



HAL
open science

Different Clinical Contexts of Use of Blood Neurofilament Light Chain Protein in the Spectrum of Neurodegenerative Diseases

Giovanni Palermo, Sonia Mazzucchi, Alessandra Della Vecchia, Gabriele Siciliano, Ubaldo Bonuccelli, Carole Azuar, Roberto Ceravolo, Simone Lista, Harald Hampel, Filippo Baldacci

► To cite this version:

Giovanni Palermo, Sonia Mazzucchi, Alessandra Della Vecchia, Gabriele Siciliano, Ubaldo Bonuccelli, et al.. Different Clinical Contexts of Use of Blood Neurofilament Light Chain Protein in the Spectrum of Neurodegenerative Diseases. *Molecular Neurobiology*, 2020, 57 (11), pp.4667-4691. 10.1007/s12035-020-02035-9 . hal-03385896

HAL Id: hal-03385896

<https://hal.sorbonne-universite.fr/hal-03385896>

Submitted on 19 Oct 2021

HAL is a multi-disciplinary open access archive for the deposit and dissemination of scientific research documents, whether they are published or not. The documents may come from teaching and research institutions in France or abroad, or from public or private research centers.

L'archive ouverte pluridisciplinaire **HAL**, est destinée au dépôt et à la diffusion de documents scientifiques de niveau recherche, publiés ou non, émanant des établissements d'enseignement et de recherche français ou étrangers, des laboratoires publics ou privés.

Dear Author

Here are the proofs of your article.

- You can submit your corrections **online** or by **fax**.
- For **online** submission please insert your corrections in the online correction form. Always indicate the line number to which the correction refers.
- Please return your proof together with the permission to publish confirmation.
- For **fax** submission, please ensure that your corrections are clearly legible. Use a fine black pen and write the correction in the margin, not too close to the edge of the page.
- Remember to note the journal title, article number, and your name when sending your response via e-mail, fax or regular mail.
- **Check** the metadata sheet to make sure that the header information, especially author names and the corresponding affiliations are correctly shown.
- **Check** the questions that may have arisen during copy editing and insert your answers/corrections.
- **Check** that the text is complete and that all figures, tables and their legends are included. Also check the accuracy of special characters, equations, and electronic supplementary material if applicable. If necessary refer to the *Edited manuscript*.
- The publication of inaccurate data such as dosages and units can have serious consequences. Please take particular care that all such details are correct.
- Please **do not** make changes that involve only matters of style. We have generally introduced forms that follow the journal's style. Substantial changes in content, e.g., new results, corrected values, title and authorship are not allowed without the approval of the responsible editor. In such a case, please contact the Editorial Office and return his/her consent together with the proof.
- If we do not receive your corrections **within 48 hours**, we will send you a reminder.

Please note

Your article will be published **Online First** approximately one week after receipt of your corrected proofs. This is the **official first publication** citable with the DOI.

Further changes are, therefore, not possible.

After online publication, subscribers (personal/institutional) to this journal will have access to the complete article via the DOI using the URL:

<http://dx.doi.org/10.1007/s12035-020-02035-9>

If you would like to know when your article has been published online, take advantage of our free alert service. For registration and further information, go to:

<http://www.springerlink.com>.

Due to the electronic nature of the procedure, the manuscript and the original figures will only be returned to you on special request. When you return your corrections, please inform us, if you would like to have these documents returned.

The **printed version** will follow in a forthcoming issue.

32	Author	Family Name	Vecchia
33		Particle	
34		Given Name	Alessandra Della
35		Suffix	
36		Organization	University of Pisa
37		Division	Department of Clinical and Experimental Medicine
38		Address	Pisa, Italy
39		e-mail	
40		Family Name	Siciliano
41		Particle	
42		Given Name	Gabriele
43		Suffix	
44	Author	Organization	University of Pisa
45		Division	Department of Clinical and Experimental Medicine
46		Address	Pisa, Italy
47		e-mail	
48		Family Name	Bonuccelli
49		Particle	
50		Given Name	Ubaldo
51		Suffix	
52	Author	Organization	University of Pisa
53		Division	Department of Clinical and Experimental Medicine
54		Address	Pisa, Italy
55		e-mail	
56		Family Name	Azuar
57		Particle	
58		Given Name	Carole
59		Suffix	
60		Organization	AP-HP - Hôpital Pitié-Salpêtrière
61	Author	Division	Centre National de Référence des Démences Rares ou Précoces, IM2A, Département de Neurologie
62		Address	Paris, France
63		Organization	Institut du Cerveau et de la Moelle épinière (ICM)
64		Division	FrontLab
65		Address	Paris, France
66		e-mail	
67	Author	Family Name	Ceravolo
68		Particle	

69		Given Name	Roberto
70		Suffix	
71		Organization	University of Pisa
72		Division	Department of Clinical and Experimental Medicine
73		Address	Pisa, Italy
74		e-mail	
<hr/>			
75		Family Name	Lista
76		Particle	
77		Given Name	Simone
78		Suffix	
79		Organization	Sorbonne University
80		Division	GRC n° 21, Alzheimer Precision Medicine (APM), AP-HP, Pitié-Salpêtrière Hospital
81	Author	Address	Boulevard de l'hôpital, Paris, France
82		Organization	Brain & Spine Institute (ICM)
83		Division	INSERM U 1127, CNRS UMR 7225
84		Address	Boulevard de l'hôpital, Paris, France
85		Organization	Pitié-Salpêtrière Hospital
86		Division	Department of Neurology, Institute of Memory and Alzheimer's Disease (IM2A)
87		Address	Paris, France
88		e-mail	
<hr/>			
89		Family Name	Hampel
90		Particle	
91		Given Name	Harald
92		Suffix	
93	Author	Organization	Institut du Cerveau et de la Moelle épinière (ICM)
94		Division	FrontLab
95		Address	Paris, France
96		e-mail	
<hr/>			
97		Received	10 May 2020
98	Schedule	Revised	
99		Accepted	22 July 2020
<hr/>			
100	Abstract	<p>One of the most pressing challenges in the clinical research of neurodegenerative diseases (NDDs) is the validation and standardization of pathophysiological biomarkers for different contexts of use (CoUs), such as early detection, diagnosis, prognosis, and prediction of treatment response. Neurofilament light chain (NFL) concentration is a particularly promising candidate, an indicator of axonal degeneration, which can be analyzed in peripheral blood with advanced ultrasensitive methods. Serum/plasma NFL</p>	

concentration is closely correlated with cerebrospinal fluid NFL and directly reflects neurodegeneration within the central nervous system. Here, we provide an update on the feasible CoU of blood NFL in NDDs and translate recent findings to potentially valuable clinical practice applications. As NFL is not a disease-specific biomarker, however, blood NFL is an easily accessible biomarker with promising different clinical applications for several NDDs: (1) early detection and diagnosis (i.e., amyotrophic lateral sclerosis, Creutzfeldt–Jakob disease, atypical parkinsonisms, sporadic late-onset ataxias), (2) prognosis (Huntington’s disease and Parkinson’s disease), and (3) prediction of time to symptom onset (presymptomatic mutation carriers in genetic Alzheimer’s disease and spinocerebellar ataxia type 3).

101	Keywords separated by ' - '	Alzheimer’s disease - Amyotrophic lateral sclerosis - Biomarkers - Creutzfeldt–Jakob disease - NFL - Parkinsonian syndromes
102	Foot note information	Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.

Different Clinical Contexts of Use of Blood Neurofilament Light Chain Protein in the Spectrum of Neurodegenerative Diseases

Giovanni Palermo¹ · Sonia Mazzucchi¹ · Alessandra Della Vecchia¹ · Gabriele Siciliano¹ · Ubaldo Bonuccelli¹ · Carole Azuar^{2,3} · Roberto Ceravolo¹ · Simone Lista^{4,5,6} · Harald Hampel³ · Filippo Baldacci^{1,3}

Received: 10 May 2020 / Accepted: 22 July 2020
© Springer Science+Business Media, LLC, part of Springer Nature 2020

Abstract

One of the most pressing challenges in the clinical research of neurodegenerative diseases (NDDs) is the validation and standardization of pathophysiological biomarkers for different contexts of use (CoUs), such as early detection, diagnosis, prognosis, and prediction of treatment response. Neurofilament light chain (NFL) concentration is a particularly promising candidate, an indicator of axonal degeneration, which can be analyzed in peripheral blood with advanced ultrasensitive methods. Serum/plasma NFL concentration is closely correlated with cerebrospinal fluid NFL and directly reflects neurodegeneration within the central nervous system. Here, we provide an update on the feasible CoU of blood NFL in NDDs and translate recent findings to potentially valuable clinical practice applications. As NFL is not a disease-specific biomarker, however, blood NFL is an easily accessible biomarker with promising different clinical applications for several NDDs: (1) early detection and diagnosis (i.e., amyotrophic lateral sclerosis, Creutzfeldt–Jakob disease, atypical parkinsonisms, sporadic late-onset ataxias), (2) prognosis (Huntington's disease and Parkinson's disease), and (3) prediction of time to symptom onset (presymptomatic mutation carriers in genetic Alzheimer's disease and spinocerebellar ataxia type 3).

Keywords Alzheimer's disease · Amyotrophic lateral sclerosis · Biomarkers · Creutzfeldt–Jakob disease · NFL · Parkinsonian syndromes

Introduction

Neurodegenerative diseases (NDDs) are currently considered as a continuum of disorders with common pathophysiological

mechanisms, including misfolded protein deposition, neuronal synaptic disruption, axonal degeneration, neuroinflammation, and oxidative stress [1–3]. Therefore, the greatest current challenge in the field of NDDs is to provide biomarkers for the pathological mechanisms underlying each clinical picture [4], in order to improve the diagnostic and prognostic stratification of the patients and to allow early diagnosis and disease monitoring as well as to test treatment efficacy.

Within this multifaceted *scenario*, neurofilament light chain (NFL) is, at the present, the most promising candidate biomarker for an early identification of a general neurodegenerative process able to support disease diagnosis, prognosis, and progression, as well as monitoring an eventual disease-modifying treatment [5–7]. It is a component of neurofilaments (NFs) that, together with glial filaments, are the main types of intermediate filaments (IFs) of the nervous system [9–11]. Its physiological function is to confer mechanical stress resistance by preserving the characteristic cellular shape, intracellular traffic regulation between axons and dendrites, and, indirectly, nerve conduction speed modulation maintaining axon diameter [11]. Recent research suggests that they are also important for normal synaptic function [12]. Axonal dysfunction and degeneration are

✉ Filippo Baldacci
filippo.baldacci@unipi.it

¹ Department of Clinical and Experimental Medicine, University of Pisa, Pisa, Italy

² Centre National de Référence des Démences Rares ou Précoces, IM2A, Département de Neurologie, AP-HP - Hôpital Pitié-Salpêtrière, Paris, France

³ FrontLab, Institut du Cerveau et de la Moelle épinière (ICM), Paris, France

⁴ GRC n° 21, Alzheimer Precision Medicine (APM), AP-HP, Pitié-Salpêtrière Hospital, Sorbonne University, Boulevard de l'hôpital, Paris, France

⁵ INSERM U 1127, CNRS UMR 7225, Brain & Spine Institute (ICM), Boulevard de l'hôpital, Paris, France

⁶ Department of Neurology, Institute of Memory and Alzheimer's Disease (IM2A), Pitié-Salpêtrière Hospital, Paris, France

52 important steps in NDD pathogenesis, occurring long before
53 neuronal cell death and often preceding detectable deposition
54 of misfolded proteins [8]. During these processes, NFL is re-
55 leased into the extracellular space and, consequently, into body
56 fluids, such as the cerebrospinal fluid (CSF) and blood.

57 Broadly speaking, mounting data reported increased CSF
58 NFL levels in NDDs [5, 6, 13]. In the mid-1990s, through the
59 first enzyme-linked immunosorbent assay (ELISA) developed
60 for NFL, Rosengren and colleagues [14] demonstrated that
61 CSF NFL concentration was increased in amyotrophic lateral
62 sclerosis (ALS), Alzheimer's disease (AD), and vascular cog-
63 nitive impairment (VCI). Although with different magnitudes,
64 further studies revealed that CSF NFL increased also in other
65 several NDDs, such as frontotemporal dementia (FTD) [15,
66 16], Parkinson's disease (PD) and atypical parkinsonisms
67 (APs) [17, 18], Huntington's disease (HD) [19], mild cog-
68 nitive impairment (MCI) [20, 21], and Creutzfeldt–Jakob dis-
69 ease (CJD) [22, 23], as well as in non-primary neurodegener-
70 ative disorders, such as multiple sclerosis (MS) [24],
71 neuroinfectious conditions [25], traumatic brain injury [26],
72 acute stroke [27], and cerebrovascular diseases [28].
73 Moreover, in NDDs, CSF NFL levels showed to correlate
74 with poorer cognition, short survival times, brain atrophy,
75 and disease severity and progression [29–31], supporting the
76 notion that it could be useful not only as a diagnostic biomark-
77 er but also as a prognostic and progression biomarker [32]. As
78 a result, it has been proposed as a dynamic biomarker for
79 axonal degeneration [5, 6, 13] with the potential capacity to
80 monitor treatment effectiveness [10, 33].

81 In the last few years, the interest in NFL research shifted
82 toward blood. An ideal biomarker should be easily measur-
83 able, accurate, quantitative, reproducible, and employable to
84 exactly categorize the population in line with a certain disease
85 [34, 35]. To this end, blood-based biomarkers would have
86 significant advantages in time efficiency and cost efficiency
87 compared to CSF and neuroimaging [36, 37]. Moreover, they
88 would offer potential applications at the population level as
89 screening tools in primary care, as well as for longitudinal
90 evaluations with repeated sampling during follow-up. It is
91 not surprising that brain pathophysiological processes are
92 reflected into the periphery. However, CSF proteins partially
93 enters the blood flow, are subsequently diluted in a greater
94 volume compared with CSF, and go through biochemical in-
95 teractions with a large amount of plasma proteins. They are
96 also cleared by blood cells and metabolized by other tissues.
97 Finally, these processes overall hamper their measurement in
98 plasma or serum using traditional techniques. Nevertheless, in
99 the past few years, the development of analytical tools for
100 ultrasensitive quantification—the immunomagnetic reduction
101 (IMR) and the single molecule array (Simoa) techniques—by
102 allowing an efficient measurement of NFL in blood [38],
103 charted a tight correlation between CSF and blood NFL in
104 different NDDs [39]. Therefore, blood NFL was suggested

105 as a proxy of any neurodegenerative process, paving the
106 way to its use in clinical practice as a reliable risk biomarker
107 for neurodegeneration [40, 41]. Nonetheless, its potential ap-
108 plication in real life remains unclear [42].

109 Biomarker is defined as “a characteristic that is objectively
110 measured and evaluated as an indicator of normal biologic and
111 pathogenic processes, or pharmacologic responses to a thera-
112 peutic intervention” [43]. From a clinical perspective, a bio-
113 marker can be also classified in further categories with some
114 practical and conceptual overlaps: (1) antecedent biomarkers
115 identifying a risk of disease development (*risk biomarkers*),
116 (2) early biomarkers screening a subclinical condition (*screen-
117 ing biomarkers*), (3) biomarkers specifically recognizing a
118 full-blown clinical picture (*diagnostic biomarkers*), (4) bio-
119 markers categorizing disease severity (*staging biomarkers*),
120 (5) biomarkers predicting future disease course (*prognostic
121 biomarkers*), and (6) biomarkers predicting treatment re-
122 sponse (*predicting or monitoring biomarkers*) [44].
123 Accordingly, it is crucial to define the context of use of a
124 certain biomarker (primary care screening, diagnostic, risk of
125 progression, disease monitoring, stratification for clinical tri-
126 als, and pharmacodynamic or treatment response monitoring).

127 This review will attempt to summarize the current literature
128 on blood (plasma or serum) NFL in NDDs, trying to translate
129 research data in practical considerations, focusing on the con-
130 text of use of blood NFL as a biomarker in the framework of
131 the NDDs (Table 1).

132 Literature Research Methods

133 We conducted a systematic review of the literature until
134 February 2020, using the key terms “NFL,” “neurofilament
135 light chain,” and “neurofilament” to interrogate the PubMed
136 database for articles published in English evaluating blood
137 NFL concentrations (serum and plasma) in NDDs. Overall,
138 we identified 38 studies. The use of internationally accepted
139 clinical diagnostic criteria for each NDD, in particular AD
140 [45–49], ALS [50–52], dementia with Lewy bodies (DLB)
141 [53–55], FTD [56–58], PD [59–61], AP [62–65], and sporadic
142 Creutzfeldt–Jakob disease (sCJD) [66, 67], has been checked
143 out for any single study. The diagnostic performance of blood
144 NFL concentrations to correctly allocate the participants to the
145 different diagnostic groups was considered as follows: “excel-
146 lent” (area under the ROC curve (AUROC) 0.90–1.00),
147 “good” (AUROC 0.80–0.89), “fair” (AUROC 0.70–0.79),
148 “poor” (AUROC 0.60–0.69), or “fail” (i.e., no discriminatory
149 capacity) (AUROC 0.50–0.59) [68].

150 AD

151 AD is the most common form of dementia in the elderly,
152 accounting for 50–70% of prevalent neurodegenerative

t1.1 **Table 1** Overview on the
t1.2 possible context of use of blood
NFL as a biomarker in NDDs
t1.3

	Diagnostic value			Prognostic value	Monitoring treatment
	Preclinical phase	Prodromal phase	Full-blown picture		
t1.4 AD	±	+	+	+	±
t1.5 PD	±	±	+	+	±
t1.6 Atypical parkinsonisms (4R tauopathies)	±	±	+	+	±
t1.7 DLB	±	±	±	±	±
t1.8 FTD	–	±	+	±	±
t1.9 ALS	±	±	+	±	±
t1.10 CJD	±	+	+	–	±
t1.11 HD	–	–	–	+	±
t1.12 SMA	–	–	–	±	+
t1.13 Sporadic late-onset ataxias	±	±	±	+	±
t1.14 NDDs as a whole	±	+	+	+	±

Plus sign (+), potential use, supportive data are available; plus–minus sign (±), unknown; negative sign, negative evidences are available

AD Alzheimer’s disease, ALS amyotrophic lateral sclerosis, CJD Creutzfeldt–Jakob disease, DLB dementia with Lewy body, FTD frontotemporal dementia, HD Huntington’s disease, NDD neurodegenerative diseases, PD Parkinson’s disease, SMA spinal muscular atrophy

153 dementia cases with an enormous health and economic impact
154 [69]. The scientific and clinical research is nowadays shifting
155 from dementia to the prodromal or even preclinical phases of
156 AD to find effective therapeutic interventions that can delay or
157 halt neurodegenerative progression [70, 71].

