- 1085 91. Mitoma, H. et al. Mechanisms for cytotoxic effects of anti-tumor necrosis factors agents
1086 on transmembrane tumor necrosis factor a-expressing cells. *Arthritis Rheum* **58**, 1248-
1087 1252 (2008).
- 1088 92. Tada, Y. et al. Collagen-induced arthritis in TNF receptor-1-deficient mice: TNF
1089 receptor-2 can modulate arthritis in the absence of TNF receptor-1. *Clinical Immunol* **99**,
1090 325-333 (2001).
- 1091 93. Lee, L.F. et al. The role of TNF-alpha in the pathogenesis of type 1 diabetes in the
1092 nonobese diabetic mouse: analysis of dendritic cell maturation. *Proc Natl Acad Sci U S A*
1093 **102**, 15995-6000 (2005).
- 1094 94. McDevitt, H., Munson, S., Ettinger, R. & Wu, A. Multiple roles for tumor necrosis
1095 factor-alpha and lymphotoxin alpha/beta in immunity and autoimmunity. *Arthritis Res* **4**
1096 **Suppl 3**, S141-52 (2002).
- 1097 95. Green, E.A. & Flavell, R.A. The temporal importance of TNFalpha expression in the
1098 development of diabetes. *Immunity* **12**, 459-69 (2000).
- 1099 96. Sakaguchi, S., Yamaguchi, T., Nomura, T. & Ono, M. Regulatory T cells and immune
1100 tolerance. *Cell* **133**, 775-87 (2008).
- 1101 97. Buckner, J.H. Mechanisms of impaired regulation by CD4(+)CD25(+)FOXP3(+) regulatory T cells in human autoimmune diseases. *Nat Rev Immunol* **10**, 849-59 (2010).
- 1102 98. Ehrenstein, M.R. et al. Compromised function of regulatory T cells in rheumatoid
1103 arthritis and reversal by anti-TNFalpha therapy. *J Exp Med* **200**, 277-85 (2004).
- 1104 99. Dige, A. et al. Adalimumab treatment in Crohn's disease does not induce early changes in
1105 regulatory T cells. *Scand J Gastroenterol* **46**, 1206-14 (2011).
- 1106 100. Li, Z. et al. Restoration of Foxp3+ Regulatory T-cell Subsets and Foxp3- Type 1
1107 Regulatory-like T Cells in Inflammatory Bowel Diseases During Anti-tumor Necrosis
1108 Factor Therapy. *Inflamm Bowel Dis* **21**, 2418-28 (2015).
- 1109

- 1110 101. Bluestone, J.A. & Tang, Q. Treg cells-the next frontier of cell therapy. *Science* **362**, 154-
1111 155 (2018).
- 1112 102. Zemmour, D. et al. Single-cell gene expression reveals a landscape of regulatory T cell
1113 phenotypes shaped by the TCR. *Nat Immunol* **19**, 291-301 (2018).
- 1114 103. Lubrano di Ricco, M. et al. Tumor necrosis factor receptor family costimulation increases
1115 regulatory T-cell activation and function via NF-kappaB. *Eur J Immunol* (2020).
- 1116 104. Vasanthakumar, A. et al. The TNF Receptor Superfamily-NF-kappaB Axis Is Critical to
1117 Maintain Effector Regulatory T Cells in Lymphoid and Non-lymphoid Tissues. *Cell Rep*
1118 **20**, 2906-2920 (2017).
- 1119 105. Chen, X. et al. Co-expression of TNFR2 and CD25 identifies more of the functional
1120 CD4(+)FoxP3(+) regulatory T cells in human peripheral blood. *Eur J Immunol* **40**, 1099-
1121 1106 (2010).
- 1122 106. Chen, X. et al. Cutting edge: expression of TNFR2 defines a maximally suppressive
1123 subset of mouse CD4+CD25+FoxP3+ T regulatory cells: applicability to tumor-
1124 infiltrating T regulatory cells. *J Immunol* **180**, 6467-71 (2008).
- 1125 107. Hamano, R., Huang, J., Yoshimura, T., Oppenheim, J.J. & Chen, X. TNF optimally
1126 activates regulatory T cells by inducing TNF receptor superfamily members TNFR2, 4-
1127 1BB and OX40. *Eur J Immunol* **41**, 2010-20 (2011).
