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## **Vaptans or voluntary increased hydration to protect the kidney: how do they compare?**

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## **Abstract**

The adverse effects of vasopressin (AVP) in diverse forms of chronic kidney disease have been well described. They depend on the antidiuretic action of AVP mediated by V2 receptors (V2R). Treatment with tolvaptan, a selective V2R antagonist, is now largely used for the treatment of patients with ADPKD. Another way to reduce the adverse effects of AVP is to reduce endogenous AVP secretion by voluntary increase in fluid intake. These two approaches differ in several ways, including the level of thirst and AVP. With voluntary increased drinking plasma osmolality will decline and so will AVP secretion. Thus, not only will V2R-mediated effects be reduced, but also those mediated by V1a (V1aR) and V1b receptors. In contrast, selective V2R antagonism will induce a loss of fluid that will stimulate AVP secretion and thus, increase AVP's influence on V1a and V1b receptors. V1aR are expressed in the luminal side of the collecting duct and in inner medullary interstitial cells, and their activation induces the production of prostaglandins, mostly PGE2. Intrarenal PGE2 have been shown to reduce sodium and water reabsorption in the collecting duct and to increase blood flow in the renal medulla, both effects contributing to increase sodium and water excretion and reduce urine concentrating activity. Conversely, non-steroidal anti-inflammatory drugs have been shown to induce a significant water and sodium retention and potentiate the antidiuretic effects of AVP. Thus, during V2R antagonism, V1aR-mediated actions may be responsible for part of the diuresis observed with this drug. These V1aR-dependent effects do not take place with voluntary increase in fluid intake. In summary, while both strategies may have beneficial effects, the information reviewed here lead us to assume that the pharmacological V2R antagonism, with resulting stimulation of V1aR and increased PGE2 production, may provide greater benefit than voluntary HWI. The influence of tolvaptan on PGE2 excretion rate and the possibility to use somewhat lower tolvaptan doses than presently prescribed remain to be evaluated.

## **Keywords**

Thirst, Prostaglandin, Polycystic kidney disease, Sodium excretion,  
Medullary interstitial cells, Chronic kidney disease

## Bullet points

### What is already known about this subject

- Vasopressin, by activation of its V2 receptors, promotes kidney disease progression in a variety of nephropathies, beyond its direct involvement in autosomal dominant polycystic kidney disease.
- This influence on CKD in general is mediated, indirectly, by modifications of the composition of the tubular fluid at the macula densa and resulting changes in the tubulo-glomerular feedback control of GFR.
- It is thus important to reduce V2 receptor-mediated actions in ADPKD, as well as in all forms of CKD; this can be achieved either by a treatment with a selective V2 receptor antagonist, or by a voluntary increase in fluid intake.

### What this study adds

- Although rarely discussed, V2 receptor blockade or voluntary increase in fluid intake differ in their consequences on endogenous vasopressin secretion: vaptans increase vasopressin secretion while high water intake reduces it.
- As a result, an increased stimulation of V1a receptors occurs with vaptans and is absent with voluntary increase in fluid intake. V1aR stimulation is known to increase PGE2 production in the collecting duct and in interstitial cells of the inner medulla.
- Because these actions reduce urine concentrating activity (by mechanisms that have been well described several decades ago), they probably contribute to amplify the aquaresis induced by V2 receptor antagonism.

### What impact this may have on practice or policy

- We suggest measurements of urinary PGE2 excretion in patients submitted to each of these two treatments.
- If indeed, this excretion is significantly greater with vaptans than with voluntary increase in water intake, it will support the hypothesis that vaptans could provide a greater benefit than voluntary increase in fluid intake by reducing V2 effects both directly and indirectly.
- Moreover, targeting a urine osmolality close to that of plasma, rather than significantly below it, which could improve tolerability without much disadvantage, deserves to be evaluated.

## **Abbreviations**

AVP	Vasopressin or antidiuretic hormone
dDAVP	Desmopressin = 1-deamino 8D-arginine vasopressin
V2R, V1aR, V1bR	AVP V2, V1a, and V1b receptors, respectively
CKD	Chronic kidney disease
ADPKD	Autosomal dominant polycystic kidney disease
HWI	Voluntary increase in water intake
CD	Collecting duct
CCD	Cortical collecting duct
IMCD	Inner medullary collecting duct
TAL	Thick ascending limb
RMIC	Renal medullary interstitial cells
PGE2	Prostaglandin E2

## 1. Introduction

The adverse effects of vasopressin (AVP) and urine concentrating activity in chronic kidney disease (CKD) in general [1, 2], and more specifically in diabetic kidney disease [3] and autosomal dominant polycystic kidney disease (ADPKD) [4, 5] have been well described. Experimental studies *in vivo* and *in vitro* have provided some insight into the mechanisms of these adverse effects. They depend on the antidiuretic action of AVP, mediated by V2 receptors (V2R). The selective non-peptide orally active V2R antagonist tolvaptan is now largely used for the treatment of patients with rapidly progressing ADPKD [6] after it was proved to bring significant benefits in the Tempo 3:4 trial [7-10].

Glomerular hyperfiltration is known to be associated with further kidney dysfunction, adverse cardiovascular events and/or death [11-17]. Experimental studies and clinical investigations have shown that the sustained action of the selective V2R agonist dDAVP induces a chronic hyperfiltration and a rise in urinary albumin excretion [18, 19] while the suppression of AVP secretion by a water load results in a decrease in GFR [18, 20, 21]. Vasopressin contributes to CKD progression and to diabetic nephropathy by V2R-dependent actions [22, 23]. In ADPKD, the adverse effects of AVP depend on two independent and additive mechanisms: 1. the well understood influence of AVP on cyst growth mediated by its second messenger, cAMP; and 2. the hyperfiltration imposed on the kidney, as described above for all forms of CKD [24].

Thus, in order to reduce AVP-dependent V2R-mediated adverse consequences in renal patients (with ADPKD and with CKD in general), a voluntary high water intake (HWI) has been proposed, instead of a pharmacologic approach [4, 25-30]. Both V2R antagonism or voluntary HWI increase urine volume and decrease urine osmolality. However, they are fundamentally different. Drinking more will reduce AVP secretion and thirst. In contrast, V2R antagonism will induce a water loss and thus, will increase AVP secretion and thirst.

The aim of this review is to comparatively describe the direct and indirect consequences of these two approaches and explain how their differences may affect the outcomes.

## 2. AVP receptors

The three AVP receptors, the V1a, V1b, V2 receptors (V1aR, V1bR, V2R) are strikingly similar in both size and amino acid sequence. The V1aR and V1bR are selectively coupled to G-proteins of the Gq/11 family, leading to the breakdown of phosphoinositide lipids. The V2R preferentially activates the G-protein Gs, resulting in the activation of adenylyl cyclase.

The AVP receptors are widely distributed in different tissues in the body, as listed in **Table 1**. **V2R** are expressed in the kidney, the vascular endothelium where they play a role in coagulation, pneumocytes, and the inner ear [31]. They are also expressed in mouse, rat and human cholangiocytes and in polycystic liver epithelium [32]. **V1aR**, besides their well-known expression in vascular smooth muscle, are expressed in the kidney, in the liver where they may influence glucoregulation [33-36] and other organs [37, 38]. **V1bR** are present in the anterior hypophysis, pancreatic islets, adrenal glands [37, 39]. With so many different target sites, it is obvious that all tissues cannot respond simultaneously and indistinctly to AVP. Differences in the level of AVP required to stimulate each of them, and possible associated permissive or antagonizing factors may allow organ-selective responses. However, the V2R-dependent antidiuretic response is by far the most sensitive of all. In healthy volunteers undergoing water diuresis, a very low rate of AVP infusion induced a significant reduction in diuresis with no influence on other target organs or even on other intrarenal target sites [40].

Within the kidney (**Table 2**), V2Rs are located in the principal cells of the collecting duct (CD) and in the thick ascending limb (TAL). Less well-known, V1aRs are expressed in the luminal membrane of CD intercalated cells [41] and in interstitial cells of the inner medulla [42]. Binding studies with a V1aR antagonist and molecular studies

[43-45] suggest V1aR expression in thin descending limbs of short looped-nephrons, which also express the urea transporter UT-A2 (**Figure 1**).

### **3. Major differences between voluntary increase in water intake or treatment with a V2R antagonist**

**Table 3** shows the similarities and differences induced by either a voluntary HWI or a treatment with a V2R antagonist. The main differences reside in the level of thirst and AVP. With voluntary HWI, thirst is largely abolished, plasma osmolality declines, and so will AVP secretion. Thus, not only will V2R-mediated effects be reduced, but also those mediated by V1a and V1b receptors. In contrast, the V2R antagonism induces a loss of water that increases thirst, plasma osmolality and AVP secretion. Tolvaptan has been shown to increase plasma AVP and its surrogate marker copeptin, about 3 to 4-fold, [46-48]. Thus, effects mediated by V1aR and V1bR should be enhanced while V2R effects are, at least largely, abolished.

During the development of V2R antagonists, a serious concern was the risk of hypertension due to V1aR-mediated vasoconstriction. The use of a mixed V1aR/V2R antagonist was proposed [49-51]. However, princeps and long-term follow-up studies with tolvaptan did not reveal any significant rise in blood pressure [7, 9]. Actually, two recent studies revealed that blood pressure declined slightly in ADPKD patients after several months or years of tolvaptan treatment, compared to placebo [52, 53]. This effect was assumed to result from the beneficial effect on disease progression and on a higher fractional excretion of sodium and urea.

Another, more recent concern was a possible rise in glycemia via hepatic V1aR [33-35]. In epidemiologic studies, AVP is associated with obesity and metabolic syndrome [36, 54, 55]. However, princeps and long-term follow-up studies with tolvaptan disclosed no significant rise in glycemia. This confirms that the concentration of AVP required to induce a sustained vasoconstrictive or a metabolic response is significantly higher than that responsible for its antidiuretic action. Similarly, increased concentration of vasopressin in ADPKD patients at baseline and during tolvaptan

treatment did not result in activation of the hypothalamic-pituitary-adrenal axis. The impaired glucocorticoid production in these patients was found to be related to their degree of kidney function impairment [56].

In addition to the differences explained above, the two treatment approaches lead to quantitatively different urine volumes. Tolvaptan given for three weeks (90/30 mg in the last week) to 27 patients with ADPKD induced an average urine volume of 5,930 mL/d, vs 2,584 mL/d at baseline. Twenty four hour osmolar excretion was strongly associated with 24-hour urine volume [57]. In contrast, in patients with voluntary HWI, much lesser water intake and urine volumes were observed. In ADPKD patients with voluntary HWI for one year, urine volume rose only from  $2.048 \pm 648$  to  $2691 \pm 710$  mL/d [58]. In CKD patients coached for one year to increase their water intake by 1.0 to 1.5 L/d above their usual consumption, urine volume went up from 1.9 to 2.5 L/d, a change distinctly lower than intended in the design of the study [28].

The discussion about tolvaptan versus voluntary HWI is only valid for counteracting vasopressin's actions in kidney patients. The situation is different for patients with metabolic diseases (diabetes mellitus, metabolic syndrome) in which adverse V1aR- or V1bR-mediated effects are suspected [54, 55, 59]. In the absence of selective V1aR and V1bR antagonists, a voluntary increase in fluid intake is the only option.

#### **4. AVP and the renal handling of sodium : opposite roles of V2 and V1a receptors. Role of V1a receptors in chronic kidney disease**

AVP or dDAVP (selective V2R agonist) increase permeability of the collecting duct to water via their influence on aquaporin 2 (AQP2). They also increase sodium reabsorption by activating ENaC. dDAVP or relatively low doses of AVP reduce sodium excretion in both rats [60] and humans [61, 62]. This effect is dependent on ENaC since it is abolished by prior administration of amiloride [63]. This sodium retention was not observed in patients with nephrogenic diabetes insipidus with loss-of-function of the

V2R, but was intact in patients with AQP2 mutations, thus showing that it is mediated by V2R [61].

Paradoxically, many studies reported a strong natriuretic action of AVP when infused *in vivo* in experimental animals or in humans (see review in [62]). How can these observations be reconciled with the sodium-retaining effect of AVP or dDAVP via ENaC? Perucca et al performed dose-response studies of AVP effect on sodium excretion in conscious rats [60]. With increasing doses, AVP turned from being antinatriuretic to being natriuretic. The natriuretic effect was prevented by a selective V1aR antagonist (**Figure 2**). This demonstrates that V1aR activation induces a natriuretic effect that, with increases in AVP secretion, overcomes the antinatriuretic V2R-dependent action [60].

In rats with chronic kidney disease (CKD) induced by renal mass reduction, Perico et al reported that treatment with the dual AVP V1aR and V2R antagonist RWJ-676070, combined with angiotensin II blockade, lowered blood pressure, proteinuria, and glomerulosclerosis only marginally over angiotensin II blockade alone [64]. Surprisingly, no increase the 24-h urine volume was observed [64]. The simultaneous blockade of V1aR and V2R thus prevented the aquaretic effect expected in response to V2R antagonism. This suggests that, in patients treated with tolvaptan, the stimulation of V1aRs by the elevated AVP might participate to the observed aquaretic effect.

In patients with CKD, the fractional urinary excretion of AVP was shown to be the major determinant of the fractional excretion of sodium [65]. This strongly suggested a role for luminal AVP (thus most likely on V1aR) as an intrinsic diuretic/natriuretic factor [65], as confirmed by *in vitro* studies (see below).