158 Biomarkers hold promise for improving early diagnosis in
159 AD and establishing a tailored approach. The use of specific
160 surrogate biomarkers (neuroimaging, blood [plasma/serum],
161 and CSF) of AD pathology has been included in revised diag-
162 nostic criteria to distinguish AD from other forms of dementia
163 since its early disease stages. However, postmortem studies
164 demonstrate a high degree of neuropathologic heterogeneity
165 in patients who received a clinical diagnosis of AD [72]. The
166 pathogenesis of AD involves interacting pathophysiological
167 cascades in which the deposition of amyloid plaques (Aβ)
168 and the formation of neurofibrillary tangles (NFTs) composed
169 of hyperphosphorylated tau protein would represent only the
170 core events. The recently established “A/T/N” scheme pro-
171 poses three binary biomarker categories which reflect AD
172 pathophysiology, where “A” refers to Aβ pathology, “T” to
173 tau pathology, and N to neurodegeneration [73, 74]. However,
174 emerging evidence stresses the existence of additional molec-
175 ular pathophysiological pathways, such as synaptic dysfunc-
176 tion and degeneration, innate immune response and neuroin-
177 flammation, vascular and cell membrane dysregulation, brain
178 metabolic dysfunction, and axonal disruption [75]. The latter
179 is prominent in AD, and it is more closely related to cognitive
180 decline than Aβ pathology [76], thus leading to propose CSF
181 NFL as a non-specific biomarker to detect early AD patho-
182 physiological alterations [77]. In addition, an increased release
183 of NFL molecules is a consequence of aging that contributes
184 to an axonal degeneration due to subclinical cerebrovascular
185 changes and neuronal atrophy [78]. In this regard, a recent

prospective community-based study enrolling a cohort of cog- 186
nitively intact subjects reported high variability of serum NFL 187
levels above 60 years [79]. 188

189 Several studies showed that CSF NFL levels are elevated in
190 AD patients when compared with healthy controls (HCs) and
191 higher NFL concentration is predictive of a rapid disease pro-
192 gression along core biomarkers of AD pathology [21].
193 Peripheral serum or plasma NFL strongly correlated with
194 CSF NFL concentration, suggesting that it reflects the same
195 pathological process [80]. In general, we found that the levels
196 of NFL are higher in serum than in plasma, but the majority of
197 studies used plasma to quantify NFL.

NFL as Diagnostic Biomarker 198

199 Current evidence revealed that plasma NFL allows to discrim-
200 inate AD patients from HC subjects with a good/excellent
201 diagnostic accuracy [77, 81] (Table 2). In addition, plasma
202 NFL levels showed to be higher in the AD dementia group
203 than in the MCI group and in Aβ-positive MCI patients than
204 HC [77]. More recently, this finding has been replicated in a
205 larger study in the Dominantly Inherited Alzheimer Network
206 (DIAN) [82]. Other studies reported higher plasma/serum
207 NFL levels in AD and MCI patients compared with controls
208 [76, 83, 84], but with conflicting results about the differences
209 between MCI and cognitively normal individuals. A recent
210 meta-analysis by Wang and colleagues [85] confirmed these
211 findings, supporting a possible contribute of plasma NFL in
212 the AD diagnostic workup. Moreover, plasma NFL levels
213 could also reflect NFT pathology (as determined by
214 NFL immunostaining) and neurodegeneration at post-
215 mortem evaluation [76].

Table 2 The utility of NFL in the diagnostic and progression evaluation of neurodegenerative diseases

First author (type of study)	Controls	Patients	Diagnostic/prognostic value of blood NFL	Notes
t2.1 AD				
t2.2 Preische, 2019 (longitudinal study)	HC (AD mutation non-carriers) <i>n</i> = 162	AD presymptomatic mutation carriers, <i>n</i> = 243	- Rate of change of serum NFL in differentiating non-mutation carriers vs presymptomatic mutation carriers, AUROC = 0.7 - Non-mutation carriers vs symptomatic non-mutation carriers, AUROC = 0.89 - Baseline serum NFL value in differentiating non-mutation carriers vs presymptomatic mutation carriers, AUROC = 0.49 (at the cutoff value of 37.4 pg/mL) - Non-mutation carriers vs symptomatic non-mutation carriers, AUROC = 0.85 (at the cutoff value of 27.9 pg/mL) - Plasma NFL in differentiating patients vs HC, AUROC = 0.85 (95% CI 0.772–0.934), unadjusted for other variables - AD vs HC, AUROC = 0.92 (95% CI 0.869–0.970), adjusted for age - At the cutoff maximizing Youden index, 25.7 pg/mL, the unconditional accuracy was 0.82	- Mutation carriers were subdivided into 3 groups: presymptomatic mutation carriers (individuals who scored 0 on the CDR scale across all visits), converters (CDR = 0 at baseline and CDR > 0 at subsequent visits), and symptomatic mutation carriers (CDR > 0 across all visits)
t2.3 Lewczuk, 2018 (cross-sectional study)	HC = 41	MCI-AD = 25 ADD = 33 AD = 58		- MCI-AD were AD patients at the stage of MCI. - ADD were AD patients at the stage of early dementia. - AD included MCI-AD + AD patients
t2.4 Mattsson, 2017 (longitudinal study)	HC = 193	AD = 180 MCI = 197 pMCI = 109 sMCI = 65	Plasma NFL in differentiating AD vs CNC, AUROC = 0.87 (correcting for age, sex, educational level, and <i>APOE</i> ε4 genotype). When only correcting for age, sex, and educational level, the AUROC was reduced to 0.79	- Patients were recruited from the ADNI cohort. - MCI patients were divided into stable MCI (sMCI, with no progression to dementia during ≥ 2-year follow-up) and progressive MCI (pMCI, with conversion to dementia)
t2.5 FTD				
t2.6 van der Ende, 2019 (longitudinal study)	Non-carriers of a mutation in GRN, C9orf72, or MAPT = 127	Presymptomatic carriers of a mutation in GRN, C9orf72, or MAPT = 149 FTD (symptomatic carriers of a mutation in GRN, C9orf72, or MAPT) = 59	- Serum NFL in differentiating symptomatic mutation carriers vs presymptomatic mutation carriers, AUROC = 0.93 (95% CI 0.90–0.97), at the cutoff concentration of 17 pg/mL - Symptomatic mutation carriers vs non-mutation carriers, AUROC = 0.95 (95% CI 0.92–0.98), at the cutoff concentration of 17 pg/mL - Baseline serum NFL converters vs presymptomatic mutation carriers, AUROC = 0.93 (95% CI 0.89–0.98), at the cutoff level of 15.0 pg/mL	- Multicenter cohort study on families with genetic FTD in Europe and Canada - At least 2 serum samples were taken with a time interval of 6 months or more. - 9 presymptomatic carriers became symptomatic during follow-up (converters). - Presymptomatic carriers or non-carrier mutations were healthy relatives of first degree at risk of FTD
t2.7 Katisko, 2019 (longitudinal study)		FTD = 91 (bvFTD = 66; nfvPPA = 16; svPPA = 4; FTD-MND = 5) PPD = 34	- Serum NFL in differentiating FTD vs PPD, AUROC = 0.85 (95% CI 0.776–0.923), at the cutoff level of 19.9 pg/mL	- The PPD group included patients with a severe late-onset psychiatric disorder (psychotic, mood disorders, or both).

t2.10 **Table 2** (continued)

First author (type of study)	Controls	Patients	Diagnostic/prognostic value of blood NFL	Notes
t2.10 Al Shweiki, 2019 (cross-sectional study)	HC = 27	bvFTD = 20 Schizophrenia = 11 Depression = 28 Bipolar disorder = 11	- bvFTD vs PPD, AUROC = 0.82 (95% CI 0.732–0.908), at the cutoff level of 19.9 pg/mL - Serum NFL in differentiating bvFTD vs HC, AUROC = 0.94 (95% CI 0.87–0.99) - bvFTD vs depression, AUROC = 0.89 (95% CI 0.8–0.98), at a cutoff level above 35.7 pg/mL - bvFTD vs bipolar disorder, AUROC = 0.94 (95% CI 0.81–1.01), at a cutoff level > 26.5 pg/mL - bvFTD vs schizophrenia, AUROC = 0.9 (95% CI 0.77–1.03), at a cutoff level > 17.7 pg/mL - Serum NFL in differentiating bvFTD vs AD, AUROC = 0.67 - bvFTD vs MCI, AUROC = 0.9 (95% CI 0.84–0.97) - bvFTD vs HC, AUROC = 0.85 (95% CI 0.72–0.97) - bvFTD vs AD subgroups selected based on CSF Aβ42 levels, AUROC = 0.79, at a cutoff value of 33 pg/mL - bvFTD vs AD subgroups selected based on both CSF Aβ42 and tau/p-tau levels, AUROC = 0.77, at a cutoff value of 34.3 pg/mL	- In the FTLD group, 26 patients had a definite diagnosis due to the C9orf72 repeat expansion - 20 bvFTD consisting of 9 possible bvFTD, 5 probable bvFTD, and 6 genetic bvFTD (4 due to C9orf72 mutations, 2 MAPT mutations) - Serum NFL levels did not differ between psychiatric and control patients - Genetic screening of 67 patients with bvFTD revealed 7 carriers for C9orf72 repeat expansion. - Analysis of MAPT and GRN in 18 patients revealed 1 carrier of the MAPT mutation and 1 carrier of the GRN mutation
t2.11 Steinacker, 2018 (longitudinal study)	HC = 15	bvFTD = 74 AD = 26 (excluding patients with normal Aβ42) AD = 11 (without the typical AD CSF biomarker pattern) MCI = 17 Subgroups were defined on the basis of both CSF Aβ42 and tau levels: - 30 bvFTD (excluding patients with decreased Aβ42) - 31 bvFTD (excluding patients with decreased Aβ42 and increased tau or p-tau) PPA = 99 (nvPPA = 40, svPPA = 38, lvPPA = 21)	- Serum NFL in differentiating FTD vs HC, AUROC = 0.81 (95% CI 0.72–0.91) at an optimal cutoff value > 36 pg/mL - ALS vs HC, AUROC = 0.99 (95% CI 0.98–1.00) - Serum NFL in differentiating FTD vs HC, AUROC = 0.97 (95% CI 0.93–1.00) - FTD vs presymptomatic carriers, AUROC = 0.93 (95% CI 0.87–0.98), at the cutoff level of 18.0 pg/mL	- Patients with PPA met only clinical diagnostic criteria without knowledge of fluid biomarker concentration - Participants were recruited as part of GENFI or ascertained before participation in GENFI. - 4 subjects became symptomatic during follow-up (converters)
t2.12 Steinacker, 2017 (longitudinal study)	HC = 35			
t2.13 Wilke, 2016 (cross-sectional study)	HC = 46	FTD = 41 ALS = 25		
t2.14 Meeter, 2016 (longitudinal study)	HC = 71	FTD (caused by a pathogenic mutation in <i>GRN</i> , <i>MAPT</i> , or C9orf72) = 101		

Table 2 (continued)

First author (type of study)	Controls	Patients	Diagnostic/prognostic value of blood NFL	Notes
t2.15 ALS		Presymptomatic carriers of a pathogenic mutation = 62	- Presymptomatic carriers vs HC, AUROC = 0.63 (95% CI 0.51–0.75), at the cutoff level of 8.3 pg/mL - Serum NFL was also associated with survival (HR in the higher tertile 3.10, 95% CI 1.09–8.76)	
t2.16 Thouvenot, 2019 (longitudinal study)	HC = 21	ALS = 198	- Serum NFL > 15.0 pg/mL in discriminating ALS vs HC, AUROC = 0.99 (95% CI 0.972–0.9999) - Patients with serum NFL levels ≥ 71.2 pg/mL had a higher risk of death (HR 4.7, 95% CI 3.0–7.4) - A cutoff NFL level of 49 pg/mL in differentiating ALS vs non-neurodegenerative controls, AUROC = 0.97 (95% CI 0.95 to 0.991) - A cutoff value of 62 pg/mL in differentiating ALS vs disease controls, AUROC = 0.87 (95% CI 0.81 to 0.935) - A cutoff value of 62 pg/mL in discriminating ALS vs all other categories considered together, AUROC = 0.88 (95% CI 0.849 to 0.926)	- Patients were prospectively followed up to 18.5 months - Disease controls included in the differential diagnosis of ALS
t2.17 Verde, 2019 (longitudinal study)	Non-neurodegenerative controls = 50	ALS = 124 Disease controls = 44 FTD = 20 AD = 20 PD = 19 CJD = 6	- Patients with serum NFL above the median (125 pg/mL) had a shorter survival than patients with NFL ≤ 125 pg/mL, with an HR of 2.39 (95% CI 1.236 to 4.63) - Serum NFL in differentiating ALS vs ALS mimic, AUROC = 0.85 (95% CI 0.79–0.90), at an optimal cutoff value of 93 pg/mL - ALS fast progressor vs ALS slow progressors, AUROC = 0.87 (95% CI 0.76–0.94), at an optimal cutoff value of 159 pg/mL - ALS vs GBS + CIDP, AUROC = 0.58 (95% CI 0.51–0.64), at an optimal cutoff value of 139 pg/mL - ALS vs HSP, AUROC = 0.84 (95% CI 0.78–0.90), at an optimal cutoff value of 55 pg/mL - ALS vs PLS, AUROC = 0.89 (95% CI 0.83–0.93), at an optimal cutoff value of 88 pg/mL - ALS vs PMA, AUROC = 0.71 (95% CI 0.63–0.78), at an optimal cutoff value of 86 pg/mL - ALS patients with serum NFL in the upper tertile had a HR = 5.34 (95% CI 1.39–20.56) vs patients with serum NFL in the low tertile.	
t2.18 Gille, 2018 (longitudinal study)		ALS = 149 (C9orf72 = 15, FTD = 15, PLS = 11, PMA = 6) ALS mimic = 19 Disease controls = 82 (GBS, CIDP, HSP)	- A subset of 16 ALS repeated serum sample showing a relative stability of NFL concentrations over time	

t2.19 **Table 2** (continued)

First author (type of study)	Controls	Patients	Diagnostic/prognostic value of blood NFL	Notes
t2.19 Feneberg, 2018 (cross-sectional study)		ALS onset ≤ 6 months = 54 ALS onset > 6 months = 135 Other MNDs (PLS, SMA, Kennedy disease) = 21 ALS mimics = 27 Neurologic disease controls = 60	<ul style="list-style-type: none"> - ALS patients with serum NFL in the mid-tertile had a HR = 4.47 (95% CI 1.08–18.63) vs patients with serum NFL in the low tertile - Serum NFL in differentiating early ALS vs neurologic disease controls, AUROC = 0.92 (95% CI 0.85–0.99, cutoff = 128 pg/mL) - Early ALS vs MND mimics, AUROC = 0.99 (95% CI 0.97–1, cutoff = 97 pg/mL) - Late ALS vs neurologic disease controls, AUROC = 0.9 (95% CI 0.83–0.97, cutoff = 116 pg/mL) - Late ALS vs MND mimics, AUROC = 0.97 (95% CI 0.94–1, cutoff = 95 pg/mL) 	<ul style="list-style-type: none"> - A minimum 3-month follow-up - Serum NFL concentrations were not significantly different between early and later symptomatic phases
t2.20 Steinacker, 2016 (longitudinal study)	HC = 28	ALS = 125	High NFL concentrations were associated with shorter survival (at least a 6-month follow-up) [numbers are not reported]	<ul style="list-style-type: none"> - Patients were subgrouped according to the progression rate at first examination into slow, intermediate, and fast progressors. - Blood NFL measures were taken every 6 months up to 3 years of follow-up and were stable over time
t2.21 Lu, 2015 (longitudinal study)	HC = 78 (London cohort = 42; Oxford cohort = 36)	ALS = 167 (London cohort = 103; Oxford cohort = 64)	<ul style="list-style-type: none"> - Serum NFL in differentiating ALS vs HC, AUROC = 0.86, at a cutoff level of 36 pg/mL (Oxford cohort) - Plasma NFL in differentiating ALS vs HC, AUROC = 0.87, at a cutoff level of 36.2 pg/mL (London cohort) - Patients with serum NFL (Oxford cohort) in the highest tertile had a HR = 6.05 (95% CI 1.68–21.87). - Patients with plasma NFL (London cohort) in the highest tertile had a HR = 3.78 (95% CI 1.68–8.50). - Patients with serum NFL (Oxford cohort) in the middle tertile had a HR = 2.68 (95% CI 0.87–8.27). - Patients with plasma NFL (London cohort) in the middle tertile had a HR = 1.91 (95% CI 0.86–4.23). - Patients with combined blood NFL in the highest tertile had a HR = 3.82 (95% CI 1.98–7.39). - Patients with combined blood NFL in the middle tertile had a HR = 2.08 (95% CI 1.09–3.97) 	<ul style="list-style-type: none"> - Multicenter study (2 cohorts) - Authors also conducted analyses of the 2 cohorts combined, using the corresponding NFL data (serum or plasma) from each cohort, using cohort-specific tertile cutoff levels, and adjusting Cox regression and Kaplan–Meier survival analyses by center
t2.22 Gaiotimo 2013 (cross-sectional study)	HC = 67	ALS = 46 AD = 20 GBS = 19	Serum NFL in differentiating ALS vs HC, with sensitivity of 91.3% and specificity of 91% at a cutoff level of 26.6 pg/mL	<ul style="list-style-type: none"> - Neurological control patients consisted of patients suffered from tension-type headache ($n = 21$), lower back pain ($n = 7$), psychiatric

Table 2 (continued)