- 1128 108. Grinberg-Bleyer, Y. et al. Pathogenic T cells have a paradoxical protective effect in
1129 murine autoimmune diabetes by boosting Tregs. *J Clin Invest* **120**, 4558-68 (2010).
- 1130 109. Baeyens, A. et al. Effector T Cells Boost Regulatory T Cell Expansion by IL-2, TNF,
1131 OX40, and Plasmacytoid Dendritic Cells Depending on the Immune Context. *J Immunol*
1132 **194**, 999-1010 (2015).
- 1133 110. Zhou, Q., Hu, Y., Howard, O.M., Oppenheim, J.J. & Chen, X. In vitro generated Th17
1134 cells support the expansion and phenotypic stability of CD4(+)Foxp3(+) regulatory T
1135 cells in vivo. *Cytokine* **65**, 56-64 (2014).
- 1136 111. Okubo, Y., Mera, T., Wang, L. & Faustman, D.L. Homogeneous expansion of human T-
1137 regulatory cells via tumor necrosis factor receptor 2. *Sci Rep* **3**, 3153 (2013).
- 1138 112. Urbano, P.C.M., Koenen, H., Joosten, I. & He, X. An Autocrine TNFalpha-Tumor
1139 Necrosis Factor Receptor 2 Loop Promotes Epigenetic Effects Inducing Human Treg
1140 Stability In Vitro. *Front Immunol* **9**, 573 (2018).
- 1141 113. Chen, X. et al. TNFR2 is critical for the stabilization of the CD4+Foxp3+ regulatory T.
1142 cell phenotype in the inflammatory environment. *J Immunol* **190**, 1076-84 (2013).
- 1143 114. Housley, W.J. et al. Natural but not inducible regulatory T cells require TNF-alpha
1144 signaling for in vivo function. *J Immunol* **186**, 6779-87 (2011).
- 1145 115. Santinon, F. et al. Involvement of Tumor Necrosis Factor Receptor Type II in FoxP3
1146 Stability and as a Marker of Treg Cells Specifically Expanded by Anti-Tumor Necrosis
1147 Factor Treatments in Rheumatoid Arthritis. *Arthritis Rheumatol* **72**, 576-587 (2020).
- 1148 116. Ban, L. et al. Strategic internal covalent cross-linking of TNF produces a stable TNF
1149 trimer with improved TNFR2 signaling. *Mol Cell Ther* **3**, 7 (2015).
- 1150 117. Chopra, M. et al. Exogenous TNFR2 activation protects from acute GvHD via host T reg
1151 cell expansion. *J Exp Med* **213**, 1881-900 (2016).
- 1152 118. He, X. et al. A TNFR2-Agonist Facilitates High Purity Expansion of Human Low Purity
1153 Treg Cells. *PLoS One* **11**, e0156311 (2016).
- 1154 119. Fischer, R. et al. Selective Activation of Tumor Necrosis Factor Receptor II Induces
1155 Antiinflammatory Responses and Alleviates Experimental Arthritis. *Arthritis Rheumatol*
1156 **70**, 722-735 (2018).
- 1157 120. Joedicke, J.J. et al. Activated CD8+ T cells induce expansion of Vbeta5+ regulatory T
1158 cells via TNFR2 signaling. *J Immunol* **193**, 2952-60 (2014).

- 1159 121. Lamontain, V. et al. Stimulation of TNF receptor type 2 expands regulatory T cells and
1160 ameliorates established collagen-induced arthritis in mice. *Cell Mol Immunol* **16**, 65-74
1161 (2019).
- 1162 122. Nagar, M. et al. TNF activates a NF-kappaB-regulated cellular program in human
1163 CD45RA- regulatory T cells that modulates their suppressive function. *J Immunol* **184**,
1164 3570-81 (2010).
- 1165 123. Wang, J. et al. TNFR2 ligation in human T regulatory cells enhances IL2-induced cell
1166 proliferation through the non-canonical NF-kappaB pathway. *Sci Rep* **8**, 12079 (2018).
- 1167 124. Bittner, S. & Ehrenschrwender, M. Multifaceted death receptor 3 signaling-promoting
1168 survival and triggering death. *FEBS Lett* **591**, 2543-2555 (2017).