## **5. V1aR-dependent prostaglandin production and actions within the kidney**

### **5. 1. Evidence for vasopressin-prostaglandin interaction in the kidney**

Prostaglandins are involved in the regulation of hormonal actions in various tissues. In the 1980s-2000s, many articles and reviews described the synthesis of prostaglandins by the kidney and their possible role in the regulation of kidney function [66-74]. But these mediators did not receive much attention in recent years.

Cyclooxygenases 1 and 2 (COX-1 and COX-2), the rate-limiting enzymes for prostaglandin synthesis from arachidonic acid, are expressed in several structures within the kidney that are targets of AVP: COX-2 in the thick ascending limb (TAL) and medullary interstitial cells (RMIC), and COX-1 in the collecting duct (CD) [75, 76]. Prostaglandins interact with G protein-coupled receptors (EP1 to EP4) [77]. EP3 is expressed in TAL and the outer medullary CD, EP1 in the inner medullary CD, and EP4 in descending vasa recta [75], sites that also express AVP receptors or may influence the antidiuretic action of AVP.

Urinary prostaglandin excretion is considered to reflect renal synthesis [78]. Inhibitors of prostaglandin synthesis have been used for elucidating their influence on kidney function. They include non-steroidal anti-inflammatory drugs (NSAID) indomethacin, meclofenamate, aspirin (acetyl salicylic acid), and more recently, selective inhibitors of either COX-1 or COX-2 [79] or selective antagonists of the different receptors [80, 81].

The natriuretic effect of AVP has been shown to depend on a V1aR-mediated influence on prostaglandin production [82]. PGE2 exerts at least two main actions in the kidney. It reduces sodium reabsorption in the collecting duct and increases blood flow in the renal medulla [71, 72, 83], both effects contributing to increase sodium excretion and reduce urine concentrating activity. Conversely, NSAID induce a significant water and sodium retention, and potentiate the antidiuretic effects of AVP [70, 84-88]. In rats, PGE2 infusion depressed the cortico-medullary gradient of NaCl, whereas indomethacin lead to a rise in medullary NaCl concentration [89, 90] without any change in renal hemodynamics. Prostaglandin synthesis may thus regulate the cortico-medullary osmotic gradient and increase natriuresis. Therefore, PGE2 produced in response to V1aR stimulation may account for the natriuresis induced by AVP.

Prostaglandins have been shown to interfere with the antidiuretic and antinatriuretic actions of AVP [91, 92]. As shown in **Figure 3**, inhibition of prostaglandin synthesis in water diuretic dogs, by indomethacin or meclofenamate, markedly potentiated the antidiuretic response to a bolus of vasopressin, without any change in GFR and solute excretion [92]. Similar results were observed in humans [93, 94]. Intrarenal prostaglandins inhibit the AVP-dependent generation of cyclic AMP *in vivo*, as previously demonstrated *in vitro* [92], an effect mediated by EP3 receptors [95]. Altogether, these observations establish the important role of prostaglandins in modulating the action of AVP.

The V1aR-dependent prostaglandin production in the kidney may explain the paradoxical observations reported in the following three studies. 1. In Brattleboro rats (lacking AVP) with 5/6th nephrectomy, AVP infusion (V2R + V1aR-mediated effects) induced much less severe signs of CKD progression than did dDAVP (V2R only). The deleterious effects of V2R activation were markedly blunted when associated with V1aR activation (**Table 4**) [96]. 2. In rats with hypertension induced by blockade of NO formation, AVP secretion is increased. Contrary to expectations, the selective V1aR antagonist worsened the rise in blood pressure and increased urinary albumin excretion [97] (**Figure 4A**). 3. The facilitated urea transporters UT-A2 and UT-A1 are expressed in the rat kidney. Infusion of dDAVP induced a marked increase in UT-A2 and UT-A1 expression, whereas almost no change was observed with an infusion of AVP [98] (**Figure 4B**). This suggests that the presence of V1aR agonism with AVP attenuated the influence of V2R agonism induced by dDAVP. Retrospectively, the effects observed in these three studies may be attributed to an influence of prostaglandins produced under the stimulation of intrarenal V1aR, that was not considered by the authors at that time.

V1aR are expressed in the intercalated cells of the CD and interstitial cells of the inner medulla (RMIC) [42]. In these two cell types, AVP stimulates the synthesis of PGE2 which is released locally. PGE2 then induces specific paracrine actions on CD principal cells (reduction of V2R-dependent osmotic water permeability and ENaC-mediated Na reabsorption), thick ascending limbs (inhibition of sodium reabsorption) and inner medullary vasculature (increase in medullary blood flow) [99]. These actions contribute to reduce the antidiuretic and antinatriuretic response mediated by V2R and to reduce

the urine concentrating ability. The simplified diagram of **Figure 5** shows the different actions of AVP, via its V2R and V1aR.

## **5.2. Collecting duct**

PGE2 attenuates the AVP-dependent effects on the CD in animals and humans [92, 93] and impairs AVP-dependent water transport in isolated perfused rabbit CCD [100] by inducing a faster degradation of cAMP. V1aRs are expressed in the lumen of the CD. Addition of AVP to the luminal perfusate of isolated perfused CDs inhibited the hydro-osmotic effect of AVP [101], and induced a sustained decrease in Na and Cl net fluxes [102]. This inhibitory action was prevented by meclofenamate, thus showing the role of PGE2 biosynthesis. A downregulation of V2R in the CD, via a V1aR-dependent pathway, represents an additional mechanism by which V1aR activation may reduce the antidiuretic action of AVP [103]. Taken together, the effects reported above explain how AVP, via its luminal V1aR in intercalated cells, significantly interferes with V2R-mediated antidiuretic action in the CD.

How high may be the concentration of AVP in the CD luminal fluid? AVP is far more concentrated in urine than in plasma. The urine/plasma ratio of AVP concentrations was 42 in a study of healthy humans [104]. When the urine concentrating ability is reduced and the rate of tubular fluid flow rate in the CD lumen is increased (by V2R antagonism or other causes), AVP concentration in the CD lumen may still be significantly higher than in plasma. Thus, luminal AVP may still induce a significant influence on V1aR in the CD.

## **5.3. Thick ascending limb**

V1aR and V1aR-mediated actions of AVP have not been reported in the TAL. However, TAL transport proteins and/or transport functions are regulated by prostaglandins. A paracrine influence on the TAL by PGE2 released by medullary CDs in the inner stripe interstitium can be favored by the close proximity of these two structures. PGE2 reduces cell cAMP content [105] and ion transport in MTAL cells by

inhibiting AVP-dependent cAMP formation [106, 107]. In isolated perfused TALs, PGE<sub>2</sub> reduced chloride transport [108] and transepithelial voltage [109], and increased HCO<sub>3</sub><sup>-</sup> reabsorption [110]. *In vivo*, endogenous PGE<sub>2</sub> reduced fractional chloride reabsorption [111]. In mouse medullary TAL cells in culture. PGE<sub>2</sub> inhibited the Na- and Cl-dependent, bumetanide-sensitive K influx, probably by downregulating the number of Na-K-2Cl cotransporters [112]. Conversely, cyclo-oxygenase inhibition increased the abundance of the Na-K-2Cl cotransporter. Thus, cyclooxygenase inhibitors enhance urinary concentrating ability in part by abolishing the inhibitory effect of PGE<sub>2</sub> on AVP-dependent cAMP production [113]. Altogether, these studies suggest that PGE<sub>2</sub> may act as a counter-regulatory factor to maintain a stable function in the MTAL during antidiuresis, when circulating AVP levels and medullary osmolality are elevated.

#### **5. 4. Inner medullary interstitial cells and medullary blood flow**

Renal medullary interstitial cells (RMICs) are very abundant in the inner medulla [114, 115]. These "lipid-laden" cells exhibit abundant lipid droplets of arachidonic acid, the precursor of prostanoids. They express COX-1, COX-2 and PGE synthase [116] and produce PGE<sub>2</sub> and PGF<sub>2</sub> alpha [117-120]. These cells also express abundantly V1aR [42]. PGE<sub>2</sub> production in RMIC is stimulated by AVP, an effect abolished by a selective antagonist of the V1aR [119, 120]. This AVP-stimulated PGE<sub>2</sub> production thus results from V1aR activation. The stimulation of V1aRs in RMIC may be very significant because AVP concentration is 20-30 times higher in medulla and papilla than in peripheral blood [121]. Urinary PGE<sub>2</sub> excretion rate is considered to reflect largely the intrarenal PGE<sub>2</sub> production by RMICs [78]. PGE<sub>2</sub> induce a significant vasodilation of the vasa recta by activation of EP2 and EP4 receptors, resulting in an increase in medullary blood [122-124].

An increased blood flow in the medullary vasa recta compromises the accumulation of solutes in the inner medulla and thus, decreases the intra-renal osmotic gradient, a crucial requirement for the urine concentrating mechanism. Urea is accumulated in the inner medulla by an AVP-dependent increase in urea permeability of the terminal inner medullary CD permitted by the activation of the facilitated urea transporter UT-A1 [125-127]. If urea accumulation in the medulla is compromised,

water reabsorption and thus urine concentration will be reduced. An increased medullary blood flow will wash out the medullary urea in ascending venous vasa recta, and thus reduce inner medullary urea concentration [99]. Moreover, PGE<sub>2</sub> may interfere with V<sub>2</sub>R-mediated influence on UT-A1 and thus, let more urea be excreted in the urine, thus reducing further its accumulation in the inner medulla and the urine concentrating capacity [128].

Intriguingly PGE<sub>2</sub> in the renal medulla exhibit a strong sex-related difference. PGE<sub>2</sub> excretion is about twice higher in female than in male rats, and ovariectomy lowers this production to the level observed in males [129]. High PGE<sub>2</sub> in females is responsible for a twofold higher medullary blood or plasma flow in female than in male rats [130, 131]. Indomethacin reduced these flows in females to values similar to those in control males [131].

Altogether, the facts described above provide evidence that AVP, through actions initiated by V<sub>1a</sub>Rs, attenuates the antidiuretic and antinatriuretic effects mediated by peritubular V<sub>2</sub>Rs and that these counter-regulatory actions are mediated by PGE<sub>2</sub> (**Figure 5**). The rise in AVP secretion induced by V<sub>2</sub>R antagonism thereby participates in the "aquaretic" effect observed during V<sub>2</sub>R antagonism. These V<sub>1a</sub>R-mediated effects in the kidney probably represent a safeguard against an excessive urine concentration that would compromise the excretory function of the kidney. Actually, a low urine flow rate markedly reduces the fractional excretion (FE) of urea [132] as well as that of other solutes, although to a lesser extent [18, 62]. In healthy humans undergoing a clinical investigation with renal clearance measurements, the FE during high and low hydration conditions (in the same subjects) were respectively 64 and 46 % for urea, 2.14 and 1.42 % for Na, and 19.3 and 15.7 % for K [21]. The FE of these solutes would most likely be even more significantly reduced without the counterbalancing influence of V<sub>1a</sub>R activation.

## **6. Possible role of AVP, via V<sub>1a</sub>R, in the high PGE<sub>2</sub> production and the high polyuria observed in monogenic tubulopathies**

In several mendelian diseases involving transporters or regulatory proteins expressed in the TAL, like antenatal Bartter syndrome, an intense polyuria is accompanied by a marked increase in PGE<sub>2</sub> urinary excretion. Inhibitors of prostaglandin synthesis significantly ameliorate the urine concentrating defect of these patients [133, 134]. Thus, in addition to the loss of function mutations in the Na-K-2Cl cotransporter [135], PGE<sub>2</sub> also contributes largely to this excessive urinary dilution [133, 134, 136].

The reason why prostaglandin synthesis is markedly increased in these diseases is not elucidated. AVP secretion is elevated in these patients, due to the loss of fluid induced by the high diuresis. Thus, the increased prostaglandin production may be explained by enhanced AVP-induced activation of renal V<sub>1a</sub>R in RMIC. This situation is similar to that observed in response to treatment with tolvaptan. Thus, the urine concentrating defect in these tubulopathies is likely due to two cumulative factors: 1, the genetic defect that impairs urine concentrating ability; 2, the influence of increased V<sub>1a</sub>R-stimulated prostaglandin synthesis that reduces the residual V<sub>2</sub>R-mediated antidiuretic action. In a mouse model of late-onset type I Bartter syndrome [137], urinary prostaglandin E<sub>2</sub> is increased, as in the human disease, and the urine concentrating defect is largely attributed to a defect in urea concentration in the urine, likely attributable to a high medullary blood flow [137].

An enhanced secretion of AVP and its resulting action on renal V<sub>1a</sub>R bring a novel pathophysiological explanation for the markedly elevated PGE<sub>2</sub> excretion rate and high diuresis observed in these monogenic diseases. V<sub>1a</sub>R antagonism could possibly represent an alternative to NSAID for reducing the urinary concentrating defect.

## **7. Complex effects of vasopressin on vascular smooth muscle cells and endothelial cells**

In arteries, V<sub>1a</sub>R are expressed on vascular smooth muscle cells [138-140] and V<sub>2</sub>R on endothelial cells [31, 141]. *Ex vivo* studies have shown that AVP can induce opposite vasomotor effects : vasoconstriction via V<sub>1a</sub>R [140, 142], and vasodilation via V<sub>2</sub>R [141]. *in vitro* studies of human HUVEC endothelial cells with heterologous V<sub>2</sub>R

expression, showed that dDAVP activates the endothelial NO-synthase eNOS via cAMP [143]. This explains the endothelial NO-dependent vasodilatation elicited by high concentrations of AVP.