First author (type of study)	Controls	Patients	Diagnostic/prognostic value of blood NFL	Notes
t2.23 Parkinsonian syndromes and degenerative ataxias		Neurological control patients = 68		disorders ($n = 26$), or miscellaneous non-specific symptoms for which no neurological explanation could be found ($n = 14$)
t2.24 Oosterveld, 2020 (cross-sectional study)	HC = 52	PD = 139	Serum NFL in differentiating PD vs HC, AUROC = 0.64 (95% CI 0.55–0.73), at the cutoff value of 15.6 pg/mL	- The inclusion of serum NFL in a panel of CSF biomarkers (phosphorylated α -synuclein/total α -synuclein ratio and oligomeric α -synuclein/total α -synuclein ratio) yielded a sensitivity of 91% and a specificity of 81% (AUC 0.90, 95% CI 0.83–0.97)
t2.25 Ng, 2020 (longitudinal study)	HC = 50	PD = 149 (76 PD patients, 30 TD and 46 PIGD, had plasma NFL measured at the 2-year mark)	- Baseline plasma NFL in differentiating PD vs HC, AUROC = 0.83 (95% CI 0.77–0.89) - At year 2, plasma NFL in discriminating TD subtype vs PIGD subtype, AUROC = 0.65 (95% CI 0.53–0.77)	- Participants were included in the Early Parkinson's Disease Longitudinal Singapore Study which is an ongoing prospective cohort study analyzing the progression of early PD over a follow-up period of 5 years. - PD patients were classified into motor subtypes of "tremor dominant" (TD), "postural instability and gait disorders" (PIGD), or "indeterminate" based on MDS-UPDRS part II and III components
t2.26 Marques, 2019 (cross-sectional study)	HC = 53	PD = 55 AP (MSA = 22, PSP = 7) = 29	- Serum NFL in differentiating AP vs PD, AUROC = 0.91 (95% CI 0.83–0.98), at the cutoff value of 14.8 ng/L - AP vs HC, AUROC = 0.88 (95% CI 0.80–0.96), at the cutoff value of 13.6 ng/L	- Patients were recruited when the clinical diagnosis was still uncertain, definite clinical diagnosis was established after 3 years of follow-up and updated again after a maximum of 12 years of follow-up data
t2.27 Li, 2019 (cross-sectional study)	HC = 100 consisting of 2 cohorts: - Cohort A = 9 - Cohort B = 91	Manifest SCA-3 = 107, consisting of 2 cohorts: - Cohort A = 17 - Cohort B = 90 Preclinical SCA-3 = 26 (all included in the cohort B)	- Serum NFL in differentiating manifest SCA-3 vs HC, AUROC = 0.98 (95% CI 0.96–1.00), at the cutoff value of 20 pg/mL - Preclinical SCA-3 vs HC, AUROC = 0.83 (95% CI 0.72–0.95), at the cutoff value of 10 pg/mL	- Participants in cohort B were classified into 3 subgroups according to the SARA scores: 26 preclinical SCA-3 individuals, 46 stage 1 SCA-3 patients, and 44 stage 2 SCA-3 patients
t2.28 Lin, 2019 (longitudinal study)	HC = 40	PD = 116 MSA = 22	- Plasma NFL in differentiating MSA vs PD, AUROC = 0.80, at the cutoff value of 24.06 pg/mL - PD vs HC, AUROC = 0.75, at the cutoff value of 12.34 pg/mL - Higher baseline NFL levels were associated with a higher risk of motor symptom progression (adjusted HR 1.03, 95% CI 1.01–1.07) and cognition progression (adjusted HR 1.03, 95% CI 1.01–1.05)	- PD patients with baseline NFL levels > 21.84 pg/mL were at higher risk of motor symptom progression. - PD patients with baseline NFL levels > 18.34 pg/mL were at higher risk of cognitive decline

Table 2 (continued)

First author (type of study)	Controls	Patients	Diagnostic/prognostic value of blood NFL	Notes
t2.29 Donker Kaat, 2018 (longitudinal study)	HC = 95	PSP = 131	- Serum NFL in differentiating PSP vs HC, AUROC = 0.87 (95% CI 0.83–0.92), at the cutoff value of 38.3 pg/mL - Higher NFL levels were associated with reduced survival (adjusted HR 1.5, 95% CI 1.1–1.9)	- Diagnosis of PSP was made according to NINDS-SPSP criteria (2003–2014); 23 patients had pathological confirmation
t2.31 Wilke, 2018 (cross-sectional study)	HC = 45	c-MSA = 25 SAOA = 25 SCA = 20	- Serum NFL in differentiating c-MSA vs SAOA, AUROC = 0.74 (95% CI 0.59–0.89) - SCA vs HC, AUROC = 0.91 (95% CI 0.81–1.00)	- SCA patients consisted of SCA-1 (<i>n</i> = 6), SCA-2 (<i>n</i> = 3), SCA-3 (<i>n</i> = 8), and SCA-6 (<i>n</i> = 3)
t2.32 Hansson, 2017 (longitudinal study)	- Lund cohort HC = 53 - London cohort HC = 26	- Lund cohort PD = 171 - Lund cohort AP = 54 (MSA = 30, PSP = 19, CBD = 5) - London cohort PD = 20 - London cohort AP = 71 (MSA = 30, PSP = 29, CBD = 12) - Early-stage disease cohort PD = 53 - Early-stage disease cohort AP = 56 (MSA = 28, PSP = 22, CBD = 6)	- Blood NFL in differentiating PD vs AP, AUROC = 0.91 (95% CI 0.87–0.95) (Lund cohort) - PD vs AP, AUROC = 0.85 (95% CI 0.72–0.98) (London cohort) - PD vs AP, AUROC = 0.81 (95% CI 0.73–0.90) (early-stage disease cohort)	- The study included 2 independent prospective cohorts of PD and AP patients and HC: the Lund (<i>n</i> = 278) and London (<i>n</i> = 117) cohorts. - The third cohort consisted of PD and AP patients with a disease duration < 3 years (early-stage disease cohort, <i>n</i> = 109)
t2.33 HD				
t2.34 Byrne, 2018 (cross-sectional study)	HC = 20	HTT mutation carriers (premanifest HD = 20; manifest HD = 40) = 60	- Plasma NFL in differentiating HTT mutation carriers vs HC, AUROC = 0.91 (95% CI 0.85–0.97) - Plasma NFL in differentiating premanifest HD vs manifest HD, AUROC = 0.93 (95% CI 0.86–0.99)	
t2.35 Byrne, 2017 (longitudinal study)	HC = 97	HTT mutation carriers (premanifest HD = 104, manifest HD = 97) = 201 They were divided into: - Early premanifest HD = 58 - Late premanifest HD = 46 - Manifest HD stage 1 = 66 - Manifest HD stage 2 = 31	Higher baseline plasma NFL levels in premanifest HD were associated with clinical disease onset during the 3-year follow-up period, adjusted HR 3.03, 95% CI 1.07–8.60	- Participants were enrolled in the TRACK-HD* study. - At enrollment, participants with <i>HTT</i> CAG expansion mutations were classified as having premanifest or manifest HD based on the UHDRS-TMS. - 18 (17%) subjects with premanifest disease at baseline were newly diagnosed as having manifest HD during the 3 years of follow-up
t2.36 CJD				
t2.37 Staffaroni et al., 2019 (longitudinal study)		sCJD = 188	Higher baseline levels of plasma NFL levels were associated with shorter survival, HR 2.08 (95% CI 1.22–3.54)	Participants included patients with probable and definite (pathology-proven) sCJD with PRNP codon 129 polymorphism data available

Table 2 (continued)

First author (type of study)	Controls	Patients	Diagnostic/prognostic value of blood NFL	Notes
t2.39 Thompson, 2018 (cross-sectional study)	HC = 24	sCJD = 45	Serum NFL in differentiating sCJD vs HC, AUROC = 1.0	- The present study included 132 autopsy cases with rapidly progressive neurological syndromes.
t2.40 Kovacs, 2017 (cross-sectional study)	HC = 18	sCJD = 65 gCJD = 21 AD = 21 Other neurological pathologies = 25	- Plasma NFL in differentiating sCJD vs HC, AUROC = 0.99 (95% CI 0.98–1.0) - gCJD vs HC, AUROC = 1.0 (CI 1.0–1.0) - AD vs HC, AUROC = 0.99 (95% CI 0.97–1.0) - Other neurological disorders vs HC, AUROC = 0.96 (CI 0.90–1.0)	- Cases with a wide range of neuropathological alterations, including cerebrovascular disease, inflammation (meningoencephalitis), primary age-related tauopathy, non-AD tauopathies or Lewy body pathology
t2.41 Steinacker, 2016 (cross-sectional study)	HC = 40	sCJD = 33 gCJD = 9 GSS mutation carrier = 1 Demented controls (DCo) = 20	Serum NFL in differentiating sCJD + gCJD vs HC + DCo, AUROC = 0.95	- 39 CJD diagnoses were neuropathologically verified. - Patients with other diseases included AD (n = 12), MCI (n = 4), FTD (n = 3), and NPH (n = 1)

A β 42 amyloid β -peptide 1–42; AD Alzheimer's disease; ADNI Alzheimer's Disease Neuroimaging Initiative; ALS amyotrophic lateral sclerosis; AP atypical parkinsonism; APOE apolipoprotein E; AUROC area under the receiver operating curve; b ν FTD behavioral variant of frontotemporal dementia; c-MSA cerebellar variant MSA; CBD corticobasal degeneration; CDR cognitive dementia rating; CI confidential interval; CIDDP chronic inflammatory demyelinating polyneuropathy; CJD Creutzfeldt–Jakob disease; CSF cerebrospinal fluid; C9ORF72 chromosome 9 open reading frame 72; DLB dementia with Lewy body; FTD frontotemporal dementia; FTD–MND frontotemporal dementia with ALS; gCJD genetic Creutzfeldt–Jakob disease; GBS Guillain-Barre syndrome; GENFI genetic FTD initiative; GSS Gerstmann–Straussler–Scheinker disease; GRN progranulin; HC healthy controls; HD Huntington's disease; HR hazard ratio; HSP hereditary spastic paraplegia; HTT huntingtin; hPPA logopenic variant PPA; MAPT microtubule-associated protein tau; MCI mild cognitive impairment; MCI-AD MCI converters to AD; MDS-UPDRS Movement Disorder Society-Unified Parkinson's Disease Rating Scale; MND motor neuron disease; MSA multiple system atrophy; NFL neurofilament light chain; nfvPPA non-fluent variant PPA; NINDS-SPSP National Institute of Neurological Disorders and Stroke and the Society for PSP; NPH normal-pressure hydrocephalus; PD Parkinson's disease; PLS primary lateral sclerosis; PPA primary progressive aphasia; PMA primary muscular atrophy; PPD primary psychiatric disorders; PRNP prion protein; PSP progressive supranuclear palsy; p-tau phospho-tau; sCJD sporadic Creutzfeldt–Jakob disease; SAOA sporadic adult-onset ataxia; SARA Scale for the Assessment and Rating of Ataxia; SCA spinocerebellar ataxia; SMA spinal muscular atrophy; svPPA semantic variant PPA; UHDRS Unified Huntington's Disease Rating Scale–Total Motor Score

*TRACK-HD is a multinational prospective observational study of HD that examines clinical and biological findings of disease progression in individuals with premanifest HD and early-stage HD

216 **NFL as Staging and Prognostic Biomarker**

217 Studies in AD and MCI-AD patients found a correlation
 218 between plasma NFL concentration and cognitive impair-
 219 ment, MRI hippocampal volume loss and brain atrophy,
 220 and cerebral ¹⁸F-FDG-PET hypometabolism [76, 81–87].
 221 Moreover, higher plasma NFL levels predicted faster cog-
 222 nitive deterioration and a higher rate of brain atrophy and
 223 hypometabolism in MCI patients over time [77]. Baseline
 224 plasma and CSF NFL levels were similarly associated with
 225 short-term declines in imaging measures of neurodegenera-
 226 tion and with global cognitive worsening, but not with
 227 change in amyloid ligand retention on PET [35], differently
 228 from CSF t-tau concentration that critically depends on ce-
 229 rebral Aβ burden [88]. Instead, increased plasma NFL was
 230 related to baseline and longitudinal glucose hypometabolism,
 231 which is an unspecific neurodegeneration marker, in AD-
 232 related regions of MCI Aβ+ individuals [87].

233 In a longitudinal analysis of NFL plasma levels in a
 234 large cohort of subjects enrolled in the Alzheimer's
 235 Disease Neuroimaging Initiative (ADNI), Mattsson and
 236 colleagues [82] found increasing rates of NFL changes
 237 from preclinical AD stage to frank AD dementia through
 238 prodromal phase, suggesting NFL as a dynamic biomarker
 239 sensitive to AD disease progression. Of note, longitudinal
 240 NFL variations correlated with several baseline AD-related
 241 features (CSF biomarkers, imaging measures, and cog-
 242 nition) in the whole population, though with significant dif-
 243 ferences regarding clinical stage. Actually, the strictest as-
 244 sociations were reported in MCI participants. Longitudinal
 245 NFL level was generally increased in patients who were
 246 classified as N+ (using temporal brain atrophy as N- indi-
 247 cator) and in those who were only T+. Therefore, NFL
 248 might reflect a neurodegenerative process that occurred
 249 independently from Aβ pathology. Noteworthy, the NFL
 250 rate of change, rather than NFL absolute concentration,
 251 was subject to a significant increase in mutation carriers
 252 compared with non-carriers. Moreover, the NFL rate of
 253 change strongly correlated with longitudinal precuneus
 254 cortical thinning in both symptomatic and presymptomatic
 255 mutation carriers [89].

256 **NFL as Risk/Screening Biomarker**

257 Blood NFL levels seem to predict the progression to AD de-
 258 mentia in patients with subjective memory complaints [77].
 259 An association between regional hypometabolism in the right
 260 hippocampus and higher plasma NFL levels was reported in
 261 cognitively normal participants from the ADNI database [87].
 262 Hu and colleagues [90] explored the predictive role to develop
 263 AD of plasma NFL at the preclinical stage. Interestingly, plas-
 264 ma NFL concentrations were already abnormally high in cog-
 265 nitively normal individuals with significant Aβ-related

pathological changes. Baseline plasma NFL levels did not
 differ in normal elderly volunteers who remain cognitively
 intact during the follow-up, independently from an initial amy-
 loid PET positivity status. Instead, a trend toward elevated
 plasma NFL concentration was observed in Aβ+ individuals
 with subjective memory complaints compared to subjects
 without memory complaints who were Aβ+, and plasma
 NFL resulted to be inversely associated with cognitive perfor-
 mance [91]. By contrast, other groups investigated the corre-
 lation between serum NFL levels with cerebral metabolism in
 MCI patients. Regional hypometabolism in bilateral
 parahippocampal gyri, right fusiform, and middle temporal
 gyri was independently predicted by plasma NFL [92].

Weston and colleagues [93] reported increased serum NFL
 concentrations also in symptomatic and presymptomatic fam-
 ilial AD (FAD) mutation carriers, showing a significant cor-
 relation with the estimated years to/from symptom onset
 across all mutation carriers as well as with cognitive decline
 and MRI atrophy. This finding suggests that increases in se-
 rum NFL precede the onset of AD symptoms. A large study in
 the DIAN cohort confirmed NFL as a sensitive marker of
 early neurodegeneration, finding significant increased serum
 NFL levels in AD mutation carriers (Aβ precursor protein
 (APP) or presenilin 1 (PSEN1) or presenilin 2 (PSEN2))
 16 years before disease onset [89]. The rate of change of
 serum NFL peaked in mutation carriers during the conversion
 phase to clinically evident cognitive impairment and reached a
 plateau in symptomatic carriers; absolute values of NFL
 showed a trend toward a slow increase over time (Table 2).
 Interestingly, the increase in plasma NFL concentration dur-
 ing the follow-up (15–30 months) in 79 elderly participants
 without dementia, including 15 subjects with MCI, was asso-
 ciated with a significant decline in both attention and global
 cognition and with an increase in cerebral amyloid PET up-
 take [32]. More recently, baseline serum NFL was shown to
 be a strong and independent predictor of brain volume loss
 and subtle cognitive changes in a longitudinal study cohort of
 neurologically intact individuals [79].

NFL as Predictive Biomarker

Although not yet in humans, transgenic mice models treated
 with a β-secretase (BACE) inhibitor showed beneficial effects
 on AD-relevant downstream markers, including reduced plas-
 ma NFL concentrations [30].

FTD

The term FTD indicates a heterogeneous spectrum of NDDs
 inexorably conveying to a dementia syndrome characterized by
 predominant behavioral—behavioral FTD (bvFTD) [94]—or
 language—primary progressive aphasia (PPA) [58]—

314 impairment. It is the third most common neurodegenerative
 315 dementia after AD and DLB and is typically diagnosed in mid-
 316 dle age [95]. bvFTD is the most prevalent phenotype (55–60%
 317 of cases) whereas PPA (40–45% of cases) can be further clas-
 318 sified as non-fluent/agrammatic variant (nfvFTD) and semantic
 319 variant (svFTD) [96]. Notably, an additional clinical PPA var-
 320 iant, named logopenic variant, can show AD pathological fea-
 321 tures in more than half of subjects [97]. Finally, a clinical over-
 322 lap between FTD and ALS is described and about 10–15% of
 323 cases with ALS report a dementia syndrome in the FTD spec-
 324 trum (ALS-FTD) [98]. The underlying pathologies in FTD,
 325 essentially abnormal accumulations of either tau or TAR
 326 DNA-binding protein 43 (TDP-43) proteins, may be recog-
 327 nized only on postmortem examination. These misfolded pro-
 328 tein aggregates lead to atrophy (structural MRI) and
 329 hypometabolism (¹⁸F-FDG-PET) of frontal and/or anterior tem-
 330 poral lobes depending on phenotype [41].

331 Unfortunately, with the exception of causative mutations in
 332 genetic forms (about 10% of cases: hexanucleotide repeat ex-
 333 pansion near the chromosome 9 open reading frame gene
 334 (C9orf72), progranulin (GRN), and microtubule-associated
 335 protein tau (MAPT) [99]), CSF poly(GP) detection in
 336 C9ORF72 expansion carriers, and decreased CSF/blood
 337 progranulin levels in GRN mutation carriers, specific patho-
 338 physiological biomarkers for FTD are completely lacking.
 339 However, the diagnosis of FTD remains very challenging,
 340 especially for behavioral variant as its diagnosis is mainly
 341 based on clinical assessment and because its symptoms show
 342 a significant overlap with primary psychiatric disorders [100].

343 In this context, blood NFL is a promising candidate bio-
 344 marker for FTD, especially for disease differential diagnosis,
 345 monitoring, and prognosis [101, 102]. Recent evidence
 346 proved that blood and CSF levels of NFL are tightly related
 347 and significantly higher in FTD subjects than in HC [103],
 348 without gender differences [104]. Furthermore, although
 349 NFL levels generally increase with aging, this association
 350 seems not present for FTD patients [16, 105–107]. Blood
 351 NFL levels tightly correlate with CSF values in almost all
 352 studies measuring this biomarker in both fluids, thus suggest-
 353 ing that the peripheral concentrations of NFL substantially
 354 reflect the pathophysiological modification within CNS lead-
 355 ing to NFL increase in CSF.

356 **NFL as Diagnostic Biomarker**

357 Serum NFL levels distinguished FTD patients from controls
 358 with good/optimal diagnostic accuracy (Table 2) [105,
 359 108–110]. Interestingly, this biomarker was higher in bvFTD
 360 individuals in comparison with psychiatric patients affected by
 361 depression, schizophrenia, and bipolar disorders [106], in
 362 which bvFTD misdiagnosis is common [100]. The diagnostic
 363 accuracy to differentiate bvFTD from psychiatric disorders was

above 80% (Table 2), independently from the specific psychi- 364
 atric condition (mood or psychotic disorders) [106, 107]. 365

366 Only one study compared both FTD and AD subjects,
 367 reporting that serum NFL levels were higher in bvFTD patients
 368 [109], and separated the two groups with a sensitivity and spec-
 369 ificity of 93% and 61%, respectively (Table 2), after a prelim-
 370 inary exclusion of bvFTD patients with an AD biomarker pro-
 371 file and clinical AD subjects without a core biomarker confir-
 372 mation. Moreover, serum NFL concentrations seemed to be
 373 higher in nfvPPA and svPPA than lvPPA subjects [105],
 374 though with only moderate accuracy. A further study showed
 375 no significant differences between PPA subtypes [110].