- 1169 125. He, T. et al. The p38 MAPK Inhibitor SB203580 Abrogates Tumor Necrosis Factor-
1170 Induced Proliferative Expansion of Mouse CD4(+)Foxp3(+) Regulatory T Cells. *Front*
1171 *Immunol* **9**, 1556 (2018).
- 1172 126. Nie, H. et al. Phosphorylation of FOXP3 controls regulatory T cell function and is
1173 inhibited by TNF-alpha in rheumatoid arthritis. *Nat Med* **19**, 322-8 (2013).
- 1174 127. Stoop, J.N. et al. Tumor necrosis factor alpha inhibits the suppressive effect of regulatory
1175 T cells on the hepatitis B virus-specific immune response. *Hepatology* **46**, 699-705
1176 (2007).
- 1177 128. Valencia, X. et al. TNF downmodulates the function of human CD4+CD25hi T-
1178 regulatory cells. *Blood* **108**, 253-61 (2006).
- 1179 129. Zanin-Zhorov, A. et al. Protein kinase C-theta mediates negative feedback on regulatory
1180 T cell function. *Science* **328**, 372-6 (2010).
- 1181 130. Pierini, A. et al. TNF-alpha priming enhances CD4+FoxP3+ regulatory T-cell
1182 suppressive function in murine GVHD prevention and treatment. *Blood* **128**, 866-71
1183 (2016).
- 1184 131. Leclerc, M. et al. Control of GVHD by regulatory T cells depends on TNF produced by T
1185 cells and TNFR2 expressed by regulatory T cells. *Blood* **128**, 1651-9 (2016).
- 1186 132. Ronin, E. et al. Tissue-restricted control of established central nervous system
1187 autoimmunity by TNF receptor 2-expressing
1188 Treg cells. *Proc Natl Acad Sci U S A* **118**, e2014043118 (2021).
- 1189 133. Zaragoza, B. et al. Suppressive activity of human regulatory T cells is maintained in the
1190 presence of TNF. *Nat Med* **22**, 16-7 (2016).
- 1191 134. Miyara, M. et al. Functional delineation and differentiation dynamics of human CD4+ T
1192 cells expressing the FoxP3 transcription factor. *Immunity* **30**, 899-911 (2009).
- 1193 135. Chen, X. et al. Expression of costimulatory TNFR2 induces resistance of CD4+FoxP3-
1194 conventional T cells to suppression by CD4+FoxP3+ regulatory T cells. *J Immunol* **185**,
1195 174-82 (2010).
- 1196 136. Yamaguchi, T., Wing, J.B. & Sakaguchi, S. Two modes of immune suppression by
1197 Foxp3(+) regulatory T cells under inflammatory or non-inflammatory conditions. *Semin*
1198 *Immunol* **23**, 424-30 (2011).
- 1199 137. Chaudhry, A. & Rudensky, A.Y. Control of inflammation by integration of
1200 environmental cues by regulatory T cells. *J Clin Invest* **123**, 939-44 (2013).
- 1201 138. Williams, L.M. & Rudensky, A.Y. Maintenance of the Foxp3-dependent developmental
1202 program in mature regulatory T cells requires continued expression of Foxp3. *Nat*
1203 *Immunol* **8**, 277-84 (2007).
- 1204 139. Chinen, T. et al. An essential role for the IL-2 receptor in T reg cell function *Nat*
1205 *Immunol* **17**, 1322-1333 (2016).
- 1206 140. Feng, Y. et al. Control of the inheritance of regulatory T cell identity by a cis element in
1207 the Foxp3 locus *Cell* **158**, 749-763 (2014).

- 1208 141. Gao, Y. et al. Inflammation negatively regulates FOXP3 and regulatory T-cell function
1209 via DBC1. *Proc Natl Acad Sci U S A* **112**, E3246-54 (2015).
- 1210 142. Xie, M. et al. NF-kappaB-driven miR-34a impairs Treg/Th17 balance via targeting
1211 Foxp3. *J Autoimmun* **102**, 96-113 (2019).
- 1212 143. Molinero, L.L., Miller, M.L., Evaristo, C. & Alegre, M.L. High TCR stimuli prevent
1213 induced regulatory T cell differentiation in a NF-kappaB-dependent manner. *J Immunol*
1214 **186**, 4609-17 (2011).