Independent of its hemodynamic implications, desmopressin increases blood concentration of vWF, Factor VIII and plasminogen activator inhibitor-1 [144-146]. These effects are present in patients with bilateral nephrectomy, and abolished in patients with inactivating mutations of V2R (X-linked diabetes insipidus), which demonstrates the direct implication of extrarenal V2R [147].

In spite of the effects reported above, it seems unlikely that the pharmacological inhibition of V2R could lead to a clinically significant disturbance of vascular physiology, because patients with X-linked diabetes insipidus do not present with hematological or vascular disorders. In addition, both randomized controlled trials and follow-up studies of tolvaptan in ADPKD did not report a significant increase in blood pressure or cardiovascular events (a small decline in blood pressure is even observed on the long term (see above). Nevertheless, detailed studies of the effects of vaptans on the cardiovascular system, and comparison with increased hydration are lacking and are currently insufficient to suggest prioritizing either attitude in this setting. In particular, whether vaptans may improve or aggravate endothelial function may be particularly relevant in ADPKD patients, in which endothelial dysfunction is present as a consequence of the disrupted polycystin complex in arteries [148].

## **8. Which strategy should be most effective for reducing V2R-mediated adverse effects?**

In ADPKD, reducing vasopressin V2R-mediated effects is a major issue because of their direct influence on cyst growth induced by V2R-dependent formation of cAMP. More generally, in CKD, reducing V2R-mediated effects is also relevant because they participate in glomerular hyperfiltration and increased albuminuria [22, 24]. It is currently difficult to conclude which of the two strategies (selective V2R antagonist or recommendations to voluntarily increased water intake) would be more effective to

slow progression of all forms of CKD. Moreover, very few results are yet available from trials based on a voluntary HWI.

A few recent trials showed the feasibility of voluntary HWI. However, they concerned only small numbers of subjects and for short durations [26, 29, 30]. A one-year trial in Canadian patients with CKD showed no benefit of HWI for kidney protection in spite of monthly coaching [28]. In this and several other HWI trials, fluid intake was lesser than prescribed [28, 149, 150]. This is also the case in patients with recurrent urolithiasis, in spite of their painful symptoms [151, 152]. There are wide inter-individual differences in the tendency to concentrate urine and thus probably in usual beverage consumption among subjects [153]. Future "HWI" studies could select "low drinkers" with high baseline copeptin and  $U_{osm}$  [35].

V1aR-mediated actions of AVP were assumed for a long time to be potentially harmful, but no related adverse effects have been reported in the long-term follow-up studies of V2R antagonists. The rise in AVP secretion induced by tolvaptan is apparently not large enough to impact extra-renal tissues expressing V1aR. However, the influence of AVP on V1aR may be significant within the kidney because AVP gets concentrated in the luminal fluid of the CD by water reabsorption, and in the medullary vasa recta supplying the RMIC because of counter-current exchanges between descending and ascending vasa recta.

A recent review described how tolvaptan treatment should be proposed in the category of patients who can best benefit of this treatment [154]. Compared to HWI, limitations of tolvaptan include the risks of some adverse effects (liver toxicity, fatigue, hyperuricemia) and cost. A modest hyperuricemia was reported in 14.8% of the patients in the Japanese branch of the TEMPO 3/4 trial [155]. The intense polyuria and nocturia are responsible for some treatment discontinuation. As discussed recently, increasing the tolerability of this treatment is a major challenge [154]. Interestingly, the doses used up to now may possibly be reduced. Indeed, an important question that has rarely been addressed is how low should  $U_{osm}$  be reduced. In a small trial in ADPKD patients, tolvaptan given for three weeks (with 90/30 mg in the last week) reduced urine osmolality from 359 {IQ 289-425} mosm/kg H<sub>2</sub>O at baseline to 139 {IQ 126-173}

mosm/kg H<sub>2</sub>O on the third week [57]. In the TEMPO 3/4 trial, the mean  $U_{osm}$  observed over the course of the study was 220-230 mosm/kg H<sub>2</sub>O (figure 3A in [156]). Based on early dose-finding studies of tolvaptan, in which efficacy was defined as the capacity to achieve  $U_{osm} < 300$  mosm/kg H<sub>2</sub>O, Torres et al deduced that further lowering of  $U_{osm}$  below 300 may provide no significant additional benefit, while decreasing quality of life [6]. It is important to note that, in this range of low  $U_{osm}$ , the difference in daily urine volumes between  $U_{osm}$  of 220 or 300 mosm/kg is very large. For a given osmolar load of 900 mosm/d, it drops by more than 1 liter (from 4.1 to 3.0 L/d). Thus, treatments with lower doses than used in previous trials [7] could antagonize V2R-mediated actions while inducing less intense side effects (thirst, polyuria, nycturia [7]), allow better compliance (and could be less expensive).

Another reason why a lower dose of tolvaptan might be appropriate is because the adverse influence of vasopressin on kidney function (including the induction of glomerular hyperfiltration) appears to be biphasic. It is restricted to situations in which urine is still concentrated above plasma osmolality. It is not observed when urine is hypo-osmotic [21, 22, 157, 158]. A recent study in 1265 CKD patients suggests that either low or high water intake may not be beneficial in CKD: the association between the rate of eGFR decline and water consumption was U-shaped [159]. These observations thus suggest that the dose of tolvaptan might be adjusted to target an  $U_{osm}$  close to that of plasma, but not significantly below it. Given the benefits in terms of quality of life and adherence to the treatment, it might be worth testing in a clinical trial if a lower dose of tolvaptan would be as efficient as the doses prescribed presently.

During V2R antagonism, we assume that a significant part of the aquaresis may result from V1aR-mediated effects and resulting PGE2 production. Unfortunately, urinary PGE2 excretion has never been evaluated in patients treated with tolvaptan. A new clinical trial could evaluate the differences in urinary PGE2 excretion rate in healthy subjects, and/or in kidney patients treated with either tolvaptan or a placebo, or in patients recommended to increase their water intake. If a significantly greater PGE2 excretion rate is observed in the tolvaptan group, and the lowest excretion rate in the group with voluntary HWI, it will suggest that PGE2 do indeed participate in the aquaresis (as it does in monogenic diseases characterized by a high PGE2 production).

In summary, while both strategies may have beneficial effects, the information reviewed above lead us to assume that the pharmacological V2R antagonism, with resulting stimulation of V1aR and increased PGE2 production, may provide greater benefit than voluntary HWI. The influence of tolvaptan on PGE2 excretion rate and the possibility to use somewhat lower tolvaptan doses than presently prescribed remain to be evaluated.

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## Figure Legends

**Figure 1. A and B.** Autoradiographies showing binding of a radio-iodinated selective V1aR antagonist in adult rat kidney. Reproduced from [43]. **A.** The four kidney zones are indicated: C, OS, IS, and IM = cortex, outer stripe and inner stripe of the outer medulla, and inner medulla, respectively. V1aR labeling is observed in the cortex, the lower half of the IS, and the IM. Calibration bar = 1 mm. **B.** Higher magnification of a tangential section through the IS shows labeled thin descending limbs of short looped-nephrons (h) apposed at the periphery of unlabeled vascular bundles (vb). In the lower part of the figure, intense labeling is also observed in collecting ducts (cd) in the upper part of the IM. Calibration bar = 200  $\mu$ m. **C and D.** Expression of the urea transporter UT-A2 in the inner stripe. **C.** Epon section of a mouse kidney through the deep IS, labeled for UT-A2. The thin descending limbs of short loops (S), lying around a vascular bundle, are labeled for UT-A2, whereas those of long loops (L), in the interbundle regions, are not. This study concludes that UT-A2 expression is restricted to the last 28% to 44% of the descending thin limbs of all short looped-nephrons. Magnification, x 80. Reproduced from [45]. **D.** In situ hybridization of UT-A2 in a rat kidney. Transverse section through the deep IS showing UT-A2 positive short loops gathered around negative vascular bundles (courtesy of Matthias Hediger, Boston Mass, USA; 1997). Note the similar pattern of labeling of UT-A2 (in D) and of V1aR (in B) in tubules gathered around vascular bundles. This suggests a possible co-localization of V1aR and UT-A2 in the thin descending limbs of short loops [43-45].

**Figure 2.** Effects of AVP (15mg/kg BW) on sodium excretion rate without or with the co-administration of the V1aR antagonist SR49059 (10 mg/kg BW). The effect of the antagonist alone and of dDAVP (selective V2 agonist) are also shown. Results of the experimental day are expressed as percentage of values observed during the previous basal day (with sham injections) in the same rats. Paired t test, experimental versus basal day : \*\* P < 0.01; \*\*\* P < 0.001. AVP injection at 15 mg/kg induced a significant rise in Na excretion, but injections at 1 or 5 mg/kg did not (not shown here). The V1aR antagonist induced a modest significant rise in urine osmolality (not shown here) and abolished the rise in sodium excretion rate due to AVP. dDAVP induced a

significant decline in sodium excretion, most likely due to increased ENaC-dependent Na reabsorption. Modified from [60].

**Figure 3.** Evidence for in vivo influence of prostaglandins on the antidiuretic response to vasopressin. Anesthetized, water diuretic dogs received a bolus of 100 mU vasopressin, twice at about a 120 min interval. The second bolus was given after the administration of 2 mg/kg indomethacin. Reproduced from [92].

**Figure 4. A.** Systolic blood pressure (SBP) in rats during an eight-week treatment with the NO synthase inhibitor N<sup>G</sup>-nitro-L-arginine (L-NNA), the selective V1aR antagonist SR 49059, or both in combination. Data are means ± SEM. L-NNA induced a significant progressive rise in SBP; 2-way ANOVA: \*\*\* P<0.001. The V1aR antagonist alone had no effect on SBP but, contrary to expectations, it significantly worsened the rise induced by L-NNA (red double arrows). This suggests that the normal activation of V1aRs is actually protective. Significant interaction of L-NNA and SR 49059: # P<0.001. Adapted from [97]. **B.** Influence of a chronic infusion of AVP (acting on both V2Rs and V1aRs) or dDAVP (acting selectively on V2Rs) on mRNA expression of facilitated urea transporters in the four medullary zones of the Brattleboro rat kidney (superficial and deep inner stripe, and base and tip of inner medulla). Adapted from [98]. Exp = Experimental, after a 5 day infusion of AVP or dDAVP. \* : p < 0.05 for dDAVP vs basal condition. dDAVP induced a marked increase in UT-A2 expression in the deep IS, and of UT-A1 in the upper IM, whereas almost no change was observed with an infusion of AVP. This suggests that the activation of the V1aRs (during AVP infusion) attenuated the influence due to activation of the V2Rs.

**Figure 5.** Effects of AVP, via V1a and V2 receptors, on different cell types in the kidney and their consequences on the urine concentrating ability. Not shown are additional effects on the thick ascending limbs via V2Rs (direct) and V1aRs (indirect, mediated by paracrine effects of PGE2 issued from surrounding CCDs). CCD and IMCD = cortical and inner medullary CDs, respectively. PG-R = PGE2 receptor.

## References

1. Bankir L, Roussel R, Bouby N. Protein- and diabetes-induced glomerular hyperfiltration: role of glucagon, vasopressin, and urea. *Am J Physiol Renal Physiol* 2015;309(1):F2-23
2. Clark WF, Sontrop JM, Huang S-H, Moist L, Bouby N, Bankir L. Hydration and Chronic Kidney Disease Progression: A Critical Review of the Evidence. *American journal of nephrology* 2016;43(4):281-292
3. Roussel R, Velho G, Bankir L. Vasopressin and diabetic nephropathy. *Current opinion in nephrology and hypertension* 2017;26(4):311-318
4. Torres VE, Bankir L, Grantham JJ. A case for water in the treatment of polycystic kidney disease. *Clinical journal of the American Society of Nephrology : CJASN* 2009;4(6):1140-1150
5. van Gastel MDA, Torres VE. Polycystic Kidney Disease and the Vasopressin Pathway. *Annals of nutrition & metabolism* 2017;70 Suppl 1:43-50
6. Chebib FT, Perrone RD, Chapman AB, Dahl NK, Harris PC, Mrug M, Mustafa RA, Rastogi A, Watnick T, Yu ASL, Torres VE. A Practical Guide for Treatment of Rapidly Progressive ADPKD with Tolvaptan. *J Am Soc Nephrol* 2018;29(10):2458-2470
7. Torres VE, Chapman AB, Devuyst O, Gansevoort RT, Grantham JJ, Higashihara E, Perrone RD, Krasa HB, Ouyang J, Czerwiec FS, Investigators TT. Tolvaptan in patients with autosomal dominant polycystic kidney disease. *The New England journal of medicine* 2012;367(25):2407-2418
8. Torres VE, Higashihara E, Devuyst O, Chapman AB, Gansevoort RT, Grantham JJ, Perrone RD, Ouyang J, Blais JD, Czerwiec FS, Investigators TT. Effect of Tolvaptan in Autosomal Dominant Polycystic Kidney Disease by CKD Stage: Results from the TEMPO 3:4 Trial. *Clinical journal of the American Society of Nephrology : CJASN* 2016;11(5):803-811
9. Torres VE, Chapman AB, Devuyst O, Gansevoort RT, Perrone RD, Koch G, Ouyang J, McQuade RD, Blais JD, Czerwiec FS, Sergeyeva O. Tolvaptan in Later-Stage Autosomal Dominant Polycystic Kidney Disease. *N Engl J Med* 2017;377(20):1930-1942
10. Torres VE, Chapman AB, Devuyst O, Gansevoort RT, Perrone RD, Dandurand A, Ouyang J, Czerwiec FS, Blais JD, Investigators TT. Multicenter, open-label, extension trial to evaluate the long-term efficacy and safety of early versus delayed treatment with tolvaptan in autosomal dominant polycystic kidney disease: the TEMPO 4:4 Trial. *Nephrology, dialysis, transplantation : official publication of the European Dialysis and Transplant Association - European Renal Association* 2018;33(3):477-489
11. Brenner BM. Nephron adaptation to renal injury or ablation. *Am J Physiol* 1985;249(3 Pt 2):F324-337