376 Finally, although FTD individuals had higher serum NFL
 377 levels than subjects with other cognitive disorders such as AD,
 378 ALS patients reported even more elevated concentration
 379 [108]. Notably, ALS subjects present a TDP-43 pathology in
 380 about 95% of cases, suggesting a potential association be-
 381 tween increased NFL concentration and TDP-43 pathology.
 382 Further confirmation of this hypothesis is supported by ob-
 383 serving that FTD forms expected to be TDP-43 positive
 384 (C9ORF72 and GRN mutation carriers, or svPPA phenotype)
 385 reported higher concentration of this biomarker than FTD
 386 subtypes expected to be tau positive (MAPT mutation carriers
 387 and nfvPPA) [16, 99, 104].

388 **NFL as Staging and Prognostic Biomarker**

389 In different studies, serum NFL increases overtime in FTD
 390 subjects, independently from the phenotypes, with the excep-
 391 tion of lvPPA [105, 109]. A longitudinal Mini-Mental State
 392 Examination (MMSE) decline was correlated with baseline
 393 serum NFL levels [107]. Disease duration was not associated
 394 with NFL concentration although one study reported a poor
 395 survival in FTD subjects in the higher tertile of serum NFL
 396 levels (Table 2).

397 Additionally, peripheral blood NFL concentrations appeared
 398 to reflect specific regional brain atrophy related to clinical phe-
 399 notypes (PPAs or bvFTD). In bvFTD patients, serum NFL was
 400 associated with a low cognitive score and a reduction in whole-
 401 brain volume and was correlated with brain atrophy, including
 402 frontal and subcortical regions [99, 104, 109, 110].

403 Serum NFL was correlated with baseline cognitive impair-
 404 ment, cognitive decline overtime, and atrophy progression of
 405 the left frontal lobe and the right middle frontal gyrus in PPA
 406 individuals and nfvPPA/svPPA subjects, respectively [105,
 407 107]. On the other hand, although only in one study, serum
 408 NFL concentration was not related to specific hypometabolic
 409 regions on ¹⁸F-FDG-PET in PPA subjects [110].

410 **NFL as Risk/Screening Biomarker**

411 Noteworthy, serum NFL was higher in genetic FTD with a
 412 full-blown clinical picture but not in presymptomatic carriers

413 compared to HC, independently from the gene mutation.
 414 Moreover, it was consistently higher in converters than in
 415 non-converter carriers [104] and increased overtime in con-
 416 verters but not in symptomatic FTD as well as non-converters.
 417 Therefore, serum NFL could differentiate genetic FTD pa-
 418 tients from presymptomatic carriers, with an excellent diag-
 419 nostic accuracy (Table 2) [16]. By contrast, the discriminatory
 420 accuracy of the biomarker relative to presymptomatic genetic
 421 carriers and controls resulted quite poor (Table 2).

422 ALS

423 ALS is a progressive neurological disease in which upper
 424 motor neuron (UMN) and lower motor neuron (LMN) degener-
 425 erate, leading to paralysis and death, typically within 3–
 426 5 years from symptom onset. To date, there is no definitive
 427 diagnostic test for ALS, and confirmation of diagnosis is
 428 based on clinical findings, electromyography results, and ex-
 429 clusion of mimics [52]. Despite efforts to increase the sensi-
 430 tivity of diagnostic criteria, often the diagnosis is made only
 431 after the onset of symptoms for both sporadic and familial
 432 ALS [111, 112]. An early diagnosis would be paramount,
 433 since it was observed that the benefit of riluzole is related to
 434 its early administration.

435 A large body of research exists on neurochemical ALS
 436 biomarkers [113, 114], among which phosphorylated neuro-
 437 filament heavy chain (pNFH) and NFL have been postulated
 438 as the most interesting candidates [115, 116]. This is not sur-
 439 prising, given the axonal impairment that characterizes the
 440 disease already at the early stage, with a large release of NF
 441 in CSF [117]. Additionally, previous studies indicate cyto-
 442 skeletal proteins as one of the key factors contributing to neu-
 443 rodegeneration in ALS [118–123]. Other evidences provide
 444 additional support that NFL aggregation is an early event in
 445 motor neuron disease [124], and that NFL is involved in the
 446 aggregation and neurotoxicity of other proteins in motor neu-
 447 rons [125]. At the same time, elevated NFL levels in ALS may
 448 be explained by the higher content of axonal proteins in motor
 449 neurons compared to other neuronal populations [126].
 450 Nonetheless, several data highlighted that NFL is able to dis-
 451 criminate ALS patients from healthy and disease controls [14,
 452 117, 127, 128], especially in cases with predominant UMN
 453 signs, and correlates with clinical disability [129], disease
 454 stage, progression, and/or prognosis [130, 131], probably
 455 reflecting the burden of motor neuron degeneration.
 456 Although CSF NFL remains the more robust fluid biomarker
 457 for ALS because of its directly reflecting alterations in the
 458 CNS, a high correlation between CSF and blood NFL concen-
 459 trations has been reported [132]. Moreover, at odds with other
 460 neurodegenerative diseases such AD, NFL concentrations do
 461 not correlate with age in ALS individuals [133–135].

NFL as Diagnostic Biomarker

462 Mounting evidence reports significantly higher blood NFL 463
 464 levels in ALS patients when compared to controls (Table 2) 464
 465 [40, 135–138]. The diagnostic performance of serum NFL in 465
 466 discriminating ALS and non-neurodegenerative subjects 466
 467 showed excellent sensitivity and specificity (Table 2) [139, 467
 468 140]. These findings led authors to propose the introduction 468
 469 of serum NFL measurement into clinical practice as support- 469
 470 ive diagnostic tool. In addition, serum NFL showed signifi- 470
 471 cantly elevated concentration in ALS even at the onset of the 471
 472 first symptoms, confirming its potential role as a biomarker 472
 473 for early detection of symptomatic sporadic ALS. In this re- 473
 474 gard, serum NFL concentrations demonstrated optimal sensi- 474
 475 tivity and specificity also in distinguishing early symptomatic 475
 476 ALS from other neurologic diseases or motor neuron disease 476
 477 mimics, independently whether diagnosis was definite, prob- 477
 478 able, or possible, following the El Escorial criteria (Table 2) 478
 479 [134]. Interestingly, Gille and colleagues [135] reported an 479
 480 increase of serum NFL as a function of the number of regions 480
 481 (i.e., cranial, cervical) affected by UMN degeneration. 481
 482 Accordingly, in a MRI-based study, elevated CSF and serum 482
 483 NFL concentrations were significantly associated with lower 483
 484 diffusion tensor imaging (DTI) fractional anisotropy and in- 484
 485 creased radial diffusivity in the corticospinal tract of ALS 485
 486 patients, as well as with clinically UMN score burden [133]. 486
 487 On the other hand, previous studies have shown that NFL 487
 488 levels were not increased in Kennedy disease and spinal mus- 488
 489 cular atrophy (strictly LMN diseases) [128, 134]. As a conse- 489
 490 quence, a subclinical involvement of the UMN is likely in 490
 491 ALS patients with isolated LMN symptoms and elevated se- 491
 492 rum NFL concentration [117]. However, the neuroanatomical 492
 493 correlate of NFL increase is not yet clear since Verde and 493
 494 colleagues [139] showed a lack of association with DTI- 494
 495 MRI measurements of the integrity of cerebral white matter 495
 496 tracts in the brain of ALS patients. Finally, serum NFL levels 496
 497 were reported relatively lower in patients with primary lateral 497
 498 sclerosis (PLS) and hereditary spastic paraplegia (HSP), two 498
 499 UMN-isolated syndromes, compared with ALS subjects, sug- 499
 500 gesting in such patients different pathophysiological processes 500
 501 and rates of neurodegenerative diseases [134, 135]. 501Q4

NFL as Staging and Prognostic Biomarker

502 Several studies reported that blood NFL levels correlate with 503
 504 disease severity parameters, such as the decline in the ALS 504
 505 Functional Rating Scale-Revised (ALSFRS-R) score and the 505
 506 ALS Milano-Torino Staging (MITOS) system score [135, 506
 507 137, 140, 141]. 507
 508 Furthermore, serum NFL levels at recruitment or at the 508
 509 time of diagnosis predicted survival independently from other 509
 510 clinical variables and were negatively associated with disease 510
 511 duration (Table 2) [136, 138, 142]. Thouvenot and colleagues 511

512 [140] evaluated the largest series ever of serum samples taken
 513 from ALS patients, finding that NFL concentration was the
 514 most important parameter related to ALS survival in multivar-
 515 iate models (Table 2). Likewise, serum NFL concentrations in
 516 the middle and high tertile were associated with an increased
 517 HR compared with those of patients in the lowest tertile
 518 (Table 2) [135]. Interestingly, unlike pNFH, NFL levels seem
 519 to change minimally throughout the course of the disease,
 520 maintaining distinct temporal profiles from controls, and a
 521 steady trajectory [136].

522 However, not all studies confirmed blood NFL as a robust
 523 prognostic biomarker in ALS patients, even if all proved low-
 524 er NFL concentration in slow disease progressors [141].
 525 Notably, the ALSFRS-R and the ALS MITOS system better
 526 correlated with CSF than serum at the baseline [137]. No
 527 significant correlation has been found between blood NFL
 528 levels and cognitive dysfunction in ALS [135].

529 NFL as Risk/Screening Biomarker

530 In contrast with AD, CSF and blood NFL levels are reported
 531 normal in presymptomatic ALS mutation carriers (C9orf72,
 532 SOD1, FUS/TLS, or TARDBP), but they increase suddenly
 533 with symptom onset in symptomatic mutation carriers as dem-
 534 onstrated by Weydt and colleagues [143]. Furthermore, using
 535 the parental age of disease onset as a proxy for assumed age of
 536 clinical onset, any trend toward an increase of NFL concen-
 537 tration was observed in asymptomatic mutation carriers.
 538 Despite recent longitudinal data on a large cohort of presymp-
 539 tomatic, SOD1 mutation carriers provided evidence that an
 540 increase in CSF and blood NFL levels occurs at least 1 year
 541 before of any clinical manifestations of the disease [144].

542 NFL as Predictive Biomarker

543 It would be interesting to determine whether riluzole reduces
 544 blood NFL levels over time given its neuroprotective effects
 545 and, although minimally, its capacity to slow disease progres-
 546 sion. Currently, no studies investigated blood NFL as an indi-
 547 cator of treatment response in ALS, and no difference in blood
 548 NFL levels between patients treated and not treated with
 549 riluzole has been reported so far [136, 139]. On the other hand,
 550 recent studies on spinal muscular atrophy (SMA), a group of
 551 severe autosomal recessively inherited neurodegenerative dis-
 552 orders characterized by degeneration of the spinal alpha motor
 553 neurons, have highlighted an emerging role of NFL in track-
 554 ing disease progression and response to treatment. Of note,
 555 recent data provided evidence that CSF NFL levels normalize
 556 and correlate with motor improvement in children with SMA
 557 treated with nusinersen, with a greatest benefit found in chil-
 558 dren who received treatment earliest during the course of dis-
 559 ease [145]. Nusinersen, an antisense oligonucleotide delivered
 560 intrathecally by a spinal tap, is the first drug clinically

approved for the treatment of all SMA types, with a rather 561
 dramatic impact on phenotype [146]. The levels of two addi- 562
 tional biomarkers of neurodegeneration (CSF tau and glial 563
 fibrillary acidic protein (GFAP), an intermediate filament 564
 present in astrocytes) decreased together with CSF NFL after 565
 nusinersen administration, indicating that the neuronal and 566
 astroglia damage can be restored by nusinersen treatment 567
 [145]. Moreover, the decrease of NFL concentration was 568
 much larger than that of tau and GFAP, suggesting NFL as 569
 an early treatment response biomarker in SMA patients, help- 570
 ful to select those patients will benefit to continue such an 571
 invasive treatment. Further studies with a long follow-up are 572
 needed, but these preliminary results in SMA indicated NFL 573
 as a promising marker for upcoming disease-modifying ther- 574
 apies in diseases beside SMA. Conversely, it is worth men- 575
 tioning that the diagnostic and monitoring value of NFL in 576
 CSF and blood has not been confirmed in adolescent and adult 577
 SMA-type (SMA types 2 and 3) patients treated with 578
 nusinersen [134, 147]. It was hypothesized that NFL release 579
 is lower in late-onset SMA than in the infantile-onset subtype. 580
 Actually, the first phenotype is characterized by a long-lasting 581
 and chronic disease course while the foster by an acute and 582
 highly aggressive onset. Additionally, subjects with infantile- 583
 onset SMA report a significantly better response to nusinersen 584
 therapy when compared to individuals with adult-onset one. 585
 In this regard, the finding of normal blood levels of NFL in 586
 SMA could be used in a diagnostic panel of biochemical 587
 markers to help differentiate patients presenting with motor 588
 neuron deficits, separating SMA from ALS. Indeed, a substan- 589
 tial proportion of patients with SMA initially receive a diag- 590
 nosis of ALS [148]. 591

592 Degenerative Parkinsonisms

593 PD is the most common degenerative parkinsonism, with ev- 593
 idence of progressive loss of dopaminergic neurons in the pars 594
 compacta of the substantia nigra. Diagnostic criteria have been 595
 recently revised to improve diagnostic accuracy imaging bio- 596
 markers as supportive features [61]. However, early diagnosis 597
 and progression prediction remain challenging for physicians. 598
 Of note, the differential diagnosis between PD and AP can be 599
 difficult, mainly at the early clinical stage. Similarly, evolution 600
 of diagnostic criteria for AP improved accuracy, but misdiag- 601
 nosis rates are still high [149–152]. The APs that most com- 602
 monly mimic PD are progressive supranuclear palsy (PSP) 603
 and multiple system atrophy (MSA), whereas among APs, 604
 the lowest diagnostic accuracy regards the corticobasal degen- 605
 eration (CBD). 606

607 Currently, biochemical biomarkers for PD and AP are an 607
 unmet need, but many CSF/serum molecules are under eval- 608
 uation. CSF NFL concentration overlaps in patients with PD, 609
 PD with dementia (PDD), and DLB and are comparable with 610

611 those in HC [31]. In contrast, it has been demonstrated that
 612 CSF NFL levels are markedly increased in AP patients.
 613 Accordingly, it might discriminate between PD and AP with
 614 a high degree of diagnostic accuracy [17, 18, 153, 154]. This
 615 is in line with the remarkable axonal degeneration of large
 616 myelinated axons occurring in AP as well as with the rapid
 617 neuronal loss in such conditions [155]. Furthermore, CSF
 618 NFL concentration correlates with measures of disease sever-
 619 ity and other clinical variables, demonstrating its capability to
 620 reflect neurodegenerative mechanisms. However, to over-
 621 come the well-known limits related to CSF examination,
 622 blood-derived NFL would be a more favorable biomarker.
 623 In this regard, the strong correlation between blood and CSF
 624 NFL levels in parkinsonian syndromes holds potential for an
 625 application in clinical practice [156].

626 **PD**

627 **NFL as Diagnostic Biomarker**

628 Similar to CSF results, NFL concentration in serum/plasma is
 629 considered useful for a differential diagnosis between PD and
 630 AP [157]. This has been tested for the first time in three inde-
 631 pendent prospective cohorts of PD, PSP, MSA, and CBD pa-
 632 tients, compared with HC. Blood NFL levels in AP were sig-
 633 nificantly elevated compared with those in PD, showing a di-
 634 agnostic accuracy ranging from good (in the early cohort with
 635 disease duration < 3 years) to excellent (in the Lund cohort)
 636 (Table 2) [156]. Conversely, blood NFL levels were not able
 637 to accurately separate PD from HC. A subsequent study in
 638 subjects with an uncertain diagnosis at the time of inclusion
 639 confirmed similar results (Table 2) [158]. Nevertheless, recent
 640 studies support the promise of plasma NFL as a diagnostic
 641 biomarker also in PD, demonstrating relatively higher NFL
 642 levels in cases vs controls [159] and a good diagnostic accuracy
 643 in differentiating PD patients from HC (Table 2) [160].
 644 Moreover, higher serum NFL levels were found even at early
 645 stages of the disease and in participants at risk of disease pro-
 646 gression (prodromal PD and symptomatic and asymptomatic
 647 mutation carriers of known PD genetic mutations), indicating
 648 the presence of active disease and potential for conversion to
 649 either PD or parkinsonian syndromes [161].

650 **NFL as Staging and Prognostic Biomarker**

651 Heterogeneous results regarding possible correlations be-
 652 tween blood NFL and PD clinical features in three indepen-
 653 dent PD cohorts have been provided in the prospective and
 654 longitudinal study of Hansson and colleagues [156]. In gen-
 655 eral, higher blood NFL levels were observed in more ad-
 656 vanced PD and, in the Lund PD cohort, a higher blood NFL
 657 concentration was associated with disease duration and more

severe motor symptoms (measured as Hoehn and Yahr 658
 (H&Y) stage, Unified Parkinson's Disease Rating Scale 659
 (UPDRS) III motor score, Timed Up and Go Test, and 660
 Tandem Gait Test). Conversely, no clinical correlations were 661
 described in the London cohort and the early-stage disease 662
 cohort [156]. However, further studies confirmed the positive 663
 relation between plasma NFL levels and motor symptom se- 664
 verity (measured as H&Y stage and UPDRS part III score) 665
 and proved a significant correlation between plasma NFL con- 666
 centration and cognitive dysfunction at MMSE [161, 162]. In 667
 another study, PDD patients reported higher plasma NFL level 668
 compared with PD subjects without dementia [84], supporting 669
 an association between plasma NFL and cognitive function in 670
 PD patients [84]. Furthermore, higher baseline plasma NFL 671
 concentrations in PD patients were found to be longitudinally 672
 associated with a higher risk of progression for both motor and 673
 cognitive symptoms, suggesting that serum NFL may be a 674
 biomarker of clinical progression in PD (Table 2) [160–162]. 675

PSP 676

NFL as Diagnostic Biomarker 677

Two other studies confirmed the diagnostic value of blood 678
 NFL in PSP patients showing good capability to discriminate 679
 between PSP and HC (Table 2) [163, 164]. In contrast, blood 680
 NFL is not suitable to separate PSP from other forms of AP 681
 [156] and similar levels are reported in patients with MSA and 682
 PSP [158]. 683

NFL as Staging and Prognostic Biomarker 684

Greater baseline NFL levels in serum/plasma seem to correlate 685
 with disease severity and clinical progression in PSP patients, 686
 though with conflicting results. Such heterogeneity may re- 687
 flect differences in study design, since PSP patients have been 688
 evaluated as a separate group in some studies but not in others 689
 where PSP, MSA, and CBD patients have been combined as a 690
 whole group. Specifically, blood NFL levels positively corre- 691
 lated with motor symptom severity, evaluated as H&Y stage 692
 and with UPDRS III motor score, but not with disease dura- 693
 tion or other clinical assessments, in the AP group (including 694
 also MSA and CBD patients) [156]. Similarly, serum NFL 695
 concentration at baseline correlated with motor performances, 696
 measured with the International Cooperative Ataxia Rating 697
 Scale score and Tandem Gait Test in another cohort of AP 698
 patients including PSP [158]. 699

In studies focusing exclusively on PSP patients, higher 700
 serum NFL levels were related to more severe motor, func- 701
 tional, and cognitive disability as well as shorter survival but 702
 not with age at symptom onset or disease duration (Table 2) 703
 [163]. Notably, NFL levels in the higher tertile were 704

705 associated with worse survival (Table 2) [164]. Higher base-
 706 line plasma NFL levels also predicted greater whole-brain
 707 and superior cerebellar peduncle volume loss at 1-year fol-
 708 low-up [163].