- 1215 144. Zhang, Q. et al. TNF-alpha impairs differentiation and function of TGF-beta-induced
1216 Treg cells in autoimmune diseases through Akt and Smad3 signaling pathway. *J Mol Cell*
1217 *Biol* **5**, 85-98 (2013).
- 1218 145. Mahmud, S.A. et al. Costimulation via the tumor-necrosis factor receptor superfamily
1219 couples TCR signal strength to the thymic differentiation of regulatory T cells. *Nat*
1220 *Immunol* **15**, 473-81 (2014).
- 1221 146. Nadkarni, S., Mauri, C. & Ehrenstein, M.R. Anti-TNF-alpha therapy induces a distinct
1222 regulatory T cell population in patients with rheumatoid arthritis via TGF-beta. *J Exp*
1223 *Med* **204**, 33-9 (2007).
- 1224 147. So, T. & Croft, M. Cutting edge: OX40 inhibits TGF-beta- and antigen-driven conversion
1225 of naive CD4 T cells into CD25+Foxp3+ T cells. *J Immunol* **179**, 1427-30 (2007).
- 1226 148. Madireddi, S. et al. SA-4-IBBL costimulation inhibits conversion of conventional CD4+
1227 T cells into CD4+ FoxP3+ T regulatory cells by production of IFN- γ *PLoS One* **7**,
1228 e42459 (2012).
- 1229 149. Khan, S.Q. et al. Cloning, expression, and functional characterization of TL1A-Ig *J*
1230 *Immunol* **190**, 1540-1550 (2013).
- 1231 150. Lina, C., Conghua, W., Nan, L. & Ping, Z. Combined treatment of etanercept and MTX
1232 reverses Th1/Th2, Th17/Treg imbalance in patients with rheumatoid arthritis. *J Clin*
1233 *Immunol* **31**, 596-605 (2011).
- 1234 151. Toubi, E. et al. Increased spontaneous apoptosis of CD4+CD25+ T cells in patients with
1235 active rheumatoid arthritis is reduced by infliximab *Ann N Y Acad Sci* **1051**, 506-514
1236 (2005).
- 1237 152. Cao, D., van Vollenhoven, R., Klareskog, L., Trollmo, C. & Malmstrom, V.
1238 CD25brightCD4+ regulatory T cells are enriched in inflamed joints of patients with
1239 chronic rheumatic disease. *Arthritis Res Ther* **6**, R335-46 (2004).
- 1240 153. Dombrecht, E.J. et al. Influence of anti-tumor necrosis factor therapy (Adalimumab) on
1241 regulatory T cells and dendritic cells in rheumatoid arthritis. *Clin Exp Rheumatol* **24**, 31-
1242 7 (2006).
- 1243 154. McGovern, J.L. et al. Th17 cells are restrained by Treg cells via the inhibition of
1244 interleukin-6 in patients with rheumatoid arthritis responding to anti-tumor necrosis
1245 factor antibody therapy. *Arthritis Rheum* **64**, 3129-38 (2012).
- 1246 155. van Amelsfort, J.M., Jacobs, K.M., Bijlsma, J.W., Lafeber, F.P. & Taams, L.S.
1247 CD4(+)CD25(+) regulatory T cells in rheumatoid arthritis: differences in the presence,
1248 phenotype, and function between peripheral blood and synovial fluid. *Arthritis Rheum*
1249 **50**, 2775-85 (2004).
- 1250 156. Cao, D. et al. Isolation and functional characterization of regulatory CD25brightCD4+ T
1251 cells from the target organ of patients with rheumatoid arthritis. *Eur J Immunol* **33**, 215-
1252 23 (2003).
- 1253 157. Herrath, J. et al. The inflammatory milieu in the rheumatic joint reduces regulatory T-cell
1254 function. *Eur J Immunol* **41**, 2279-90 (2011).
- 1255 158. Komatsu, N. et al. Pathogenic conversion of Foxp3+ T cells into TH17 cells in
1256 autoimmune arthritis. *Nat Med* **20**, 62-8 (2014).

- 1257 159. Chen, Z. et al. The ubiquitin ligase Stub1 negatively modulates regulatory T cell
1258 suppressive activity by promoting degradation of the transcription factor Foxp3.