12. Moriya T, Tsuchiya A, Okizaki S, Hayashi A, Tanaka K, Shichiri M. Glomerular hyperfiltration and increased glomerular filtration surface are associated with renal function decline in normo- and microalbuminuric type 2 diabetes. *Kidney Int* 2012;81(5):486-493
13. Ruggenti P, Porrini EL, Gaspari F, Motterlini N, Cannata A, Carrara F, Cella C, Ferrari S, Stucchi N, Parvanova A, Iliev I, Dodesini AR, Trevisan R, Bossi A, Zaletel J, Remuzzi G. Glomerular hyperfiltration and renal disease progression in type 2 diabetes. *Diabetes Care* 2012;35(10):2061-2068
14. Bjornstad P, Cherney DZ, Snell-Bergeon JK, Pyle L, Rewers M, Johnson RJ, Maahs DM. Rapid GFR decline is associated with renal hyperfiltration and impaired GFR in adults with Type 1 diabetes. *Nephrol Dial Transplant* 2015;30(10):1706-1711
15. Park M, Yoon E, Lim YH, Kim H, Choi J, Yoon HJ. Renal hyperfiltration as a novel marker of all-cause mortality. *J Am Soc Nephrol* 2015;26(6):1426-1433
16. Oh SW, Yang JH, Kim MG, Cho WY, Jo SK. Renal hyperfiltration as a risk factor for chronic kidney disease: A health checkup cohort study. *PLoS One* 2020;15(9):e0238177
17. Penno G, Orsi E, Solini A, Bonora E, Fondelli C, Trevisan R, Vedovato M, Cavalot F, Gruden G, Laviola L, Nicolucci A, Pugliese G. Renal hyperfiltration is independently associated with increased all-cause mortality in individuals with type 2 diabetes: a prospective cohort study. *BMJ Open Diabetes Res Care* 2020;8(1)
18. Bouby N, Ahloulay M, Nsegbe E, Dechaux M, Schmitt F, Bankir L. Vasopressin increases glomerular filtration rate in conscious rats through its antidiuretic action. *J Am Soc Nephrol* 1996;7(6):842-851
19. Bardoux P, Bichet DG, Martin H, Gallois Y, Marre M, Arthus MF, Lonergan M, Ruel N, Bouby N, Bankir L. Vasopressin increases urinary albumin excretion in rats and humans: involvement of V2 receptors and the renin-angiotensin system. *Nephrol Dial Transplant* 2003;18(3):497-506
20. Hadj-Aissa A, Bankir L, Fraysse M, Bichet DG, Laville M, Zech P, Pozet N. Influence of the level of hydration on the renal response to a protein meal. *Kidney Int* 1992;42(5):1207-1216
21. Anastasio P, Cirillo M, Spitali L, Frangiosa A, Pollastro RM, De Santo NG. Level of hydration and renal function in healthy humans. *Kidney Int* 2001;60(2):748-756
22. Bankir L, Bouby N, Ritz E. Vasopressin: a novel target for the prevention and retardation of kidney disease? *Nat Rev Nephrol* 2013;9(4):223-239
23. Clark WF, Sontrop JM, Huang SH, Moist L, Bouby N, Bankir L. Hydration and Chronic Kidney Disease Progression: A Critical Review of the Evidence. *Am J Nephrol* 2016;43(4):281-292
24. Bankir L, Bichet DG. What can copeptin tell us in patients with autosomal dominant polycystic disease? *Kidney Int* 2019;96(1):19-22

25. Negoianu D, Goldfarb S. Just add water. *J Am Soc Nephrol* 2008;19(6):1041-1043
26. Wang CJ, Creed C, Winklhofer FT, Grantham JJ. Water prescription in autosomal dominant polycystic kidney disease: a pilot study. *Clin J Am Soc Nephrol* 2011;6(1):192-197
27. Wang CJ, Grantham JJ, Wetmore JB. The medicinal use of water in renal disease. *Kidney international* 2013;84(1):45-53
28. Clark WF, Sontrop JM, Huang S-H, Gallo K, Moist L, House AA, Cuerden MS, Weir MA, Bagga A, Brimble S, Burke A, Muirhead N, Pandeya S, Garg AX. Effect of Coaching to Increase Water Intake on Kidney Function Decline in Adults With Chronic Kidney Disease: The CKD WIT Randomized Clinical Trial. *JAMA* 2018;319(18):1870-1879
29. Wong ATY, Mannix C, Grantham JJ, Allman-Farinelli M, Badve SV, Boudville N, Byth K, Chan J, Coulshed S, Edwards ME, Erickson BJ, Fernando M, Foster S, Haloob I, Harris DCH, Hawley CM, Hill J, Howard K, Howell M, Jiang SH, Johnson DW, Kline TL, Kumar K, Lee VW, Lonergan M, Mai J, McCloud P, Peduto A, Rangan A, Roger SD, Sud K, Torres V, Vilayur E, Rangan GK. Randomised controlled trial to determine the efficacy and safety of prescribed water intake to prevent kidney failure due to autosomal dominant polycystic kidney disease (PREVENT-ADPKD). *BMJ Open* 2018;8(1):e018794
30. Enhörning S, Brunkwall L, Tasevska I, Ericson U, Persson Tholin J, Persson M, Lemetais G, Vanhaecke T, Dolci A, Perrier ET, Melander O. Water Supplementation Reduces Copeptin and Plasma Glucose in Adults With High Copeptin: The H2O Metabolism Pilot Study. *J Clin Endocrinol Metab* 2019;104(6):1917-1925
31. Juul KV, Bichet DG, Nielsen S, Nørgaard JP. The physiological and pathophysiological functions of renal and extrarenal vasopressin V2 receptors. *Am J Physiol Renal Physiol* 2014;306(9):F931-940
32. Mancinelli R, Franchitto A, Glaser S, Vetusch A, Venter J, Sferra R, Pannarale L, Olivero F, Carpino G, Alpini G, Onori P, Gaudio E. Vasopressin regulates the growth of the biliary epithelium in polycystic liver disease. *Lab Invest* 2016;96(11):1147-1155
33. Taveau C, Chollet C, Waeckel L, Desposito D, Bichet DG, Arthus M-F, Magnan C, Philippe E, Paradis V, Fougelle F, Hainault I, Enhörning S, Velho G, Roussel R, Bankir L, Melander O, Bouby N. Vasopressin and hydration play a major role in the development of glucose intolerance and hepatic steatosis in obese rats. *Diabetologia* 2015;58(5):1081-1090
34. Taveau C, Chollet C, Bichet DG, Velho G, Guillon G, Corbani M, Roussel R, Bankir L, Melander O, Bouby N. Acute and chronic hyperglycemic effects of vasopressin in normal rats: involvement of V(1A) receptors. *American journal of physiology. Endocrinology and metabolism* 2017;312(3):E127-E135
35. Enhörning S, Tasevska I, Roussel R, Bouby N, Persson M, Burri P, Bankir L, Melander O. Effects of hydration on plasma copeptin, glycemia and gluco-regulatory hormones: a water intervention in humans. *European journal of nutrition* 2019;58(1):315-324

36. Carroll HA, James LJ. Hydration, Arginine Vasopressin, and Glucoregulatory Health in Humans: A Critical Perspective. *Nutrients* 2019;11(6):1201
37. Koshimizu TA, Nakamura K, Egashira N, Hiroyama M, Nonoguchi H, Tanoue A. Vasopressin V1a and V1b receptors: from molecules to physiological systems. *Physiol Rev* 2012;92(4):1813-1864
38. Ostrowski NL, Young WS, 3rd, Knepper MA, Lolait SJ. Expression of vasopressin V1a and V2 receptor messenger ribonucleic acid in the liver and kidney of embryonic, developing, and adult rats. *Endocrinology* 1993;133(4):1849-1859
39. Lolait SJ, O'Carroll AM, Mahan LC, Felder CC, Button DC, Young WS, 3rd, Mezey E, Brownstein MJ. Extrahypothalamic expression of the rat V1b vasopressin receptor gene. *Proc Natl Acad Sci U S A* 1995;92(15):6783-6787
40. Andersen LJ, Andersen JL, Schutten HJ, Warberg J, Bie P. Antidiuretic effect of subnormal levels of arginine vasopressin in normal humans. *Am J Physiol* 1990;259(1 Pt 2):R53-60
41. Yasuoka Y, Kobayashi M Fau - Sato Y, Sato Y Fau - Zhou M, Zhou M Fau - Abe H, Abe H Fau - Okamoto H, Okamoto H Fau - Nonoguchi H, Nonoguchi H Fau - Tanoue A, Tanoue A Fau - Kawahara K, Kawahara K. The intercalated cells of the mouse kidney OMCD(is) are the target of the vasopressin V1a receptor axis for urinary acidification. *Clin Exp Nephrol* 2013;17(6):783-792 LID - 710.1007/s10157-10013-10783-y [doi]
42. Zhuo JL. Renomedullary interstitial cells: a target for endocrine and paracrine actions of vasoactive peptides in the renal medulla. *Clin Exp Pharmacol Physiol* 2000;27(7):465-473
43. Arpin-Bott MP, Waltisperger E, Freund-Mercier MJ, Stoeckel ME. Histoautoradiographic localization, pharmacology and ontogeny of V(1a) vasopressin binding sites in the rat kidney. *Nephron* 1999;83(1):74-84
44. Trinh-Trang-Tan MM, Bankir L. Integrated function of urea transporters in the mammalian kidney. *Exp Nephrol* 1998;6(6):471-479
45. Zhai XY, Fenton RA, Andreasen A, Thomsen JS, Christensen EI. Aquaporin-1 is not expressed in descending thin limbs of short-loop nephrons. *J Am Soc Nephrol* 2007;18(11):2937-2944
46. Al Therwani S, Rosenbæk JB, Mose FH, Bech JN, Pedersen EB. Effect of tolvaptan on renal water and sodium excretion and blood pressure during nitric oxide inhibition: a dose-response study in healthy subjects. *BMC nephrology* 2017;18(1):86-86
47. Zitteva D, Boertien WE, van Beek AP, Dullaart RPF, Franssen CFM, de Jong PE, Meijer E, Gansevoort RT. Vasopressin, copeptin, and renal concentrating capacity in patients with autosomal dominant polycystic kidney disease without renal impairment. *Clinical journal of the American Society of Nephrology : CJASN* 2012;7(6):906-913
48. Gansevoort RT, van Gastel MDA, Chapman AB, Blais JD, Czerwiec FS, Higashihara E, Lee J, Ouyang J, Perrone RD, Stade K, Torres VE, Devuyst O, Investigators T. Plasma

copeptin levels predict disease progression and tolvaptan efficacy in autosomal dominant polycystic kidney disease. *Kidney international* 2019;96(1):159-169

49. Xiang MA, Rybczynski PJ, Patel M, Chen RH, McComsey DF, Zhang H-C, Gunnet JW, Look R, Wang Y, Minor LK, Zhong HM, Villani FJ, Demarest KT, Damiano BP, Maryanoff BE. Next-generation spirobenzazepines: identification of RWJ-676070 as a balanced vasopressin V1a/V2 receptor antagonist for human clinical studies. *Bioorganic & medicinal chemistry letters* 2007;17(23):6623-6628

50. Gunnet JW, Wines P, Xiang M, Rybczynski P, Andrade-Gordon P, de Garavilla L, Parry TJ, Cheung W-M, Minor L, Demarest KT, Maryanoff BE, Damiano BP. Pharmacological characterization of RWJ-676070, a dual vasopressin V(1A)/V(2) receptor antagonist. *European journal of pharmacology* 2008;590(1-3):333-342

51. Mondritzki T, Kolkhof P, Sabbah HN, Gheorghide M, Fürstner C, Schmeck C, Siedentop H, Schaefer S, Truebel H. Differentiation of arginine vasopressin antagonistic effects by selective V2 versus dual V2/V1a receptor blockade in a preclinical heart failure model. *American journal of therapeutics* 2011;18(1):31-37

52. Heida JE, Gansevoort RT, Torres VE, Devuyst O, Perrone RD, Lee J, Li H, Ouyang J, Chapman AB. The Effect of Tolvaptan on BP in Polycystic Kidney Disease: A Post Hoc Analysis of the TEMPO 3:4 Trial. *J Am Soc Nephrol* 2021;Apr 22:ASN.2020101512. (doi: 10.1681/ASN.2020101512. Epub ahead of print. PMID: 33888577.)