709 **MSA and Degenerative Ataxias**

710 **NFL as Diagnostic Biomarker**

711 As for PSP, studies indicate elevated blood NFL concentration
 712 in MSA patients, suggesting its use in discrimination of MSA
 713 from PD and HC with a good diagnostic accuracy (Table 2).
 714 However, as aforementioned, NFL cannot discriminate
 715 among APs [158].

716 Moreover, blood NFL was proposed to improve the differ-
 717 ential diagnosis of degenerative ataxias. In a pilot study eval-
 718 uating serum NFL levels in patients with a clinical diagnosis
 719 of probable cerebellar-MSA (c-MSA) subtype, sporadic adult-
 720 onset ataxia (SAOA), and frequent repeat-expansion
 721 spinocerebellar ataxias (SCAs 1, 2, 3, and 6) and in HCs,
 722 serum NFL concentration was found to be higher in SCA
 723 patients and in the c-MSA group compared with controls.
 724 This is probably the result of the diffuse involvement of
 725 spinocerebellar and corticospinal tracts in these multisystemic
 726 neurodegenerative ataxias [165]. However, the performance
 727 of serum NFL differentiating c-MSA from SCAOA was only
 728 moderate (Table 2), in contrast with a higher accuracy previ-
 729 ously reported for CSF NFL (AUC = 0.93) [166]. NFL levels
 730 were significantly lower in SCAOA and comparable with those
 731 of HC. A further study investigated serum NFL concentration
 732 in large cohorts of SCA-3 subjects and demonstrated higher
 733 levels in both preclinical and manifest SCA-3 individuals
 734 compared with HC [167]. Serum NFL levels discriminated
 735 manifest SCA-3 from HC with excellent accuracy, and the
 736 diagnostic performance remained good in distinguishing pre-
 737 clinical SCA-3 subjects from HC (Table 2) [167]. Recently,
 738 plasma NFL concentrations resulted higher also in patients
 739 affected by Friedreich's ataxia (FA), which is the most com-
 740 mon autosomal recessive ataxia caused by CAG repeat expan-
 741 sion in the ATXN3/MJD1 gene, compared with aged-
 742 matched controls [168].

743 **NFL as Staging and Prognostic Biomarker**

744 Serum NFL in c-MSA patients does not seem to correlate with
 745 clinical disease severity (as assessed by the Scale for the
 746 Assessment and Rating of Ataxia (SARA)) or disease progres-
 747 sion [165]. Similarly, a recent study in 99 patients with geneti-
 748 cally confirmed FA did not find a correlation with disease
 749 severity (as defined by SARA score), age at onset, or disease
 750 duration [169]. Moreover, serum NFL concentration remains

stable in a subgroup of 14 FA patients who received a 2-year
 follow-up evaluation [169].

Conversely, serum NFL concentration increased with dis-
 ease severity in a large cohort of SCA-3 patients, including
 manifest and preclinical individuals, and correlated with both
 clinical scales (according to SARA and International
 Cooperative Ataxia Rating Scale (ICARS) scores) and reduc-
 tion of cerebellar and brainstem volume [167]. Preclinical
 SCA-3 group was divided in early and late preclinical sub-
 groups using the median predicted number of years to onset of
 manifest disease. Serum NFL concentrations resulted higher
 in manifest than preclinical SCA-3 subjects and in late pre-
 clinical SCA-3 subjects compared with early preclinical SCA-
 3 individuals. However, no differences were observed be-
 tween early preclinical subjects and HC [167]. Despite CAG
 repeat count is a well-known prognostic factor for SCA-3 and
 FA, a correlation between serum NFL and CAG repeat lengths
 has been inconsistently reported [168, 169].

NFL as Risk/Screening Biomarker

In their study on 133 SCA-3 patients, Li and colleagues [167]
 demonstrated higher serum NFL concentrations in 26 preclini-
 cal ATXN3 mutation carriers (patients with SARA score < 3)
 compared with controls. Moreover, a correlation between mo-
 tor symptoms, neuroimaging markers, and serum NFL was
 found in all ATXN3 mutation carriers, suggesting that NFL
 may serve to track neurodegeneration and disease progression
 already in pre and prodromal SCA-3 phases.

HD

Among the most common neurodegenerative diseases, HD is
 unique, since the major part (~99%) of individuals presenting
 a HD phenotype have a mutation in the same gene [170].
 Indeed, HD is an autosomal dominant inherited neurodegen-
 erative disease with the typical manifestations of involuntary
 movements, psychiatric symptoms, and cognitive decline.
 The etiological basis is the deleterious expansion of polyglu-
 tamine encoding CAG repeats in the huntingtin (HTT) gene,
 leading to the expression of neurotoxic mutant huntingtin
 (mHTT) and extensive degeneration of neurons primarily
 occurring in the striatum and cortex [171]. The disease usu-
 ally starts in midlife, with age of onset inversely correlat-
 ing to CAG repeat number [171]. Although the cause is
 known, disease-modifying treatments are not yet available.
 In HD, a reliable genetic test confirms a clinical diagnosis in
 symptomatic people or predicts disease onset in asymptomatic
 mutation carriers [172]. As a consequence, a novel biomarker
 should be directed to track disease progression and predict a
 treatment response to targeted therapies. Although not conclu-
 sive, the results of the available studies display that blood NFL

799 could have a role in this context. Previous reports on CSF
 800 NFL indicate elevated concentrations in HD subjects [19,
 801 173, 174]. This is not surprising because mtHTT interacts with
 802 other proteins altering their function and finally leading to
 803 abnormal protein aggregation and impaired axonal transport
 804 [175]. Furthermore, the level of misfolded mtHTT protein
 805 correlates with NFL concentration in CSF, thus suggesting a
 806 contemporary releasing of both proteins from damaged neu-
 807 rons [176].

808 **NFL as Staging and Prognostic Biomarker**

809 In the first retrospective study investigating NFL concentra-
 810 tions in the blood of premanifest HD (preHD) and early-stage
 811 HD patients enrolled in the TRACK-HD cohort, Byrne and
 812 colleagues [177] showed higher baseline NFL levels in 201
 813 HTT mutation carriers, including 58 with early premanifest
 814 and 46 subjects with late premanifest disease, than in controls.
 815 Moreover, NFL concentration reflected baseline motor and
 816 cognitive deficits in HD patients and differed significantly
 817 with increasing disease stage. Positive associations were
 818 found between plasma NFL concentration, age, and CAG
 819 triplet repeat counts, with higher CAG lengths being associat-
 820 ed with earlier and steeper increases in plasma NFL [177].
 821 Therefore, NFL is the first biofluid marker showing a direct
 822 relationship with a causative gene expansion [178].

823 Of note, baseline plasma NFL predicted rates of brain at-
 824 rophy, cognitive decline, and worsening of functional ability
 825 and motor performance in mtHTT carriers [177].
 826 Interestingly, it was closely associated with the rate of
 827 whole-brain atrophy than with the rate of striatal one, suggest-
 828 ing that plasma NFL reflects more the rate of global neuronal
 829 degeneration than that of a specific brain area [177].
 830 Additionally, in the same TRACK-HD cohort, Johnson and
 831 colleagues [179] showed voxel-wise region-specific associa-
 832 tions between plasma NFL levels and both cross-sectional and
 833 longitudinal MRI cortical thinning and white matter volume
 834 reduction, highlighting the value of NFL as a dynamic and
 835 robust marker of brain atrophy. Notably, higher concentra-
 836 tions of NFL in plasma were associated with lower volume
 837 in regions known to be affected in HD and predicted subse-
 838 quent occipital gray matter atrophy and widespread white mat-
 839 ter reduction over the 3-year follow-up, independently of age
 840 and CAG length repeats [179]. NFL increased significantly
 841 from baseline both in individuals with premanifest HD and
 842 in those with manifest HD [179]. Remarkably, in a subsequent
 843 study, Byrne and colleagues [180], combining CSF/plasma
 844 NFL and CSF mutant huntingtin protein (mHTT_p) as biofluid
 845 biomarkers, demonstrated that NFL levels were more accurate
 846 than mHTT_p to discriminate between premanifest and mani-
 847 fested HDs and correlated with severity of symptoms better than
 848 mHTT_p in manifest HD.

CJD

849 CJD is the most common human prion disease. Approximately 850
 851 85% of cases are sCJD, but in a minority of cases, CJD can be
 852 genetically determined (gCJD) [181]. The disease is a rapidly
 853 progressive and fatal neurodegenerative condition, whose differ-
 854 ent phenotypes depend, at least in part, by polymorphisms on
 855 the gene encoding prion protein (PrP) [182]. Diagnosis is fre-
 856 quently tardive and relies on clinical World Health Organization
 857 (WHO) criteria supported by detection of the 14-3-3 protein
 858 and, more recently, t-tau in the CSF [66, 183, 184]. Also, CSF
 859 NFL recently demonstrated to be a reliable biomarker in the
 860 CJD diagnostic workup. Although few studies explored its role
 861 as a biomarker in CJD patients so far, CSF NFL levels are
 862 significantly increased in CJD (including those with more slow-
 863 ly progressive and atypical disease course) compared with AD,
 864 FTD, other NDDs (dementia), and controls, indicating a massive
 865 synaptic degeneration and neuroaxonal damage in CJD [22, 23,
 866 185, 186]. Additionally, it is noteworthy that NFL concentra-
 867 tions in CSF appear highly variable among different sCJD sub-
 868 types, with higher NFL levels in those with more rapidly
 869 progressing disease [187]. Importantly, plasma NFL correlates
 870 with CSF NFL concentration and recent studies suggest that
 871 blood NFL can accurately reflect the massive neurodegeneration
 872 in CJD patients.

NFL as Diagnostic Biomarker

873 Diagnostic accuracy of serum NFL for discrimination be- 874
 875 tween CJD and controls was excellent (Table 2) [185]. 875
 876 These findings have been independently replicated in other 876
 877 two studies. Serum NFL distinguished patients from controls 877
 878 with 100% sensitivity and 100% specificity in 45 sCJD pa- 878
 879 tients enrolled in the National Prion Monitoring Cohort [188]. 879
 880 Noteworthy, Kovacs and colleagues [189] reported high sen- 880
 881 sitivity and specificity of plasma NFL concentration in dis- 881
 882 criminating CJD subjects from non-CJD controls in a cohort 882
 883 of 132 pathologically classified patients (sCJD, gCJD, and 883
 884 AD cases) showing a rapidly progressive neurological picture. 884
 885 However, in this study, the diagnostic value in the differenti- 885
 886 ation between prion and other disease cases resulted lower 886
 887 than previously reported investigations (Table 2) [189]. 887
 888 Moreover, serum NFL values have been elevated since the 888
 889 early phases of the disease, suggesting a possible role as a 889
 890 screening biomarker [188]. Conversely, serum NFL concen- 890
 891 tration overlapped between ALS and CJD patients in a recent 891
 892 prospective study, even though the size of CJD group was 892
 893 very small [139]. 893

NFL as Staging and Prognostic Biomarker

894 Longitudinal changes in serum tau and NFL levels were in- 895
 896 vestigated in the aforementioned study of Thompson and 896

897 colleagues [188]. However, despite a trend toward increasing
898 concentrations of both tau and NFL over the last 12 months
899 before death, at variance with tau, no association was found
900 between serum NFL concentration and speed of decline on the
901 Medical Research Council (MRC) Prion Disease Rating Scale
902 [188]. Also, a recent study strengthens the tight association of
903 plasma tau levels with the rate of disease progression and
904 survival time in sCJD [190].

905 NFL as Risk/Screening Biomarker

906 gCJD forms are linked to mutations in the prion protein gene
907 (PRNP) inherited with an autosomal dominant pattern and
908 variable penetrance [191]. The relationship between genotype
909 and phenotype remains a matter of debate, and index cases do
910 not always have a family history [192]. Under this scenario,
911 the discovery of biomarkers serving as surrogates of outcome
912 in clinical trials may be crucial because *PRNP* mutation carriers
913 might benefit from a presymptomatic intervention. PrP-
914 lowering therapeutics are now in preclinical development, and
915 to this end, CSF total PrP has demonstrated to be a strong
916 candidate fluid biomarker showing stable low levels in an
917 ongoing natural history study including presymptomatic mu-
918 tation carriers and normal controls [193].

919 Conclusions

920 The impact of physiological variables on blood NFL concen-
921 tration, such as sex and age, has not been systematically in-
922 vestigated across published studies. Other variables possibly
923 affecting its modification in peripheral blood, including sys-
924 temic comorbidities and concomitant drug therapies, were not
925 taken into account. Liver and renal clearance as well as blood
926 cell counts and plasma protein composition could affect bio-
927 marker concentrations [194]; however, these factors were not
928 investigated. Indeed, blood NFL concentration alterations as-
929 sociated with renal and hepatic dysfunctions remain unknown,
930 thus representing a potential relevant methodological bias,
931 especially in subjects with NDDs, being generally old and
932 frequently exhibiting vascular comorbidities. In general,
933 blood NFL concentrations correlate with aging due to a subtle
934 axonal degeneration and vascular changes in elderly.
935 However, such an association was clearly absent in CJD,
936 ALS, and AP. This suggests that the probable contribution
937 of aging on NFL concentrations and neurodegeneration be-
938 comes trivial in highly aggressive forms of NDDs [195].

939 Blood NFL concentrations are reported to be massively
940 elevated in ALS patients, even in early disease stages, indicat-
941 ing its value as an efficacious, yet unspecific, biomarker in the
942 differential diagnosis of ALS from ALS mimics. Moreover,
943 blood NFL concentrations can reflect disease severity and/or
944 progression in ALS, suggesting that peripheral NFL could

945 contribute to support ALS prognosis. Other confounding var- 945
946 variables, such as the discordant disease progression in the dif- 946
947 ferent clinical subtypes of ALS, should be elucidated in fur- 947
948 ther studies. Actually, blood NFL could help better stratify the 948
949 multifaceted clinical presentation of ALS phenotypes. 949
950 However, further studies are needed to confirm its prognostic 950
951 value. In contrast with genetic forms of AD, blood NFL con- 951
952 centrations are not increased in presymptomatic ALS muta- 952
953 tion carriers appearing to tightly link to the symptomatic phase 953
954 of the disease. 954

955 Another NDD where blood NFL increases result impres- 955
956 sively is CJD. Because of early diagnosis remains challeng- 956
957 ing, NFL might be a reliable screening blood-based biomarker 957
958 with a potentially high negative predictive value for CJD sub- 958
959 jects, ruling out more common and less aggressive neurode- 959
960 generative dementia, such as AD. Definitely, blood NFL 960
961 could be of usefulness as a first-step examination to promptly 961
962 detect CJD during its prodromal phase and to start future 962
963 disease-modifying treatments. On the other hand, preliminary 963
964 findings do not support a potential use of NFL as a predictor of 964
965 longitudinal disease progression in CJD, and its specificity 965
966 should be further substantiated in comparison with other high- 966
967 ly aggressive forms of NDD. 967

968 All reported studies showed a good or even excellent diag- 968
969 nostic performance of blood NFL in distinguishing patients 969
970 with neurodegenerative disorders from HC. The potential con- 970
971 tribution of this biomarker candidate to discriminate between 971
972 different dementia disorders remains ambiguous given the 972
973 lack of pathognomonic specificity. However, mounting data 973
974 suggest that blood NFL could be a useful diagnostic tool in the 974
975 diagnostic workup of FTD, to distinguish FTD (especially 975
976 FTD with a TDP-43 pathology) from AD patients and to iden- 976
977 tify PPA with a likely underlying AD pathology, including 977
978 lvPPA. Moreover, it can represent a biomarker tracking the 978
979 disease progression and potentially identifying the transition 979
980 phase from the presymptomatic to the symptomatic stage of 980
981 the genetic forms of the disease. Most importantly, serum 981
982 NFL is assumed to be a promising screening tool to rule out 982
983 an underlying neurodegenerative disease in individuals with 983
984 psychiatric disorders. 984

985 In AD, blood NFL may predict progression to dementia 985
986 in individuals with MCI at high risk and identify preclin- 986
987 ical AD before the conversion phase. Moreover, NFL cap- 987
988 tures early neurodegenerative changes in presymptomatic 988
989 familiar AD mutation carriers in which blood NFL con- 989
990 centrations correlate with the predicted time to symptom 990
991 onset. Further studies are crucial to calculate the negative 991
992 predictive value of blood NFL as a screening tool in large 992
993 and selected cohorts of individuals at risk of neurodegen- 993
994 eration and AD, such as individuals with subjective mem- 994
995 ory complaints and/or decline, late-onset psychiatric dis- 995
996 orders, cerebrovascular disease, and diabetes, as well as in 996
997 aging and elderly individuals in general. 997

998 Because of the high negative predictive value of elevated
 999 blood NFL concentrations in excluding PD, this candidate
 1000 biomarker can represent a valid screening tool for clinicians
 1001 in the early differential diagnosis between PD and AP in cases
 1002 with confounding clinical presentations. In contrast, blood
 1003 NFL measurements do not suffice to differentiate PD patients
 1004 from controls and cannot be used to separate PSP, MSA, and
 1005 CBD from each other. Nevertheless, blood NFL may be a
 1006 prognostic tool in clinical practice in both PD and AP patients.
 1007 Currently, to the best of our knowledge, there are no studies
 1008 investigating blood NFL in patients with DLB, and few stud-
 1009 ies are available for PDD patients.

1010 Regarding other relevant clinical neurological presenta-
 1011 tions, blood NFL may support the classification of sporadic
 1012 late-onset ataxias, notably helping in differentiating c-MSA-C
 1013 from SAOA. In choreic patients, blood NFL appears to be a
 1014 robust prognostic biomarker of HD disease onset and progres-
 1015 sion and holds potential as a predictive biomarker of response
 1016 to disease-modifying agents in clinical trials.

1017 Finally, blood NFL seems to be a promising candidate
 1018 predictor of the timing of clinical phenoconversion in pre-
 1019 symptomatic mutation carriers with AD, HD, and SCA-3.
 1020 Conversely, blood NFL concentrations are mostly normal in
 1021 premanifest ALS and FTD mutation carriers but promptly
 1022 increase with the onset of clinical symptoms.