1259 *Immunity* **39**, 272-85 (2013).
- 1260 160. van Loosdregt, J. et al. Stabilization of the transcription factor Foxp3 by the
1261 deubiquitinase USP7 increases Treg-cell-suppressive capacity. *Immunity* **39**, 259-71
1262 (2013).
- 1263 161. Huang, Z. et al. Anti-TNF-alpha therapy improves Treg and suppresses Teff in patients
1264 with rheumatoid arthritis. *Cell Immunol* **279**, 25-9 (2012).
- 1265 162. Blache, C. et al. Number and phenotype of rheumatoid arthritis patients' CD4+CD25hi
1266 regulatory T cells are not affected by adalimumab or etanercept. *Rheumatology (Oxford)*
1267 **50**, 1814-22 (2011).
- 1268 163. Hvas, C.L. et al. Discrete changes in circulating regulatory T cells during infliximab
1269 treatment of Crohn's disease. *Autoimmunity* **43**, 325-33 (2010).
- 1270 164. Boschetti, G. et al. Therapy with anti-TNFalpha antibody enhances number and function
1271 of Foxp3(+) regulatory T cells in inflammatory bowel diseases. *Inflamm Bowel Dis* **17**,
1272 160-70 (2011).
- 1273 165. Kato, K. et al. Infliximab therapy impacts the peripheral immune system of
1274 immunomodulator and corticosteroid naive patients with Crohn's disease. *Gut Liver* **5**,
1275 37-45 (2011).
- 1276 166. Li, Z. et al. Reciprocal changes of Foxp3 expression in blood and intestinal mucosa in
1277 IBD patients responding to infliximab. *Inflamm Bowel Dis* **16**, 1299-310 (2010).
- 1278 167. Ricciardelli, I., Lindley, K.J., Londei, M. & Quarantino, S. Anti tumour necrosis-alpha
1279 therapy increases the number of FOXP3 regulatory T cells in children affected by Crohn's
1280 disease. *Immunology* **125**, 178-83 (2008).
- 1281 168. Veltkamp, C. et al. Apoptosis of regulatory T lymphocytes is increased in chronic
1282 inflammatory bowel disease and reversed by anti-TNFalpha treatment. *Gut* **60**, 1345-53
1283 (2011).
- 1284 169. Calleja, S. et al. Adalimumab specifically induces CD3(+) CD4(+) CD25(high) Foxp3(+)
1285 CD127(-) T-regulatory cells and decreases vascular endothelial growth factor plasma
1286 levels in refractory immuno-mediated uveitis: a non-randomized pilot intervention study.
1287 *Eye (Lond)* **26**, 468-77 (2012).
- 1288 170. Sugita, S., Yamada, Y., Kaneko, S., Horie, S. & Mochizuki, M. Induction of regulatory T
1289 cells by infliximab in Behcet's disease. *Invest Ophthalmol Vis Sci* **52**, 476-84 (2011).
- 1290 171. Xueyi, L. et al. Levels of circulating Th17 cells and regulatory T cells in ankylosing
1291 spondylitis patients with an inadequate response to anti-TNF-alpha therapy. *J Clin*
1292 *Immunol* **33**, 151-61 (2013).
- 1293 172. Wehrens, E.J. et al. Anti-tumor necrosis factor alpha targets protein kinase B/c-Akt-
1294 induced resistance of effector cells to suppression in juvenile idiopathic arthritis. *Arthritis*
1295 *Rheum* **65**, 3279-3284 (2013).
- 1296 173. Verwoerd, A. et al. Infliximab therapy balances regulatory T cells, tumour necrosis factor
1297 receptor 2 (TNFR2) expression and soluble TNFR2 in sarcoidosis. *Clin Exp Immunol*
1298 **185**, 263-70 (2016).
- 1299 174. Nguyen, D.X. et al. Regulatory T cells as a biomarker for response to adalimumab in
1300 rheumatoid arthritis. *J Allergy Clin Immunol* **142**, 978-980 e9 (2018).
- 1301 175. Chen, X., Oppenheim, J.J., Winkler-Pickett, R.T., Ortaldo, J.R. & Howard, O.M.