53. Minami S, Hamano T, Iwatani H, Mizui M, Kimura Y, Isaka Y. Tolvaptan promotes urinary excretion of sodium and urea: a retrospective cohort study. *Clin Exp Nephrol* 2018;22(3):550-561

54. Enhörning S, Bankir L, Bouby N, Struck J, Hedblad B, Persson M, Morgenthaler NG, Nilsson PM, Melander O. Copeptin, a marker of vasopressin, in abdominal obesity, diabetes and microalbuminuria: the prospective Malmö Diet and Cancer Study cardiovascular cohort. *International journal of obesity (2005)* 2013;37(4):598-603

55. Enhörning S, Sjögren M, Hedblad B, Nilsson PM, Struck J, Melander O. Genetic vasopressin 1b receptor variance in overweight and diabetes mellitus. *European journal of endocrinology* 2016;174(1):69-75

56. Heida JE, Minović I, van Faassen M, Kema IP, Boertien WE, Bakker SJL, van Beek AP, Gansevoort RT. Effect of Vasopressin on the Hypothalamic-Pituitary-Adrenal Axis in ADPKD Patients during V2 Receptor Antagonism. *Am J Nephrol* 2020;51(11):861-870

57. Kramers BJ, van Gastel MDA, Boertien WE, Meijer E, Gansevoort RT. Determinants of Urine Volume in ADPKD Patients Using the Vasopressin V2 Receptor Antagonist Tolvaptan. *Am J Kidney Dis* 2019;73(3):354-362

58. Higashihara E, Nutahara K, Tanbo M, Hara H, Miyazaki I, Kobayashi K, Nitatori T. Does increased water intake prevent disease progression in autosomal dominant polycystic kidney disease? *Nephrol Dial Transplant* 2014;29(9):1710-1719

59. Enhörning S, Leosdottir M, Wallström P, Gullberg B, Berglund G, Wirfält E, Melander O. Relation between human vasopressin 1a gene variance, fat intake, and diabetes. *Am J Clin Nutr* 2009;89(1):400-406
60. Perucca J, Bichet DG, Bardoux P, Bouby N, Bankir L. Sodium excretion in response to vasopressin and selective vasopressin receptor antagonists. *J Am Soc Nephrol* 2008;19(9):1721-1731
61. Bankir L, Fernandes S, Bardoux P, Bouby N, Bichet DG. Vasopressin-V2 receptor stimulation reduces sodium excretion in healthy humans. *Journal of the American Society of Nephrology : JASN* 2005;16(7):1920-1928
62. Bankir L, Bichet DG, Bouby N. Vasopressin V2 receptors, ENaC, and sodium reabsorption: a risk factor for hypertension? *American journal of physiology. Renal physiology* 2010;299(5):F917-F928
63. Blanchard A, Frank M, Wuerzner G, Peyrard S, Bankir L, Jeunemaitre X, Azizi M. Antinatriuretic effect of vasopressin in humans is amiloride sensitive, thus ENaC dependent. *Clin J Am Soc Nephrol* 2011;6(4):753-759
64. Perico N, Zoja C, Corna D, Rottoli D, Gaspari F, Haskell L, Remuzzi G. V1/V2 Vasopressin receptor antagonism potentiates the renoprotection of renin-angiotensin system inhibition in rats with renal mass reduction. *Kidney Int* 2009;76(9):960-967
65. Nonoguchi H, Takayama M, Owada A, Ujiie K, Yamada T, Nakashima O, Sakuma Y, Koike J, Terada Y, Marumo F, Tomita K. Role of urinary arginine vasopressin in the sodium excretion in patients with chronic renal failure. *Am. J. Med. Sci.* 1996;312(5):195-201
66. Frolich JCE, Nies ASE, Schrier RWE. Proceedings of a conference on prostaglandins and the kidney. Stuttgart, Germany, July 23-24, 1980. *Kidney Int* 1981;19(6):755-880
67. Gross PA, Schrier RW, Anderson RJ. Prostaglandins and water metabolism: a review with emphasis on in vivo studies. *Kidney Int* 1981;19(6):839-850
68. Handler JS. Vasopressin-prostaglandin interactions in the regulation of epithelial cell permeability to water. *Kidney Int* 1981;19(6):831-838
69. Kramer HJ, Glanzer K, Dusing R. Role of prostaglandins in the regulation of renal water excretion. *Kidney Int* 1981;19(6):851-859
70. Dunn MJ, Hood VL. Prostaglandins and the kidney. *The American journal of physiology* 1977;233(3):169-184
71. Schlondorff D. Renal prostaglandin synthesis. Sites of production and specific actions of prostaglandins. *Am. J. Med.* 1986;81(2B):1-11
72. Bonvalet JP, Pradelles P, Farman N. Segmental synthesis and actions of prostaglandins along the nephron. *Am. J. Physiol.* 1987;253(3 Pt 2):F377-F387

73. Breyer MD, Breyer RM. Prostaglandin E receptors and the kidney. *American journal of physiology. Renal physiology* 2000;279(1):F12-F23
74. Harris RC. Cyclooxygenase-2 and the kidney: functional and pathophysiological implications. *Journal of hypertension. Supplement : official journal of the International Society of Hypertension* 2002;20(6):S3-S9
75. Hao C-M, Breyer MD. Physiological regulation of prostaglandins in the kidney. *Annual review of physiology* 2008;70:357-377
76. Nørregaard R, Kwon T-H, Frøkiær J. Physiology and pathophysiology of cyclooxygenase-2 and prostaglandin E2 in the kidney. *Kidney research and clinical practice* 2015;34(4):194-200
77. Olesen ET, Fenton RA. Is there a role for PGE2 in urinary concentration? *J Am Soc Nephrol* 2013;24(2):169-178
78. Frölich JC, Wilson TW, Sweetman BJ, Smigel M, Nies AS, Carr K, Watson JT, Oates JA. Urinary prostaglandins. Identification and origin. *The Journal of clinical investigation* 1975;55(4):763-770
79. Kremer J. From prostaglandin replacement to specific COX-2 inhibition: a critical appraisal. *The Journal of rheumatology. Supplement* 2000;60:9-12
80. Breyer MD. Prostaglandin receptors in the kidney: a new route for intervention? *Experimental nephrology* 1998;6(3):180-188
81. Hayashi S, Ueno N, Murase A, Takada J. Design, synthesis and structure-activity relationship studies of novel and diverse cyclooxygenase-2 inhibitors as anti-inflammatory drugs. *Journal of enzyme inhibition and medicinal chemistry* 2014;29(6):846-867
82. Kompanowska-Jeziarska E, Emmeluth C, Grove L, Christensen P, Sadowski J, Bie P. Mechanism of vasopressin natriuresis in the dog: role of vasopressin receptors and prostaglandins. *Am. J. Physiol.* 1998;274(6 Pt 2):R1619-R1625
83. Chou SY, Porush JG, Faubert PF. Renal medullary circulation: hormonal control. *Kidney Int.* 1990;37(1):1-13
84. Dixey JJ, Williams TD, Lightman SL, Lant AF, Brewerton DA. The effect of indomethacin on the renal response to arginine vasopressin in man. *Clin Sci (Lond)* 1986;70(5):409-416
85. Breyer MD, Jacobson HR, Hebert RL. Cellular mechanisms of prostaglandin E2 and vasopressin interactions in the collecting duct. *Kidney Int* 1990;38(4):618-624
86. Sharma JN, Jawad NM. Adverse effects of COX-2 inhibitors. *ScientificWorldJournal* 2005;5:629-645
87. Nasrallah R, Zimpelmann J, Eckert D, Ghossein J, Geddes S, Beique JC, Thibodeau JF, Kennedy CRJ, Burns KD, Hébert RL. PGE(2) EP(1) receptor inhibits vasopressin-

dependent water reabsorption and sodium transport in mouse collecting duct. *Lab Invest* 2018;98(3):360-370

88. Lieberthal W, Vasilevsky ML, Valari R, Levinsky N. Interactions between ADH and prostaglandins in isolated erythrocyte-perfused rat kidney. *Am. J. Physiol.* 1987;252:F331-F337
89. Ganguli M, Tobian L, Azar S, O'Donnell M. Evidence that prostaglandin synthesis inhibitors increase the concentration of sodium and chloride in rat renal medulla. *Circulation research* 1977;40(5 Suppl 1):I135-I139
90. Haylor J, Lote CJ. The influence of prostaglandin E2 and indomethacin on the renal corticomedullary solute gradient in the rat. *The Journal of pharmacy and pharmacology* 1983;35(5):299-305
91. Fejes-Tóth G, Magyar A, Walter J. Renal response to vasopressin after inhibition of prostaglandin synthesis. *The American journal of physiology* 1977;232(5):F416-F423
92. Anderson RJ, Berl T, McDonald KD, Schrier RW. Evidence for an in vivo antagonism between vasopressin and prostaglandin in the mammalian kidney. *J Clin Invest* 1975;56(2):420-426
93. Berl T, Raz A, Wald H, Horowitz J, Czaczkes W. Prostaglandin synthesis inhibition and the action of vasopressin: studies in man and rat. *Am. J. Physiol.* 1977;232(6):F529-F537
94. Haylor J. Prostaglandin synthesis and renal function in man. *The Journal of Physiology* 1980;298(1):383-396
95. Fleming EF, Athirakul K, Oliverio MI, Key M, Goulet J, Koller BH, Coffman TM. Urinary concentrating function in mice lacking EP3 receptors for prostaglandin E2. *The American journal of physiology* 1998;275(6):F955-F961
96. Bardoux P, Martin H, Ahloulay M, Schmitt F, Bouby N, Trinh-Trang-Tan MM, Bankir L. Vasopressin contributes to hyperfiltration, albuminuria, and renal hypertrophy in diabetes mellitus: study in vasopressin-deficient Brattleboro rats. *Proc Natl Acad Sci U S A* 1999;96(18):10397-10402
97. Loichot C, Cazaubon C, Grima M, De Jong W, Nisato D, Imbs JL, Barthelmebs M. Vasopressin does not effect hypertension caused by long-term nitric oxide inhibition. *Hypertension* 2000;35(2):602-608
98. Promeneur D, Bankir L, Hu MC, Trinh-Trang-Tan MM. Renal tubular and vascular urea transporters: influence of antidiuretic hormone on mRNA expression in Brattleboro rats. *J. Am. Soc. Nephrol.* 1998;9:1359-1366
99. Stokes JB. Integrated actions of renal medullary prostaglandins in the control of water excretion. *The American journal of physiology* 1981;240(6):F471-F480