1023 In general, given the rapid advances in elucidating the path-
 1024 ophysiological mechanisms of diseases, at the molecular diag-
 1025 nostic level, biomarkers are excellent flexible tools to im-
 1026 prove and inform all phases of drug discovery and develop-
 1027 ment by enabling validation of mechanisms of actions [196,
 1028 197]. For this reason, NFL is assumed to act as an innovative
 1029 molecular mechanistic biomarker supporting *in vivo* detection
 1030 and the measurement of definite pathophysiological mecha-
 1031 nisms across the spectrum of different NDDs. Together with
 1032 other innovative molecular indicators, NFL will help establish
 1033 panels of biomarkers—i.e., molecular signatures—
 1034 encompassing the entire spectrum of molecular events of the
 1035 NDD spectrum disorders. Applying these molecular signa-
 1036 tures in longitudinal investigations will be critical to provide
 1037 information to depict the pathophysiological processes char-
 1038 acterizing different NDDs [198]. These innovative biomarkers
 1039 will enable the selection of the most appropriate therapies for
 1040 individual patients by defining which molecular pathophysio-
 1041 logical events account for the patient’s clinical symptoms at
 1042 different stages of the disease [199, 200]. This will establish
 1043 the grounds to develop effective targeted treatment strate-
 1044 gies—i.e., “molecularly” targeted therapies—for the accurate
 1045 treatment of specific molecular pathophysiological pathways.
 1046 Future developments in investigating NDD heterogeneity will
 1047 allow clinicians to deliver targeted interventions that are “cus-
 1048 tomized,” i.e., tailored, to the definite profiles of the individual
 1049 NDD patient, according to the precision medicine paradigm.
 1050 Such a precision medicine-based strategy is now increasingly

facing the clinical and biological/genetic complexity and het- 1051
 erogeneity of the various forms of NDD [198]. Precision med- 1052
 icine emphasizes the need of clinical medicine to focus on the 1053
 pathophysiology of the individual patient, with his/her own 1054
 distinctive, diverse, and complex matrix of multisystem fea- 1055
 tures [200]. Concerted global efforts will pave the way for a 1056
 future of neurology, in which drugs will timely and effectively 1057
 support the prevention and treatment of diseases with very 1058
 precise biomarker-guided targeted approaches for the right 1059
 patient at the right time [201]. 1060

Acknowledgments H.H. is an employee of Eisai Inc. This work has been 1061
 performed during his previous position at Sorbonne University, Paris, 1062
 France. At Sorbonne University, he was supported by the AXA 1063
 Research Fund, the “Fondation partenariale Sorbonne Université,” and 1064
 the “Fondation pour la Recherche sur Alzheimer,” Paris, France. 1065
 1066

Compliance with Ethical Standards 1067

Competing Interests S.L. received lecture honoraria from Roche and 1068
 Servier. 1069

H.H. is an employee of Eisai Inc. and serves as Senior Associate 1070
 Editor for the Journal Alzheimer’s & Dementia and does not receive 1071
 any fees or honoraria since May 2019; before May 2019, he had received 1072
 lecture fees from Servier, Biogen, and Roche; research grants from Pfizer, 1073
 Avid, and MSD Avenir (paid to the institution); travel funding from 1074
 Functional Neuromodulation, Axovant, Eli Lilly and company, Takeda 1075
 and Zinfandel, GE Healthcare and Oryzon Genomics; and consultancy 1076
 fees from Qynapse, Jung Diagnostics, Cytox Ltd., Axovant, Anavex, 1077
 Takeda and Zinfandel, GE Healthcare, Oryzon Genomics, and 1078
 Functional Neuromodulation, and he also participated in scientific advi- 1079
 sory boards of Functional Neuromodulation, Axovant, Eisai, Eli Lilly and 1080
 company, Cytox Ltd., GE Healthcare, Takeda and Zinfandel, Oryzon 1081
 Genomics, and Roche Diagnostics. 1082

He is co-inventor in the following patents as a scientific expert and has 1083
 received no royalties: 1084

- In Vitro Multiparameter Determination Method for the Diagnosis 1085
 and Early Diagnosis of Neurodegenerative Disorders Patent 1086
 Number: 8916388 1087
- In Vitro Procedure for Diagnosis and Early Diagnosis of 1088
 Neurodegenerative Diseases Patent Number: 8298784 1089
- Neurodegenerative Markers for Psychiatric Conditions Publication 1090
 Number: 20120196300 1091
- In Vitro Multiparameter Determination Method for the Diagnosis 1092
 and Early Diagnosis of Neurodegenerative Disorders Publication 1093
 Number: 20100062463 1094
- In Vitro Method for the Diagnosis and Early Diagnosis of 1095
 Neurodegenerative Disorders Publication Number: 20100035286 1096
- In Vitro Procedure for Diagnosis and Early Diagnosis of 1097
 Neurodegenerative Diseases Publication Number: 20090263822 1098
- In Vitro Method for the Diagnosis of Neurodegenerative Diseases 1099
 Patent Number: 7547553 1100
- CSF Diagnostic In Vitro Method for Diagnosis of Dementias and 1101
 Neuroinflammatory Diseases Publication Number: 20080206797 1102
- In Vitro Method for the Diagnosis of Neurodegenerative Diseases 1103
 Publication Number: 20080199966 1104
- Neurodegenerative Markers for Psychiatric Conditions Publication 1105
 Number: 20080131921 1106

1107 G.P., S.M., A.D.V., G.S., U.B., C.A., R.C., and F.B. declare that they
1108 have no conflicts of interest relevant to this work.

q5 1109 **References**

1110 1. Glass CK, Saijo K, Winner B, Marchetto MC, Gage FH (2010)
1111 Mechanisms underlying inflammation in neurodegeneration. *Cell*
1112 140(6):918–934
1113 2. Heemels MT (2016) Neurodegenerative diseases. *Nature*
1114 539(7628):179
1115 3. Chi H, Chang HY, Sang TK (2018) Neuronal cell death mecha-
1116 nisms in major neurodegenerative diseases. *Int J Mol Sci* 19(10):
1117 3082
1118 4. Hampel H, Toschi N, Baldacci F et al (2018) Alzheimer’s disease
1119 biomarker-guided diagnostic workflow using the added value of
1120 six combined cerebrospinal fluid candidates: A β_{1-42} , total-tau,
1121 phosphorylated-tau, NFL, neurogranin, and YKL-40. *Alzheimers Dement* 14(4):492–501
1122 5. Khalil M, Teunissen CE, Otto M et al (2018) Neurofilaments as
1123 biomarkers in neurological disorders. *Nat Rev Neurol* 14(10):
1124 577–589
1125 6. Gaetani L, Blennow K, Calabresi P, Di Filippo M, Parnetti L,
1126 Zetterberg H (2019) Neurofilament light chain as a biomarker in
1127 neurological disorders. *J Neurol Neurosurg Psychiatry* 90(8):870–
1128 881
1129 7. Zhao Y, Xin Y, Meng S, He Z, Hu W (2019) Neurofilament light
1130 chain protein in neurodegenerative dementia: a systematic review
1131 and network meta-analysis. *Neurosci Biobehav Rev* 102:123–138
1132 8. Kanaan NM, Pigino GF, Brady ST, Lazarov O, Binder LI, Morfini
1133 GA (2013) Axonal degeneration in Alzheimer’s disease: when
1134 signaling abnormalities meet the axonal transport system. *Exp*
1135 *Neurol* 246:44–53
1136 9. Yuan A, Rao MV, Veeranna NRA (2012) Neurofilaments at a
1137 glance. *J Cell Sci* 125:3257–3263
1138 10. Zetterberg H (2016) Neurofilament light: a dynamic cross-disease
1139 fluid biomarker for neurodegeneration. *Neuron* 91(1):1–3
1140 11. Liu Q, Xie F, Siedlak SL et al (2004) Neurofilament proteins in
1141 neurodegenerative diseases. *Cell Mol Life Sci* 61(24):3057–3075
1142 12. Yuan A, Rao MV, Veeranna NRA (2017) Neurofilaments and
1143 neurofilament proteins in health and disease. *Cold Spring Harb*
1144 *Perspect Biol* 9(4):a018309
1145 13. Petzold A, Keir G, Warren J, Fox N, Rossor MN (2007) A sys-
1146 tematic review and meta-analysis of CSF neurofilament protein
1147 levels as biomarkers in dementia. *Neurodegener Dis* 4(2–3):
1148 185–194
1149 14. Rosengren LE, Karlsson JE, Karlsson JO, Persson LI, Wikkelso C
1150 (1996) Patients with amyotrophic lateral sclerosis and other neu-
1151 rodegenerative diseases have increased levels of neurofilament
1152 protein in CSF. *J Neurochem* 67(5):2013–2018
1153 15. Landqvist Waldo M, Frizell Santillo A et al (2013) Cerebrospinal
1154 fluid neurofilament light chain protein levels in subtypes of
1155 frontotemporal dementia. *BMC Neurol* 13:54
1156 16. Meeter LH, Dopfer EG, Jiskoot LC et al (2016) Neurofilament
1157 light chain: a biomarker for genetic frontotemporal dementia. *Ann*
1158 *Clin Transl Neurol* 3(8):623–636
1159 17. Hall S, Öhrfelt A, Constantinescu R et al (2012) Accuracy of a
1160 panel of 5 cerebrospinal fluid biomarkers in the differential diag-
1161 nosis of patients with dementia and/or parkinsonian disorders.
1162 *Arch Neurol* 69(11):1445–1452
1163 18. Magdalinou NK, Paterson RW, Schott JM et al (2015) A panel of
1164 nine cerebrospinal fluid biomarkers may identify patients with
1165 atypical parkinsonian syndromes. *J Neurol Neurosurg Psychiatry*
1166 86(11):1240–1247

1167 19. Vinther-Jensen T, Börnsen L, Budtz-Jørgensen E et al (2016) 1168
1169 Selected CSF biomarkers indicate no evidence of early neuroin-
1170 flammation in Huntington disease. *Neurol Neuroimmunol*
1171 *Neuroinflamm* 3(6):e287
1172 20. Lista S, Toschi N, Baldacci F et al (2017) Diagnostic accuracy of 1172
1173 CSF neurofilament light chain protein in the biomarker-guided
1174 classification system for Alzheimer’s disease. *Neurochem Int*
1175 108:355–360
1176 21. Zetterberg H, Skillbäck T, Mattsson N et al (2016) Association of 1176
1177 cerebrospinal fluid neurofilament light concentration with
1178 Alzheimer disease progression. *Jama Neurol* 73(1):60–67
1179 22. van Eijk JJ, van Everbroeck B, Abdo WF, Kremer BP, Verbeek 1179
1180 MM (2010) CSF neurofilament proteins levels are elevated in
1181 sporadic Creutzfeldt-Jakob disease. *J Alzheimers Dis* 21(2):569–
1182 576
1183 23. Kanata E, Golanska E, Villar-Piqué A et al (2019) Cerebrospinal 1183
1184 fluid neurofilament light in suspected sporadic Creutzfeldt-Jakob
1185 disease. *J Clin Neurosci* 60:124–127
1186 24. Lycke JN, Karlsson JE, Andersen O, Rosengren LE (1998) 1186
1187 Neurofilament protein in cerebrospinal fluid: a potential marker
1188 of activity in multiple sclerosis. *J Neurol Neurosurg Psychiatry*
1189 64:402–404
1190 25. Yilmaz A, Blennow K, Hagberg L et al (2017) Neurofilament 1190
1191 light chain protein as a marker of neuronal injury: review of its
1192 use in HIV-1 infection and reference values for HIV-negative
1193 controls. *Expert Rev Mol Diagn* 17(8):761–770
1194 26. Shahim P, Tegner Y, Gustafsson B et al (2016) Neurochemical 1194
1195 aftermath of repetitive mild traumatic brain injury. *JAMA Neurol*
1196 73(11):1308–1315
1197 27. Pujol-Calderón F, Portelius E, Zetterberg H, Blennow K, 1197
1198 Rosengren LE, Höglund K (2019) Neurofilament changes in se-
1199 rum and cerebrospinal fluid after acute ischemic stroke. *Neurosci*
1200 *Lett* 698:58–63
1201 28. Pawlitzki M, Butryn M, Kirchner F et al (2019) CSF 1201
1202 Neurofilament light chain level predicts axonal damage in cere-
1203 bral vasculitis. *Ann Clin Transl Neurol* 6(6):1134–1137
1204 29. Skillbäck T, Farahmand B, Bartlett JW et al (2014) CSF neurofilam- 1204
1205 ent light differs in neurodegenerative diseases and predicts se-
1206 verity and survival. *Neurology* 83(21):1945–1953
1207 30. Bacioglu M, Maia LF, Preische O et al (2016) Neurofilament light 1207
1208 chain in blood and CSF as marker of disease progression in mouse
1209 models and in neurodegenerative diseases. *Neuron* 91:56–66
1210 31. Bridel C, van Wieringen WN, Zetterberg H et al (2019) 1210
1211 Diagnostic value of cerebrospinal fluid neurofilament light pro-
1212 tein in neurology: a systematic review and meta-analysis. *Jama*
1213 *Neurol* 76(9):1035–1048
1214 32. Mielke MM, Syrjanen JA, Blennow K et al (2019) Plasma and 1214
1215 CSF neurofilament light: relation to longitudinal neuroimaging
1216 and cognitive measures. *Neurology* 93(3):e252–e260
1217 33. Molinuevo JL, Ayton S, Batrla R et al (2018) Current state of 1217
1218 Alzheimer’s fluid biomarkers. *Acta Neuropathol* 136(6):821–853
1219 34. Hampel H, Lista S, Khachaturian ZS (2012) Development of bio- 1219
1220 markers to chart all Alzheimer’s disease stages: the royal road to
1221 cutting the therapeutic Gordian Knot. *Alzheimers Dement* 8(4):
1222 312–336
1223 35. Baldacci F, Lista S, Garaci F, Bonuccelli U, Toschi N, Hampel H 1223
1224 (2016) Biomarker-guided classification scheme of neurodegener-
1225 ative diseases. *J Sport Health Sci* 5(4):383–387
1226 36. Hampel H, Frank R, Broich K et al (2010) Biomarkers for 1226
1227 Alzheimer’s disease: academic, industry and regulatory perspec-
1228 tives. *Nat Rev Drug Discov* 9(7):560–574
1229 37. Blennow K, Hampel H, Weiner M, Zetterberg H (2010) 1229
1230 Cerebrospinal fluid and plasma biomarkers in Alzheimer disease.
1231 *Nat Rev Neurol* 6(3):131–144

1232 38. Zetterberg H, Blennow K (2018) From cerebrospinal fluid to
1233 blood: the third wave of fluid biomarkers for Alzheimer's disease.
1234 *J Alzheimers Dis* 64(s1):S271–S279

1235 39. Kuhle J, Barro C, Andreasson U et al (2016) Comparison of three
1236 analytical platforms for quantification of the neurofilament light
1237 chain in blood samples: ELISA, electrochemiluminescence immu-
1238 noassay and Simoa. *Clin Chem Lab Med* 54(10):1655–1661

1239 40. Gaiottino J, Norgren N, Dobson R et al (2013) Increased neuro-
1240 filament light chain blood levels in neurodegenerative neurologi-
1241 cal diseases. *PLoS One* 8(9):e75091

1242 41. Baldacci F, Mazzucchi S, Della Vecchia A et al (2020) The path to
1243 biomarker-based diagnostic criteria for the spectrum of neurode-
1244 generative diseases. *Expert Rev Mol Diagn* 20(4):421–441

1245 42. Califf RM (2018) Biomarker definitions and their applications.
1246 *Exp Biol Med (Maywood)* 243(3):213–221

1247 43. Biomarkers Definitions Working Group (2001) Biomarkers and
1248 surrogate endpoints: preferred definitions and conceptual frame-
1249 work. *Clin Pharmacol Ther* 69(3):89–95

1250 44. Chen XH, Huang S, Kerr D (2011) Biomarkers in clinical medi-
1251 cine. *IARC Sci Publ* 163:303–322

1252 45. McKhann G, Drachman D, Folstein M, Katzman R, Price D,
1253 Stadlan EM (1984) Clinical diagnosis of Alzheimer's disease:
1254 report of the NINCDS-ADRDA Work Group under the auspices
1255 of Department of Health and Human Services Task Force on
1256 Alzheimer's Disease. *Neurology* 34:939–944

1257 46. McKhann GM, Knopman DS, Chertkow H et al (2011) The diag-
1258 nosis of dementia due to Alzheimer's disease: recommendations
1259 from the National Institute on Aging-Alzheimer's Association
1260 workgroups on diagnostic guidelines for Alzheimer's disease.
1261 *Alzheimers Dement* 7:263–269

1262 47. Sperling RA, Aisen PS, Beckett LA et al (2011) Toward defining
1263 the preclinical stages of Alzheimer's disease: recommendations
1264 from the National Institute on Aging-Alzheimer's Association
1265 workgroups on diagnostic guidelines for Alzheimer's disease.
1266 *Alzheimers Dement* 7:280–292

1267 48. Albert MS, DeKosky ST, Dickson D et al (2011) The diagnosis of
1268 mild cognitive impairment due to Alzheimer's disease: recom-
1269 mendations from the National Institute on Aging-Alzheimer's
1270 Association workgroups on diagnostic guidelines for
1271 Alzheimer's disease. *Alzheimers Dement* 7:270–279

1272 49. Dubois B, Feldman HH, Jacova C et al (2014) Advancing research
1273 diagnostic criteria for Alzheimer's disease: the IWG-2 criteria.
1274 *Lancet Neurol* 13:614–629

1275 50. Brooks BR (1994) El Escorial World Federation of Neurology
1276 criteria for the diagnosis of amyotrophic lateral sclerosis.
1277 Subcommittee on Motor Neuron Diseases/Amyotrophic Lateral
1278 Sclerosis of the World Federation of Neurology Research Group
1279 on Neuromuscular Diseases and the El Escorial "Clinical limits of
1280 amyotrophic lateral sclerosis" workshop contributors. *J Neurol Sci*
1281 124(Suppl):96–107

1282 51. Costa J, Swash M, de Carvalho M (2012) Awaji criteria for the
1283 diagnosis of amyotrophic lateral sclerosis: a systematic review.
1284 *Arch Neurol* 69:1410–1416

1285 52. Al-Chalabi A, Hardiman O, Kiernan MC, Chiò A, Rix-Brooks B,
1286 van den Berg LH (2016) Amyotrophic lateral sclerosis: moving
1287 towards a new classification system. *Lancet Neurol* 15(11):1182–
1288 1194

1289 53. McKeith IG, Galasko D, Kosaka K et al (1996) Consensus guide-
1290 lines for the clinical and pathologic diagnosis of dementia with
1291 Lewy bodies (DLB): report of the consortium on DLB interna-
1292 tional workshop. *Neurology* 47(5):1113–1124