1302 Glucocorticoid amplifies IL-2-dependent expansion of functional
1303 FoxP3(+)CD4(+)CD25(+) T regulatory cells in vivo and enhances their capacity to
1304 suppress EAE. *Eur J Immunol* **36**, 2139-49 (2006).
- 1305 176. Kim, D. et al. Anti-inflammatory Roles of Glucocorticoids Are Mediated by Foxp3 +
1306 Regulatory T Cells via a miR-342-Dependent Mechanism. *Immunity* **53**, 581-596 (2020).

- 1307 177. Byng-Maddick, R. & Ehrenstein, M.R. The impact of biological therapy on regulatory T
1308 cells in rheumatoid arthritis. *Rheumatology (Oxford)* **54**, 768-75 (2015).
- 1309 178. Di Sabatino, A. et al. Peripheral regulatory T cells and serum transforming growth factor-
1310 beta: relationship with clinical response to infliximab in Crohn's disease. *Inflamm Bowel*
1311 *Dis* **16**, 1891-7 (2010).
- 1312 179. Zou, H., Li, R., Hu, H., Hu, Y. & Chen, X. Modulation of Regulatory T Cell Activity by
1313 TNF Receptor Type II-Targeting Pharmacological Agents. *Front Immunol* **9**, 594 (2018).
- 1314 180. Fischer, R., Kontermann, R.E. & Pfizenmaier, K. Selective Targeting of TNF Receptors
1315 as a Novel Therapeutic Approach. *Front Cell Dev Biol* **8**, 401 (2020).
- 1316 181. Mukai, Y. et al. Structure-function relationship of tumor necrosis factor (TNF) and its
1317 receptor interaction based on 3D structural analysis of a fully active TNFR1-selective
1318 TNF mutant. *J Mol Biol* **385**, 1221-9 (2009).
- 1319 182. Ando, D. et al. Creation of mouse TNFR2-selective agonistic TNF mutants using a phage
1320 display technique. *Biochem Biophys Res* **7**, 309-315 (2016).
- 1321 183. Van Ostade, X., Vandenabeele, P., Tavernier, J. & Fiers, W. Human tumor necrosis
1322 factor mutants with preferential binding to and activity on either the R55 or R75 receptor.
1323 *Eur J Biochem* **220**, 771-9 (1994).
- 1324 184. McCann, F.E. et al. Selective tumor necrosis factor receptor I blockade is
1325 antiinflammatory and reveals immunoregulatory role of tumor necrosis factor receptor II
1326 in collagen-induced arthritis. *Arthritis Rheumatol* **66**, 2728-38 (2014).
- 1327 185. Williams, S.K. et al. Anti-TNFR1 targeting in humanized mice ameliorates disease in a
1328 model of multiple sclerosis. *Sci Rep* **8**, 13628 (2018).
- 1329 186. Steeland, S. et al. TNFR1 inhibition with a Nanobody protects against EAE development
1330 in mice. *Sci Rep* **7**, 13646 (2017).
- 1331 187. Williams, S.K. et al. Antibody-mediated inhibition of TNFR1 attenuates disease in a
1332 mouse model of multiple sclerosis. *PLoS One* **9**, e90117 (2014).
- 1333 188. Zalevsky, J. et al. Dominant-negative inhibitors of soluble TNF attenuate experimental
1334 arthritis without suppressing innate immunity to infection. *J Immunol* **179**, 1872-83
1335 (2007).
- 1336 189. Shibata, H. et al. The treatment of established murine collagen-induced arthritis with a
1337 TNFR1-selective antagonistic mutant TNF. *Biomaterials* **30**, 6638-47 (2009).
- 1338 190. Brambilla, R. et al. Inhibition of soluble tumour necrosis factor is therapeutic in
1339 experimental autoimmune encephalomyelitis and promotes axon preservation and
1340 remyelination. *Brain* **134**, 2736-54 (2011).
- 1341 191. Nomura, T. et al. Therapeutic effect of PEGylated TNFR1-selective antagonistic mutant
1342 TNF in experimental autoimmune encephalomyelitis mice. *J Control Release* **149**, 8-14
1343 (2011).
- 1344 192. Taoufik, E. et al. Transmembrane tumour necrosis factor is neuroprotective and regulates
1345 experimental autoimmune encephalomyelitis via neuronal nuclear factor-kappaB *Brain*
1346 **134**, 2722-2735 (2011).