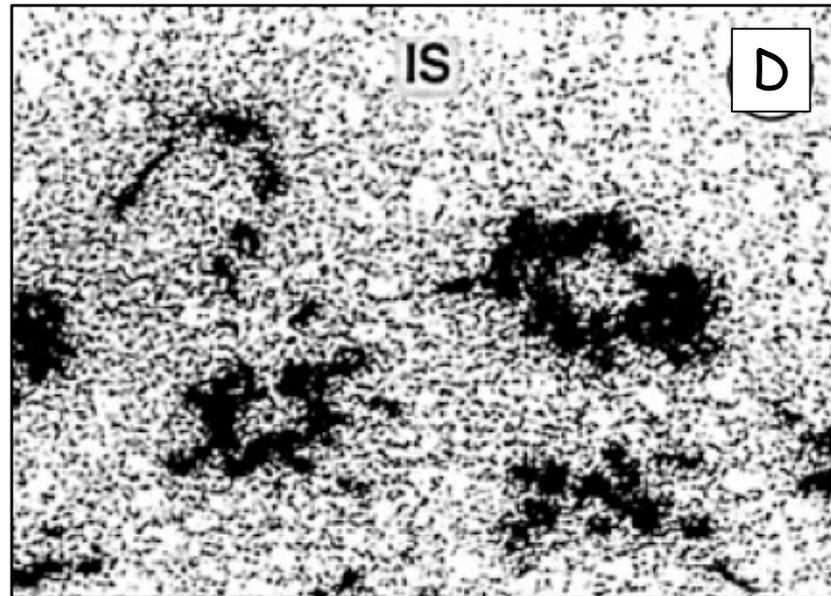
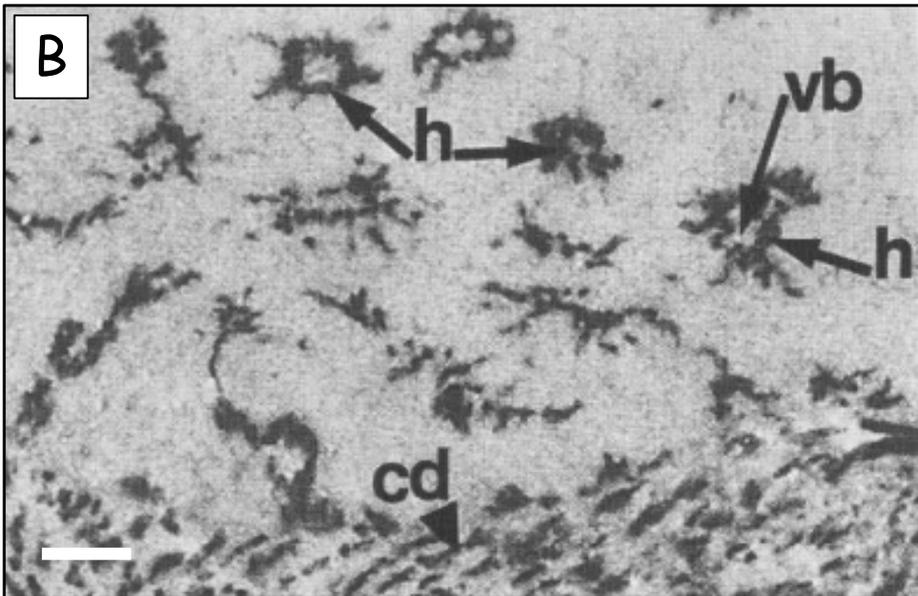
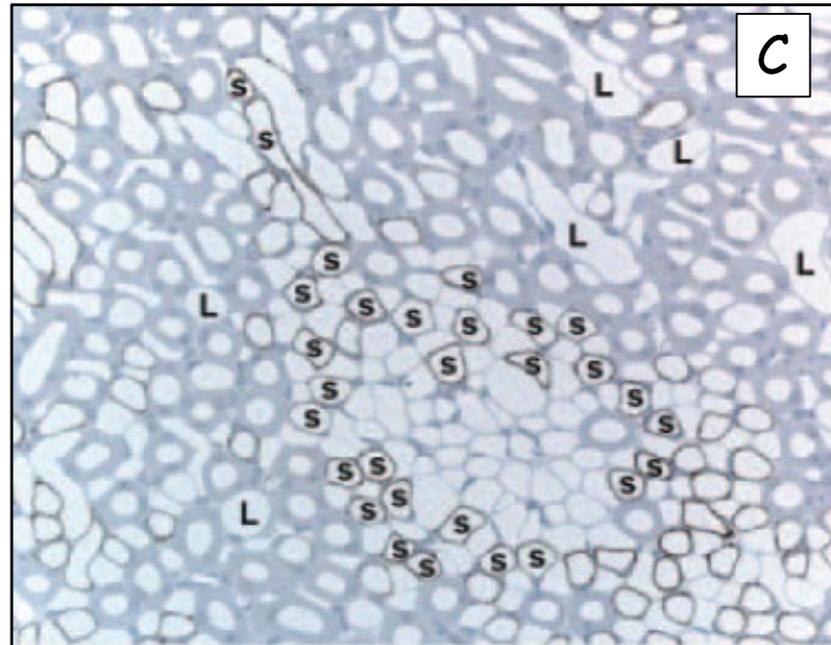
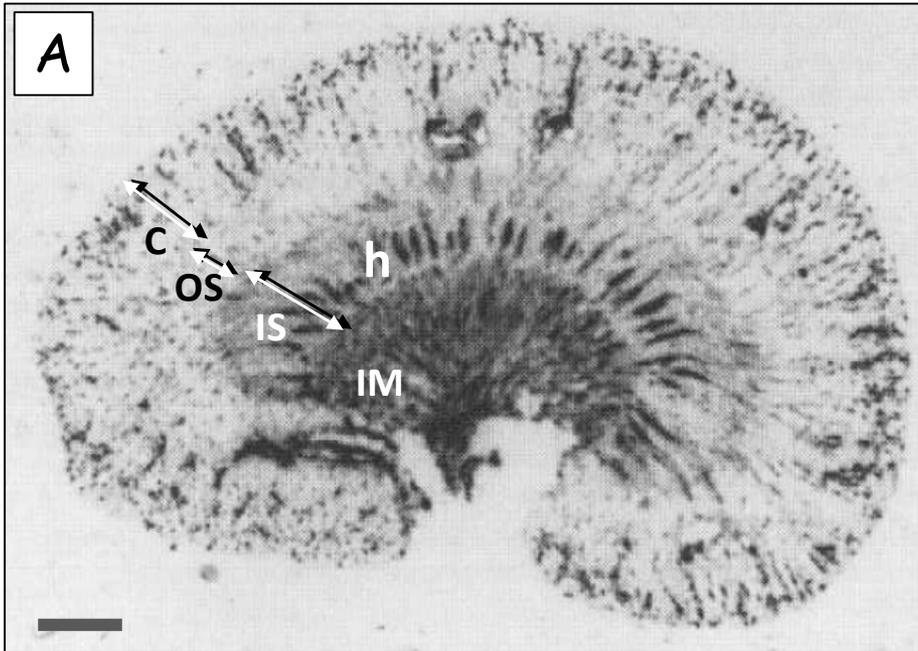
100. Orloff J, Zusman R. Role of prostaglandin E (PGE) in the modulation of the action of vasopressin on water flow in the urinary bladder of the toad and mammalian kidney. *The Journal of membrane biology* 1978;40 Spec No:297-304
101. Ando Y, Asano Y. Functional evidence for an apical V1 receptor in rabbit cortical collecting duct. *Am. J. Physiol.* 1993;264(3 Pt 2):F467-F471
102. Holt WF, Lechene C. ADH-PGE2 interactions in cortical collecting tubule. I. Depression of sodium transport. *The American journal of physiology* 1981;241(4):F452-F460
103. Izumi Y, Nakayama Y, Mori T, Miyazaki H, Inoue H, Kohda Y, Inoue T, Nonoguchi H, Tomita K. Downregulation of vasopressin V2 receptor promoter activity via V1a receptor pathway. *Am. J. Physiol. Renal Physiol.* 2007;292(5):F1418-F1426
104. Schmitt F, Bresson JL, Beressi N, Bichet DG, Chauveau D, Bankir L. Influence of plasma amino acid level on vasopressin secretion. *Diabetes & metabolism* 2003;29(4 Pt 1):352-361
105. Torikai S, Kurokawa K. Effect of PGE2 on vasopressin-dependent cell cAMP in isolated single nephron segments. *Am J Physiol* 1983;245(1):F58-66
106. Nakao A, Allen ML, Sonnenburg WK, Smith WL. Regulation of cAMP metabolism by PGE2 in cortical and medullary thick ascending limb of Henle's loop. *Am J Physiol* 1989;256(3 Pt 1):C652-657
107. Abdullah HI, Pedraza PL, McGiff JC, Ferreri NR. Calcium-sensing receptor signaling pathways in medullary thick ascending limb cells mediate COX-2-derived PGE2 production: functional significance. *Am J Physiol Renal Physiol* 2008;295(4):F1082-1089
108. Stokes JB. Effect of prostaglandin E2 on chloride transport across the rabbit thick ascending limb of Henle. Selective inhibitions of the medullary portion. *The Journal of clinical investigation* 1979;64(2):495-502
109. Culpepper RM, Andreoli TE. Interactions among prostaglandin E2, antidiuretic hormone, and cyclic adenosine monophosphate in modulating Cl<sup>-</sup> absorption in single mouse medullary thick ascending limbs of Henle. *The Journal of clinical investigation* 1983;71(6):1588-1601
110. Good DW, George T. Regulation of HCO<sub>3</sub><sup>-</sup> absorption by prostaglandin E2 and G proteins in rat medullary thick ascending limb. *The American journal of physiology* 1996;270(5 Pt 2):F711-F717
111. Peterson LN, McKay AJ, Borzecki JS. Endogenous prostaglandin E2 mediates inhibition of rat thick ascending limb Cl reabsorption in chronic hypercalcemia. *The Journal of clinical investigation* 1993;91(6):2399-2407
112. Kaji DM, Chase HS, Jr., Eng JP, Diaz J. Prostaglandin E2 inhibits Na-K-2Cl cotransport in medullary thick ascending limb cells. *The American journal of physiology* 1996;271(1 Pt 1):C354-C361

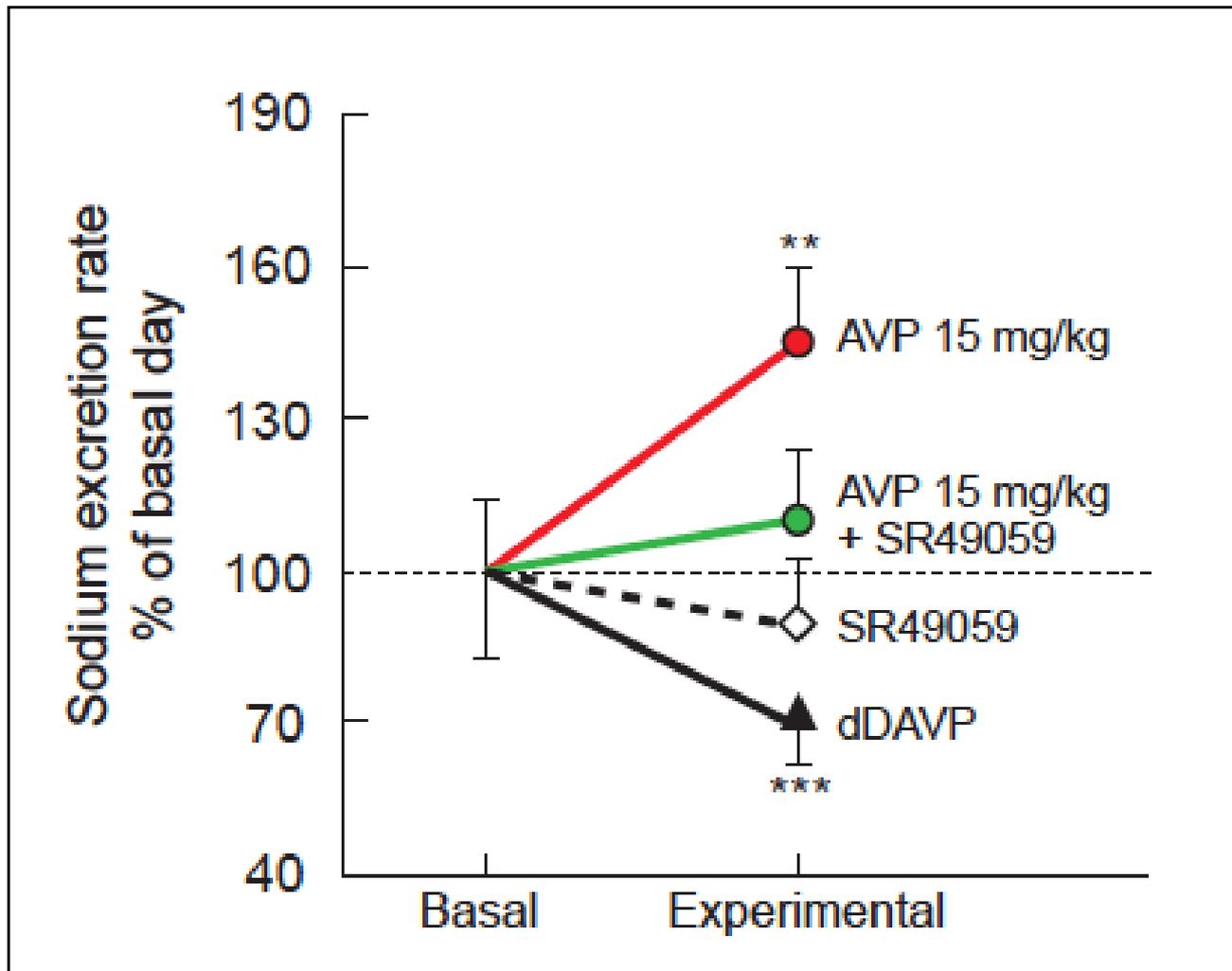
113. Fernández-Llama P, Ecelbarger CA, Ware JA, Andrews P, Lee AJ, Turner R, Nielsen S, Knepper MA. Cyclooxygenase inhibitors increase Na-K-2Cl cotransporter abundance in thick ascending limb of Henle's loop. *Am J Physiol* 1999;277(2):F219-226
114. Bohman SO, Jensen PK. The interstitial cells in the renal medulla of rat, rabbit, and gerbil in different states of diuresis. *Cell Tissue Res* 1978;189(1):1-18
115. Lemley KV, Kriz W. Anatomy of the renal interstitium. *Kidney international* 1991;39(3):370-381
116. Campean V, Theilig F Fau - Paliege A, Paliege A Fau - Breyer M, Breyer M Fau - Bachmann S, Bachmann S. Key enzymes for renal prostaglandin synthesis: site-specific expression in rodent. *Am J Physiol Renal Physiol* 2003;285(1):F19-32
117. Dunn MJ, Staley RS, Harrison M. Characterization of prostaglandin production in tissue culture of rat renal medullary cells. *Prostaglandins* 1976;12(1):37-49
118. Bohman SO. Demonstration of prostaglandin synthesis in collecting duct cells and other cell types of the rabbit renal medulla. *Prostaglandins* 1977;14(4):729-744
119. Zusman RM, Keiser HR. Prostaglandin biosynthesis by rabbit renomedullary interstitial cells in tissue culture. Stimulation by angiotensin II, bradykinin, and arginine vasopressin. *J Clin Invest* 1977;60(1):215-223
120. Beck TR, Hassid A, Dunn MJ. The effect of arginine vasopressin and its analogs on the synthesis of prostaglandin E2 by rat renal medullary interstitial cells in culture. *J. Pharmacol. Exp. Ther.* 1980;215(1):15-19
121. Shimizu K, Nakao A, Nonaka T, Oka H. Identification of vasopressin and determination of its corticomedullary levels in rat kidney tissue. *Kidney Int* 1984;26(6):785-790
122. Lemley KV, Schmitt SL, Holliger C, Dunn MJ, Robertson CR, Jamison RL. Prostaglandin synthesis inhibitors and vasa recta erythrocyte velocities in the rat. *The American journal of physiology* 1984;247(4 Pt 2):F562-F567
123. Yoshida M, Ueda S, Soejima H, Tsuruta K, Ikegami K. Effects of prostaglandin E2 and I2 on renal cortical and medullary blood flow in rabbits. *Arch Int Pharmacodyn Ther* 1986;282(1):108-117
124. Gomez SI, Strick DM, Romero JC. Role of nitric oxide and prostaglandin in the maintenance of cortical and renal medullary blood flow. *Braz J Med Biol Res* 2008;41(2):170-175
125. Fenton RA. Urea transporters and renal function: lessons from knockout mice. *Curr Opin Nephrol Hypertens* 2008;17(5):513-518
126. Yang B, Bankir L. Urea and urine concentrating ability: new insights from studies in mice. *Am J Physiol Renal Physiol* 2005;288(5):F881-896