1293 54. McKeith IG, Dickson DW, Lowe J et al (2005) Diagnosis and
1294 management of dementia with Lewy bodies: third report of the
1295 DLB Consortium. *Neurology* 65(12):1863–1872

55. McKeith IG, Boeve BF, Dickson DW et al (2017) Diagnosis and 1296
management of dementia with Lewy bodies: fourth consensus 1297
report of the DLB Consortium. *Neurology* 89(1):88–100 1298

56. Neary D, Snowden JS, Gustafson L et al (1998) Frontotemporal 1299
lobar degeneration: a consensus on clinical diagnostic criteria. 1300
Neurology 51:1546–1554 1301

57. Piguet O, Hornberger M, Mioshi E, Hodges JR (2011) 1302
Behavioural-variant frontotemporal dementia: diagnosis, clinical 1303
staging, and management. *Lancet Neurol* 10:162–172 1304

58. Gorno-Tempini ML, Hillis AE, Weintraub S et al (2011) 1305
Classification of primary progressive aphasia and its variants. 1306
Neurology 76:1006–1014 1307

59. Gibb WR, Lees AJ (1988) The relevance of the Lewy body to the 1308
pathogenesis of idiopathic Parkinson's disease. *J Neurol* 1309
Neurosurg Psychiatry 51:745–752 1310

60. Gelb DJ, Oliver E, Gilman S (1999) Diagnostic criteria for 1311
Parkinson disease. *Arch Neurol* 56(1):33–39 1312

61. Postuma RB, Berg D, Stern M et al (2015) MDS clinical diagnos- 1313
tic criteria for Parkinson's disease: MDS-PD clinical diagnostic 1314
criteria. *Mov Disord* 30(12):1591–1601 1315

62. Litvan I, Agid Y, Calne D et al (1996) Clinical research criteria for 1316
the diagnosis of progressive supranuclear palsy (Steele- 1317
Richardson-Olszewski syndrome): report of the NINDS-SPSP in- 1318
ternational workshop. *Neurology* 47:1–9 1319

63. Gilman S, Wenning GK, Low PA et al (2008) Second consensus 1320
statement on the diagnosis of multiple system atrophy. *Neurology* 1321
71(9):670–676 1322

64. Armstrong MJ, Litvan I, Lang AE et al (2013) Criteria for the 1323
diagnosis of corticobasal degeneration. *Neurology* 80(5):496–503 1324

65. Hoglinger GU, Respondek G, Stamelou M et al (2017) Clinical 1325
diagnosis of progressive supranuclear palsy: the movement disorder 1326
society criteria. *Mov Disord* 32:853–864 1327

66. World Health Organization (2003) WHO manual for surveillance 1328
of human transmissible spongiform encephalopathies, including 1329
variant Creutzfeldt-Jakob disease [https://apps.who.int/iris/handle/](https://apps.who.int/iris/handle/10665/42656) 1330
[10665/42656](https://apps.who.int/iris/handle/10665/42656) [Accessed 30 Jan 2003] 1331

67. Manix M, Kalakoti P, Henry M et al (2015) Creutzfeldt-Jakob 1332
disease: updated diagnostic criteria, treatment algorithm, and the 1333
utility of brain biopsy. *Neurosurg Focus* 39:E2 1334

68. Xia J, Broadhurst DI, Wilson M, Wishart DS (2013) Translational 1335
biomarker discovery in clinical metabolomics: an introductory 1336
tutorial. *Metabolomics* 9(2):280–299 1337

69. Winblad B, Amouyel P, Andrieu S et al (2016) Defeating 1338
Alzheimer's disease and other dementias: a priority for 1339
European science and society. *Lancet Neurol* 15(5):455–532 1340

70. Fogel DB (2018) Factors associated with clinical trials that fail and 1341
opportunities for improving the likelihood of success: a review. 1342
Contemp Clin Trials Commun 11:156–164 1343

71. Dubois B (2018) The emergence of a new conceptual framework 1344
for Alzheimer's disease. *J Alzheimers Dis* 62(3):1059–1066 1345

72. Beach TG, Monsell SE, Phillips LE, Kukull W (2012) Accuracy 1346
of the clinical diagnosis of Alzheimer disease at National Institute 1347
on Aging Alzheimer Disease Centers, 2005–2010. *J Neuropathol* 1348
Exp Neurol 71(4):266–273 1349

73. Jack CR Jr, Bennett DA, Blennow K et al (2016) A/T/N: an un- 1350
biased descriptive classification scheme for Alzheimer disease 1351
biomarkers. *Neurology* 87(5):539–547 1352

74. Jack CR Jr, Bennett DA, Blennow K et al (2018) NIA-AA re- 1353
search framework: toward a biological definition of Alzheimer's 1354
disease. *Alzheimers Dement* 14(4):535–562 1355

75. Hampel H, O'Bryant SE, Molinuevo JL et al (2018) Blood-based 1356
biomarkers for Alzheimer disease: mapping the road to the clinic. 1357
Nat Rev Neurol 14(11):639–652 1358

76. Ashton NJ, Leuzy A, Lim YM et al (2019) Increased plasma 1359
neurofilament light chain concentration correlates with severity 1360

1361 of post-mortem neurofibrillary tangle pathology and neurodegener- 1426
 1362 ation. *Acta Neuropathol Commun* 7(1):5 1427
 1363 77. Mattsson N, Andreasson U, Zetterberg H, Blennow K, 1428
 1364 Alzheimer's Disease Neuroimaging Initiative (2017) 1429
 1365 Association of plasma neurofilament light with neurodegenera- 1430
 1366 tion in patients with Alzheimer disease. *Jama Neurol* 74(5):557– 1431
 1367 566 1432
 1368 78. Vågberg M, Norgren N, Dring A et al (2015) Levels and age 1433
 1369 dependency of neurofilament light and glial fibrillary acidic 1434
 1370 protein in healthy individuals and their relation to the brain parenchy- 1435
 1371 mal fraction. *PLoS One* 10(8):e0135886 1436
 1372 79. Khalil M, Pirpamer L, Hofer E et al (2020) Serum neurofilament 1437
 1373 light levels in normal aging and their association with morpholog- 1438
 1374 ic brain changes. *Nat Commun* 11(1):812 1439
 1375 80. Forgrave LM, Ma M, Best JR, DeMarco ML (2019) The diagnos- 1440
 1376 tic performance of neurofilament light chain in CSF and blood for 1441
 1377 Alzheimer's disease, frontotemporal dementia, and amyotrophic 1442
 1378 lateral sclerosis: a systematic review and meta-analysis. 1443
 1379 *Alzheimers Dement (Amst)* 11:730–743 1444
 1380 81. Lewczuk P, Ermann N, Andreasson U et al (2018) Plasma neuro- 1445
 1381 filament light as a potential biomarker of neurodegeneration in 1446
 1382 Alzheimer's disease. *Alzheimers Res Ther* 10(1):71 1447
 1383 82. Mattsson N, Cullen NC, Andreasson U, Zetterberg H, Blennow K 1448
 1384 (2019) Association between longitudinal plasma neurofilament 1449
 1385 light and neurodegeneration in patients with Alzheimer disease. 1450
 1386 *JAMA Neurol* 76(7):791–799 1451
 1387 83. Pereira JB, Westman E, Hansson O, Alzheimer's Disease 1452
 1388 Neuroimaging Initiative (2017) Association between cerebrospinal 1453
 1389 fluid and plasma neurodegeneration biomarkers with brain 1454
 1390 atrophy in Alzheimer's disease. *Neurobiol Aging* 58:14–29 1455
 1391 84. Lin YS, Lee WJ, Wang SJ, Fuh JL (2018) Levels of plasma neuro- 1456
 1392 filament light chain and cognitive function in patients with 1457
 1393 Alzheimer or Parkinson disease. *Sci Rep* 8(1):17368 1458
 1394 85. Wang SY, Chen W, Xu W et al (2019) Neurofilament light chain 1459
 1395 in cerebrospinal fluid and blood as a biomarker for neurodegener- 1460
 1396 ative diseases: a systematic review and meta-analysis. *J* 1461
 1397 *Alzheimers Dis* 72(4):1353–1361 1462
 1398 86. Zhou W, Zhang J, Ye F et al (2017) Plasma neurofilament light 1463
 1399 chain levels in Alzheimer's disease. *Neurosci Lett* 650:60–64 1464
 1400 87. Benedet AL, Ashton NJ, Pascoal TA et al (2019) Plasma neuro- 1465
 1401 filament light associates with Alzheimer's disease metabolic 1466
 1402 decline in amyloid-positive individuals. *Alzheimers Dement (Amst)* 1467
 1403 11:679–689 1468
 1404 88. Timmers M, Tesseur I, Bogert J et al (2019) Relevance of the 1469
 1405 interplay between amyloid and tau for cognitive impairment in 1470
 1406 early Alzheimer's disease. *Neurobiol Aging* 79:131–141 1471
 1407 89. Preische O, Schultz SA, Apel A et al (2019) Serum neurofilament 1472
 1408 dynamics predicts neurodegeneration and clinical progression in 1473
 1409 presymptomatic Alzheimer's disease. *Nat Med* 25(2):277–283 1474
 1410 90. Hu H, Chen KL, Ou YN et al (2019) Neurofilament light chain 1475
 1411 plasma concentration predicts neurodegeneration and clinical pro- 1476
 1412 gression in nondemented elderly adults. *Aging (Albany NY)* 1477
 1413 11(17):6904–6914 1478
 1414 91. Chatterjee P, Goozee K, Sohrabi HR et al (2018) Association of 1479
 1415 plasma neurofilament light chain with neocortical amyloid- β load 1480
 1416 and cognitive performance in cognitively normal elderly partici- 1481
 1417 pants. *J Alzheimers Dis* 63(2):479–487 1482
 1418 92. Mayeli M, Mirshahvalad SM, Aghamollai V, Tafakhori A, 1483
 1419 Abdolalizadeh A, Rahmani F (2019) Plasma neurofilament light 1484
 1420 chain levels are associated with cortical hypometabolism in 1485
 1421 Alzheimer disease signature regions. *J Neuropathol Exp Neurol:* 1486
 1422 nlz054 1487
 1423 93. Weston PSJ, Poole T, Ryan NS et al (2017) Serum neurofilament 1488
 1424 light in familial Alzheimer disease: a marker of early neurodegen- 1489
 1425 eration. *Neurology* 89(21):2167–2175 1490
 94. Rascovsky K, Hodges JR, Knopman D et al (2011) Sensitivity of 1426
 revised diagnostic criteria for the behavioural variant of 1427
 frontotemporal dementia. *Brain* 134:2456–2477 1428
 95. Erkinen MG, Kim MO, Geschwind MD (2018) Clinical neuro- 1429
 logic and epidemiology of the major neurodegenerative diseases. 1430
Cold Spring Harb Perspect Biol 10(4):a033118 1431
 96. Bang J, Spina S, Miller BL (2015) Frontotemporal dementia. 1432
Lancet 386:1672–1682 1433
 97. Spinelli EG, Mandelli ML, Miller ZA et al (2017) Typical and 1434
 atypical pathology in primary progressive aphasia variants. *Ann* 1435
Neurol 81:430–443 1436
 98. Strong MJ, Abrahams S, Goldstein LH et al (2017) Amyotrophic 1437
 lateral sclerosis - frontotemporal spectrum disorder (ALS-FTSD): 1438
 revised diagnostic criteria. *Amyotroph Lateral Scler Frontotemporal* 1439
Degener 18:153–174 1440
 99. Rademakers R, Neumann M, Mackenzie IR (2012) Advances in 1441
 understanding the molecular basis of frontotemporal dementia. 1442
Nat Rev Neurol 8(8):423–434 1443
 100. Ducharme S, Dols A, Laforce R et al (2020) Recommendations to 1444
 distinguish behavioural variant frontotemporal dementia from 1445
 psychiatric disorders. *Brain* 143(6):1632–1650 1446
 101. Meeter LH, Kaat LD, Rohrer JD, van Swieten JC (2017) Imaging 1447
 and fluid biomarkers in frontotemporal dementia. *Nat Rev Neurol* 1448
 13(7):406–419 1449
 102. Zetterberg H, van Swieten JC, Boxer AL, Rohrer JD (2019) 1450
 Review: Fluid biomarkers for frontotemporal dementias. 1451
Neuropathol Appl Neurobiol 45(1):81–87 1452
 103. Pijnenburg YA, Verwey NA, van der Flier WM, Scheltens P, 1453
 Teunissen CE (2015) Discriminative and prognostic potential of 1454
 cerebrospinal fluid phosphoTau/tau ratio and neurofilaments for 1455
 frontotemporal dementia subtypes. *Alzheimers Dement (Amst)* 1456
 1(4):505–512 1457
 104. Ende E, Meeter L, Poos J et al (2019) Serum neurofilament light 1458
 chain in genetic frontotemporal dementia: a longitudinal, 1459
 multicentre cohort study. *Lancet Neurol* 18:1103–1111 1460
 105. Steinacker P, Semler E, Anderl-Straub S et al (2017) 1461
 Neurofilament as a blood marker for diagnosis and monitoring 1462
 of primary progressive aphasia. *Neurology* 88:961–969 1463
 106. Al Shweiki MR, Steinacker P, Oeckl P et al (2019) Neurofilament 1464
 light chain as a blood biomarker to differentiate psychiatric disor- 1465
 ders from behavioural variant frontotemporal dementia. *J* 1466
Psychiatr Res 113:137–140 1467
 107. Katisko K, Cajanus A, Jääskeläinen O et al (2020) Serum neuro- 1468
 filament light chain is a discriminative biomarker between 1469
 frontotemporal lobar degeneration and primary psychiatric disor- 1470
 ders. *J Neurol* 267(1):162–167 1471
 108. Wilke C, Preische O, Deuschle C et al (2016) Neurofilament light 1472
 chain in FTD is elevated not only in cerebrospinal fluid, but also in 1473
 serum. *J Neurol Neurosurg Psychiatry* 87:1270–1272 1474
 109. Steinacker P, Anderl-Straub S, Diehl-Schmid J et al (2018) Serum 1475
 neurofilament light chain in behavioral variant frontotemporal 1476
 dementia. *Neurology* 91(15):e1390–e1401 1477
 110. Matias-Guiu JA, Gomez-Pinedo U, Forero L et al (2019) Plasma 1478
 neurofilament light chain in primary progressive aphasia and re- 1479
 lated disorders: clinical significance and metabolic correlates. *J* 1480
Alzheimers Dis 72:1–10 1481
 111. Cellura E, Spataro R, Taiello AC, La Bella V (2012) Factors 1482
 affecting the diagnostic delay in amyotrophic lateral sclerosis. 1483
Clin Neurol Neurosurg 114(6):550–554 1484
 112. Turner MR, Kiernan MC, Leigh PN, Talbot K (2009) Biomarkers 1485
 in amyotrophic lateral sclerosis. *Lancet Neurol* 8(1):94–109 1486
 113. Lehnert S, Costa J, de Carvalho M et al (2014) Multicentre quality 1487
 control evaluation of different biomarker candidates for amyotro- 1488
 phic lateral sclerosis. *Amyotroph Lateral Scler Frontotemporal* 1489
Degener 15(5–6):344–350 1490

Q6

1491 114. Turner MR, Benatar M (2015) Ensuring continued progress in
1492 biomarkers for amyotrophic lateral sclerosis. *Muscle Nerve*
1493 51(1):14–18

1494 115. Xu Z, Henderson RD, David M, McCombe PA (2016)
1495 Neurofilaments as biomarkers for amyotrophic lateral sclerosis:
1496 a systematic review and meta-analysis. *PLoS One* 11(10):
1497 e0164625

1498 116. Turner MR, Gray E (2016) Are neurofilaments heading for the
1499 ALS clinic? *J Neurol Neurosurg Psychiatry* 87(1):3–4

1500 117. Brettschneider J, Petzold A, Süßmuth SD, Ludolph AC, Tumani
1501 H (2006) Axonal damage markers in cerebrospinal fluid are in-
1502 creased in ALS. *Neurology* 66(6):852–856

1503 118. Lin H, Schlaepfer WW (2006) Role of neurofilament aggregation
1504 in motor neuron disease. *Ann Neurol* 60(4):399–406

1505 119. Munoz DG, Greene C, Perl DP, Selkoe DJ (1988) Accumulation
1506 of phosphorylated neurofilaments in anterior horn motoneurons of
1507 amyotrophic lateral sclerosis patients. *J Neuropathol Exp Neurol*
1508 47(1):9–18

1509 120. Collard JF, Côté F, Julien JP (1995) Defective axonal transport in
1510 a transgenic mouse model of amyotrophic lateral sclerosis. *Nature*
1511 375(6526):61–64

1512 121. Lee MK, Marszalek JR, Cleveland DW (1994) A mutant neuro-
1513 filament subunit causes massive, selective motor neuron death:
1514 implications for the pathogenesis of human motor neuron disease.
1515 *Neuron* 13(4):975–988

1516 122. Cañete-Soler R, Silberg DG, Gershon MD, Schlaepfer WW
1517 (1999) Mutation in neurofilament transgene implicates RNA pro-
1518 cessing in the pathogenesis of neurodegenerative disease. *J*
1519 *Neurosci* 19(4):1273–1283

1520 123. Nie Z, Wu J, Zhai J et al (2002) Untranslated element in neurofil-
1521 ament mRNA has neuropathic effect on motor neurons of trans-
1522 genic mice. *J Neurosci* 22(17):7662–7670

1523 124. Zhai J, Lin H, Julien JP, Schlaepfer WW (2007) Disruption of
1524 neurofilament network with aggregation of light neurofilament
1525 protein: a common pathway leading to motor neuron degeneration
1526 due to Charcot-Marie-Tooth disease-linked mutations in NFL and
1527 HSPB1. *Hum Mol Genet* 16(24):3103–3116

1528 125. Lin H, Zhai J, Schlaepfer WW (2005) RNA-binding protein is
1529 involved in aggregation of light neurofilament protein and is im-
1530 plicated in the pathogenesis of motor neuron degeneration. *Hum*
1531 *Mol Genet* 14(23):3643–3659

1532 126. Watson D (1991) Regional variation in the abundance of axonal
1533 cytoskeletal proteins. *J Neurosci Res* 30(1):226–231

1534 127. Reijn TS, Abdo WF, Schelhaas HJ, Verbeek MM (2009) CSF
1535 neurofilament protein analysis in the differential diagnosis of
1536 ALS. *J Neurol* 256(4):615–619

1537 128. Steinacker P, Feneberg E, Weishaupt J et al (2016) Neurofilaments
1538 in the diagnosis of motoneuron diseases: a prospective study on 455
1539 patients. *J Neurol Neurosurg Psychiatry* 87(1):12–20

1540 129. Tortelli R, Ruggieri M, Cortese R et al (2012) Elevated cerebro-
1541 spinal fluid neurofilament light levels in patients with amyotrophic
1542 lateral sclerosis: a possible marker of disease severity and progres-
1543 sion. *Eur J Neurol* 19(12):1561–1567