- 1347 193. Maier, O., Fischer, R., Agresti, C. & Pfizenmaier, K. TNF receptor 2 protects
1348 oligodendrocyte progenitor cells against oxidative stress. *Biochem Biophys Res Commun*
1349 **440**, 336-41 (2013).
- 1350 194. Rauert, H. et al. Membrane tumor necrosis factor (TNF) induces p100 processing via
1351 TNF receptor-2 (TNFR2). *J Biol Chem* **285**, 7394-7404 (2009).
- 1352 195. Dong, Y. et al. Essential protective role of tumor necrosis factor receptor 2 in
1353 neurodegeneration. *Proc Natl Acad Sci U S A* **113**, 12304-12309 (2016).
- 1354 196. Fischer, R. et al. TNFR2 promotes Treg-mediated recovery from neuropathic pain across
1355 sexes. *Proc Natl Acad Sci U S A* **116**, 17045-17050 (2019).

- 1356 197. Klatzmann, D. & Abbas, A.K. The promise of low-dose interleukin-2 therapy for
1357 autoimmune and inflammatory diseases. *Nat Rev Immunol* **15**, 283-94 (2015).
- 1358 198. Grinberg-Bleyer, Y. et al. IL-2 reverses established type 1 diabetes in NOD mice by a
1359 local effect on pancreatic regulatory T cells. *J Exp Med* **207**, 1871-8 (2010).
- 1360 199. Tang, Q. et al. Central role of defective interleukin-2 production in the triggering of islet
1361 autoimmune destruction. *Immunity* **28**, 687-97 (2008).
- 1362 200. Mori, L., Iselin, S., De Libero, G. & Lesslauer, W. Attenuation of collagen-induced
1363 arthritis in 55-kDa TNF receptor type 1 (TNFR1)-IgG1-treated and TNFR1-deficient
1364 mice. *J Immunol* **157**, 3178-82 (1996).
- 1365 201. Tseng, W.Y. et al. TNF receptor 2 signaling prevents DNA methylation at the Foxp3
1366 promoter and prevents pathogenic conversion of regulatory T cells. *Proc Natl Acad Sci U*
1367 *SA* **116**, 21666-21672 (2019).
- 1368 202. Bluml, S. et al. Antiinflammatory effects of tumor necrosis factor on hematopoietic cells
1369 in a murine model of erosive arthritis. *Arthritis Rheum* **62**, 1608-19 (2010).
- 1370 203. Eugster, H.P. et al. Severity of symptoms and demyelination in MOG-induced EAE
1371 depends on TNFR1. *Eur J Immunol* **29**, 626-32 (1999).
- 1372 204. Suvannavejh, G.C. et al. Divergent roles for p55 and p75 tumor necrosis factor receptors
1373 in the pathogenesis of MOG(35-55)-induced experimental autoimmune
1374 encephalomyelitis. *Cell Immunol* **205**, 24-33 (2000).
- 1375 205. Kassiotis, G. & Kollias, G. Uncoupling the proinflammatory from the
1376 immunosuppressive properties of tumor necrosis factor (TNF) at the p55 TNF receptor
1377 level: implications for pathogenesis and therapy of autoimmune demyelination. *J Exp*
1378 *Med* **193**, 427-34 (2001).
- 1379 206. Miller, P.G., Bonn, M.B., Franklin, C.L., Ericsson, A.C. & McKarns, S.C. TNFR2
1380 Deficiency Acts in Concert with Gut Microbiota To Precipitate Spontaneous Sex-Biased
1381 Central Nervous System Demyelinating Autoimmune Disease. *J Immunol* **195**, 4668-84
1382 (2015).
- 1383 207. Wang, Y.L. et al. Targeting pre-ligand assembly domain of TNFR1 ameliorates
1384 autoimmune diseases - an unrevealed role in downregulation of Th17 cells. *J Autoimmun*
1385 **37**, 160-70 (2011).
- 1386 208. Fischer, R. et al. Exogenous activation of tumor necrosis factor receptor 2 promotes
1387 recovery from sensory and motor disease in a model of multiple sclerosis. *Brain Behav*
1388 *Immun* **81**, 247-259 (2019).
- 1389
1390