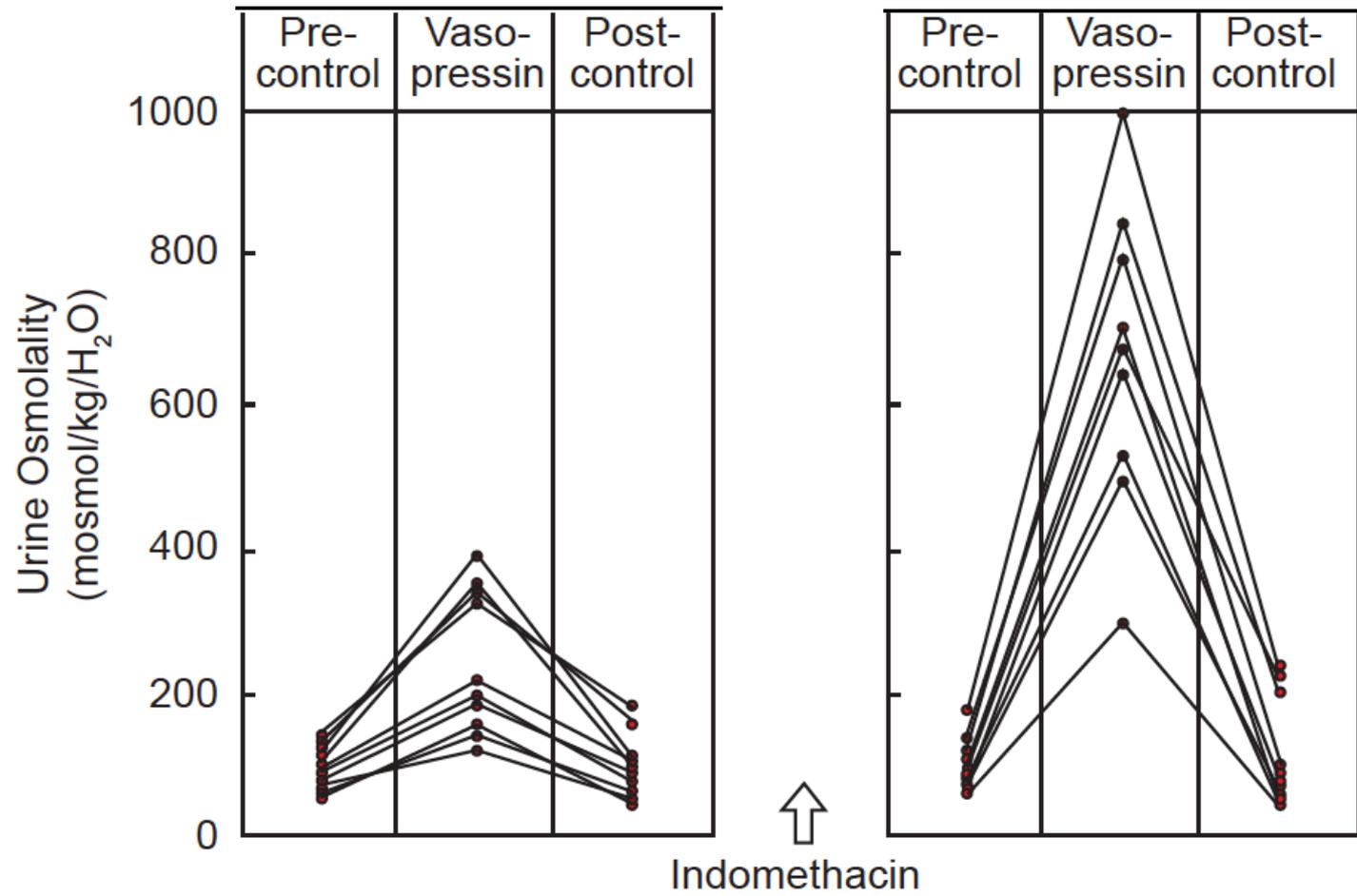
127. Geng X, Zhang S, He J, Ma A, Li Y, Li M, Zhou H, Chen G, Yang B. The urea transporter UT-A1 plays a predominant role in a urea-dependent urine-concentrating mechanism. *J Biol Chem* 2020;295(29):9893-9900
128. Roman RJ, Lechene C. Prostaglandin E2 and F2 alpha reduces urea reabsorption from the rat collecting duct. *Am J Physiol* 1981;241(1):F53-60
129. Sullivan JC, Sasser Jm Fau - Pollock DM, Pollock Dm Fau - Pollock JS, Pollock JS. Sexual dimorphism in renal production of prostanoids in spontaneously. *Hypertension* 2005;45(3):406-411
130. Eloy L, Grunfeld JP, Bayle F, Bankir L, Ramos-Frendo B, Trinh-Trang-Tan MM. Papillary plasma flow in rats. II. Hormonal control. *Pflugers Arch* 1983;398(3):253-258
131. Parekh N, Zou Ap Fau - Jungling I, Jungling I Fau - Endlich K, Endlich K Fau - Sadowski J, Sadowski J Fau - Steinhausen M, Steinhausen M. Sex differences in control of renal outer medullary circulation in rats: role of. *Am J Physiol* 1993;264(4 Pt 2):F629-636
132. Bankir L, Bouby N, Trinh-Trang-Tan MM, Ahloulay M, Promeneur D. Direct and indirect cost of urea excretion. *Kidney Int* 1996;49(6):1598-1607
133. Nüsing RM, Treude A, Weissenberger C, Jensen B, Bek M, Wagner C, Narumiya S, Seyberth HW. Dominant role of prostaglandin E2 EP4 receptor in furosemide-induced salt-losing tubulopathy: a model for hyperprostaglandin E syndrome/antenatal Bartter syndrome. *J Am Soc Nephrol* 2005;16(8):2354-2362
134. Reinalter SC, Jeck N, Brochhausen C, Watzer B, Nüsing RM, Seyberth HW, Kömhoff M. Role of cyclooxygenase-2 in hyperprostaglandin E syndrome/antenatal Bartter syndrome. *Kidney Int* 2002;62(1):253-260
135. Guay-Woodford LM. Bartter syndrome: unraveling the pathophysiologic enigma. *Am J Med* 1998;105(2):151-161
136. Nüsing RM, Seyberth HW. The role of cyclooxygenases and prostanoid receptors in furosemide-like salt losing tubulopathy: the hyperprostaglandin E syndrome. *Acta Physiol Scand* 2004;181(4):523-528
137. Kemter E, Rathkolb B, Bankir L, Schrewe A, Hans W, Landbrecht C, Klaften M, Ivandic B, Fuchs H, Gailus-Durner V, Hrabé de Angelis M, Wolf E, Wanke R, Aigner B. Mutation of the Na(+)-K(+)-2Cl(-) cotransporter NKCC2 in mice is associated with severe polyuria and a urea-selective concentrating defect without hyperreninemia. *American journal of physiology. Renal physiology* 2010;298(6):F1405-F1415
138. Park F, Mattson DL, Skelton MM, Cowley AW, Jr. Localization of the vasopressin V1a and V2 receptors within the renal cortical and medullary circulation. *Am J Physiol* 1997;273(1 Pt 2):R243-251
139. Aldasoro M, Medina P, Vila JM, Otero E, Martinez-León JB, Lluch S. Endothelium-dependent relaxation of human saphenous veins in response to vasopressin and desmopressin. *J Vasc Surg* 1997;25(4):696-703

140. Vila JM, Aldasoro M, Segarra G, Martínez-León JB, Mauricio MD, Lluch S, Medina P. Contractile responses of human thyroid arteries to vasopressin. *Life Sci* 2013;93(15):525-529
141. Medina P, Segarra G, Vila JM, Chuan P, Domenech C, Lluch S. V2-receptor-mediated relaxation of human renal arteries in response to desmopressin. *Am J Hypertens* 1999;12(2 Pt 1):188-193
142. Ohlstein EH, Berkowitz BA. Human vascular vasopressin receptors: analysis with selective vasopressin receptor antagonists. *J Pharmacol Exp Ther* 1986;239(3):737-741
143. Kaufmann JE, Iezzi M, Vischer UM. Desmopressin (DDAVP) induces NO production in human endothelial cells via V2 receptor- and cAMP-mediated signaling. *J Thromb Haemost* 2003;1(4):821-828
144. Mannucci PM, Aberg M, Nilsson IM, Robertson B. Mechanism of plasminogen activator and factor VIII increase after vasoactive drugs. *Br J Haematol* 1975;30(1):81-93
145. Mannucci PM, Ruggeri ZM, Pareti FI, Capitanio A. 1-Deamino-8-d-arginine vasopressin: a new pharmacological approach to the management of haemophilia and von Willebrand's diseases. *Lancet* 1977;1(8017):869-872
146. Kaufmann JE, Oksche A, Wollheim CB, Günther G, Rosenthal W, Vischer UM. Vasopressin-induced von Willebrand factor secretion from endothelial cells involves V2 receptors and cAMP. *J Clin Invest* 2000;106(1):107-116
147. Bichet DG, Razi M, Lonergan M, Arthus MF, Papukna V, Kortas C, Barjon JN. Hemodynamic and coagulation responses to 1-desamino[8-D-arginine] vasopressin in patients with congenital nephrogenic diabetes insipidus. *N Engl J Med* 1988;318(14):881-887
148. Lorthioir A, Joannidès R, Rémy-Jouet I, Fréguin-Bouilland C, Iacob M, Roche C, Monteil C, Lucas D, Renet S, Audrézet MP, Godin M, Richard V, Thuillez C, Guerrot D, Bellien J. Polycystin deficiency induces dopamine-reversible alterations in flow-mediated dilatation and vascular nitric oxide release in humans. *Kidney Int* 2015;87(2):465-472
149. El-Damanawi R, Lee M, Harris T, Cowley LB, Bond S, Pavey H, Sandford RN, Wilkinson IB, Karet Frankl FE, Hiemstra TF. High water vs. ad libitum water intake for autosomal dominant polycystic kidney disease: a randomized controlled feasibility trial. *Qjm* 2020;113(4):258-265
150. Nakamura Y, Watanabe H, Tanaka A, Yasui M, Nishihira J, Murayama N. Effect of Increased Daily Water Intake and Hydration on Health in Japanese Adults. *Nutrients* 2020;12(4)
151. Daudon M, Hennequin C, Boujelben G, Lacour B, Jungers P. Serial crystalluria determination and the risk of recurrence in calcium stone formers. *Kidney international* 2005;67(5):1934-1943