1544 130. Zetterberg H, Jacobsson J, Rosengren L, Blennow K, Andersen
1545 PM (2007) Cerebrospinal fluid neurofilament light levels in amy-
1546 otrophic lateral sclerosis: impact of SOD1 genotype. *Eur J Neurol*
1547 14(12):1329–1333

1548 131. Tortelli R, Copetti M, Ruggieri M et al (2015) Cerebrospinal fluid
1549 neurofilament light chain levels: marker of progression to gener-
1550 alized amyotrophic lateral sclerosis. *Eur J Neurol* 22(1):215–218

1551 132. Poesen K, Van Damme P (2019) Diagnostic and prognostic per-
1552 formance of neurofilaments in ALS. *Front Neurol* 9:1167

1553 133. Menke RA, Gray E, Lu CH et al (2015) CSF neurofilament light
1554 chain reflects corticospinal tract degeneration in ALS. *Ann Clin*
1555 *Transl Neurol* 2(7):748–755

1556 134. Feneberg E, Oeckl P, Steinacker P et al (2018) Multicenter eval-
1557 uation of neurofilaments in early symptom onset amyotrophic lat-
1558 eral sclerosis. *Neurology* 90(1):e22–e30

1559 135. Gille B, De Schaepdryver M, Goossens J et al (2019) Serum
1560 neurofilament light chain levels as a marker of upper motor neuron
1561 degeneration in patients with amyotrophic lateral sclerosis.
1562 *Neuropathol Appl Neurobiol* 45(3):291–304

1563 136. Lu CH, Macdonald-Wallis C, Gray E et al (2015) Neurofilament
1564 light chain: a prognostic biomarker in amyotrophic lateral sclero-
1565 sis. *Neurology* 84(22):2247–2257

1566 137. Gaiani A, Martinelli I, Bello L et al (2017) Diagnostic and prog-
1567 nostic biomarkers in amyotrophic lateral sclerosis: neurofilament
1568 light chain levels in definite subtypes of disease. *JAMA Neurol*
1569 74(5):525–532

1570 138. Steinacker P, Huss A, Mayer B et al (2017) Diagnostic and prog-
1571 nostic significance of neurofilament light chain NF-L, but not
1572 progranulin and S100B, in the course of amyotrophic lateral scler-
1573 osis: data from the German MND-net. *Amyotroph Lateral Scler*
1574 *Frontotemporal Degener* 18(1–2):112–119

1575 139. Verde F, Steinacker P, Weishaupt JH et al (2019) Neurofilament
1576 light chain in serum for the diagnosis of amyotrophic lateral scler-
1577 osis. *J Neurol Neurosurg Psychiatry* 90(2):157–164

1578 140. Thouvenot E, Demattei C, Lehmann S et al (2020) Serum neuro-
1579 filament light chain at time of diagnosis is an independent prog-
1580 nostic factor of survival in amyotrophic lateral sclerosis. *Eur J*
1581 *Neurol* 27(2):251–257

1582 141. Poesen K, De Schaepdryver M, Stubendorff B et al (2017)
1583 Neurofilament markers for ALS correlate with extent of upper
1584 and lower motor neuron disease. *Neurology* 88(24):2302–2309

1585 142. De Schaepdryver M, Lunetta C, Tarlarini C et al (2020)
1586 Neurofilament light chain and C reactive protein explored as pre-
1587 dictors of survival in amyotrophic lateral sclerosis. *J Neurol*
1588 *Neurosurg Psychiatry* 91(4):436–437

1589 143. Weydt P, Oeckl P, Huss A et al (2016) Neurofilament levels as
1590 biomarkers in asymptomatic and symptomatic familial amyotro-
1591 phic lateral sclerosis. *Ann Neurol* 79(1):152–158

1592 144. Benatar M, Wu J, Andersen PM, Lombardi V, Malaspina A
1593 (2018) Neurofilament light: a candidate biomarker of presymp-
1594 tomatic amyotrophic lateral sclerosis and phenoconversion. *Ann*
1595 *Neurol* 84(1):130–139

1596 145. Olsson B, Alberg L, Cullen NC et al (2019) NFL is a marker of
1597 treatment response in children with SMA treated with nusinersen.
1598 *J Neurol* 266(9):2129–2136

1599 146. Corey DR (2017) Nusinersen, an antisense oligonucleotide drug
1600 for spinal muscular atrophy. *Nat Neurosci* 20(4):497–499

1601 147. Wurster CD, Günther R, Steinacker P et al (2019) Neurochemical
1602 markers in CSF of adolescent and adult SMA patients undergoing
1603 nusinersen treatment. *Ther Adv Neurol Disord* 12:
1604 1756286419846058

1605 148. Fratta P, Nirmalanathan N, Masset L et al (2014) Correlation of
1606 clinical and molecular features in spinal bulbar muscular atrophy.
1607 *Neurology* 82(23):2077–2084

1608 149. Respondek G, Roeber S, Kretschmar H et al (2013) Accuracy of
1609 the National Institute for Neurological Disorders and Stroke/Society
1610 for Progressive Supranuclear Palsy and neuroprotection and natural
1611 history in Parkinson plus syndromes criteria for the diagnosis of
1612 progressive supranuclear palsy. *Mov Disord* 28:504–509

1613 150. Rizzo G, Copetti M, Arcuti S, Martino D, Fontana A, Logroscino
1614 G (2016) Accuracy of clinical diagnosis of Parkinson disease: a
1615 systematic review and meta-analysis. *Neurology* 86(6):566–576

1616 151. Rizzo G, Arcuti S, Copetti M et al (2018) Accuracy of clinical
1617 diagnosis of dementia with Lewy bodies: a systematic review and
1618 meta-analysis. *J Neurol Neurosurg Psychiatry* 89(4):358–366

1619 152. Miki Y, Foti SC, Asi YT et al (2019) Improving diagnostic accu-
1620 racy of multiple system atrophy: a clinicopathological study. *Brain*
1621 142(9):2813–2827

1622 153. Abdo WF, Bloem BR, Van Geel WJ, Esselink RA, Verbeek MM 1688
 1623 (2007) CSF neurofilament light chain and tau differentiate multi- 1689
 1624 ple system atrophy from Parkinson's disease. *Neurobiol Aging* 1690
 1625 28(5):742–747
 1626 154. Herbert MK, Aerts MB, Beenes M et al (2015) CSF neurofilament 1691
 1627 light chain but not FLT3 ligand discriminates parkinsonian disor- 1692
 1628 ders. *Front Neurol* 6:91
 1629 155. Tsukamoto K, Matsusue E, Kanasaki Y et al (2012) Significance 1694
 1630 of apparent diffusion coefficient measurement for the differential 1695
 1631 diagnosis of multiple system atrophy, progressive supranuclear 1696
 1632 palsy, and Parkinson's disease: evaluation by 3.0-T MR imaging. 1697
 1633 *Neuroradiology* 54:947–955
 1634 156. Hansson O, Janelidze S, Hall S et al (2017) Blood-based NfL: a 1699
 1635 biomarker for differential diagnosis of parkinsonian disorder. 1700
 1636 *Neurology* 88:930–937
 1637 157. Parnetti L, Gaetani L, Eusebi P et al (2019) CSF and blood bio- 1701
 1638 markers for Parkinson's disease. *Lancet Neurol* 18(6):573–586
 1639 158. Marques TM, van Rumund A, Oeckl P et al (2019) Serum NFL 1702
 1640 discriminates Parkinson disease from atypical parkinsonisms. 1703
 1641 *Neurology* 92(13):e1479–e1486
 1642 159. Oosterveld LP, Verberk IMW, Majbour NK et al (2020) CSF or 1704
 1643 serum neurofilament light added to α -synuclein panel discrimi- 1705
 1644 nates Parkinson's from controls. *Mov Disord* 35(2):288–295
 1645 160. Ng ASL, Tan YJ, Yong ACW et al (2020) Utility of plasma 1706
 1646 neurofilament light as a diagnostic and prognostic biomarker of 1707
 1647 the postural instability gait disorder motor subtype in early 1708
 1648 Parkinson's disease. *Mol Neurodegener* 15(1):33
 1649 161. Mollenhauer B, Dakna M, Liu T-Y et al (2019) Validation of 1709
 1650 serum neurofilament light chain as a biomarker of Parkinson's 1710
 1651 disease progression. *bioRxiv*. Cold Spring Harbor Laboratory; 1711
 1652 Epub:762237. <https://doi.org/10.1101/762237>
 1653 162. Lin CH, Li CH, Yang KC et al (2019) Blood NfL: a biomarker for 1712
 1654 disease severity and progression in Parkinson disease. *Neurology* 1713
 1655 93(11):e1104–e1111
 1656 163. Rojas JC, Karydas A, Bang J et al (2016) Plasma neurofilament 1714
 1657 light chain predicts progression in progressive supranuclear palsy. 1715
 1658 *Ann Clin Transl Neurol* 3:216–225
 1659 164. Donker Kaat L, Meeter LH, Chiu WZ et al (2018) Serum neuro- 1716
 1660 filament light chain in progressive supranuclear palsy. 1717
 1661 *Parkinsonism Relat Disord* 56:98–101
 1662 165. Wilke C, Bender F, Hayer SN et al (2018) Serum neurofilament 1718
 1663 light is increased in multiple system atrophy of cerebellar type and 1719
 1664 in repeat-expansion spinocerebellar ataxias: a pilot study. *J Neurol* 1720
 1665 265(7):1618–1624
 1666 166. Abdo WF, van de Warrenburg BP, Munneke M et al (2006) CSF 1721
 1667 analysis differentiates multiple-system atrophy from idiopathic 1722
 1668 late-onset cerebellar ataxia. *Neurology* 67:474–479
 1669 167. Li QF, Dong Y, Yang L et al (2019) Neurofilament light chain is a 1723
 1670 promising serum biomarker in spinocerebellar ataxia type 3. *Mol* 1724
 1671 *Neurodegener* 14(1):39
 1672 168. Zeitberger AM, Thomas-Black G, Garcia-Moreno H et al (2018) 1725
 1673 Plasma markers of neurodegeneration are raised in Friedreich's 1726
 1674 ataxia. *Front Cell Neurosci* 12:366
 1675 169. Hayer SN, Liepelt I, Barro C et al (2020) NfL and pNfH are 1727
 1676 increased in Friedreich's ataxia. *J Neurol* 267(5):1420–1430
 1677 170. Johnson CD, Davidson BL (2010) Huntington's disease: progress 1728
 1678 toward effective disease-modifying treatments and a cure. *Hum* 1729
 1679 *Mol Genet* 19(R1):R98–R102
 1680 171. Bates GP, Dorsey R, Gusella JF et al (2015) Huntington disease. 1730
 1681 *Nat Rev Dis Primers* 1:15005
 1682 172. Craufurd D, MacLeod R, Frontali M et al (2015) Diagnostic ge- 1731
 1683 netic testing for Huntington's disease. *Pract Neurol* 15(1):80–84
 1684 173. Constantinescu R, Romer M, Oakes D, Rosengren L, Kiebert K 1732
 1685 (2009) Levels of the light subunit of neurofilament triplet protein 1733
 1686 in cerebrospinal fluid in Huntington's disease. *Parkinsonism Relat* 1734
 1687 *Disord* 15(3):245–248
 174. Niemelä V, Landtblom AM, Blennow K, Sundblom J (2017) Tau 1735
 or neurofilament light-which is the more suitable biomarker for 1736
 Huntington's disease? *PLoS One* 12(2):e0172762
 175. Ross CA, Aylward EH, Wild EJ et al (2014) Huntington disease: 1737
 natural history, biomarkers and prospects for therapeutics. *Nat* 1738
Rev Neurol 10(4):204–216
 176. Wild EJ, Boggio R, Langbehn D et al (2015) Quantification of 1739
 mutant huntingtin protein in cerebrospinal fluid from 1740
 Huntington's disease patients. *J Clin Invest* 125(5):1979–1986
 177. Byrne LM, Rodrigues FB, Blennow K et al (2017) Neurofilament 1741
 light protein in blood as a potential biomarker of neurodegenera- 1742
 tion in Huntington's disease: a retrospective cohort analysis. 1743
Lancet Neurol 16(8):601–609
 178. Rodrigues FB, Byrne LM, Wild EJ (2018) Biofluid biomarkers in 1744
 Huntington's disease. *Methods Mol Biol* 1780:329–396
 179. Johnson EB, Byrne LM, Gregory S et al (2018) Neurofilament 1745
 light protein in blood predicts regional atrophy in Huntington dis- 1746
 ease. *Neurology* 90(8):e717–e723
 180. Byrne LM, Rodrigues FB, Johnson EB et al (2018) Evaluation of 1747
 mutant huntingtin and neurofilament proteins as potential markers 1748
 in Huntington's disease. *Sci Transl Med* 10(458):eaat7108
 181. Uttley L, Carroll C, Wong R, Hilton DA, Stevenson M (2020) 1749
 Creutzfeldt-Jakob disease: a systematic review of global inci- 1750
 dence, prevalence, infectivity, and incubation. *Lancet Infect Dis* 1751
 20(1):e2–e10
 182. Parchi P, Giese A, Capellari S et al (1999) Classification of spo- 1752
 radic Creutzfeldt-Jakob disease based on molecular and phenotypic 1753
 analysis of 300 subjects. *Ann Neurol* 46(2):224–233
 183. Zerr I, Kallenberg K, Summers DM et al (2009) Updated clinical 1754
 diagnostic criteria for sporadic Creutzfeldt-Jakob disease. *Brain* 1755
 132:2659–2668
 184. Zanusso G, Fiorini M, Ferrari S et al (2011) Cerebrospinal fluid 1756
 markers in sporadic Creutzfeldt-Jakob disease. *Int J Mol Sci* 12: 1757
 6281–6292
 185. Steinacker P, Blennow K, Halbgebauer S et al (2016) 1758
 Neurofilaments in blood and CSF for diagnosis and prediction 1759
 of onset in Creutzfeldt-Jakob disease. *Sci Rep* 6:38737
 186. Zerr I, Schmitz M, Karch A et al (2018) Cerebrospinal fluid neu- 1760
 rofilament light levels in neurodegenerative dementia: evaluation 1761
 of diagnostic accuracy in the differential diagnosis of prion dis- 1762
 eases. *Alzheimers Dement* 14(6):751–763
 187. Abu-Rumeileh S, Capellari S, Stanzani-Maserati M et al (2018) 1763
 The CSF neurofilament light signature in rapidly progressive neu- 1764
 rodegenerative dementias. *Alzheimers Res Ther* 10(1):3
 188. Thompson AGB, Luk C, Heslegrave AJ et al (2018) 1765
 Neurofilament light chain and tau concentrations are markedly 1766
 increased in the serum of patients with sporadic Creutzfeldt- 1767
 Jakob disease, and tau correlates with rate of disease progression. 1768
J Neurol Neurosurg Psychiatry 89(9):955–961
 189. Kovacs GG, Andreasson U, Liman V et al (2017) Plasma and 1769
 cerebrospinal fluid tau and neurofilament concentrations in rapidly 1770
 progressive neurological syndromes: a neuropathology-based co- 1771
 hort. *Eur J Neurol* 24(11):1326–1e77
 190. Staffaroni AM, Kramer AO, Casey M et al (2019) Association of 1772
 blood and cerebrospinal fluid tau level and other biomarkers with 1773
 survival time in sporadic Creutzfeldt-Jakob disease. *JAMA* 1774
Neurol 76(8):969–977
 191. Ladogana A, Kovacs GG (2018) Genetic Creutzfeldt-Jakob dis- 1775
 ease. *Handb Clin Neurol* 153:219–242
 192. Kim MO, Takada LT, Wong K, Forner SA, Geschwind MD 1776
 (2018) Genetic PrP prion diseases. *Cold Spring Harb Perspect* 1777
Biol 10(5):a033134
 193. Vallabh SM, Minikel EV, Schreiber SL, Lander ES (2020) 1778
 Towards a treatment for genetic prion disease: trials and bio- 1779
 markers. *Lancet Neurol* 19(4):361–368 1780

- 1753 194. Baldacci F, Lista S, Vergallo A, Palermo G, Giorgi FS, Hampel H
1754 (2019) A frontline defense against neurodegenerative diseases: the
1755 development of early disease detection methods. *Expert Rev Mol*
1756 *Diagn* 19(7):559–563
- 1757 195. Ashton NJ, Hye A, Rajkumar AP et al (2020) An update on blood-
1758 based biomarkers for non-Alzheimer neurodegenerative disorders.
1759 *Nat Rev Neurol* 16(5):265–284
- 1760 196. Hampel H, Lista S (2013) Use of biomarkers and imaging to
1761 assess pathophysiology, mechanisms of action and target engage-
1762 ment. *J Nutr Health Aging* 17(1):54–63
- 1763 197. Hampel H, O'Bryant SE, Castrillo JI et al (2016) Precision med-
1764 icine - the golden gate for detection, treatment and prevention of
1765 Alzheimer's disease. *J Prev Alzheimers Dis* 3(4):243–259
- 1766 198. Hampel H, Toschi N, Babiloni C et al (2018) Revolution of
1767 Alzheimer precision neurology. *Passageway of systems biology*
1768 and neurophysiology. *J Alzheimers Dis* 64(s1):S47–S105
199. Hampel H, O'Bryant SE, Durrleman S et al (2017) A precision
1769 medicine initiative for Alzheimer's disease: the road ahead to
1770 biomarker-guided integrative disease modeling. *Climacteric*
1771 20(2):107–118
200. Hampel H, Vergallo A, Aguilar LF et al (2018) Precision pharma-
1773 cology for Alzheimer's disease. *Pharmacol Res* 130:331–365
1774
201. Hampel H, Vergallo A, Perry G, Lista S, Alzheimer Precision
1775 Medicine Initiative (APMI) (2019) The Alzheimer Precision
1776 Medicine Initiative. *J Alzheimers Dis* 68(1):1–24
1777
- Publisher's Note** Springer Nature remains neutral with regard to jurisdic-
1778 tional claims in published maps and institutional affiliations.
1779

UNCORRECTED PROOF

AUTHOR QUERIES

AUTHOR PLEASE ANSWER ALL QUERIES.

- Q1. Please check if the affiliations are captured and presented correctly.
- Q2. Please check if the section headings are assigned to appropriate levels.
- Q3. Please check the provided expanded form for the abbreviation “MMSE” if correct.
- Q4. Please check the word “diseases” inserted after the word “neurodegenerative,” which can be found in the sentence starting “Finally, serum NFL levels were reported relatively lower,” for completeness if correct.
- Q5. References [19, 174], [22, 186], [200, 203] based on original manuscript we received were identical. Hence, the latter was deleted and reference list and citations were adjusted. Please check if appropriate.
- Q6. Please provide complete bibliographic details of this reference.

UNCORRECTED PROOF