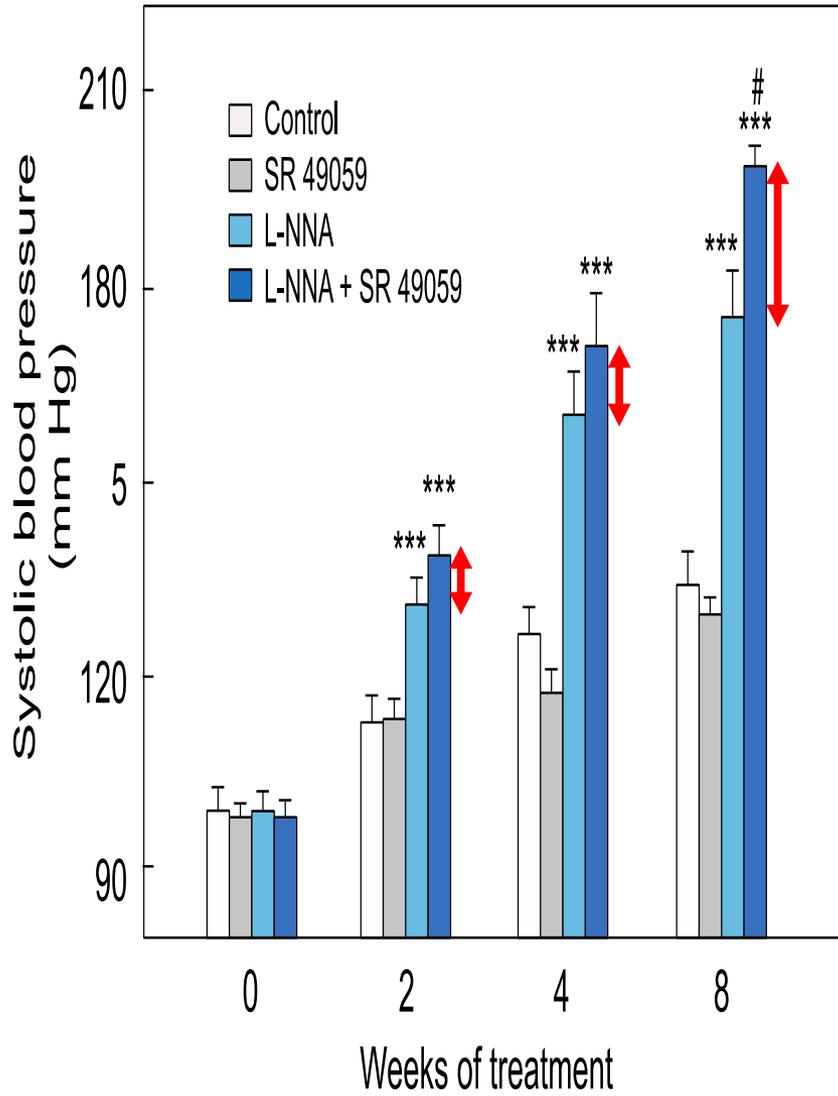
152. Bankir L, Daudon M. Recurrent (as opposed to non-recurrent) stone formers failed to increase urine volume significantly over a 3 year period in spite of recommendations to drink more. Abstract # TH-PO817. ASN Annual meeting, Abstract book 2008:294A
153. Perucca J, Bouby N, Valeix P, Bankir L. Sex difference in urine concentration across differing ages, sodium intake, and level of kidney disease. *Am J Physiol Regul Integr Comp Physiol* 2007;292(2):R700-705
154. Meijer E, Gansevoort RT. Vasopressin V2 receptor antagonists in autosomal dominant polycystic kidney disease: efficacy, safety, and tolerability. *Kidney Int* 2020;98(2):289-293
155. Muto S, Okada T, Yasuda M, Tsubouchi H, Nakajima K, Horie S. Long-term safety profile of tolvaptan in autosomal dominant polycystic kidney disease patients: TEMPO Extension Japan Trial. *Drug Healthc Patient Saf* 2017;9:93-104
156. Devuyst O, Chapman AB, Gansevoort RT, Higashihara E, Perrone RD, Torres VE, Blais JD, Zhou W, Ouyang J, Czerwiec FS. Urine Osmolality, Response to Tolvaptan, and Outcome in Autosomal Dominant Polycystic Kidney Disease: Results from the TEMPO 3:4 Trial. *Journal of the American Society of Nephrology : JASN* 2017;28(5):1592-1602
157. Min HK, Sung SA, Lee SY, Lee SW. Sub-morbid dehydration-associated glomerular hyperfiltration: An emerging reality? *Kidney Res Clin Pract* 2019;38(2):196-204
158. Bankir L. The antidiuretic effect of AVP leads to glomerular hyperfiltration. But this effect is biphasic. *Néphrologie & Thérapeutique* 2019;15(5):368
159. Wagner S, Merklung T, Metzger M, Bankir L, Laville M, Frimat L, Combe C, Jacquelinet C, Fouque D, Massy ZA, Stengel B. Water intake and progression of chronic kidney disease: the CKD-REIN cohort study. *Nephrol Dial Transplant* 2021



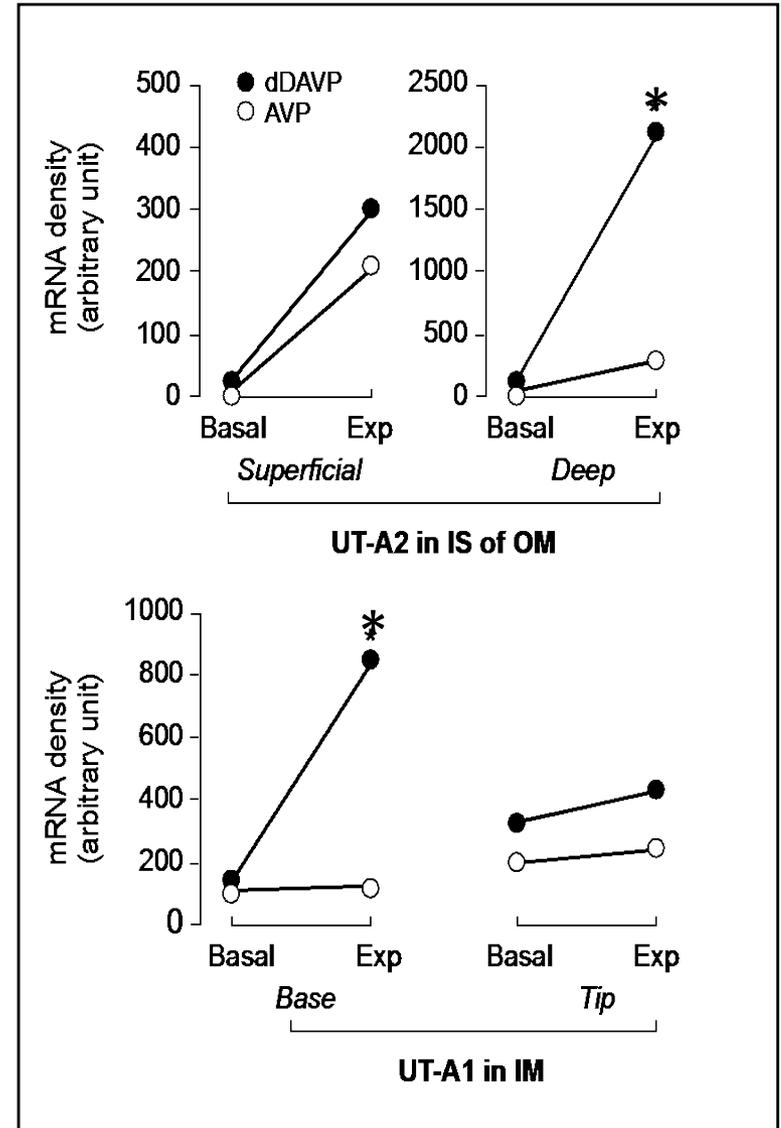




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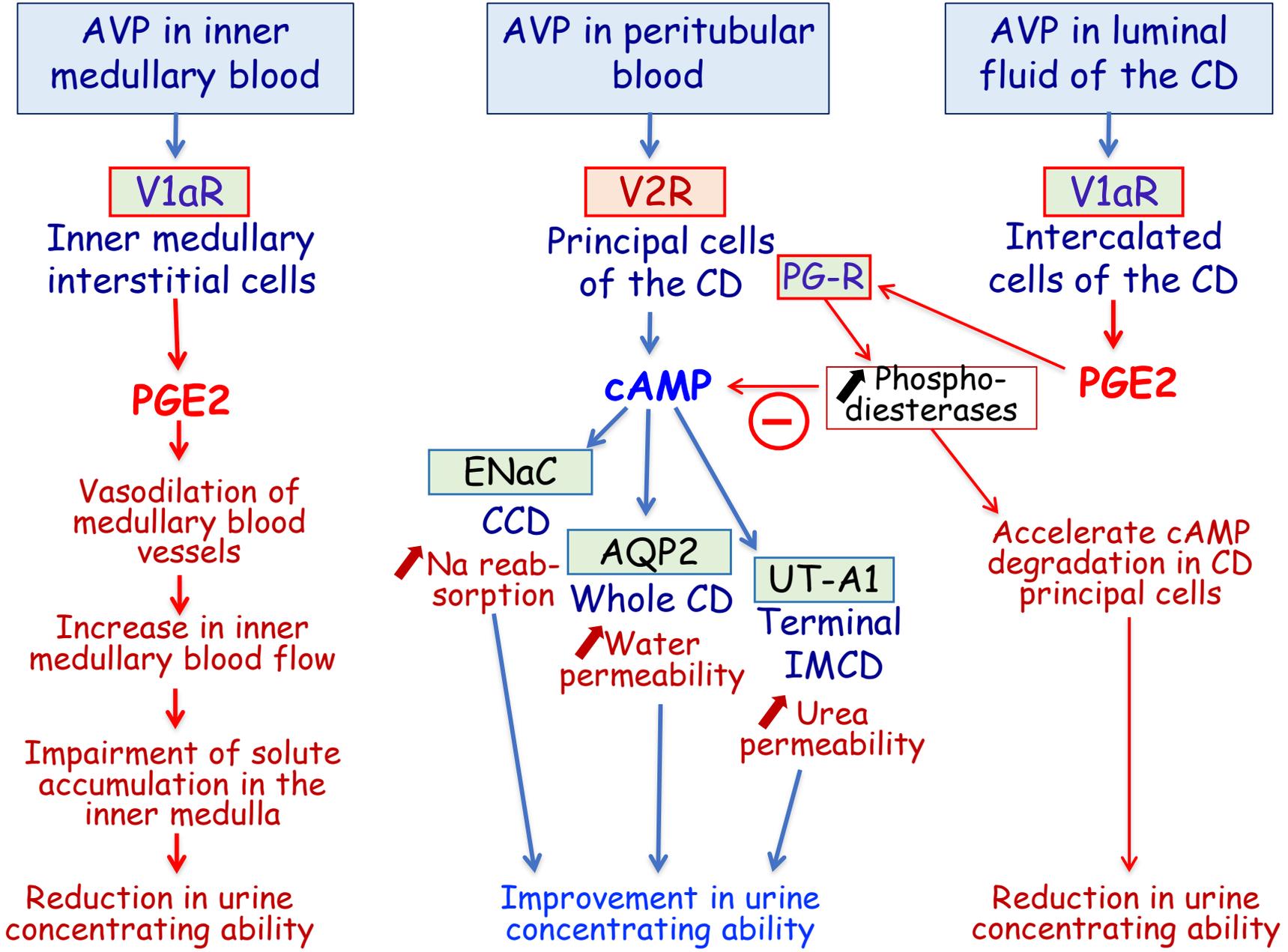


Table 1. Target sites of circulating vasopressin in the body and the receptors involved (target sites in the central nervous system are not considered here).

	<b>V2 receptors</b>	<b>V1a receptors</b>	<b>V1b = V3 receptors</b>
Vascular smooth muscle cells		Yes	
Endothelium	Yes		
Platelets		Yes	
Heart			Yes
Kidney	Yes	Yes	
Liver		Yes	
Adipocytes		Yes	
Spleen			Yes
Lung (pneumocytes)	Yes		
Inner ear	Yes		
Pancreatic islets			Yes
Adrenal gland		Yes	Yes
Anterior hypophysis			Yes
Thymus			Yes
Uterus			Yes
Breast			Yes

Table 2. Main structures expressing V2 and V1a receptors in the kidney, and resulting main effects

<b>Structures</b>	<b>Main effects</b>
<b><u>V2 receptors</u></b>	
Thick ascending limb (basolat. mbne)	Stimulation of Na-K-2Cl mediated transport
Connecting tubule (basolat. mbne)	Insertion of AQP2 in the luminal membrane
CD principal cells (basolat. mbne)	Insertion of AQP2 in the luminal membrane Stimulation of ENaC-mediated Na reabsorption Activation of UT-A1 in the terminal IMCD
<b><u>V1a receptors</u></b>	
Arterial vasa recta smooth muscle cells	Vasoconstriction (not significant in vivo for usual AVP concentrations)
CD intercalated cells (luminal membrane)	Stimulation of prostaglandin production
Inner medullary interstitial cells	Stimulation of prostaglandin production, leading to vasodilation of medullary vasculature

basolat. mbne = basolateral membrane

Table 3. Similarities and differences induced by voluntary increase in hydration or treatment with a vaptan

	Voluntary Increase in Hydration	Treatment with a Vaptan
Water intake	Increased	Increased
Urine volume	Increased	Increased
Urine osmolality	Decreased	Decreased
Plasma osmolality	Decreased	Increased
Thirst	Decreased	Increased
Vasopressin secretion	Decreased	Increased
V1aR- and V1bR-mediated effects	Decreased	Increased
Cause of higher water intake	Voluntary	Increased thirst
Observance	Difficult to drink in excess of thirst	Easy to take a pill

V1aR and V1bR = vasopressin V1a and V1b receptors, respectively.

Table 4. Influence of either AVP or dDAVP infusion on CKD progression in Brattleboro rats. Results observed during the third month after 5/6<sup>th</sup> nephrectomy and initiation of the AVP or dDAVP infusion.

	Control (DI)	AVP (V1a & V2 agonism)		dDAVP (V2 agonism)	
Body weight (g)	339 ± 7	364 ± 12		370 ± 18	
Food intake (g/d)	13.8 ± 0.5	13.8 ± 1.4		14.5 ± 1.8	
Urine flow rate (ml/d)	105 ± 8	20 ± 3	§	32 ± 6	§
Urine osmolality (mosm/kg H <sub>2</sub> O)	199 ± 13	997 ± 141	§	694 ± 129	§
Urinary protein excretion (mg/d)	23 ± 3	21 ± 4		49 ± 10	§ #
Systolic blood pressure (mm Hg)	194 ± 12	176 ± 15		204 ± 12	
Hematocrit (%)	43 ± 1	40 ± 2		36 ± 2	§
Plasma sodium concentration (mmol/L)	154 ± 1	146 ± 3	§	148 ± 2	§
Plasma potassium concentration (mmol/L)	3.36 ± 0.05	3.87 ± 0.17	§	3.95 ± 0.13	§
Plasma urea concentration (mmol/L)	15.4 ± 1	15.6 ± 2.0		30.9 ± 7.3	§ #
Plasma creatinine concentration (µmol/L)	107 ± 3	100 ± 6		127 ± 11	§ #
Kidney weight (mg/100g BW)	331 ± 16	541 ± 72	§	671 ± 50	§

Comparison of the three groups by a one-way ANOVA followed by Fisher post-hoc test.

AVP or dDAVP versus control : § = p < 0.05 or less; dDAVP versus AVP : # = p < 0.05 or less

Data from reference 24.